



Published in final edited form as:

*Trends Endocrinol Metab.* 2016 October ; 27(10): 706–718. doi:10.1016/j.tem.2016.06.001.

## The Ca<sup>2+</sup>/Calmodulin/CaMKK2 Axis: Nature's Metabolic CaMshaft

Kathrina L. Marcelo<sup>1</sup>, Anthony R. Means<sup>1,2,\*\*,\*</sup>, and Brian York<sup>1,2,\*\*,\*</sup>

<sup>1</sup>Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, TX

<sup>2</sup>Dan L. Duncan Cancer Center, Baylor College of Medicine, Houston, TX

### Abstract

Calcium (Ca<sup>2+</sup>) is an essential ligand that binds its primary intracellular receptor Calmodulin (CaM) to trigger a variety of downstream processes and pathways. Central to the actions of Ca<sup>2+</sup>/CaM is the activation of a highly conserved Ca<sup>2+</sup>/CaM kinase (CaMK) cascade, which amplifies Ca<sup>2+</sup> signals through a series of subsequent phosphorylation events. Proper regulation of Ca<sup>2+</sup> flux is necessary for whole-body metabolism and disruption of Ca<sup>2+</sup> homeostasis has been linked to a variety of metabolic diseases. Herein, we provide a synthesis of recent advances that highlight the roles of the Ca<sup>2+</sup>/CaM kinase axis in key metabolic tissues. An appreciation of this information is critical in order to understand the mechanisms by which Ca<sup>2+</sup>/CaM-dependent signaling contributes to metabolic homeostasis and disease.

### Keywords

Calcium; Calmodulin; CaMKK2; CaMKIV; AMPK

### Calmodulin is the intracellular receptor for Ca<sup>2+</sup>

Calcium (Ca<sup>2+</sup>) serves as one of the most ubiquitous and critical second messengers that regulate a plethora of downstream cellular signaling events. In response to an external stimulus, Ca<sup>2+</sup> release from intracellular stores or influx from the extracellular space through ion channels, leads to its binding to target proteins and subsequent regulation of their activity. Ca<sup>2+</sup> is a crucial regulatory ligand, and among its many binding partners, calmodulin (CaM) serves as its primary intracellular receptor for controlling essentially every aspect of cellular biology (Box 1). The Ca<sup>2+</sup>/CaM complex controls the activity of over 120 enzymes and proteins, including but not limited to transcription factors, ion channels and pumps, phosphatases, and kinases [1].

\*\*Correspondence: Brian York, Ph.D., Department of Molecular and Cellular Biology, Baylor College of Medicine, M810b DeBakey Building, One Baylor Plaza, Houston, TX 77030, Phone: 713-798-4178, york@bcm.edu. Anthony R. Means, Ph.D., Department of Molecular and Cellular Biology, Baylor College of Medicine, M829 DeBakey Building, One Baylor Plaza, Houston, TX 77030, Phone: 713-798-4178, armeans@bcm.edu.

\*These authors contributed equally to this work

**Disclosure Statement:** The authors have nothing to disclose.

**Publisher's Disclaimer:** This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

**Box 1****The Ca<sup>2+</sup>/CaM Structure-Function Relationship**

Since Ca<sup>2+</sup> is such a pervasive signal, it is essential for its receptor CaM to have structural plasticity to dynamically regulate the panoply of Ca<sup>2+</sup>-dependent target proteins and downstream pathways. Over the nearly forty years of research on CaM, there have been a number of excellent reviews that summarize details of its structure and physicochemical properties [1–3]. CaM is a small, highly conserved, dumbbell-shaped protein expressed by all eukaryotic cells. In humans, CaM is encoded by three distinct genes (*CALM1*, *CALM2*, *CALM3*), each having unique alternative regulation, tissue-specificity, and alternative splicing, yet surprisingly, producing identical proteins [4]. Structurally, each of its two lobes (N- and C-terminus) consists of a pair of EF hand motifs, each of which can bind two Ca<sup>2+</sup> ions. In the absence of Ca<sup>2+</sup>, the N-terminus of CaM adopts a closed conformation while the C-terminal lobe remains in a semi-open conformation with a slightly exposed hydrophobic patch. A rise in intracellular Ca<sup>2+</sup> leads to saturation of Ca<sup>2+</sup> binding to CaM, which induces remodeling of both lobes into a more open conformation that exposes hydrophobic patches that promote its stable interaction with target proteins [2,3]. A helix connecting the two lobular domains of CaM also confers incredible flexibility to the molecule, allowing the Ca<sup>2+</sup>/CaM complex to adopt an infinite variety of conformations, each of which is unique for recognition and regulation of a specific binding partner [1].

**A key Ca<sup>2+</sup>/CaM-dependent kinase cascade emerges**

While the majority of early CaM studies were focused on characterizing its sequence, structure and cellular functions (Box 1), subsequent efforts resulted in the discovery and characterization of key Ca<sup>2+</sup>/CaM downstream targets (Figure I in Box 2). The compendium of Ca<sup>2+</sup>/CaM target proteins has been stratified into six (6) classes according to the nature of their interaction with the ligand-receptor complex and its effect on their activity [1]. In this review we primarily focus on class E and F targets, which are activated upon binding Ca<sup>2+</sup>/CaM, and whose regulation and activities collectively define the Ca<sup>2+</sup>/CaM-dependent kinase cascade (Fig. 1). In response to external stimuli, pulsatile increases in intracellular Ca<sup>2+</sup> are sensed by CaM and the Ca<sup>2+</sup>/CaM complex adopts an increased affinity for class E and F target proteins [1]. However, class E targets, which include Ca<sup>2+</sup>/CaM-dependent kinases (CaMK) I and IV, require further phosphorylation by class F proteins Ca<sup>2+</sup>/CaM-dependent kinase kinase 1 and 2 (CaMKK1, CaMKK2). Activated CaMKK2 also phosphorylates the  $\alpha$  subunit of AMP-activated protein kinase (AMPK), and forms a multimeric protein complex comprised of Ca<sup>2+</sup>/CaM, CaMKK2, and the AMPK $\alpha$  and  $\beta$  subunits [5] (Fig. 1). Importantly, the regulatory gamma ( $\gamma$ ) subunit of AMPK that senses changes in the ATP:AMP ratio is not required for this interaction, so that activation of AMPK in this scenario is unique to fluctuations in intracellular Ca<sup>2+</sup>. Moreover, the CaMKK2 and AMPK $\alpha$  interaction occurs via their kinase domains, and CaMKK2 must exist in its activated conformation to interact with AMPK $\alpha$ , but not with its other kinase substrates, CaMKI and CaMKIV [6]. In this manner, full activation of the Ca<sup>2+</sup>/CaM-

dependent kinase cascade effector molecules culminates in appropriate regulation of their cellular functions.

