

Synthesis, Herbicidal, Fungicidal and Insecticidal Evaluation of 3-(Dichlorophenyl)-isocoumarins and (±)-3-(Dichlorophenyl)-3,4-dihydroisocoumarins

Ghulam Qadeer,^a Nasim Hasan Rama,^{*a} Zhi-Jin Fan,^b Bin Liu^b and Xiu-Feng Liu^b

^aDepartment of Chemistry, Quaid-I-Azam University, Islamabad 45320, Pakistan

^bState Key Laboratory of Elemento-Organic Chemistry, Tianjin Key Laboratory of Pesticide Science, Nankai University, Tianjin, 300071, P. R. China

Este é o primeiro relato mostrando que 3-(diclorofenil)isocumarinas e (±)-3,4-(dihidroisocumarinas) são inibidores do crescimento de plantas e fungos. 3-Diclorofenil-isocumarinas foram sintetizadas pela condensação de ácido homoftálico com cloretos de diclorobenzoíla. A hidrólise alcalina destas isocumarinas formou ceto ácidos. 3-(Diclorofenil)-3,4-dihidroisocumarinas racêmicas foram obtidas pela redução de ceto ácidos a hidróxi-ácidos racêmicos, seguida pela ciclodehidratação, usando anidrido acético. As atividades herbicida, fungicida e inseticida dos compostos sintetizados foram avaliadas. Alguns dos compostos sintetizados mostraram excelentes atividades fungicida e herbicida, mas nenhum deles apresentou efeito inseticida nos insetos usados nos testes. Este estudo sugere que isocumarinas e compostos relacionados podem ser usados como guias para o planejamento de herbicidas e fungicidas bioativos.

This is the first report showing that 3-(dichlorophenyl)isocoumarins and (±)-3,4-dihydroisocoumarins are plant and plant fungus growth inhibitors. 3-Dichlorophenylisocoumarins were synthesized by condensation of homophthalic acid with dichlorobenzoyl chlorides. The alkaline hydrolysis of these isocoumarins afforded keto acids. Racemic 3-(Dichlorophenyl)-3,4-dihydroisocoumarins were obtained by reduction of keto acids to racemic hydroxy acids, followed by cyclodehydration using acetic anhydride. The herbicidal, fungicidal and insecticidal activities of the synthesized compounds have been evaluated. Some of the synthesized compounds show excellent herbicidal and fungicidal activities but none of the synthesized compounds presented any insecticidal effects on the test insects. The findings of this study suggest that isocoumarins and related compounds may serve as lead compounds towards the design of bioactive herbicides and fungicides.

Keywords: 3-(dichlorophenyl)isocoumarins, (±)-3-(dichlorophenyl)-3,4-dihydroisocoumarin, herbicide, fungicide, insecticide

Introduction

Weeds compete with crops for sunshine, water, nutrients, and physical space and are thus capable of greatly influencing the growth of crops and undermining both crop quality and yield. Also, many weeds harbor or nest pathogens, viruses, and pests, which may result in the occurrence and spread of plant diseases and insect pests in crops. Herbicides, as the main weed control tool, play a very important role in modern agriculture. Since the discovery of the herbicide 2,4-dichlorophenoxyacetic acid (2,4-D) by Zimmerman and Hitchcock, the agrochemical industry has successfully developed a wide array of

herbicides with various chemical structures and modes of action.¹ However, an inevitable problem associated with the use of herbicides is the occurrence of herbicide-resistant weeds.² For example, the widespread use of herbicides, such as chlorsulfuron, atrazine, diclofop-methyl, and paraquat, has caused herbicide resistance in many weeds. Therefore, it is necessary to develop efficient herbicides with novel structures and modes of action to overcome the resistance of weeds.

