

The Calcium-Sensing Receptor: A Window into the Physiology and Pathophysiology of Mineral Ion Metabolism

NAIBEDYA CHATTOPADHYAY*, AMBRISH MITHAL, AND EDWARD M. BROWN†

Endocrine-Hypertension Division, Department of Medicine, Brigham and Women's Hospital, Boston, Massachusetts 02115

- I. Introduction
- II. Physiology of Calcium Homeostasis
- III. Calcium-Sensing by Various Cell Types
 - A. Parathyroid cells
 - B. Kidney cells
 - C. C-cells
 - D. $[Ca^{2+}]_o$ -sensing in other cell types
- IV. Molecular Cloning of a $[Ca^{2+}]_o$ -Sensing Receptor (CaR)
 - A. Bovine parathyroid $[Ca^{2+}]_o$ -sensing receptor
 - B. Cloning of additional species homologs of the CaR
 - C. Receptor-effector coupling of the CaR
 - D. Tissue distribution of the CaR
 - E. Regulation of the CaR
 - F. The CaR gene
- V. Are There Additional Forms of $[Ca^{2+}]_o$ -Sensors/Receptors?
- VI. Syndromes of Extracellular Calcium Resistance
 - A. Familial hypocalciuric hypercalcemia (FHH)
 1. Genetics of FHH
 2. The CaR in FHH: implications for regulation of parathyroid and renal function by $[Ca^{2+}]_o$
 - B. Neonatal severe hyperparathyroidism (NSHPT)
 1. Genetics of NSHPT: association with FHH
 2. Mutations in the CaR in NSHPT
 3. Spectrum of NSHPT
- VII. Activating Mutations of the CaR: Autosomal Dominant Hypocalcemia
- VIII. Alterations in CaR Function in Other Disease States
 - IX. Diagnostic Implications
 - X. Therapeutic Implications
 - XI. Summary

I. Introduction

CALCIUM $^{1}(Ca^{2+})$ ions are of critical importance for a variety of vital bodily functions (1–4). Intra- and extracellular Ca^{2+} act in distinct but sometimes complementary

ways to regulate a multitude of biological processes. The cytosolic free calcium concentration ($[Ca^{2+}]_i$) plays a pivotal role in controlling cellular processes such as secretion, differentiation, and motility (2, 5). Calcium ions act as key intracellular second messengers and also as cofactors for a number of enzymes. Although $[Ca^{2+}]_i$ generally exists at much lower levels than the extracellular calcium concentration ($[Ca^{2+}]_o$), it can undergo large, rapid changes due to either influx through the cell membrane or release from intracellular stores (2, 5). $[Ca^{2+}]_o$, on the other hand, remains remarkably constant, varying by only a few percentage points over much of a lifetime under normal circumstances (1, 3, 4). $[Ca^{2+}]_o$ likewise plays an essential role in numerous processes, including blood clotting, neuromuscular excitability, and maintenance of the integrity of the skeletal system.

The near constancy of $[Ca^{2+}]_o$ is the result of a complex homeostatic system, which, in mammals and other tetrapods, primarily involves the parathyroid glands, calcitonin (CT)-producing C cells, kidney, bone, and intestine (1, 3, 4). It has been known for many years that calcium ions move across cell membranes via ion channels and other transport processes, but the actual mechanism(s) that enables cells to "sense" (*i.e.* recognize and respond to) $[Ca^{2+}]_o$ was for many years poorly understood. The parathyroid glands of mammals are remarkably sensitive to $[Ca^{2+}]_o$, and studies on this cell type provided the first evidence that $[Ca^{2+}]_o$ modulates $[Ca^{2+}]_i$ and other second messengers without actually crossing the cell membrane, utilizing a mechanism similar to that of ligands acting through the so-called "calcium-mobilizing," G protein-linked receptors (1, 6–8). That this was actually the case was confirmed recently with the cloning of a G protein-coupled, $[Ca^{2+}]_o$ -sensing receptor (CaR) from bovine parathyroid gland (9). Subsequently CaRs from various species have been cloned from several different tissues, including not only those involved in Ca^{2+} homeostasis, such as the kidney (10) and CT-secreting C cells (11), but also tissues, such as the brain (12), not thought to be involved directly in mineral ion homeostasis.

Soon after the cloning of the CaR, it was recognized that inherited mutations in this receptor could produce clinical disorders of mineral ion metabolism, such as familial hypocalciuric hypercalcemia (FHH) (13), neonatal severe hyperparathyroidism (NSHPT) (13), and autosomal dominant hypocalcemia (ADH) (14). It also facilitated the development of "calcimimetic" agents targeted at the receptor, which

Address reprint requests to: Naibedya Chattopadhyay, Ph.D., Endocrine-Hypertension Division, Brigham and Women's Hospital, 221 Longwood Avenue, Boston, Massachusetts 02115.

* Present address: Department of Endocrinology, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, India.

† Supported by USPHS Grants DK-41415, DK-44588, DK-48330, and DK-46422 as well as by grants from NPS Pharmaceuticals, Inc., and the St. Giles Foundation.

could potentially be of substantial use in the management of disorders with abnormal $[Ca^{2+}]_o$ -sensing, such as primary and secondary hyperparathyroidism (15). This review will provide an update on these recent developments, outlining the discovery, cloning, and characterization of the receptor, as well as its known functions in various tissues and the way it is thought to malfunction in hyper- and hypocalcemic states.

II. Physiology of Calcium Homeostasis

The maintenance of $[Ca^{2+}]_o$ within the normal limits of 1.1–1.3 mM found in humans and other mammals is accomplished through the complex interplay of the various hormones and effector tissues that constitute the essential elements of the calcium homeostatic system (1, 3, 4) (Fig. 1). The most important of these hormones are PTH and vitamin D in humans, while CT also exerts a physiologically relevant hypocalcemic action in some species, such as the rat. Alterations in $[Ca^{2+}]_o$ result in activation of both short term (minutes to hours) and long term (days to weeks or longer) homeostatic responses. Lowering of $[Ca^{2+}]_o$, for instance, results in rapid release of preformed PTH from the chief cells of the parathyroid glands, which is followed within 15–30 min by an increase in net PTH production due to reduced intracellular degradation of the hormone (16, 17). Within hours, the level of the mRNA for prepro-PTH increases, which is a result of an increase in gene transcription (18) as well as a result of posttranslational mechanisms (19).

The hypocalcemia-elicited increase in the circulating level of PTH normalizes $[Ca^{2+}]_o$ by virtue of its actions on kidney, bone, and (indirectly through changes in 1,25-dihydroxyvitamin D [$1,25-(OH)_2D_3$] production) intestine (1, 3, 4). PTH

binds to its high-affinity cell surface receptors in the kidney to increase the reabsorption of Ca^{2+} from the distal tubule and promote phosphaturia by inhibiting proximal tubular reabsorption of phosphate within a matter of minutes (4). In bone, PTH probably acts initially on the osteoblast (20), which, through mechanisms involving cellular contact and/or local humoral mediators, stimulates preformed osteoclasts to release calcium and phosphate ions into the extracellular fluid within 1–2 h by enhancing bone resorption. If hypocalcemia persists for longer periods, PTH can increase the formation of osteoclasts from their mononuclear precursors and also enhance the activity of the enzyme, 25-hydroxyvitamin D_3 [$25(OH)D_3$] 1-hydroxylase, in cells of the renal proximal tubule to catalyze the conversion of the prohormone, $25(OH)D_3$, to the active hormone, $1,25-(OH)_2D_3$ (1, 3, 4). The latter acts via its specific receptor in intestinal epithelial cells to promote absorption of Ca^{2+} and also to enhance release of skeletal Ca^{2+} (21). Long-term hypocalcemia, sustained for days to weeks, leads to hyperplasia of the parathyroid glands, which can result in greatly increased rates of PTH secretion (22, 23).

The homeostatic system has negative feedback elements that either reestablish the original steady state of the system or promote a new steady state if, for example, there were a persistent stress on the system, such as a change in dietary calcium availability. The restoration of $[Ca^{2+}]_o$ toward normal directly inhibits PTH synthesis and secretion, while $1,25-(OH)_2D_3$ also exerts potent inhibitory effects of PTH gene expression (24–26), parathyroid cellular proliferation (27, 28), and PTH secretion (29) that provide an additional feedback loop.

Hypercalcemia reverses the homeostatic alterations in the $[Ca^{2+}]_o$ -elevating system just described and, to a lesser extent, activates a hypocalcemic effector mechanism built around the actions of CT. High $[Ca^{2+}]_o$ acutely inhibits PTH release, an action that is opposite to the usual role of calcium in stimulus-secretion coupling. More prolonged elevations in $[Ca^{2+}]_o$ result in suppression of PTH gene expression (30). The resultant reduction in the circulating level of PTH decreases $[Ca^{2+}]_o$ by lowering its renal tubular reabsorption (4), diminishing Ca^{2+} mobilization from bone, and depressing the absorption of Ca^{2+} from the intestine due to reduced PTH-mediated synthesis of $1,25-(OH)_2D_3$. Hypercalcemia also directly stimulates CT secretion from the C cells of the thyroid gland (31). CT is a potent inhibitor of osteoclast activity and can, therefore, reduce the flux of Ca^{2+} into the extracellular fluid from bone. In addition, CT exerts a calciuric effect, which contributes to its hypocalcemic action. However, CT has very modest hypocalcemic effects under normal circumstances in normal adult humans, although it acts more potently in this regard in states of elevated bone turnover, such as hypercalcemia or Paget's disease of bone (3). Therefore, in humans $[Ca^{2+}]_o$ -mediated changes in PTH secretion and, in turn, in $1,25-(OH)_2D_3$ production represent the dominant determinants of the activity of the tissues involved in movements of calcium (and phosphate) ions into or out of the extracellular fluid.

Discussions of calcium homeostasis have generally focused on the direct actions of $[Ca^{2+}]_o$ on parathyroid and C cells as being the major sites where $[Ca^{2+}]_o$ sensing regulates

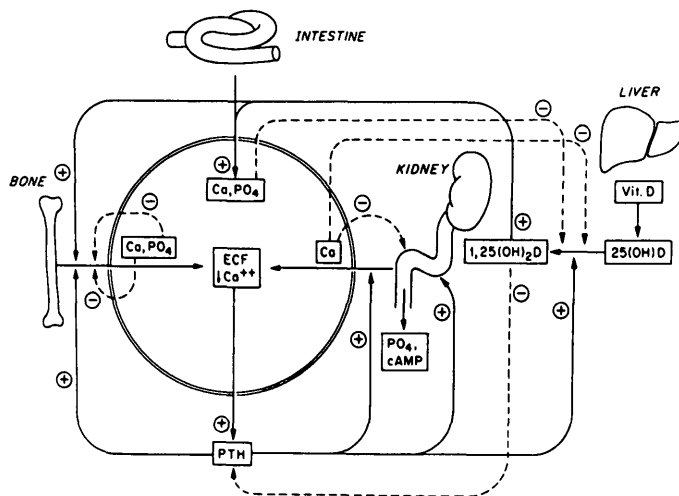


FIG. 1. Schematic diagram illustrating the regulation of $[Ca^{2+}]_o$ homeostasis. The solid lines and arrows indicate the actions of PTH and $1,25-(OH)_2D_3$; the dotted lines and arrows demonstrate examples of direct effects of Ca^{2+} or phosphate ions. Abbreviations are as follows: Ca^{2+} , calcium; PO_4 , phosphate; ECF, extracellular fluid; $1,25-(OH)_2D$, $1,25$ -dihydroxyvitamin D; $25(OH)D$, 25 -hydroxyvitamin D; + signs indicate positive actions while - signs indicate inhibitory effects. [Reproduced with permission from Brown EM, Pollak M, Hebert SC 1994 Cloning and characterization of extracellular Ca^{2+} -sensing receptors from parathyroid and kidney: molecular physiology and pathophysiology of Ca^{2+} -sensing. *The Endocrinologist* 4:419–426.]

the system. As outlined in subsequent sections, however, the capacity of cells to monitor and respond to changes in $[Ca^{2+}]_o$ is a more widespread property than generally recognized (1), and in several cases this $[Ca^{2+}]_o$ sensing is mediated by the recently cloned $[Ca^{2+}]_o$ -sensing receptor. This provides an even more sophisticated mechanism for regulating mineral ion metabolism, where cells use the informational content of the local extracellular ionic environment to modify their functions in homeostatically appropriate ways. The property of $[Ca^{2+}]_o$ sensing is also used as a means of integrating homeostatic systems (e.g. those regulating calcium, magnesium, and water metabolism) in ways that balance the needs of free-living terrestrial organisms for divalent cations and water (32).

III. Calcium Sensing by Various Cell Types

A. Parathyroid cells

The parathyroid cell is remarkably sensitive to alterations in $[Ca^{2+}]_o$, readily detecting changes on the order of a few percent and responding with alterations in hormonal secretion within seconds (1, 6–8). A steep inverse sigmoidal relationship between PTH secretion and $[Ca^{2+}]_o$ has been demonstrated both *in vivo* (33, 34) and *in vitro* (1, 35). A computer model can be employed to fit this relationship (Fig. 2) (36). These curves can be defined in terms of four parameters (A through D), as follows: A represents the maximal rate of PTH release and is equivalent to the secretory reserve of the parathyroid on being exposed to a maximal, acute hypocalcemic stimulus. B represents the slope of the curve at its midpoint. This curve is steepest at this point, so that large changes in PTH secretion take place in response to minor alterations in $[Ca^{2+}]_o$ around the midpoint. C refers to the set point, which is defined as the calcium concentration producing half of the maximal inhibition of PTH secretion; it has values of about 1.0 mM $[Ca^{2+}]_o$ *in vitro* (36) and 1.1–1.3 mM *in vivo* (33) in humans. Others have defined the set point as the level of $[Ca^{2+}]_o$ at which the rate of PTH secretion is one half of its maximal value (37). With the latter definition, the relative position of C on the curve will vary as a function of the suppressibility of the gland (i.e. if maximal suppressibility is only slightly more than 50%, the set point will be very close to the bottom of the curve, where PTH secretion is maximally suppressed). The set point of the parathyroid gland plays a key role in "setting" the level of $[Ca^{2+}]_o$, although the set point for $[Ca^{2+}]_o$ (38) (i.e. the level at which the serum ionized calcium concentration is set) is generally slightly higher than that for the parathyroid gland, such that the circulating level of intact PTH is usually about 20–25% of the maximal level achieved during a maximal hypocalcemic stimulus (33). Not surprisingly, changes in the set point of the parathyroid (e.g. a reset "calciostat") produce major changes in PTH secretion at any given level of $[Ca^{2+}]_o$ (36) and, in turn, the steady state level of the serum calcium concentration. Elevations in set point are typically seen in hyperparathyroid states (36) and in FHH (39, 40), as well as in parathyroid cells maintained in culture (28, 41), implying varying degrees of calcium "resistance" of the parathyroid $[Ca^{2+}]_o$ -sensing mechanism in these conditions. The fourth parameter, D, represents the

