THE ACTIVITY OF PEPTIDES OF THE CALCITONIN FAMILY IN BONE

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Naot D, Musson DS, Cornish J. The Activity of Peptides of the Calcitonin Family in Bone. Physiol Rev 99: 781-805, 2019. Published December 12, 2018; doi: 10.1152/physrev.00066.2017.—Calcitonin was discovered over 50 yr ago as a new hormone that rapidly lowers circulating calcium levels. This effect is caused by the inhibition of calcium efflux from bone, as calcitonin is a potent inhibitor of bone resorption. Calcitonin has been in clinical use for conditions of accelerated bone turnover, including Paget's disease and osteoporosis; although in recent years, with the development of drugs that are more potent inhibitors of bone resorption, its use has declined. A number of peptides that are structurally similar to calcitonin form the calcitonin family, which currently includes calcitonin gene-related peptides (α CGRP and β CGRP), amylin, adrenomedullin, and intermedin. Apart from being structurally similar, the peptides signal through related receptors and have some overlapping biological activities, although other activities are peptide specific. In bone, in vitro studies and administration of the peptides to animals generally found inhibitory effects on osteoclasts and bone resorption and positive effects on osteoblasts and bone formation. Surprisingly, studies in genetically modified mice have demonstrated that the physiological role of calcitonin appears to be the inhibition of osteoblast activity and bone turnover, whereas amylin inhibits osteoclast activity. The review article focuses on the activities of peptides of the calcitonin family in bone and the challenges in understanding the relationship between the pharmacological effects and the physiological roles of these peptides.

Ι.	INTRODUCTION	781
П.	THE ACTIVITY OF THE	786
III.	THE PHYSIOLOGICAL ROLE	793
IV.	CLINICAL USE OF CALCITONIN	796
V.	SUMMARY	798

I. INTRODUCTION

A. The Calcitonin Family of Peptides

Calcitonin is a peptide hormone secreted from the parafollicular cells of the thyroid gland in response to an increase in serum calcium. The calcitonin family of peptides includes a number of peptide hormones that are structurally related to calcitonin and signal through common receptors, although they are expressed in different tissues and respond to different signals. Although some of the biological activities of the family members overlap, others are unique, and the main physiological functions of the peptide hormones are diverse.

1. Calcitonin

Calcium homeostasis is mainly controlled by the skeleton, the gut, and the kidney. The coordinated actions of these organs maintain the circulating concentrations of calcium within the narrow range of 8.5 and 10.5 mg/dL (2.12–2.62 mM). The tight control of circulating calcium levels is essential for major physiological processes, including muscle contraction, neuronal excitation, glycogen metabolism, and coagulation. At the cellular level, calcium ions play key roles in basic functions, such as cell adhesion and division (69). The importance of calcium homeostasis and the detrimental consequences of calcium imbalance have been recognized by physicians for hundreds of years.

The discovery of the parathyroid glands in the late 19th century was central to the understanding of the hormonal mechanisms that control calcium homeostasis (168). The physiological function of the gland was described later, in 1925, when Collip (39) showed that acid extracts of the parathyroid gland can reverse the tetany caused by parathyroidectomy. He concluded that the gland is an endocrine organ that secretes a parathyroid hormone (PTH), which responds to low calcium levels and acts to restore them back to the normal range. With the improvement of methods for PTH extraction, the mechanism of action of PTH could be further investigated, and it was found that the hormone mobilizes calcium from bone (13, 147).

Calcium homeostasis, and in particular the role of the thyroid–parathyroid in the regulation of calcium levels, was investigated by Copp and Cheney in a perfusion system in anesthetized dogs (42). Perfusion of the thyroid parathyroid glands with high levels of calcium caused a rapid fall (within 15 min) in the level of calcium in the blood. As PTH was the only hormone known to be secreted from the gland, the researchers hypothesized that the perfused high calcium concentrations suppressed PTH secretion, which, in turn, caused the observed fall in systemic calcium levels. To test this hypothesis, the investigators removed the thyroidparathyroid from the dogs, expecting the same effect on circulating calcium levels in the absence of PTH. However, to their surprise, systemic calcium levels remained high. They concluded that hypercalcemia does not merely suppress the production of PTH, but stimulates the production of a hormone that lowers calcium levels in the blood (41, 42). The name "calcitonin" was suggested for the putative agent that controls calcium tone. Subsequent studies established that calcitonin is produced by the thyroid gland, and a number of investigators suggested the alternative name "thyrocalcitonin" (95).

Biological assays that had been developed to investigate of the mechanisms of action of PTH in controlling circulating calcium levels were used to study calcitonin. Using one of these experimental systems, Friedman and Raisz (73) injected pregnant rats with ⁴⁵Ca and then placed embryonic bone in tissue culture in a bioassay used to determine the release of radioactively labeled calcium as a measure of bone resorption. The effects of partially purified calcitonin from extracts of rat thyroid gland were measured in baseline conditions and in combination with PTH, which induced bone resorption. The results of the study showed that calcitonin inhibits both basal and PTH-stimulated bone resorption, thus identifying the underlying mechanism of calcitonin's hypocalcemic effect (73). Further evidence for the direct inhibition of bone resorption by calcitonin came from an in vivo study that measured the collagen breakdown product hydroxyproline in urine of rats treated with highly purified porcine calcitonin (142). The study showed that calcitonin rapidly diminished urinary hydroxyproline excretion, indicating direct inhibition of bone resorption and collagen breakdown.

Calcitonin was subsequently purified from several species: mammals, birds, and fish (230). Purification of calcitonin from human thyroid gland proved to be challenging, because it is produced at very low levels. Human calcitonin (hCT) was eventually purified by Neher et al. (161) from patients with tumors of the C cells of the thyroid, who produced high levels of calcitonin. The study determined the amino acid sequence of hCT and found considerable variability between pig calcitonin and hCT, although both are 32–amino acid peptides that contain a disulfide bridge near the amino-terminal and have an amidated carboxyterminal. Linkage analysis mapped the hCT gene *CALCA* to the short arm of chromosome 11 (122). Although C cells of the thyroid are undoubtedly the major source of circulating calcitonin in humans, calcitonin-like immunoreactivity has also been identified in the prostate gland, the gastrointestinal tract, thymus, bladder, lung, and central nervous system (70). Interestingly, there is evidence to suggest the presence of calcitonin-like immunoreactivity in blood and urine samples from patients who had total thyroidectomy, indicating extrathyroidal secretion of calcitonin.

2. Calcitonin gene-related peptide

The organization of eukaryotic genes in exons and intervening "silent" introns, which are cleaved in the process of RNA maturation, was discovered simultaneously by a number of groups (76). Numerous studies demonstrating alternative splicing as a mechanism of post-transcriptional regulation followed this discovery. In a study of rat calcitoninproducing medullary thyroid carcinoma line, Rosenfeld et al. (180) found that in the process of serial transplantation of the carcinoma cell line, some of the tumors changed their phenotype and produced only very low levels of calcitonin. Analysis of these cell lines showed that with the change into low or no production of calcitonin, a new mRNA species, larger than calcitonin mRNA, appeared in the cytoplasm. Further investigations determined that the calcitonin gene CALCA encodes two alternative mRNA species, calcitonin and calcitonin gene-related peptide (CGRP), and that this alternative splicing was responsible for the decline in calcitonin expression in cell lines that switched to CGRP synthesis instead. Under normal physiological conditions, the processing of the CALCA gene transcript is tissue specific. In the thyroid C cells, mRNA transcribed from the calcitonin gene encodes a precursor of the hormone calcitonin, whereas in neuronal tissue, alternative splicing of the RNA generates mRNA encoding the precursor of the neuropeptide CGRP (181). A second, separate gene that encodes CGRP was identified later in both humans and rodents (8, 99). The product of this gene was named β CGRP, with the previously discovered CALCA-encoded product becoming α CGRP. β CGRP is the only mature transcript of the CALCB gene.

An additional product of the CALCA gene is procalcitonin, a 116-amino acid precursor of calcitonin. Under normal physiological conditions, the expression and processing of CALCA mRNA into the form that encodes procalcitonin is mostly restricted to C cells of the thyroid, where the precursor protein is quickly cleaved to generate mature calcitonin. The concentration of procalcitonin in the circulation in these conditions is very low (63). In the presence of a bacterial infection, the expression of the CALCA gene is induced in many organs and tissues, producing a rapid and substantial increase in the circulating levels of procalcitonin. Studies have shown that procalcitonin production is stimulated directly by bacterial endotoxins and lipopolysaccharides and indirectly by inflammatory mediators. However, the function of procalcitonin synthesized under microbial infection is still unclear. The rapid increase of procalcitonin in the circulation is specific to bacterial infection and is not seen during viral infection or inflammation of other causes. Therefore, procalcitonin is currently in clinical use as a biomarker for assessing risk of sepsis and septic shock (208). Procalcitonin has also been examined as a biomarker for bone and joint infection, and a meta-analysis of studies including a total of 583 patients concluded that it may be a useful predictor of osteomyelitis or septic arthritis (188). Recently, procalcitonin has been shown to inhibit osteoclast differentiation in bone marrow cultures, and interestingly, this activity was not mediated by the calcitonin receptor (CTR) (119).

3. Amylin

One of the pathophysiological features of type 2 diabetes is the presence of amyloid in the pancreatic islets of Langerhans. Amylin, or islet amyloid polypeptide (IAPP), was purified from pancreatic deposits of patients with type 2 diabetes (40) and from human insulinoma (217). Amylin is a 37-amino acid peptide that is structurally similar to CGRP, with ~50% identity between the two peptides. Human amylin monomers are soluble but can aggregate to form amyloid in type 2 diabetes patients and also spontaneously, in a concentration-dependent manner in vitro, in which they form amylin fibrils (124). The role of the amylin aggregates in the development of β cell lesions in type 2 diabetes is not entirely clear yet, although there is evidence that they contribute to cell death and the loss of islet β cell mass. Recent results from transplantation of human and transgenic animal islets suggest that oligomers of amylin play a key role in the progressive failure of β cells (25, 216). Shortly after the purification of amylin from the amyloid deposits, amylin was also found to be present in β cells of the healthy pancreas. Amylin is stored in the same cellular granules that contain insulin, although the concentration of amylin in the granules is only ~1%-2% of that of insulin. Amylin is cosecreted with insulin; hyperglycemia stimulates amylin secretion, whereas hypoglycemia reduces it (6, 164). The effect of amylin on glucose metabolism is opposite to that of insulin, as it stimulates glycogen breakdown in skeletal muscle (221). Apart from pancreatic β cells, amylin is produced in the gastrointestinal tract and the nervous system (226). In healthy individuals, circulating amylin levels are 5-10 pmol/L, rising to 10-20 pmol/L following a meal (67). Higher levels of circulating amylin were found in humans and animal models of obesity and type 2 diabetes (24, 88, 151, 184).

4. Adrenomedullin

Adrenomedullin was discovered by Kitamura et al., who isolated the peptide hormone from human pheochromocytoma by screening for factors that elevate cAMP levels in platelets (121). The same study showed that the peptide is also highly expressed in normal adrenal medulla and that it elicits a potent and long-lasting hypotensive effect, suggesting that adrenomedullin was a newly identified hormone that regulates blood pressure. Further studies demonstrated high expression levels of adrenomedullin throughout the vasculature and in cardiovascular organs, an expression pattern that was considered to reflect its central role as a vasodilator (28, 145). Other groups found that adrenomedullin is expressed in many tissues, and marked elevation of circulating adrenomedullin was found to be triggered by essential hypertension, renal failure, sepsis, and normal pregnancy (28). Adrenomedullin regulates the proliferation, migration, and differentiation of various cell types and affects a large number of physiological and pathological processes, including hormone release, inflammation, and oxidative stress (189).

5. Intermedin (adrenomedullin 2)

In fish, five different genes (Adm1-5) that encode a family of five adrenomedullin peptides have been identified, whereas in mammals, only one additional adrenomedullin family member has been found (195). In 2004, two groups discovered the mammalian gene ADM2, which encodes a peptide with high similarity to adrenomedullin, and was named intermedin or adrenomedullin 2 (177, 195). Takei et al. identified intermedin in human, mouse, and rat and determined that, in mice, intermedin mRNA was expressed in the submaxillary gland, kidney, stomach, ovary, lymphoid tissues, and pancreas, but not in the adrenal medulla. Intravenous injection of intermedin in mice decreased arterial pressure more potently than adrenomedullin. Roh et al. found that, in the rat, intermedin is primarily expressed in the pituitary and gastrointestinal tract and showed that intraperitoneal administration of intermedin decreased blood pressure in both normal and spontaneously hypertensive rats (98, 177).

6. CTR-stimulating peptide

CTR-stimulating peptide (CRSP) was isolated from porcine brain (114). The isolation of two additional similar peptides from porcine established a family of CRSP-1–3, with high degree of similarity to CGRP (115). CRSP-1 is expressed mainly in the thyroid gland and the central nervous system. Administration of CRSP into anesthetized rats decreased serum calcium, but did not alter their blood pressure. CRSPs have been identified in other mammals, including cattle, dog, and horse, but are absent in human, rat, and mouse (115).

B. Synthesis and Structure

Peptides of the calcitonin family in human are encoded by five genes with high degree of homology, suggesting that the family was derived from a primordial gene through duplication and mutation. Three of the genes, *CALCA*, *CALCB*, and *ADM*, are located on chromosome 11, whereas *IAPP* and *ADM2* are on chromosome 12 and 22, respectively. *CALCA* spans ~5.6 kb and contains six exons; exon IV encodes mature calcitonin and exon V encodes mature α CGRP, whereas exons I–III are included in both calcitonin and α CGRP mRNA (22). The splicing of the *CALCA* gene is tissue specific; in thyroid C cells, 99% of the primary RNA transcript is processed to produce calcitonin mRNA, whereas in neuronal tissue 95% is processed to encode α CGRP mRNA (137). The *CALCB* gene has similar structure to *CALCA*; however, because of sequence variation, the only mature peptide encoded by this gene is β CGRP, which differs from α CGRP by one amino acid in the rat and three amino acids in human. *ADM* encodes adrenomedullin and includes four exons, with exon IV encoding for the mature adrenomedullin peptide. The *IAPP* gene that encodes amylin and the *ADM*2 gene that encodes adrenomedullin 2 contain three exons each, with exon III encoding the mature peptide.

All the calcitonin-family peptides are synthesized as precursor proteins that undergo sequential proteolytic processing steps and post-translational modifications to produce the mature peptides. The peptides of the calcitonin family share a number of common structural features: a ring structure formed by a disulfide bridge between two cysteine residues at the NH₂-terminal, an α -helical mid-region, and an amidated amino acid at the carboxyl-terminal (FIGURE 1). The NH₂-terminal and COOH-terminal are highly conserved among calcitonin, CGPR, and amylin, whereas the middle region is more divergent (21). Studies of structure/function have established that the ring structure at the NH₂-terminal of the peptides is essential for receptor activation (17). Thus, linear peptides produced by truncation of the NH₂terminal domains, including amylin₈₋₃₇, α CGRP₈₋₃₇, and adrenomedullin₂₂₋₅₂, have been used as antagonists of the parent molecules, as they bind the receptors but fail to activate them (21). In contrast, the NH₂-terminal ring structures (amylin₁₋₈, α CGRP₁₋₈, and ADM₁₅₋₂₂) have been shown to retain some of the biological activity of the parent molecules (47, 51).

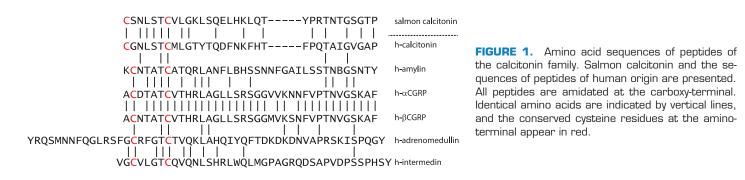
Calcitonin has been identified in a great number of species. In nonmammalian species, including birds, fish, and reptiles, calcitonin is derived from the ultimobranchial body (70, 162). Interestingly, in all the species examined, the cysteine residues at positions 1 and 7 and the overall length of 32 amino acids are conserved. Calcitonin preparations from two teleost species have been developed and used clinically. Elcatonin (Asu1–7 eel calcitonin analog) is a derivative of eel calcitonin, in which the NH₂-terminal amino

group was replaced by a hydrogen atom and the disulfidebond replaced by ethylene linkage (153). These modifications increased the stability of elcatonin in comparison to the parent molecule while retaining the biological activity. Elcatonin has been trialed for a number of indications and was recently shown to alleviate pain and inhibit bone resorption in patients with osteoporotic vertebral fractures (198). Salmon calcitonin (sCT) differs from eel calcitonin by three amino acids and shares 50% sequence identity with the human peptide. sCT has much higher biological potency in humans than hCT, and has, by far, been the most widely used preparation in clinical practice (33).

C. Receptors

CTR belongs to the seven-transmembrane domain class II (family B) G protein-coupled receptors (GPCRs), a group that includes other receptors that bind regulatory peptides, including PTH/PTH-related peptide, glucagon, vasoactive intestinal polypeptide, and secretin. The first CTR cDNA was cloned from porcine (134), followed by the cloning of the receptors from human and rat (7, 81, 127, 187). CTR is expressed in several tissues, including epithelial kidney cells, the central nervous system, and mature osteoclasts (172). The gene encoding the human CTR, CALCR, is located on chromosome 7 and contains 14 exons spanning 150 kb. A number of different isoforms of CTR are produced by alternative splicing; the most common ones differ from each other by the presence or absence of a 16-amino acid sequence in intracellular domain 1 (70). Two isoforms are also found in rodents, with a difference in a 37-amino acid region in extracellular domain 2. The insert negative form is the predominant one in both humans and rodents. The alternative splicing that produces the two isoforms, which appears to be cell type specific, has functional implications, as it affects both the affinity of the receptors to ligands of the calcitonin family and the downstream signaling mechanisms (70).

CTR-like receptor (CRLR) was cloned independently by two groups as a novel receptor with high sequence homology to the CTR (31, 163). The gene encoding CRLR in humans, *CALCRL*, is located on chromosome 2 and contains 15 exons spanning over 103 kb of genomic DNA. For



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several years, CRLR was an "orphan receptor," with no known ligands (72), until McLatchie et al. (146) discovered the family of receptor-modifying proteins (RAMPs) that form dimers with CRLR to produce specific receptors for the calcitonin-family peptides. Three RAMPs have been identified (RAMP1–3), and although they share only ~30% amino acid sequence identity, all have a similar structure: an extracellular NH₂-terminal, a single transmembrane α -helix, and a short intracellular COOH-terminal. The extracellular domain contains six cysteine residues in RAMP1 and 3, but only four residues in RAMP2. Evidence suggests that disulfide bridges between cysteines 2–4 and 3–6 are essential for CRLR/RAMP complex stability and function (90).

