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Research Article

Synthesis and Antimicrobial Activities of 5-Arylidene-thiazolidine-2,4-dione Derivatives

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Antibiotic resistance is considered one of the world's major public health concerns. The main cause of bacterial resistance is the improper and repeated use of antibiotics. To alleviate this problem, new chemical substances against microorganisms are being synthesized and tested. Thiazolidines are compounds having many pharmacological activities including antimicrobial activities. For this purpose some thiazolidine derivatives substituted at position 5 in the thiazolidine nucleus were synthesized and tested against several microorganisms. Using a disc diffusion method, antimicrobial activity was verified against Gram-positive, Gramnegative, and alcohol acid resistant bacteria and yeast. The minimum inhibition concentrations (MIC) and minimum bactericidal concentrations (MBC) were determined. All derivatives showed antimicrobial activity mainly against Gram-positive bacteria, with MIC values ranging from 2 to $16 \,\mu g/mL$.

1. Introduction

Thiazolidine is a class of compounds which merit special attention because it belongs to a group of substances with activity in medicinal chemistry. This nucleus is associated with antibacterial, antifungal, antiviral, antituberculosis, anticancer, and antiparasite biological activities [1–4]. The use of new synthetic methods and structure-activity relationship studies has made possible a broad study of new drugs with different actions. The computational search for possible mechanisms of 4-thiazolidinones anticancer activity has been studied together with the medical chemistry which permits the optimization of existing drugs [5].

Currently, very important groups of heterocyclic compounds such as thiazolidine-2,4-dione, 2-imino-4-thiazolidinone, 4-thioxo-thiazolidine-2,4-dione, and 2-thioxo-1,3-thiazolidine-4-one (rhodanine) have attracted the attention of researchers in their search for new agents with specific pharmacological properties [6–14].

Multiple antibiotic resistant bacteria represent a challenge in the treatment of infections. It is imperative, therefore, that new substances with antimicrobial properties be found to fight these microorganisms [15]. To be considered a bacterium resistant to a certain antibiotic, the microorganism should be able to grow in vitro when subjected to an inhibitory concentration equal to that obtained in the blood.

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However, the concentration of several antibiotics in the bloodstream can be much lower than that achieved by the same antibiotic in other body tissues or fluids. Thus, a bacterium could be "resistant" to a certain antibiotic when it is present in the bloodstream but "sensitive" when it is in the urinary tract or vice versa [16].

Bacteria become resistant to chemotherapeutic agents by three main mechanisms: destruction or inactivation of the drug, prevention of the penetration of the target site within the microbe, and alteration of drug target sites. There may be variations in these mechanisms [17]. With increasing bacterial resistance to antibiotics, attention has become focused on the development of new derivatives to be used as antimicrobial therapy in infection control [18]. Antibiotics are substances produced synthetically by bacteria and fungi with the function of suppressing the growth of microorganisms [19]. Currently, new antibiotics are needed for the treatment of multidrug resistant bacteria. The clinical use of new drugs has decreased since the 1980s, due to a reduction in the discovery of new, more efficient, and less toxic drugs by pharmaceutical companies around the world. Other research groups are worried about the rise in recurrence of many infectious diseases and the lack of new drugs and development of new antimicrobial products in the face of increasing resistance to existing agents [20].

The literature reports on the results of a number of biological activities when the substituents and their positions on the thiazolidine ring are changed [21]. In this case, medicinal chemistry is an important aid in the discovery of new active molecules using small heterocyclic rings to increase the biological activity of certain nuclei [22]. Due to the importance of the core of the thiazolidine ring, eleven compounds have been synthesized by our research group, some products already known and others unknown by introducing arylidene groups at the position 5 of the thiazolidine ring in order to test the antimicrobial activity of each compound against nine different microorganisms.

2. Material and Methods

2.1. Chemistry. The chemical reagents were supplied by Sigma-Aldrich (USA) and were used without further purification. Purity of the compounds was checked using thin layer chromatography (TLC) plates (silica gel G) in the appropriated system for each compound. The spots were located with short (254 nm)/long (365 nm) UV wavelength.