Over the last two decades, there has been a dramatic increase in the rate of superficial and invasive fungal infections.³⁻⁷ Modern agriculture relies on effective control of fungal diseases to increase crop yield and quality and consequently increase crop value.⁸ No single fungicide can be used for all disease situations and the widespread

*e-mail: nasimhrama@yahoo.com

use of fungicides can select for fungicide resistant pathogens.⁹ Therefore, there is need for safer and more cost-effective fungicides, which are easier to use and provide better performance against resistant pathogens.

Isocoumarins and 3,4-dihydroisocoumarins are the secondary metabolites of a wide variety of fungi, lichens, molds, bacteria, higher plants and insects. The majority of isocoumarins have been isolated from various species of the fungal genera *Artemisia*, *Aspergillus*, *Ceratocystis*, *Fusarium*, *Penicillium*, and *Streptomyces* etc. A number of them are constituents of a few higher plant families. Naturally occurring isocoumarins containing halogens have been seldom reported. Examples of naturally occurring isocoumarins containing fluorine are not known yet. However, a few examples of naturally occurring chlorine- and bromine-containing isocoumarins have been reported. Laresenb,¹⁰ has isolate chlorine-containing metabolite dichlorodiaportin from the cheese-associated cultures of *Penicillium nalgiovense*. 4-Chloro-3-[(4'-fluorophenyl) methoxy]isocoumarin has been found,¹¹ to be quite an effective inhibitor for human Q31 granzyme A, murine and human granzyme A, isolated from cytotoxic T lymphocytes. This isocoumarin derivative has also been found,¹² to be useful in the treatment of emphysema as serine protease inhibitor. 6-(2'-Chloro-4'-trifluoromethylphenoxy)-3,4-dihydroisocoumarin has been used,¹³ as a herbicide, which almost totally controlled the growth of *Schinochloa crusgall*, *Sinapis alba* and other weeds. 7-Amino-4-chloro-3-(2'-bromoethoxy)isocoumarin has been synthesized,¹⁴ and evaluated as a potent inhibitor of human leuko elastase and several blood coagulation enzymes.¹⁵ 7-Amino-3-(2'-bromopropoxy)-4-chloroisocoumarin and 7-Amino-3-(3'-bromopropoxy)-4-chloroisocoumarin have been patented,¹⁶ as an ascapain inhibitor in the inhibition and treatment of neurodegeneration.

In continuation of our previous studies,¹⁷⁻¹⁹ and biological activities associated with chloro-substituted isocoumarins

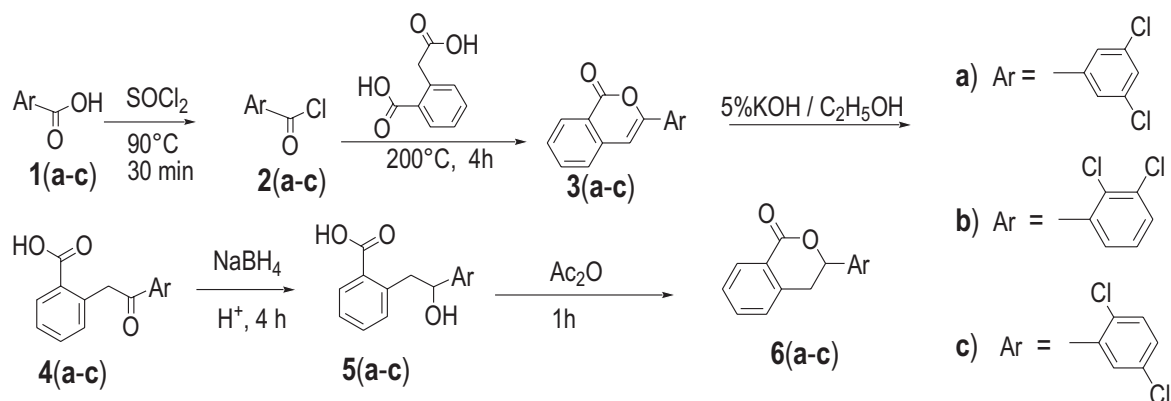
prompted us to synthesize some new 3-(dichlorophenyl) isocoumarins and their conversion to the corresponding (\pm)-3-(dichlorophenyl)-3,4-dihydroisocoumarins in order to check their herbicidal, fungicidal and insecticidal activities.