Dimerization of CRLR with RAMP1 forms a specific, highaffinity receptor for CGRP, whereas CRLR dimerization with either RAMP2 or RAMP3 produces receptors with high affinity to adrenomedullin (146). Later studies have shown that CRLR/RAMP complex also functions as receptor for intermedin, which binds in a nonselective manner to either of the three RAMPs/CRLR dimers (177). In addition, RAMPs were found to interact and modulate the pharmacology of CTR (11, 37, 156). Although CTR itself binds calcitonin, complexes of CTR with either of the three RAMPs form specific amylin receptors. Thus, the combinations of the CTR, CRLR, and RAMP1–3 have been recognized as the receptors for all the peptides of the calcitonin family, and a consensus nomenclature has been established **(FIGURE 2)** (37, 156, 169). Pharmacological studies of the interactions between peptides of the calcitonin family and their receptors have shown that each of the receptor combinations typically binds one of the calcitonin-family peptides with high affinity, whereas other members of the family can bind to the same receptor dimer with lower affinities (21). The specific binding affinities are also dependent on the experimental system (155). The cross-reactivity of members of the calcitonin family with the various receptor combinations presents a challenge for interpretation of experimental results, as deficiency in one specific component can be masked by interactions of the remaining members of the peptide and receptor families.

The regulation of GPCRs is a very active field of research, as GPCRs are the most commonly used drug targets, and it is estimated that 30%-50% of all medications currently in clinical use act through ~80 members of this receptor family. Pharmacological and molecular studies of the RAMP family uncovered novel mechanisms of regulation of GPCRs (89, 123). RAMPs have been found to regulate the activity of CTR and CRLR through a number of mechanisms: 1) RAMPs act as pharmacological switches, modifying the binding specificities of the GPCRs to their ligands. The crystal structure of CRLR/RAMP1 and CRLR/RAMP2 has been recently solved, and it was found that RAMPs alter GPCR ligand binding pocket first by allosteric changes and then by providing distinct contact sites for ligand interaction (20). 2) Receptor trafficking: CRLR by itself does not function as an independent receptor, as it is not expressed

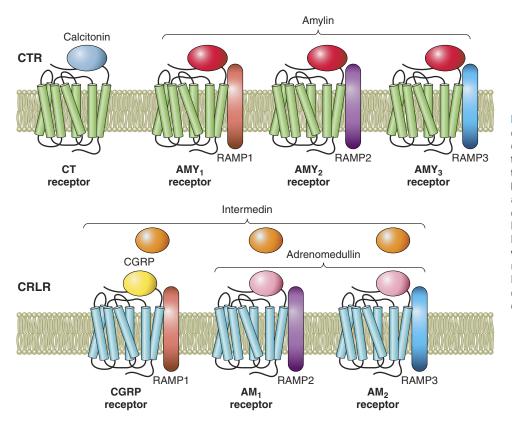


FIGURE 2. Receptors and ligands of the calcitonin family. The seven-transmembrane calcitonin receptor (CTR) by itself binds calcitonin, and when in a dimer with one of the three receptor activity-modifying proteins (RAMPs), it produces a receptor with high affinity to amylin. Calcitonin receptor–like receptor (CRLR) requires a RAMP partner to be presented on the membrane and bind its ligands, forming a high affinity CGRP receptor when associated with RAMP1 and adrenomedullin receptor with RAMP2 or RAMP3. Intermedin binds to dimers of CRLR with any of the RAMPs. The official names of the receptors are indicated below each diagram.

on the cell membrane. RAMPs are necessary to chaperone CRLR to the cell membrane. 3) Receptor desensitization: following ligand binding, the complex of GPCR and RAMP are typically internalized by the cell. A study of CRLR in complex with the three different RAMPs found that while in complex with RAMP1 and RAMP2, internalization of the receptor is followed by degradative pathways; RAMP3 causes CRLR to be recycled back to the cell surface, allowing for rapid receptor resensitization (19, 123). 4) Signaling: studies of the amylin receptors (AMY_{1-3}) found that downstream signaling pathways of intracellular calcium production depend on the specific interacting RAMP and suggested that RAMP-complexed receptors have a different signaling profile to that of CTR expressed in the absence of RAMPs (152). A recent study of the pharmacology of CRLR/RAMP receptor complex demonstrated that the receptors display both ligand- and RAMP-dependent signaling bias among the downstream $G\alpha$ subunit activation (218).

Pharmacological studies of receptor/ligand interactions of the calcitonin family explored the mechanisms that contribute to the greater potency of sCT in comparison to hCT. A recent study demonstrated that these ligands are equipotent during short-term stimulation. However, long-term stimulation with sCT results in sustained activation of CTR, whereas hCT loses its activity much earlier (9). Pharmacological studies have established that binding of sCT to rat and human CTRs is essentially irreversible, whereas hCT rapidly dissociates from the receptor. This dissociation was found to be independent of G protein coupling and related to the ability of the ligand to form amphipathic α -helical secondary structure. The higher potential of sCT to form these structures, in comparison to hCT, appears to depend on its NH₂-terminal residues. In structure/function studies, chimeras of sCT and hCT were made, and sCT peptides, which had the NH₂ terminus substituted by the 13-21 residues of hCT, completely dissociated from receptors, whereas combinations of sCT (1-16) and hCT (17-32) retained the irreversible binding of sCT (92). Recently, studies that compared the binding of sCT and hCT to the CTR have found that the different ligands not only affect receptor conformation, but also modulate the bound G protein and the kinetics of downstream signaling (9, 75).

II. THE ACTIVITY OF THE CALCITONIN-FAMILY PEPTIDES IN BONE: IN VITRO STUDIES AND IN VIVO STUDIES IN WILD-TYPE ANIMALS

A. The Role of Endogenous Calcitonin in Hypercalcemia

The finding that calcitonin reduces the level of calcium in the circulation led to the hypothesis that its physiological role might be in restoring normal levels of serum calcium in

states of hypercalcemia. This hypothesis was tested in a number of early in vivo studies in rats that were either parathyroidectomized (PTX) to remove PTH-secreting cells or thyroparathyroidectomized (TPTX) to remove both PTH- and calcitonin-secreting C cells. When hypercalcemia was induced directly by injection or infusion of calcium, the presence of the thyroid gland was necessary to lower the levels of circulating calcium (78, 196). Similar results were seen when hypercalcemia was induced by injection of parathyroid extract or with partially purified PTH preparation, as serum calcium levels were significantly higher in TPTX rats, confirming the thyroid origin of calcitonin and its role in acute hypercalcemia (94). Secondary hyperparathyroidism and resistance to the calcemic action of PTH are a common finding in renal failure. Rodriguez et al. have shown that the presence of the thyroid gland and the production of endogenous calcitonin are important in decreasing the calcemic response to PTH in rats in both PTHinduced hypercalcemia in the context of renal failure and in diet-induced hyperparathyroidism (176). A physiological role for calcitonin in protection against bone loss, induced by other hormones, has also been suggested. Calcitonindeficient TPTX rats that were treated with PTH to induce hypercalcemia showed a significant cancellous bone loss in the proximal tibia, whereas calcitonin-sufficient PTX rats had no bone loss, suggesting a protective role for calcitonin in these conditions (225). Following observations that ovariectomy (OVX)-induced osteopenia in rats is associated with a decrease in circulating calcitonin, it has been suggested that calcitonin mediates estrogen deficiency-induced bone loss. However, experimental evidence has not provided support for this hypothesis. For example, the OVX-induced decrease in femur density and calcium content were similar in thyroidectomized rats and in animals with intact thyroid (109).

The main strength of the studies described above is the use of in vivo animal models that allowed the careful manipulation of hormone levels by removal (and in some cases, autotransplantation) of the thyroid, parathyroid, and the ovaries. These studies confirmed that the thyroid is the main source of calcitonin and demonstrated the capacity of endogenous calcitonin to respond to hypercalcemia induced by different protocols. Despite these obvious advances in scientific knowledge, these studies did not provide the answer to the fundamental question of the physiological role of CT, but rather defined its role in a pathological environment. In later years, the development of genetically modified mouse models have provided a much better experimental platform for further investigations of the role of endogenous calcitonin in situations of calcium stress as well as the physiological role of calcitonin.

B. Osteoclasts and Bone Resorption

Mature osteoclasts, formed by fusion of hematopoietic precursor cells, are unique multinuclear cells whose primary

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function is to resorb bone. The differentiation and activity of osteoclasts are regulated by a large number of local and systemic factors. Early stages of osteoclast differentiation are initiated by the binding of macrophage colony-stimulating factor (M-CSF) to its receptor c-FMS, which, in turn, induces the expression of receptor activator of nuclear factor kappa-B (RANK), a membrane protein expressed by preosteoclasts. The interaction between RANK and RANK ligand (RANKL), which is expressed by cells of the osteoblast lineage, is the major trigger of osteoclast differentiation and activation. Osteoprotegerin (OPG), a decoy receptor secreted from cells of the osteoblast lineage, binds to RANKL and inhibits the RANK/RANKL interaction in a competitive manner. Thus, the ratio between the levels of OPG and RANKL is a key factor in the control of osteoclast activation and bone resorption (120). Mature osteoclasts degrade bone extracellular matrix through a specialized mechanism, which has been studied in detail. When fully differentiated osteoclasts come into contact with mineralized bone matrix, they form a "sealing zone," an F-actinrich, ring-like structure that surrounds the enclosed space of the resorption lacuna. The membrane of the osteoclast within the sealing zone is folded and forms a ruffled border through which protons and matrix-degrading enzymes are released in the process of bone resorption.

Studies carried out shortly after the discovery of calcitonin established that inhibition of bone resorption is the main mechanism for the rapid drop in circulating calcium levels induced by calcitonin (230). In the following years, the activity of calcitonin in osteoclasts has been described in detail, and with the discovery of the additional members of the calcitonin family, their activities in osteoclasts have also been investigated and compared with that of calcitonin (53). The expression of CTR, CRLR, and RAMP1–3 has been detected in osteoclasts, suggesting that all the members of the calcitonin family can bind to these cells (84).

1. Calcitonin

Binding of calcitonin to its receptor on osteoclasts has major and immediate consequences: within minutes, there is a loss of the ruffled border, followed by cell retraction and arrest of cell motility and bone resorption (29, 30). A number of signaling mechanisms mediate the activity of calcitonin in osteoclasts; the inhibition of motility and induction of a quiescent state are cAMP dependent, whereas the retraction and disruption of the resorption activity in the sealed zone are mediated through intracellular calcium signaling (3, 229). Studies of the mechanisms involved in the detachment of the sealing zone found that calcitonin modulates the phosphorylation and intracellular distribution of Src and of the tyrosine kinase Pyk2, which is highly expressed in osteoclasts and is localized mainly in the sealing zone (190, 231). A short pretreatment of mature mouse osteoclasts with sCT had no effect on the number of mononuclear or multinuclear osteoclasts that developed in culture but reduced the capacity of the pretreated cells to resorb bone (212). The cells pretreated with sCT produced pits of smaller size than control cells, suggesting a lasting inhibitory effect of sCT on osteoclast motility.

A number of studies investigated the effect of calcitonin on early stages of osteoclast differentiation. Cornish et al. have shown that in murine bone marrow cultures stimulated to generate osteoclasts by 1,25(OH)₂D₃, calcitonin dose dependently decreased the number of tartrate-resistant acid phosphatase (TRAP)-positive multinucleated cells and substantially reduced the ratio of TRAP-positive multinucleated to mono- and binucleated cells, indicating an inhibitory effect on fusion of osteoclast precursors (44). In contrast, an earlier study of murine bone marrow cultures treated with $1,25(OH)_2D_3$ found that the presence of sCT had no effect on the number of mononucleated and multinucleated TRAP-positive cells (106). In cultures treated with sCT, the cells had very low expression of CTR in comparison to the controls, but the overall level of TRAP expression was unaffected by sCT. When the number of nuclei within the TRAP-positive multinucleated cells were counted, the mean number was lower with sCT treatment, indicating a subtle negative effect of sCT on cell fusion. The main difference between the two studies was the protocol of sCT supplementation. In the latter study by Ikegame et al., a continuous treatment protocol was used with sCT added daily at 0.1 nmol/L. In the study by Cornish et al., sCT, at concentrations ranging from 0.1 pmol/L to 1 nmol/L, was replenished every second day, and it is therefore possible that the cells were exposed to fluctuating levels of sCT. The discrepancies between the results of the two studies suggest tight regulation of osteoclast differentiation in response to sCT, which is likely to be controlled by complex downstream signaling mechanisms.

The use of bone marrow cultures to investigate the effect of sCT on osteoclast differentiation cannot differentiate between direct activity on osteoclast precursors and indirect effect mediated by cells of mesenchymal origin that are present in the bone marrow. The potential direct effect of sCT on osteoclast precursors was studied in two experimental systems: in cells isolated from mouse spleen and in a highly purified preparation of macrophages from mouse bone marrow (83). In cultures of mouse spleen cells, and in bone marrow macrophages (BMM) induced to differentiate by M-CSF and RANKL, sCT inhibited the formation of TRAP-positive multinucleated cells and resorption pits. The cultures contained a large number of TRAP-positive mononucleated cells, indicating the inhibition of cell fusion. Surprisingly, no changes were identified in the levels of expression of a number of genes important for osteoclast progenitor cell differentiation. The inhibitory effect of sCT in this experimental system could be reproduced by activation of cAMP and protein kinase A (PKA) (83).

2. CGRP

Similar to the effects of calcitonin, injection of CGRP into rabbits and rats produced hypocalcemia (179, 202). A large number of studies found that CGRP inhibits osteoclast activity and bone resorption, although CGRP's potency is consistently much lower than that of calcitonin. In cells cultured from mouse bone marrow and induced to differentiate into osteoclasts by M-CSF and RANKL, the area of resorption pits was reduced by CGRP in a concentration of 0.1 nM or higher, whereas higher concentrations of 10 nM CGRP were required for the inhibition of TRAP-positive cell formation, suggesting that CGRP acts with higher potency to inhibit osteoclast activity than osteoclast differentiation (213). In mouse bone marrow cultures treated with 1,25(OH)₂D₃, CGRP also inhibited the formation of TRAP-positive mono- and binuclear cells and the subsequent fusion of these cells to form multinucleated osteoclasts (44). Reduced formation of osteoclasts was also observed in human bone marrow cultures following treatment with CGRP (2). In this experimental system, CGRP was shown to bind specifically to osteoclast precursors and regulate osteoclast development.

The effect of CGRP on bone resorption has also been assessed in preclinical animal models. OVX is a standard procedure performed in rodents and used as a model for estrogen deficiency–related osteoporosis. When injected into OVX rats, CGRP had an inhibitory effect on the increased indices of bone resorption (207). In this experiment, CGRP was less effective than sCT in suppressing bone resorption, although it was tested at a concentration that was 500 times higher than that of sCT. The reduced magnitude and the low potency suggest that this might be a nonspecific effect of CGRP and not necessarily an activity elicited by binding of CGRP to its own receptor (207). In a local injection model, application of CGRP over the calvaria of adult mice had no significant effect on bone resorption (45, 54).

Recent studies have suggested a role for CGRP in protecting the bone against detrimental effects associated with bone implants. Following joint arthroplasty, aseptic loosening can occur as a result of wear particles that are released from the implant and induce local increase in bone resorption. This process of particle-induced osteolysis is a common cause of early implant failure. The presence of CGRP in the skeleton and nerve fibers adjacent to sites of periprosthetic osteolysis led a number of research groups to test the hypothesis that CGRP has a protective role by inhibiting osteolysis by osteoclasts. The effect of CGRP on the catabolic activity of ultra-high molecular weight polyethylene (UHMWPE) particles was studied in vitro in primary human osteoblasts and in the human osteoblast-like cell line MG-63 (117, 224). In both cell types, UHMWPE particles induced RANKL expression and inhibited the expression of OPG, whereas CGRP reduced UHMWPE-

induced RANKL expression, suggesting an indirect inhibition of particle-induced bone resorption.

3. Amylin

Early studies have found that, similar to calcitonin and CGRP, injection of amylin strongly induced hypocalcemia with a potency that was either equal or lower than that of calcitonin (59, 77, 138, 220, 228). In mouse bone marrow cultures that were stimulated to generate osteoclasts by $1,25(OH)_2D_3$, amylin inhibited the formation of TRAP-positive mono- and binuclear cells as well as the fusion of these cells and the formation of multinucleated osteoclast-like cells (44). A later study found that the activity of amylin to inhibit cell fusion required the activation of the extracellular signal-regulated protein kinase 1/2 (ERK1/2) in osteoclast precursors (58).

Amylin also inhibits bone resorption by mature osteoclasts, as determined by the reduced number of resorptive pits per TRAP-positive multinucleated cell in bone marrow cells cultured on bone slices (44). In an organ culture system of neonatal mouse calvariae, in which bone resorption was stimulated by $1,25(OH)_2D_3$, amylin increased cAMP levels and reduced both basal and PTH-stimulated resorption (51, 166). Amylin was also shown to inhibit bone resorption in organ cultures of fetal mouse long bones, in which its potency was similar to that of CGRP, yet 60-fold less than that of hCT (197).

In vivo, daily administration of amylin to adult mice, either locally over the calvariae for five days or systemically for one month, produced a reduction of 60%–70% in indices of bone resorption (45, 48). In a study of the activity of amylin in the context of estrogen deficiency in rats, amylin was injected for 30 days starting 60 days after the OVX surgery. In this experimental model, amylin reduced urinary excretion of deoxypyridinoline and reduced trabecular bone loss, whereas it had no effect on cortical bone indices (101).

4. Adrenomedullin

Despite the presence of adrenomedullin receptors on osteoclasts and the ability of adrenomedullin to induce the synthesis of cAMP in these cells, studies have consistently shown that adrenomedullin does not affect the differentiation and activity of osteoclasts. This has been demonstrated in bone marrow cultures stimulated to generate osteoclasts with $1,25(OH)_2D_3$, or M-CSF and RANKL, and in ex vivo calvarial cultures stimulated with adrenomedullin (43, 46, 82). This is in contrast to all other members of the calcitonin family.

Nevertheless, in certain pathological situations that induce bone loss, adrenomedullin appears to act indirectly to inhibit bone resorption through the modulation of the inflammatory environment. In an in vitro model, rheumatoid synovial fibroblasts were treated with the proinflammatory factors IL-1 β and TNF- α and then cocultured with peripheral blood mononuclear cells. The treated cells stimulated osteoclast formation, whereas the addition of adrenomedullin in this system modulated RANKL and OPG expression in the rheumatoid synovial fibroblasts and attenuated the increased osteoclast formation (227). In an experimental model of collagen-induced arthritis in mice, adrenomedullin and its truncated form, ADM₂₂₋₅₂, reduced TNF- α , IL-6, and IL-17 expression in the joints and increased the expression of IL-4 and IL-10. This immune regulation was associated with decreased cartilage degradation and systemic bone loss, suggesting a protective skeletal effect (1).