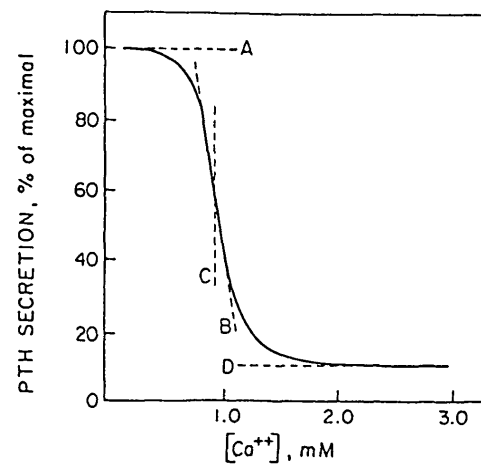
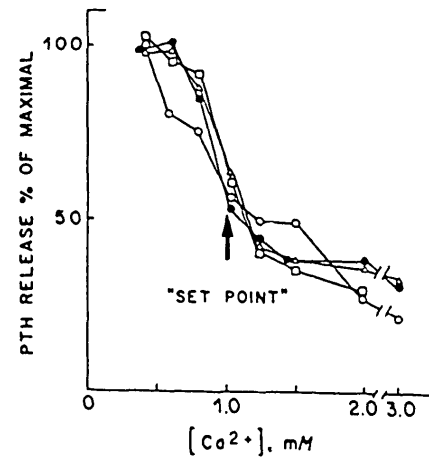


FIG. 2. Inverse sigmoidal relationship between $[Ca^{2+}]_o$ and PTH release and four-parameter model describing such curves. The data in the upper panel represent results obtained using dispersed normal human parathyroid cells and are expressed as the percent of the maximal rate of PTH release observed at 0.3 mM $[Ca^{2+}]_o$. The set point represents the level of $[Ca^{2+}]_o$ at which PTH release is half-maximally suppressed. The lower panel illustrates the four parameters that may be used to describe such curves. Details are given in the text. [Reproduced with permission from Brown EM 1982 PTH secretion *in vivo* and *in vitro*: regulation by calcium and other secretagogues. *Miner Electrolyte Metab* 8:130–150.]

minimum rate of PTH secretion that persists even at very high levels of $[Ca^{2+}]_o$, due to the existence of a basal non-suppressible component of PTH release (42). The computer model used to fit the sigmoidal relationship between PTH and $[Ca^{2+}]_o$ was based on a similar model that had been used to describe the binding of hormones to their cell surface receptors, although at the time it had not been appreciated that $[Ca^{2+}]_o$ regulates parathyroid function through an entirely analogous, receptor-mediated mechanism.

A large body of indirect evidence supported the existence of a "receptor-like mechanism" that confers upon parathyroid cells the capacity to sense $[Ca^{2+}]_o$. Raising $[Ca^{2+}]_o$, for example, activates phospholipase C (PLC) in dispersed bovine parathyroid cells, leading to accumulation of inositol 1,4,5-trisphosphate (IP_3) due to activation of PLC (43) and consequent release of Ca^{2+} from its intracellular stores (6).

This results in an initial, transient "spike" in $[Ca^{2+}]_i$, followed by a sustained increase due to influx of Ca^{2+} through what are most likely voltage-insensitive channels (6, 44), although some studies have suggested that parathyroid cells (45), like C cells, have voltage-sensitive calcium channels (46). In addition, high levels of $[Ca^{2+}]_o$ produce a pertussis toxin-sensitive inhibition of agonist-stimulated cAMP accumulation in bovine parathyroid cells (47). In contrast, pertussis toxin has little, if any, effect on high $[Ca^{2+}]_o$ -evoked changes in $[Ca^{2+}]_i$ or inositol phosphates (48). These studies also showed the capacity of this $[Ca^{2+}]_o$ -sensing apparatus to recognize not only extracellular calcium ions *per se* but also a variety of other inorganic divalent cations (*e.g.* Mg^{2+} , Ba^{2+}), trivalent cations (*e.g.* La^{3+} and Gd^{3+}), and even polyvalent cations such as polylysine, protamine, and neomycin (49–51). The lectin concanavalin A, which binds to sugar moieties on the cell surface, reduces the effects of elevated $[Ca^{2+}]_o$ on $[Ca^{2+}]_i$ and PTH release, suggesting that the putative receptor was a glycoprotein (52). All these observations suggested that the apparatus for $[Ca^{2+}]_o$ sensing was probably a G protein-coupled cell surface receptor, linked to inhibition of adenylate cyclase via G_i and to activation of PLC through a pertussis toxin-insensitive G protein, presumably a member of the G_q family.

B. Kidney cells

$[Ca^{2+}]_o$ exerts direct actions on a variety of cell types in the kidney. Elevated levels of $[Ca^{2+}]_o$ directly inhibit the 1-hydroxylation of 25-hydroxyvitamin D, independent of the associated inhibition of PTH secretion (53), since this effect can be observed in experimental animals in which PTH is "clamped" by infusion of the hormone either into intact or parathyroidectomized animals. In the medullary thick ascending limb (MTAL), but not in the proximal tubule, elevated levels of $[Ca^{2+}]_o$ produce a pertussis toxin-sensitive inhibition of cAMP accumulation (54, 55), analogous to that seen in parathyroid cells (47). $[Ca^{2+}]_o$ also exerts a direct inhibitory action on NaCl reabsorption in the thick ascending limb (TAL) (56), and elevated peritubular, but not luminal, concentrations of either $[Ca^{2+}]_o$ or $[Mg^{2+}]_o$ also inhibit the reabsorption of both calcium and magnesium ions in the TAL (57). In the distal tubule, $[Ca^{2+}]_o$ acts synergistically with $1,25-(OH)_2D_3$ to increase the level of the mRNA for calbindin D_{28K} (58).

It has long been recognized that hypercalcemia produces nephrogenic diabetes insipidus (NDI) in some patients by reducing the action of vasopressin on the MTAL and/or collecting duct of the kidney (59), possibly contributing to the nephrogenic DI that can be observed in this setting. Other effects of $[Ca^{2+}]_o$ on renal function include inhibition of renin secretion from the juxtaglomerular cells (60), renal vasoconstriction (61), and reduction in glomerular filtration rate through as yet unexplained mechanisms (62). Thus, at least in the case of $[Ca^{2+}]_o$ -evoked changes in vasopressin-stimulated cAMP accumulation in the TAL, some of the effects of $[Ca^{2+}]_o$ on renal function could potentially be exerted by a $[Ca^{2+}]_o$ -sensing mechanism similar to that thought to be present in parathyroid cells.

C. C Cells

As discussed previously, elevations in $[Ca^{2+}]_o$ acutely elicit secretion of CT from the thyroidal C cells, which, in turn, inhibits bone resorption (although this action is of minor physiological relevance in normal adult humans) and promotes renal calcium excretion (31). C cells are highly sensitive to $[Ca^{2+}]_o$, suggesting that they too have a specific mechanism for sensing changes in extracellular calcium ions; however, in contrast to parathyroid cells, they show stimulation rather than inhibition of secretion in response to increases in $[Ca^{2+}]_o$ (63). Studies on $[Ca^{2+}]_o$ sensing in C cells suggested fundamental differences between the mechanisms used by parathyroid and C cells to detect changes in $[Ca^{2+}]_o$. Similar to parathyroid cells, C cells respond to small increases in $[Ca^{2+}]_o$ with elevations in $[Ca^{2+}]_i$, but when studied at the level of individual cells, they show slow oscillations in $[Ca^{2+}]_i$ that can be abolished by blockers of voltage-sensitive Ca^{2+} channels (46, 64). Thus release of Ca^{2+} from internal stores appears to play little, if any, role in the high $[Ca^{2+}]_o$ -evoked changes in $[Ca^{2+}]_i$ dynamics in C cells, and the type(s) of Ca^{2+} channels mediating influx of extracellular calcium ions also appears to differ from those activated by elevated levels of $[Ca^{2+}]_o$ in parathyroid cells. On the basis of these results, some investigators suggested that a voltage-sensitive Ca^{2+} channel might represent the principal $[Ca^{2+}]_o$ -sensing mechanism in C cells (64). The resultant high $[Ca^{2+}]_o$ -activated increases in $[Ca^{2+}]_i$ would then stimulate CT secretion through the direct relationship between $[Ca^{2+}]_i$ and exocytosis of CT found in this cell type, as in most secretory cells.

D. $[Ca^{2+}]_o$ -sensing in other cell types

$[Ca^{2+}]_o$ acts directly on additional cell types, some that are involved in mineral ion metabolism but others that are not, implying that they too have $[Ca^{2+}]_o$ -sensing mechanisms potentially differing from those described to this point. For example, elevations in $[Ca^{2+}]_o$ inhibit osteoclastic bone resorption (65, 66) and produce changes in $[Ca^{2+}]_i$ in osteoclasts somewhat reminiscent of those elicited in parathyroid cells by high levels of $[Ca^{2+}]_o$. The pharmacological profile of the polycations that exert such effects differs distinctly from that for the $[Ca^{2+}]_o$ -sensing mechanism in parathyroid, kidney, and C cells. In addition, $[Ca^{2+}]_o$ and other polycations stimulate the proliferation of some osteoblast-like cells and elicit changes in intracellular mediators (67) that are clearly different from those evoked by $[Ca^{2+}]_o$ in parathyroid, kidney, or C cells.

The placenta plays an important role in transporting calcium and other minerals from the mother to the developing fetus, and PTH-related protein (PTHrP) may play an important role in regulating this process (68). Not surprisingly, certain placental cells, the cytotrophoblasts, show regulation of PTHrP secretion by $[Ca^{2+}]_o$ (69). Raising $[Ca^{2+}]_o$ inhibits the proliferation of keratinocytes and promotes their differentiation (70). $[Ca^{2+}]_o$ also modulates the growth and/or differentiation of cultured mammary cells and various other epithelial cells (for review, see Ref. 1). Therefore, $[Ca^{2+}]_o$ sensing is an attribute that is not limited to cells involved in

mineral ion metabolism and might potentially be present in a wider range of cell types and regulate a larger variety of cellular functions than currently recognized.

IV. Molecular Cloning of a $[Ca^{2+}]_o$ -Sensing Receptor

A. Bovine parathyroid $[Ca^{2+}]_o$ -sensing receptor

Racke *et al.* (71) and Shoback and co-workers (72) independently showed that *Xenopus laevis* oocytes became responsive to $[Ca^{2+}]_o$ -sensing receptor agonists, such as Ca^{2+} or Gd^{3+} , after being injected with parathyroid mRNA. Subsequently, Brown *et al.* (9) used a similar assay as a means for screening a bovine parathyroid cDNA library in an attempt to isolate full-length, functional clones of the $[Ca^{2+}]_o$ -sensing receptor from this tissue source. The rationale for choosing expression cloning in *X. laevis* oocytes was that after translation of the functional receptor protein by the oocytes from mRNA synthesized *in vitro* from CaR-encoding cDNA clones of a parathyroid cell library, the receptor protein will couple to the endogenous G protein-activated PLC of the oocyte. The resultant CaR agonist-dependent increases in IP_3 and release of Ca^{2+} from intracellular stores will stimulate Ca^{2+} -activated chloride (Cl^-) currents, producing large currents that are readily measurable by standard electrophysiological means. The membrane impermeant gadolinium ion was chosen over Ca^{2+} as the CaR agonist used to screen for clones encoding functional receptor, since the latter could conceivably confound interpretation of the results by entering the

oocytes through expressed channel proteins and/or transporters (9).

Use of this approach to screen a directional cDNA library prepared from size-fractionated bovine parathyroid poly(A⁺) RNA exhibiting maximal Gd^{3+} -activated Cl^- currents in oocytes (of 4–6 kb in size) resulted in isolation of a single 5.3-kb clone containing the entire coding sequence of the CaR [bovine parathyroid Ca^{2+} -sensing receptor (BoPCaR)(9)]. The deduced amino acid sequence of BoPCaR predicts three major domains (Fig. 3): 1) a large hydrophilic extracellular domain at the amino terminus, consisting of 613 amino acids; 2) a hydrophobic transmembrane domain consisting of 250 amino acids with seven membrane-spanning segments characteristic of the superfamily of G protein-coupled receptors; and 3) a cytosolic carboxy-terminal tail predicted to have 222 amino acids. BoPCaR has nine predicted N-linked glycosylation sites within the extracellular amino-terminal domain, consistent with the native protein being expressed as a glycoprotein. Within the intracellular loops and carboxy-terminal tail of the protein, BoPCaR contains four predicted protein kinase C (PKC) phosphorylation sites, which may contribute to the PKC-mediated uncoupling of the receptor from activation of PLC in bovine parathyroid cells (73).

Among the G protein-coupled receptors, the $[Ca^{2+}]_o$ -sensing receptor only shares amino acid sequence homology with the metabotropic glutamate receptors (mGluRs) (9), which are G protein-coupled receptors present in the central nervous system that respond to glutamate, the

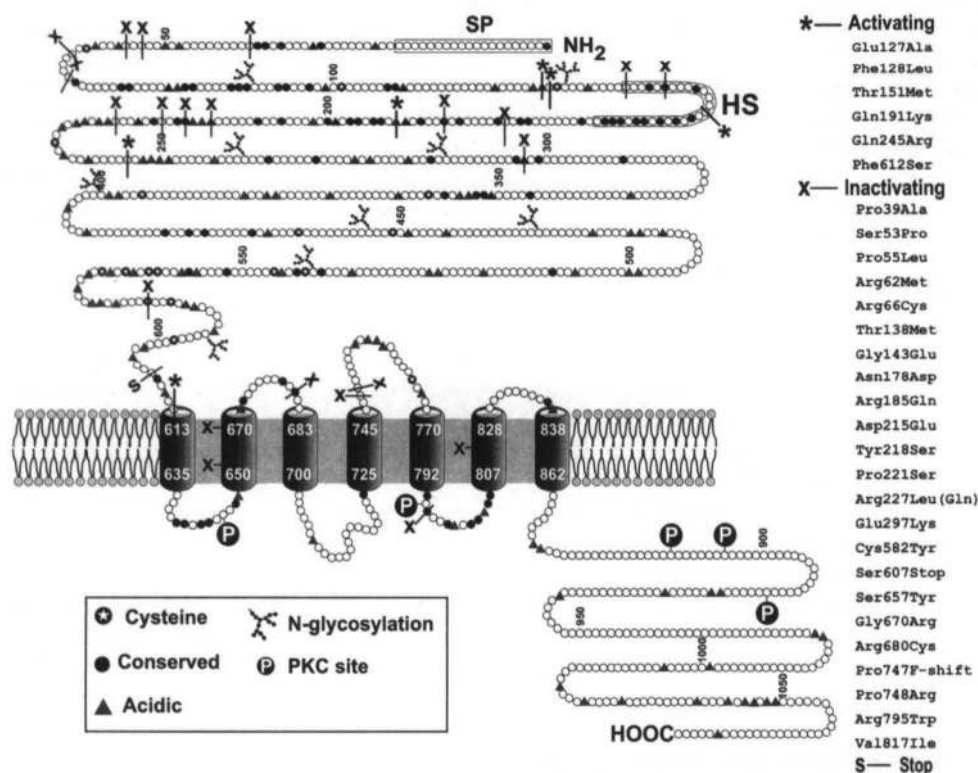


FIG. 3. Schematic diagram of the predicted structure of the $[Ca^{2+}]_o$ -sensing receptor, illustrating inactivating and activating mutations. Symbols are explained in key. Additional abbreviations are: SP, predicted signal peptide; HS, hydrophobic segment. The mutations shown are from Refs. 13, 14, 98, 119, 121, 122, 151, and 152. [Modified with permission from The New England Journal of Medicine E. M. Brown *et al.*: *N Engl J Med* 333:234–240, 1995 (32). © 1995 Massachusetts Medical Society. All rights reserved].

