1	Synthesis of phenylalalinol-derived oxazolopyrrolidone
2	lactams and evaluation as NMDA receptor antagonists
3	Nuno A. L. Pereira • Francesc X. Sureda • Mireia Turch • Mercedes
4	Amat • Joan Bosch • Maria M. M. Santos
5	
6	Dedication – Dedicated to Professor Sundaresan Prabhakar on occasion of
7	his 75th anniversary.
8	
9	Abstract N-methyl-D-aspartate (NMDA) receptor antagonists are known
10	to rescue neuronal cell death caused by excessive activation of glutamate
11	receptors. This phenomenon, known as excitotoxicity, is implicated in the
12	pathogenesis of several neurodegenerative disorders including ischemia,
13	Alzheimer's disease, Parkinson's disease, and Huntington's disease.
14	Unfortunately, some antagonists of NMDA receptor have been tested in
15	clinical trials with discouraging results. However, recent advances in the
16	physiology and pharmacology of the NMDA receptor have kept the interest
17	alive to modulate NMDA receptors for therapeutic intervention.
18	We present here the synthesis of a small library of phenylalalinol-derived
19	oxazolopyrrolidone lactams and their evaluation as NMDA receptor
20	antagonists. The compounds were easily synthesized in yields up to 92%.
21	In addition, one of the compounds has an IC_{50} of 62 μM and offers
22	potential to develop more potent NMDA receptor antagonists.

```
1
 2
     Keywords Amino alcohols • Chiral auxiliaries • NMDA receptor
 3
 4
     antagonists • pyrrolidones • lactams
 5
 6
     M. M. M. Santos (\boxtimes)
 7
 8
     Research Institute for Medicines and Pharmaceutical Sciences (iMed.UL),
 9
     Faculty of Pharmacy, University of Lisbon. Av. Prof. Gama Pinto, 1649-
10
     003 Lisbon, Portugal
11
     e-mail: mariasantos@ff.ul.pt
12
     Telephone: +351 217946451
13
     Fax: +351 217946470
14
15
     N. A. L. Pereira ● M. M. M. Santos (⊠)
16
     Research Institute for Medicines and Pharmaceutical Sciences (iMed.UL),
17
     Faculty of Pharmacy, University of Lisbon. Av. Prof. Gama Pinto, 1649-
18
     003 Lisbon, Portugal
19
20
     F. X. Sureda • M. Turch
21
     Pharmacology Unit, Faculty of Medecine and Health Sciences, Universitat
22
     Rovira i Virgili, c./St. Llorenç 21, 43201 Reus (Tarragona), Spain
23
24
     M. Amat • J. Bosch
25
     Laboratory of Organic Chemistry, Faculty of Pharmacy and Institute of
26
     Biomedicine (IBUB), University of Barcelona, Av. Joan XXIII, s/n, 08028
27
     Barcelona, Spain
28
29
```

1 Introduction

N-Methyl-*D*-aspartate receptors (NMDAR) are part of the ionotropic
glutamate receptors family. After activation by their co-agonists, glycine
and glutamate, they allow the neural influx of Ca²⁺ through a membrane
ion pore thus playing an important role in the postsynaptic depolarization
[1-4].

7 Apart of their involvement on synaptic plasticity, which has been 8 postulated as the neurochemical basis of learning and memory, NMDA 9 receptors have been implicated in neuronal death [5]. High levels of 10 glutamate have been found in brain trauma and other neurodegenerative 11 diseases, so it is thought that NMDA receptors are potential targets for 12 neuroprotective compounds [6]. In fact, the adamantanes amantadine and 13 memantine develop their neuroprotective effects through blockade at the 14 NMDA receptor. Specifically, memantine is authorized in Western 15 countries and used therapeutically to slow-down the progression of 16 Alzheimer's disease [7].