5. Intermedin

The activity of intermedin in osteoclasts differs from that of adrenomedullin and is similar to that of calcitonin, CGRP, and amylin. In cultures of BMM stimulated with M-CSF and RANKL, intermedin inhibited the formation of multinucleated osteoclasts as well as their ability to resorb bone, as measured by the number of pits formed when cultured on bovine bone slices. Intermedin's activity in this experimental system was mediated by cAMP (82). Recent studies suggested that intermedin might be able to inhibit osteoclast formation indirectly, as MC3T3 osteoblastic cells treated with intermedin had reduced expression of RANKL and M-CSF and increased expression of OPG (175).

6. Comparative studies

A number of studies directly compared the activity of different peptides of the calcitonin family on osteoclast formation and activity. sCT, human amylin, and human CGRP all inhibited osteoclast formation in mouse bone marrow cultures treated by $1,25(OH)_2D_3$ (44). sCT inhibited osteoclast differentiation at concentrations of 0.1 pmol/L and above, whereas CGRP and amylin were much less potent and were active at concentrations of 1 nmol/L and above. sCT, but not CGRP or amylin, reduced the ratio of TRAPpositive multinucleated to mono-/binucleated cells (44). Similar results were found by Granholm et al. (83), who demonstrated that sCT and hCT inhibit osteoclast formation in mouse spleen and mouse BMM, with half maximal inhibition at 1 pmol/L for sCT and 3–4 orders of magnitude higher for hCT (83).

The activity of calcitonin-family peptides on osteoclast formation was also investigated in mouse BMM that were either treated with M-CSF alone to expand the population of osteoclast progenitors or with M-CSF and RANKL to induce further differentiation into TRAP-positive multinucleated, osteoclast-like cells (84). In BMM cells treated with M-CSF, the mRNA and proteins of CRLR and RAMP1–3 were expressed, and treatment of the cells with amylin, CGRP, adrenomedullin, or intermedin induced the production of cAMP (84). However, the addition of RANKL was necessary to induce the expression of CTR, and only when CTR was expressed the cells became responsive to sCT. It is interesting to note that amylin was active in BMM cells treated with M-CSF alone, although CTR was not expressed in these cells, suggesting that, in these cells, amylin signals through a receptor other than CTR/RAMP1–3.

The effects of peptides of the calcitonin family on PTHstimulated bone resorption were determined by measuring ⁴⁵Ca release from mouse calvaria in organ cultures (82). Comparison of the inhibitory effects of the peptides on bone resorption clearly demonstrated that sCT had the highest potency, with half-maximal inhibition (IC₅₀) at 3 pmol/L, whereas CGRP and amylin had IC₅₀ at 10–30 nmol/L, intermedin at 300 nmol/L, and adrenomedullin was without effect (82).

The effects of calcitonin, CGRP, and amylin on isolated osteoclasts have also been compared through the use of time-lapse video microscopy and image analysis. CGRP and amylin reduced osteoclast motility, but only calcitonin reduced both motility and osteoclast retraction. This study demonstrated that the effect on cell motility is mediated via the cAMP signaling pathway, whereas the effect on retraction appears to be mediated by changes in intracellular calcium. The authors concluded that CGRP and amylin activate only the cAMP pathway, whereas calcitonin also acts on osteoclasts via changes in intracellular calcium and suggest that this may contribute to the greater potency of calcitonin in inhibiting bone resorption (4, 5).

7. The "escape phenomenon"

Early studies have demonstrated that when calcitonin was used clinically to inhibit bone resorption in hypercalcemic cancer patients, the effects of calcitonin were lost after a few days, an effect later termed the "escape phenomenon" (158, 219). In vitro, this effect was first identified in organ cultures, in which, initially, calcitonin inhibited the stimulation of bone resorption by PTH, but after prolonged exposure to calcitonin, its inhibitory effect was lost (215). The desensitization of the cells to calcitonin was found to be the result of a ligand-induced internalization of the CTR, as well as inhibition of de novo synthesis of the receptor, and has been confirmed in both human and murine osteoclast cultures (194, 212). In cultures of mouse bone marrow treated with 1,25(OH)₂D₃, in which multinucleated TRAPpositive cells were abundant, a brief, one-hour treatment with sCT markedly reduced the expression of CTR mRNA, which remained reduced for the 72 h it was monitored (173). Subsequent studies established that the escape phenomenon is not unique to calcitonin, and can be induced by other members of the calcitonin family. When amylin was tested in the experimental system of neonatal mouse calvaria, it was also found to produce a transient inhibition of bone resorption by PTH (166). More recently, it has been shown that all members of the calcitonin family downregulated the expression of CTR mRNA, whereas CRLR levels remained unchanged (82). This observation indicates that the downregulation of CTR expression is not limited to peptides that directly bind CTR, but can also be induced, probably through an indirect mechanism, by calcitoninfamily peptides that bind CRLR/RAMPs (82).

C. Osteoblasts and Bone Formation

Osteoblasts, the mononuclear bone–forming cells, differentiate from mesenchymal stem cells in the bone marrow. Under a tightly coordinated sequence of molecular signaling, preosteoblasts differentiate into mature osteoblasts that produce and lay down the extracellular bone matrix and subsequently mineralize it. Following a period of active bone formation, the osteoblast can either become embedded within the bone matrix and undergo terminal differentiation into osteocyte, remain on the bone surface in a quiescent state, or die through apoptosis.

1. Calcitonin

A number of studies in vitro and in vivo examined the effect of calcitonin on osteoblast proliferation and bone formation. In one of the earlier studies, calcitonin was found to stimulate osteoblast proliferation and bone formation when administered during the initial phases of bone formation, but when administered after the initiation of bone formation, it had an inhibitory effect (214). Subsequent studies found a direct stimulatory effect of calcitonin on osteoblast proliferation as well as increase in indices of bone formation (68, 209). In contrast, calcitonin had no effect on proliferation of primary rat osteoblasts in culture and did not alter bone formation indices in a local injection in adult mice (45, 47). The understanding of any direct effects of calcitonin in osteoblasts is challenged by the reproducible experimental evidence showing that CTR is not expressed in these cells (160). A possible mechanism for the demonstrated activity of calcitonin in osteoblasts could be nonspecific interactions of calcitonin with receptors other than CTR because of the high peptide concentrations used. In addition, it is possible that the primary osteoblast cultures contained some proportion of other bone cells and an indirect effect in osteoblast was measured. Later studies, using genetically modified animals, produced evidence showing that calcitonin does, in fact, regulate bone formation, but rather than a direct effect in osteoblasts, this activity is mediated via osteoclasts and osteocytes (see below).

2. CGRP

Unlike calcitonin, CGRP appears to have a positive effect on osteoblasts in vitro, but does not affect bone formation in vivo. Specific binding of CGRP to cells of rat calvaria was demonstrated (186). CGRP treatment increased cAMP levels in the UMR 106-01 rat osteosarcoma cell line and in other osteoblastic cell lines and primary osteoblast cultures (18, 148, 200). In primary bone cell cultures from neonatal chicken, rat, and mouse calvariae, CGRP increased cAMP levels, whereas calcitonin had no effect on these cells (149). Intracellular calcium is another second messenger involved in CGRP signaling in osteoblast-like cells, as CGRP treatment increased intracellular calcium levels in UMR 106-01 osteosarcoma cells and in two human osteoblastic cell lines: MG-63 and OHS-4 (23, 65, 118). CGRP induced the proliferation of primary osteoblasts, but its potency was much lower than that of amylin (49). Furthermore, the proliferative effect of CGRP was not inhibited by its antagonist $CGRP_{8-37}$ (210). This has raised the possibility that the effects of CGRP and amylin on osteoblast proliferation are mediated by a common receptor, which has a higher affinity for amylin (49). α CGRP was also shown to stimulate osteoblast differentiation in cultured rat bone marrow cells. whereas in this experimental system, β CGRP had no osteogenic effect (96).

In primary cultures of human osteoblast-like cells, CGRP has been shown to inhibit apoptosis through a Wnt/β catenin-dependent pathway and stimulate differentiation by increasing BMP-2 expression (154, 201). There is also evidence that CGRP induces differentiation of osteoblast precursors (157, 200). Recent studies demonstrated that CGRP induced the differentiation of bone marrow stromal cells into mineralizing osteoblasts in cells derived from either healthy or OVX rats (133, 213). The effect of CGRP on differentiation of bone marrow stromal cells into osteoblasts appears to be mediated via induction of the Wnt/ β -catenin pathway (232). Studies of osteoblast differentiation in three-dimensional cultures have also demonstrated that CGRP promotes osteogenesis of primary rat osteoblast-like cells cultured on Bio-Oss bone graft substitute and the differentiation of adipose-derived stem cells cultured within calcium alginate gels (102, 132). Despite the abundance of evidence that CGRP has positive effects on osteoblasts in vitro, local injection of CGRP in adult mice had no effect on osteoblast indices (45), whereas OVX rats treated with CGRP had no change in bone formation rates (207).

3. Amylin

Early studies have shown that amylin stimulates cAMP production in a pre-osteoblastic cell line and in both primary fetal rat osteoblasts and primary human osteoblasts, thus identifying bone as a potential target of amylin (45, 197, 211). In studies exploring the signaling pathways responsible for amylin's proliferative effect, it was demonstrated that phosphorylation of ERK1/2, most likely through activation of G_i proteins, was required for the mitogenic effect of amylin in rat osteoblast-like cells, as amylin's proliferative effect was inhibited by PD-98059, a specific inhibitor of this pathway (52). Surprisingly, the proliferative effect of amylin in osteoblasts required the presence of an IGF-1 receptor despite the fact there was no direct binding of amylin to IGF-1 receptor, nor was there a paracrine effect of osteoblast-derived IGF-1 (52).

Structure/function studies of amylin found that the NH₂terminal octapeptide fragment $\operatorname{amylin}_{1-8}$ stimulated primary rat osteoblast proliferation and thymidine incorporation in ex vivo cultures of neonatal mouse calvariae (50, 51). The availability of a short peptide that retains the beneficial bone effects, but is devoid of amylin's activity on energy and carbohydrate metabolism, provides an opportunity for the development of amylin-based therapy for bone. Stable analogs of $\operatorname{amylin}_{1-8}$ for potential use in osteoporosis are under development (125, 126).

In vivo, amylin acts to stimulate bone formation. When administered locally over mouse calvariae daily for 5 days, amylin induced two- to fourfold increases in histomorphometric indices of osteoblast activity (45). Systemic administration of amylin to adult mice also produced a 30%-100% increase in these indices (48), whereas in healthy rats, and in osteopenic OVX rats, amylin injections induced an increase of serum osteocalcin (100, 178). Amylin infusion has also produced osteogenic effects in diabetic rat models, although its efficacy varied depending on the model (86). Local injection of $\operatorname{amylin}_{1-8}$ over the calvariae of healthy female mice had a positive effect on bone formation, which was greater than that of an equimolar dose of human PTH_{1-34} , whereas systemic administration to adult male mice produced a near twofold increase in histomorphometric indices of osteoblast activity (47). In contrast, $\operatorname{amylin}_{1-8}$ administration to OVX rats had no effect on parameters of bone formation (66).

4. Adrenomedullin

Similar to amylin, adrenomedullin induced the proliferation of primary human and rat osteoblasts in vitro and in an ex vivo neonatal mouse calvaria culture (46, 87). Adrenomedullin is quite highly expressed in osteoblasts, and therefore could be acting in these cells through a paracrine/ autocrine mechanism (159). Unlike other members of the CT family, adrenomedullin has only modest effects on cAMP concentrations in osteoblasts, with studies indicating that adrenomedullin activates ERK1/2 and voltage-dependent calcium channels in these cells (199, 206). Although adrenomedullin does not bind directly to IGF-1R, similar to amylin, its proliferative effect in osteoblasts appears to depend on the presence of IGF-1R (52). Adrenomedullin acts as a survival factor and inhibits osteoblast apoptosis, likely through the ERK1/2 signaling pathway, CREB activation, and the Wnt signaling pathway (130, 206).

When injected locally over the calvaria of adult male mice for five days, adrenomedullin increased indices of osteoblastic activity two- to fourfold and significantly increased bone area (46). Furthermore, systemic treatment with the ADM_{27-52} fragment increased trabecular bone volume and cortical width in the tibia as well as enhancing bone strength in the humerus (43).

5. Intermedin

Intermedin is the least-studied peptide of the CT family in regard to its effects on osteoblasts and bone formation. In vitro studies have demonstrated that intermedin does not affect MC3T3 osteoblast-like cell proliferation or differentiation, yet it does appear to inhibit dexamethasone and serum starvation-induced apoptosis, suggesting an overall positive effect in these cells (175). Further studies of the effect of intermedin in osteoblasts are required to better understand its activity in these cells and its potential effect on bone formation.

D. Osteocytes

Osteocytes are terminally differentiated osteoblasts that reside in lacunae within the mineralized bone tissue and communicate with each other and with other cells and tissues through dendritic processes across a network of interconnected canaliculi. Osteocytes play central roles in the regulation of bone turnover and mineral metabolism. Recent studies found that osteocytes develop osteoclast-like properties and are able to remove bone matrix in their immediate surroundings in a process of osteolysis (171). Osteocytes were found to express genes that had been previously considered as osteoclastic markers, including TRAP and cathepsin K (171). One of the best-studied osteocyte-derived factors is sclerostin, a glycoprotein encoded by the SOST gene. Inactivating mutations in SOST were identified as the cause of rare diseases characterized by bone overgrowth and general osteosclerosis (10). Sclerostin binds to LRP4 chaperone and LRP5/6 coreceptors and inhibits the Wnt/βcatenin signaling pathway (64). Studies in transgenic animals found that sclerostin potently inhibits bone formation and stimulates bone resorption through a combination of direct effects on osteoclast precursors and indirect effects through the RANKL/OPG pathway. Currently, a humanized monoclonal antibody-targeting sclerostin is in advanced stages of development as an antiosteoporotic drug (55, 182).

The MLO/Y4 cell line was established from bones of transgenic mice expressing the SV40 large T antigen under the control of the osteocalcin promoter (116). MLO/Y4 cells produce extensive dendritic processes when cultured and present other osteocyte-like properties and have, therefore, been used by many research teams as a model for the study of osteocytes. The activity of calcitonin in MLO/Y4 cells has been studied by a number of groups. Expression of the CTR in MLO/Y4 cells was suggested by the specific binding of [¹²⁵I]sCT to these cells and the downstream induction of phosphorylation of ERK1/2 and increase in the intracellular levels of cAMP (167). Induction of apoptosis in MLO/Y4 cells by etoposide, TNF- α , or glucocorticoids was prevented by pretreatment of the cells with 5-10 ng/mL sCT. The authors suggest that the therapeutic effect of calcitonin in diseases such as glucocorticoid-induced osteoporosis may be partly due to inhibition of osteocyte apoptosis (167). Later studies demonstrated that the activity of calcitonin in osteocytes is not restricted to the MLO/Y4 cell line. In an animal study that examined the importance of osteoclast activity to the anabolic effect of PTH, young rats receiving anabolic PTH regimen were injected with sCT to transiently inhibit osteoclast activity (80). Unexpectedly, the study found that sCT induced the expression of Sost and inhibited the expression of other osteocyte genes: Mepe and *Dmp1*. A possible direct effect of calcitonin in osteocytes was suggested and supported by the finding that CTR was expressed in osteocytes that were freshly isolated from rat calvaria, although it was undetectable in cultured primary osteocytes (80). Extension of the study to older animals found that in 6-mo-old rats, sCT had no effect on the expression of Sost, Dmp1, and Mepe (79). Calcr mRNA was expressed in osteocytes isolated from 3-wk-old mice, but its level declined as mice aged and was undetectable at 49 wk of age. The authors concluded that the activity of calcitonin in osteocytes is likely to be physiologically relevant in young rodents (79). Expression of the CTR was also demonstrated in osteocytes isolated from young DMP1/green fluorescence protein transgenic mice (36). Using specific lineage surface markers to isolate a highly purified preparation of osteocytes, the study confirmed that Calcr, Ctsk, and Acp5 (TRAP), all considered as osteoclast-marker genes, are, in fact, also expressed in osteocytes (36).

E. Interpretation of In Vitro and In Vivo Studies in Wild-Type Animals

FIGURE 3 presents a summary diagram of the activity of the calcitonin-family peptides in bone cells in the context of bone remodeling. Although the activities of the peptides vary in both specificity and potency, it is evident that an overall positive effect is produced by the calcitonin-family peptides in bone. It is important to note that the experiments discussed in the current section under the title "The activity of the calcitonin-family peptides in bone in vitro studies and in vivo studies in wild-type animals" were conducted using a vast range of peptide concentrations. Under normal physiological conditions, serum levels of the calcitonin-family peptides in humans are within the picomole per liter range. In the studies described above, peptides were used from this low physiological concentration and up to concentrations as high as micromole per liter. With the exception of CGRP, which can reach high local concentrations in the bone microenvironment, experiments testing peptides of the calcitonin family at high pharmacological concentrations have to be considered as such and are not necessarily directly relevant to the physiological activities of these peptides. Furthermore, the fact that sCT is widely used in experimental systems of mouse, rat, and human certainly restricts the interpretation of the results to pharmacological, rather than physiological, conclusions. However, although the physiological relevance might be questionable, the study of sCT and amylin at high concentrations is clinically relevant given the use of sCT as a drug for different indications and the current use of the synthetic analog of human amylin, pramlintide, in patients with diabetes (170). As discussed below, the development of genetically modified animal models was critical in advancing our understanding of the physiological bone activity of the calcitonin-family peptides.

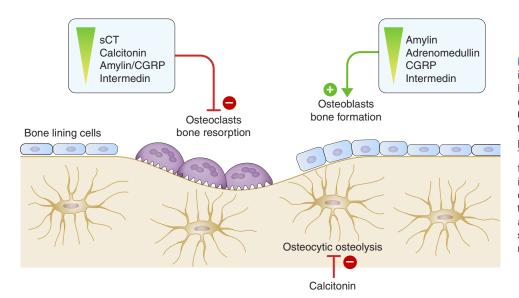


FIGURE 3. The effects of calcitonin-family peptides on bone cells. In vitro studies have established an overall positive effect of the calcitonin-family peptides in bone. Calcitonin, CGRP, and amylin are inhibitory to osteoclasts, and CGRP and amylin also promote osteoblast formation and activity. The osteocyte appears to be a target cell for calcitonin, as studies in genetically modified mice suggested a protective role for calcitonin in osteocytic osteolysis. The peptides are listed in order of potency, which is also indicated by the color gradient. sCT, salmon calcitonin; CGRP, calcitonin generelated peptide.