## Box 2

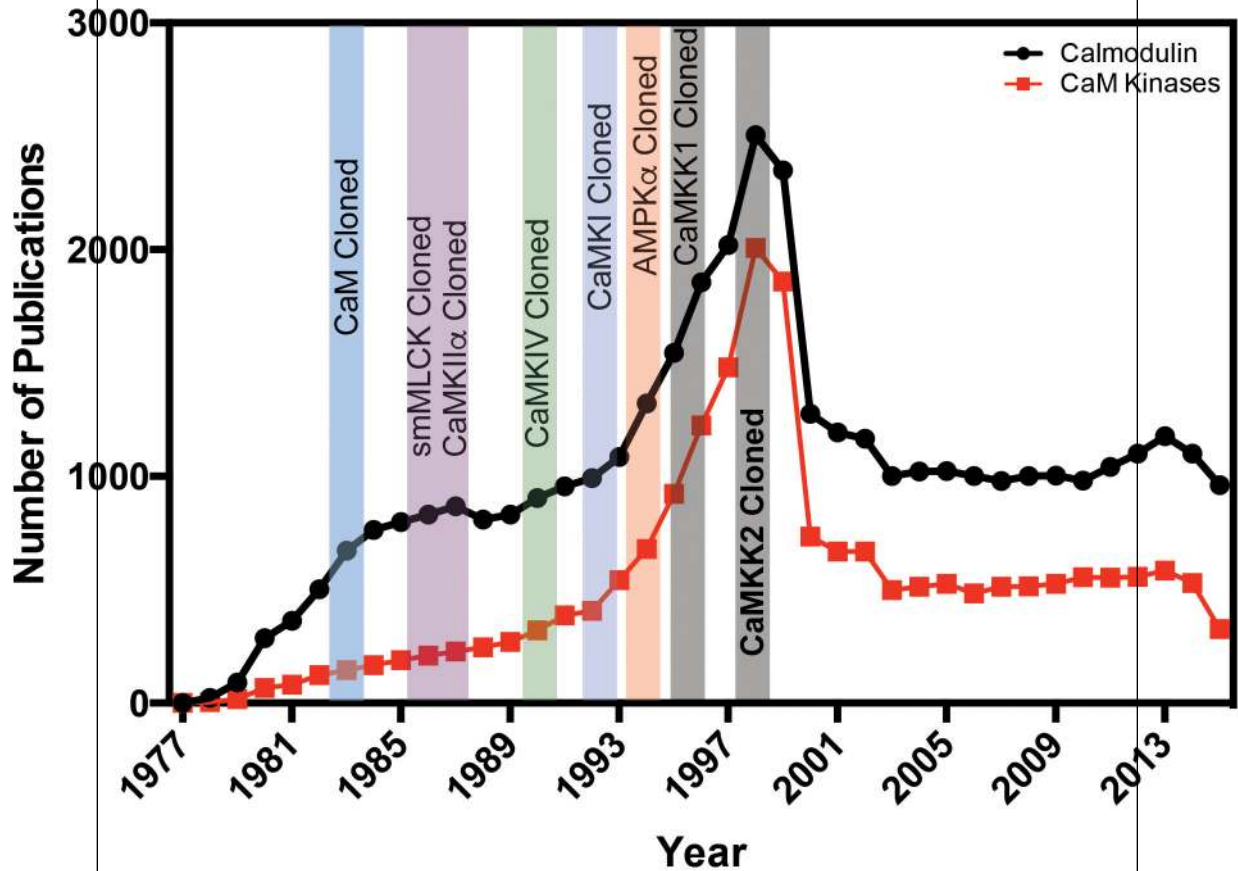
### CaM Kinase Historical Overview

Since calcium ( $\text{Ca}^{2+}$ ) is the 5<sup>th</sup> most abundant element in the earth's crust and the human body, its emergence as a ubiquitous and pervasive second messenger was unpredicted. While  $\text{Ca}^{2+}$  exists at mM concentrations in biological fluids, a similar elevation of free  $\text{Ca}^{2+}$  in any living cell would be catastrophic. Therefore, free  $\text{Ca}^{2+}$  within cells is tightly regulated in the nM range, and cells have adapted multiple mechanisms to accomplish this, including the evolutionary emergence of numerous  $\text{Ca}^{2+}$  pumps and channels associated with plasma, mitochondrial and intracellular membranes as well as a plethora of  $\text{Ca}^{2+}$ -binding proteins [56]. Many of these proteins function to buffer, sense or monitor the frequency, amplitude or diffusion of  $\text{Ca}^{2+}$  within the cell. However, only one  $\text{Ca}^{2+}$ -binding protein, calmodulin (CaM), is essential, ubiquitously expressed and serves as the primary intracellular receptor for  $\text{Ca}^{2+}$  thereby mediating most of the ion's second messenger functions [1].

CaM was originally identified in snake venom as an activator of cyclic nucleotide phosphodiesterase (PDE) [57] and subsequently shown to require [59] and bind  $\text{Ca}^{2+}$  [60]. Shortly thereafter, the protein was independently "discovered" by several investigators working on different biological systems and given different names. The name calmodulin was proposed to investigators in this fledgling field in 1978 as a unifying name and adopted within a year or two. For a review on its early history please see [2,61]. Since its discovery, CaM has been the subject of almost 32,000 papers cited in PubMed (Figure I in Box 2). Over 120 CaM-binding partners have been identified and include members of virtually every major class of cellular proteins.

Early roles for CaM in endocrinology and metabolism were demonstrated by its participation in the action of FSH in testicular Sertoli cells by regulating a cyclic AMP PDE [62] and as an integral, regulatory subunit of the heterotetrameric metabolic enzyme phosphorylase kinase (a complex also regulated by cAMP) [63], respectively. These early findings presaged the subsequent discovery that, in addition to cAMP-mediated pathways, CaM regulates a variety of other intracellular signaling cascades involving nitric oxide, soluble phospholipids, nuclear receptors and membrane receptors/ion channels [56]. Other CaM targets include several  $\text{Ca}^{2+}$  pumps/channels, the protein phosphatase, calcineurin, and a number of CaM-dependent protein kinases (CaMK) [64]. While the early CaM field was driven by studies on CaM (Figure I in Box 2) [65] the large upswing in CaM publications mirrors the cloning and subsequent mechanistic studies on the CaMKs [64,66,67]. Biochemical studies of CaMKIV [68] and CaMKI [69], revealed that each required not only CaM binding, but also phosphorylation of a Thr on the activation loop to exhibit maximal activity. These findings led to the discovery of two upstream CaM kinase kinases (CaMKK) CaMKK1 [70] and CaMKK2 [71]. The requirement for one protein kinase to regulate another was reminiscent of the MAP kinase cascade and by analogy named the CaM kinase cascade. CaMKK2 was shown to

activate not only CaMKI and CaMKIV but also the master metabolic protein kinase, AMPK [72]. Thus, CaMKK2 has emerged as the primary CaMK participating in regulation of metabolic homeostasis.



**Figure I, Box 2. Timeline of CaM- and CaMK-related publications**

Graphic representation of the historical increase in the number of PubMed® publications on CaM (black line) and the Ca<sup>2+</sup>/CaM-dependent kinases (CaMK) (red line).

Discoveries highlighted include cloning of the genes for the following cascade components: CaM - 1981 [65]; smooth muscle myosin light chain kinase (smMLCK) - 1986 [66]; CaM kinase II α (CaMKIIα) - 1987 [67]; CaMKIV - 1991 [68]; CaMKI - 1993 [69]; AMP-activated protein kinase α (AMPKα) - 1995 [72]; CaMK kinase 1 (CaMKK1) - 1995 [70]; CaMKK2 - 1997 [71].

### Tissue-specific metabolic regulation by the Ca<sup>2+</sup>/CaM-dependent kinase cascade

Communication among different metabolic tissues and organs ensures proper energy flow and balanced caloric intake and utilization to maintain organismal energy homeostasis. Ca<sup>2+</sup>/CaM signaling is critical for mediating the downstream effects of hormones, metabolites, inflammatory agents and neuroendocrine signals that coordinate tissue crosstalk

to balance the energy equation. Epidemiological studies have demonstrated that several hallmarks of metabolic syndrome, including obesity and diabetes, are associated with abnormal serum  $\text{Ca}^{2+}$  levels [7,8]. As discussed previously, the  $\text{Ca}^{2+}/\text{CaM}$  complex mediates the amplification of  $\text{Ca}^{2+}$  signals through activation of a tightly regulated set of protein kinases, which themselves have critical functions in key metabolic tissues. An overview of the recent developments in  $\text{Ca}^{2+}/\text{CaM}$ -dependent kinase cascade research as they pertain to metabolic homeostasis is discussed below and summarized in Fig. 2. Since virtually all of the physiologic actions relevant to metabolism illustrated in Fig. 2 are common to cascades using CaMKK2 as the most upstream kinase, this review will focus exclusively on these pathways.

## Brain

The brain consumes nearly 70% of total body glucose and its normal functions depend on an optimal range of glucose levels. As part of its role in providing centralized control over the entire organism, the vertebrate brain, particularly the hypothalamus, is responsible for integrating metabolic signals from peripheral organs and tissues [9]. CaMKK2 is highly expressed in the arcuate nucleus (ARC) of the hypothalamus and plays a fundamental role in whole body energy homeostasis [10]. In the fasted state, the orexigenic peptide ghrelin is released from ghrelinergic cells of the intestine and binds its cognate receptor (GHSR, or growth hormone secretagogue receptor) on presynaptic neurons that express the orexigenic hormones Neuropeptide Y (NPY) and Agouti-Related Protein (AgRP), resulting in release of  $\text{Ca}^{2+}$  from intracellular stores. Orexin A, an appetite-stimulating neuropeptide, acts via voltage-gated L-type  $\text{Ca}^{2+}$  channels to increase intracellular  $\text{Ca}^{2+}$  in NPY-producing hypothalamic neurons [11]. Together these  $\text{Ca}^{2+}$  signals lead to activation of CaMKK2 and phosphorylation of AMPK, which are necessary for regulating the hypothalamic expression of NPY and AgRP. These findings provide a mechanism for how two orexigenic signals, ghrelin and orexin A, function through CaMKK2 and its target AMPK in specific cell populations of the hypothalamus to control appetite and energy homeostasis [10, 11]. In fact, mice devoid of *CaMKK2* fail to respond to ghrelin, have reduced *NPY* expression, and consequently consume less food when compared to pair-fed wild-type (WT) mice. These findings partially explain why *CaMKK2*-null mice are protected against high fat diet (HFD)-induced weight gain, insulin resistance, and glucose intolerance [10].

Brain-derived serotonin is a neurotransmitter that modulates appetite and energy expenditure upon binding Htr1a and Htr2b (5-hydroxytryptamine (serotonin) receptor 1A and 2B, respectively) receptors on hypothalamic ARC neurons, triggering cAMP response element-binding protein (CREB)-mediated regulation of genes that favor feeding [12,13]. Conversely, serotonin binding to Htr2c receptors on ventromedial hypothalamic (VMH) neurons induces CaMKIV phosphorylation via CaMKK2 [12]. Active CaMKIV phosphorylates CREB, which in this case promotes expression of genes that decrease sympathetic tone [14]. Since the sympathetic nervous system inhibits osteoblast proliferation and promotes bone resorption by osteoclasts [15], *CaMKK2* deletion in VMH neurons decreases peripheral bone mass accrual [14]. A recent study described a dual regulatory role by which CaMKK2 activation of CaMKIV prevents premature differentiation of mesenchymal stem cells into bone matrix-producing osteoblasts, while CaMKK2 activity

leads to CREB phosphorylation and subsequent differentiation of osteoclasts. The increase in trabecular bone mass and osteoblast number and the accompanying decrease in osteoclasts were therefore explained by the shift from catabolic to anabolic pathways in *CaMKK2*-deficient mice during bone remodeling and homeostasis [16]. The phenotypic differences observed in bone mass accrual arising from *CaMKK2* loss are best explained by its cell intrinsic functions. The serotonin studies utilized VMH-specific *CaMKK2* deletion [14], leaving *CaMKK2* function intact in osteoclast and osteoblast progenitors. In whole body *CaMKK2* ablation studies [16], the cell autonomous role of *CaMKK2* in the osteoblast and osteoclast progenitors has perhaps shifted the balance towards osteoblast differentiation to such an extent that the contribution of sympathetic tone on bone mass accrual becomes secondary or even negligible.