The aim of our study was to investigate the importance of these compounds in agriculture. The general synthetic scheme is shown as follows.

Results and Discussion

Synthesis

Condensation of the acid chloride with homophthalic acid is useful for the preparation of 3-substituted isocoumarins.¹⁷⁻¹⁹ A short and efficient synthesis of 3-(dichlorophenyl) isocoumarins **3(a-c)** by using this method and their conversion into the corresponding racemic 3-(dichlorophenyl)-3,4-dihydroisocoumarins **6(a-c)** were achieved and the herbicidal, fungicidal and insecticidal activities of the compounds were examined. Dichlorobenzoic acids **1(a-c)** were converted into their respective acid chlorides **2(a-c)** by reaction with thionyl chloride. Direct condensation of the acid chlorides **2(a-c)** with homophthalic acid at 200 °C afforded the 3-(dichlorophenyl)isocoumarins **3(a-c)**, which were purified by column chromatography (eluant was pet. ether that corresponds to the fraction with a boiling range of 40-80 °C) and showed a single spot on TLC. These isocoumarins **3(a-c)** exhibited a characteristic ¹H-singlet at δ 6.95, 6.89 and 7.01 ppm respectively for the C₄-H in the ¹H NMR. The aromatic hydrogens appeared in the expected region, *i.e.* 7.29-8.33 ppm. In the IR spectra of isocoumarins **3(a-c)**, the lactone carbonyl absorptions were observed at 1708, 1704 and 1703 cm⁻¹ respectively. The molecular ion peak in the mass spectrum of the isocoumarins **3(a-c)** was observed at *m/z* 290. The alkaline hydrolysis of isocoumarins **3(a-c)** afforded the



Scheme 1. The synthesis of the target compounds.

was separated, concentrated and chromatographed on silica gel using petroleum ether (40-80 °C) as eluent to afford 3-(dichlorophenyl)isocoumarins **3(a-c)** as solid, which was further purified by recrystallization from methanol.

3-(3',5'-Dichlorophenyl)isocoumarin (**3a**)

Yield: 81%; mp 208-210 °C; IR ν_{\max} / cm^{-1} : 3156(C-H), 2935 (Ar-H), 1708 (C=O), 1590 (C=C arom), 1099 (C-Cl). EI-MS m/z (rel. int.): 290 (M^+ 100), 292 (M^+ +2, 67.5), 294 (M^+ +4, 13); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 6.95 (1H, s, H-4), 7.39 (1H, t, J 1.2 Hz, H-4'), 7.50 (1H, m, H-7), 7.53 (1H, m, H-6), 7.55 (1H, dd, J 2.2, 7.6 Hz, H-5), 7.75 (2H, t, J 1.2 Hz, H-2' and 6'), 8.31 (1H, dd, J 1.9, 7.9 Hz, H-8); Elemental analysis Found: C 61.83, H 2.73, Cl 24.30; Calculated: C 62.07, H 2.76, Cl 24.48.

3-(2',3'-Dichlorophenyl)isocoumarin (**3b**)

Yield: 80%; mp 180-182 °C; IR ν_{\max} / cm^{-1} : 3152 (C-H), 2931(Ar-H), 1710 (C=O), 1593 (C=C arom), 1096 (C-Cl). EI-MS m/z (rel. int.): 290 (M^+ 28), 292 (M^+ +2, 17.5), 294 (M^+ +4, 3.7); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 6.89 (1H, s, H-4), 7.29 (1H, dd, J 1.9, 7.9 Hz, H-4'), 7.33 (2H, t, J 7.8 Hz, H-5'), 7.50 (2H, t, J 7.8 Hz, H-6'), 7.54 (1H, m, H-7), 7.56 (1H, dd, J 2.2, 8.0 Hz, H-5), 7.74 (1H, m, H-6), 8.33 (1H, dd, J 2.1, 7.9 Hz, H-8); Elemental analysis Found: C 61.99, H 2.69, Cl 24.33; Calculated: C 62.07, H 2.76, Cl 24.48.