major excitatory neurotransmitter in the brain (74). BoPCaR has only modest identity in its amino acid sequence with the mGluRs (18–24%), but it shares striking topological similarity. Both classes of receptors possess a large amino-terminal, extracellular domain as well as a total of 20 strictly conserved cysteine residues (17 within the extracellular domain and three in transmembrane segments or extracellular loops). Studies using chimeric receptors constructed from mGluRs with differing ligand specificities (75) or chimeras in which the extracellular domains are interchanged between an mGluR and the CaR (76) have established that the principal determinants of ligand binding to both classes of receptors reside within the large amino-terminal domain. The conserved cysteines may contribute to organizing this domain of the receptor into a binding pocket appropriate for interacting with these small charged ligands (*e.g.* glutamate or Ca^{2+} and other polyvalent cations for the mGluRs and CaR, respectively). O'Hara and co-workers (77) and Conklin and Bourne (78) have suggested that the mGluRs and BoPCaR bear structural homology to the bacterial periplasmic binding proteins. The latter are involved in sensing extracellular ligands as part of the processes of chemotaxis or cellular uptake of extracellular ligands in bacteria and have a bilobate structure with the binding pocket for the ligand located within the cleft between the two lobes of the protein (79). Upon binding of the ligand, the protein undergoes a conformational change in which the two lobes approximate one another, largely obliterating the remaining space between them (hence the basis for the "venus fly trap" model of ligand binding by these proteins). O'Hara *et al.* (77) have created a model of the extracellular domain of the mGluRs that successfully predicts certain features of the binding of mGluR agonists to the receptor based on an overall three-dimensional structure postulated to be similar to that of the periplasmic binding proteins. A detailed understanding of how their respective ligands interact with the mGluRs and BoPCaR, however, will require determination of the structure of their extracellular domains by x-ray crystallography. It is of interest that BoPCaR possesses several highly acidic regions within the extracellular domain that are similar to those thought to be involved in the binding of calcium ions to other low affinity calcium-binding proteins, such as calsequestrin and calreticulin, but are not present in the mGluRs (9). There are no predicted high-affinity calcium-binding motifs in this domain of the protein, such as EF hands (the high-affinity Ca^{2+} -binding domain of calmodulin and related intracellular Ca^{2+} -binding proteins), which would be inappropriate for $[\text{Ca}^{2+}]_o$ sensing as they would be persistently occupied by the ambient concentrations within the extracellular fluids.

B. Cloning of additional species homologs of the CaR

Recently, two cDNA clones (4.0 and 5.2 kb) have been isolated from a cDNA library prepared from a human parathyroid adenoma (80). Expression in *X. laevis* oocytes confirmed that both clones encode functional human parathyroid CaRs [human parathyroid calcium receptor (HuPCaR)].

An essentially identical $[\text{Ca}^{2+}]_o$ -sensing receptor was subsequently isolated from a human kidney cDNA library (81). Highly similar, if not identical, rat $[\text{Ca}^{2+}]_o$ -sensing receptors have been cloned from kidney outer medulla [rat kidney calcium-sensing receptor (RaKCaR)] (10), the striatum of the brain (12), and a C cell line derived from medullary thyroid carcinoma (rMTC 44–2) cells (11). All are almost certainly derived from a single gene. The CaRs from different species likewise all show a high degree of homology with BoPCaR (>90% amino acid identity), strongly suggesting they are homologs of the same ancestral gene. Most of the differences between the receptors cloned from different species reside in their carboxy-terminal tails. The predicted glycosylation and PKC phosphorylation sites are more or less conserved between the bovine, rat, and human receptors, although the human and rat receptors have one and two additional PKC phosphorylation sites, respectively. In addition, the human receptor has one predicted protein kinase A (PKA) phosphorylation site and the rat receptor has two such sites, while BoPCaR has no predicted PKA sites. The functional significance of these sites and the interspecies differences between them are at present unknown. Agents stimulating PKC activity are known to uncouple the CaR from activation of PLC, as will be described in detail later (see Section IV.E). Activators of PKA have no obvious effect on $[\text{Ca}^{2+}]_o$ sensing in bovine parathyroid cells, consistent with the lack of predicted PKA sites in the bovine receptor. Similar studies have not yet been reported for the rat and human receptors.

There are several additional notable differences between the predicted amino acid and nucleotide sequences in the various species homologs of the $[\text{Ca}^{2+}]_o$ -sensing receptor. Within the coding sequence, 10 amino acids are inserted between codons 536–545 in one fully characterized clone (HuPCaR-5.2) of two full-length human CaRs isolated from a cDNA library prepared from a parathyroid adenoma (80). None of the other forms of the receptor isolated to date have shown this insertion. Therefore, this insert presumably represents a rare splice variant in which a portion of an intron is incorporated into the coding sequence, without any apparent effect on the function of the expressed receptor (80). There is, in addition, a benign polymorphism (or possibly a cloning artifact) in another of the CaR clones isolated from this same cDNA library (HuPCaR-4.0), with replacement of the normal Arg at codon 990 (as assessed by its presence in BoPCaR) with Gly (80). Again, this change in amino acid sequence has no obvious impact on the function of the expressed receptor. There are additional differences in the non-coding sequence of the receptor. In their 5'-untranslated regions, the two HuPCaR clones diverge 242 bp upstream of the translational start site, with the two distinct sequences 5' to this site presumably representing splice variants within noncoding sequences that are of uncertain physiological significance.

C. Receptor-effector coupling of the CaR

In bovine parathyroid cells, high levels of $[\text{Ca}^{2+}]_o$ activate PLC in a pertussis toxin-insensitive manner (48), unlike its actions on cAMP accumulation (47), suggesting that the CaR is coupled to PLC through a member of the G_q family (82).

In *X. laevis* oocytes expressing the CaR, high $[Ca^{2+}]_o$, $[Gd^{3+}]_o$, or neomycin levels also increased the accumulation of inositol 1,4,5-trisphosphate and $[Ca^{2+}]_i$ by severalfold, the latter arising primarily from intracellular Ca^{2+} stores (9). Unlike the case in bovine parathyroid cells, however, high concentrations of pertussis toxin (10 $\mu\text{g/ml}$) markedly attenuated the high $[Ca^{2+}]_o$ -elicited increases in IP_3 in oocytes. Therefore, unlike parathyroid cells, the CaR activated PLC in the oocyte via a pertussis toxin-sensitive G protein. The specificity for G protein coupling of other receptors can also vary when they are expressed in oocytes compared with the cell of origin (83). Of interest, the high $[Ca^{2+}]_o$ -elicited increase in inositol phosphate accumulation in AtT-20 cells, a mouse pituitary cell line expressing the CaR, is also pertussis toxin-sensitive (84), indicating that the specificity of the G protein through which the CaR couples to PLC can vary in mammalian cells as well.

The expressed CaR confers high $[Ca^{2+}]_o$ -mediated inhibition of cAMP accumulation on human embryonic kidney cells (HEK 293) stably expressing the receptor (85). Recent studies in tubules from the MTAL, however, have raised the possibility that high levels of $[Ca^{2+}]_o$ inhibit agonist-stimulated cAMP accumulation through an indirect mechanism involving arachidonic acid (86). Direct addition of arachidonic acid to tubule suspensions leads to a pertussis toxin-sensitive inhibition of cAMP accumulation. It remains to be determined whether the high $[Ca^{2+}]_o$ -evoked inhibition of cAMP accumulation in cells stably expressing the cloned CaR involves a similar mechanism or whether the CaR can directly couple to inhibition of adenylate cyclase via G_i .

D. Tissue distribution of the CaR

By Northern blot analysis, transcripts for the $[Ca^{2+}]_o$ -sensing receptor are present in a variety of tissues. The sizes of these transcripts vary both within and between species. In the bovine species, the major transcript is 5.6 kb in length (9), with a minor transcript of 9.5 kb, while in rat the major transcript is 7.5–8.5 kb with a minor transcript of 4.1 kb (10–12). The significance of the presence of several CaR transcripts is at present unknown. Some of the tissues expressing transcripts for the CaR play an important role in maintaining calcium homeostasis and are known to sense $[Ca^{2+}]_o$, including the parathyroid and thyroid glands (where the mRNA for the receptor is present exclusively in the C cells) as well as several regions of the kidney. The use of *in situ* hybridization and immunohistochemistry, utilizing antibodies raised against synthetic peptides within the extracellular domain of the CaR, has permitted more detailed localization of the receptor within these tissues. In the parathyroid gland, CaR mRNA is present within the chief cells. The receptor protein is present on the cell surface of the chief cell but can also be visualized in some cases over the cytoplasm (87). It is not yet clear whether the latter represents receptor protein within the biosynthetic pathway or that resident within intracellular organelles, such as the endoplasmic reticulum and secretory vesicles. The total calcium concentration within such organelles is generally maintained within the millimolar range (2) and $[Ca^{2+}]_o$ sensing via the CaR could potentially be involved in regulating the state of filling of the

organelles or other aspects of their calcium homeostasis. C cells likewise contain abundant CaR protein on the plasma membrane (11).

The location of CaR mRNA and protein within the kidney has recently been clarified not only by the use of *in situ* hybridization and immunohistochemistry but also by performing PCR on reverse-transcribed RNA isolated from individually microdissected and identified segments of the nephron (88). In this way it has been possible to identify which of the segments of the nephron contain transcripts for the CaR. mRNA for the receptor is present within nearly all of them, including the glomerulus, proximal convoluted and straight tubules, MTAL and cortical thick ascending limb, distal convoluted tubule, and cortical, outer medullary, and inner medullary collecting ducts. In the TAL, the CaR has a similar, if not identical, distribution to that of the PTH/PTHrP receptor and is present predominantly on the basolateral surface of the cells. Since it is known that high $[Ca^{2+}]_o$ inhibits both PTH-induced cAMP production and Ca^{2+} reabsorption by this segment of the nephron, the presence of both the CaR and PTH receptors in TAL and also in distal convoluted tubule is strong supporting evidence for a crucial role of the $[Ca^{2+}]_o$ -sensing receptor in calciotropic hormone-regulated mineral ion homeostasis (32). Moreover, it implies that the CaR carries out this function not only by regulating PTH secretion but also by modulating its action(s) on the kidney.

In brain, immunolocalization of the CaR protein revealed the presence of the receptor throughout the central nervous system with particular abundance in the cerebellum and hippocampus, olfactory bulbs, ependymal zones of the cerebral ventricles, and cerebral arteries (12). The high level of expression of the CaR in several types of cerebellar cells (*e.g.* Purkinje and granule cells) may suggest that it has a role in motor coordination, while its presence in the CA1 to CA3 layers of the hippocampus raises the possibility that the CaR may also have some role(s) in cognitive functions, such as learning and memory, in this region of the central nervous system (12, 89). It is significant that the structurally and functionally homologous mGluRs are also present abundantly in the hippocampus, where they are also thought to play an important role in the generation of long-term potentiation and other processes related to cognition (74). The site of CaR immunostaining included nerve fibers and terminals in addition to a lesser degree of staining of neuronal cell bodies (*e.g.* in the hippocampal pyramidal cells). Cerebral arteries also displayed prominent CaR staining within a network of branching nerve fibers, a pattern resembling the distribution of neuronal nitric oxide synthase (12). The authors suggested a role for the CaR in detecting local changes in $[Ca^{2+}]_o$ concentration in the vicinity of neurons, including within synaptic clefts, thereby potentially regulating release of neurotransmitters or other aspects of neuronal function.

Alterations in serum calcium concentration are reflected in changes in the level of calcium in the cerebrospinal fluid. Could the CaR in the ventricular ependymal cells play a role in sensing changes in $[Ca^{2+}]_o$ or alterations in the gradient for $[Ca^{2+}]_o$ between its systemic level and that within the cerebrospinal fluid and/or brain, thereby contributing to regulation of the calcium concentration in the cerebrospinal fluid?

Finally, it is possible that the CaR in brain responds to ligands other than calcium (e.g. polycations, such as spermine) and may play a more general role as an ion sensor than in the parathyroid.

E. Regulation of the CaR

To date there have been relatively few studies on the regulation of the function and/or expression of the CaR. Activators of PKC blunt substantially the high $[Ca^{2+}]_o$ -elicited increases in inositol phosphate accumulation and $[Ca^{2+}]_i$ in bovine parathyroid cells (1, 73, 90). The presence of predicted sites for PKC-mediated phosphorylation on intracellular domains of the CaR suggest that these effects of altering PKC activity in parathyroid cells may be the result of PKC-mediated phosphorylation of one or more of these sites (9–12, 81). PKC could also, of course, exert additional effects on the PLC/IP₃ pathway by phosphorylating G protein(s), PLC, or other elements within the signal transduction pathway. Direct documentation of PKC-mediated phosphorylation of the CaR as well as determination of the functional consequences of removal of its PKC sites will enable more detailed dissection of the mechanisms involved and the importance of these sites in regulating the receptor's function. Activators of cAMP-dependent PKA, in contrast, have no apparent effect on high $[Ca^{2+}]_o$ -elicited elevations in the levels of inositol phosphates and $[Ca^{2+}]_i$ in bovine parathyroid cells (91).

Studies on the regulation of the level of expression of the CaR are likewise at an early stage. Treatment of rats with 1,25-(OH)₂D *in vivo* produced a modest increase in the level of CaR mRNA in the parathyroid in one study (92) but not in another (93), while chronic decreases or increases in serum calcium concentration had no effect on CaR mRNA levels in the parathyroid in either study. In contrast, elevated levels of $[Ca^{2+}]_o$ produced ~2-fold increases in CaR mRNA levels in AtT-20 cells (84). The physiological relevance of the latter effect is unclear, but it raises the possibility that the regulation of the receptor in tissues involved in calcium homeostasis, such as parathyroid cells, may differ from that in tissues such as the brain that are not directly involved in mineral ion metabolism. Of interest, calcium infusion does elevate ACTH levels *in vivo* in normal human volunteers (94, 95), suggesting that the presence of the CaR in AtT-20 cells is probably not simply due to ectopic expression of the receptor in a tumor cell line.

Bovine parathyroid cells placed into culture undergo a rapid and marked decrease in CaR transcript and protein expression, with 75–80% reductions in both parameters within 18 h and 36–48 h, respectively (87). This is accompanied by a progressive increase in the set point for $[Ca^{2+}]_o$ -regulated PTH secretion followed by a loss of the high $[Ca^{2+}]_o$ -mediated suppression of PTH release. Therefore, the level of expression of the CaR may play a key role in determining both parameters C and D in the four-parameter model of $[Ca^{2+}]_o$ -regulated PTH release (see Section III.A). In contrast, there is a dramatic up-regulation of CaR mRNA and protein expression in rat kidney during the first postnatal week of life (96). This correlates temporally with development of the TAL and reflects, therefore, the tissue-specific expression of the receptor in the kidney. Thus large changes

in the expression of the CaR can take place both *in vivo* and *in vitro*; however, the mechanisms underlying these changes as well as the determinants of the tissue-specific expression of the CaR remain to be clarified.