Higher order brain functions are essential for satisfying the desire to feed—the brain facilitates behavioral adaptations required for food accrual while generating a memory of the events for the next round of feeding [17,18]. The  $\text{Ca}^{2+}$ /CaM kinase cascade up-regulates and activates CREB to induce downstream gene expression programs necessary for long-term memory (LTM) formation and different types of learning [19–22]. *CaMKK2* is required during spatial memory formation and for CREB up-regulation and activation but not for contextual fear LTM [19,23]. Downstream of *CaMKK2* action lies *CaMKIV*, which is necessary for contextual LTM [21]. *CaMKIV*'s role during spatial learning remains largely undefined. Since *CaMKIV* is activated after LTP at hippocampal CA1 synapses [24], and *CaMKK2* is indispensable during hippocampal memory formation [23], it is likely that *CaMKIV* acts downstream of *CaMKK2* during spatial learning to establish late CA1 LTP.

In addition to learning and memory, metabolic syndrome and cerebral ischemia are both associated with impaired energy utilization and dysregulated  $\text{Ca}^{2+}$  signaling [25]. It is not surprising that metabolic syndrome is a major risk factor for stroke, which also includes cognitive decline as a persistent pathology [26].  $\text{Ca}^{2+}$ /CaM-dependent kinases are associated with endogenous protective functions against cerebrovascular damage. *CaMKK2*- and *CaMKIV*-null mice present aggravated brain injuries and behavioral deficits following middle cerebral artery occlusion (MCAO) [27,28]. *CaMKIV*-knockout mice presented more severe ischemic injuries and displayed higher mortality rates than *CaMKK2*<sup>-/-</sup> mice, suggesting that perhaps *CaMKIV* is downstream of *CaMKK2*. Phosphorylation of *CaMKIV* leads to its nuclear translocation and subsequent phosphorylation of histone deacetylase 4 (HDAC4), which is exported from the nucleus, allowing the transcription of CREB, a neuronal survival factor [27,28]. In a similar fashion, both *CaMKK2* and *CaMKIV* are instrumental for the proliferation and survival of cerebellar granule cell neurons, by phosphorylating CREB [29]. *CaMKK2*/*CaMKIV*-regulated CREB expression and activation [29] promotes transcription of pro-survival genes, including *Bdnf* (brain derived neurotrophic factor) during granule cell development [29] and *Bcl2* during ischemia [27,28].

Interestingly, both *CaMKK2* and *CaMKIV* preserve the integrity of the blood-brain barrier (BBB). MCAO-treated *CaMKK2*<sup>-/-</sup> and *CaMKIV*<sup>-/-</sup> mice displayed loss of BBB collagen IV protein and reduced expression of extracellular matrix-degrading,  $\text{Ca}^{2+}$ -dependent matrix metalloproteinases (MMP) 2/9 [27,28]. It was speculated that *CaMKK2* confers protection to endothelial cells that primarily constitute the BBB proper [28]. *CaMKK2* phosphorylates

both AMPK and SIRT1, a class III histone deacetylase, in mouse aortic endothelium. This dual activation induces anti-inflammatory events in the cytoplasm and nuclear transcription of antioxidants [30]. The mechanism involving the  $\text{Ca}^{2+}$ /CaM kinase cascade that confers endothelial cell homeostasis in the BBB remains to be fully elucidated, although ischemic models strongly suggest it involves CaMKIV activation by CaMKK2 [27,28] in contrast to that observed in the aortic endothelium that requires AMPK [30].

Distinct from CaMKIV, the role of AMPK is controversial in neuronal ischemic injury. Although stroke induces increased phosphorylation of AMPK, no differences were observed in activated AMPK levels between sham-treated mice and those treated with a selective CaMKK2 inhibitor (STO-609) prior to MCAO [27]. These findings suggest that AMPK activation during stroke is independent of CaMKK2 activity, and that the neuroprotective effects of CaMKK2 are not mediated via AMPK. As reviewed in Ronnett et al. [31], numerous studies provide conflicting claims regarding the benefits and disadvantages of AMPK activation following stroke. However, these reports failed to specifically examine AMPK phosphorylation by CaMKK2. A recent study on neonatal hypoxia-ischemic brain injury demonstrated the pathological activation of the CaMKK2-AMPK axis in neurons [32]. Interestingly, while inactivation of AMPK during the course of ischemic injury promotes neuronal survival, inhibiting AMPK prior to the insult sensitizes the neurons and exacerbates cell death [31]. Collectively, these studies highlight the importance of external inputs, cell types and developmental events that in part reflect the metabolic contributions of the  $\text{Ca}^{2+}$ /CaM kinase cascade in the brain and the choice of effector molecules that confer cell-specific responses that are impacted by perturbations in metabolic homeostasis.

## Liver

While hypothalamic CaMKK2 plays a central role in controlling whole body energy homeostasis [10], little was known about its function in peripheral metabolic tissues. In a study by Anderson et al. [33], WT mice were pair fed to match the consumption level of *CaMKK2*-null mice maintained on HFD, given that *CaMKK2* deletion reduces appetite [10]. Although pair feeding marginally slowed weight gain in WT mice, it failed to improve glucose tolerance [33]. Reduced food intake therefore is insufficient to fully account for the improved metabolic status of *CaMKK2*<sup>-/-</sup> mice, suggesting a role for  $\text{Ca}^{2+}$ /CaM signaling via CaMKK2 in peripheral metabolic tissues.

During prolonged starvation, the liver provides glucose and ketone bodies, both of which are energy sources for the brain [34]. The liver is the central metabolic organ responsible for maintenance of plasma glucose levels and regulating the level of metabolites in blood, including amino acids and fatty acids (FAs) packaged as very low-density lipoproteins (VLDL) for transport [34]. Hepatocytes, which constitute the parenchymal cells of the liver, are essential for its metabolic capacity and express CaMKK2. Acute deletion of hepatic *CaMKK2* is sufficient to lower blood glucose and improve glucose tolerance in fed and fasted mice [33]. Primary hepatocytes (PH) isolated from *CaMKK2*-null mice produced less glucose, which was accompanied by reduced expression of the gluconeogenic genes glucose-6-phosphatase (*G6pc*) and phosphoenolpyruvate carboxykinase (*PEPCK*), owing to the cell autonomous role of CaMKK2 in phosphorylating hepatic HDAC5 [33]. Similar to

HDAC4 function in neurons, HDAC5 phosphorylation relieves transcriptional repression of peroxisome proliferator-activated receptor  $\gamma$  coactivator 1- $\alpha$  (PGC1- $\alpha$ ) and activates the expression of gluconeogenesis-related genes [33]. *CaMKK2*<sup>-/-</sup> mice also displayed higher acyl-carnitine levels compared to those of WT, as the mice progressed from the fed to fasted states. *In vitro* studies confirmed that *CaMKK2* ablation increased both lipogenesis and lipid oxidation, concomitant with a decrease in glucose utilization in the liver, both of which are consistent with a switch from glucose to fat metabolism [33]. The increased FA oxidation observed in *CaMKK2*-knockout mice could also be a compensatory response to the overall reduced glucose utilization in these animals, which would account for their steatotic livers observed in response to prolonged fasting that are unaccompanied by other pathophysiological symptoms [33]. Thus, the contribution of Ca<sup>2+</sup>/CaM signaling through CaMKK2 in regulating energy homeostasis not only involves hypothalamic control of appetite [10], but also a coordinated function in peripheral tissues such as the liver for maintenance of basal glucose levels by controlling hepatic glucose production [33] (Fig. 3).

Hepatocellular carcinoma (HCC) is among the leading causes of cancer-related deaths worldwide and, particularly in the U.S., its rise in incidence has been directly linked to the onset of adult obesity [38]. Ca<sup>2+</sup>, as previously described, has been epidemiologically linked to obesity, and several studies have also implicated its role in HCC. *CaMKK2* expression is markedly up-regulated in HCC compared to matched normal tissue and similarly in several human and murine hepatic cancer cell lines [35]. Survival of HCC patients also inversely correlates with high *CaMKK2* expression. Consistent with these findings, knockdown or pharmacological inhibition of CaMKK2 greatly reduced colony formation and proliferation of liver cancer cells, which was rescued by overexpression of exogenous *CaMKK2*, but not kinase mutant versions of the protein. CaMKK2 activity is therefore instrumental in the cell growth machinery of liver cancer cells [35]. CaMKK2 contributes to hepatic cancer cell growth primarily through CaMKIV, which together with CaMKK2 provides a scaffold for the coordinated regulation of the mammalian target of rapamycin/ribosomal protein S6 kinase, 70 kDa (mTOR/S6K) proteins that respond to nutrient status to drive protein synthesis [35] (Fig. 3). *CaMKIV* has been previously shown to be up-regulated in human and rat HCC [34]. Ablation of *CaMKIV* in liver cancer cell lines phenocopies *CaMKK2* deletion, and although rescue experiments with *CaMKK2* re-expression restored protein synthesis and proliferation defects in both *CaMKK2*- or *CaMKIV*-knockdown cells, *CaMKIV* overexpression was insufficient to completely rescue the loss of CaMKK2 [35].

*In vivo* xenograft studies revealed that loss of *CaMKK2* in PHM1 cells (p53-null/Myc-transformed liver cancer cells) subcutaneously injected into nude mice retarded tumor growth and reduced tumor volume compared to WT PHM1-derived tumors [35]. Furthermore, pharmacological treatment of nude mice harboring WT PHM1-derived tumors with STO-609 was sufficient to suppress tumor outgrowth. In a diethylnitrosamine (DEN) murine model for hepatic cancer, STO-609 treatment resulted in a 20% reduction in tumor volume while vehicle-treated mice showed a 50% increase in tumor volume as measured via PET/CT imaging [35]. CaMKK2 activity is required for aggressive liver cancer growth, and its inhibition is sufficient to block the downstream signaling machinery, leading to attenuation of tumor growth *in vivo*. Given that *CaMKK2*<sup>-/-</sup> mice are resistant to HFD-induced obesity [33], a major risk factor for HCC, and are protected from LPS-induced



fulminant hepatitis [37], it is not surprising that CaMKK2 significantly contributes to cancer cell proliferation. Perhaps even more significant is the potential for targeting the CaMKK2/CaMKIV signaling axis by using STO-609 as a therapeutic recourse. Several studies have focused on Ca<sup>2+</sup> signaling components, and CaMKK2 in particular, to explore new therapeutic avenues for combatting fatty liver disease and cancer [39,40]. Baicalin, a polyphenolic compound, decreased fat, liver, and whole body weights in HFD-fed mice by inhibiting the CaMKK2-AMPK pathway [39], while bortezomib, a proteasome inhibitor and multiple myeloma therapeutic agent, activated CaMKK2 protein synthesis in human breast cancer cell lines [40]. Only time, further studies, and eventually human clinical trials will reveal the feasibility of such a strategy.