3-(2',5'-Dichlorophenyl)isocoumarin (**3c**)

Yield: 81%; mp 187-188 °C; IR ν_{\max} / cm^{-1} : 3155(C-H), 2935(Ar-H), 1710 (C=O), 1593 (C=C arom), 1093 (C-Cl). EI-MS m/z (rel. int.): 290 (M^+ 100), 292 (M^+ +2, 69.6), 294 (M^+ +4, 13.7); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.01(1H, s, H-4), 7.32 (1H, dd, J 2.47, 8.56 Hz, H-4'), 7.41 (1H, d, J 8.56 Hz, H-3'), 7.50 (1H, dd, J 2.1, 7.80 Hz, H-5), 7.56 (1H, m, H-7), 7.72 (1H, d, J 2.02 Hz, H-6'), 7.75 (1H, m, H-6), 8.32 (1H, dd, J 2.7, 7.9 Hz, H-8); Elemental analysis Found: C 61.89, H 2.71, Cl 24.40; Calculated: C 62.07, H 2.76, Cl 24.48.

General procedure for 2-(dichlorobenzoylmethyl) benzoic acid **4(a-c)**

A solution of isocoumarins **3(a-c)** in ethanol (50 mL) and 5% potassium hydroxide (100 mL) were refluxed for 4 h. Ethanol was removed from the reaction mixture by distillation. Ice cold water (20 mL) was added and the reaction mixture was acidified with hydrochloric acid. The reaction mixtures were then

extracted with dichloromethane (3 × 20 mL). The extracts were dried (Na_2SO_4) and evaporated to yield crude solids **4(a-c)**, which were recrystallized from methanol.

2-(3',5'-Dichlorobenzoylmethyl) benzoic acid (**4a**)

Yield: 81%; mp 187-188 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3155 (C-H), 2935 (Ar-H), 1710 (C=O), 1593 (C=C arom), 1093 (C-Cl); EI-MS m/z (rel. int.): 308 (M^+ 100), 310 (M^+ +2, 65.6), 312 (M^+ +4, 11.7); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 4.62 (2H, s, H-1'), 7.18 (1H, dd, J 1.56, 7.99 Hz, H-3), 7.24 (1H, m, H-4), 7.52 (1H, m, H-5), 7.56 (1H, dd, J 2.41 Hz, H-4'') 7.94 (2H, d, J 1.76 Hz, H-2'' and 6''), 8.15 (1H, dd, J 1.2, 7.9 Hz, H-6). Elemental Analysis Found: C 58.23, H 3.28, Cl 25.60; Calculated: C 58.44, H 3.25, Cl 25.26.

2-(2',3'-Dichlorobenzoylmethyl)benzoic acid (**4b**)

Yield: 77%; mp 160-162 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3151 (C-H), 2935 (Ar-H), 1712 (C=O), 1590 (C=C arom), 1091 (C-Cl); EI-MS m/z (rel. int.): 308 (M^+ 97), 310 (M^+ +2, 55), 312 (M^+ +4, 13); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 4.64 (1H, s, H-1'), 7.06 (1H, t, J 7.9 Hz, H-5''), 7.20 (1H, dd, J 1.56, 7.99 Hz, H-3), 7.29 (1H, m, H-4), 7.42 (1H, t, J 8.17 Hz, H-4''), 7.63 (1H, m, H-5), 7.81 (1H, dd, J 1.43, 7.78 Hz, H-6''), 8.02 (1H, dd, J 2.23, 8.01 Hz, H-6); Elemental Analysis Found: C 58.30, H 3.23, Cl 25.46; Calculated: C 58.44, H 3.25, Cl 25.26.