F. The CaR gene

The gene encoding the $[Ca^{2+}]_o$ -sensing receptor resides on chromosome 3 in humans (97). It is known to contain six exons comprising the coding region of the gene (13) as well as at least one additional upstream, noncoding exon (98). Five of these exons encode the amino-terminal, extracellular domain of the CaR, while the sixth codes for the entire transmembrane domain and the carboxy-terminal tail. This exonic structure provides indirect support for the concept (78) that the CaR represents the fusion of a seven membrane-spanning, signal-transducing motif with additional exons involved in determining the $[Ca^{2+}]_o$ -sensing properties of the receptor. Virtually nothing is presently known about the regions upstream of the translational start site of the receptor gene that are involved in regulating the level of expression of the CaR and the tissue specificity of its expression.

V. Are There Additional Forms of $[Ca^{2+}]_o$ -Sensors/Receptors?

It is probable that there are $[Ca^{2+}]_o$ -sensors or receptors in addition to the CaR. cDNAs encoding a putative $[Ca^{2+}]_o$ -sensor, which is related to the low density lipoprotein receptor superfamily and expressed on parathyroid and placental cells as well as on the cells of the kidney proximal tubules, have recently been isolated but have not yet been fully characterized in terms of their $[Ca^{2+}]_o$ -sensing properties (99, 100). The $[Ca^{2+}]_o$ -sensing mechanism mediating the effects of calcium on osteoclasts also differs from the CaR in its properties, generally only responding to levels of $[Ca^{2+}]_o$ higher than those needed to modulate the activity of the CaR in parathyroid and kidney cells and also being responsive to divalent cations (e.g. Cd²⁺) that have no effect on the cloned and expressed CaR (65, 66). In addition, much more work will be required to define fully the nature of the $[Ca^{2+}]_o$ -receptors/sensors that mediate the actions of $[Ca^{2+}]_o$ on the proliferation and differentiation of epithelial cells and other cell types (for review, see Ref. 1). It is entirely possible that there are diseases of $[Ca^{2+}]_o$ sensing that result from dysfunction of such additional $[Ca^{2+}]_o$ -receptors/sensors.

VI. Syndromes of Extracellular Calcium Resistance — Experiments of Nature Elucidating the Functions of the CaR *in Vivo*

A. Familial hypocalciuric hypercalcemia (FHH)

FHH (first named familial benign hypercalcemia or FBH) is a rare, heritable disorder of mineral metabolism, first described in 1972 (101). FHH is characterized by lifelong, moderate (<12 mg/dl) but generally asymptomatic hypercalcemia (102, 103). The degree of hypercalcemia is similar to that seen in primary hyperparathyroidism, but in FHH it is characteristically accompanied by inappropriately low urinary

calcium excretion (a calcium to creatinine clearance ratio of <0.01) (102, 103), along with inappropriately "normal" circulating levels of PTH in the face of hypercalcemia, which are often midnormal and may, in some cases, be in the lower part of the normal range. It is sometimes difficult to differentiate patients with FHH from those with mild primary hyperparathyroidism, particularly those individuals with hyperparathyroidism who have levels of PTH in the upper part of the normal range (104). Hypophosphatemia is variable, and mild elevations in serum magnesium level may also be present (102, 105, 106). Serum levels of 25(OH)D and 1,25-(OH)₂D are normal (107–109). The disorder is generally considered to be benign, as patients do not develop complications of hypercalcemia such as nephrolithiasis and NDI. Older patients with FHH, however, show a seemingly higher prevalence of chondrocalcinosis than would be expected in the normal population (102). Moreover, Marx and co-workers (102) described three cases of pancreatitis in their FHH kindreds. Two of these had known predisposing factors (102), however, and Law and Heath (103) have described an apparently increased incidence of gallstones in FHH. Thus it is probably unlikely that pancreatitis is a direct consequence of FHH (103, 110). Other nonspecific symptoms reported in earlier series (102) could not be attributed to FHH in later studies (103, 111), perhaps because ascertainment bias attributed symptoms to the disease process in probands that were not confirmed in more detailed analyses of entire kindreds. An unusual variant of the disease has been reported in a family from Oklahoma, which showed an age-related, progressive elevation in PTH levels along with hypophosphatemia and osteomalacia (112).

Individuals with FHH show abnormal parathyroid and renal responsiveness to extracellular calcium (113). It has been shown using intravenous calcium infusion that patients with FHH have an elevated set point for calcium-regulated PTH release (39, 40). That is, for any given level of serum calcium, FHH patients have a higher concentration of PTH than normal subjects. To suppress the circulating PTH level to 50% of its maximal value, these individuals require a serum calcium level slightly higher than that necessary to achieve a comparable degree of suppression in normals. These findings suggest that the parathyroid glands of patients with FHH show a defect in $[Ca^{2+}]_o$ sensing, exhibiting mild to moderate resistance to the inhibitory effects of extracellular calcium on PTH secretion.

Calcium handling by the kidney is also abnormal in individuals with FHH, with these patients failing to show a hypercalciuric response to hypercalcemia (102, 105). Indeed, they are often overtly hypocalciuric (102, 106). This phenomenon is PTH-independent, as these individuals remain hypocalciuric even after total parathyroidectomy (107, 114). Interestingly, some persons with FHH reported by Heath and co-workers (108) displayed normal gastrointestinal absorption of calcium and normal levels of 1,25-(OH)₂D despite being on a low calcium diet, suggesting a blunted homeostatic response to low calcium intake. By the late 1980s, convincing evidence had accumulated that FHH is a generalized disorder of calcium ion sensing with wide variability in its phenotypic presentation (106, 108).

1. *Genetics of FHH.* FHH is inherited as an autosomal dominant disorder, with greater than 90% penetrance (113). A severe variety of hyperparathyroidism that was found in some neonates from FHH families has been shown to represent most commonly the homozygous form of FHH and will be discussed in detail later. Over the past few years, several studies have confirmed that, in the majority of FHH families, the disease gene could be linked to a locus on the long arm of chromosome 3 (97). Genetic analysis of four families with FHH mapped the gene to band 3q21–24 (115), and subsequent studies have confirmed that the great majority of families with FHH exhibit this same genetic linkage. In one family, however, the disorder maps to chromosome 19 (band 19p13.3) (115). Furthermore, the disorder in the Oklahoma kindred does not show linkage to either chromosomes 3 or 19, confirming the genetic heterogeneity of FHH (116).

Since FHH seemed to be a disorder of calcium ion sensing, the need to look for abnormalities in the newly cloned CaR was obvious. Pollak *et al.* (13) mapped the human homolog of the bovine CaR gene to chromosome 3. Using ribonuclease A protection assays, they screened the human calcium-sensing receptor gene in affected individuals from three unrelated FHH families and found unique missense mutations in each family (*i.e.* point mutations in which a change in a single nucleotide base substitutes a new amino acid for the one originally coded for). None of the observed mutations were found in the genomic DNA of 50 normocalcemic individuals. All three mutations occurred in amino acid residues conserved between the human and bovine receptor genes. Two of the three families exhibited a mutation in the amino-terminal extracellular domain [R185Q (inadvertently described as R185E in the original paper) and E297K] of the CaR, which is thought to be involved in the binding of extracellular calcium. Based on homology with the mGluRs, the authors inferred that one of the extracellular mutations (R185Q) could potentially disrupt a salt bridge that contributes to the general structure of the binding pocket. The third family showed a mutation (R795W) in the third intracellular loop, which could be important for signal transduction, as seen in other G protein-linked receptors. This intracellular residue is also part of a consensus PKC phosphorylation site, and disruption of the latter could potentially also contribute to malfunction of the mutated receptor. Furthermore, *X. laevis* oocytes, injected with synthetic mRNA encoding a CaR engineered to contain the R795W missense mutation, exhibited a markedly blunted response to $[Ca^{2+}]_o$, $[Gd^{3+}]_o$, and neomycin, providing convincing evidence that this mutation impaired $[Ca^{2+}]_o$ sensing (13). Studies on other G protein-linked receptors have suggested the possibility that mutant receptors may interfere with the function of the wild type receptor on the cell surface or that mutant receptors could decrease the number of the receptors produced from the normal allele that are able to reach the cell surface (*i.e.* exert a dominant negative effect) (117). For example, a mutation in the human thromboxane A2 receptor appears to cause a familial bleeding disorder through a dominant negative mechanism (118). It is possible that this mechanism also contributes to abnormal $[Ca^{2+}]_o$ sensing, particularly in families with more severe hypercalcemia in heterozygotes.

The same group later studied eight families with FHH and reported missense mutations in the extracellular domain in five of them, each mutation producing a nonconservative amino acid change (119). However, three of the families studied did not show a detectable mutation within the coding sequence of the CaR protein, despite showing linkage to the FHH locus on chromosome 3. The methods that were employed to detect mutations are generally able to detect more than 95% of all point mutations, and the riboprobes used contained all intron and exon boundaries except those flanking exons 5 and 6. Therefore, failure to identify mutations that interrupt consensus splice sites, thereby disrupting processing of mRNA precursors, is highly unlikely. From these results, it appears that the defect in these three families could lie in noncoding sequences of the CaR gene (*i.e.* promoter or enhancer sequences). These defects may result in alterations in CaR gene transcription or expression that reduce the number of otherwise normal receptors reaching the cell surface and thereby produce functional "resistance" to $[Ca^{2+}]_o$. The authors' attempts to predict phenotype based on the different mutations observed were not successful, and one family member, who was homozygous for a mutation that in heterozygotes exhibited very mild hypercalcemia, suffered from NSHPT (119).

The list of CaR mutations reported in FHH is growing rapidly (13, 98, 119–122). So far, with few exceptions (98, 121), each family has been found to have its own unique mutation. Most of these mutations have been detected in the large extracellular domain (exons 2 and 3), and it is likely that they interfere with ligand binding. A recent report from Japan describes a mutation at codon 39 (P39A) in a hypercalcemic proband from a family with FHH (122). The proband was homozygous for this mutation, while the consanguineous parents, who had borderline elevated serum calcium concentrations, were heterozygous for the defect. Normocalcemic family members did not show any CaR mutation. Although the authors did not perform expression studies, this mutation appeared to result in a very mild defect in calcium sensing in heterozygotes and manifests overt hypercalcemia only in the homozygous state (122). Several clinical features of the proband are noteworthy in this regard. First, she had serum magnesium concentration ~50% above the upper limit of normal, which was similar to the degree of elevation of the serum calcium concentration (15–17 mg/dl). Moreover, the presence of hypermagnesemia supports the role of the CaR in contributing to "setting" $[Mg^{2+}]_o$. Finally, this report serves to highlight once again the genetic and phenotypic heterogeneity of the condition. The aforementioned reports also emphasize the importance of the extracellular domain in determining the responsiveness of the CaR to $[Ca^{2+}]_o$, a point that has been further established with the recognition of activating mutations in the same region causing hypocalcemia (14), as detailed later.

The remainder of the missense mutations described to date fall within exon 7 of the CaR gene, which encodes the transmembrane domains and cytoplasmic, carboxy-terminal tail of the receptor (13, 98, 119–122). A novel type of mutation has been described recently in two presumptively related families from Nova Scotia whose family members express either FHH in heterozygotes or NSHPT in homozygotes (120, 123).

The authors found an unusually large exon 7 in all affected individuals. This is due to the insertion of a 383-bp Alu repetitive sequence at codon 877. This sequence is in opposite orientation to the CaR gene and contains an exceptionally long poly A tract. Stop signals are present in all three reading frames within the Alu sequence, leading to a predicted truncation of the CaR protein after a long stretch of repeated phenylalanines (encoded by the triplet AAA), which would likely result in production of a nonfunctional protein that might well fail to reach the cell surface (120). Interestingly, this insertion seems to have expanded in a subsequent generation of this family (123). In addition to the functionally significant mutations described to this point, three benign polymorphisms that are not associated with any abnormalities in mineral ion homeostasis have been described in the carboxy-terminal tail of the CaR (121).

Based on these studies pertaining to FHH, we conclude the following: 1) FHH is characterized by genotypic and phenotypic heterogeneity. 2) Approximately one half to two thirds of all individuals with FHH, independent of their genetic locus, show inactivating mutations within the coding region of the recently cloned CaR (*e.g.* because of the rarity of the disorder arising from the other loci). However, each family generally has its own unique mutation. The majority of mutations occur within the extracellular domain and likely reduce the affinity of the receptor for extracellular Ca^{2+} . Some occur in transmembrane or cytoplasmic domains and might conceivably disrupt signal transduction. 3) Of the remaining one third to one half of individuals with FHH, the defect maps to the CaR gene in most, although there are no detectable mutations in the coding region. These individuals could have defects in promoter or enhancer sequences of the CaR gene, which have not yet been characterized in detail. 4) In some families the defect maps to other, as yet undefined, genes either on chromosome 19p or elsewhere. Because of the generally benign clinical course of patients with FHH, parathyroidectomy should not be performed except in very unusual clinical circumstances.

2. The CaR in FHH. Implications for regulation of parathyroid and renal function by $[Ca^{2+}]_o$

As discussed in preceding sections, several lines of evidence suggest that $[Ca^{2+}]_o$ exerts direct actions on parathyroid function as well as renal mineral and water handling, potentially through cellular mechanisms mediated by the CaR (32). The recognition of an abnormal CaR in FHH provides useful experiments in nature that have elucidated the role of the CaR in regulating the normal function of parathyroid and kidney.

CaR and $[Ca^{2+}]_o$ -regulated parathyroid function. As noted above, individuals with FHH have a modest increase in their set point for $[Ca^{2+}]_o$ -regulated PTH release (39, 40), while *in vitro* studies on parathyroid glands from two individuals with NSHPT exhibited much more severe resistance to $[Ca^{2+}]_o$ (2-fold or greater increases in the value of the set point) (124, 125). In addition, similar results have recently been reported in mice heterozygous or homozygous for targeted deletion of the murine homolog of the CaR gene (126). Thus, the alterations in $[Ca^{2+}]_o$ -regulated parathyroid function in FHH and NSHPT, as well as in mice homozygous or

heterozygous for "knockout" of the CaR, provide strong evidence that this receptor plays a key role in mediating the inhibitory effects of $[Ca^{2+}]_o$ on PTH secretion. Recent data also suggest that the CaR mediates the high $[Ca^{2+}]_o$ -induced reduction in expression of the PTH gene (127). It remains to be determined whether the $[Ca^{2+}]_o$ -sensing receptor is also responsible for additional effects of $[Ca^{2+}]_o$ on parathyroid function (for review, see Ref. 1), including inhibition of parathyroid cellular proliferation (the parathyroid hyperplasia in NSHPT provides indirect support for a role for the receptor in regulating this aspect of parathyroid function), alterations in the intracellular degradation of PTH as well as changes in cellular respiration, and the activity of the hexose monophosphate shunt.

