

Synthesis of silver nanoparticles by sophorolipids: Effect of temperature and sophorolipid structure on the size of particles[†]

M B KASTURE, P PATEL, A A PRABHUNE, C V RAMANA, A A KULKARNI* and B L V PRASAD*

National Chemical Laboratory, Dr. Homi Bhabha Road, Pune 411 008

e-mail: aa.kulkarni@ncl.res.in; pl.bhagavatula@ncl.res.in

Abstract. We report *in situ* synthesis of silver nanoparticles using biosurfactants called sophorolipids as reducing and capping agents. We further study the effect of temperature and the structure of sophorolipid on the size of silver nanoparticles obtained. The silver nanoparticles were characterized by UV-visible, transmission electron microscope (TEM) and light scattering (DLS) analysis techniques.

Keywords. Sophorolipid; nucleation and growth; dynamic light scattering; silver nanoparticles.

1. Introduction

Physicochemical properties of metal nanoparticles have a strong dependence on their shape¹ and size,² which make them strong candidate for various applications such as catalysis,³ biosensing,^{4,5} recording media⁶ and optics.⁷ Silver nanoparticles have been widely studied due to their applications of optical properties⁸ and application such as antimicrobial agent,^{9,21} substrate for surface enhanced Raman spectroscopy (SERS)¹⁰ and biosensing.¹¹ Their aforementioned interesting properties and enormous application potential has led to numerous methods being developed for synthesis of nanoparticles of various shapes and sizes. Typically, the methods employed for the synthesis of these nanomaterials include solution based method,¹² micelle,^{13,14} sol gel,¹⁵ chemical precipitation,¹⁶ hydrothermal synthesis,¹⁷ pyrolysis¹⁸ and vapour deposition.¹⁹

Nowadays, molecules, which exhibit dual nature such as capping and reducing in one go,²⁰ are preferred since the reaction takes place in one step and thereby the need for external reducing agents is eliminated. This may enable us to exert greater control on the reaction parameters and condense the number of steps involved in synthesis so that the lab scale syntheses could be scaled up greatly enhancing their application potential. Another focused area of research is accomplishing the nanoparticle formation

in the aqueous media.²¹ This is also an important stipulation from the environmental point of view if the large scale nanoparticle synthesis needs to be accomplished. The great hurdle to overcome in the large scale synthesis of nanomaterials, though, is the concentrations of reagents that we can employ. Since, the higher concentrations may lead to irreversible aggregations; the synthesis is always performed in mM concentrations. A way forward from this stumbling block is the usage of continuous flow reactions²² where again molecules that can accomplish reduction and capping in one go are preferred. However, a thorough understanding of the various factors that lead to the formation of nanoparticles with desired parameters (size, shape, particle size distribution, etc.) is needed to convert a laboratory batch process into a continuous process.

As a first step towards the above aim, here we report the effect of different parameters that control the synthesis of the silver nanoparticles under consideration. Using sophorolipids we have recently started exploring the utility of bio-surfactant called sophorolipid as capping and reducing agent in nanoparticle synthesis.²³ These have been shown to result in the reduction of Ag⁺ ions to Ag nanoparticles and cap the ensuing nanoparticles. Sophorolipids are molecules with a sophorose – a dimeric glucose – attached to ω or $\omega - 1$ carbon of fatty acids (oleic acid, stearic acid, etc.). The final sophorolipid capped nanoparticles that we obtain fall in the category of glyconanoparticles. These glyconanoparticles are being investigated with great fervour these days for their widespread applications as cell mimicks, to

[†]Dedicated to Prof. C N R Rao on his 75th birthday

*For correspondence

understand protein–carbohydrate interactions and in various biomedical applications also.²⁴ Here in the current study, we present the results on particle size variation as a function of time and temperature when two different types of sophorolipids are used for the synthesis of Ag NPs. The objective is to identify the suitable conditions that yield specific particle sizes with other reaction parameters being the same. The understanding from these results will have great bearing on further work where these reactions will be carried out in continuous flow processes. The nanoparticles are characterized by UV-visible, transmission electron microscopy and dynamic light scattering (DLS) studies.