2-(2',5'-dichlorobenzoylmethyl) benzoic acid (**4c**)

Yield: 71%; mp 140-142 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3157 (C-H), 2933 (Ar-H), 1711 (C=O), 1590 (C=C arom), 1094 (C-Cl); EI-MS m/z (rel. int.): 308 (M^+ 100), 310 (M^+ +2, 79), 312 (M^+ +4, 19); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 4.63 (1H, s, H-1'), 7.32 (1H, m, H-4), 7.34 (1H, dd, J 1.99, 7.9 Hz, H-3), 7.41 (1H, d, J 8.9 Hz, H-3''), 7.44 (1H, d, J 8.8 Hz, H-4'), 7.58 (1H, m, H-5), 7.97 (1H, d, J 1.78 Hz, H-6''), 8.17 (1H, dd, J 1.67, 7.9 Hz, H-6); Elemental Analysis Found: C 58.33, H 3.20, Cl 25.25; Calculated: C 58.44, H 3.25, Cl 25.26.

General procedure for the preparation of (\pm)-3-(dichlorophenyl)-3,4-dihydroisocoumarins **6(a-c)**

To a solution of the keto acids **4(a-c)** (2.07 mmol) dissolved in 1% potassium hydroxide solution (25 mL), and sodium borohydride (0.25 g) was added. The reaction mixture was then stirred for 1 h at room temperature. After acidification with hydrochloric acid, the reaction

mixture was extracted with ethyl acetate (2 × 50 mL). The usual workup gave the hydroxy acids **5(a-c)**, which were purified by recrystallization by ethyl acetate. The hydroxyl acids **5(a-c)** were dissolved in acetic anhydride (1 mL) and heated under reflux for 2 h. The reaction mixture was cooled, water (25 mL) was added and the reaction mixture was stirred overnight. The crystals that deposited were collected by filtration and the filtrates were extracted with dichloromethane (2 × 20 mL). The solvent was removed under reduced pressure and the crude dihydroisocoumarins **6(a-c)** were purified by column chromatography on silica gel using petroleum ether (40-80 °C) as an eluent.

2-[2'-Hydroxy-2'-(3'', 5''-dichlorophenyl)ethyl] benzoic acid (5a)

Yield: 77%, mp 138-140 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3157 (C-H), 2933 (Ar-H), 1711 (C=O), 1590 (C=C arom), 1094 (C-Cl): EI-MS m/z (rel. int.): 310 (M^+ 100), 312 (M^+ +2, 79), 314 (M^+ +4, 19); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 2.25 (1H, dd, J 8.22, 15.6 Hz, H-1'b), 2.60 (1H, dd, J 6.24, 15.6 Hz, H-1'a), 4.10 (1H, dd, J 7.12, 14.20 Hz, H-2'), 7.05 (3H, t, J 1.76 Hz, H-2'', 4'' and 6''), 7.57 (1H, m, H-5), 7.73 (1H, dd, J 1.78, 8.9 Hz, H-3), 7.78 (1H, m, H-4), 8.16 (1H, dd, J 1.77, 7.9 Hz, H-6); Elemental Analysis Found: C 57.91, H 3.88, Cl 15.41; Calculated: C 58.06, H 3.87, Cl 15.17.

2-[2'-Hydroxy-2'-(2'', 3''-dichlorophenyl)ethyl] benzoic acid (5b)

Yield: 70 %; mp 150-152 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3159 (C-H), 2936 (Ar-H), 1713 (C=O), 1593 (C=C arom), 1094 (C-Cl): EI-MS m/z (rel. int.): 310 (M^+ 86), 312 (M^+ +2, 65), 314 (M^+ +4, 11); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 2.22 (1H, dd, J 8.22, 15.6 Hz, H-1'b), 2.63 (1H, dd, J 6.24, 15.6 Hz, H-1'a), 4.17 (1H, dd, J 7.12, 14.20 Hz, H-2'), 7.55 (1H, dd, J 1.76, 8.1 Hz, H-4''), 7.59 (1H, t, J 7.72 Hz, H-5''), 7.63 (1H, dd, J 1.76, 7.78 Hz, H-6''), 7.57 (1H, m, H-5), 7.73 (1H, dd, J 1.87, 8.9 Hz, H-3), 7.78 (1H, m, H-4), 8.13 (1H, dd, J 2.1, 7.9 Hz, H-6); Elemental Analysis Found: C 58.12, H 3.77, Cl 15.11; Calculated: C 58.06, H 3.87, Cl 15.17).