The CaR and $[Ca^{2+}]_o$ regulation of the function of the TAL. As discussed above, FHH is characterized by excessive, inappropriate reabsorption of calcium ions by the kidney, which persists after parathyroidectomy (107, 114). Studies by Attie *et al.* (114) suggested that this abnormality in renal calcium handling takes place in the TAL, since a loop diuretic, ethacrynic acid, was able to increase urinary calcium excretion in individuals with FHH. The CaR is present in cells of the TAL on the basis of studies employing *in situ* hybridization, immunohistochemistry, and RT-PCR (88). Moreover, increases in the peritubular, but not luminal, levels of $[Ca^{2+}]_o$ to which the TAL is exposed inhibit reabsorption of calcium ions (57). Therefore, the alteration in tubular handling of calcium in FHH provides additional evidence that the CaR is responsible for high $[Ca^{2+}]_o$ -mediated modulation of Ca^{2+} reabsorption by this segment of the nephron. In effect, the TAL, like the parathyroid cell, is "resistant" to $[Ca^{2+}]_o$ in persons with FHH, limiting their capacity to up-regulate urinary calcium excretion in the face of an increase in $[Ca^{2+}]_o$, which, in turn, contributes to maintenance of their hypercalcemia.

The CaR and the control of $[Mg^{2+}]_o$ homeostasis. Individuals with FHH show mild hypermagnesemia due, in large part, to excessive magnesium reabsorption in the distal nephron (102). It has been shown that elevated peritubular levels of not only $[Ca^{2+}]_o$ but also $[Mg^{2+}]_o$ inhibit reabsorption of both ions in the TAL (57). Moreover, both the cloned parathyroid (BoPCaR) (9) and renal (RaKCaR) (10) CaRs respond to Mg^{2+} when expressed in *X. laevis* oocytes, albeit at somewhat higher concentrations than for Ca^{2+} . It is likely that the concentration of Mg^{2+} in the MTAL is higher than in the initial glomerular filtrate, since relatively little Mg^{2+} is reabsorbed before this region of the nephron (57). The concentrations achieved may be sufficient to be sensed by the CaR, which could thereby play an important, although as yet unproven, role in the regulation of renal Mg^{2+} reabsorption by the TAL. In FHH, the presence of abnormal CaRs with reduced activity leads to excessive Mg^{2+} reabsorption and consequent hypermagnesemia (102, 105). However, unlike Ca^{2+} reabsorption, the presence of PTH may be necessary for this inappropriate Mg^{2+} reabsorption in FHH, as this abnormality does not persist after parathyroidectomy (114). Of interest, mice that are heterozygous or homozygous for deletion of the CaR gene also show statistically significant increases in $[Mg^{2+}]_o$ relative to wild type mice, providing additional evidence for a role of the CaR in Mg^{2+} homeostasis (126).

The CaR and urinary concentrating ability. One of the well described effects of hypercalcemia on the kidney is to decrease maximal urinary concentrating ability, leading to hyposthenuria and, in some cases, overt NDI (59). This could occur in two ways: 1) elevations in extracellular calcium are known to inhibit NaCl absorption in the TAL (56), where the CaR is located (88). NaCl reabsorption in the TAL is crucial for generation of the countercurrent gradient that is needed for subsequent hydrosmotic water flow in the collecting duct under the influence of vasopressin. 2) Hypercalcemia likewise exerts an inhibitory effect on the action of vasopressin on water transport in the collecting duct (128). Recent data indicate that the CaR is present on the apical surface of cells of the collecting ducts, and, therefore, this effect on vasopressin action might also be CaR-mediated (96).

Individuals with FHH show an alteration in their maximal urinary concentrating ability in response to hypercalcemia, which provides additional indirect support for a role for the CaR in regulating water metabolism. In contrast to most hypercalcemic patients, persons with FHH concentrate their urine normally despite being hypercalcemic (129). This could occur because of deranged $[Ca^{2+}]_o$ sensing in either MTAL or collecting duct resulting from reduced activity of mutant CaRs. Impaired $[Ca^{2+}]_o$ sensing in the MTAL could lead to a reduced or absent inhibitory effect of hypercalcemia on NaCl reabsorption and consequent generation of the medullary countercurrent gradient. Alternatively or in addition, the presence of CaRs with reduced activity in the collecting duct could block the inhibitory action of hypercalcemia on vasopressin action in this nephron segment, likewise mitigating the development of NDI. In normal humans, the excretion of a dilute urine in the setting of hypercalciuria may be a protective mechanism that reduces the risk of developing nephrolithiasis and/or nephrocalcinosis when concentrated urine is being elaborated in states of dehydration (Fig. 4).

B. Neonatal severe hyperparathyroidism (NSHPT)

Neonatal primary hyperparathyroidism can be defined as symptomatic hypercalcemia with hyperparathyroid bone disease occurring in children under the age of 6 months (113). A recent analysis of 49 cases of neonatal severe primary hyperparathyroidism showed that most cases present at birth or within the first week of life (113). Failure to thrive, anorexia, constipation, and respiratory distress are common presenting features. Chest deformity, craniotables, dysmorphic facies, and anovaginal or rectovaginal fistulas have also been described (130–140). Respiratory complications due to thoracic deformity are a major cause of morbidity (113, 137).

Skeletal radiographs show marked demineralization, with fractures of the ribs and long bones, subperiosteal erosions, metaphyseal widening, and, occasionally, rickets (137). The hypercalcemia is severe, typically ranging from 14–20 mg/dl, although values as high as 30.8 mg/dl have been described (138). Interestingly, relative hypocalciuria has been documented in some cases, even in the absence of a family history of FHH (141). Serum PTH levels were high in the cases in which they were measured, although the degree of elevation can be mild (124, 141–144). Histological examina-

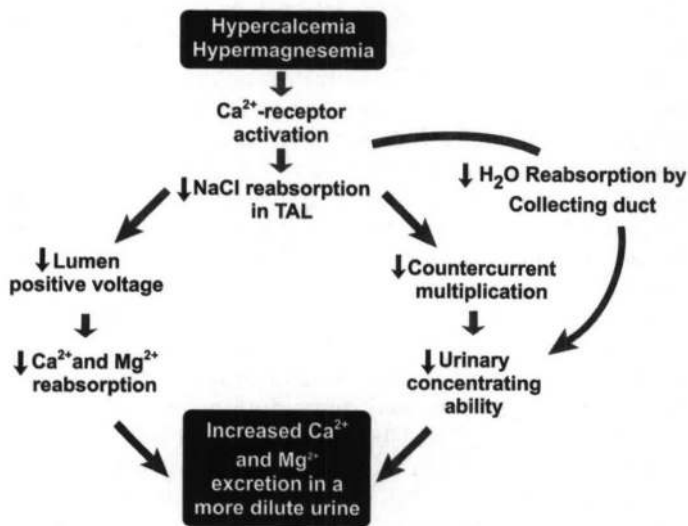


FIG. 4. Role of the CaR in mediating interactions between the renal handling of divalent cations and water. With a systemic calcium load, reduced Ca^{2+} reabsorption in the TAL increases the urinary Ca^{2+} concentration. The high $[\text{Ca}^{2+}]_o$ -mediated reduction in maximal urinary concentrating ability, however, mitigates the risk of deposition of calcium salts in the final urine. [Reproduced with permission from Brown EM, Hebert SC 1996 Novel insights into the physiology and pathophysiology of Ca^{2+} -sensing receptor. *Regul Pept Lett*, in press.]

tion of bone may show typical osteitis fibrosa cystica (136). The disorder is often fatal if aggressive medical and surgical management is delayed, although occasionally the disease runs a self-limited course, reverting to milder hypercalcemia by the age of 6 to 7 months with conservative treatment (137, 145). Initial management includes hydration and aggressive respiratory support. If the clinical condition is very severe or deteriorates during treatment, however, total parathyroidectomy with autotransplantation of part of one of the glands is recommended during the first month of life (113). Some authors recommend total parathyroidectomy with lifelong management of the resultant hypoparathyroidism (124, 136). There is a dramatic improvement after parathyroidectomy, and rapid healing of the skeletal changes is usually observed, even though the hypercalcemia recurs with less than total parathyroidectomy or after autotransplantation (113, 136). In all cases where parathyroid glands have been removed, they have been enlarged, sometimes up to 10 times the mass of the normal parathyroid glands in infants of this age. Histological examination has shown chief cell or water-clear cell hyperplasia (113, 136, 146).

1. Genetics of NSHPT: association with FHH. Since the initial descriptions of FHH and NSHPT, the occurrence of NSHPT in FHH families has been noted by several investigators (102, 135, 136, 147). In 1981 Marx *et al.* (102) reported their analysis of 15 kindreds with FHH, where they found that three patients from two families had severe primary hyperparathyroidism in the neonatal period. One of the explanations they offered was that NSHPT may be the homozygous form of FHH. The same group later studied a family with two children affected by NSHPT: their consanguineous parents had mild elevations in serum ionized calcium levels (total calcium levels were normal) as well as hypocalciuria, while multiple family members had borderline, sometimes inter-

mittent, hypercalcemia (147). These clinical findings lent support to the belief that NSHPT is actually the homozygous form of FHH and that some of the apparently sporadic occurrences of NSHPT could be due to failure to recognize very mild hypercalcemia in family members heterozygous for the defect. (It is also entirely possible that in some cases the alteration in $[\text{Ca}^{2+}]_o$ sensing by the mutant CaR could be so mild that rather than hypercalcemia *per se*, the serum calcium concentration is increased but remains within the normal range). After the localization of the gene for FHH to chromosome 3q in four families (97) and to chromosome 19p (115) in a single family, Pollak *et al.* (148) studied 11 families with FHH and demonstrated that the disease gene mapped to chromosome 3q2 in all of them (148). Four of these families had consanguineous marriages with affected NSHPT offspring. Analysis of genetic markers closely linked to the FHH gene showed that the pattern of inheritance of these markers was consistent with NSHPT representing the homozygous form of the same disease.

2. Mutations in the CaR in NSHPT. After the confirmation that NSHPT is the homozygous form of FHH, analysis of the CaR gene in families where both of these disorders coexisted confirmed that NSHPT patients inherit mutated CaR genes from both parents (13, 119, 120). Thus these patients do not possess a copy of the normal CaR, and as a result show much more severe hypercalcemia, elevated PTH, and parathyroid cell hyperplasia, as well as a markedly altered set point for calcium-regulated PTH secretion. All of these findings indicate a state of severe, albeit variable, parathyroid resistance to $[\text{Ca}^{2+}]_o$.

3. Spectrum of NSHPT. It is unlikely, however, that all cases of NSHPT represent the homozygous form of FHH. Cases of NSHPT have been reported to occur sporadically or in FHH families with only one affected parent (132, 143, 145). Indeed, recent studies have documented the occurrence of heterozygous, *de novo* CaR mutations in two sporadic cases (98) (*i.e.* caused by a single *de novo* CaR mutation in the child of normal parents), both of whom had evidence of hyperparathyroid bone disease, but with less severe hypercalcemia than is seen in typical NSHPT. This may provide an explanation for the observed clinical spectrum in NSHPT, which ranges from the typically severe, life-threatening manifestations (in patients homozygous for mutated CaR) to a more benign, possibly even self-limited form of the disorder (*i.e.* in patients with *de novo* heterozygous CaR mutations).

NSHPT that occurs in families where one parent has FHH could sometimes reflect unrecognized FHH in the other parent due to intermittent hypercalcemia. Another possible explanation for this observation could be the level of the maternal serum calcium level to which the developing fetus is exposed *in utero*. Calcium is actively transported across the placenta from the mother to the fetus, resulting in a higher concentration of serum calcium in the fetus (68). A mother with a normal serum calcium concentration would expose the fetus with FHH to a relatively hypocalcemic environment, which could conceivably "overstimulate" the fetal parathyroid glands and lead to the development of more

severe, "secondary" fetal/neonatal hyperparathyroidism superimposed upon the abnormal FHH-associated $[Ca^{2+}]_o$ sensing already present. This hypothesis is supported by the finding that cases of NSHPT with autosomal dominant inheritance have been reported in which the father had FHH while the mother was normocalcemic (106, 113, 136). Some of these cases have had a relatively benign, self-limited course, lending further credence to this theory. The possibility that an unidentified factor/protein may interact with the mutated FHH gene has also been suggested. Incomplete penetrance or uniparental disomy are other possible mechanisms.

These studies on NSHPT, which can, in effect, represent a "knockout" of the human CaR, provide the following perspective on the function of the receptor in humans: 1) They highlight the importance of the CaR in fetal and neonatal calcium homeostasis. 2) They suggest a possible role for the CaR in the regulation of parathyroid cellular proliferation, since there is marked parathyroid hyperplasia in NSHPT.

VII. Activating Mutations of the CaR: Autosomal Dominant Hypocalcemia

ADH refers to a familial syndrome characterized by mild hypocalcemia with few or no symptoms and detectable PTH levels (14). Since these patients appear to tolerate their hypocalcemia very well, it was hypothesized that they could have an altered set point for recognizing extracellular calcium, *i.e.* their parathyroid glands could be "hyperresponsive" to extracellular calcium (14, 149). Thus their calcium homeostatic system might be "reset" to maintain extracellular calcium at subnormal concentrations, the converse of FHH, where the system has been reset upward due to loss-of-function mutations in the CaR gene. Interestingly, in one such family, EDTA infusion was able to provoke PTH secretion, suggesting that the parathyroid glands were able to secrete PTH, but there was indeed a downward shift in the set point for $[Ca^{2+}]_o$ -regulated PTH secretion (149).

After the cloning of BoPCaR and the recognition of inactivating CaR mutations in families with FHH and NSHPT, and in light of the fact that activating mutations in other G protein-coupled receptors had been demonstrated in several disorders (32), it was thought that an "overactive" CaR in families with ADH could provide an explanation for this syndrome. Therefore, Pollak *et al.* (14) searched for activating mutations in the CaR in two families with ADH. In one of the probands, they demonstrated a missense mutation (E127A) in the CaR gene (14). This mutation was found in all hypocalcemic members of the family. Moreover, *X. laevis* oocytes expressing the mutant receptor exhibited severalfold higher levels of IP_3 at both low and high $[Ca^{2+}]_o$ compared with oocytes expressing the wild type receptor. They concluded that this mutation in the extracellular domain increased CaR activity "inappropriately" at low extracellular calcium concentrations, causing hypocalcemia in individuals heterozygous for such a mutation. No mutation in the CaR could be detected in the second family.

Since this report, another family with "autosomal domi-

nant hypoparathyroidism" has been reported to have a missense mutation in the extracellular domain of the CaR (150, 151). Pearce *et al.* (152) recently studied six families with ADH on the basis of clinical criteria. In addition to having asymptomatic hypocalcemia with detectable PTH levels, affected family members also had hypomagnesemia and hyperphosphatemia. The reason for classifying these families as ADH rather than familial isolated hypoparathyroidism was their unusual response to vitamin D treatment. Attempts to normalize their serum calcium levels with calcitriol resulted in marked hypercalciuria, which led to nephrocalcinosis and renal impairment in several cases (153). Some of them also complained of thirst and polyuria, possibly due to the development of NDI, with normalization of their serum calcium concentrations. It thus appears that the calcium homeostatic system in these individuals is adjusted to a lower than normal serum calcium concentration, and attempts to make them normocalcemic result in "hypercalcemic" manifestations (*i.e.* hypercalciuria, NDI). Four of these six families demonstrated mutations in the extracellular domain of the CaR that presumably produce inappropriate activation of the receptor at normal or even low levels of $[Ca^{2+}]_o$. Each of the six families with mutations so far reported have had unique mutations in the extracellular domain (two families in the latter study did not exhibit any mutations in the CaR gene), although several additional families have been reported in preliminary studies with autosomal dominant hypocalcemia associated with the presence of activating mutations within transmembrane domains of the receptor. Interestingly, the homozygous form of ADH, which would presumably present with more severe hypocalcemia, has not been identified.