In 2009, some oral active oxazolidine derivatives were described to act as NMDA antagonists by preventing the binding of the NMDAR ligands [8]. Based on this information and due to our interest in the synthesis of oxazolo lactams [9-10] we decided to extend our research to the synthesis of enantiopure oxazolopyrrolidone lactams using (*S*)-

1 phenylalaninol as a chiral inductor. Since the biological activity is greatly 2 affected by the absolute stereo-outcome of the compounds, a series of (R)-3 phenylalaninol derivatives was also prepared. 4 **Results and Discussion** 5 6 7 **Synthesis** 8 Recently, our research group has been interested in the synthesis of 9 phenylalalinol-derived oxazolopyrrolidone lactams to be evaluated as 10 NMDA receptor antagonists. The first series of compounds was 11 synthesized by cyclocondensation of (S)-phenylalaninol 1 with oxoacids 12 2a-e (Scheme 1). In turn, tricyclic lactams 3a-c were prepared from 2-13 acylbenzoic acid derivatives **2a-c** via reflux in toluene under Dean-Stark 14 conditions (Table 1). Starting from oxoacids 2d-e and using the same 15 reaction conditions we obtained the bicyclic lactams **3d-e** in 72-73% yields 16 (Table 1). In all cases, only one diastereoisomer product was observed.



4 To study the effect of the corresponding enantiomers as antagonists 5 at the NMDA receptor, lactams **3a'-c'** were also synthesized starting from (*R*)-phenylalaninol with 62-85% yields (Scheme 2, Table 1). 6



Scheme 2 8

Aminoalcohol	R	Reaction time/h	Product	Yield (%)
(S)-phenylalaninol	Н	16	3 a	70
(S)-phenylalaninol	Me	16	3b	92
(S)-phenylalaninol	Ph	16	3c	85
(S)-phenylalaninol	Me	48	3d	73
(S)-phenylalaninol	Ph	48	3e	72
(R)-phenylalaninol	Н	16	3 a'	71
(R)-phenylalaninol	Me	16	3 b'	85
(R)-phenylalaninol	Ph	16	3 c'	74

1 **Table 1** Reaction of phenylalaninol enantiomers **1** and **1**' with oxo acids **2**.

2

3 NMR spectroscopy

The most important features of the ¹H NMR spectra of these compounds
are the resonances of the H-3, H-2, and CH₂Ar protons. The H-3 signal
appears as a multiplet around 4.33–4.48 ppm. The diastereotopic H-2
protons appear as double of doublets around 4.08-4.33 ppm and 3.58-4.07
ppm. The methylene CH₂Ar protons appear as double of doublets around
2.94-3.19 ppm and 2.30-2.99 ppm.

Furthermore, in the ¹³C NMR spectra of compounds **3a-c** the newly formed
C-9b chiral center appears around 100ppm. This signal moves downfield as

the electronic demand of the substituent is increased: δ = 90.78 (H); 98.93
 (CH₃); 101.04 (Ph) ppm.

3 Compound 3b' underwent NOESY experiments and it was possible to 4 observe the correlations depicted in figure 1. As expected and accordingly 5 with published results with very similar compounds [9] synthesized via 6 ciclocondensation with an enantiopure aminoalcohols, the stereo-outcome 7 doesn't seem to be affected by the size of the keto-acid R substituent.

н

0=

'''/CH₃



- 9
- 10

11

12

13 Figure 1.- NOESY correlations observed for **3b**'.

14

15 NMDA receptor antagonist activity

16 The NMDA receptor activity of the compounds was evaluated by 17 measuring their ability to inhibit the intracellular calcium increase induced 18 by NMDA in cultured cerebellar granule neurons. Addition of glutamate or 19 NMDA (100 μ M) in the presence of glycine (10 μ M) produced a robust 20 and stable increase in intracellular calcium that was challenged with 21 cumulative additions of the compounds to be tested.

In our assays, memantine (used as a positive control) yielded an IC_{50} value 1 2 in the low micromolar range (1.48 μ M). As it is shown on figure 2, only 3 three out of the eight synthesized compounds showed an inhibitory activity higher than the 50% of the maximal effect. Specifically, 3c and 3d showed 4 an IC₅₀ in the high micromolar range (> 250 μ M), while 3e showed a 5 higher potency as a NMDA antagonist, giving an IC₅₀ of 62.0 µM. Related 6 7 to compound **3c**, which showed an IC₅₀ of 309.7 μ M, the enantiomer **3c'** 8 was inactive, so it seems that the stereochemistry at the 3 position is 9 important for activity.