## Adipose

Following feeding stimulated by orexigenic signals in the brain, glucose and VLDL produced by the liver are converted into triacylglycerides (TAG) in adipose tissue. During fasting or exercise, TAGs are hydrolyzed into glycerol and free fatty acids (FFAs), which are then transported to the liver for use during gluconeogenesis. Adipocytes therefore largely function to balance lipid storage and release, to coincide with the feeding to fasting transition [41]. Obesity, metabolic syndrome, and their associated complications are often characterized by adipose tissue dysfunction. These dysfunctions include increased macrophage recruitment, abnormal microvasculature, and perturbed adipokine production [42]. Disrupted Ca<sup>2+</sup> signaling accompanying obesity is also associated with attenuated differentiation of pre-adipocyte progenitors [43]. *CaMKK2*-null mice display increased adiposity when fed standard chow due to both increases in size and number of adipocytes [44]. *CaMKK2* expression is suppressed once pre-adipocytes differentiate into mature adipocytes, while *in vitro* ablation or pharmacological inhibition of CaMKK2 increased adipogenesis [44]. Taken together, these findings suggested that CaMKK2 is essential during this differentiation process. Increased intracellular Ca<sup>2+</sup> leads to CaMKK2 activation of AMPK in pre-adipocytes, which together maintain expression of preadipocyte factor-1 (*Pref-1*). *Pref-1*, in turn, maintains expression of *Sox9* through phosphorylation of ERK1/2. *Sox9* inhibits transcription of early adipogenic factors such as *C/EBPβ* and *C/EBPδ* and eventually blunts other downstream events that maintain a differentiated adipocyte state [44]. CaMKK2 and AMPK therefore play cell autonomous roles in pre-adipocytes to maintain an undifferentiated state [44], similar to their roles in bone remodeling [16], cerebellar granule cell development and granulopoiesis [45]. Thus, Ca<sup>2+</sup>/CaM signaling consistently functions within several progenitor cell populations to maintain their undifferentiated, proliferative state, and employs a cell-autonomous and unique mechanism involving CaMKK2.

Although *CaMKK2* is not expressed in mature adipocytes [44], studies have reported such expression in adipose exposed to exogenous hormonal stimulation [46,47]. In response to thyroid hormone, increased Ca<sup>2+</sup> concentration and AMPK activation via CaMKK2 was observed in adipocytes hypomorphic for *Lkb1*, an alternative upstream kinase for AMPK activation [47]. Meanwhile, CaMKK2 reportedly regulates AMPK action in response to glucagon, which then phosphorylates acetyl-CoA carboxylase 1 (ACC1) in adipocytes. Glucagon-induced phosphorylation of ACC1 attenuates its activity during FA synthesis [46]. Since increased intracellular Ca<sup>2+</sup> within adipocytes was not induced by glucagon, the

mechanism for AMPK activation remains to be clarified and reconciled with previous findings. One possibility is that glucagon up-regulates *CaMKK2* expression, as this gene is a stimulus-regulated event in a variety of cell types [35,48–50].

## Pancreas

Two key regulators of liver metabolism are the pancreatic hormones glucagon and insulin. Although the balance between  $\text{Ca}^{2+}$  influx and efflux critical to the regulation of insulin production by pancreatic beta ( $\beta$ ) cells is well appreciated [51], the molecular underpinnings that contribute to this process are poorly understood. Recent reports demonstrate the key roles of  $\text{Ca}^{2+}$  signaling and  $\text{Ca}^{2+}$ /CaM kinase cascade components in insulin production and secretion. *CaMKK2* expression is largely correlated with  $\beta$  cells and only marginally with glucagon-producing  $\alpha$  cells. Moreover, CaMKK2 is not required for maintenance of islet structure,  $\beta$  cell polarity, or hormone production [52]. In fact, loss of *CaMKK2* led to increased insulin content per islet and increased pancreatic islet insulin secretion. Therefore, it appears that CaMKK2 is an important component of the  $\text{Ca}^{2+}$ -mediated response to glucose signaling in  $\beta$  cells, acting as a molecular rheostat to regulate insulin secretion and perhaps production. The activity of CaMKK2 facilitates pancreatic overproduction and over-secretion of insulin during chronic metabolic stress such as HFD feeding, which contributes to insulin resistance and glucose intolerance in WT mice. *CaMKK2*-null mice fed a standard chow have plasma insulin levels that are marginally higher than WT mice, but still within normal physiological levels. Chronic metabolic stress fails to further induce insulin production or secretion because CaMKK2, which is required for these processes, is absent [52]. Combined with its roles in controlling satiety (brain) [10], metabolic conversion (liver) [33] and storage (adipose) [44], these newly appreciated functions of CaMKK2 in the pancreas also fractionally contribute to the protection from diet-induced glucose intolerance and insulin resistance observed in *CaMKK2*<sup>-/-</sup> mice.

The exact mechanism by which CaMKK2 regulates insulin secretion and/or production remains to be explored. Perhaps loss of *CaMKK2* results in increased secretory granules and/or increased number of granules poised for secretion at the plasma membrane. Other reports suggest that following an increase in intracellular  $\text{Ca}^{2+}$ , CaMKK2 acts through AMPK to regulate insulin secretion and  $\beta$  cell survival in response to either uridine diphosphate [53] or to adipose tissue derived leptin [54]. Nevertheless, the mechanism by which CaMKK2 regulates insulin secretion is an area ripe for investigation and could even be exploited for treatment of T2DM.

CaMKK2 is also involved in lineage programming of pancreatic  $\alpha$  to  $\beta$  cells [55], which is an attractive therapeutic strategy for type 1 diabetes (T1D) that is characterized by loss of  $\beta$  cell mass. Analysis of transcriptomic, proteomic, and phosphoproteomic profiles from  $\alpha$  and  $\beta$  cells found that *CaMKK2* had decreased expression and phosphorylation in  $\beta$  cells compared to  $\alpha$  cells [55]. *In vitro* knockdown of *CaMKK2* in  $\alpha$  cells or treatment with STO-609 led to the increased expression of insulin and other key  $\beta$  cell markers, although glucagon expression was also increased. These data present a strong potential for inhibition of this kinase during  $\alpha$ -to- $\beta$  transdifferentiation [55]. Pancreatic  $\alpha$  and  $\beta$  cells derive from the same endocrine progenitor during development. Knowing that CaMKK2 functions in

cell fate decisions in cerebellar granule cells, adipose [44], bone [16], and granulocyte-monocyte progenitors [45], it is likely that CaMKK2 also plays a role during the differentiation of these pancreatic cell types. *CaMKK2*-null mice had pancreatic islets that appeared morphologically normal and retained glucagon-expressing cells that were presumably  $\alpha$  cells [52]. However, the expression pattern of other definitive  $\alpha$  and  $\beta$  cells were not particularly scored [52], and would be an interesting follow up to investigate the role of CaMKK2 in pancreatic islet cell differentiation. It is plausible that deletion of *CaMKK2* in endocrine progenitors either endowed  $\alpha$  cells the ability to produce insulin and/or skewed the differentiation towards  $\beta$  cell progenitors.

## Concluding Remarks and Future Perspectives

In this review, we provide a synthesis of the most recent and critical reports highlighting the roles of the  $\text{Ca}^{2+}$ /CaM-dependent kinase cascade in organismal metabolic regulation. The kinase cascade triggered by the ligand-receptor binding of  $\text{Ca}^{2+}$  to CaM is a tightly regulated mechanism ubiquitous to every key metabolic tissue, which confers pleiotropic actions ranging from regulation of food intake to energy expenditure and fuel storage. Aside from rekindling these previously underappreciated concepts, this review emphasizes the potential of inhibiting members of this kinase cascade for treatment of metabolic diseases such as obesity, diabetes, and even cancer. Of the kinases in this  $\text{Ca}^{2+}$ /CaM-dependent pathway, CaMKK2 currently appears to be the most attractive therapeutic target. Inhibition of CaMKK2 in mice results in attenuated food intake, promoted weight loss, and generally improved whole body glucose homeostasis [10,33]. CaMKK2 also inhibits progenitor differentiation into mature adipocytes [44]. Investigation into the action of CaMKK2 in pancreatic cell types suggests a prophylactic role against developing insulin resistance [52]. The potential for targeting this kinase to induce  $\alpha$ -to- $\beta$  cell transdifferentiation could impact T1D treatment strategies [55]. Also, the cell autonomous functions of CaMKK2 in macrophages [37], which play key roles in a variety of metabolic tissues, offers an alternative strategy for attenuating the pleiotropic actions of this kinase by pharmacological inhibition. Finally, *in vivo* administration of STO-609 to mice harboring hepatic tumors significantly regressed tumor burden [35]. STO-609 is currently the most selective and potent pharmacologic inhibitor against CaMKK2. The limited availability of information regarding the pharmacokinetic properties and toxicity of STO-609 as well as the optimal method of packaging and administration of the inhibitor are all issues that must be addressed before one can consider STO-609 as a candidate therapeutic agent for pathologies arising from metabolic dysfunction.