The mechanism(s) by which these mutations could activate the CaR is presently unclear. Activating point mutations in other G protein-coupled receptors, like the TSH and LH receptors, are present in transmembrane domains and presumably enhance the processes of signal transduction or mimic the active state of the receptor after ligand binding if the activating mutation is truly ligand-independent (154, 155). In the CaR, it is likely that mutations within the extracellular domain enhance the affinity of the CaR for extracellular calcium or mimic the ligand-bound state of the extracellular domain, thereby imitating subsequent events in signal transduction at inappropriately low levels of extracellular calcium or even in the total absence of calcium.

These studies provide additional evidence for the importance of the CaR in setting the responsiveness of parathyroid and kidney to changes in $[Ca^{2+}]_o$. The parathyroid glands of individuals with ADH are hyperresponsive to extracellular calcium. In addition, normalization of serum calcium in these individuals leads to hypercalciuria that may be in excess of that expected in hypoparathyroid patients as individuals with ADH seem excessively prone to the complications of hypercalciuria and hypercalcemia even at low-to-normal levels of $[Ca^{2+}]_o$ (153). These observations complement the earlier ones in individuals with inactivating CaR mutations (FHH), in which there is inappropriate hypocalciuria despite hypercalcemia, and confirms a major role for the CaR in regulating tubular reabsorption of calcium. In addition, the apparently reciprocal abnormalities in water metabolism in

ADH and FHH provide additional indirect evidence that the CaR is intimately involved in coordinating renal handling of divalent cations and water.

Furthermore, these studies suggest the existence of ADH as an entity distinct from typical hypoparathyroidism and could lead to recognition of a larger number of cases with ADH, many of which may previously have been classified as mild cases of familial isolated hypoparathyroidism. This distinction is of great clinical importance, since treating ADH patients with calcitriol to normalize their serum calcium concentration may result in irreversible renal damage.

VIII. Alterations in CaR Function in Other Disease States

After the detection of mutations in the CaR in inherited disorders of calcium metabolism, it was of interest to determine whether there were alterations in CaR expression and/or function in acquired states of parathyroid dysfunction. One study has shown loss of heterozygosity encompassing the CaR locus in 10% of parathyroid adenomas (156). A detailed study of the CaR gene in various forms of primary and secondary hyperparathyroidism, including cases of adenoma, carcinoma, and hyperplastic glands from patients with sporadic or familial hyperparathyroidism or those with uremic hyperparathyroidism (157), however, failed to find any evidence for mutations in the CaR similar to those found in FHH. Furthermore, we found a substantial reduction in the immunoreactivity of the CaR protein in parathyroid adenomas as well as in uremic hyperparathyroidism as assessed by immunohistochemistry using a polyclonal antiserum raised against a peptide derived from the extracellular domain of the CaR (158). These alterations in CaR expression could possibly contribute to the altered set point for $[Ca^{2+}]_o$ -regulated PTH secretion seen in various forms of primary hyperparathyroidism as well as in severe uremic hyperparathyroidism (36).

IX. Diagnostic Implications

Detection of mutations in the CaR in patients with sporadic asymptomatic hypercalcemia could clearly be helpful in diagnosing FHH, although the size of the CaR-coding sequence makes this a substantial undertaking if direct sequencing were performed. More rapid screening procedures, such as denaturing gradient gel electrophoresis or the use of ribonuclease protection, could facilitate the process of screening for point mutations (98). A negative screen for mutations is not of much value, however, as not all FHH patients show CaR mutations, even when the disorder is linked to the chromosome 3 locus. Therefore, the diagnosis of FHH will likely continue to be established by the traditional approach of documenting an autosomal dominant inheritance of asymptomatic hypercalcemia in family members other than the proband that is accompanied by relative hypocalciuria (calcium/creatinine clearance ratio of <0.01). In our experience, it is not uncommon for patients with mild hyperparathyroidism to restrict their calcium intake voluntarily to the point where separation of their clearance ratios from those

encountered in FHH patients can be problematic. In this situation, particularly when first-degree relatives are not readily available for family screening, dietary supplementation to a total of 1000 mg elemental calcium/day can be very helpful, as it will usually increase urinary calcium excretion in patients with true primary hyperparathyroidism well above the levels seen with FHH. Other causes of apparent hypocalciuria in otherwise typical primary hyperparathyroidism are vitamin D deficiency, the use of hypocalciuric agents such as lithium or thiazides, and, occasionally, hypothyroidism. Hypercalcemia screening of as many family members as possible of patients with a provisional diagnosis of FHH is important since even borderline hypercalcemic patients may harbor mutations. Mutational screening of the spouse of a hypercalcemic patient could potentially be of value in predicting risk for NSHPT in the offspring. In conjunction with family screening for hypercalcemia, mutational analysis could also be useful for the diagnosis of cases presenting with hyperparathyroidism in the neonatal period, especially in the absence of a family history of hypercalcemia.

The recognition of cases with autosomal dominant hypocalcemia due to activating mutations of the CaR is important, since many of these individuals could be "over" treated with calcium/vitamin D with deleterious, sometimes irreversible, renal consequences if the disorder is not identified. The documentation of this condition requires careful clinical and genetic characterization. The clinician should carefully consider this diagnosis in individuals with the presumed diagnosis of familial hypoparathyroidism, particularly those who develop marked hypercalciuria and/or renal impairment when treated with vitamin D, so that complications such as nephrocalcinosis and renal failure can be prevented.

X. Therapeutic Implications

The recognition that $[Ca^{2+}]_o$ sensing by parathyroid cells might involve a cell surface, G protein-coupled receptor led to attempts directed at the development of drugs targeted at the receptor. Drugs that mimic the actions of high levels of $[Ca^{2+}]_o$ ("calcimimetics") on the receptor, for example, could conceivably be employed to lower PTH levels in states of hyperparathyroidism. Indeed, drugs of the latter type were developed more or less simultaneously with the cloning of the CaR. This made it possible to employ transfection of cells normally lacking the receptor to document that calcimimetics actually acted at the level of the CaR to enhance the degree of receptor activation at any given level of $[Ca^{2+}]_o$ (15). One such calcimimetic, NPS R-568, is the most promising drug developed to date and is currently undergoing clinical trials in hyperparathyroid states (159). NPS R-568 is a small organic molecule that inhibits PTH secretion, resulting in sustained hypocalcemia in normal rats. This hypocalcemia is thought to be due to a reduced Ca^{2+} efflux from bone resulting from decreased PTH secretion. In rats with mild chronic renal failure due to partial nephrectomy, NPS R-568 treatment for 4 weeks prevented the development of secondary hyperparathyroidism and an increase in bone formation rate, indicating its potential use in this condition in

humans (160). Preliminary data indicate that the drug also lowers both PTH and serum Ca^{2+} levels acutely in normal human subjects (159). Short-term use of the drug caused no discernible side effects. NPS R-568, therefore, could prove to be a major advance in the modalities available for treatment of both primary and secondary hyperparathyroidism.

Recent data suggest that NPS R-568 may inhibit not only PTH secretion but also hormonal synthesis. Cultured bovine parathyroid cells show a substantial reduction in PTH mRNA level when incubated overnight in the presence of NPS R-568 as well as an elevated level of $[\text{Ca}^{2+}]_o$ (127). The reduction in the level of PTH mRNA was greater than that seen with high calcium alone. The capacity of calcimimetics to suppress PTH synthesis could have broad implications both therapeutically as well as for understanding CaR-PTH interactions.

XI. Summary

The recent cloning of a $[\text{Ca}^{2+}]_o$ -sensing receptor from several different tissues in several species directly demonstrates that a variety of cells can directly recognize and respond to small changes in their ambient level of $[\text{Ca}^{2+}]_o$ through a G protein-coupled, cell surface receptor. This finding directly documents that $[\text{Ca}^{2+}]_o$ can act as an extracellular, first messenger in addition to subserving its better known role as an intracellular second messenger. Several of the tissues expressing the CaR are important elements in the calcium homeostatic system that have long been known to be capable of sensing $[\text{Ca}^{2+}]_o$, such as parathyroid and thyroidal C cells. The presence of the receptor in the kidney, however, provides strong evidence that several of the long-recognized but poorly understood direct actions of $[\text{Ca}^{2+}]_o$ on renal function could be mediated by the CaR. These actions include the up-regulation of urinary calcium and magnesium excretion in the setting of hypercalcemia, which complements the indirect inhibition of renal tubular reabsorption of calcium that results from high $[\text{Ca}^{2+}]_o$ -mediated inhibition for PTH secretion. The impaired renal concentrating capacity in hypercalcemia is likely a manifestation of a homeostatically important interaction between the regulation of renal calcium and water handling that reduces the risk of pathological deposition of calcium in the kidney when there is a need to dispose of excess calcium in the urine. In this regard, the availability of human syndromes of $[\text{Ca}^{2+}]_o$ "resistance" or "overresponsiveness" due to loss-of-function or gain-of-function mutations in the CaR, respectively, have provided useful experiments in nature that have clarified the importance of the receptor in both abnormal and normal physiology. Much remains to be learned, however, about the role of the CaR in locations, such as the brain, where it likely responds to local rather than systemic levels of $[\text{Ca}^{2+}]_o$. In such sites, it may represent an important modulator of neuronal function, responding to $[\text{Ca}^{2+}]_o$ as a neuromodulator or even neurotransmitter. The development of therapeutics that either activate or inhibit the function of the CaR may be useful for treating a variety of conditions in which the receptor is either under- or overactive. Finally, it would not be surprising to discover additional receptors for $[\text{Ca}^{2+}]_o$ or for other

ions (the CaR may, in fact, be an important $[\text{Mg}^{2+}]_o$ -sensor) that could function abnormally in certain disease states and be amenable to pharmacological manipulation with ion receptor-based therapeutics.

References

1. Brown EM 1991 Extracellular Ca^{2+} sensing, regulation of parathyroid cell function, and role of Ca^{2+} and other ions as extracellular (first) messengers. *Physiol Rev* 71:371-411
2. Pietrobon D, DiVirgilio F, Pozzan T 1990 Structural and functional aspects of calcium homeostasis in eukaryotic cells. *Eur J Biochem* 193:599-622
3. Stewart AF, Broadus AE 1987 Mineral Metabolism. In: Felig P, Baxter JD, Broadus AE, Frohman LA (eds) *Endocrinology, Metabolism*, ed 2. McGraw-Hill, New York, pp 1317-1453
4. Kurokawa K 1994 The kidney and calcium homeostasis. *Kidney Int* 45[Suppl 44]:S97-S105
5. Berridge MJ, Irvine RF 1984 Inositol trisphosphate, a novel second messenger in cellular signal transduction. *Nature* 312:315-321
6. Nemeth EF, Scarpa A 1986 Cytosolic Ca^{++} and the regulation of secretion in parathyroid cells. *FEBS Lett* 203:15-19
7. Shoback DM, Membreno LA, McGhee JG 1988 High calcium and other divalent cations increase inositol trisphosphate in bovine parathyroid cells. *Endocrinology* 123:382-389
8. Juhlin C, Lundgren S, Johansson H, Lorentzen J, Rask L, Larsson E, Rastad J, Akerstrom G, Klareskog L 1990 500 Kilodalton calcium sensor regulating cytoplasmic Ca^{2+} in cytotrophoblast cells of human placenta. *J Biol Chem* 265:8275-8279
9. Brown EM, Gamba G, Riccardi D, Lombardi D, Butters R, Kifor O, Sun A, Hediger MA, Lytton J, Hebert SC 1993 Cloning and characterization of an extracellular Ca^{2+} -sensing receptor from bovine parathyroid. *Nature* 366:575-580
10. Riccardi D, Park J, Lee W-S, Gamba G, Brown EM, Hebert SC 1995 Cloning and functional expression of a rat kidney extracellular calcium-sensing receptor. *Proc Natl Acad Sci USA* 92:131-135
11. Garrett JE, Tamir H, Kifor O, Simin RT, Rogers KV, Mithal A, Gagel RF, Brown EM 1995 Calcitonin-secreting cells of the thyroid gland express an extracellular calcium-sensing receptor gene. *Endocrinology* 136:5202-5211
12. Ruat M, Molliver ME, Snowman AM, Snyder SH 1995 Calcium sensing receptor: molecular cloning in rat and localization to nerve terminals. *Proc Natl Acad Sci USA* 92:3161-3165
13. Pollak MR, Brown EM, Chou Y-H, Hebert SC, Marx SJ, Steinmann B, Levi T, Seidman CE, Seidman JG 1993 Mutations in the human Ca^{2+} -sensing receptor gene cause familial hypocalcemic hypercalcemia and neonatal severe hyperparathyroidism. *Cell* 75:1297-1303
14. Pollak MR, Brown EM, Estep HL, McLaine PN, Kifor O, Park J, Hebert SC, Seidman CE, Seidman JG 1994 Autosomal dominant hypocalcaemia caused by a Ca^{2+} -sensing receptor gene mutation. *Nature Genet* 8:303-307
15. Steffey ME, Fox J, VanWagenen BC, Delmar EG, Balandrin MF, Nemeth EF 1993 Calcimimetics: structurally and mechanistically novel compounds that inhibit hormone secretion from parathyroid cells. *J Bone Miner Res* 8[Suppl 1]:S175 (Abstract)
16. Morrissey JJ, Hamilton JW, MacGregor RR, Cohn DV 1980 The secretion of parathormone fragments 34-84 and 37-84 by dispersed porcine parathyroid cells. *Endocrinology* 107:164-171
17. Hanley DA, Takatsuki K, Sultan JM, Schneider AB, Sherwood LM 1978 Direct release of parathyroid hormone fragments from functioning bovine parathyroid glands *in vitro*. *J Clin Invest* 62:1247-1254
18. Naveh-Manly T, Silver J 1990 Regulation of parathyroid hormone gene expression by hypocalcemia, hypercalcemia and vitamin D in the rat. *J Clin Invest* 86:1313-1319
19. Hawa NS, O'Riordan JL, Farrow SM 1993 Post-transcriptional regulation of bovine parathyroid hormone synthesis. *J Mol Endocrinol* 10:43-49
20. Rodan GA, Martin TJ 1981 Role of osteoblasts in hormonal control of bone resorption - a hypothesis. *Calcif Tissue Int* 33:349-351