III. THE PHYSIOLOGICAL ROLE OF CALCITONIN PEPTIDES IN BONE: STUDIES OF GENETICALLY MODIFIED ANIMALS

A. Calcitonin: Physiological Role

Since the discovery of calcitonin over 50 yr ago (42), its pharmacological activity to lower circulating calcium levels through the inhibition of bone resorption has been thoroughly investigated and described in great detail. It is, therefore, surprising that the physiological role of calcitonin is still unclear. Calcium metabolism and bone mineral density are unaffected in patients with medullary thyroid carcinoma who have long-term excess of endogenous calcitonin levels, or in patients who had undergone thyroidectomy and have undetectable circulating calcitonin (105, 223). Therefore, as deficiency or excess of calcitonin secretion do not appear to have any pathological outcomes, it has been suggested that calcitonin has no physiological role in mammals. One of the codiscoverers of calcitonin expressed the view that calcitonin might be in the process of becoming vestigial, as it appears to be important for survival in fish, but it has no physiological effect on calcium homeostasis or bone metabolism in mammals (93). Similar views questioning the physiological function of calcitonin in mammals were subsequently published under the following titles: "Calcitonin guardian of the mammalian skeleton or is it just a fish story?" (150) and "Calcitonin: physiology or fantasy?" (60). Notwithstanding these skeptical views, the current consensus is that calcitonin has an important role in protection of the skeleton under conditions of calcium stress.

B. Genetically Modified Mice: Calcitonin, CGRP, and CTR

Laboratory investigations in vitro and in vivo established that calcitonin strongly inhibits bone resorption, and therefore, calcitonin-deficient mice were expected to have reduced bone mass in comparison to wild-type (WT) controls as a result of increased bone resorption. However, the bone phenotype of calcitonin-deficient mice proved to be entirely different than expected. In 2002, Hoff et al. generated a mouse strain deficient of the coding sequences of both calcitonin and aCGRP [Calca knockout (KO) or CT/ CGRP^{-/-}] (97). The mice had no developmental defects and had normal baseline-circulating calcium levels. Surprisingly, the Calca KO mice had significantly greater trabecular bone volume resulting from increased bone formation. Calca KO mice were less sensitive to OVX, maintaining their bone mass, whereas WT mice lost one-third of bone mass over 2 mo after OVX. The phenotype of the Calca KO mice suggested that the osteoblast is the primary target of calcitonin/CGRP in bone and that calcitonin/CGRP inhibit

bone formation. However, as the mice were deficient in both calcitonin and α CGRP, it was unclear which of the two peptides was responsible for the phenotype (97). Shortly after the original publication, the phenotype of α CGRP KO mice that had intact calcitonin was described by the same group (185). α CGRP KO mice had normal levels of circulating calcitonin, and unlike the Calca KO, they had reduced bone formation rate and developed osteopenia. Thus, the high bone mass phenotype of the Calca KO mice was a result of calcitonin deficiency, whereas α CGRP appears to increase bone formation. A subsequent long-term study of Calca KO mice showed that the bone phenotype developed with age. In animals 12 and 18 mo of age, the increased bone formation was accompanied by a parallel increase in bone resorption, with an overall accelerated bone turnover rate that produced hyperostotic lesions in 20% of the Calca KO mice. Therefore, the bone phenotype of the older KO mice suggested a physiological role for calcitonin as an inhibitor of bone remodeling (104). In contrast to the age-dependent progression of the phenotype in Calca KO mice, α CGRP KO showed a stable phenotype of osteopenia at all the ages analyzed (104). The analysis of the phenotype of the Calca KO and α CGRP KO mice is confounded by the presence of an intact Calcb gene that encodes BCGRP. However, a study of the bone phenotype of Calcb KO mice found no differences between these mice and control WT mice and led the authors to conclude that β CGRP does not have an important role in regulating bone remodeling (103).

Mechanical loading of the ulna can be used as a model for the study of bone adaptive response to mechanical loading. Sample et al. (183) used this model to test the hypothesis that CGRP, released from sensory fibers in bone, contributes to the adaptive response to load. Mechanically induced activation of periosteal mineralization was seen in WT mice but not in α CGRP KO mice. Interestingly, the phenotype of β CGRP KO mice was similar to that of the WT (183).

Transgenic mice overexpressing CGRP were produced and their bone phenotype determined (15). CGRP was introduced under the osteocalcin promoter to direct the overexpression of the peptide specifically to osteoblasts. Increased bone formation rate as well as higher trabecular bone density and volume were determined in transgenic mice overexpressing CGRP in comparison to WT littermates. These findings suggest a positive effect of CGRP in osteoblasts and are consistent with the study showing reduced bone formation rate and osteopenia in α CGRP KO mice (15, 185).

Further investigations of the physiological role of peptides of the calcitonin family in bone focused on the phenotype of receptor-deficient mice. As global deletion of CTR proved to be embryonic lethal, alternative KO animal models were developed to study the physiological effect of CTR deficiency. In hemizygote CTR KO mice $(Calcr^{+/-})$, the expres-

sion of CTR in osteoclasts was half of that found in the WT, and thus, these animals were used as a model of Calcr haploinsufficiency (58). Similar to the phenotype described for the Calca-deficient mice (97), $Calcr^{+/-}$ mice had high bone mass caused by increased bone formation without changes in bone resorption (58). These results provide further evidence for the physiological role of calcitonin as an inhibitor of bone formation and suggest that its activity is mediated via CTR. A second animal model was generated using the Cre/loxP system. Unlike the global CTR KO mice, this strain, which had an incomplete deletion of the Calcr gene (with some residual level of expression), was viable (62). In this animal model, only male mice had a small increase in bone formation rate, indicating that CTR plays a minor role in the physiological regulation of bone and calcium homeostasis in the basal state. Recently, using a modified technique to knockout the expression of CTR, Keller et al. managed to produce a new strain of viable global Calcr KO mice and used it to investigate the underlying mechanisms of CTR bone activity (119). Similar to the previous models of CTR deficiency, the Calcr KO mice had high bone mass because of increased bone formation, a phenotype that was then reproduced in mice that had the gene deletion restricted to the osteoclast lineage. The study found that the loss of CTR in osteoclasts increased the levels of sphingolipid transporter 2 (SPNS2), an exporter protein required for the secretion of sphingosine-1-phosphate (S1P), which is a potent inducer of bone formation. Therefore, according to this study, calcitonin binding to CTR on osteoclasts inhibits the expression of SPNS2, causing a drop in S1P secretion and inhibition of osteoblast activity caused by the reduced levels of S1P (119).

C. Calcitonin, CGRP, and CTR: Conclusions and Remaining Uncertainties

The studies of genetically modified animals described above contributed greatly to the understanding of the significance of calcitonin and its receptor to skeletal biology. However, although some conclusions can be drawn from the integration of these studies, other questions remain unanswered and clearly warrant further investigations. In general, both Calca KO mice and Calcr KO mice were consistently found to have high bone mass because of increased bone formation, although Calcr is not expressed in osteoblasts. This apparent contradiction had been resolved by the finding that the skeletal phenotype of an osteoclast-specific Calcr KO had increased bone formation, similar to that of the global Calcr KO, and that calcitonin increased the levels of osteocyte-secreted sclerostin. Therefore, calcitonin effect in osteoblasts is likely indirect and mediated via the two types of bone cells that express Clacr: osteoclasts and osteocytes. Among the questions that require further investigations is the skeletal phenotype of a mouse deficient of calcitonin. So far, the activity of calcitonin has been inferred from the skeletal phenotype of mice deficient of both calcitonin and α CGRP and mice deficient of α CGRP alone; however, a direct investigation of a calcitonin-deficient mouse is missing. Another important problem that had been overlooked is amylin signaling in the Calcr KO mouse. Because all the specific amylin receptors known at present include CTR, the analysis and interpretation of the Calcr KO phenotype should consider the possibility of detrimental effect on amylin signaling in this mouse model. It is interesting to note that although the pharmacological function of calcitonin in bone cells is to inhibit bone resorption, studies of genetically modified animals determined that calcitonin's physiological role is to inhibit bone formation. A similar discrepancy was identified in studies of the effect of PTH in bone, as PTH is a physiological stimulator of bone resorption that stimulates bone formation in pharmacological use (143).

D. The Role of Calcitonin and CTR in Situations of Calcium Stress

1. Hypercalcemia

The CTR KO mouse model described above (62) was used to study the role of CTR in maintaining calcium homeostasis in hypercalcemia. When hypercalcemia was induced by $1,25(OH)_2D_3$, the peak in serum total calcium was significantly greater in the CTR KO mice, suggesting that CTR is important in situations of calcium stress (62). The same animal model was used to investigate the contribution of osteoclast-expressed CTR to the protective effect against hypercalcemia (204). The study compared the response to hypercalcemia in three mice strains: WT, global CTR KO, and a Cre/loxP mouse model in which CTR was specifically deleted in osteoclasts. The two KO strains had similar responses to hypercalcemia, with peak serum calcium levels 18% higher than the WT because of increased bone resorption. These results suggest that calcitonin protects against hypercalcemia predominantly through inhibition of osteoclast activity.

2. Lactation

Transgenic animal models were used to study the role of calcitonin and CTR during lactation, a physiological state in which the maternal skeleton rapidly demineralizes to supply calcium to the milk. In the *Calca* KO model, deficient in both calcitonin and CGRP, spine bone mineral content dropped during 21 days of lactation by over 50%, whereas in the WT controls, the drop was only of 23.6% (222). Thirteen days after weaning, spine bone mineral content returned to baseline values in the WT mice, whereas in the *Calca* KO mice, it took 18 days to reach these values. To determine whether the effect in this double-KO mouse model was due to the deficiency in calcitonin or in CGRP, groups of mice were treated with either sCT or CGRP to replace the missing peptide. Injection of sCT normalized the

bone parameters, whereas CGRP was without effect, indicating that calcitonin is the peptide that plays an important role in calcium balance and preservation of the maternal skeleton during lactation and its recovery after weaning (222).

A physiological role for calcitonin in protecting the maternal skeleton from excessive resorption during lactation has also been suggested by a study of global CTR KO mice (38). At the end of lactation, measures of bone resorption by osteoclasts were similar between the CTR KO mice and the WT controls. However, gene expression analysis of mRNA extracted from whole tibiae found increased levels of a number of genes, including Catk and Mmp13, in the CTR KO mice. As increased osteocytic osteolysis had been previously demonstrated in mice during lactation, the authors hypothesized that calcitonin deficiency induced the expression of osteolytic genes in osteocytes. Histomorphometric analysis of the femurs at the end of lactation found that osteocyte lacunar area in CTR KO mice was larger than in WT, suggesting a role for calcitonin in inhibition of osteocytic osteolysis and protection of the maternal skeleton during lactation (38).

E. Genetically Modified Mice: Amylin

The bone phenotype of amylin-deficient mice was investigated by Dacquin et al. (58). Amylin deficiency had no effect on the regulation of food intake, body weight, or glucose metabolism in this experimental model. At the age of 24 wk, both male and female amylin KO mice were osteoporotic with vertebral bone volume over tissue volume (BV/TV) ~50% lower than that of WT and trabecular and cortical bone thickness reduced. The number of osteoblasts was similar between amylin KO and WT mice, and dynamic histomorphometry using calcein double labeling demonstrated a similar bone formation rate. The amylin KO mice had an increased number of osteoclasts and an increase in degradation products of collagen in the urine, suggesting that the osteoporotic phenotype was a result of accelerated bone resorption (58). The study also investigated the role of CTR in mediating the bone effects of amylin. Unlike amylin KO mice, hemizygous $Calcr^{+/-}$ animals had high bone mass, and compound hemizygous $Calcr^{+/-}$ Amylin^{+/-} mice displayed a combination of abnormalities identified in each of the individual hemizygous KOs. Therefore, CTR is unlikely to be mediating the amylin bone effects. The receptor that mediates amylin's effects in bone tissue is yet to be identified (58). Further studies of the same strain of amylin KO mice compared the bone phenotype between young and adult male and female mice (61). Amylin KO males had increased trabecular thickness at 4 and 6 wk of age and increased femoral length at 26 wk, whereas female mice were no different from the WT controls (61).

In summary, the effects of amylin on osteoclast and bone resorption were generally similar in the different experi-

mental systems; amylin deficiency produced an osteoporotic phenotype because of increased bone resorption, and amylin was shown to inhibit osteoclast differentiation and activity in vitro and reduce bone resorption when administered into WT animals. On the other hand, although positive effects of amylin on osteoblasts and bone formation were demonstrated in vitro and with administration of the peptide into animals, amylin deficiency did not modify indices of bone formation. Although the reason for this discrepancy is not clear, it could perhaps reflect the difference between the physiological effect of amylin inferred from the skeletal phenotype of the amylin KO mice and the high, pharmacological concentrations of amylin used in vitro. In addition, it is important to note that the bone phenotype of the amylin KO mouse is age- and gender-dependent, and it is further complicated by the observation that the amylin activity in bone is not mediated by CTR, although currently there is no known amylin receptor that does not include CTR. Thus, a number of unresolved questions remain, and a better understanding of the physiological and pharmacological effects of amylin in bone would require further investigations in existing experimental models and the development of novel ones.

F. Genetically Modified Mice: Adrenomedullin, CRLR, and Ramp1–3

Development of mice strains deficient of adrenomedullin, *Calcrl*, and *Ramp1*, *Ramp2*, and *Ramp3* produced interesting and some unexpected results (108, 189). Genetic deficiency in either adrenomedullin, *Calcrl*, or *Ramp2* (27, 57, 74) is embryonic lethal at midgestation because of cardiovascular defects and a hypoproliferative lymphatic vasculature, whereas *Ramp1* KO and *Ramp3* KO are viable (57, 131, 203). These findings demonstrate that despite the structural similarity among the three RAMPS, RAMP1 and 3 are unable to compensate for the loss of RAMP2.

The skeletal effects of adrenomedullin deficiency were determined in a conditional KO mouse model that was produced using a doxycycline-dependent Cre/Lox excision of the adrenomedullin gene (144). For unknown reasons, rather than being entirely absent, the levels of circulating adrenomedullin were only reduced by ~50% following doxycycline treatment. Analysis of femora by histology and micro-CT determined increased bone mass and density in adrenomedullin-deficient mice in comparison to WT controls. These findings were unexpected, as previously in vitro and local injection experiments had shown a direct positive effect of adrenomedullin on bone formation (46, 159, 205). The authors suggested that the increased bone mass and density in the adrenomedullin-deficient mice were caused by an indirect mechanism. Candidates that could be involved in such indirect effects are CGRP and ghrelin, two peptides that promote bone formation and had increased circulating levels in the adrenomedullin-deficient mice. The

results from the mouse model produced in this study are confounded by the fact that the KO mice gained weight rapidly after exposure to doxycycline (144). In the second part of the study, the authors investigated the effect of an inhibitor of adrenomedullin (the small molecule 16311) on bone loss in OVX mice. The molecule had no effect on BV/TV in femora of sham controls but protected the OVX mice from reduction of BV/TV, suggesting a role for adrenomedullin in mediating the OVX-induced bone loss (144).

Hemizygous $Ramp2^{+/-}$ mice are viable, although they have severe reproductive defects, fetal growth restriction, enlarged pituitary glands, and a number of additional abnormalities (107). Haploinsufficiency for RAMP2 caused significant skeletal defects, including delayed development and reduced mineral content and mineral density in femora of 18-wk-old females in comparison to WT controls. Tibiae of $Ramp2^{+/-}$ mice were significantly longer than those of WT controls. Although $Ramp2^{+/-}$ mice had an interesting skeletal phenotype, it is difficult to delineate the molecular mechanisms responsible for the abnormalities because of the complex endocrine phenotype of these mice. The skeletal phenotype of Ramp1 KO and Ramp3 KO has not been described in detail yet. A study that may have relevance to bone, and specifically to fracture healing, examined wound healing in Ramp1 KO mice (128). The study found that wound healing and wound-induced angiogenesis were significantly suppressed in Ramp1 KO mice in comparison to WT controls. Wound healing was also delayed in chimeric mice produced by transplantation of bone marrow from Ramp1 KO into WT mice, indicating a crucial role for hematopoietic cells recruited into the wound.

IV. CLINICAL USE OF CALCITONIN PEPTIDES FOR BONE DISEASES

The discovery of calcitonin as a potent inhibitor of bone resorption by osteoclasts identified it as a potential therapeutic for conditions of excessive resorption. Although neither the absence nor pathologically high levels of endogenous calcitonin appear to affect bone mass, pharmacological use of calcitonin was considered a promising option for the inhibition of bone turnover, preservation of bone mass, and fracture prevention (105, 223).

A. sCT Administration

Among the various calcitonin preparations that have been used in clinical practice, sCT is the most widely used, as it is ~50 times more potent than hCT. sCT has been shown to be safe, and although a large proportion of patients treated with sCT develop antibodies against it, it is generally believed that these have minimal clinical impact (85). sCT was first introduced to the market in 1974 and was subsequently approved by the United States Food and Drug Administra-

tion for use in the treatment of postmenopausal osteoporosis, hypercalcemia, and Paget's disease (33, 165). sCT was initially commercially available as an injectable formulation for intramuscular or subcutaneous use. Calcitonin injections had benign, but uncomfortable, side effects, and compliance to long-term parenteral therapy was low (174). The development of calcitonin as a nasal spray provided a much more convenient option for patients, and nasal spray has been used in the clinic and in many studies of calcitonin in humans. The main disadvantage of the nasal spray formulation is its low bioavailability, estimated to be only 10%–25% in comparison to parenteral injections (112). The latest stage in the development of sCT as a drug is an oral formulation, which promises to improve both bioavailability and compliance (16). In the oral formulation, sCT is linked to a drug delivery agent that interacts with the peptide weakly and noncovalently, increasing the ability of sCT to cross the gastrointestinal epithelium and providing a partial protection from degradation by enzymes in the upper digestive tract (113).

B. sCT for Osteoporosis and Fracture Prevention

sCT was one of the first drugs used as an antiresorptive therapy for osteoporosis. A number of small studies of intranasal or subcutaneous use of sCT showed increases in bone density, reduced number of vertebral fractures, and an association with reduction of the risk of hip fracture (26, 33, 110, 139), whereas others found no bone effects (12). The Prevent Recurrence of Osteoporotic Fractures study was a large, multicenter, prospective, 5-yr, randomized, placebo-controlled study that determined the effect of sCT nasal spray on the risk of new vertebral fractures in postmenopausal women with osteoporosis (35). The study found that a daily dose of 200 IU significantly reduced the risk of new vertebral fractures in the study population. However, this conclusion was later challenged, mainly because of the high dropout rate (59%) that compromised the analysis and because only the 200 IU/day dose appeared to be effective, but a higher dose was ineffective (56). A later study used a combination of noninvasive MRI technology and iliac crest bone biopsies to determine the effects of intranasal sCT on parameters of trabecular microarchitecture at multiple skeletal sites (34). This 2-yr, placebo-controlled trial of 91 postmenopausal osteoporotic women suggested a therapeutic benefit of sCT in maintaining trabecular microarchitecture at some skeletal sites, but not others. In a more recent large, randomized, double-blind, placebo-controlled study, oral sCT was tested for the treatment of postmenopausal osteoporosis in a total of 4,665 women over 36 mo (91). sCT induced modest increases in vertebral, femur, neck, and hip bone mineral density in the treatment group but had no significant effects on the proportion of patients with new vertebral, hip, or nonvertebral fractures. Because of this lack of efficacy in preventing frac-

tures, the development of the orally formulated calcitonin was terminated (91). sCT was initially approved for the treatment of postmenopausal osteoporosis at a time when the only other pharmaceutical treatment available was hormone therapy. At present, sCT use declined, and it is no longer considered an appropriate treatment option for osteoporosis for two main reasons: 1) a number of studies suggested an association between sCT use and cancer incidence (165). Although the evidence is not strong, it led to a 2012 ruling by the European Medicines Agency that "the benefits of calcitonin-containing medicines did not outweigh their risks in the treatment of osteoporosis and that they should no longer be used for this condition." There is some controversy around this ruling, as it is regarded as being based on unconvincing evidence (71). 2) The development of bisphosphonates and other effective drugs provide much better options for osteoporosis treatment (165).