2. Experimental

Silver nitrate and potassium hydroxide were purchased from SRL and Merck respectively. Oleic acid and linoleic acid were purchased from Aldrich and the chemicals used as media in synthesis of sophorolipid like malt extract, glucose, yeast extract and peptone were purchased from Hi Media. All the chemicals were used as received without further purification.

2.1 Characterization techniques

2.1a *UV-visible*: UV-visible measurements were carried out on Jasco V-570 UV/VIS Spectrophotometer at the resolution of 1 nm.

2.1b *Transmission electron spectroscopy*: TEM measurements were performed on FEI model Technai G² F-30 operated at an accelerating voltage of 300 kV.

2.1c *Dynamic light scattering measurements*: DLS measurements were carried out on Brookhaven Instrument model 90 Plus Particle Size Analyzer.

2.2 Methodology

The preparation of sophorolipids from oleic acid and linoleic acid was carried out following reported procedures²⁵ and the spectroscopic characterization concurs with the reported data.²³

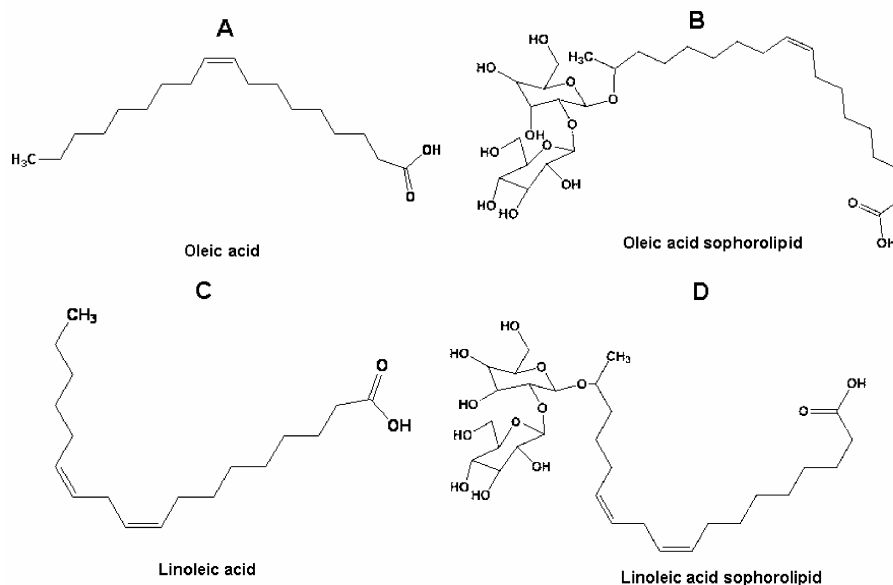
Synthesis of sophorolipid capped Ag NPs was carried out at different temperatures. In typical experiment when one or two drops of concentrated

KOH solution is added to 10 mL of solution containing sophorolipid (1×10^{-3} M) and AgNO₃ (1×10^{-3} M) colour of the solution changes to yellow indicating the formation of Ag nanoparticles. This experiment was repeated at five different temperatures viz. 30 (room temperature; RT), 40, 60, 80 and 90°C and with two different sophorolipids viz. oleic acid and linoleic acid derived sophorolipid for a period of 5 min. The temperature was maintained constant using a thermostat (Julabo, Germany). For the characterization using light scattering, the experiments were carried out at above-mentioned different temperatures while the solution was kept at that temperature inside the light scattering experiment also using a thermostat. Control experiments carried out in exactly the same manner described above but with pure oleic acid and linoleic acid molecules did not lead to the formation of AgNPs indicating that the sophorose moiety is necessary for the reductive synthesis of AgNPs.

3. Results and discussion

Scheme 1 shows the structures of oleic acid (A), oleic acid sophorolipid (B), linoleic acid (C), and linoleic acid sophorolipid (D). Oleic acid (OA) has one *cis* double bond while the linoleic acid (LOA) has two *cis* double bonds.