2-[2'-Hydroxy-2'-(2'', 5''-dichlorophenyl)ethyl] benzoic acid (5c)

Yield: 72 %; mp 172-174 °C; IR ν_{\max} / cm^{-1} : 3300-3250 (-OH), 3153 (C-H), 2936 (Ar-H), 1716 (C=O), 1593 (C=C arom), 1099 (C-Cl): EI-MS m/z (rel. int.): 310 (M^+ 100), 312 (M^+ +2, 75), 314 (M^+ +4, 18); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 2.21 (1H, dd, J 8.22, 15.6 Hz, H-1'b), 2.61

(1H, dd, J 6.24, 15.6 Hz, H-1'a), 4.14 (1H, dd, J 7.12, 14.20 Hz, H-2'), 7.23 (1H, d, J 8.9 Hz, H-3''), 7.24 (1H, dd, J 1.56, 7.8 Hz, H-4''), 7.27 (1H, d, J 1.0 Hz, H-6''), 7.42 (1H, m, H-5), 7.73 (1H, dd, J 2.3, 8.9 Hz, H-3), 7.78 (1H, m, H-4), 8.15 (1H, dd, J 1.87, 7.79 Hz, H-6); Elemental Analysis Found: C 57.99, H 3.58, Cl 15.21; Calculated: C 58.06, H 3.87, Cl 15.17.

(±)-3-(3',5'-Dichlorophenyl)-3,4-dihydroisocoumarin (6a)

Yield: 73 %; mp 184-186 °C; IR ν_{\max} / cm^{-1} : 3153 (C-H), 2935 (Ar-H), 1708 (C=O), 1593 (C=C arom), 1099 (C-Cl): EI-MS m/z (rel. int.): 292 (M^+ , 11.3), 294 (M^+ +2, 9.2), 296 (M^+ +4, 3); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 3.09 (1H, dd, J 12.10, 16.25 Hz, H-4b), 3.29 (1H, dd, J 2.90, 16.36 Hz, H-4a), 5.93 (1H, dd, J 2.90, 12.03 Hz, H-3), 7.30 (2H, d, J 2.6 Hz, H-2' and 6'), 7.44 (1H, d, J 2.3 Hz, H-4'), 7.48 (1H, m, H-7), 7.58 (1H, dd, J 2.24, 7.60 Hz, H-5), 7.66 (1H, m, H-6), 8.15 (1H, dd, J 1.34, 7.6 Hz, H-8); Elemental Analysis Found: C 61.44, H 3.44, Cl 10.95; Calculated: C 61.64, H 3.08, Cl 10.96.

(±)-3-(2',3'-Dichlorophenyl)-3,4-dihydroisocoumarin (6b)

Yield: 77 %; mp 136-138 °C; IR ν_{\max} / cm^{-1} : 3151 (C-H), 2939 (Ar-H), 1710 (C=O), 1597 (C=C arom), 1097 (C-Cl): EI-MS m/z (rel. int.): 292 (M^+ 57.5), 294 (M^+ +2, 18), 296 (M^+ +4, 13); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 3.10 (1H, dd, J 12.11, 16.20 Hz, H-4b), 3.27 (1H, dd, J 2.90, 16.38 Hz, H-4a), 5.93 (1H, dd, J 2.86, 12.02 Hz, H-3), 7.29 (1H, dd, J 2.6, 7.4 Hz, H-6'), 7.30 (1H, t, J 5.91 Hz, H-5'), 7.32 (1H, dd, J 1.76, 7.9 Hz, H-4'), 7.45 (1H, m, H-7), 7.58 (1H, m, H-6), 7.66 (1H, dd, J 2.6, 7.73 Hz, H-5), 8.15 (1H, dd, J 2.6, 7.61 Hz, H-8); Elemental Analysis Found: C 61.54, H 3.04, Cl 10.99; Calculated: C 61.64, H 3.08, Cl 10.96.