C. Paget's Disease of Bone

Calcitonin was the first inhibitor of osteoclasts used for treatment of Paget's disease of bone, a common condition characterized by localized increases in bone turnover (129).

A number of observational studies, carried out in the 1970– 1980s, established the efficacy and safety of sCT in Paget's disease. sCT was effective in inhibiting bone turnover, providing pain relief, and improving bone structure, healing osteolytic lesions and supporting the formation of normal lamellar bone (33). However, calcitonin produced only partial and short-lived improvements, as resistance appeared to develop following its repeated use. Similar to osteoporosis treatment, bisphosphonates offer better options for management of Paget's disease. The European Medicines Agency recommends that sCT be used for the shortest possible time, at the minimum effective dose in Paget's disease, in patients who do not respond to alternative treatments.

D. Osteoarthritis

Another potential indication for sCT is osteoarthritis (OA), a degenerative joint disease that involves cartilage, subchondral bone, and many of the surrounding tissues (136). A number of mechanisms have been suggested for the beneficial effect of sCT in OA, including the inhibition of subchondral bone turnover and subsequent periarticular bone degradation, a direct activity in chondrocytes to induce the

Gene	Protein	Main Activity in Bone Cells in Vitro	References	Skeletal Phenotype of Knockout Mice	References
<i>Calca</i> Calc	Calcitonin	Osteoclasts: inhibition of activity and differentiation. Osteoblasts: inconsistent findings.	(3, 29, 30, 44, 82, 83, 190) (45, 68, 209)	<i>Calca</i> KO (CT/CGRP ^{-/-}): High bone mass, increased bone formation rate, and trabecular bone volume, no change in indices of bone resorption.	(97)
		Osteocytes: inhibition of apoptosis, increased expression of the Wnt- signaling inhibitor sclerostin.	(80, 167)	In older mice (12–18 mo), parallel increase in resorption, overall accelerated bone remodeling rate.	(104)
Calca	αCGRP	Osteoclasts: inhibition of activity and differentiation. Osteoblasts: stimulation of proliferation and differentiation.	(2, 44, 82, 213) (49, 102, 133, 154, 201, 210, 213, 232)	Reduced bone formation rate, osteopenia.	(185)
Calcb	βCGRP	No effect on osteogenesis in bone marrow cultures.	(96)	No skeletal phenotype.	(103, 183)
lapp	Amylin	Osteoclasts: inhibition of activity and differentiation. Osteoblasts: stimulation of proliferation and differentiation.	(4, 44, 51, 58, 82, 166, 197) (45, 49, 51, 52, 210, 211)	Osteoporosis caused by accelerated bone resorption. No effect on bone formation rate.	(58)
Adm	Adrenomedullin	Osteoclasts: no effect. Osteoblasts: stimulation of proliferation and differentiation.	(82) (46, 52, 87, 130, 159)	Embryonic lethal Partial-KO model: increased bone mass.	(27) (144)
Adm2	Intermedin	Osteoclasts: inhibition of activity and differentiation.	(82)	NA	
		Osteoblasts: no effect on proliferation or differentiation.	(175)		

Table I. The main effects of members of the calcitonin family on bone cells and the skeletal phenotypes of peptide-deficient mice

CGRP, calcitonin gene-related peptide; KO, knockout; NA, not available.

production and secretion of cartilage extracellular proteins, and providing an analgesic effect (33).

A direct effect of calcitonin on chondrocytes has been demonstrated in a number of studies. In a model of degradation in bovine articular cartilage explants, calcitonin induced cAMP production in chondrocytes and inhibited degradation of type II collagen and the increase in matrix metalloproteinase activity (193). In vivo, calcitonin was shown to attenuate type II collagen degradation and to reduce cartilage erosion, extracellular matrix degradation, and subchondral bone damage in experimental animal models of arthritis (32, 140, 192). In addition, OA induced by destabilization of the medial meniscus produced a fivefold increase in cartilage erosion index, whereas transgenic mice overexpressing sCT had ~60% lower erosion index than WT mice (191). A direct effect of calcitonin in chondrocytes requires the expression of CTR in these cells. However, although some studies demonstrate the expression of CTR in chondrocytes (193), others find no expression (135).

In humans, oral sCT reduced circulating carboxy-terminal collagen cross-links II and improved functional disability in patients, albeit in a small test group, selected for active disease (14, 141). However, recently published results of two phase III–randomized, double-blind, placebo-controlled trials that evaluated the effect of oral sCT on symptomatic knee OA found no significant impact on joint space narrowing in the treatment groups (111).

V. SUMMARY

The calcitonin-family peptides and their receptors form a complex network of interacting molecules. Although the

primary physiological roles of the peptides are diverse and they affect a wide range of cell types and tissues, the skeletal phenotype of mice deficient of either calcitonin, amylin, α CGRP, or CTR indicates an important role for the calcitonin family in skeletal physiology. The main effects of the calcitonin-family peptides and the characteristics of the skeletal phenotype of mice deficient of the peptides and receptors are summarized in **TABLES 1** and **2**.

A number of questions regarding the bone activities of the peptides are yet to be resolved. One major challenge is the discrepancy between the pharmacological effects observed in bone cells in vitro and in animals injected with the peptides and the physiological roles inferred from the skeletal phenotype of genetically modified mice and clinical observations in humans. A large part of these discrepancies are undoubtedly due to the inherent differences in physiology and genetics between species. To date, the fundamental target of calcitonin-family peptides in bone appears to be the osteoclast. It is not surprising to see discrepancies between animals and humans, as well as between animals of different ages, as the activity of osteoclasts, along with the rate of bone remodeling, are vastly different. There are also differences in pharmacokinetics between species such that pharmacological responses in mice to certain drug concentrations cannot be easily related to responses in humans. Similarly, response to pharmacological interventions changes depending on age, health, and a number of environmental factors that cannot be translated from the laboratory. Despite this, major progress achieved in recent years, including the discovery of the osteocyte as a target cell for calcitonin and the development of animal models with tissue-specific gene KO, has helped explain some of the apparent contradictory observations. Additional answers will undoubtedly

Gene	Protein	Skeletal Phenotype of Knockout Mice	References
Calcr	Calcitonin receptor (CTR)	 Calcr^{+/-} (hemizygous): high bone mass, increased bone formation. 	(58)
		2) Incomplete KO model: increased bone formation.	(62)
		Global Calcr KO: high bone mass, increased bone formation.	(119)
		 Osteoclast-specific Calcr KO: high bone mass, increased bone formation. 	(119)
Calcrl	Calcitonin receptor–like receptor (CRLR)	Embryonic lethal	(57, 74)
Ramp1	Receptor activity-modifying protein (RAMP)1	NA	
Ramp2	RAMP2	Embryonic lethal	(74)
		Ramp2 ^{+/-} (hemizygous): delayed bone development, reduced mineral content, and mineral density.	(107)
Ramp3	RAMP3	NA	

KO, knockout; NA, not available.

Physiol Rev • VOL 99 • JANUARY 2019 • www.prv.org

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be provided in the future with further development of genetically modified animals and thorough investigations of their phenotype at different ages, in states of calcium stress, and in models of accelerated bone turnover.

The use of calcitonin for the management of conditions of accelerated bone turnover was expected to increase with the development of nasal and oral formulations of sCT. However, with the availability of more effective drugs for longterm inhibition of bone resorption, sCT use has declined, and there are no current trials for its use for skeletal disorders. Despite this, there is still a great interest in the calcitonin family of peptides and their skeletal effects, with current studies designed to clarify the activities and physiological roles of this complex and intriguing family of peptides.

ACKNOWLEDGMENTS

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GRANTS

The authors receive research funding from the Health Research Council of New Zealand.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

REFERENCES

- Ah Kioon MD, Asensio C, Ea HK, Velard F, Uzan B, Rullé S, Bazille C, Marty C, Falgarone G, Nguyen C, Collet C, Launay JM, Cohen-Solal M, Lioté F. Adrenomedullin(22-52) combats inflammation and prevents systemic bone loss in murine collageninduced arthritis. *Arthritis Rheum* 64: 1069–1081, 2012. doi:10.1002/art.33426.
- Akopian A, Demulder A, Ouriaghli F, Corazza F, Fondu P, Bergmann P. Effects of CGRP on human osteoclast-like cell formation: a possible connection with the bone loss in neurological disorders? *Peptides* 21: 559–564, 2000. doi:10.1016/S0196-9781(00)00185-6.
- Alam AS, Bax CM, Shankar VS, Bax BE, Bevis PJ, Huang CL, Moonga BS, Pazianas M, Zaidi M. Further studies on the mode of action of calcitonin on isolated rat osteoclasts: pharmacological evidence for a second site mediating intracellular Ca2+ mobilization and cell retraction. J Endocrinol 136: 7–15, 1993. doi:10.1677/joe.0.1360007.
- Alam AS, Moonga BS, Bevis PJ, Huang CL, Zaidi M. Amylin inhibits bone resorption by a direct effect on the motility of rat osteoclasts. *Exp Physiol* 78: 183–196, 1993. doi:10.1113/expphysiol.1993.sp003679.
- Alam AS, Moonga BS, Bevis PJ, Huang CL, Zaidi M. Selective antagonism of calcitonininduced osteoclastic quiescence (Q effect) by human calcitonin gene-related peptide-(Val8Phe37). Biochem Biophys Res Commun 179: 134–139, 1991. doi:10.1016/0006-291×(91)91345-D.
- Alam T, Chen L, Ogawa A, Leffert JD, Unger RH, Luskey KL. Coordinate regulation of amylin and insulin expression in response to hypoglycemia and fasting. *Diabetes* 41: 508–514, 1992. doi:10.2337/diab.41.4.508.
- 7. Albrandt K, Brady EM, Moore CX, Mull E, Sierzega ME, Beaumont K. Molecular cloning and functional expression of a third isoform of the human calcitonin receptor

and partial characterization of the calcitonin receptor gene. *Endocrinology* 136: 5377–5384, 1995. doi:10.1210/endo.136.12.7588285.

- Amara SG, Arriza JL, Leff SE, Swanson LW, Evans RM, Rosenfeld MG. Expression in brain of a messenger RNA encoding a novel neuropeptide homologous to calcitonin gene-related peptide. *Science* 229: 1094–1097, 1985. doi:10.1126/science.2994212.
- Andreassen KV, Hjuler ST, Furness SG, Sexton PM, Christopoulos A, Nosjean O, Karsdal MA, Henriksen K. Prolonged calcitonin receptor signaling by salmon, but not human calcitonin, reveals ligand bias. *PLoS One* 9: e92042, 2014. doi:10.1371/journal. pone.0092042.
- Appelman-Dijkstra NM, Papapoulos SE. From disease to treatment: from rare skeletal disorders to treatments for osteoporosis. *Endocrine* 52: 414–426, 2016. doi:10. 1007/s12020-016-0888-7.
- Armour SL, Foord S, Kenakin T, Chen WJ. Pharmacological characterization of receptor-activity-modifying proteins (RAMPs) and the human calcitonin receptor. J Pharmacol Toxicol Methods 42: 217–224, 1999. doi:10.1016/S1056-8719(00)00074-5.
- Arnala I, Saastamoinen J, Alhava EM. Salmon calcitonin in the prevention of bone loss at perimenopause. Bone 18: 629–632, 1996. doi:10.1016/8756-3282(96)00084-1.
- Aurbach GD. Isolation of parathyroid hormone after extraction with phenol. J Biol Chem 234: 3179–3181, 1959.
- 14. Bagger YZ, Tankó LB, Alexandersen P, Karsdal MA, Olson M, Mindeholm L, Azria M, Christiansen C. Oral salmon calcitonin induced suppression of urinary collagen type II degradation in postmenopausal women: a new potential treatment of osteoarthritis. *Bone* 37: 425–430, 2005. doi:10.1016/j.bone.2005.04.032.
- Ballica R, Valentijn K, Khachatryan A, Guerder S, Kapadia S, Gundberg C, Gilligan J, Flavell RA, Vignery A. Targeted expression of calcitonin gene-related peptide to osteoblasts increases bone density in mice. *J Bone Miner Res* 14: 1067–1074, 1999. doi:10.1359/jbmr.1999.14.7.1067.
- Bandeira L, Lewiecki EM, Bilezikian JP. Pharmacodynamics and pharmacokinetics of oral salmon calcitonin in the treatment of osteoporosis. *Expert Opin Drug Metab Toxicol* 12: 681–689, 2016. doi:10.1080/17425255.2016.1175436.
- Barwell J, Gingell JJ, Watkins HA, Archbold JK, Poyner DR, Hay DL. Calcitonin and calcitonin receptor-like receptors: common themes with family B GPCRs? Br J Pharmacol 166: 51–65, 2012. doi:10.1111/j.1476-5381.2011.01525.x.
- Bjurholm A, Kreicbergs A, Schultzberg M, Lerner UH. Neuroendocrine regulation of cyclic AMP formation in osteoblastic cell lines (UMR-106-01, ROS 17/2.8, MC3T3-E1, and Saos-2) and primary bone cells. *J Bone Miner Res* 7: 1011–1019, 1992. doi:10. 1002/jbmr.5650070903.
- Bomberger JM, Parameswaran N, Hall CS, Aiyar N, Spielman WS. Novel function for receptor activity-modifying proteins (RAMPs) in post-endocytic receptor trafficking. J Biol Chem 280: 9297–9307, 2005. doi:10.1074/jbc.M413786200.
- Booe JM, Walker CS, Barwell J, Kuteyi G, Simms J, Jamaluddin MA, Warner ML, Bill RM, Harris PW, Brimble MA, Poyner DR, Hay DL, Pioszak AA. Structural Basis for Receptor Activity-Modifying Protein-Dependent Selective Peptide Recognition by a G Protein-Coupled Receptor. *Mol Cell* 58: 1040–1052, 2015. doi:10.1016/j.molcel. 2015.04.018.
- Bower RL, Hay DL. Amylin structure-function relationships and receptor pharmacology: implications for amylin mimetic drug development. Br J Pharmacol 173: 1883– 1898, 2016. doi:10.1111/bph.13496.
- Breimer LH, MacIntyre I, Zaidi M. Peptides from the calcitonin genes: molecular genetics, structure and function. *Biochem J* 255: 377–390, 1988. doi:10.1042/ bj2550377.
- Burns DM, Stehno-Bittel L, Kawase T. Calcitonin gene-related peptide elevates calcium and polarizes membrane potential in MG-63 cells by both cAMP-independent and -dependent mechanisms. *Am J Physiol Cell Physiol* 287: C457–C467, 2004. doi:10. 1152/ajpcell.00274.2003.
- Butler PC, Chou J, Carter WB, Wang YN, Bu BH, Chang D, Chang JK, Rizza RA. Effects of meal ingestion on plasma amylin concentration in NIDDM and nondiabetic humans. *Diabetes* 39: 752–756, 1990. doi:10.2337/diab.39.6.752.
- Cao P, Abedini A, Wang H, Tu LH, Zhang X, Schmidt AM, Raleigh DP. Islet amyloid polypeptide toxicity and membrane interactions. *Proc Natl Acad Sci USA* 110: 19279– 19284, 2013. doi:10.1073/pnas.1305517110.

- Cardona JM, Pastor E. Calcitonin versus etidronate for the treatment of postmenopausal osteoporosis: a meta-analysis of published clinical trials. Osteoporos Int 7: 165– 174, 1997. doi:10.1007/BF01622285.
- Caron KM, Smithies O. Extreme hydrops fetalis and cardiovascular abnormalities in mice lacking a functional Adrenomedullin gene. *Proc Natl Acad Sci USA* 98: 615–619, 2001. doi:10.1073/pnas.98.2.615.
- Caron KM, Smithies O. Multiple roles of adrenomedullin revealed by animal models. Microsc Res Tech 57: 55–59, 2002. doi:10.1002/jemt.10046.
- Chambers TJ, Athanasou NA, Fuller K. Effect of parathyroid hormone and calcitonin on the cytoplasmic spreading of isolated osteoclasts. J Endocrinol 102: 281–286, 1984. doi:10.1677/joe.0.1020281.
- Chambers TJ, Magnus CJ. Calcitonin alters behaviour of isolated osteoclasts. J Pathol 136: 27–39, 1982. doi:10.1002/path.1711360104.
- Chang CP, Pearse RV II, O'Connell S, Rosenfeld MG. Identification of a seven transmembrane helix receptor for corticotropin-releasing factor and sauvagine in mammalian brain. *Neuron* 11: 1187–1195, 1993. doi:10.1016/0896-6273(93)90230-O.
- Cheng T, Zhang L, Fu X, Wang W, Xu H, Song H, Zhang Y. The potential protective effects of calcitonin involved in coordinating chondrocyte response, extracellular matrix, and subchondral trabecular bone in experimental osteoarthritis. *Connect Tissue Res* 54: 139–146, 2013. doi:10.3109/03008207.2012.760549.
- Chesnut CH III, Azria M, Silverman S, Engelhardt M, Olson M, Mindeholm L. Salmon calcitonin: a review of current and future therapeutic indications. Osteoporos Int 19: 479–491, 2008. doi:10.1007/s00198-007-0490-1.
- Chesnut CH III, Majumdar S, Newitt DC, Shields A, Van Pelt J, Laschansky E, Azria M, Kriegman A, Olson M, Eriksen EF, Mindeholm L. Effects of salmon calcitonin on trabecular microarchitecture as determined by magnetic resonance imaging: results from the QUEST study. J Bone Miner Res 20: 1548–1561, 2005. doi:10.1359/JBMR. 050411.
- Chesnut CH III, Silverman S, Andriano K, Genant H, Gimona A, Harris S, Kiel D, LeBoff M, Maricic M, Miller P, Moniz C, Peacock M, Richardson P, Watts N, Baylink D; PROOF Study Group. A randomized trial of nasal spray salmon calcitonin in postmenopausal women with established osteoporosis: the prevent recurrence of osteoporotic fractures study. *Am J Med* 109: 267–276, 2000. doi:10.1016/S0002-9343(00)00490-3.
- Chia LY, Walsh NC, Martin TJ, Sims NA. Isolation and gene expression of haematopoietic-cell-free preparations of highly purified murine osteocytes. *Bone* 72: 34–42, 2015. doi:10.1016/j.bone.2014.11.005.
- Christopoulos G, Perry KJ, Morfis M, Tilakaratne N, Gao Y, Fraser NJ, Main MJ, Foord SM, Sexton PM. Multiple amylin receptors arise from receptor activity-modifying protein interaction with the calcitonin receptor gene product. *Mol Pharmacol* 56: 235–242, 1999. doi:10.1124/mol.56.1.235.
- Clarke MV, Russell PK, Findlay DM, Sastra S, Anderson PH, Skinner JP, Atkins GJ, Zajac JD, Davey RA. A Role for the Calcitonin Receptor to Limit Bone Loss During Lactation in Female Mice by Inhibiting Osteocytic Osteolysis. *Endocrinology* 156: 3203–3214, 2015. doi:10.1210/en.2015-1345.
- Collip JB. The extraction of a parathyroid hormone which will prevent or control parathyroid tetany and which regulates the level of blood calcium. J Biol Chem 63: 395–438, 1925.
- Cooper GJ, Willis AC, Clark A, Turner RC, Sim RB, Reid KB. Purification and characterization of a peptide from amyloid-rich pancreases of type 2 diabetic patients. Proc Natl Acad Sci USA 84: 8628–8632, 1987. doi:10.1073/pnas.84.23.8628.
- Copp DH, Cameron EC, Cheney BA, Davidson AG, Henze KG. Evidence for calcitonin–a new hormone from the parathyroid that lowers blood calcium. *Endocrinology* 70: 638–649, 1962. doi:10.1210/endo-70-5-638.
- Copp DH, Cheney B. Calcitonin-a hormone from the parathyroid which lowers the calcium-level of the blood. Nature 193: 381–382, 1962. doi:10.1038/193381a0.
- Cornish J, Callon KE, Bava U, Coy DH, Mulvey TB, Murray MA, Cooper GJ, Cooper GJ, Reid IR. Systemic administration of adrenomedullin(27-52) increases bone volume and strength in male mice. *J Endocrinol* 170: 251–257, 2001. doi:10.1677/joe.0. 1700251.