All melting points were measured in a capillary tube on a Quimis apparatus. Infrared spectra of 1% KBr pellets were recorded using a Bruker IFS66 spectrometer. ¹H NMR and ¹³C NMR spectra were measured on a VARIAN VNMRS 400-MR, using 400 MHz for ¹H and 75.4 MHz for ¹³C in CDCl₃ and acetone-d₆ maintained at 25°C using Me₄Si (TMS) as an internal standard. The chemical shifts were reported in δ units and the coupling constants (*J*) were reported in hertz. The following abbreviations were used to indicate the peak multiplicity: s (singlet), d (doublet), dd (double doublet), t (triplet), and m (multiplet). C, H, N, and S analyses were performed with a Carlo Erba elemental analyzer, model

(a)
$$H_2N$$
- C - NH_2 + $CICH_2$ - $COOH$ H_2O $Reflux$ O

(b)
$$\begin{array}{c} O \\ N \end{array}$$
 $\begin{array}{c} H \\ CHO \\ \hline Piperidine \end{array}$ $\begin{array}{c} H \\ R \end{array}$ $\begin{array}{c} O \\ N \end{array}$ $\begin{array}{c} H \\ C \\ S \end{array}$ $\begin{array}{c} O \\ O \end{array}$

$$\label{eq:R2} \begin{split} R=&\textbf{2a})\ 3\text{-}OCH_3,\ 4\text{-}OH;\ \textbf{2b})\ 2,4\text{-}Cl_2;\ \textbf{2c})\ 3,4\text{-}Cl;\ \textbf{2d})\ 4\text{-}OH;\\ \textbf{2e})\ 3\text{-}OCH_3;\ \textbf{2f})\ 3\text{-}CH_3;\ \textbf{2g})\ 3\text{-}NO_2;\ \textbf{2h})\ 4\text{-}NO_2;\ \textbf{2i})\ 3\text{-}Cl \end{split}$$

Scheme 1: Substituent in the synthesis of 5-arylidene-thiazolidine-2,4-dione (2a-i).

EA1108. Mass spectra were recorded on a Varian MAT 711 spectrometer at an electron impact of 70 eV. The synthesized compounds are shown in Scheme 1, showing the respective substituent.

- 2.2. Procedure for Preparation of the Synthesis of Thiazolidine-2,4-dione (1). Compound (1) (thiazolidine-2,4-dione) was synthesized by refluxing monochloroacetic acid and thiourea in water. This compound was synthesized according to a published procedure [16]. Molecular formula $C_3H_3O_2NS$; yield 78%; mp 118–120°C; Rf 0.48 (0.9:0.1 CHCl₃/MeOH). Recrystallization: water.
- 2.3. General Method for the Synthesis of 5-Arylidene-thiazolidine-2,4-dione (2a-i). The compound (2a-i) was synthesized from a mixture of thiazolidine-2,4-dione (1) (2.5 g, 21.36 mmol), aldehyde (21.36 mmol), piperidine (14.11 mmol), and ethanol (150 mL). The reaction mixture was heated under reflux and continuously stirred for a period of 8-9 h. The course of the reaction was monitored by TLC. The reaction mixture was poured into water and acidified with acetic acid. The resulting precipitate was filtered off and recrystallized from acetic acid to give (2a-i).
- 2.4. Microbiological Activity. Gram-positive, Gram-negative, and alcohol acid resistant bacteria and yeast were selected for the examination of antimicrobial activity in vitro. For the evaluation of the antimicrobial activity, first a disc diffusion test was used to screen the antimicrobial activity of all compounds. The compounds that had inhibition zones greater than 10 mm of diameter were submitted to a second test for the determination of minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC). The tests were performed as follows.
- 2.4.1. Disk Diffusion Method. Antimicrobial activity was evaluated by disc diffusion method according to Bauer et al. [23]

against Gram-positive bacteria (Staphylococcus aureus—DAUFPE 01, Micrococcus luteus—DAUFPE 06, Bacillus subtilis—DAUFPE 16, and Enterococcus faecalis—DAUFPE 138), Gram-negative bacteria (Pseudomonas aeruginosa—DAUFPE 39, Escherichia coli—DAUFPE 224, and Serratia marcescens—DAUFPE 398), acid alcohol resistant bacteria (Mycobacterium smegmatis—DAUFPE 71), and yeast (Candida albicans—DAUFPE 1007) obtained from the culture collection of the Department of Antibiotics at the Federal University of Pernambuco (UFPE), Brazil.