UV-Visible spectra were recorded for Ag NPs synthesized using OA and LOA sophorolipid as capping/reducing agent. Figure 1A shows the UV-visible spectra for OA-sophorolipid capped/reduced Ag NPs synthesized at different temperatures while figure 1B corresponds to LOA sophorolipid reduced/capped Ag NPs. Curves 1–5 corresponds to different temperatures employed i.e. 30°C, 40°C, 60°C, 80°C and 90°C respectively. In both the cases Ag NPs display a very distinct yellowish-brown colour and an absorbance in the region 400–420 nm accredited to the collective oscillations of conduction electrons and termed as surface plasmon resonance (SPR).²⁶ From the UV-visible curves we observe that at room temperature no distinct surface plasmon resonance peak is observed when both OA and LOA derived sophorolipids are used as reducing and capping agents. As the temperature increases surface plasmon peak begins to evolve and at 90°C a well defined and sharp peak is observed. This trend is same in both OA and LOA cases; however in case of LOA derived sophorolipid no clear SPR is observed up to 80°C while at 90°C it becomes very apparent. Figure



Scheme 1. Structure of oleic acid (**A**), oleic acid sophorolipid (**B**), linoleic acid (**C**) and linoleic acid sophorolipid (**D**).

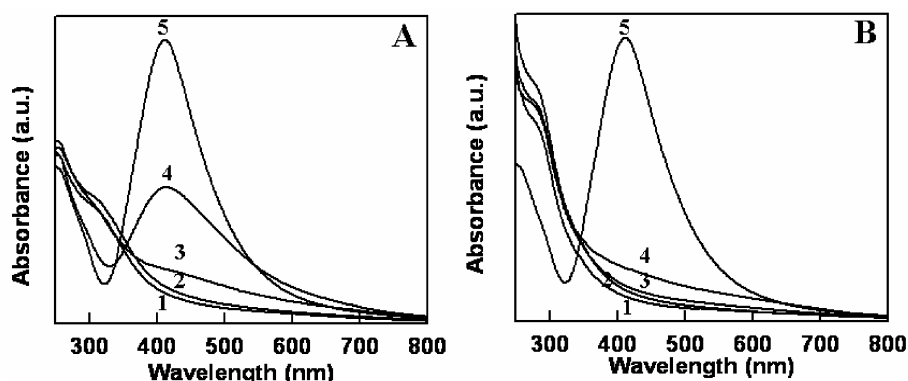


Figure 1. Shows UV-visible spectra for silver nanoparticles synthesized using OA sophorolipid (**A**) and LOA sophorolipid (**B**) as capping and reducing agent. Curves 1–5 were recorded from the samples at different temperatures at which the experiments were carried out in both cases.

2A represents the TEM image of OA sophorolipid capped/reduced Ag NPs synthesized at 40°C. It is observed from the TEM images that the particles are poly-dispersed. The particle size distribution is shown in figure 2B and the estimated mean particle size is close to 20 nm. Figure 2D shows the TEM image for OA sophorolipid capped/reduced Ag NPs synthesized at 90°C. In this case, the particles are spherical almost mono-dispersed and also smaller in size than the particles synthesized at 40°C. The average particle size in this case is 5.5 nm (figure 2E). For LOA sophorolipid similar trends are observed. Figure 3A shows TEM image of LOA capped/reduced Ag nano-

particles synthesized at 40°C. Particles are again polydispersed with average particle size as 22 nm (figure 3B). For reaction carried out at higher temperature i.e. 90°C particles are spherical and well defined with average particle size of 11 nm (figure 3E).