- Cornish J, Callon KE, Bava U, Kamona SA, Cooper GJ, Reid IR. Effects of calcitonin, amylin, and calcitonin gene-related peptide on osteoclast development. *Bone* 29: 162–168, 2001. doi:10.1016/S8756-3282(01)00494-X.
- Cornish J, Callon KE, Cooper GJ, Reid IR. Amylin stimulates osteoblast proliferation and increases mineralized bone volume in adult mice. *Biochem Biophys Res Commun* 207: 133–139, 1995. doi:10.1006/bbrc.1995.1163.
- Cornish J, Callon KE, Coy DH, Jiang NY, Xiao L, Cooper GJ, Reid IR. Adrenomedullin is a potent stimulator of osteoblastic activity in vitro and in vivo. *Am J Physiol* 273: E1113–E1120, 1997.
- Cornish J, Callon KE, Gasser JA, Bava U, Gardiner EM, Coy DH, Cooper GJ, Reid IR. Systemic administration of a novel octapeptide, amylin-(1–8), increases bone volume in male mice. *Am J Physiol Endocrinol Metab* 279: E730–E735, 2000. doi:10.1152/ ajpendo.2000.279.4.E730.
- Cornish J, Callon KE, King AR, Cooper GJ, Reid IR. Systemic administration of amylin increases bone mass, linear growth, and adiposity in adult male mice. *Am J Physiol* 275: E694–E699, 1998.
- Cornish J, Callon KE, Lin CQ, Xiao CL, Gamble GD, Cooper GJ, Reid IR. Comparison of the effects of calcitonin gene-related peptide and amylin on osteoblasts. J Bone Miner Res 14: 1302–1309, 1999. doi:10.1359/jbmr.1999.14.8.1302.
- Cornish J, Callon KE, Lin CQ, Xiao CL, Mulvey TB, Cooper GJ, Reid IR. Trifluoroacetate, a contaminant in purified proteins, inhibits proliferation of osteoblasts and chondrocytes. *Am J Physiol* 277: E779–E783, 1999.
- Cornish J, Callon KE, Lin CQ, Xiao CL, Mulvey TB, Coy DH, Cooper GJ, Reid IR. Dissociation of the effects of amylin on osteoblast proliferation and bone resorption. *Am J Physiol* 274: E827–E833, 1998.
- Cornish J, Grey A, Callon KE, Naot D, Hill BL, Lin CQ, Balchin LM, Reid IR. Shared pathways of osteoblast mitogenesis induced by amylin, adrenomedullin, and IGF-1. *Biochem Biophys Res Commun* 318: 240–246, 2004. doi:10.1016/j.bbrc.2004.04.020.
- Cornish J, Naot D. Amylin and adrenomedullin: novel regulators of bone growth. Curr Pharm Des 8: 2009–2021, 2002. doi:10.2174/1381612023393341.
- Cornish J, Reid IR. Skeletal effects of amylin and related peptides. *Endocrinologist* 9: 183–189, 1999. doi:10.1097/00019616-199905000-00004.
- 55. Cosman F, Crittenden DB, Adachi JD, Binkley N, Czerwinski E, Ferrari S, Hofbauer LC, Lau E, Lewiecki EM, Miyauchi A, Zerbini CA, Milmont CE, Chen L, Maddox J, Meisner PD, Libanati C, Grauer A. Romosozumab Treatment in Postmenopausal Women with Osteoporosis. N Engl J Med 375: 1532–1543, 2016. doi:10.1056/ NEJMoa1607948.
- Cummings SR, Chapurlat RD. What PROOF proves about calcitonin and clinical trials. *Am J Med* 109: 330–331, 2000. doi:10.1016/S0002-9343(00)00539-8.
- Dackor R, Fritz-Six K, Smithies O, Caron K. Receptor activity-modifying proteins 2 and 3 have distinct physiological functions from embryogenesis to old age. J Biol Chem 282: 18094–18099, 2007. doi:10.1074/jbc.M703544200.
- Dacquin R, Davey RA, Laplace C, Levasseur R, Morris HA, Goldring SR, Gebre-Medhin S, Galson DL, Zajac JD, Karsenty G. Amylin inhibits bone resorption while the calcitonin receptor controls bone formation in vivo. J Cell Biol 164: 509–514, 2004. doi:10.1083/jcb.200312135.
- Datta HK, Zaidi M, Wimalawansa SJ, Ghatei MA, Beacham JL, Bloom SR, MacIntyre I. In vivo and in vitro effects of amylin and amylin-amide on calcium metabolism in the rat and rabbit. *Biochem Biophys Res Commun* 162: 876–881, 1989. doi:10.1016/0006-291×(89)92391-7.
- Davey RA, Findlay DM. Calcitonin: physiology or fantasy? J Bone Miner Res 28: 973– 979, 2013. doi:10.1002/jbmr.1869.
- Davey RA, Moore AJ, Chiu MW, Notini AJ, Morris HA, Zajac JD. Effects of amylin deficiency on trabecular bone in young mice are sex-dependent. *Calcif Tissue Int* 78: 398–403, 2006. doi:10.1007/s00223-005-0286-2.
- Davey RA, Turner AG, McManus JF, Chiu WS, Tjahyono F, Moore AJ, Atkins GJ, Anderson PH, Ma C, Glatt V, MacLean HE, Vincent C, Bouxsein M, Morris HA, Findlay DM, Zajac JD. Calcitonin receptor plays a physiological role to protect against hypercalcemia in mice. *J Bone Miner Res* 23: 1182–1193, 2008. doi:10.1359/jbmr. 080310.

800

BONE ACTIVITY OF CALCITONIN-FAMILY PEPTIDES

- 63. Davies J. Procalcitonin. J Clin Pathol 68: 675–679, 2015. doi:10.1136/jclinpath-2014-202807.
- Delgado-Calle J, Sato AY, Bellido T. Role and mechanism of action of sclerostin in bone. Bone 96: 29–37, 2017. doi:10.1016/j.bone.2016.10.007.
- Drissi H, Lieberherr M, Hott M, Marie PJ, Lasmoles F. Calcitonin gene-related peptide (CGRP) increases intracellular free Ca2+ concentrations but not cyclic AMP formation in CGRP receptor-positive osteosarcoma cells (OHS-4). *Cytokine* 11: 200–207, 1999. doi:10.1006/cyto.1998.0415.
- Ellegaard M, Thorkildsen C, Petersen S, Petersen JS, Jørgensen NR, Just R, Schwarz P, Ramirez MT, Stahlhut M. Amylin(1-8) is devoid of anabolic activity in bone. *Calcif Tissue Int* 86: 249–260, 2010. doi:10.1007/s00223-010-9338-3.
- Eller LK, Ainslie PN, Poulin MJ, Reimer RA. Differential responses of circulating amylin to high-fat vs. high-carbohydrate meal in healthy men. *Clin Endocrinol (Oxf)* 68: 890– 897, 2008. doi:10.1111/j.1365-2265.2007.03129.x.
- Farley JR, Tarbaux NM, Hall SL, Linkhart TA, Baylink DJ. The anti-bone-resorptive agent calcitonin also acts in vitro to directly increase bone formation and bone cell proliferation. *Endocrinology* 123: 159–167, 1988. doi:10.1210/endo-123-1-159.
- Favus M, Goltzman D. Regulation of calcium and magnesium. In: Primer on the Metabolic Bone Diseases and Disorders of Mineral Metabolism, edited by Rosen CJ. United States: Wiley-Blackwell, 2013. doi:10.1002/9781118453926.ch22.
- Findlay DM, Sexton PM. Calcitonin. Growth Factors 22: 217–224, 2004. doi:10.1080/ 08977190410001728033.
- Findlay DM, Sexton PM, Martin JE. Calcitonin. Oxford, UK: Elsevier Health Sciences, 2015.
- Flühmann B, Muff R, Hunziker W, Fischer JA, Born W. A human orphan calcitonin receptor-like structure. *Biochem Biophys Res Commun* 206: 341–347, 1995. doi:10. 1006/bbrc.1995.1047.
- Friedman J, Raisz LG. Thyrocalcitonin: inhibitor of bone resorption in tissue culture. Science 150: 1465–1467, 1965. doi:10.1126/science.150.3702.1465.
- Fritz-Six KL, Dunworth WP, Li M, Caron KM. Adrenomedullin signaling is necessary for murine lymphatic vascular development. J Clin Invest 118: 40–50, 2008. doi:10. 1172/JCI33302.
- Furness SGB, Liang YL, Nowell CJ, Halls ML, Wookey PJ, Dal Maso E, Inoue A, Christopoulos A, Wootten D, Sexton PM. Ligand-dependent modulation of G protein conformation alters drug efficacy. *Cell* 167: 739–749.e11, 2016. doi:10.1016/j.cell. 2016.09.021.
- 76. Gilbert W. Why genes in pieces? Nature 271: 501, 1978. doi:10.1038/271501a0.
- 77. Gilbey SG, Ghatei MA, Bretherton-Watt D, Zaidi M, Jones PM, Perera T, Beacham J, Girgis S, Bloom SR. Islet amyloid polypeptide: production by an osteoblast cell line and possible role as a paracrine regulator of osteoclast function in man. *Clin Sci (Lond)* 81: 803–808, 1991. doi:10.1042/cs0810803.
- Gittes RF, Irvin GL III. Roles of thyroxine and thyrocalcitonin in the response to hypercalcemia in rats. *Endocrinology* 79: 1033–1039, 1966. doi:10.1210/endo-79-6-1033.
- Gooi JH, Chia LY, Walsh NC, Karsdal MA, Quinn JM, Martin TJ, Sims NA. Decline in calcitonin receptor expression in osteocytes with age. J Endocrinol 221: 181–191, 2014. doi:10.1530/JOE-13-0524.
- Gooi JH, Pompolo S, Karsdal MA, Kulkarni NH, Kalajzic I, McAhren SH, Han B, Onyia JE, Ho PW, Gillespie MT, Walsh NC, Chia LY, Quinn JM, Martin TJ, Sims NA. Calcitonin impairs the anabolic effect of PTH in young rats and stimulates expression of sclerostin by osteocytes. *Bone* 46: 1486–1497, 2010. doi:10.1016/j.bone.2010.02. 018.
- Gorn AH, Rudolph SM, Flannery MR, Morton CC, Weremowicz S, Wang TZ, Krane SM, Goldring SR. Expression of two human skeletal calcitonin receptor isoforms cloned from a giant cell tumor of bone. The first intracellular domain modulates ligand binding and signal transduction. *J Clin Invest* 95: 2680–2691, 1995. doi:10.1172/ JCI117970.
- Granholm S, Henning P, Lerner UH. Comparisons between the effects of calcitonin receptor-stimulating peptide and intermedin and other peptides in the calcitonin family on bone resorption and osteoclastogenesis. J Cell Biochem 112: 3300–3312, 2011. doi:10.1002/jcb.23256.

- Granholm S, Lundberg P, Lerner UH. Calcitonin inhibits osteoclast formation in mouse haematopoetic cells independently of transcriptional regulation by receptor activator of NF-kappaB and c-Fms. J Endocrinol 195: 415–427, 2007. doi:10.1677/ JOE-07-0338.
- Granholm S, Lundberg P, Lerner UH. Expression of the calcitonin receptor, calcitonin receptor-like receptor, and receptor activity modifying proteins during osteoclast differentiation. J Cell Biochem 104: 920–933, 2008. doi:10.1002/jcb.21674.
- Grauer A, Ziegler R, Raue F. Clinical significance of antibodies against calcitonin. Exp Clin Endocrinol Diabetes 103: 345–351, 1995. doi:10.1055/s-0029-1211376.
- Gutiérrez-Rojas I, Lozano D, Nuche-Berenguer B, Moreno P, Acitores A, Ramos-Álvarez I, Rovira A, Novials A, Martín-Crespo E, Villanueva-Peñacarrillo ML, Esbrit P. Amylin exerts osteogenic actions with different efficacy depending on the diabetic status. *Mol Cell Endocrinol* 365: 309–315, 2013. doi:10.1016/j.mce.2012.11.013.
- Hamada H, Kitamura K, Chosa E, Eto T, Tajima N. Adrenomedullin stimulates the growth of cultured normal human osteoblasts as an autocrine/paracine regulator. *Peptides* 23: 2163–2168, 2002. doi:10.1016/S0196-9781(02)00259-0.
- Hartter E, Svoboda T, Ludvik B, Schuller M, Lell B, Kuenburg E, Brunnbauer M, Woloszczuk W, Prager R. Basal and stimulated plasma levels of pancreatic amylin indicate its co-secretion with insulin in humans. *Diabetologia* 34: 52–54, 1991. doi:10. 1007/BF00404025.
- Hay DL, Pioszak AA. Receptor Activity-Modifying Proteins (RAMPs): New Insights and Roles. Annu Rev Pharmacol Toxicol 56: 469–487, 2016. doi:10.1146/annurevpharmtox-010715-103120.
- Hay DL, Poyner DR, Sexton PM. GPCR modulation by RAMPs. *Pharmacol Ther* 109: 173–197, 2006. doi:10.1016/j.pharmthera.2005.06.015.
- 91. Henriksen K, Byrjalsen I, Andersen JR, Bihlet AR, Russo LA, Alexandersen P, Valter I, Qvist P, Lau E, Riis BJ, Christiansen C, Karsdal MA; SMC021 investigators. A randomized, double-blind, multicenter, placebo-controlled study to evaluate the efficacy and safety of oral salmon calcitonin in the treatment of osteoporosis in postmenopausal women taking calcium and vitamin D. *Bone* 91: 122–129, 2016. doi:10.1016/j.bone. 2016.07.019.
- Hilton JM, Dowton M, Houssami S, Sexton PM. Identification of key components in the irreversibility of salmon calcitonin binding to calcitonin receptors. *J Endocrinol* 166: 213–226, 2000. doi:10.1677/joe.0.1660213.
- Hirsch PF, Baruch H. Is calcitonin an important physiological substance? Endocrine 21: 201–208, 2003. doi:10.1385/ENDO:21:3:201.
- Hirsch PF, Munson PL. Importance of the thyroid gland in the prevention of hypercalcemia in rats. *Endocrinology* 79: 655–658, 1966. doi:10.1210/endo-79-3-655.
- Hirsch PF, Voelkel EF, Munson PL. Thyrocalcitonin: Hypocalcemic Hypophosphatemic Principle of the Thyroid Gland. Science 146: 412–413, 1964. doi:10.1126/ science.146.3642.412.
- Hirt D, Bernard GW. CGRP-beta unlike CGRP-alpha has no osteogenic stimulatory effect in vitro. Peptides 18: 1461–1463, 1997. doi:10.1016/S0196-9781(97)00199-X.
- Hoff AO, Catala-Lehnen P, Thomas PM, Priemel M, Rueger JM, Nasonkin I, Bradley A, Hughes MR, Ordonez N, Cote GJ, Amling M, Gagel RF. Increased bone mass is an unexpected phenotype associated with deletion of the calcitonin gene. J Clin Invest 110: 1849–1857, 2002. doi:10.1172/JCI200214218.
- Hong Y, Hay DL, Quirion R, Poyner DR. The pharmacology of adrenomedullin 2/intermedin. Br J Pharmacol 166: 110–120, 2012. doi:10.1111/j.1476-5381.2011. 01530.x.
- Höppener JW, Steenbergh PH, Zandberg J, Geurts van Kessel AH, Baylin SB, Nelkin BD, Jansz HS, Lips CJ. The second human calcitonin/CGRP gene is located on chromosome 11. *Hum Genet* 70: 259–263, 1985. doi:10.1007/BF00273453.
- Horcajada-Molteni MN, Chanteranne B, Lebecque P, Davicco MJ, Coxam V, Young A, Barlet JP. Amylin and bone metabolism in streptozotocin-induced diabetic rats. J Bone Miner Res 16: 958–965, 2001. doi:10.1359/jbmr.2001.16.5.958.
- Horcajada-Molteni MN, Davicco MJ, Lebecque P, Coxam V, Young AA, Barlet JP. Amylin inhibits ovariectomy-induced bone loss in rats. J Endocrinol 165: 663–668, 2000. doi:10.1677/joe.0.1650663.
- Huang CZ, Yang XN, Liu DC, Sun YG, Dai XM. Calcitonin Gene-Related Peptide-Induced Calcium Alginate Gel Combined with Adipose-Derived Stem Cells Differen-

tiating to Osteoblasts. *Cell Biochem Biophys* 73: 609–617, 2015. doi:10.1007/s12013-015-0630-8.