10 More importantly, the phenyl derivative 3e is more potent as NMDA 11 receptor antagonism than amantadine (92.0 μ M).



13 Figure 2.- Inhibitory effect of the synthesized compounds and the14 adamantanes memantine and amantadine on NMDA-induced intracellular

calcium increase in cultured cerebellar granule neurons. The compounds
 were tested from 0.1 µM up to the highest possible concentration. Data are
 the mean of three different experiments, carried out on three different
 batches of cultured cells.

5

6 In summary, we have synthesized and fully characterized several 7 phenylalalinol-derived oxazolopyrrolidone lactams. In addition we describe 8 here the potential use of lactam **3e** as a hit compound to develop NMDA 9 receptor antagonists. The data now obtained provides a basis for exploring 10 if related derivatives have enhanced activity. The synthesis and biological 11 evaluation of more **3e** related compounds are in progress.

12

13 **Experimental**

14

15 Chemistry

All reagents and solvents were obtained from commercial suppliers and
were used without further purification. Melting points were determined
using a Kofler camera Bock monoscope M. The infrared spectra were
collected on a Shimadzu IRAffinity-1 FTIR infrared spectrophotometer.
Low resolution mass spectra (MS) were performed in LCLEM, Faculdade
de Farmácia, Universidade de Lisboa. Merck Silica Gel 60 F₂₅₄ plates were

1	used as analytical TLC; flash column chromatography was performed on
2	Merck Silica Gel (200-400 mesh). ¹ H and ¹³ C NMR spectra were recorded
3	on a Bruker 400MHz Ultra-Shield. Proton nuclear magnetic resonance
4	spectra were recorded at 400 MHz. Carbon nuclear magnetic resonance
5	spectra were recorded at 100 MHz. ¹ H and ¹³ C NMR chemical shifts are
6	expressed in δ (ppm) referenced to the solvent used and the proton coupling
7	constants J in hertz (Hz). Spectras were assigned using appropriate COSY,
8	DEPT and HMQC sequences.
9	
10	General procedure for the cyclocondensation reaction of (S) -2-
11	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e:
11 12	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling
11 12 13	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1
11 12 13 14	 amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (S)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total
 11 12 13 14 15 	 amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total consumption of the starting aminoalcohol. The solvent was evaporated and
 11 12 13 14 15 16 	 amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total consumption of the starting aminoalcohol. The solvent was evaporated and the crude residue was purified by column chromatography using ethyl
 11 12 13 14 15 16 17 	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total consumption of the starting aminoalcohol. The solvent was evaporated and the crude residue was purified by column chromatography using ethyl acetate/ <i>n</i> -hexane as eluent. The solid products were recrystallized in diethyl
 11 12 13 14 15 16 17 18 	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (<i>S</i>)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total consumption of the starting aminoalcohol. The solvent was evaporated and the crude residue was purified by column chromatography using ethyl acetate/ <i>n</i> -hexane as eluent. The solid products were recrystallized in diethyl ether/ <i>n</i> -hexane.
 11 12 13 14 15 16 17 18 19 	amino-3-phenylpropan-1-ol 1 with keto-acids 2a-e: To a stirred solution of (S)-2-amino-3-phenylpropan-1-ol in boiling toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1 eq. of the desired oxo-acid. The mixture was refluxed until total consumption of the starting aminoalcohol. The solvent was evaporated and the crude residue was purified by column chromatography using ethyl acetate/n-hexane as eluent. The solid products were recrystallized in diethyl ether/n-hexane. (3S,9bR)-3-benzyl-2,3-dihydrooxazolo[2,3-a]isoindol-5(9bH)-one