Particle size distribution obtained from dynamic light scattering (DLS) is shown in figure 2(C, F) and in figure 3 (C, F). Figures 2(C, F) represent particle size distribution for OA sophorolipid capped/reduced Ag nanoparticles at 40°C and 90°C respectively. The average particle size obtained from DLS data (35 nm and 3 nm for reactions done at 40°C and 90°C

respectively) were compared with the results obtained by TEM analysis (20 nm and 5.5 nm for reactions done at 40°C and 90°C respectively). Keeping in view that TEM and DLS analyse particle sizes differently (DLS analysis includes the ligand shell and determines the hydrodynamic size whereas in TEM we can look at only metallic core) the values described above from DLS and TEM at these two temperatures can be considered to be in good agreement with each other. Similarly figures 3 (C, F) represent particles size distribution obtained from DLS for LOA sophorolipid as capping/reducing agent at 40°C and 90°C. At these two temperatures (40°C and 90°C) the average particle sizes obtained are 28 nm and 14 nm, respectively. The values obtained in this case are also in accordance to the values obtained by TEM measurements. Figure 4 shows graph of average particle size versus the temperature. This information can be used further to carry out the reaction

in continuous flow experiments. For the range of temperatures under consideration, at lower temperatures the particles are smaller and with increasing temperatures, the average particle size goes through a maximum and becomes smaller again towards higher temperature. While one would expect a continuous decrease in particle size with increase in temperature and the present result is unusual. One of the reasons for this phenomenon could be the fact that the reaction time for each case was identical. Since the reaction is relatively slow at lower temperatures, in the time for which the reaction was observed and analysed, the particles must not have formed and grown completely. While, for higher temperatures, the reaction is relatively fast and particle formation and growth is complete. In order to confirm this speculation, we carried out the synthesis at room temperature and observed the UV spectrum at every hour for almost 21 h. It was seen that the, surface

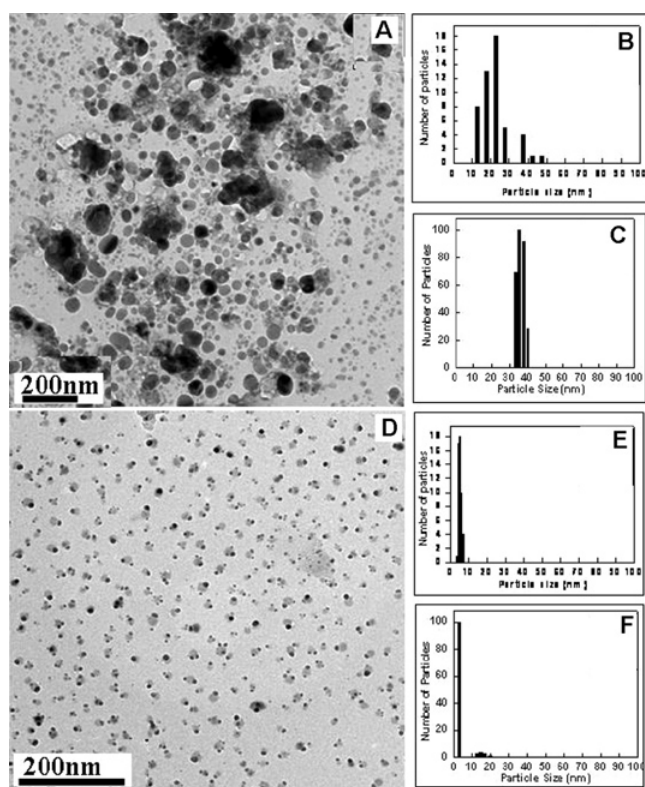


Figure 2. TEM images and particle size distribution for silver nanoparticles synthesized using OA sophorolipid as capping/reducing agent. (A) TEM image for the experiments carried out at 40°C and (D) TEM image for reaction carried out at 90°C. B and C are particle size distribution form TEM and DLS respectively for reaction temperature 40°C. E and F are particle size distribution plots obtained from TEM and DLS respectively for reaction temperature 90°C.