Paper discs (Whatman number 2) with 6 mm diameters were impregnated with 20 μ L of a 15,000 μ g/mL solution of the synthesized compounds and dissolved in DMSO. The discs were then placed in medium sown with one of the microorganisms. The following standard drugs were used as controls: ketoconazole (Neoquímica, Brazil), for yeast and cefalexin (Eurofarma, Brazil) and kanamycin (Centro de Controle e Produtos para Diagnósticos—CECON, Brazil) for bacteria. The tests were carried out 3 times, with the results expressed (in millimeters) as mean \pm SEM of the diameters of the inhibition zones formed around the discs. The negative control test was carried out with DMSO soaked discs.

2.4.2. Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC). A multiwell plate (96 wells) was used to determine the minimum inhibitory concentration (MIC) following the criteria adopted by the Clinical and Laboratory Standards Institute (CLSI) [24]. Tests were carried out in Müeller-Hinton broth and Sabouraud dextrose broth (Difco, USA) at pH 7.4 and the twofold serial dilution technique was applied. A 1,280 µg/mL stock solution was prepared from the product. A standardized suspension of microorganisms was prepared for use with a 0.5 tube on the McFarland scale. The plate was incubated for 18 hours and thereafter an indicator dye resazurin (Sigma-Aldrich, USA) was applied to show if there was microbial growth in the well. MIC was determined as the concentration of the last well where there was no microbial growth. From this experiment, the content of the wells was sown on plates with Müeller-Hinton agar medium to establish the minimum bactericidal concentration (MBC), which is the concentration where there is no colony growth. All analyses were performed in triplicate.

3. Result and Discussion

3.1. Synthesis

5-(3-Methoxy-4-hydroxy-arylidene)-thiazolidine-2,4-dione (2a). Yield 70%; mp 260°C; Rf 0.50 (CHCl₃/MeOH 9:1). Recrystallization: ethanol; IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1566 (C=C); 1730–1670 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.48 (s 1H, NH); 7.71 (s 1H, CH=); 3.77 (s 3H, OCH₃); 6.02 (s 1H, OH). 6.92 (s 1H₍₂₎); 6.86 (d 1H₍₅₎ J = 8.33); 6.91 (d 1H₍₆₎ J = 8.34); ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.81 (C=O₂); 166.89 (C=O₄); 119.55 (C₅, heterocycle); 143.28 (CH=); 56.50 (OCH₃); 129.20 (C₁); 112.58 (C₂), 148.06 (C₃); 144.76 (C₄); 115.56 (C₅); 122.30 (C₆). Anal. Calcd. for

 $C_{11}H_9NO_4S$: C, (52.58%); H, (3.61%); N, (5.57%). Found: C, 53.01%; H, 3.83%; N, 5.25%. HRMS⁺: calcd 251.0252; found 251.0253.

5-(2,4-Dichloro-arylidene)-thiazolidine-2,4-dione (2b). Yield 65%; mp 203°C; Rf 0.51 (CHCl₃/MeOH 9.6 : 0.4). Recrystallization: ethanol; IR (KBr 1%, $\nu_{\rm max}$ cm $^{-1}$) 1570 (C=C); 1737–1675 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.50 (s 1H, NH); 7.35 (s 1H, CH=); 7.55 (s 1H, H₍₃₎7.44 (d 1H, H₍₅₎ J=7.90); 7.39 (d 1H (H₍₆₎ J=7.90). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.85 (C=O₂); 166.86 (C=O₄); 120.20 (C₅ heterocycle); 140.23 (CH=); 133.61 (C₁); 134.84 (C₂); 129.52 (C₃); 134.93 (C₄); 127.52 (C₅); 135.80 (C₆). Anal. Calcd. for C₁₀H₅Cl₂NO₂S: C, 43.81%; H, 1.84%; N, 5.11%. Found: C, 44.01%; H, 1.96%; N, 5.05%. HRMS⁺: calcd 272.9418; found 272.9419.