- Huebner AK, Keller J, Catala-Lehnen P, Perkovic S, Streichert T, Emeson RB, Amling M, Schinke T. The role of calcitonin and alpha-calcitonin gene-related peptide in bone formation. Arch Biochem Biophys 473: 210–217, 2008. doi:10.1016/j.abb.2008.02. 013.
- Huebner AK, Schinke T, Priemel M, Schilling S, Schilling AF, Emeson RB, Rueger JM, Amling M. Calcitonin deficiency in mice progressively results in high bone turnover. J Bone Miner Res 21: 1924–1934, 2006. doi: 10.1359/jbmr.060820.
- 105. Hurley DL, Tiegs RD, Wahner HW, Heath H III. Axial and appendicular bone mineral density in patients with long-term deficiency or excess of calcitonin. N Engl J Med 317: 537–541, 1987. doi:10.1056/NEJM198708273170904.
- 106. Ikegame M, Rakopoulos M, Martin TJ, Moseley JM, Findlay DM. Effects of continuous calcitonin treatment on osteoclast-like cell development and calcitonin receptor expression in mouse bone marrow cultures. J Bone Miner Res 11: 456–465, 1996. doi:10.1002/jbmr.5650110406.
- 107. Kadmiel M, Fritz-Six K, Pacharne S, Richards GO, Li M, Skerry TM, Caron KM. Research resource: Haploinsufficiency of receptor activity-modifying protein-2 (RAMP2) causes reduced fertility, hyperprolactinemia, skeletal abnormalities, and endocrine dysfunction in mice. *Mol Endocrinol* 25: 1244–1253, 2011. doi:10.1210/me. 2010-0400.
- Kadmiel M, Fritz-Six KL, Caron KM. Understanding RAMPs through genetically engineered mouse models. Adv Exp Med Biol 744: 49–60, 2012. doi:10.1007/978-1-4614-2364-5_5.
- 109. Kalu DN, Hardin RR. Evaluation of the role of calcitonin deficiency in ovariectomy-induced osteopenia. *Life Sci* 34: 2393–2398, 1984. doi:10.1016/0024-3205(84)90427-2.
- 110. Kanis JA, Johnell O, Gullberg B, Allander E, Dilşen G, Gennari C, Lopes Vaz AA, Lyritis GP, Mazzuoli G, Miravet L. Evidence for efficacy of drugs affecting bone metabolism in preventing hip fracture. *BMJ* 305: 1124–1128, 1992. doi:10.1136/bmj.305.6862. 1124.
- 111. Karsdal MA, Byrjalsen I, Alexandersen P, Bihlet A, Andersen JR, Riis BJ, Bay-Jensen AC, Christiansen C; CSMC021C2301/2 investigators. Treatment of symptomatic knee osteoarthritis with oral salmon calcitonin: results from two phase 3 trials. Osteoarthritis Cartilage 23: 532–543, 2015. doi:10.1016/j.joca.2014.12.019.
- 112. Karsdal MA, Byrjalsen I, Riis BJ, Christiansen C. Optimizing bioavailability of oral administration of small peptides through pharmacokinetic and pharmacodynamic parameters: the effect of water and timing of meal intake on oral delivery of Salmon Calcitonin. BMC Clin Pharmacol 8: 5, 2008. doi:10.1186/1472-6904-8-5.
- 113. Karsdal MA, Henriksen K, Bay-Jensen AC, Molloy B, Arnold M, John MR, Byrjalsen I, Azria M, Riis BJ, Qvist P, Christiansen C. Lessons learned from the development of oral calcitonin: the first tablet formulation of a protein in phase III clinical trials. J Clin Pharmacol 51: 460–471, 2011. doi:10.1177/0091270010372625.
- 114. Katafuchi T, Kikumoto K, Hamano K, Kangawa K, Matsuo H, Minamino N. Calcitonin receptor-stimulating peptide, a new member of the calcitonin gene-related peptide family. Its isolation from porcine brain, structure, tissue distribution, and biological activity. J Biol Chem 278: 12046–12054, 2003. doi:10.1074/jbc.M207970200.
- 115. Katafuchi T, Yasue H, Osaki T, Minamino N. Calcitonin receptor-stimulating peptide: Its evolutionary and functional relationship with calcitonin/calcitonin gene-related peptide based on gene structure. *Peptides* 30: 1753–1762, 2009. doi:10.1016/j. peptides.2009.06.012.
- 116. Kato Y, Windle JJ, Koop BA, Mundy GR, Bonewald LF. Establishment of an osteocytelike cell line, MLO-Y4. J Bone Miner Res 12: 2014–2023, 1997. doi:10.1359/jbmr.1997. 12.12.2014.
- 117. Kauther MD, Bachmann HS, Neuerburg L, Broecker-Preuss M, Hilken G, Grabellus F, Koehler G, von Knoch M, Wedemeyer C. Calcitonin substitution in calcitonin deficiency reduces particle-induced osteolysis. *BMC Musculoskelet Disord* 12: 186, 2011. doi:10.1186/1471-2474-12-186.
- 118. Kawase T, Howard GA, Roos BA, Burns DM. Diverse actions of calcitonin generelated peptide on intracellular free Ca2+ concentrations in UMR 106 osteoblastic cells. Bone 16, Suppl: 379S–384S, 1995. doi:10.1016/S8756-3282(95)80457-9.

- 119. Keller J, Catala-Lehnen P, Huebner AK, Jeschke A, Heckt T, Lueth A, Krause M, Koehne T, Albers J, Schulze J, Schilling S, Haberland M, Denninger H, Neven M, Hermans-Borgmeyer I, Streichert T, Breer S, Barvencik F, Levkau B, Rathkolb B, Wolf E, Calzada-Wack J, Neff F, Gailus-Durner V, Fuchs H, de Angelis MH, Klutmann S, Tsourdi E, Hofbauer LC, Kleuser B, Chun J, Schinke T, Amling M. Calcitonin controls bone formation by inhibiting the release of sphingosine 1-phosphate from osteoclasts. Nat Commun 5: 5215, 2014. doi:10.1038/ncomms6215.
- Khosla S. Minireview: the OPG/RANKL/RANK system. Endocrinology 142: 5050– 5055, 2001. doi:10.1210/endo.142.12.8536.
- 121. Kitamura K, Kangawa K, Kawamoto M, Ichiki Y, Nakamura S, Matsuo H, Eto T. Adrenomedullin: a novel hypotensive peptide isolated from human pheochromocytoma. *Biochem Biophys Res Commun* 192: 553–560, 1993. doi:10.1006/bbrc.1993. 1451.
- 122. Kittur SD, Hoppener JW, Antonarakis SE, Daniels JD, Meyers DA, Maestri NE, Jansen M, Korneluk RG, Nelkin BD, Kazazian HH Jr. Linkage map of the short arm of human chromosome II: location of the genes for catalase, calcitonin, and insulin-like growth factor II. Proc Natl Acad Sci USA 82: 5064–5067, 1985. doi:10.1073/pnas.82.15.5064.
- 123. Klein KR, Matson BC, Caron KM. The expanding repertoire of receptor activity modifying protein (RAMP) function. *Crit Rev Biochem Mol Biol* 51: 65–71, 2016. doi: 10.3109/10409238.2015.1128875.
- 124. Konarkowska B, Aitken JF, Kistler J, Zhang S, Cooper GJ. The aggregation potential of human amylin determines its cytotoxicity towards islet beta-cells. *FEBS J* 273: 3614– 3624, 2006. doi:10.1111/j.1742-4658.2006.05367.x.
- 125. Kowalczyk R, Brimble MA, Callon KE, Watson M, Cornish J. How to blast osteoblasts? Novel dicarba analogues of amylin-(1-8) to treat osteoporosis. *Bioorg Med Chem* 20: 6011–6018, 2012. doi:10.1016/j.bmc.2012.08.053.
- 126. Kowalczyk R, Harris PW, Brimble MA, Callon KE, Watson M, Cornish J. Synthesis and evaluation of disulfide bond mimetics of amylin-(1-8) as agents to treat osteoporosis. *Bioorg Med Chem* 20: 2661–2668, 2012. doi:10.1016/j.bmc.2012.02.030.
- 127. Kuestner RE, Elrod RD, Grant FJ, Hagen FS, Kuijper JL, Matthewes SL, O'Hara PJ, Sheppard PO, Stroop SD, Thompson DL, . Cloning and characterization of an abundant subtype of the human calcitonin receptor. *Mol Pharmacol* 46: 246–255, 1994.
- 128. Kurashige C, Hosono K, Matsuda H, Tsujikawa K, Okamoto H, Majima M. Roles of receptor activity-modifying protein 1 in angiogenesis and lymphangiogenesis during skin wound healing in mice. FASEB J 28: 1237–1247, 2014. doi:10.1096/fj.13-238998.
- 129. Langston AL, Ralston SH. Management of Paget's disease of bone. Rheumatology (Oxford) 43: 955–959, 2004. doi:10.1093/rheumatology/keh243.
- Lausson S, Cressent M. Signal transduction pathways mediating the effect of adrenomedullin on osteoblast survival. J Cell Biochem 112: 3807–3815, 2011. doi:10.1002/ jcb.23311.
- 131. Li M, Wetzel-Strong SE, Hua X, Tilley SL, Oswald E, Krummel MF, Caron KM. Deficiency of RAMP1 attenuates antigen-induced airway hyperresponsiveness in mice. PLoS One 9: e102356, 2014. doi:10.1371/journal.pone.0102356.
- 132. Li Y, Yang L, Zheng Z, Li Z, Deng T, Ren W, Wu C, Guo L. Bio-Oss[®] modified by calcitonin gene-related peptide promotes osteogenesis in vitro. Exp Ther Med 14: 4001–4008, 2017.
- Liang W, Zhuo X, Tang Z, Wei X, Li B. Calcitonin gene-related peptide stimulates proliferation and osteogenic differentiation of osteoporotic rat-derived bone mesenchymal stem cells. *Mol Cell Biochem* 402: 101–110, 2015. doi:10.1007/s11010-014-2318-6.
- 134. Lin HY, Harris TL, Flannery MS, Aruffo A, Kaji EH, Gorn A, Kolakowski LF Jr, Yamin M, Lodish HF, Goldring SR. Expression cloning and characterization of a porcine renal calcitonin receptor. *Trans Assoc Am Physicians* 104: 265–272, 1991.
- Lin Z, Pavlos NJ, Cake MA, Wood DJ, Xu J, Zheng MH. Evidence that human cartilage and chondrocytes do not express calcitonin receptor. *Osteoarthritis Cartilage* 16: 450–457, 2008. doi:10.1016/j.joca.2007.08.003.
- Litwic A, Edwards MH, Dennison EM, Cooper C. Epidemiology and burden of osteoarthritis. Br Med Bull 105: 185–199, 2013. doi:10.1093/bmb/lds038.
- Lou H, Gagel RF, Berget SM. An intron enhancer recognized by splicing factors activates polyadenylation. Genes Dev 10: 208–219, 1996. doi:10.1101/gad.10.2.208.

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BONE ACTIVITY OF CALCITONIN-FAMILY PEPTIDES

- 138. MacIntyre I. Amylinamide, bone conservation, and pancreatic beta cells. Lancet 2: 1026-1027, 1989. doi:10.1016/S0140-6736(89)91028-3.
- 139. MacIntyre I, Stevenson JC, Whitehead MI, Wimalawansa SJ, Banks LM, Healy MJ. Calcitonin for prevention of postmenopausal bone loss. *Lancet* 1: 900–902, 1988. doi:10.1016/S0140-6736(88)91712-6.
- 140. Mancini L, Paul-Clark MJ, Rosignoli G, Hannon R, Martin JE, Macintyre I, Perretti M. Calcitonin and prednisolone display antagonistic actions on bone and have synergistic effects in experimental arthritis. Am J Pathol 170: 1018–1027, 2007. doi:10.2353/ ajpath.2007.060830.
- 141. Manicourt DH, Azria M, Mindeholm L, Thonar EJ, Devogelaer JP. Oral salmon calcitonin reduces Lequesne's algofunctional index scores and decreases urinary and serum levels of biomarkers of joint metabolism in knee osteoarthritis. *Arthritis Rheum* 54: 3205–3211, 2006. doi:10.1002/art.22075.
- 142. Martin TJ, Robinson CJ, MacIntyre I. The mode of action of thyrocalcitonin. Lancet 1: 900–902, 1966. doi:10.1016/S0140-6736(66)91577-7.
- 143. Martin TJ, Sims NA. Calcitonin physiology, saved by a lysophospholipid. J Bone Miner Res 30: 212–215, 2015. doi:10.1002/jbmr.2449.
- 144. Martínez-Herrero S, Larrayoz IM, Ochoa-Callejero L, Fernández LJ, Allueva A, Ochoa I, Martínez A. Prevention of Bone Loss in a Model of Postmenopausal Osteoporosis through Adrenomedullin Inhibition. *Front Physiol* 7: 280, 2016. doi:10.3389/fphys. 2016.00280.
- Martínez A. Biology of adrenomedullin. Introduction. *Microsc Res Tech* 57: 1–2, 2002. doi:10.1002/jemt.10045.
- 146. McLatchie LM, Fraser NJ, Main MJ, Wise A, Brown J, Thompson N, Solari R, Lee MG, Foord SM. RAMPs regulate the transport and ligand specificity of the calcitoninreceptor-like receptor. *Nature* 393: 333–339, 1998. doi:10.1038/30666.
- 147. McLean FC. The parathyroid hormone and bone. Clin Orthop 9: 46-60, 1957.
- 148. Michelangeli VP, Findlay DM, Fletcher A, Martin TJ. Calcitonin gene-related peptide (CGRP) acts independently of calcitonin on cyclic AMP formation in clonal osteogenic sarcoma cells (UMR 106-01). *Calcif Tissue Int* 39: 44–48, 1986. doi:10.1007/ BF02555739.
- 149. Michelangeli VP, Fletcher AE, Allan EH, Nicholson GC, Martin TJ. Effects of calcitonin gene-related peptide on cyclic AMP formation in chicken, rat, and mouse bone cells. *J Bone Miner Res* 4: 269–272, 1989. doi:10.1002/jbmr.5650040220.
- 150. Miller S. Calcitonin-guardian of the Mammalian skeleton or is it just a fish story? Endocrinology 147: 4007–4009, 2006. doi:10.1210/en.2006-0599.
- 151. Mitsukawa T, Takemura J, Asai J, Nakazato M, Kangawa K, Matsuo H, Matsukura S. Islet amyloid polypeptide response to glucose, insulin, and somatostatin analogue administration. *Diabetes* 39: 639–642, 1990. doi:10.2337/diab.39.5.639.
- 152. Morfis M, Tilakaratne N, Furness SG, Christopoulos G, Werry TD, Christopoulos A, Sexton PM. Receptor activity-modifying proteins differentially modulate the G protein-coupling efficiency of amylin receptors. *Endocrinology* 149: 5423–5431, 2008. doi:10.1210/en.2007-1735.
- 153. Morikawa T, Munekata E, Sakakibara S, Noda T, Otani M. Synthesis of eel-calcitonin and (asu I,7)-eel-calcitonin: contribution of the disulfide bond to the hormonal activity. Experientia 32: 1104–1106, 1976. doi:10.1007/BF01927568.
- 154. Mrak E, Guidobono F, Moro G, Fraschini G, Rubinacci A, Villa I. Calcitonin generelated peptide (CGRP) inhibits apoptosis in human osteoblasts by β-catenin stabilization. J Cell Physiol 225: 701–708, 2010. doi:10.1002/jcp.22266.
- 155. Muff R, Born W, Lutz TA, Fischer JA. Biological importance of the peptides of the calcitonin family as revealed by disruption and transfer of corresponding genes. *Peptides* 25: 2027–2038, 2004. doi:10.1016/j.peptides.2004.08.007.
- 156. Muff R, Bühlmann N, Fischer JA, Born W. An amylin receptor is revealed following co-transfection of a calcitonin receptor with receptor activity modifying proteins-1 or -3. Endocrinology 140: 2924–2927, 1999. doi:10.1210/endo.140.6.6930.
- 157. Mullins MW, Ciallella J, Rangnekar V, McGillis JP. Characterization of a calcitonin gene-related peptide (CGRP) receptor on mouse bone marrow cells. *Regul Pept* 49: 65–72, 1993. doi:10.1016/0167-0115(93)90385-L.

- 158. Mundy GR, Wilkinson R, Heath DA. Comparative study of available medical therapy for hypercalcemia of malignancy. Am J Med 74: 421–432, 1983. doi:10.1016/0002-9343(83)90961-0.
- 159. Naot D, Callon KE, Grey A, Cooper GJ, Reid IR, Cornish J. A potential role for adrenomedullin as a local regulator of bone growth. *Endocrinology* 142: 1849–1857, 2001. doi:10.1210/endo.142.5.8152.
- 160. Naot D, Cornish J. The role of peptides and receptors of the calcitonin family in the regulation of bone metabolism. *Bone* 43: 813–818, 2008. doi:10.1016/j.bone.2008. 07.003.
- 161. Neher R, Riniker B, Maier R, Byfield PG, Gudmundsson TV, MacIntyre I. Human calcitonin. Nature 220: 984–986, 1968. doi:10.1038/220984a0.
- 162. Niall HD, Keutmann HT, Copp DH, Potts JT Jr. Amino acid sequence of salmon ultimobranchial calcitonin. Proc Natl Acad Sci USA 64: 771–778, 1969. doi:10.1073/ pnas.64.2.771.
- Njuki F, Nicholl CG, Howard A, Mak JC, Barnes PJ, Girgis SI, Legon S. A new calcitonin-receptor-like sequence in rat pulmonary blood vessels. *Clin Sci (Lond)* 85: 385– 388, 1993. doi:10.1042/cs0850385.
- 164. O'Brien TD, Westermark P, Johnson KH. Islet amyloid polypeptide and insulin secretion from isolated perfused pancreas of fed, fasted, glucose-treated, and dexamethasone-treated rats. *Diabetes* 40: 1701–1706, 1991. doi:10.2337/diab.40.12.1701.
- 165. Overman RA, Borse M, Gourlay ML. Salmon calcitonin use and associated cancer risk. Ann Pharmacother 47: 1675–1684, 2013. doi:10.1177/1060028013509233.
- 166. Pietschmann P, Farsoudi KH, Hoffmann O, Klaushofer K, Hörandner H, Peterlik M. Inhibitory effect of amylin on basal and parathyroid hormone-stimulated bone resorption in cultured neonatal mouse calvaria. *Bone* 14: 167–172, 1993. doi:10.1016/8756-3282(93)90244-5.
- 167. Plotkin LI, Weinstein RS, Parfitt AM, Roberson PK, Manolagas SC, Bellido T. Prevention of osteocyte and osteoblast apoptosis by bisphosphonates and calcitonin. J Clin Invest 104: 1363–1374, 1999. doi:10.1172/JCI6800.
- Potts JT. Parathyroid hormone: past and present. J Endocrinol 187: 311–325, 2005. doi:10.1677/joe.1.06057.
- 169. Poyner DR, Sexton PM, Marshall I, Smith DM, Quirion R, Born W, Muff R, Fischer JA, Foord SM. International Union of Pharmacology. XXXII. The mammalian calcitonin gene-related peptides, adrenomedullin, amylin, and calcitonin receptors. *Pharmacol Rev* 54: 233–246, 2002. doi:10.1124/pr.54.2.233.
- 170. Qiao YC, Ling W, Pan YH, Chen YL, Zhou D, Huang YM, Zhang XX, Zhao HL. Efficacy and safety of pramlintide injection adjunct to insulin therapy in patients with type I diabetes mellitus: a systematic review and meta-analysis. *Oncotarget* 8: 66504– 66515, 2017. doi:10.18632/oncotarget.16008.
- 171. Qing H, Ardeshirpour L, Pajevic PD, Dusevich V, Jähn K, Kato S, Wysolmerski J, Bonewald LF. Demonstration of osteocytic perilacunar/canalicular remodeling in mice during lactation. J Bone Miner Res 27: 1018–1029, 2012. doi:10.1002/jbmr.1567.
- Quinn JM, Morfis M, Lam MH, Elliott J, Kartsogiannis V, Williams ED, Gillespie MT, Martin TJ, Sexton PM. Calcitonin receptor antibodies in the identification of osteoclasts. *Bone* 25: 1–8, 1999. doi:10.1016/S8756-3282(99)00094-0.
- 173. Rakopoulos M, Ikegame M, Findlay DM, Martin TJ, Moseley JM. Short treatment of osteoclasts in bone marrow culture with calcitonin causes prolonged suppression of calcitonin receptor mRNA. *Bone* 17: 447–453, 1995. doi:10.1016/8756-3282(95)00280-8.
- 174. Reginster JY, Franchimont P. Side effects of synthetic salmon calcitonin given by intranasal spray compared with intramuscular injection. *Clin Exp Rheumatol* 3: 155– 157, 1985.
- 175. Ren H, Ren H, Li X, Yu D, Mu S, Chen Z, Fu Q. Effects of intermedin on proliferation, apoptosis and the expression of OPG/RANKL/M-CSF in the MC3T3-EI osteoblast cell line. *Mol Med Rep* 12: 6711–6717, 2015. doi:10.3892/mmr.2015.4328.
- 176. Rodriguez M, Felsenfeld AJ, Torres A, Pederson L, Llach F. Calcitonin, an important factor in the calcemic response to parathyroid hormone in the rat. *Kidney Int* 40: 219–225, 1991. doi:10.1038/ki.1991.203.
- 177. Roh J, Chang CL, Bhalla A, Klein C, Hsu SY. Intermedin is a calcitonin/calcitonin gene-related peptide family peptide acting through the calcitonin receptor-like recep-

tor/receptor activity-modifying protein receptor complexes. J Biol Chem 279: 7264–7274, 2004. doi:10.1074/jbc.M305332200.