Despite all the information summarized in this review regarding the critical roles of  $\text{Ca}^{2+}$ /CaM-dependent kinase cascade components in whole body energy balance, a number of key questions remain unanswered (see Outstanding Questions). CaMKK2's function remains largely undefined in a variety of metabolic tissues, such as brown adipose tissue, cardiac and skeletal muscle, spleen, and kidney. Within skeletal muscle in particular, although CaMKK2 does not appear to be expressed in this tissue, congenic ablation of *CaMKK2* results in overt muscle hypertrophy. These data suggest CaMKK2 might regulate myogenic humoral factors, which represents yet another area that is primed for further investigation.

### Outstanding Questions Box

What are the roles of CaMKK2 signaling in unexplored metabolic tissues (i.e. brown adipose tissue, skeletal and cardiac muscle, spleen and kidney)?

What signals (i.e. hormones, metabolites) control the gene expression of members of the Ca<sup>2+</sup>/CaM/CaMK axis (i.e. *Calm*, *CaMKK2*, *AMPK*, *CaMKIV*) in response to physiologic or pathophysiological conditions? Is the expression of these pathway components coordinately regulated?

What are the cell-specific functions of the Ca<sup>2+</sup>/CaM-dependent kinase pathway components? Are the known functions of Ca<sup>2+</sup>/CaM-dependent kinases attributed to common cell types that are ubiquitously present in all metabolic organs (i.e. macrophages)?

What is the subcellular localization of CaMKK2? Is CaMKK2 bound or tethered to intracellular membranes? Does CaMKK2 shuttle between the cytoplasm and nucleus? What mechanisms control the intracellular trafficking and/or retention of CaMKK2?

Can alternate and/or additional targets/substrates for CaMKK2 be identified? If so, what are their cell- or tissue-specific functions downstream of Ca<sup>2+</sup>/CaM/CaMKK2 action?

Does CaMKK2 contribute to metabolic reprogramming by controlling fuel substrate utilization?

Aside from consensus binding sequences within the 5' untranslated region of the human *CaMKK2* gene for the transcription factors Ikaros, Runx1, and GATA1 [5], very little is known about regulation of *CaMKK2* and *CaMKIV* gene expression. How are the *CaMKK2* and *CaMKIV* genes controlled? An up-regulation in *CaMKK2* expression was observed over the course of hepatic tumor progression in human and mouse samples [35], and *CaMKIV* expression was likewise increased in human and rat liver tumors [36]. It will be interesting to determine the distinct mechanisms for regulating the expression of these genes during pathologic versus physiologic conditions, or whether these events are merely the result of compensatory processes. Moreover, both CaMKK2 and CaMKIV have pleiotropic and cell-autonomous functions in various metabolic organs. Are there tissue- or cell-specific mechanisms in place to regulate the expression of these kinases?

Analysis of bulk tissue tends to dilute the contribution of individual and underrepresented cell types relative to that of the parenchymal cell type. It was previously demonstrated in the liver that fractionation of the hepatic cell types prior to transcriptomic analyses revealed unique, cell-centric gene expression patterns generated in response to oncogenic or chronic metabolic stress [56]. Performing OMICs-based analyses on individual cell populations will not only uncover signaling nodes specific for that cell type, but are likely to unveil cell-specific functions for proteins and/or protein families. Fractionation of tissues into individual cell types could help determine whether any of the Ca<sup>2+</sup>/CaM-dependent kinase pathway components, particularly CaMKK2, have cell-specific functions.

The precise distribution of the CaMKK2 protein within the cell is poorly understood. Determining the subcellular localization of the active versus inactive forms of this kinase, in basal versus stimulated states, will shed light on unappreciated functions of CaMKK2. Various studies implicate CaMKK2 activity with phagocytosis [37], secretion [52], and autophagy [57], which involve trafficking of membrane-associated and soluble proteins. Whether CaMKK2 is tethered to any intracellular membranes and what mechanisms are in place to regulate its trafficking within the cell are important questions that warrant further investigation.

Only recently has it been discovered that a kinase other than Lkb1 could phosphorylate AMPK. CaMKK2 activates the AMPK $\alpha$  subunit in response to increases in intracellular Ca<sup>2+</sup> concentration, which is then followed by complex formation involving CaMKK2, AMPK $\alpha$  and  $\beta$  subunits and Ca<sup>2+</sup>/CaM [5]. As CaMKI, CaMKIV and AMPK represent the most well characterized substrates of CaMKK2, it is likely that additional substrates exist that may open doors for unappreciated physiological functions of this kinase. The availability of specific pharmacological inhibitors for any new CaMKK2 substrates will be greatly beneficial, especially for assessing their potential as putative therapeutic agents.

## Acknowledgments

This work was supported by NIH GM033976 to A.R.M. The authors acknowledge the joint participation by Adrienne Helis Malvin Medical Research Foundation through its direct engagement in the continuous active conduct of medical research in conjunction with Baylor College of Medicine and the “CaMKK2 inhibitors for therapeutic treatment of hepatic cancer” program.

## References

1. Chin D, Means AR. Calmodulin: a prototypical calcium sensor. *Trends in cell biology*. 2000; 10:322–328. [PubMed: 10884684]
2. Means AR, Dedman JR. Calmodulin--an intracellular calcium receptor. *Nature*. 1980; 285:73–77. [PubMed: 6990273]
3. Clapham DE. Calcium signaling. *Cell*. 2007; 131:1047–1058. [PubMed: 18083096]
4. Berchtold MW, Egli R, Rhyner JA, Hameister H, Strehler EE. Localization of the human bona fide calmodulin genes CALM1, CALM2, and CALM3 to chromosomes 14q24-q31, 2p21.1-p21.3, and 19q13.2-q13.3. *Genomics*. 1993; 16:461–465. [PubMed: 8314583]
5. Racioppi L, Means AR. Calcium/calmodulin-dependent protein kinase kinase 2: roles in signaling and pathophysiology. *J Biol Chem*. 2012; 287:31658–31665. [PubMed: 22778263]
6. Green MF, Anderson KA, Means AR. Characterization of the CaMKKbeta-AMPK signaling complex. *Cell Signal*. 2011; 23:2005–2012. [PubMed: 21807092]
7. dos Santos LC, de Padua Cintra I, Fisberg M, Martini LA. Calcium intake and its relationship with adiposity and insulin resistance in post-pubertal adolescents. *Journal of human nutrition and dietetics: the official journal of the British Dietetic Association*. 2008; 21:109–116. [PubMed: 18339051]
8. Torres MR, da Ferreira TS, Carvalho DC, Sanjuliani AF. Dietary calcium intake and its relationship with adiposity and metabolic profile in hypertensive patients. *Nutrition*. 2011; 27:666–671. [PubMed: 20934855]
9. Coll AP, Yeo GSH. The hypothalamus and metabolism: integrating signals to control energy and glucose homeostasis. *Current Opinion in Pharmacology*. 2013; 13:970–976. [PubMed: 24075719]
10. Anderson KA, Ribar TJ, Lin F, Noeldner PK, Green MF, et al. Hypothalamic CaMKK2 Contributes to the Regulation of Energy Balance. *Cell Metabolism*. 2008; 7:377–388. [PubMed: 18460329]

11. Wu WN, Wu PF, Zhou J, Guan XL, Zhang Z, et al. Orexin-A activates hypothalamic AMP-activated protein kinase signaling through a Ca(2)(+)-dependent mechanism involving voltage-gated L-type calcium channel. *Mol Pharmacol*. 2013; 84:876–887. [PubMed: 24068427]
12. Yadav VK, Oury F, Suda N, Liu ZW, Gao XB, et al. A serotonin-dependent mechanism explains the leptin regulation of bone mass, appetite, and energy expenditure. *Cell*. 2009; 138:976–989. [PubMed: 19737523]
13. Yadav VK, Oury F, Tanaka KF, Thomas T, Wang Y, et al. Leptin-dependent serotonin control of appetite: temporal specificity, transcriptional regulation, and therapeutic implications. *J Exp Med*. 2011; 208:41–52. [PubMed: 21187319]
14. Oury F, Yadav VK, Wang Y, Zhou B, Liu XS, et al. CREB mediates brain serotonin regulation of bone mass through its expression in ventromedial hypothalamic neurons. *Genes Dev*. 2010; 24:2330–2342. [PubMed: 20952540]
15. Takeda S, Karsenty G. Molecular bases of the sympathetic regulation of bone mass. *Bone*. 2008; 42:837–840. [PubMed: 18295563]
16. Cary RL, Waddell S, Racioppi L, Long F, Novack DV, et al. Inhibition of Ca(2)(+)/calmodulin-dependent protein kinase kinase 2 stimulates osteoblast formation and inhibits osteoclast differentiation. *J Bone Miner Res*. 2013; 28:1599–1610. [PubMed: 23408651]
17. Bracis C, Gurarie E, Van Moorter B, Goodwin RA. Memory Effects on Movement Behavior in Animal Foraging. *PLoS ONE*. 2015; 10:e0136057. [PubMed: 26288228]
18. Janmaat KRL, Ban SD, Boesch C. Chimpanzees use long-term spatial memory to monitor large fruit trees and remember feeding experiences across seasons. *Animal Behaviour*. 2013; 86:1183–1205.
19. Peters M, Mizuno K, Ris L, Angelo M, Godaux E, et al. Loss of Ca2+/Calmodulin Kinase Kinase  $\beta$  Affects the Formation of Some, But Not All, Types of Hippocampus-Dependent Long-Term Memory. *The Journal of Neuroscience*. 2003; 23:9752–9760. [PubMed: 14586002]
20. Mizuno K, Ris L, Sánchez-Capelo A, Godaux E, Giese KP. Ca2+/calmodulin kinase kinase alpha is dispensable for brain development but is required for distinct memories in male, though not in female, mice. *Mol Cell Biol*. 2006;26.
21. Ho N, Liauw JA, Blaeser F, Wei F, Hanissian S, et al. Impaired synaptic plasticity and cAMP response element-binding protein activation in Ca2+/calmodulin-dependent protein kinase type IV/Gr-deficient mice. *J Neurosci*. 2000; 20:6459–6472. [PubMed: 10964952]
22. Kang H, Sun LD, Atkins CM, Soderling TR, Wilson MA, et al. An important role of neural activity-dependent CaMKIV signaling in the consolidation of long-term memory. *Cell*. 2001; 106:771–783. [PubMed: 11572782]
23. Mizuno K, Antunes-Martins A, Ris L, Peters M, Godaux E, et al. Calcium/calmodulin kinase kinase beta has a male-specific role in memory formation. *Neuroscience*. 2007;145. [PubMed: 17220885]
24. Minichiello L, Calella AM, Medina DL, Bonhoeffer T, Klein R, et al. Mechanism of TrkB-mediated hippocampal long-term potentiation. *Neuron*. 2002; 36:121–137. [PubMed: 12367511]
25. Pivovarova NB, Andrews SB. Calcium-dependent mitochondrial function and dysfunction in neurons. *FEBS J*. 2010; 277:3622–3636. [PubMed: 20659161]
26. Tatemichi TK, Desmond DW, Stern Y, Paik M, Sano M, et al. Cognitive impairment after stroke: frequency, patterns, and relationship to functional abilities. *J Neurol Neurosurg Psychiatry*. 1994; 57:202–207. [PubMed: 8126506]
27. McCullough LD, Tarabishy S, Liu L, Benashski S, Xu Y, et al. Inhibition of calcium/calmodulin-dependent protein kinase kinase  $\alpha$  and calcium/calmodulin-dependent protein kinase IV is detrimental in cerebral ischemia. *Stroke*. 2013;44.
28. Liu L, McCullough L, Li J. Genetic deletion of calcium/calmodulin-dependent protein kinase kinase  $\beta$  (CaMKK  $\beta$ ) or CaMK IV exacerbates stroke outcomes in ovariectomized (OVXed) female mice. *BMC Neuroscience*. 2014; 15:1–10. [PubMed: 24380503]
29. Kokubo M, Nishio M, Ribar TJ, Anderson KA, West AE, et al. BDNF-mediated cerebellar granule cell development is impaired in mice null for CaMKK2 or CaMKIV. *J Neurosci*. 2009; 29:8901–8913. [PubMed: 19605628]