5-(3,4-Dichloro-arylidene)-thiazolidine-2,4-dione (2c). Yield 59%; mp 174°C; Rf 0.50 (CHCl₃/MeOH 9.5:0.5). Recrystallization: ethanol; IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1571 (C=C); 1735–1676 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.50 (s 1H, NH); 7.77 (s 1H, CH=); 7.52 (s 1H, H₍₂₎; 7.38 (d 1H, H₍₅₎ J=8.91); 7.52 (d 1H (H₍₆₎ J=8.92). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.89 (C=O₄); 119.17 (C₅ heterocycle); 143.13 (CH=); 134.42 (C₁); 129.49 (C₂); 133.15 (C₃); 131.79 (C₄); 130.71 (C₅); 127.26 (C₆). Anal. Calcd. for C₁₀H₅Cl₂NO₂S: C, 43.81%; H, 1.84%; N, 5.11%. Found: C, 43.91%; H, 1.97%; N, 5.07%. HRMS⁺: calcd 272.9418; found 272.9419.

5-(4-Hidroxy-arylidene)-thiazolidine-2,4-dione (2d). Yield 45%; mp 149-150°C; Rf 0.49 (CH₂Cl₂/Hex. 5:5). Recrystallization: methanol; IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1570 (C=C); 1734–1677 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.49 (s 1H, NH); 7.54 (s 1H, CH=); 6.40 (s 1H, OH); 7.31 (d 1H, H₍₂₎-H₍₆₎ J=8.65); 7.21 (d 1H, H₍₃₎-H₍₅₎ J=8.65). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.89 (C=O₄); 119.55 (C₅ heterocycle); 140.25 (CH=); 129.15 (C₁); 129.15 (C₂)-(C₆); 115.96 (C₃)-(C₅); 157.71 (C₄). Anal. Calcd. for C₁₀H₇NO₃S: C, 54.29%; H, 3.19%; N, 6.33%. Found: C, 54.93%; H, 3.23%; N, 6.11%. HRMS⁺: calcd 221.0147; found 272.0148.

5-(4-Methoxy-arylidene)-thiazolidine-2,4-dione (2e). Yield 40%; mp 210–212°C; Rf 0.48 (CHCl₃/MeOH 9.6:0.4). Recrystallization: CHCl₃; IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1575 (C=C); 1734–1676 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.49 (s 1H, NH); 7.74 (s 1H, CH=); 3.65 (s 3H, OCH₃); 7.35 (d 1H, H₍₂₎-H₍₆₎ J = 8.65); 6.75 (d 1H (H₍₃₎-H₍₅₎ J = 8.65). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.89 (C=O₄); 119.50 (C₅ heterocycle); 142.55 (CH=); 55.67 (OCH₃); 128.42 (C₁); 129.37 (C₂)-(C₆); 114.32 (C₃)-(C₅); 160.31 (C₄). Anal. Calcd. for C₁₁H₉NO₃S: C, 56.16%; H, 3.86%; N; 5.95%. Found: C, 56.98%; H, 3.99%; N, 6.45%. HRMS⁺: calcd 235.0303; found 235.0303.

5-(3-Methyl-arylidene)-thiazolidine-2,4-dione (2f). Yield 52%; mp 195-196°C; Rf 0.50 (Hex./Ethyl acetate 8.5:1.5).