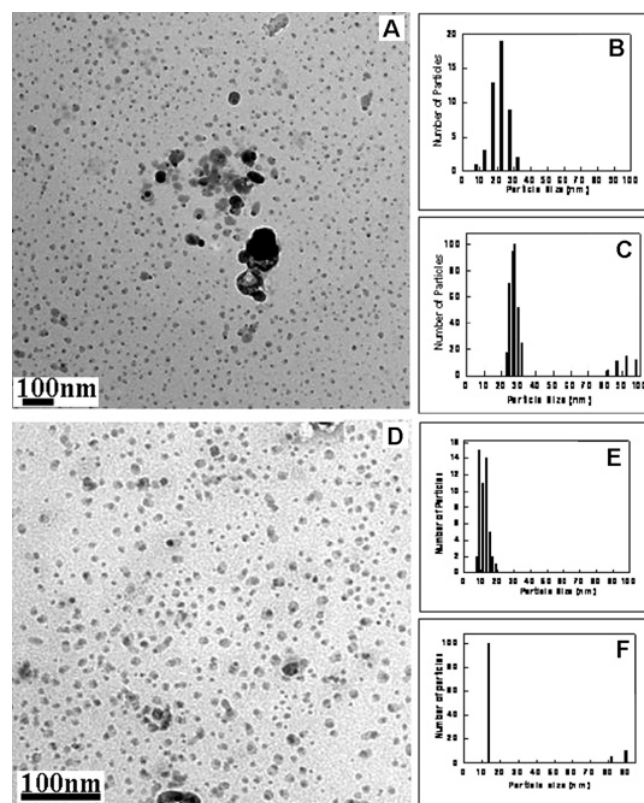


Figure 3. TEM images and particle size distribution for silver nanoparticles synthesized using LOA sophorolipid as capping/reducing agent. (A) TEM image for the experiments carried out at 40°C and (D) TEM image for reaction carried out at 90°C. B and C are particle size distribution plots form TEM and DLS respectively for reaction temperature 40°C. E and F are particle size distribution plots obtained from TEM and DLS respectively for reaction temperature 90°C.

plasmon peak was steadily increasing with time and at 21 h, it was the highest. Thus at lower temperatures particle formation process is slow and this would certainly lead to smaller particles in the initial period of observation thereby showing a trend that passes through a peak. The observations from longer time clearly suggest that we would observe a steady decrease in the particle size with increasing temperature. This observation was evident for both the sophorolipids.

The decreased particle size and narrower size distribution with increasing temperature is a well known phenomenon.²⁷ It is normally attributed to the increased reaction rates at higher temperatures. As the reaction rate increases the silver ions are consumed faster thus leaving less possibility for particle size growth and hence smaller particles and narrower size distributions at higher temperatures. The difference in particle sizes between OA derived sophorolipid and LOA derived sophorolipid is little intriguing. However, the roots for this apparently intriguing result may lie in the formation of olefin-silver ion complex formation. It is a well established fact that olefin silver complex depends on various factors such as chain length of olefins and on increasing number of substituents at the double bond. The stability of this complex decreases in the order $R-CH=CH_2 > R_2C=CH_2 > cis\ R.CH=CH.R > trans\ R.CH=CH.R > R_2C=CH.R > R_2C=CR_2$.²⁸ In our case we use OA and LOA to prepare the sophorolipid.

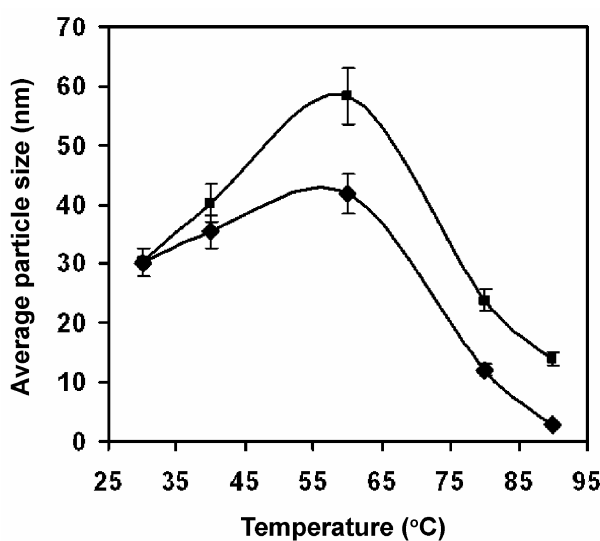


Figure 4. Average particle sizes obtained from DLS data for OA sophorolipid (1) and LOA sophorolipids reduced/capped (2) silver nanoparticles.