30. Wen L, Chen Z, Zhang F, Cui X, Sun W, et al. Ca<sup>2+</sup>/calmodulin-dependent protein kinase kinase  $\beta$  phosphorylation of Sirtuin 1 in endothelium is atheroprotective. *Proceedings of the National Academy of Sciences*. 2013; 110:E2420–E2427.
31. Ronnett GV, Ramamurthy S, Kleman AM, Landree LE, Aja S. AMPK in the brain: its roles in energy balance and neuroprotection. *Journal of Neurochemistry*. 2009; 109:17–23. [PubMed: 19393004]
32. Rousset CI, Leiper FC, Kichev A, Gressens P, Carling D, et al. A dual role for AMP-activated protein kinase (AMPK) during neonatal hypoxic–ischemic brain injury in mice. *Journal of Neurochemistry*. 2015; 133:242–252. [PubMed: 25598140]
33. Anderson KA, Lin F, Ribar TJ, Stevens RD, Muehlbauer MJ, et al. Deletion of CaMKK2 from the Liver Lowers Blood Glucose and Improves Whole-Body Glucose Tolerance in the Mouse. *Molecular Endocrinology*. 2012; 26:281–291. [PubMed: 22240810]
34. Nordlie RC, Foster JD, Lange AJ. Regulation of glucose production by the liver. *Annu Rev Nutr*. 1999; 19:379–406. [PubMed: 10448530]
35. Lin F, Marcelo KL, Rajapakse K, Coarfa C, Dean A, et al. The camKK2/camKIV relay is an essential regulator of hepatic cancer. *Hepatology*. 2015; 62:505–520. [PubMed: 25847065]
36. Tamura N, Tai Y, Sugimoto K, Kobayashi R, Konishi R, et al. Enhanced expression and activation of Ca<sup>2+</sup>/calmodulin-dependent protein kinase IV in hepatocellular carcinoma. *Cancer*. 2000; 89:1910–1916. [PubMed: 11064347]
37. Racioppi L, Noeldner PK, Lin F, Arvai S, Means AR. Calcium/Calmodulin-dependent Protein Kinase Kinase 2 Regulates Macrophage-mediated Inflammatory Responses. *Journal of Biological Chemistry*. 2012; 287:11579–11591. [PubMed: 22334678]
38. Blonski W, Kotlyar DS, Forde KA. Non-viral causes of hepatocellular carcinoma. *World Journal of Gastroenterology: WJG*. 2010; 16:3603–3615. [PubMed: 20677332]
39. Xi Y, Wu M, Li H, Dong S, Luo E, et al. Baicalin Attenuates High Fat Diet-Induced Obesity and Liver Dysfunction: Dose-Response and Potential Role of CaMKK $\beta$ /AMPK/ACC Pathway. *Cellular Physiology and Biochemistry*. 2015; 35:2349–2359. [PubMed: 25896320]
40. Deshmukh RR, Dou QP. Proteasome inhibitors induce AMPK activation via CaMKK $\beta$  in human breast cancer cells. *Breast Cancer Research and Treatment*. 2015; 153:79–88. [PubMed: 26227473]
41. Rosen ED, Spiegelman BM. Adipocytes as regulators of energy balance and glucose homeostasis. *Nature*. 2006; 444:847–853. [PubMed: 17167472]
42. Hajer GR, van Haeften TW, Visseren FL. Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *Eur Heart J*. 2008; 29:2959–2971. [PubMed: 18775919]
43. Jensen B, Farach-Carson MC, Kenaley E, Akanbi KA. High extracellular calcium attenuates adipogenesis in 3T3-L1 preadipocytes. *Exp Cell Res*. 2004; 301:280–292. [PubMed: 15530863]
44. Lin F, Ribar TJ, Means AR. The Ca<sup>2+</sup>/Calmodulin-Dependent Protein Kinase Kinase, CaMKK2, Inhibits Preadipocyte Differentiation. *Endocrinology*. 2011; 152:3668–3679. [PubMed: 21862616]
45. Teng EC, Racioppi L, Means AR. A cell-intrinsic role for CaMKK2 in granulocyte lineage commitment and differentiation. *J Leukoc Biol*. 2011; 90:897–909. [PubMed: 21816924]
46. Peng I-C, Chen Z, Sun W, Li Y-S, Marin TL, et al. Glucagon regulates ACC activity in adipocytes through the CaMKK $\beta$ /AMPK pathway. *American Journal of Physiology -Endocrinology and Metabolism*. 2012; 302:E1560–E1568. [PubMed: 22454291]
47. Gormand A, Henriksson E, Ström K, Jensen TE, Sakamoto K, et al. Regulation of AMP-activated protein kinase by LKB1 and CaMKK in adipocytes. *Journal of Cellular Biochemistry*. 2011; 112:1364–1375. [PubMed: 21312243]
48. Massie CE, Lynch A, Ramos-Montoya A, Boren J, Stark R, et al. The androgen receptor fuels prostate cancer by regulating central metabolism and biosynthesis. *The EMBO Journal*. 2011; 30:2719–2733. [PubMed: 21602788]
49. Krebs J, Means RL, Honegger P. Induction of calmodulin kinase IV by the thyroid hormone during the development of rat brain. *J Biol Chem*. 1996; 271:11055–11058. [PubMed: 8626646]
50. Liu YY, Brent GA. A complex deoxyribonucleic acid response element in the rat Ca(2+)/calmodulin-dependent protein kinase IV gene 5'-flanking region mediates thyroid hormone