(±)-3-(2',5'-Dichlorophenyl)-3,4-dihydroisocoumarin (6c)

Yield: 77 %; mp 121-124 °C; IR ν_{\max} / cm^{-1} : 3154 (C-H), 2933 (Ar-H), 1712 (C=O), 1598 (C=C arom), 1097 (C-Cl): EI-MS m/z (rel. int.): 292 (M^+ 35), 294 (M^+ +2, 21), 296 (M^+ +4, 11); $^1\text{H NMR}$ (300 MHz, CDCl_3): δ 3.09 (1H, dd, J 12.10, 16.25 Hz, H-4b), 3.29 (1H, dd, J 2.90, 16.36 Hz, H-4a), 5.93 (1H, dd, J 2.90, 12.03 Hz, H-3), 7.24 (1H, dd, J 7.6 Hz, H-3'), 7.30 (1H, dd, J 2.6 Hz, H-6'), 7.44 (1H, dd, J 2.3, 8.1 Hz, H-4'), 7.48 (1H, m, H-7), 7.58 (1H, m, H-6), 7.66 (1H, dd, J 1.24, 7.78 Hz, H-5), 8.15 (1H, J 1.23, 7.6 Hz, H-8). Elemental Analysis Found: C 61.44, H 3.44, Cl 10.95; Calculated: C 61.64, H 3.08, Cl 10.96.

Herbicidal assays²⁰

Compound with certain concentration was dissolved in acetone or DMF and diluted with water, and the solutions

was added to certain amount of soil as soil treatment in order that the concentration of test compound was 750 kg ha⁻¹, after this, the weed seeds were germinated, each pot contains 20-30 seeds and each experiment triplicated. The plant was cultured in the green house at 25-27 °C. After 4 weeks of growth, the fresh weight was measured and the inhibition percentage was calculated according to the corresponding control. For foliate spray, after the growth of weed for 2 weeks, 750 kg ha⁻¹ of solution was sprayed, and the weeds were cultured for another 2 weeks, again fresh weight was measured and the inhibition percentage was calculated according to the corresponding control. The weeds used for this experiment were *Brassica campestris*, *Echinochloa crusgalli*, *Amaranthus retroflexus* L and *Digitaria sanguinalis*(L.)Scop.

Fungicidal assays

Using fungi growth inhibition method for fungicide activity determination as described by Fan.²¹ Compound with 500 µg mL⁻¹ of concentration was dissolved in water by 0.1mL of assistance of DMF and then 500 µg mL⁻¹ of compound in agar plate was prepared, the fungi was inoculated and cultured in the culture tank at 24-26 °C, the diameter of fungi spread was measured two days later, growth inhibition was calculated by corresponding control.

Insecticidal assays

Weighing 10 mg sample into a 50 mL of glass beaker, then 20 mL of acetone was added, the maize leaf was dipped in the sample solution for 5 seconds, and the leaf was put in a petri dish with 10 cm of diameter to evaporate all solvents. Nine pieces of maize leaves were cut short and put into the petri dish of 10 cm diameter containing 10 *Mythimna separata* with 4 instars, the death rate of insect was detected 24 h and 96 h later experiment. For *Culex pipiens pallens* larva experiment, the 4 instars of insect was dipped into the water solution for 24 h, the death rate of insect was detected. None of the compound shows insecticidal activity.