1	Starting from 90 mg of (S)-2-amino-3-phenylpropan-1-ol in 10 mL
2	of toluene. The obtained residue was purified by column chromatography
3	(AcOEt: <i>n</i> -hexane 3:7). Recrystallization from diethyl ether/ <i>n</i> -hexane
4	afforded 110 mg (70%) 3a . ¹ H NMR spectra was found to be identical with
5	the one described in Ref. [11].
6	(3S,9bR)-3-benzyl-9b-methyl-2,3-dihydrooxazolo[2,3-a]isoindol-
7	<i>5(9bH)-one</i> (3b , C ₁₈ H ₁₇ NO ₂)
8	Starting from 330 mg of (S)-2-amino-3-phenylpropan-1-ol in 30 mL
9	of toluene. The obtained residue was purified by column chromatography
10	(AcOEt: <i>n</i> -hexane 3:7) affording 560 mg (92%) of a colorless oil. 3b . 1 H
11	NMR (400 MHz, CDCl ₃): δ = 7.69 (d, J= 7.4 Hz, 1H, H-Ar), 7.54 (m, 1H,
12	H-Ar); 7.45 (m, 2H, H-Ar); 7.27 (m, 4H, H-Ar); 7.20 (m, 1H, H-Ar); 4.39
13	(m, 1H, H-3); 4.21 (dd, <i>J</i> = 8.9, 7.4 Hz, 1H, H-2), 4.07 (dd, <i>J</i> = 8.9, 6.5 Hz,
14	1H, H-2), 3.21 (dd, <i>J</i> = 13.8, 5.8 Hz, 1H, CH ₂ -Ph), 2.95 (dd, <i>J</i> = 13.8, 8.6
15	Hz, 1H, CH ₂ -Ph), 1.69 (s, 3H, CH ₃) ppm; ¹³ C NMR (100 MHz, CDCl ₃): δ
16	= 174.20 (C=O), 147.26 (Cq), 137.27 (Cq), 133.22 (CH-Ar), 131.60 (Cq),
17	130.16 (CH-Ar), 129.45 (2CH-Ar), 128.59 (2CH-Ar), 126.80 (CH-Ar),
18	124.33 (CH-Ar), 122.10 (CH-Ar), 98.93 (C-9b), 74.06 (CH ₂), 56.77 (CH),
19	40.89 (CH ₂ -Ph), 23.02 (CH ₃) ppm; IR (NaCl): $\overline{\nu}$ = 1715 (C=O) cm ⁻¹ ; MS
20	(ESI, CP 3.0 kV, SP 30V): m/z calc. = 279 [M] ⁺ , m/z found 280 [M+H] ⁺ ; R _f
21	(ethyl acetate: n -hexane 1:1) = 0.769.

1 (3S,9bR)-3-benzyl-9b-phenyl-2,3-dihydrooxazolo[2,3-a]isoindol-

2 *5(9bH)-one* (**3c**, C₂₃H₁₉NO₂)

3 Starting from 100 mg of (S)-2-amino-3-phenylpropan-1-ol in 7 mL of toluene. The obtained residue was purified by column chromatography 4 5 (AcOEt:n-hexane 1:9). Recrystallization from diethyl ether/n-hexane afforded 193 mg (86%) **3c**. M.p.: 92-94 °C; ¹H NMR (400 MHz, CDCl₃): δ 6 = 7.80 - 7.75 (m, 1H, H-Ar), 7.68 - 7.62 (m, 2H, H-Ar), 7.50 - 7.37 (m, 7 8 5H, H-Ar), 7.31 – 7.20 (m, 4H, H-Ar), 7.17 – 7.14 (m, 2H, H-Ar), 4.66 – 9 4.52 (m, 1H, H-3), 4.44 (dd, J = 8.6, 7.5 Hz, 1H, H-2), 3.96 (dd, J = 8.7, 10 6.6 Hz, 1H, H-2), 3.02 (dd, J = 13.8, 6.8 Hz, 1H, CH2-Ph), 2.51 (dd, J =13.8, 8.7 Hz, 1H, CH2-Ph). ppm; ¹³C NMR (101 MHz, CDCl₃): $\delta = 174.52$ 11 12 (C=O), 147.27 (Cq), 138.95 (Cq), 137.61 (Cq), 133.41 (CH-Ar), 131.10 13 (Cq), 130.23 (CH-Ar), 129.11 (2CH-Ar), 128.94 (2CH-Ar), 128.86 (CH-14 Ar), 128.64 (2CH-Ar), 126.77 (CH-Ar), 125.96 (2CH-Ar), 124.52 (CH-15 Ar), 123.56 (CH-Ar), 101.04 (C-9b), 75.91 (CH₂), 56.80 (CH), 40.54 (CH₂-Ph) ppm; IR (KBr): $\overline{\nu}$ = 1721 (C=O) cm⁻¹; MS (ESI, CP 3.0 kV, SP 16 30V): m/z calc. = 341 [M]⁺, m/z found = 342 [M+H]⁺; R_f (ethyl acetate: n-17 18 hexane 3:7) = 0.607.