Recrystallization: acetic acid; IR (KBr 1%, $ν_{\rm max}$ cm⁻¹) 1573 (C=C); 1730–1739 (C=O); ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.49 (s 1H, NH); 7.73 (s 1H, CH=); 2.36 (s, 3H CH₃); 7.25 (s 1H H₍₂₎); 6.95 (d 1H H₍₄₎ J=7.71); 7.31 (t, 2H H₅, J=7.72); 7.31 (d 1H H₆ J=7.52). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 167.89 (C=O₄); 119.55 (C₅ heterocycle); 143.18 (CH=); 21.20 CH₃; 136.18 (C₁); 128.93 (C₂); 133.87 (C₃); 126.60 (C₄); 129.51 (C₅); 125.93 (C₆). Anal. Calcd. for C₁₁H₉NO₂S: C, 60.26%; H, 4.14%; N, 6.39%. Found: C, 60.66%; H, 4.45%; N, 6.11%; HRMS⁺: calcd 219.0354; found 272.0353.

5-(3-Nitro-arylidene)-thiazolidine-2,4-dione (2**g**). Yield 80%; mp 250°C; Rf 0.51 (CHCl₃/MeOH 9.6 : 0.4). Recrystallization: acetic acid. IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1571 (C=C); 1740–1667 (C=O); ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.49 (s 1H, NH); 7.37 (s 1H, CH=); 8.35 (s 1H H₍₂₎) 7.92 (d 1H H₍₄₎, J=8.0); 7.54 (t, 2H H₅, J=7.72); 7.77 (d 1H H₆ J=7.7). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.88 (C=O₄); 119.55 (C₅ heterocycle); 143.65 (CH=); 136.25 (C₁); 123.06 (C₂); 148.74 (C₃); 122.14 (C₄); 130.16 (C₅); 131.91 (C₆). Anal. Calcd. for C₁₀H₆N₂O₄S: C, 48.00%; H, 2.42%; N, 11.20%. Found: C, 48.35%; H, 2.1%; N, 11.01%; HRMS⁺: calcd 250.0048; found 250.0051.

5-(4-Nitro-arylidene)-thiazolidine-2,4-dione (2h). Yield 69%; mp 220–223°C; Rf 0.52 (CHCl₃/MeOH 9.6 : 0.4). Recrystallization: THF; IR (KBr 1%, $\nu_{\rm max}$ cm⁻¹) 1575 (C=C); 1734–1676 (C=O). ¹H NMR (CDCl₃,300 MHz, δ ppm): 8.49 (s 1H, NH); 7.74 (s 1H, CH=); 7.63 (d 1H, H₍₂₎–H₍₆₎ J=8.65); 8.09 (d 1H (H₍₃₎–H₍₅₎ J=8.65). ¹³C NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.89 (C=O₄); 119.50 (C₅ heterocycle); 142.55 (CH=); 141.50 (C₁); 129.37 (C₂)–(C₆); 124.13 (C₃)–(C₅); 147.21 (C₄); Anal. Calcd. for C₁₀H₆N₂O₄S: C, 48.00%; H, 2.42%; N, 11.20%. Found: C, 48.35%; H, 2.1%; N, 11.01%; HRMS⁺: calcd 250.0048; found 250.0049.

5-(3-Chloro-arylidene)-thiazolidine-2,4-dione (2i). Yield 54%; mp 208-209°C; Rf 0.46 (Hexane/Ethyl acetate 4:1). Recrystallization: ethyl acetate; IR (KBr 1%, $\nu_{\rm max}$ cm $^{-1}$) 1582 (C=C); 1729–1672 (C=O). $^1{\rm H}$ NMR (CDCl₃,300 MHz, δ ppm): 8.48 (s 1H, NH); 7.77 (s 1H, C=H); 7.34 (s 1H₍₂₎); 7.18 (d 1H₍₄₎ (*J* = 8.07); 7.36 (t 2H₍₅₎ (*J* = 8.07); 7.36 (d 1H₍₆₎ *J* = 2.27); $^{13}{\rm C}$ NMR (Acetone-d₆, 75.4 MHz, δ ppm): 169.86 (C=O₂); 166.46 (C=O₄); 119.55 (C₅ heterocycle); 143.09 (CH=); 135.81 (C₁); 127.72 (C₂); 133.85 (C₃); 128.62 (C₄); 130.25 (C₅); 125.78 (C₆). Anal. Calcd. for C₁₀H₆ClNO₂S: C, 50.11%; H, 2.52%; N, 5.84%. Found: C, 50.55%; H, 2.67%; N, 5.57%. HRMS⁺: calcd 238.9808; found 238.9810.