21. Marx SJ, Liberman UA, Eil CA 1983 Calciferols: actions and deficiencies in actions. *Vitam Horm* 40:235-308
22. Raisz LG 1965 Regulation by calcium of parathyroid growth and secretion *in vitro*. *Nature* 44:103-110
23. Capen CC, Rowland GN 1968 Ultrastructural evaluation of the parathyroid glands of young cats with experimental hyperparathyroidism. *Z Zellforsch Mikrosk Anat* 90:495-506
24. Lopez-Hilker S, Galceran T, Chan Y-L, Rapp N, Martin KJ, Slatopolsky E 1986 Hypocalcemia may not be essential for the development of secondary hyperparathyroidism in chronic renal failure. *J Clin Invest* 78:1097-1102
25. Slatopolsky E, Berkoben M, Kelber J, Brown A, Delmez J 1992 Effects of calcitriol and non-calcemic vitamin D analogs on secondary hyperparathyroidism. *Kidney Int* 42[Suppl 38]:S43-S49
26. Silver J, Russell J, Sherwood LM 1985 Regulation by vitamin D metabolites of messenger ribonucleic acid for preproparathyroid hormone in isolated bovine parathyroid cells. *Proc Natl Acad Sci USA* 82:4270-4273
27. Kremer R, Bolivar I, Goltzman D, Hendy GN 1989 Influence of calcium and 1,25-dihydroxycholecalciferol on proliferation and proto-oncogene expression in primary cultures of bovine parathyroid cells. *Endocrinology* 125:935-941
28. Nygren P, Larsson R, Johansson H, Ljunghall S, Rastad J, Akerstrom G 1988 1,25(OH)₂D inhibits hormone secretion and proliferation but not functional dedifferentiation of cultured bovine parathyroid cells. *Calcif Tissue Int* 43:213-218
29. Cantley LK, Russell J, Lettieri D, Sherwood LM 1985 1,25-Dihydroxyvitamin D₃ suppresses parathyroid hormone secretion from bovine parathyroid cells in tissue culture. *Endocrinology* 117:2114-2119
30. Russell J, Lettieri D, Sherwood LM 1983 Direct regulation by calcium of cytoplasmic messenger ribonucleic acid coding for preproparathyroid hormone in isolated bovine parathyroid cells. *J Clin Invest* 72:1851-1855
31. Austin LA, Heath III H 1981 Calcitonin: physiology and pathophysiology. *N Engl J Med* 304:269-278
32. Brown EM, Pollak M, Seidman CE, Seidman JG, Chou Y-HW, Riccardi D, Hebert SC 1995 Calcium-ion-sensing cell-surface receptors. *N Engl J Med* 333:234-240
33. Brent GA, LeBoff MS, Seely EW, Conlin PR, Brown EM 1988 Relationship between the concentration and rate of change of calcium and serum intact parathyroid hormone levels in normal humans. *J Clin Endocrinol Metab* 67:944-950
34. Mayer GP, Hurst JG 1978 Sigmoidal relationship between parathyroid hormone secretion rate and plasma calcium concentration in calves. *Endocrinology* 102:1036-1042
35. Habener JF, Potts Jr JT 1976 Relative effectiveness of magnesium and calcium on the secretion and biosynthesis of parathyroid hormone *in vitro*. *Endocrinology* 98:197-202
36. Brown EM 1983 Four parameter model of the sigmoidal relationship between parathyroid hormone release and extracellular calcium concentration in normal and abnormal parathyroid tissue. *J Clin Endocrinol Metab* 56:572-581
37. Felsenfeld AJ, Llach F 1993 Parathyroid gland function in chronic renal failure. *Kidney Int* 43:771-789
38. Parfitt AM 1987 Bone and plasma calcium homeostasis. *Bone* 8 [Suppl 1]:1-8
39. Auwerx J, Demedts M, Bouillon R 1984 Altered parathyroid set point to calcium in familial hypocalciuric hypercalcemia. *Acta Endocrinol (Copenh)* 106:215-218
40. Khosla S, Ebeling PR, Firek AF, Burritt MM, Kao PC, Heath III H 1993 Calcium infusion suggests a "set-point" abnormality of parathyroid gland function in familial benign hypercalcemia and more complex disturbances in primary hyperparathyroidism. *J Clin Endocrinol Metab* 76:715-720
41. LeBoff MS, Shoback D, Brown EM, Thatcher J, Leombruno R, Beaudoin D, Henry M, Wilson R, Pallotta J, Marynick S, Stock J, Leight G 1985 Regulation of parathyroid hormone release and cytosolic calcium by extracellular calcium in dispersed and cultured bovine and pathological human parathyroid cells. *J Clin Invest* 75:49-57
42. Habener JF, Rosenblatt M, Potts Jr JT 1984 Parathyroid hormone: biochemical aspects of biosynthesis, secretion, action, and metabolism. *Physiol Rev* 64:985-1053
43. Brown EM, Enyedi P, LeBoff M, Rothberg J, Preston J, Chen C 1987 High extracellular Ca²⁺ and Mg²⁺ stimulate accumulation of inositol phosphates in bovine parathyroid cells. *FEBS Lett* 218:113-118
44. Muff R, Nemeth EF, Haller-Brem S, Fischer JA 1988 Regulation of hormone secretion and cytosolic Ca²⁺ by extracellular Ca²⁺ in parathyroid cells and C-cells: role of voltage-sensitive Ca²⁺ channels. *Arch Biochem Biophys* 265:128-135
45. Pocotte SL, Ehrenstein G, Fitzpatrick LA 1995 Role of calcium channels in parathyroid hormone secretion. *Bone* 16:S365-372
46. Fajtova VT, Quinn SJ, Brown EM 1991 Cytosolic calcium responses of single rMTC 44-2 cells to stimulation with external calcium and potassium. *Am J Physiol* 261:E151-158
47. Chen CJ, Barnett JV, Congo DA, Brown EM 1989 Divalent cations suppress 3',5'-adenosine monophosphate accumulation by stimulating a pertussis toxin-sensitive guanine nucleotide binding protein in cultured bovine parathyroid cells. *Endocrinology* 124:233-239
48. Hawkins D, Enyedi P, Brown EM 1989 The effects of high extracellular Ca²⁺ and Mg²⁺ concentrations on the levels of inositol 1,3,4,5-tetrakisphosphate in bovine parathyroid cells. *Endocrinology* 124:838-844
49. Brown EM, Fuleihan GE-H, Chen CJ, Kifor O 1990 A comparison of the effects of divalent and trivalent cations on parathyroid hormone release, 3',5'-cyclic-adenosine monophosphate accumulation, and the levels of inositol phosphates in bovine parathyroid cells. *Endocrinology* 127:1064-1071
50. Ridefelt P, Hellman P, Wallfelt C, Akerstrom G, Rastad J, Gylfe E 1992 Neomycin interacts with Ca²⁺-sensing of normal and adenomatous parathyroid cells. *Mol Cell Endocrinol* 83:211-218
51. Brown EM, Katz C, Butters R, Kifor O 1991 Polyarginine, polylysine, and protamine mimic the effects of high extracellular calcium concentrations on dispersed bovine parathyroid cells. *J Bone Miner Res* 6:1217-1225
52. Brown EM, Butters R, Katz C, Kifor O, Fuleihan GE 1992 A comparison of the effects of concanavalin A and TPA on the modulation of parathyroid function by extracellular calcium and neomycin in dispersed bovine parathyroid cells. *Endocrinology* 130:3143-3151
53. Weisinger JR, Favus MJ, Langman CB, Bushinsky DA 1989 Regulation of 1,25-dihydroxyvitamin D₃ by calcium in the parathyroidectomized, parathyroid hormone replete rat. *J Bone Miner Res* 4:929-935
54. Takaichi K, Kurokawa K 1986 High Ca²⁺ inhibits peptide hormone-dependent cAMP production specifically in thick ascending limbs of Henle. *Miner Electrolyte Metab* 12:342-346
55. Takaichi K, Kurokawa K 1988 Inhibitory guanosine triphosphate-binding protein-mediated regulation of vasopressin action in isolated single medullary tubules of mouse kidney. *J Clin Invest* 82:1437-1444
56. Hebert SC, Andreoli TE 1984 Control of NaCl transport in the thick ascending limb. *Am J Physiol* 246:F745-F756
57. Quamme GA 1989 Control of magnesium transport in the thick ascending limb. *Am J Physiol* 256:F197-F210
58. Clemens TL, McGlade SA, Garrett KP, Craviso GL, Hendy GN 1989 Extracellular calcium modulates vitamin D-dependent calbindin-D_{28k} gene expression in chick kidney cells. *Endocrinology* 124:1582-1584
59. Suki WN, Eknayan G, Rector Jr FC, Seldin DW 1969 The renal diluting and concentrating mechanism in hypercalcemia. *Nephron* 6:50-61
60. Fray JCS, Park CS, Valentine AN 1987 Calcium and the control of renin secretion. *Endocr Rev* 8:53-93
61. Edvall CA 1958 Renal function in hyperparathyroidism: a clinical study of 30 cases with special reference to selective renal clearance and renal vein catheterization. *Acta Chir Scand* 229:1-54
62. Humes HD, Ichikawa I, Troy JL, Brenner BM 1978 Evidence for a parathyroid hormone-dependent influence of calcium on the glomerular ultrafiltration coefficient. *J Clin Invest* 61:32-40
63. Fried RM, Tashjian Jr AH 1986 Unusual sensitivity of cytosolic free

- Ca²⁺ to changes in extracellular Ca²⁺ in rat C-cells. *J Biol Chem* 261:7669-7674
64. Scherubl H, Schultz G, Hescheler J 1991 Electrophysiological properties of rat calcitonin-secreting cells. *Mol Cell Endocrinol* 82:293-301
 65. Zaidi M, Datta HK, Patchell A, Moonga B, MacIntyre I 1989 "Calcium-activated" intracellular calcium elevation: a novel mechanism of osteoclast regulation. *Biochem Biophys Res Commun* 163:1461-1465
 66. Malgaroli A, Meldolesi J, Zallone AZ, Teti A 1989 Control of cytosolic free calcium in rat and chicken osteoclasts. The role of extracellular calcium and calcitonin. *J Biol Chem* 264:14342-14347
 67. Hartle JE, Arthur JM, Raymond JT, Quarles LD 1994 Evidence for a cation sensing receptor in osteoblasts coupled to G-protein activation and DNA synthesis. *J Bone Miner Res* 9:S159 (Abstract 156)
 68. Rodda CP, Caple IW, Martin TJ 1992 Role of PTHrP in fetal and neonatal physiology. In: Halloran BP, Nissenson RA (eds) *Parathyroid Hormone Related Protein: Normal Physiology and Its Role in Cancer*. CRC Press, Boca Raton, FL, pp 169-196
 69. Hellman P, Ridefelt P, Juhlin C, Akerstrom G, Rastad J, Gylfe E 1992 Parathyroid-like regulation of parathyroid-hormone-related protein release and cytoplasmic calcium in cytotrophoblast cells of the human placenta. *Arch Biochem Biophys* 293:174-180
 70. Hennings H, Michael D, Cheng C, Steinert P, Holbrook K, Yuspa SH 1980 Calcium regulation of growth and differentiation of mouse epidermal cells in culture. *Cell* 19:245-254
 71. Racke FK, Hammerland JG, Dubyak GR, Nemeth EF 1993 Functional expression of the parathyroid cell calcium receptor in *Xenopus* oocytes. *FEBS Lett* 333:132-136
 72. Chen T-H, Pratt SA, Shoback DM 1994 Injection of bovine parathyroid poly(A)⁺ RNA into *Xenopus* oocytes confers sensitivity to extracellular calcium. *J Bone Miner Res* 9:293-300
 73. Kifor O, Congo D, Brown EM 1990 Phorbol esters modulate the high Ca²⁺-stimulated accumulation of inositol phosphates in bovine parathyroid cells. *J Bone Miner Res* 5:1003-1011
 74. Nakanishi S 1992 Molecular diversity of glutamate receptors and implications for brain function. *Science* 258:597-603
 75. Takahashi K, Tsuchida K, Tanabe Y, Masu M, Nakanishi S 1993 Role of the large extracellular domain of metabotropic glutamate receptors in agonist selectivity determination. *J Biol Chem* 268:19341-19345
 76. Hammerland LG, Krapcho KJ, Alasti N, Garrett JE, Capuano IV, Hung BCP, Fuller FH 1995 Cation binding determinants of the calcium receptor revealed by functional analysis of chimeric receptors and a deletion mutant. *J Bone Miner Res* 10:S156 (Abstract 69)
 77. O'Hara PJ, Sheppard PO, Thogersen H, Venezia D, Haldeman BA, McGrane V, Houamed KM, Thomsen C, Gilbert TL, Mulvihill ER 1993 The ligand binding domain in metabotropic glutamate receptors is related to bacterial periplasmic binding proteins. *Neuron* 11:41-52
 78. Conklin BR, Bourne HR 1994 Homeostatic signals: marriage of the flytrap and the serpent. *Nature* 367:22
 79. Sharff AJ, Rodseth LE, Spurlino JC, Quioco FA 1992 Crystallographic evidence for a large ligand-induced hinge-twist motion between the two domains of the maltodextrin binding protein involved in active transport and chemotaxis. *Biochemistry* 31:10657-10663
 80. Garrett JE, Capuano IV, Hammerland LG, Hung BCP, Brown EM, Hebert SC, Nemeth EF, Fuller F 1995 Molecular cloning and functional expression of human parathyroid calcium receptor cDNAs. *J Biol Chem* 270:12919-12925
 81. Aida K, Koishi S, Tawata M, Onaya T 1995 Molecular cloning of a putative Ca²⁺-sensing receptor cDNA from human kidney. *Biochem Biophys Res Commun* 214:524-529
 82. Varrault A, Pena MS, Goldsmith PK, Mithal A, Brown EM, Spiegel AM 1995 Expression of G-protein alpha-subunits in bovine parathyroid. *Endocrinology* 136:4390-4396
 83. Moriarty TM, Sealoff SC, Carty DJ, Roberts JL, Iyengar R, Landau EM 1989 Coupling of exogenous receptors to phospholipase C in *Xenopus laevis* oocytes through pertussis toxin-sensitive and -insensitive pathways. Cross-talk between heterotrimeric G-proteins. *J Biol Chem* 264:13524-13530
 84. Emanuel RL, Adler GK, Krapcho K, Fuller F, Quinn SJ, Brown EM 1996 Calcium-sensing receptor expression, regulation in AtT-20 pituitary cell line. *Mol Endocrinol* 10:555-565
 85. Rogers KV, Dunn CK, Hebert SC, Brown EM, Nemeth EF 1995 Pharmacological comparison of bovine parathyroid, human parathyroid, and rat kidney calcium receptors expressed in HEK 293 cells. *J Bone Miner Res* 10:S483 (Abstract T516)
 86. Firsov D, Aarab L, Mandon B, Siaume-Perez S, de Rouffignac C, Chabardes D 1995 Arachidonic acid inhibits hormone-stimulated cAMP accumulation in the medullary thick ascending limb of the rat kidney by a mechanism sensitive to pertussis toxin. *Pflügers Arch* 429:636-646
 87. Mithal A, Kifor O, Kifor I, Vassilev P, Butters R, Krapcho K, Simin R, Fuller F, Hebert SC, Brown EM 1995 The reduced responsiveness of cultured bovine parathyroid cells to extracellular Ca²⁺ is associated with marked reduction in the expression of extracellular Ca²⁺-sensing receptor mRNA and protein. *Endocrinology* 136:3087-3092
 88. Riccardi D, Plotkin MD, Lee W-S, Lee K, Segre GV, Brown EM, Hebert SC 1995 Colocalization of the Ca²⁺-sensing receptor and PTH/PTHrP receptor in rat kidney. *J Am Soc Nephrol* 6:954 (Abstract)
 89. Dunn CK, Brown EM, Hebert SC, Rogers KV, Localization of calcium receptor mRNA in the adult rat central nervous system by *in situ* hybridization. Abstracts of the Neuroscience Meeting, 1994, p 1061 (Abstract)
 90. Racke FK, Nemeth EF 1993 Cytosolic calcium homeostasis in bovine parathyroid cells and its modulation by protein kinase C. *J Physiol (Lond)* 468:141-162
 91. Brown EM, Redgrave J, Thatcher JT 1984 Effect of the phorbol ester TPA on PTH secretion. Evidence for a role for protein kinase C in the control of PTH release. *FEBS Lett* 175:72-75
 92. Zhong M, Finch J, McCracken R, McCracken R, Morrissey J, Slatopolsky E, Brown AJ 1994 Rat parathyroid gland calcium receptor: regulation by 1,25-dihydroxyvitamin D₃ but not by dietary hyperparathyroidism. *J Bone Miner Res* 9[Suppl 1]:S217 (Abstract A436)
 93. Rogers KV, Dunn CK, Conklin RL, Hadfield S, Petty BA, Brown EM, Hebert SC, Nemeth EF, Fox J 1995 Calcium receptor messenger ribonucleic acid levels in the parathyroid glands and kidney of vitamin D deficient rats are not regulated by plasma calcium or 1,25-dihydroxyvitamin D₃. *Endocrinology* 136:499-504
 94. Isaac R, Raymond J-P, Rainfray M, Ardaillou R 1984 Effects of an acute calcium load on plasma ACTH, cortisol, aldosterone and renin activity in man. *Acta Endocrinol (Copenh)* 105:251-257
 95. Fuleihan GE-H, Brown EM, Gleason R, Scott J, Adler G 1996 Calcium modulation of adrenocorticotrophic hormone levels in women. *J Clin Endocrinol Metab* 81:932-936
 96. Baum MA, Chattopadhyay N, Brown EM, Ruddy MK, Hosselet C, Riccardi D, Hebert S, Harris HW 1995 Perinatal expression of aquaporins-2 and 3 and the calcium receptor in developing rat kidney collecting ducts. *J Am Soc Nephrol* 6:319 (Abstract)
 97. Chou Y-H, Brown EM, Levi T, Crowe G, Atkinson AB, Arnqvist HJ, Toss G, Fuleihan GE, Seidman JG, Seidman CE 1992 The gene responsible for familial hypocalciuric hypercalcemia maps to chromosome 3 in four unrelated families. *Nature Genet* 1:295-300
 98. Pearce SHS, Trump D, Wooding C, Besser GM, Chew SL, Grant DB, Heath DA, Hughes IA, Paterson CR, Whyte MP, Thakker RV 1995 Calcium-sensing receptor mutations in familial benign hypercalcemia and neonatal hyperparathyroidism. *J Clin Invest* 96:2683-2692
 99. Lundgren S, Hjalm G, Hellman P, Ek B, Juhlin C, Rastad J, Klareskog L, Akerstrom G, Rask L 1994 A protein involved in calcium sensing of the human parathyroid and placental cytotrophoblast cells belongs to the LDL-receptor protein superfamily. *Exp Cell Res* 212:344-350
 100. Saito A, Pietromonaco S, Loo AK, Farquhar MG 1994 Complete cloning and sequencing of rat gp330/"megalin," a distinctive member of the low density lipoprotein receptor gene family. *Proc Natl Acad Sci USA* 91:9725-9729
 101. Foley Jr TP, Harrison HC, Arnaud CD, Harrison HE 1972 Familial benign hypercalcemia. *J Pediatr* 81:1060-1067
 102. Marx SJ, Attie MF, Levine MA, Spiegel AM, Downs Jr RW, Lasker