While OA has one *cis* double bond and LOA has two *cis* double bonds (see scheme 1). So in case of LOA sophorolipid there is a probability that silver ions bind to the two double bonds more strongly or more number of Ag^+ ions bind to the two double bonds thus slowing down the reduction rate. Therefore, the LOA sophorolipid reduces Ag^+ ions to Ag nanoparticles more slowly. As mentioned previously, a slower reduction rate leads to bigger particles and broader particle size distributions. Further work on quantifying these aspects in terms of different relative rates is in progress.

Thus the main features of our above results can be summarized as follows. Both OA derived sophorolipid and LOA derived sophorolipids are capable of reducing silver ions to Ag NPs and the ensuing Ag NPs are capped by the sophorolipid or the oxidized form of the sophorolipid. The temperature dependent reduction of Ag^+ ions by both these sophorolipids presents us the following scenarios: (i) at low temperatures both these result in bigger particles (TEM and DLS results) and broad particle size distributions. Again in both cases as the temperature is raised the particle size becomes smaller as well as the distribution gets narrower and (ii) a more interesting observation is that when compared at each temperature condition, OA derived sophorolipid always results in smaller particle sizes as compared to LOA derived sophorolipid.

4. Conclusions

The formation of silver nanoparticles is reported using the OA derived sophorolipid and LOA derived sophorolipid which have the capability to act as reducing and capping agents in one go. The reactions are carried out at various temperature conditions and at lower reaction temperatures bigger particles with broad size distributions are observed. With increasing temperature, the particles were seen to become smaller with a narrow size distribution. The apparently intriguing result of LOA derived sophorolipid always leading to bigger particles in comparison to OA derived sophorolipid is explained on the basis of stronger complexation of silver ions to the two double bonds present in the former. This reduces the reaction rate and hence to slower nucleus formation and greater particle size growth. Efforts are underway to perform these reactions in continuous flow mode so that synthesis of these nanoparticles at relatively larger scale can be accomplished.

Acknowledgements

M B K thanks Department of Science and Technology (DST) UNANST for financial support. This work was funded by DST-UNANST scheme and Department of Biotechnology (DBT) and is gratefully acknowledged. A A K thanks the Consortium on Microreaction Technology (C μ R) www.ncl-india.org/cm/r/ for financial support.