The compounds (2a-i) were synthesized by Knoevenagel condensation with nine aromatic aldehydes yielding 5-arylidene-thiazolidine-2,4-dione derivatives. The infrared spectrum of these compoundsshowed a strong absorption band of the functional group ranging between 1566 at 1775 cm⁻¹ corresponding to C=C and 1775–1676 cm⁻¹ concerning the carbonyl in positions 2 and 4 of the thiazolidine-2,4-dione ring [25]. The molecular structures of the compounds were established by ¹H NMR and ¹³C NMR

spectra which exhibited the presence of the signals corresponding to hydrogen and carbons in the molecule. The compound (2a) presented a singlet at 3.77 ppm of 3 hydrogen referent to OCH3 and a singlet at 6.02 concerning the absorption of the OH group. This substituent also presented absorption at 56.50 ppm of the OCH₃ in ¹³C NMR. The group OCH₃in (2d) presented signal in 3.77 ppm to hydrogen and 56.50 to absorption in ¹³C NMR. The absorption of hydrogen as a singlet at 3.65 ppm of the OCH₃ group at position 4 of arylidene and 55.67 to ¹³C NMR was sufficient to characterize the compound (2e). The compound (2f) presented absorption as a singlet at 2.36 ppm to hydrogen in methyl group at position 3 and 20.15 ppm to ¹³C NMR in the molecule. The ¹H NMR spectra of compounds showed only one kind of proton absorption for CH= in the range 7.71-7.78. According to the literature, this proton absorption is the compound represented by the configuration Z, as illustrated by the absorption of the CH= group. Isomer (Z) appears to be more thermodynamically stable than isomers E [26].

The ¹H NMR spectrum of compounds in CDCl₃-d6 revealed the presence of methyne moiety and the position of the hydrogen of the aromatic group. In addition, the structures were supported by their mass spectrum which revealed a molecular ion peak. The elemental analysis was also important to supplement the characterization of the chemical structure.

3.2. Antimicrobial Activity. The results of the disc diffusion test are shown in Table 1. Most of the synthesized compounds were active against Gram-positive bacteria. MIC and MBC values are shown in Table 2.

Compound (1) thiazolidine-2,4-dione showed no antimicrobial activity against any of the tested microorganisms [12]. When it reacted with aromatic benzaldehydes forming derivatives substituted at position 5, however, all derivatives showed antimicrobial activity. This affirmation is similar to what is in the literature, showing that substitution at position 5 with arylidene groups has an important effect on enhancing the antimicrobial properties [14].

This shows the importance of the dual CH= bond at position 5, thus introducing a pharmacophore group in the molecule. This group generally influences the specific structural changes in biological effects, stereochemistry, size, and shape of the carbon chain as well as the nature of the other substituent. The compound (2a) with substituents 3-OCH₃ and 4-OH of the arylidene ring was the most active of all, showing better inhibition for almost all microorganisms tested. This compound inhibited all Gram-positive bacteria tested and, additionally, was the only one that inhibited the growth of Gram-negative bacteria, alcohol acid resistant bacteria, and also the yeast, as shown in Table 1. Generally, 4-thiazolidinediones derivatives have demonstrated antimicrobial activity against Gram-positive and Gram-negative bacteria [14].

Compounds (**2b**) and (**2c**), containing the substituents 2,4-Cl₂ and 3,4-Cl₂, respectively, inhibited all Gram-positive bacteria tested and also the alcohol acid resistant bacteria.

TABLE 1: Antimicrobial activity of compounds tested against different microorganisms (diameter of inhibition halo in mm).