19 (3S,7aR)-3-benzyl-7a-methyltetrahydropyrrolo[2,1-b]oxazol-5(6H)20 one (3d)

1	Starting from 100 mg of (S)-2-amino-3-phenylpropan-1-ol in 10 mL
2	of toluene. The obtained residue was purified by column chromatography
3	(AcOEt: <i>n</i> -hexane 1:1) affording 111 mg (73%) of a colorless oil. 3d . 1 H
4	NMR spectra was found to be identical with the one described in Ref. [12]
5	(3S,7aS)-3-benzyl-7a-phenyltetrahydropyrrolo[2,1-b]oxazol-5(6H)-
6	one (3e , C ₁₉ H ₁₉ NO ₂)
7	Starting from 100 mg of (S)-2-amino-3-phenylpropan-1-ol in 10 mL
8	of toluene. The obtained residue was purified by column chromatography
9	(AcOEt: <i>n</i> -hexane 3:7). Recrystallization from diethyl ether/ <i>n</i> -hexane
10	afforded 140 mg (72%) 3e . M.p.: 55-56 °C; ¹ H NMR (400 MHz, CDCl ₃): δ
11	7.51 (d, <i>J</i> = 7.4 Hz, 2H, H-Ar), 7.44 – 7.38 (m, 3H, H-Ar), 7.28 – 7.21 (m,
12	3H, H-Ar), 7.08 (d, J = 7.3 Hz, 2H, H-Ar), 4.50 – 4.35 (m, 1H, H-3), 4.13
13	(t, $J = 8.1$ Hz, 1H, H-2), $3.65 - 3.49$ (m, 1H, H-2), 2.94 (dd, $J = 13.7$, 6.2
14	Hz, 1H, CH ₂ -Ph), 2.89 – 2.77 (m, 1H, H-6), 2.63 – 2.45 (m, 2H, H-6 & H-
15	7), $2.35 - 2.18$ (m, 2H, CH ₂ -Ph & H-7) ppm; ¹³ C NMR (100 MHz, CDCl ₃):
16	δ 179.87 (C=O), 142.55 (Cq), 137.26 (Cq), 128.90 (2CH-Ar), 128.70
17	(2CH-Ar), 128.49 (2CH-Ar), 128.31 (CH-Ar), 126.60 (CH-Ar), 125.07
18	(2CH-Ar), 102.27 (C-7a), 72.26 (CH ₂), 56.44 (CH), 39.92 (CH ₂ -Ph), 35.05
19	(C-7), 32.57 (C-6) ppm; IR (KBr): $\overline{\nu} = 1721$ (C=O) cm ⁻¹ ; MS (ESI, CP 3.0
20	kV, SP 30V): m/z calc. = 293 [M] ⁺ , m/z found = 294 [M+H] ⁺ ; R _f (ethyl
21	acetate: <i>n</i> -hexane $3:7$) = 0.313.