References

- Alivisatos A P 1996 *J. Phys. Chem.* **100** 13226
- (a) Jin R, Cao Y W, Mirkin C A, Kelly K L, Schatz G C and Zheng J G 2001 *Science* **294** 1901; (b) Aizpurua J, Hanarp P, Sutherland D S, Käll M, Bryant G W and García de Abajo F J 2003 *Phys. Rev. Lett.* **90** 057401
- Moreno-Manas M and Pleixats R 2003 *Acc. Chem. Res.* **36** 638
- Mirkin C A, Letsinger R L, Mucic R C and Storchhoff J J 1996 *Nature* **382** 607
- Han M, Gao X, Su J Z and Nie S 2001 *Nature Biotechnol.* **19** 631
- Sun S, Murray C B, Weller D, Folks L and Moser A 2000 *Science* **287** 1989
- Kamat P V 2002 *J. Phys. Chem.* **B106** 7729
- (a) Jin R C, Cao Y W, Mirkin C A, Kelly K L, Schatz G C and Zheng J G 2001 *Science* **294** 1901; (b) Silva T J, Schultz S and Weller D 1994 *Appl. Phys. Lett.* **65** 658; (c) Schultz S, Smith D R, Mock J J and Schultz D A 2000 *Proc. Natl. Acad. Sci. USA* **97** 996
- (a) Sanpui P, Murugadoss A, Durga Prasad P V, Ghosh S S and Chattopadhyay A 2008 *Int. J. Food Microbiol.* **124** 142; (b) Ruparelia J P, Chatterjee A K, Duttagupta S P and Mukherji S 2008 *Acta Biomaterialia* **4** 3 707; (c) Kumar A, Vemula P K, Ajayan P M and John G 2008 *Natl. Mater.* **3** 236
- (a) Nie S and Emory S R 1997 *Science* **275** 1102; (b) Dick L A, McFarland A D, Haynes C L and P van Duyne R 2002 *J. Phys. Chem.* **B106** 853
- (a) Link S and El-Sayed M A 1999 *J. Phys. Chem.* **B103** 8410; (b) Kottmann J P, Martin O J F, Smith D R and Schultz S 2001 *Chem. Phys. Lett.* **341** 1
- (a) Qu L, Peng Z A and Peng X 2001 *Nano Lett.* **1** 333; (b) Peng Z A and Peng X 2002 *J. Am. Chem. Soc.* **124** 3343
- (a) Lisiecki I, Billoudet F and Pileni M P 1996 *J. Phys. Chem.* **100** 4160; (b) Hao E, Bailey R C, Schatz G C, Hupp J T and Li S 2004 *Nano Lett.* **4** 327
- (a) Turkevich J, Stevenson P C and Hillier J 1951 *Discuss. Faraday Soc.* **11** 55; (b) Taleb A, Petit C and Pileni M P 1997 *Chem. Mater.* **9** 950; (c) Barnickel P and Wokaun A 1990 *Mol. Phys.* **69** 1; (d) Chen Z J, Qu X M, Fang F Q and Jiang L 1996 *Colloids Surf.* **B7** 173
- Mondelaers D, Vanhoyland G, Van den Rul H, D'Haen J, Van Bael M K, Mullens J and Van Poucke L C 2002 *Mater. Res. Bull.* **37** 901
- Spanhel L, Haase M, Weller H and Henglein A 1987 *J. Am. Chem. Soc.* **109** 5649
- Sun Y-P, Guduru R, Lin F and Whiteside T *Indian Eng. Chem. Res.* 2000 **39** 4663
- (a) Cai W and Zhang L 1997 *J. Phys. Condensed Matter* **9** 7257; (b) Maya L, Paranthaman M, Thundat T and Bauer M L 1996 *J. Vac. Sci. Technol.* **B14** 15
- Liu Y, Zhu W, Tse M S and Shen Y 1995 *J. Mater. Sci. Lett.* **14** 1185
- (a) Si S and Mandal T K 2007 *Chem. Eur. J.* **13** 3160; (b) Si S, Bhattacharjee R R, Banerjee A and Mandal T K 2006 *Chem. Eur. J.* **12** 1256; (c) Bhattacharjee R R, Das A K, Haldar D, Si S, Banerjee A and Mandal T K 2005 *J. Nanosci. Nanotechnol.* **5** 1141
- Dahl J A, Maddux B L S and Hutchison J E 2007 *Chem. Rev.* **107** 2228
- (a) Jahn A, Reiner J E, Vreeland W N, DeVoe D L, Locascio L E and Gaitan M 2008 *J. Nanopart. Res.* **10** 925; (b) Song Y, Hormes J and Kumar C S S R 2008 *Small* **4** 698
- (a) Kasture M, Singh S, Patel P, Joy P A, Prabhune A A, Ramana C V and Prasad B L V 2007 *Langmuir* **23** 1409; (b) Singh S, Jaisawal S, Patel P, Prabhune A A, Ramana C V and Prasad B L V *New J. Chem.* 2008 (accepted)
- de la Fuente J M and Penadés S 2006 *Biochim. Biophys. Acta* **1760** 636
- (a) Inge N A, Bogaert V, Saerens K, De Muynck C, Develter D, Soetaert W and Vandamme E J 2007 *Appl. Microbiol. Biotechnol.* **76** 23; (b) Shah S and Prabhune A A, 2007 *Biotechnol. Lett.* **29** 267
- (a) Henglein A 1993 *J. Phys. Chem.* **97** 547; (b) Mulvaney P I 1996 *Langmuir* **12** 788
- Park J, Joo J, Kwon G S, Jang Y and Hyeon T 2007 *Angew. Chem. Int. Ed.* **46** 4630
- de Ligny, C L 1976 in *Advances in chromatography* (eds) J C Giddings, E Grushka, J Cazes and P R Brown (New York: Marcel Dekker) vol. 14, pp 265–304