						Microorganisms				
						•			Alcohol acid	
Synthetized c	Synthetized compounds		Gram-positive bacteria	ıcteria		Gram-	Gram-negative bacteria		resistant bacteria	Yeast
3000	() (II3C)	S. aureus	M. luteus	E. faecalis	B. subtilis	P. aeruginosa	E. coli	S. marcescens	M. smegmatis	C. albicans
и	Subs.	DAUFPE	DAUFPE	DAUFPE	DAUFPE	DAUFPE	DAUFPE	DAUFPE	DAUFPE	DAUFPE
		01	90	138	16	39	224	398	71	1007
1	1	0	0	0	0	0	0	0	0	0
2a	$3-0$ CH $_3$	19.3 ± 0.47	27.8 ± 0.16	11.2 ± 0.62	15.0 ± 0.16	15.0 ± 0.16	0	0	19.3 ± 0.77	16.2 ± 0.82
2 b	2,4-Cl	11.7 ± 0.56	32.0 ± 0.81	18.8 ± 0.85	30.7 ± 0.47	0	0	0	19.2 ± 0.41	0
2c	3,4-Cl	19.4 ± 0.08	34.0 ± 0.80	8.4 ± 0.17	25.4 ± 0.05	0	0	0	12.1 ± 0.09	0
2d	4-OH	0	13.8 ± 0.56	11.0 ± 0.82	0	0	0	0	0	0
2e	$3-0$ CH $_3$	0	12.7 ± 0.56	0	0	0	0	0	0	0
2f	3-CH_3	18.3 ± 0.47	0	18.8 ± 0.16	17.8 ± 0.62	0	0	0	0	0
2g	$3-NO_2$	21.8 ± 0.16	0	22.8 ± 0.70	23.7 ± 0.16	0	0	0	0	0
2h	$4-NO_2$	23.8 ± 0.16	0	21.9 ± 0.17	18.6 ± 0.15		0	0	0	0
2i	3-CI	20.8 ± 0.27	24.8 ± 0.16	21.8 ± 0.16	27.7 ± 0.16	0	0	0	0	0
×	Kan (30 µg/disc)	23.0 ± 0.82	28.3 ± 0.09	0	14.0 ± 0.82	20.0 ± 0.12	15.1 ± 0.19	15.0 ± 0.12	40.0 ± 0.12	I
)	Cef (30 μ g/disc)	35.3 ± 0.09	54.0 ± 0.12	24.1 ± 0.19	45.0 ± 0.12	0	24.3 ± 0.09	0	0	I
K	Ket (300 µg/disc)	Ι	Ι	I	1	I	1	I	I	24.1 ± 0.19

Standard compounds: Cef: cephalexin; Ket: ketoconazole; Kan: kanamycin. —: not tested; X: medium; δ : standard deviation.

Table 2: Inhibition and bactericidal concentrations of compounds (2a-i), with respect to different microorganisms ($\mu g/mL$).

Country attend						Microorganisms	ganisms					
symmetized	S. au	reus	M. luteus	ens	B. subt		M. smegn	ıatis	E. faec	alis	C. albi	cans
compounds	DAUFPE 01	PE 01	DAUFPE 06	SE 06	DAUFPE 16	E 16	DAUFPE 71	E 71	DAUFPE 138	E 138	DAUFP	E 1007
	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MBC	MIC	MIC MBC
2a	16	32	4	8	4	8	16	32	64	>128	∞	32
2b	8	16	2	4	2	4	2	4	16	32	I	I
2c	8	16	2	4	2	4	2	4	16	32	I	I
2d	l		4	8				1			I	I
2e	1		4	8				1			I	I
2f	16	32	16	32	4	∞		1			I	I
2g	8	16	∞	16	16	32	I	I	I	I	I	
2 h	8	16	∞	16	I	I	I	1	1	I	I	I
2 i	8	16	16	32	4	∞		1		1	I	I
Cefalexin	8	16	<2	2	<2>	<2	>128	>128	>128	>128	I	
Ketoconazole											<2	2

MIC: minimum inhibitory concentration; MBC: minimum bactericidal concentration; —: not tested.

However, compound (2c) showed halos slightly smaller than (2b).