4

2	General procedure for the cyclocondensation reaction of (R) -2-
3	amino-3-phenylpropan-1-ol 1' with keto-acids 2a-c:
4	To a stirred solution of (R) -2-amino-3-phenylpropan-1-ol in boiling
5	toluene under inert atmosphere and a Dean-Stark apparatus, was added 1,1
6	eq. of the desired oxo-acid. The mixture was refluxed until total
7	consumption of the starting aminoalcohol. The solvent was evaporated and
8	the crude residue was purified by column chromatography using ethyl
9	acetate/ <i>n</i> -hexane as eluent.
10	(3R,9bS)-3-benzyl-2,3-dihydrooxazolo[2,3-a]isoindol-5(9bH)-one
11	(3a')
12	Starting from 100 mg of (R)-2-amino-3-phenylpropan-1-ol in 15 mL
13	of toluene. The obtained residue was purified by column chromatography
14	(AcOEt: <i>n</i> -hexane 2:8). Recrystallization from diethyl ether/ <i>n</i> -hexane
15	afforded 125 mg (71%) 3a'. ¹ H NMR spectra was found to be identical
16	with the one described in Ref. [11].
17	(3R,9bS)-3-benzyl-9b-methyl-2,3-dihydrooxazolo[2,3-a]isoindol-
18	<i>5(9bH)-one</i> (3b' , C ₁₈ H ₁₇ NO ₂)
19	Starting from 100 mg of (R)-2-amino-3-phenylpropan-1-ol in 15 mL
20	of toluene. The obtained residue was purified by column chromatography
21	(AcOEt: <i>n</i> -hexane 2:8) affording 157 mg (85%) of a colorless oil. ¹ H NMR,

¹³C NMR and IR spectra were found to be identical with the ones described
 for compound **3b**.

3 (3R,9bS)-3-benzyl-9b-phenyl-2,3-dihydrooxazolo[2,3-a]isoindol5 (9bH)-one (3c', C₂₃H₁₉NO₂)
5 Starting from 100 mg of (*R*)-2-amino-3-phenylpropan-1-ol in 15 mL
6 of toluene. The obtained residue was purified by column chromatography
7 (AcOEt:*n*-hexane 2:8). Recrystallization from diethyl ether/*n*-hexane
8 afforded 167 mg (74%) of 3c'. ¹H NMR, ¹³C NMR and IR spectra were
9 found to be identical with the ones described for compound 3c.

10

11 NMDA receptor antagonist activity

12 The activity of the synthesized compounds as NMDA receptor antagonists 13 was evaluated using primary cultures of rat cerebellar neurons, as described 14 previously [13]. Briefly, cultures were prepared from 7-8 day-old Wistar rats (Charles River, France). Cerebella were dissected, minced and 15 16 trypsinized, and after several sedimentations, cells were plated on poly-17 lysinized coverslips placed in 24-well plates at a density of 1•106 cells/mL. 18 Plates were kept at 37°C in a cell incubator (Heraeus, Germany). After 16-19 18h, 10 µM cytosine arabinoside (Sigma-Aldrich, USA) was added to 20 avoid excessive proliferation of astrocytes. Cultures prepared in this manner are ready to be used in the NMDA receptor activity assays from the
 7th to the 11th day in vitro.

3 Activity at the NMDA receptor was assessed using the calcium-4 sensitive probe Fura-2 (Invitrogen, USA). After incubation with 6 µM 5 Fura-2 acetoxymethyl ester (Fura-2 AM) for 30-45 min at 37°C, a coverslip 6 was transferred to a plastic holder that was inserted in a quartz cuvette for 7 fluorescence measurements. Recordings of Fura-2 fluorescence were 8 performed using a PerkinElmer LS50B luminiscence spectrometer, both at 9 340 and 380 nm excitation wavelengths, and at 510 nm of emission. The 10 ratio of F340/F380 (R) is proportional to intracellular calcium. All the 11 measurements were made at 37°C and under mild stirring. Once the 12 recording was started, glycine (10 µM) and NMDA (100 µM) were added 13 to the cuvette, at 50 and 100 s respectively. This produced a sustained 14 increase in F_{340}/F_{380} , indicating that the NMDA receptors were activated 15 and that the intracellular calcium concentration was high. This intracellular 16 calcium increase was challenged with cumulative concentrations of the compounds under investigation, (from $1 \cdot 10^{-7}$ M up to up to $3 \cdot 10^{-4}$ M). If 17 18 the compounds would act as antagonists at the NMDA receptor this would be detected as a decrease in the value F_{340}/F_{380} . Experiments were 19 20 performed in triplicate. Memantine was used as a positive control.