- induction and chicken ovalbumin upstream promoter transcription factor 1 repression. *Mol Endocrinol.* 2002; 16:2439–2451. [PubMed: 12403833]
51. Gromada J, Frokjaer-Jensen J, Dissing S. Glucose stimulates voltage- and calcium-dependent inositol trisphosphate production and intracellular calcium mobilization in insulin-secreting beta TC3 cells. *Biochem J.* 1996; 314(Pt 1):339–345. [PubMed: 8660305]
52. Marcelo KL, Ribar T, Means CR, Tsimelzon A, Stevens RD, et al. Research Resource: Roles for Calcium/Calmodulin-Dependent Protein Kinase Kinase 2 (CaMKK2) in Systems Metabolism. *Mol Endocrinol.* 2016:me20161021.
53. Balasubramanian R, Maruoka H, Jayasekara PS, Gao Z-G, Jacobson KA. AMP-activated protein kinase as regulator of P2Y6 receptor-induced insulin secretion in mouse pancreatic  $\beta$ -cells. *Biochemical Pharmacology.* 2013; 85:991–998. [PubMed: 23333427]
54. Park S-H, Ryu S-Y, Yu W-J, Han YE, Ji Y-S, et al. Leptin promotes KATP channel trafficking by AMPK signaling in pancreatic  $\beta$ -cells. *Proceedings of the National Academy of Sciences.* 2013; 110:12673–12678.
55. Choudhary A, Hu He K, Mertins P, Udeshi ND, Dančik V, et al. Quantitative-Proteomic Comparison of Alpha and Beta Cells to Uncover Novel Targets for Lineage Reprogramming. *PLoS ONE.* 2014; 9:e95194. [PubMed: 24759943]
56. Means, AR.; Kahl, CR.; Crenshaw, DG.; Dayton, JS. Traversing the Cell Cycle: The Calcium/Calmodulin Connection. In: Carafoli, E.; Klee, C., editors. *Calcium as a Cellular Regulator.* New York: Oxford University Press; 1999.
57. Crawford SE, Estes MK. Viroporin-mediated calcium-activated autophagy. *Autophagy.* 2013; 9:797–798. [PubMed: 23442576]
58. Cheung WY. Cyclic 3',5'-nucleotide phosphodiesterase. Demonstration of an activator. *Biochem Biophys Res Commun.* 1970; 38:533–538. [PubMed: 4315350]
59. Kakiuchi S, Yamazaki R. Calcium dependent phosphodiesterase activity and its activating factor (PAF) from brain studies on cyclic 3',5'-nucleotide phosphodiesterase (3). *Biochem Biophys Res Commun.* 1970; 41:1104–1110. [PubMed: 4320714]
60. Wang JH, Desai R. A brain protein and its effect on the Ca<sup>2+</sup>-and protein modulator-activated cyclic nucleotide phosphodiesterase. *Biochem Biophys Res Commun.* 1976; 72:926–932. [PubMed: 186066]
61. Cheung WY. Calmodulin plays a pivotal role in cellular regulation. *Science.* 1980; 207:19–27. [PubMed: 6243188]
62. Fakunding JL, Tindall DJ, Dedman JR, Mena CR, Means AR. Biochemical actions of follicle-stimulating hormone in the sertoli cell of the rat testis. *Endocrinology.* 1976; 98:392–402. [PubMed: 174898]
63. Shenolikar S, Cohen PT, Cohen P, Nairn AC, Perry SV. The role of calmodulin in the structure and regulation of phosphorylase kinase from rabbit skeletal muscle. *Eur J Biochem.* 1979; 100:329–337. [PubMed: 159817]
64. Colomer-Font, J.; Means, AR. Physiological roles of the Ca<sup>2+</sup>/CaM-dependent protein kinase cascade in Health and Disease. In: Carafoli, E.; Brini, M., editors. *Calcium Signalling and Disease.* Dordrecht, The Netherlands: Springer Science; 2007. p. 169-214.
65. Putkey JA, Ts'ui KF, Tanaka T, Lagace L, Stein JP, et al. Chicken calmodulin genes. A species comparison of cDNA sequences and isolation of a genomic clone. *J Biol Chem.* 1983; 258:11864–11870. [PubMed: 6137485]
66. Guerriero V Jr, Russo MA, Olson NJ, Putkey JA, Means AR. Domain organization of chicken gizzard myosin light chain kinase deduced from a cloned cDNA. *Biochemistry.* 1986; 25:8372–8381. [PubMed: 3030394]
67. Hanley RM, Means AR, Ono T, Kemp BE, Burgin KE, et al. Functional analysis of a complementary DNA for the 50-kilodalton subunit of calmodulin kinase II. *Science.* 1987; 237:293–297. [PubMed: 3037704]
68. Means AR, Cruzalegui F, LeMagueresse B, Needleman DS, Slaughter GR, et al. A novel Ca<sup>2+</sup>/calmodulin-dependent protein kinase and a male germ cell-specific calmodulin-binding protein are derived from the same gene. *Mol Cell Biol.* 1991; 11:3960–3971. [PubMed: 1649385]



69. Picciotto MR, Czernik AJ, Nairn AC. Calcium/calmodulin-dependent protein kinase I. cDNA cloning and identification of autophosphorylation site. *J Biol Chem.* 1993; 268:26512–26521. [PubMed: 8253780]
70. Tokumitsu H, Enslen H, Soderling TR. Characterization of a Ca<sup>2+</sup>/calmodulin-dependent protein kinase cascade. Molecular cloning and expression of calcium/calmodulin-dependent protein kinase kinase. *J Biol Chem.* 1995; 270:19320–19324. [PubMed: 7642608]
71. Anderson KA, Means RL, Huang QH, Kemp BE, Goldstein EG, et al. Components of a calmodulin-dependent protein kinase cascade. Molecular cloning, functional characterization and cellular localization of Ca<sup>2+</sup>/calmodulin-dependent protein kinase kinase beta. *J Biol Chem.* 1998; 273:31880–31889. [PubMed: 9822657]
72. Gao G, Widmer J, Stapleton D, Teh T, Cox T, et al. Catalytic subunits of the porcine and rat 5' - AMP-activated protein kinase are members of the SNF1 protein kinase family. *Biochim Biophys Acta.* 1995; 1266:73–82. [PubMed: 7718624]
73. Wayman GA, Kaech S, Grant WF, Davare M, Impey S, et al. Regulation of axonal extension and growth cone motility by calmodulin-dependent protein kinase I. *J Neurosci.* 2004; 24:3786–3794. [PubMed: 15084659]
74. Kahl CR, Means AR. Regulation of cyclin D1/Cdk4 complexes by calcium/calmodulin-dependent protein kinase I. *J Biol Chem.* 2004; 279:15411–15419. [PubMed: 14754892]
75. Scott JW, Park E, Rodriguiz RM, Oakhill JS, Issa SM, et al. Autophosphorylation of CaMKK2 generates autonomous activity that is disrupted by a T85S mutation linked to anxiety and bipolar disorder. *Sci Rep.* 2015; 5:14436. [PubMed: 26395653]
76. Erhardt A, Lucae S, Unschuld PG, Ising M, Kern N, et al. Association of polymorphisms in P2RX7 and CaMKKb with anxiety disorders. *Journal of Affective Disorders.* 2007; 101:159–168. [PubMed: 17197037]

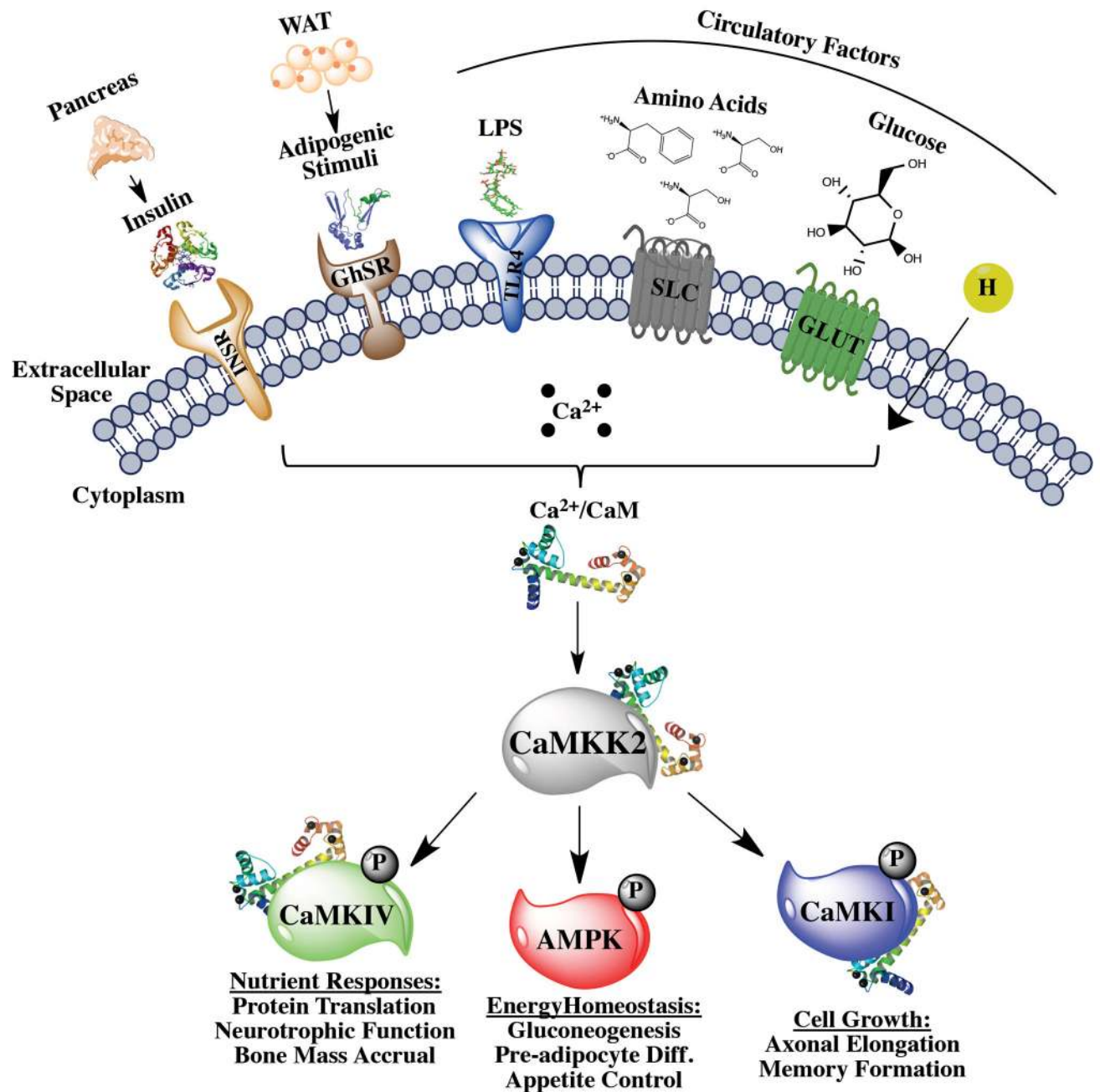
### Trends Box

Calmodulin is the intracellular receptor for  $\text{Ca}^{2+}$  that controls the downstream actions of the CaMK cascade that contributes to metabolic homeostasis.

CaMKK2 contributes to the maintenance of organismal energy homeostasis such that *CaMKK2*<sup>-/-</sup> mice are protected from diet-induced obesity, glucose intolerance, and insulin resistance.

CaMKK2 activates CaMKIV to regulate sympathetic tone and bone mass accrual, protect neurons from ischemia-related injuries, maintain the integrity of the blood-brain barrier, and regulate protein synthesis to control hepatic tumor cell growth.