- 178. Romero DF, Bryer HP, Rucinski B, Isserow JA, Buchinsky FJ, Cvetkovic M, Liu CC, Epstein S. Amylin increases bone volume but cannot ameliorate diabetic osteopenia. *Calcif Tissue Int* 56: 54–61, 1995. doi:10.1007/BF00298745.
- 179. Roos BA, Fischer JA, Pignat W, Alander CB, Raisz LG. Evaluation of the in vivo and in vitro calcium-regulating actions of noncalcitonin peptides produced via calcitonin gene expression. *Endocrinology* 118: 46–51, 1986. doi:10.1210/endo-118-1-46.
- Rosenfeld MG, Amara SG, Roos BA, Ong ES, Evans RM. Altered expression of the calcitonin gene associated with RNA polymorphism. *Nature* 290: 63–65, 1981. doi: 10.1038/290063a0.
- 181. Rosenfeld MG, Mermod JJ, Amara SG, Swanson LW, Sawchenko PE, Rivier J, Vale WW, Evans RM. Production of a novel neuropeptide encoded by the calcitonin gene via tissue-specific RNA processing. *Nature* 304: 129–135, 1983. doi:10.1038/ 304129a0.
- 182. Saag KG, Petersen J, Brandi ML, Karaplis AC, Lorentzon M, Thomas T, Maddox J, Fan M, Meisner PD, Grauer A. Romosozumab or Alendronate for Fracture Prevention in Women with Osteoporosis. N Engl J Med 377: 1417–1427, 2017. doi:10.1056/ NEJMoa1708322.
- 183. Sample SJ, Heaton CM, Behan M, Bleedorn JA, Racette MA, Hao Z, Muir P. Role of calcitonin gene-related peptide in functional adaptation of the skeleton. *PLoS One* 9: e113959, 2014. doi:10.1371/journal.pone.0113959.
- 184. Sanke T, Hanabusa T, Nakano Y, Oki C, Okai K, Nishimura S, Kondo M, Nanjo K. Plasma islet amyloid polypeptide (Amylin) levels and their responses to oral glucose in type 2 (non-insulin-dependent) diabetic patients. *Diabetologia* 34: 129–132, 1991. doi:10.1007/BF00500385.
- 185. Schinke T, Liese S, Priemel M, Haberland M, Schilling AF, Catala-Lehnen P, Blicharski D, Rueger JM, Gagel RF, Emeson RB, Amling M. Decreased bone formation and osteopenia in mice lacking alpha-calcitonin gene-related peptide. *J Bone Miner Res* 19: 2049–2056, 2004. doi:10.1359/jbmr.040915.
- 186. Seitz PK, Thomas ML, Cooper CW. Binding of calcitonin and calcitonin gene-related peptide to calvarial cells and renal cortical membranes. J Bone Miner Res 1: 51–56, 1986. doi:10.1002/jbmr.5650010109.
- 187. Sexton PM, Houssami S, Hilton JM, O'Keeffe LM, Center RJ, Gillespie MT, Darcy P, Findlay DM. Identification of brain isoforms of the rat calcitonin receptor. *Mol Endocrinol* 7: 815–821, 1993.
- 188. Shen CJ, Wu MS, Lin KH, Lin WL, Chen HC, Wu JY, Lee MC, Lee CC. The use of procalcitonin in the diagnosis of bone and joint infection: a systemic review and meta-analysis. Eur J Clin Microbiol Infect Dis 32: 807–814, 2013. doi:10.1007/s10096-012-1812-6.
- 189. Shindo T, Sakurai T, Kamiyoshi A, Ichikawa-Shindo Y, Shimoyama N, Iinuma N, Arai T, Miyagawa S. Regulation of adrenomedullin and its family peptide by RAMP system– lessons from genetically engineered mice. *Curr Protein Pept Sci* 14: 347–357, 2013. doi:10.2174/13892037113149990052.
- 190. Shyu JF, Shih C, Tseng CY, Lin CH, Sun DT, Liu HT, Tsung HC, Chen TH, Lu RB. Calcitonin induces podosome disassembly and detachment of osteoclasts by modulating Pyk2 and Src activities. Bone 40: 1329–1342, 2007. doi:10.1016/j.bone.2007. 01.014.
- 191. Sondergaard BC, Catala-Lehnen P, Huebner AK, Bay-Jensen AC, Schinke T, Henriksen K, Schilling S, Haberland M, Nielsen RH, Amling M, Karsdal MA. Mice overexpressing salmon calcitonin have strongly attenuated osteoarthritic histopathological changes after destabilization of the medial meniscus. *Osteoarthritis Cartilage* 20: 136– 143, 2012. doi:10.1016/j.joca.2011.11.004.
- 192. Sondergaard BC, Oestergaard S, Christiansen C, Tankó LB, Karsdal MA. The effect of oral calcitonin on cartilage turnover and surface erosion in an ovariectomized rat model. Arthritis Rheum 56: 2674–2678, 2007. doi:10.1002/art.22797.
- 193. Sondergaard BC, Wulf H, Henriksen K, Schaller S, Oestergaard S, Qvist P, Tankó LB, Bagger YZ, Christiansen C, Karsdal MA. Calcitonin directly attenuates collagen type II degradation by inhibition of matrix metalloproteinase expression and activity in articular chondrocytes. Osteoarthritis Cartilage 14: 759–768, 2006. doi:10.1016/j.joca. 2006.01.014.

- Takahashi S, Goldring S, Katz M, Hilsenbeck S, Williams R, Roodman GD. Downregulation of calcitonin receptor mRNA expression by calcitonin during human osteoclastlike cell differentiation. J Clin Invest 95: 167–171, 1995. doi:10.1172/JCI117634.
- 195. Takei Y, Inoue K, Ogoshi M, Kawahara T, Bannai H, Miyano S. Identification of novel adrenomedullin in mammals: a potent cardiovascular and renal regulator. FEBS Lett 556: 53–58, 2004. doi:10.1016/S0014-5793(03)01368-1.
- Talmage RV, Neuenschwander J, Kraintz L. Evidence for the Existence of Thyrocalcitonin in the Rat. Endocrinology 76: 103–107, 1965. doi:10.1210/endo-76-1-103.
- 197. Tamura T, Miyaura C, Owan I, Suda T. Mechanism of action of amylin in bone. J Cell Physiol 153: 6–14, 1992. doi:10.1002/jcp.1041530103.
- 198. Tanaka S, Yoshida A, Kono S, Oguma T, Hasegawa K, Ito M. Effectiveness of elcatonin for alleviating pain and inhibiting bone resorption in patients with osteoporotic vertebral fractures. J Bone Miner Metab. In press.
- 199. Tazaki M, Endoh T, Kobayashi H, Nobushima H, Shibukawa Y, Tsumura M, Sato M, Ubaidus S, Sueishi K. Adrenomedullin facilitates calcium channel currents in osteoblasts. *Bull Tokyo Dent Coll* 53: 203–206, 2012. doi:10.2209/tdcpublication.53.203.
- Thiebaud D, Akatsu T, Yamashita T, Suda T, Noda T, Martin RE, Fletcher AE, Martin TJ. Structure-activity relationships in calcitonin gene-related peptide: cyclic AMP response in a preosteoblast cell line (KS-4). *J Bone Miner Res* 6: 1137–1142, 1991. doi:10.1002/jbmr.5650061016.
- Tian G, Zhang G, Tan YH. Calcitonin gene-related peptide stimulates BMP-2 expression and the differentiation of human osteoblast-like cells in vitro. Acta Pharmacol Sin 34: 1467–1474, 2013. doi:10.1038/aps.2013.41.
- Tippins JR, Morris HR, Panico M, Etienne T, Bevis P, Girgis S, MacIntyre I, Azria M, Attinger M. The myotropic and plasma-calcium modulating effects of calcitonin generelated peptide (CGRP). *Neuropeptides* 4: 425–434, 1984. doi:10.1016/0143-4179(84)90118-5.
- 203. Tsujikawa K, Yayama K, Hayashi T, Matsushita H, Yamaguchi T, Shigeno T, Ogitani Y, Hirayama M, Kato T, Fukada S, Takatori S, Kawasaki H, Okamoto H, Ikawa M, Okabe M, Yamamoto H. Hypertension and dysregulated proinflammatory cytokine production in receptor activity-modifying protein I-deficient mice. *Proc Natl Acad Sci USA* 104: 16702–16707, 2007. doi:10.1073/pnas.0705974104.
- 204. Turner AG, Tjahyono F, Chiu WS, Skinner J, Sawyer R, Moore AJ, Morris HA, Findlay DM, Zajac JD, Davey RA. The role of the calcitonin receptor in protecting against induced hypercalcemia is mediated via its actions in osteoclasts to inhibit bone resorption. *Bone* 48: 354–361, 2011. doi:10.1016/j.bone.2010.09.013.
- Uzan B, de Vernejoul MC, Cressent M. RAMPs and CRLR expressions in osteoblastic cells after dexamethasone treatment. *Biochem Biophys Res Commun* 321: 802–808, 2004. doi:10.1016/j.bbrc.2004.07.037.
- Uzan B, Villemin A, Garel JM, Cressent M. Adrenomedullin is anti-apoptotic in osteoblasts through CGRP1 receptors and MEK-ERK pathway. J Cell Physiol 215: 122–128, 2008. doi:10.1002/jcp.21294.
- Valentijn K, Gutow AP, Troiano N, Gundberg C, Gilligan JP, Vignery A. Effects of calcitonin gene-related peptide on bone turnover in ovariectomized rats. *Bone* 21: 269–274, 1997. doi:10.1016/S8756-3282(97)00142-7.
- Vijayan AL, Vanimaya, Ravindran S, Saikant R, Lakshmi S, Kartik R, G M. Procalcitonin: a promising diagnostic marker for sepsis and antibiotic therapy. J Intensive Care 5: 51, 2017. doi:10.1186/s40560-017-0246-8.
- Villa I, Dal Fiume C, Maestroni A, Rubinacci A, Ravasi F, Guidobono F. Human osteoblast-like cell proliferation induced by calcitonin-related peptides involves PKC activity. Am J Physiol Endocrinol Metab 284: E627–E633, 2003. doi:10.1152/ajpendo.00307. 2002.
- Villa I, Melzi R, Pagani F, Ravasi F, Rubinacci A, Guidobono F. Effects of calcitonin gene-related peptide and amylin on human osteoblast-like cells proliferation. *Eur J Pharmacol* 409: 273–278, 2000. doi:10.1016/S0014-2999(00)00872-4.
- Villa I, Rubinacci A, Ravasi F, Ferrara AF, Guidobono F. Effects of amylin on human osteoblast-like cells. *Peptides* 18: 537–540, 1997. doi:10.1016/S0196-9781(97)00056-9.
- Wada S, Udagawa N, Nagata N, Martin TJ, Findlay DM. Calcitonin receptor downregulation relates to calcitonin resistance in mature mouse osteoclasts. *Endocrinology* 137: 1042–1048, 1996. doi:10.1210/endo.137.3.8603572.

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BONE ACTIVITY OF CALCITONIN-FAMILY PEPTIDES

- 213. Wang L, Shi X, Zhao R, Halloran BP, Clark DJ, Jacobs CR, Kingery WS. Calcitoningene-related peptide stimulates stromal cell osteogenic differentiation and inhibits RANKL induced NF-kappaB activation, osteoclastogenesis and bone resorption. *Bone* 46: 1369–1379, 2010. doi:10.1016/j.bone.2009.11.029.
- Weiss RE, Singer FR, Gorn AH, Hofer DP, Nimni ME. Calcitonin stimulates bone formation when administered prior to initiation of osteogenesis. J Clin Invest 68: 815–818, 1981. doi:10.1172/JCI110319.
- Wener JA, Gorton SJ, Raisz LG. Escape from inhibition or resorption in cultures of fetal bone treated with calcitoninand parathyroid hromone. *Endocrinology* 90: 752–759, 1972. doi:10.1210/endo-90-3-752.
- Westermark P, Andersson A, Westermark GT. Islet amyloid polypeptide, islet amyloid, and diabetes mellitus. *Physiol Rev* 91: 795–826, 2011. doi:10.1152/physrev. 00042.2009.
- 217. Westermark P, Wernstedt C, Wilander E, Hayden DW, O'Brien TD, Johnson KH. Amyloid fibrils in human insulinoma and islets of Langerhans of the diabetic cat are derived from a neuropeptide-like protein also present in normal islet cells. *Proc Natl Acad Sci USA* 84: 3881–3885, 1987. doi:10.1073/pnas.84.11.3881.
- 218. Weston C, Winfield I, Harris M, Hodgson R, Shah A, Dowell SJ, Mobarec JC, Woodlock DA, Reynolds CA, Poyner DR, Watkins HA, Ladds G. Receptor Activity-modifying Protein-directed G Protein Signaling Specificity for the Calcitonin Gene-related Peptide Family of Receptors. *J Biol Chem* 291: 21925–21944, 2016. doi:10.1074/jbc.M116.751362. A correction to this article is available at http://dx.doi.org/10.1074/jbc.A116.751362.
- Wilkinson R. Treatment of hypercalcaemia associated with malignancy. Br Med J (Clin Res Ed) 288: 812–813, 1984. doi:10.1136/bmj.288.6420.812.
- 220. Wimalawansa SJ. Amylin, calcitonin gene-related peptide, calcitonin, and adrenomedullin: a peptide superfamily. *Crit Rev Neurobiol* 11: 167–239, 1997. doi:10.1615/ CritRevNeurobiol.v11.i2-3.40.
- 221. Wimalawansa SJ, Gunasekera RD, Datta HK. Hypocalcemic actions of amylin amide in humans. J Bone Miner Res 7: 1113–1116, 1992. doi:10.1002/jbmr.5650070915.
- Woodrow JP, Sharpe CJ, Fudge NJ, Hoff AO, Gagel RF, Kovacs CS. Calcitonin plays a critical role in regulating skeletal mineral metabolism during lactation. *Endocrinology* 147: 4010–4021, 2006. doi:10.1210/en.2005-1616.

- 223. Wüster C, Raue F, Meyer C, Bergmann M, Ziegler R. Long-term excess of endogenous calcitonin in patients with medullary thyroid carcinoma does not affect bone mineral density. J Endocrinol 134: 141–147, 1992. doi:10.1677/joe.0.1340141.
- 224. Xu J, Kauther MD, Hartl J, Wedemeyer C; Study was performed at the University of Duisburg Essen, Germany. Effects of alpha-calcitonin gene-related peptide on osteoprotegerin and receptor activator of nuclear factor-κB ligand expression in MG-63 osteoblast-like cells exposed to polyethylene particles. J Orthop Surg 5: 83, 2010. doi:10.1186/1749-799X-5-83.
- Yamamoto I, Kitamura N, Aoki J, Shigeno C, Hino M, Asonuma K, Torizuka K, Fujii N, Otaka A, Yajima H. Human calcitonin gene-related peptide possesses weak inhibitory potency of bone resorption in vitro. *Calcif Tissue Int* 38: 339–341, 1986. doi:10.1007/ BF02555747.
- Young A. Tissue expression and secretion of amylin. Adv Pharmacol 52: 19–45, 2005. doi:10.1016/S1054-3589(05)52002-7.
- Yun HJ, Lee EG, Lee SI, Chae HJ, Yoo WH. Adrenomedullin inhibits MAPK pathwaydependent rheumatoid synovial fibroblast-mediated osteoclastogenesis by IL-1 and TNFalpha. *Rheumatol Int* 29: 1161–1168, 2009. doi:10.1007/s00296-008-0832-0.
- Zaidi M, Datta HK, Bevis PJ, Wimalawansa SJ, MacIntyre I. Amylin-amide: a new bone-conserving peptide from the pancreas. *Exp Physiol* 75: 529–536, 1990. doi:10. 1113/expphysiol.1990.sp003429.
- Zaidi M, Datta HK, Moonga BS, MacIntyre I. Evidence that the action of calcitonin on rat osteoclasts is mediated by two G proteins acting via separate post-receptor pathways. J Endocrinol 126: 473–481, 1990. doi:10.1677/joe.0.1260473.
- Zaidi M, Inzerillo AM, Moonga BS, Bevis PJ, Huang CL. Forty years of calcitonin– where are we now? A tribute to the work of lain Macintyre, FRS. *Bone* 30: 655–663, 2002. doi:10.1016/S8756-3282(02)00688-9.
- Zhang Z, Neff L, Bothwell AL, Baron R, Horne WC. Calcitonin induces dephosphorylation of Pyk2 and phosphorylation of focal adhesion kinase in osteoclasts. *Bone* 31: 359–365, 2002. doi:10.1016/S8756-3282(02)00834-7.
- 232. Zhou R, Yuan Z, Liu J, Liu J. Calcitonin gene-related peptide promotes the expression of osteoblastic genes and activates the WNT signal transduction pathway in bone marrow stromal stem cells. *Mol Med Rep* 13: 4689–4696, 2016. doi:10.3892/mmr.2016.5117.