The compound (2d) with 4-OH group was active only for two Gram-positive bacteria, M. Luteus and E. faecalis, while (2e), substituted with 3-OCH₃, was only active against M. luteus. This fact indicates that the groups OH and OCH₃ separately do not contribute to a significant antimicrobial activity. This fact is reinforced by the activity of compound (2a), which had both groups (3-OCH₃ and 4-OH), against all tested microorganisms, including the Gram-negative bacterium P. aeruginosa. The compounds (2f, 2g, and 2h) were active only for Gram-positive bacteria, except E. faecalis, with halos between 17.8 \pm 0.62 and 23.8 \pm 0.16 mm. The compound (2i) also inhibited the growth of all Gram-positive bacteria.

In general, all compounds tested were active against Gram-positive bacteria presenting values of 16 and 32 µg/mL for S. aureus and 4 and 8 µg/mL for M. luteus and B. subtilis, for MIC and MBC, respectively. MIC and MBC values for *M*. smegmatis were 16 and 32 µg/mL, smaller than the standard drugs. The compounds (2b, 2c, 2g, 2h, and 2i) showed MIC and MBC values for S. aureus near those obtained for the standard drug, cefalexin. The compounds (2b) and (2c) showed MIC and MBC for M. Luteus and B. subtilis also near the standard drug. The results for M. smegmatis 16 and 32 µg/mL were lower than the standard Kanamycin (128 μ g/mL) showing significant activity in inhibiting these microorganisms. Recent studies with 5-hydroxy-arylidene derivatives showed MIC values higher than 128 µg/mL for two Gram-negative bacteria, E. coli and P. aeruginosa, and 128 μg/mL for *E. faecalis* and *S. aureus* [7].

These results showed that these substituents separately did not have significant antimicrobial activity. The groups chlorine, nitro, and methyl at position 3 on the thiazolidine ring seem to favor this activity for other positions [7]. This affirmation reveals that electrons donating or withdrawing groups in molecules increase the lipophilicity of the compounds, which alters the permeability across the bacterial cell membrane [27]. These molecules showed the greatest inhibition among the tested compounds. The compounds (2b, 2c, and 2i) containing chlorine atoms in the molecule were active for all Gram-positive bacteria, indicating the power of chlorine in antimicrobial activity.

This demonstrates that the substitution of arylidene at position 5 on the thiazolidine ring enhances the antimicrobial activity of the pharmacophore group in these compounds. The condensation of the compounds 5-arylidenethiazolidine-2,4-dione was crucial to generate considerable activity in the synthesized derivatives [8]. The results of this study demonstrate significant antimicrobial activity for compounds 5-arylidene-thiazolidine-2,4-dione substituted (2a-i), with values similar to the drugs tested cephalexin >128, confirming the antimicrobial activity of thiazolidine compounds as described in the literature [1, 27]. These compounds probably act on the synthesis of peptidoglycan from the cell wall, which explains the higher antimicrobial effects on Gram-positive bacteria instead of Gram-negative ones [2, 28].

4. Conclusion

In order to find an antimicrobial agent, nine compounds were synthesized and tested against Gram-positive, Gramnegative, and alcohol acid resistant bacteria and yeast. All compounds were active against all Gram-positive bacteria. One of the compounds was the most active, inhibiting Grampositive, Gram-negative, and alcohol acid resistant bacteria and also the yeast. The other compounds had varied activity between classes of Gram-positive and acid alcohol resistant bacteria. Compounds containing chlorine in the molecule showed the best antibacterial activity, thus demonstrating the power of this atom on the bacteria. The compounds with substituents nitro and methyl also showed significant activity. All the compounds presented the *Z* configuration. The chemical structures of the compounds were determined by physical methods IR, ¹HNMR, ¹³CNMR and mass spectrometry.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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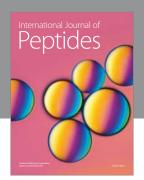
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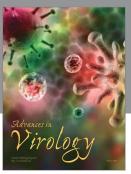
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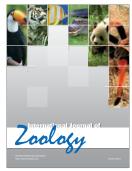
















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