The pleiotropic functions of CaMKK2 downstream of  $\text{Ca}^{2+}$ /CaM for coordinating energy homeostasis underscore the importance of developing suitable inhibitors against CaMKK2 for treatment of metabolic-related diseases.



**Figure 1. The  $Ca^{2+}/CaM$ -dependent kinase cascade mediates pleiotropic metabolic responses to physiologic and pathophysiologic stimuli**  
 Upstream extracellular signals such as insulin from the pancreas, adipogenic stimuli from white adipose tissue (WAT), and lipopolysaccharide (LPS), amino acids, hormones and glucose from the circulation bind to their respective receptors that trigger a rise in intracellular  $Ca^{2+}$  concentration and accumulation of  $Ca^{2+}/CaM$  targets such as CaMKK2. The increased affinity and binding of  $Ca^{2+}/CaM$  for CaMKK2 results in an increase in CaMKK2 kinase activity, which phosphorylates and activates CaMKIV, AMPK and CaMKI. Activation of CaMKI is involved in regulation of cell growth, as observed in neurite

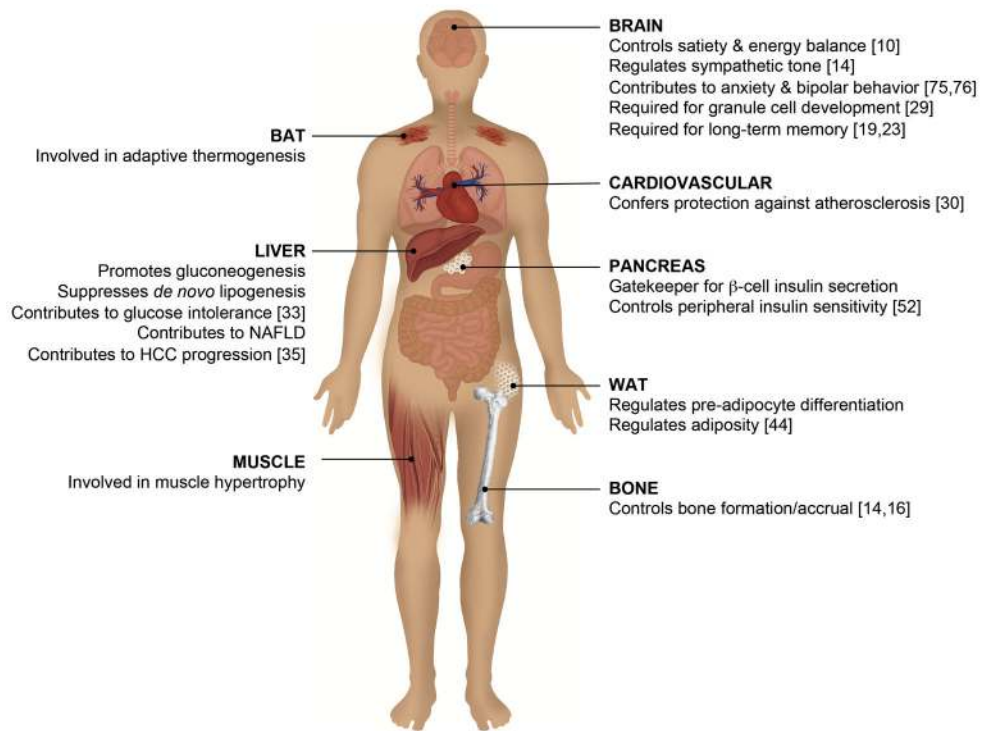
elongation and branching [73] as well as during cell cycle control [74]. CaMKK2-dependent activation of AMPK leads to regulation of energy balance, particularly in the brain [10], liver [33] and adipose [44]. Regulation of CaMKIV activity results in control of protein synthesis and gene expression programs responsive to nutrients [35] and hormones [14].

Author Manuscript

Author Manuscript

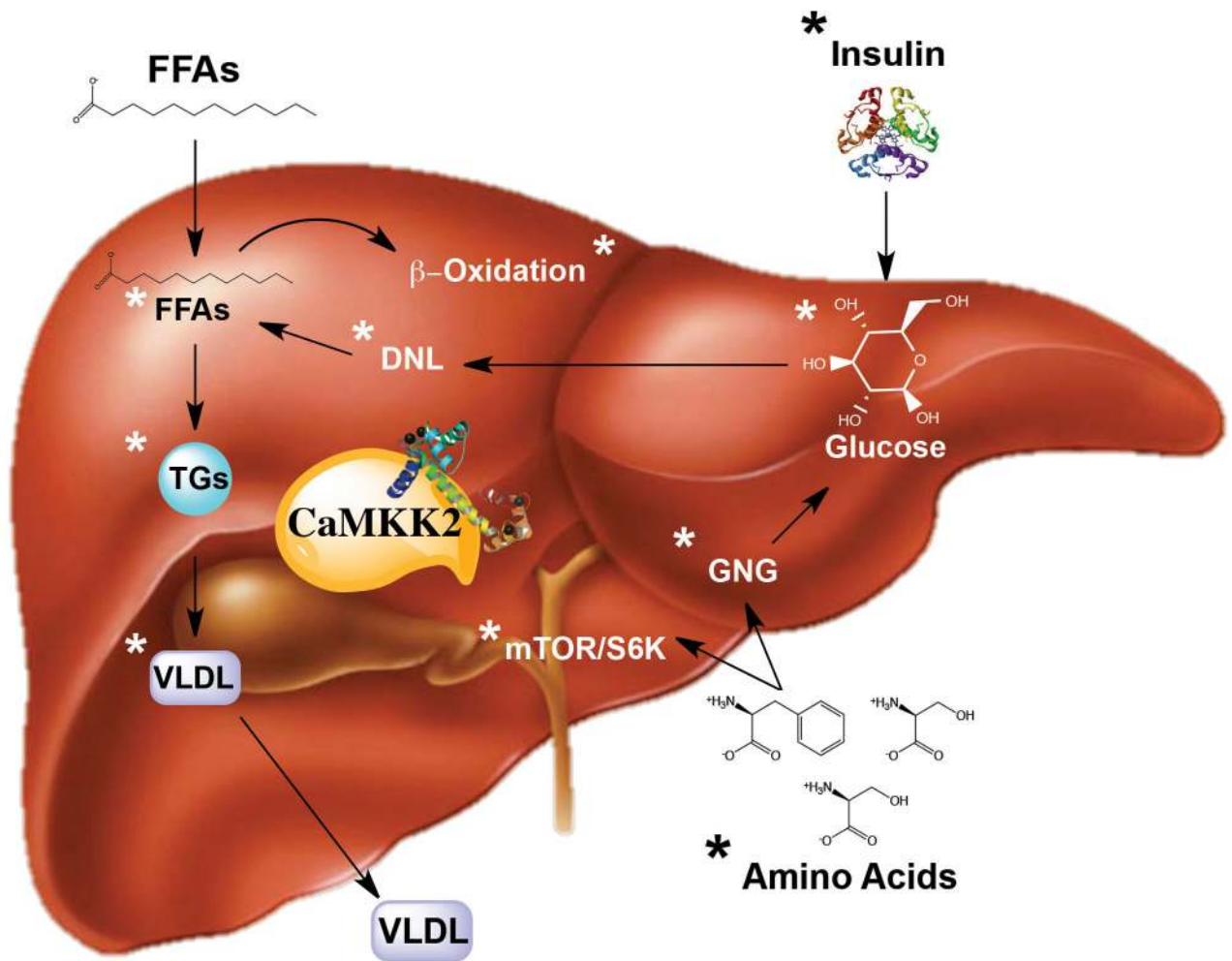
Author Manuscript

Author Manuscript



**Figure 2. CaMKK2 regulates whole body energy homeostasis through coordinating the actions of key metabolic tissues**

In the brain, CaMKK2 and AMPK function to control appetite and energy homeostasis [10], whereas CaMKK2-mediated activation of CaMKIV is required for cerebellar granule cell development [29]. CaMKK2 is required for regulation of sympathetic tone [14] and long-term memory formation [19,23], and is implicated in anxiety and bipolar disorder [75,76]. Within the liver, CaMKK2 regulates AMPK to promote gluconeogenesis while suppressing *de novo* lipogenesis [33]. The CaMKK2/CaMKIV axis in the liver also contributes to non-alcoholic fatty liver disease (NAFLD) and is instrumental during progression of hepatocellular carcinoma [35]. In white adipose tissue (WAT), CaMKK2 phosphorylates AMPK to regulate adiposity and pre-adipocyte differentiation [44]. In brown adipose tissue (BAT), CaMKK2 plays a role in adaptive thermogenesis. In the pancreas, CaMKK2 acts as a gatekeeper for  $\beta$ -cell insulin secretion and controls peripheral insulin sensitivity [52]. Within the cardiovascular system, CaMKK2 confers protection against atherosclerosis [30]. In bone, CaMKK2 also functions to regulate bone mass accrual [14,16].



### Figure 3. Liver-centric metabolic functions of CaMKK2

In the liver, ablation of *CaMKK2* impacts the metabolic flexibility of numerous pathways as indicated by the asterisks (\*). Loss of *CaMKK2* in response to high fat diet feeding reduces levels of free fatty acids (FFAs), triglycerides (TGs) and very low-density lipoproteins (VLDL) as well as low density lipoproteins and cholesterol (not depicted) [52]. Consistent with these observations, hepatic-specific ablation of *CaMKK2* is sufficient to confer improved insulin sensitivity [33]. Primary hepatocytes devoid of *CaMKK2* show increased *de novo* lipogenesis along with increased  $\beta$ -oxidation [33]. In the context of hepatic cancer, *CaMKK2* is essential for driving mTOR/S6K signaling to promote protein synthesis that increases cancer cell proliferation [35].