1	When a minimum of 50% of inhibition was reached, the IC_{50} value
2	was calculated using non-linear regression with GraphPad Prism 5.0.
3	
4	Acknowledgements
5	Fundação para a Ciência e Tecnologia (Portugal) is acknowledged for
6	financial support (PTDC/QUI-QUI/111664/2009; PEst-
7	OE/SAU/UI4013/2011; REDE/1518/REM/2005). We also thank the
8	Portuguese-Spanish- Integrated Action (E-07/11 and AIB2010PT-00324)
9	and the Spanish and Ministerio de Ciencia e Innovación (MICINN)
10	(CTQ2009-07021/BQU) for financial support.
11	
12	References
12 13	References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or
12 13 14	References Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen
12 13 14 15	References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194.
12 13 14 15 16	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440:
12 13 14 15 16 17	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440: 456–462.
12 13 14 15 16 17 18	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440: 456–462. 3. Kaczor AA, Matosiuk D (2010) Molecular structure of ionotropic
12 13 14 15 16 17 18 19	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440: 456–462. 3. Kaczor AA, Matosiuk D (2010) Molecular structure of ionotropic glutamate receptors. Curr Med Chem 17(24): 2608-2635.
 12 13 14 15 16 17 18 19 20 	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440: 456–462. 3. Kaczor AA, Matosiuk D (2010) Molecular structure of ionotropic glutamate receptors. Curr Med Chem 17(24): 2608-2635. 4. Paoletti P (2011) Molecular basis of NMDA receptor functional
 12 13 14 15 16 17 18 19 20 21 	 References 1. Wollmuth LP, Talukdu I (2011) Local constraints in either the GluN1 or GluN2 subunit equally impair NMDA receptor pore opening. J Gen Physiol 138(2): 179-194. 2. Mayer ML (2006) Glutamate recetors at atomic resolution. Nature 440: 456–462. 3. Kaczor AA, Matosiuk D (2010) Molecular structure of ionotropic glutamate receptors. Curr Med Chem 17(24): 2608-2635. 4. Paoletti P (2011) Molecular basis of NMDA receptor functional diversity. Eur J Neurosci 33(8): 1351-1365.

1	5. Danysz W, Parsons CG (2012) Alzheimer's disease, beta-amyloid,
2	glutamate, NMDA receptors and memantine - searching for the
3	connections. Br J Pharmacol 167(2): 324-352.
4	6. Lau A, Tymianski M (2010) Glutamate receptors, neurotoxicity and
5	neurodegeneration. Pflugers Arch 460:525-542.
6	7. Parsons CG, Stoffler A, Danysz W (2007) Memantine: a NMDA
7	receptor antagonist that improves memory by restoration of
8	homeostasis in the glutarnatergic system - too little activation is bad,
9	too much is even worse. Neuropharmacology 53: 699-723.
10	8. Ip NY, Zhu H-J, Ip FC-F (2009) Oxazolidine as NMDA receptor
11	antagonists. WO2009092324, July 30, 2009.
12	9. Santos MMM (2011) Tryptophanol-Derived Oxazolopiperidone
13	Lactams: Valuable Building Blocks for the Enantioselective Synthesis
14	of Piperidine-Containing Alkaloids. In: Heterocyclic Targets in
15	Advanced Organic Synthesis, Research Signpost, India, 69-82.
16	10. Amat M, Pérez M, Bosch J (2011) Enantioselective Synthesis of Indole
17	Alkaloids from Chiral Lactams. Synlett 2: 143-160.
18	11. Sikoraiová J, Chihab-Eddine A, Marchalín S, Daïch A (2002)
19	Diastereoselective Access to Chiral Non-Racemic [1,3]Oxazolo-[2,3-
20	a]isoindol-5-one Ring Systems via O-Cationic Cyclization. J
21	Heterocyclic Chem 39: 383-390.

1	12. Allin SM, James SL, Martin WP, Smith TAD, Elsegood, MRJ (2001)
2	Stereoselective synthesis of the pyrroloisoquinoline ring system. J
3	Chem Soc, Perkin Trans 1: 3029-3036.
4	13. Torres E, Duque MD, López-Querol M, Taylor MC, Naesens L, Ma C,
5	Pinto LH, Sureda FX, Kelly JM, Vázquez S. (2012) Synthesis of
6	benzopolycyclic cage amines: NMDA receptor antagonist, trypanocidal
7	and antiviral activities. Bioorg Med Chem 20:942-948.
8	

2 Graphics for use in the Table of Contents