Acknowledgments

The authors gratefully acknowledge funds from the URF project at Quaid-i-Azam University, Islamabad, Pakistan and the National Natural Science Foundation of China (No. 20672062), the National Key Project for Basic Research (No. 2003CB114402) and the Tianjin Natural Science Foundation (No. 07JCYBJC01200).

References

- Böger, P.; Wakabayashi, K.; Hirai, K.; *Herbicide Classes in Development*, Springer-Verlag: Berlin, Germany, 2002, p. 364 (ISBN 0540431470).
- Heap, I.; <http://www.weedscience.org>, accessed Sept 21, 2005.
- Beck-Sague, C. M.; Jarvis, W. R.; *J. Infect. Dis.* **1993**, *167*, 1247.
- Diekema, D. J.; Messer, S. A.; Brueggemann, A. B.; Coffman, S. L.; Doern, G. V.; Herwaldt, L. A.; Pfaller, M. A.; *J. Clin. Microbiol.* **2002**, *40*, 1298.
- Fridkin, S. K.; Jarvis, W. R.; *Clin. Microbiol. Rev.* **1996**, *9*, 499.
- Wenzel, R. P.; *Clin. Infect. Dis.* **1995**, *20*, 1531.
- Wingard, J. R.; *Clin. Infect. Dis.* **1995**, *20*, 115.
- Irvine, N. M.; Ricks, M. J.; Ross, R.; Bryan, K.; Klittich, C. J. R.; *CAN 142: 392441*; *Dow Agro sciences LLC: PCT Int. Appl.*, *2005033095* **2005**.
- Yamaguchi, I.; Fujimura, M.; *J. Pestic. Sci.* **2005**, *30*, 67.
- Thomas, O. L.; Jens, B.; *J. Nat. Prod.* **1999**, *62*, 1182.
- Odake, C. M.; Kam, L.; Narasimhan, M.; Poe, J. T.; Blake, O.; Krahenbuhl, J.; Powers, J. C.; *Biochemistry* **1991**, *30*, 2217.
- Hudig, D.; Allison, N. J.; Kam C. M. and Powers J. C.; *Mol. Immunol.* **1989**, *26*, 793. (CA 111: 192909h)
- Clark, M. T.; Gilmore, I. J.; *U. K. Pat. Appl.*, GB 2207425. (CA, 111, 19479v)
- Kerrigan, J. E.; Oleksyszyn, J.; Kam, C. M.; Selzler J. and Powers, J. C.; *J. Med. Chem.* **1995**, *38*, 544.
- Kam, C. M.; Kerrigan, J. E.; Plaskon, R. R.; Duffy, E. J.; Lollar, P.; Suddath, F. L.; Powers, J. C.; *J. Med. Chem.* **1994**, *37*, 1298.
- Bartus, R. T.; Eveleth, D. D. J.; Lynch G. S.; Powers, J. C.; *PCT Int. Appl.*, WO 99, 11,850. (CA 118, 822z)
- Ahmad, H.B.; Rama, N.H.; M. Hussain, M.; Hussain, M.T.; Qasim, M. M.; Hameed, S.; Malana. M.A.; Malik, A.; *Indian J. Chem.* **2003**, *42B*, 611.
- Shafiq, Z.; Arfan, M.; Rama, N. H.; Hameed, S.; Abbas,G.; Hussain, M.T.; *Turk. J. Chem.* **2005**, *29*, 321.
- Zhijin, F.; Qadeer, G.; Rama, N.H.; *Faming Zhuanli Shenqing Gongkai Shuomingshu*, Patent, Application No. CN 1012-2353 200,510,122,353. Priority: CAN 145:188619 AN 2006:593756 CAPLUS, p.24.
- Chen Nian-chun; *Beijing Agricultural University Press*, 1991.
- Zhi-Jin, F.; Bin, L.; Xiu-Feng, L.; Bin, Z.; Chang-Ling, L.; Zheng-Ming, L.; *Chemical Journal of Chinese Universities* **2004**, *25*, 663.

Received: December 8, 2006

Web Release Date: September 12, 2007