- RD 1981 The hypocalciuric or benign variant of familial hypercalcemia: clinical and biochemical features in fifteen kindreds. *Medicine (Baltimore)* 60:397-412
103. Law Jr WM, Heath III H 1985 Familial benign hypercalcemia (hypocalciuric hypercalcemia). Clinical and pathogenetic studies in 21 families. *Ann Intern Med* 102:511-519
 104. Gunn IR, Wallace JR 1992 Urine calcium and serum ionized calcium, total calcium and parathyroid hormone concentrations in the diagnosis of primary hyperparathyroidism and familial benign hypercalcemia. *Ann Clin Biochem* 29:52-58
 105. Marx SJ, Spiegel AM, Brown EM, Koehler JO, Gardner DG, Brennan MF, Aurbach GD 1978 Divalent cation metabolism. Familial hypocalciuric hypercalcemia vs. typical primary hyperparathyroidism. *Am J Med* 65:235-242
 106. Heath III H 1989 Familial benign (hypocalciuric) hypercalcemia. A troublesome mimic of primary hyperparathyroidism. *Endocrinol Metab Clin North Am* 18:723-740
 107. Davies M, Adams PH, Lumb GA, Berry JL, Loveridge N 1984 Familial hypocalciuric hypercalcemia: evidence for continued enhanced renal tubular reabsorption of calcium following total parathyroidectomy. *Acta Endocrinol (Copenh)* 106:499-504
 108. Law Jr WM, Bollman S, Kumar R, Heath III H 1984 Vitamin D metabolism in familial benign hypercalcemia (hypocalciuric hypercalcemia) differs from that in primary hyperparathyroidism. *J Clin Endocrinol Metab* 58:744-747
 109. Kristiansen JH, Rodbro P, Christiansen C, Brochner Mortensen J, Carl J 1985 Familial hypocalciuric hypercalcemia. II. Intestinal calcium absorption and vitamin D metabolism. *Clin Endocrinol (Oxf)* 23:511-515
 110. Stuckey BGA, Gutteridge DH, Kent GN, Reed WD 1990 Familial hypocalciuric hypercalcemia and pancreatitis: no causal link proven. *Aust NZ J Med* 20:718-719
 111. Heath DA 1989 Familial benign hypercalcemia. *Trends Endocrinol Metab* 1:6-9
 112. McMurtry CT, Schranck FW, Walkenhorst DA, Murphy WA, Kocher DB, Teitelbaum SL, Rupich RC, Whyte MP 1992 Significant developmental elevation in serum parathyroid hormone levels in a large kindred with familial benign (hypocalciuric) hypercalcemia. *Am J Med* 93:247-258
 113. Heath DA 1994 Familial hypocalciuric hypercalcemia. In: Bilezikian JP, Marcus R, Levine MA (eds) *The Parathyroids*, Raven Press, New York, pp 699-710
 114. Attie MF, Gill Jr JR, Stock JL, Spiegel AM, Downs Jr RW, Levine MA, Marx SJ 1983 Urinary calcium excretion in familial hypocalciuric hypercalcemia. Persistence of relative hypocalciuria after induction of hypoparathyroidism. *J Clin Invest* 72:667-676
 115. Heath III H, Jackson CE, Otterud B, Leppert MF 1993 Genetic linkage analysis in familial benign (hypocalciuric) hypercalcemia: evidence for locus heterogeneity. *Am J Hum Genet* 53:193-200
 116. Trump D, Whyte MP, Wooding C, Pang JT, Pearce SHS, Kocher DB, Thakker RV 1995 Linkage studies in a kindred from Oklahoma with familial benign (hypocalciuric) hypercalcemia (FBH) and developmental elevations in serum parathyroid hormone levels, indicate a third locus for FBH. *Hum Genet* 96:183-187
 117. Maggio R, Vogel Z, Wess J 1993 Coexpression studies with mutant muscarinic/adrenergic receptors provide evidence for intermolecular "cross-talk" between G protein linked receptors. *Proc Natl Acad Sci USA* 90:3103-3107
 118. Hirata T, Kakizuka A, Ushikubi F, Fuse I, Okuma M, Narumiya S 1994 Arg 60 to Leu mutation of the human thromboxane A2 receptor in a dominantly inherited bleeding disorder. *J Clin Invest* 94:1662-1667
 119. Chou YH, Pollak MR, Brandi ML, Toss G, Arnqvist H, Atkinson AB, Papapoulos SE, Marx SJ, Brown EM, Seidman JG, Seidman CE 1995 Mutations in the human Ca²⁺-sensing receptor gene that cause familial hypocalciuric hypercalcemia. *Am J Hum Genet* 56:1075-1079
 120. Janicic N, Pausova Z, Cole DEC, Hendy GN 1995 Insertion of an Alu sequence in the Ca²⁺-sensing receptor gene in familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. *Am J Hum Genet* 56:880-886
 121. Heath III H, Odelberg S, Jackson CE, Teh BT, Hayward N, Larsson C, Buist NRM, Krapcho KJ, Hung BC, Capuano IV, Garrett JE, Leppert MF 1996 Clustered inactivating mutations and benign polymorphisms of the calcium receptor gene in familial benign hypocalciuric hypercalcemia suggest receptor functional domains. *J Clin Endocrinol Metab* 81:1312-1317
 122. Aida K, Koishi S, Inoue M, Nakazato M, Tawata M, Onaya T 1995 Familial hypocalciuric hypercalcemia associated with mutation in the human Ca²⁺-sensing receptor gene. *J Clin Endocrinol Metab* 80:2594-2598
 123. Janicic N, Pausova Z, Cole DEC, Hendy GN 1995 De novo expansion of an alu insertion mutation of the Ca²⁺-sensing receptor gene in familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. *J Bone Miner Res* 10[Suppl 1]:S191
 124. Marx SJ, Lasker RD, Brown EM, Fitzpatrick LA, Sweezey NB, Goldbloom RB, Gillis DA, Cole DE 1986 Secretory dysfunction in parathyroid cells from a neonate with severe primary hyperparathyroidism. *J Clin Endocrinol Metab* 62:445-449
 125. Cooper L, Wertheimer J, Levey R, Brown E, LeBoff M, Wilkinson R, Anast CS 1986 Severe primary hyperparathyroidism in a neonate with two hypercalcemic parents: management with parathyroidectomy and heterotopic autotransplantation. *Pediatrics* 78:263-268
 126. Ho C, Conner DA, Pollak MR, Ladd DJ, Kifor O, Warren HB, Brown EM, Seidman JG, Seidman CE 1995 A mouse model of human familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. *Nature Genet* 11:389-394
 127. Garrett JE, Steffey ME, Nemeth EF 1995 The calcium receptor agonist R-568 suppresses PTH mRNA levels in cultured bovine parathyroid cells. *J Bone Miner Res* 10:S387 (Abstract M539)
 128. Jones SM, Frindt G, Windhager EE 1988 Effect of peritubular [Ca] or ionomycin on hydrosmotic response of CCTs to ADH or cAMP. *Am J Physiol* 254:F240-F253
 129. Marx SJ, Attie MF, Stock JL, Spiegel AM, Levine MA 1981 Maximal urine-concentrating ability: familial hypocalciuric hypercalcemia vs. typical primary hyperparathyroidism. *J Clin Endocrinol Metab* 52:736-740
 130. Randall C, Lauchlan SC 1963 Parathyroid hyperplasia in an infant. *Am J Dis Child* 105:364-367
 131. Garcia-Bunuel R, Kutchemeshgi A, Brandes D 1974 Hereditary hyperparathyroidism: the fine structure of the parathyroid gland. *Arch Pathol* 97:399-403
 132. Spiegel AM, Harrison HE, Marx SJ, Brown EM, Aurbach GD 1977 Neonatal primary hyperparathyroidism with autosomal dominant inheritance. *J Pediatr* 90:269-272
 133. Proesmans W, Dhondt F, Logghe N 1977 Congenital hyperparathyroidism. Case report and review of the literature. *Acta Pediatr Belg* 30:45-52
 134. Thompson NW, Carpenter LC, Kessler DL, Nishiyama RH 1978 Hereditary neonatal hyperparathyroidism. *Arch Surg* 113:100-103
 135. Matsuo M, Okita K, Takemine H, Fujita T 1982 Neonatal primary hyperparathyroidism in familial hypocalciuric hypercalcemia. *Am J Dis Child* 136:728-731
 136. Marx SJ, Attie MF, Spiegel AM, Levine MA, Lasker RD, Fox M 1982 An association between neonatal severe primary hyperparathyroidism and familial hypocalciuric hypercalcemia in three kindreds. *N Engl J Med* 306:257-264
 137. Eftekhari F, Yousefzadeh DK 1982 Primary infantile hyperparathyroidism: clinical, laboratory and radiographic features in 21 cases. *Skeletal Radiol* 8:201-208
 138. Gaudelus J, Dandine M, Nathanson M, Perelman R, Hassan M 1983 Rib cage deformity in neonatal hyperparathyroidism [letter]. *Am J Dis Child* 137:408-409
 139. Pomeranz A, Wolach B, Raz A, Ben Ari Y 1992 Neonatal hyperparathyroidism. Conservative treatment with intravenous and oral rehydration solutions. *Child Nephrol Urol* 12:55-58
 140. Corbeel L, Casaer P, Malvaux P, Lormans J, Bourgeois N 1968 Hyperparathyroidie congenitale. *Arch Fr Pediatr* 25:879-891
 141. Mallete LA 1994 The functional and pathologic spectrum of parathyroid abnormalities in hyperparathyroidism. In: Bilezikian JP, Marcus R, Levine MA (eds) *The Parathyroids*, Raven Press, New York, pp 423-455
 142. Lutz P, Kane O, Pfersdorff A, Seiller F, Sauvage P, Levy JM 1986 Neonatal primary hyperparathyroidism: total parathyroidectomy

- with autotransplantation of cryopreserved parathyroid tissue. *Acta Paediatr Scand* 75:179-182
143. **Harris SS, D'Ercole AJ** 1989 Neonatal hyperparathyroidism: the natural course in the absence of surgical intervention. *Pediatrics* 83:53-56
 144. **Fujimoto Y, Hazama H, Oku K** 1990 Severe primary hyperparathyroidism in a neonate having a parent with hypercalcemia: treatment by total parathyroidectomy and simultaneous heterotopic autotransplantation. *Surgery* 108:933-938
 145. **Page LA, Haddow JE** 1987 Self-limited neonatal hyperparathyroidism in familial hypocalciuric hypercalcemia. *J Pediatr* 111:261-264
 146. **Fujita T, Watanabe N, Fukase M, Tsutsumi M, Fukami T, Imai Y, Sakaguchi K, Okada S, Matsuo M, Takemine H** 1983 Familial hypocalciuric hypercalcemia involving four members of a kindred including a girl with severe neonatal primary hyperparathyroidism. *Miner Electrolyte Metab* 9:51-54
 147. **Marx SJ, Fraser D, Rapoport A** 1985 Familial hypocalciuric hypercalcemia. Mild expression of the gene in heterozygotes and severe expression in homozygotes. *Am J Med* 78:15-22
 148. **Pollak M, Chou YH, Marx SJ, Steinmann B, Cole DE, Brandi ML, Papapoulos SE, Menko FH, Hendy GN, Brown EM, Seidman CE, Seidman JG** 1994 Familial hypocalciuric hypercalcemia and neonatal severe hyperparathyroidism. Effects of mutant gene dosage on phenotype. *J Clin Invest* 93:1108-1112
 149. **Estep HL, Mistry Z, Burke PK**, Familial idiopathic hypocalcemia. Program and Abstracts of the 63rd Annual Meeting of The Endocrine Society, Cincinnati, OH, 1981, p 275 (Abstract 750)
 150. **Finegold DN, Armitage MM, Galiani M, Matise TC, Pandian MR, Perry YM, Deka R, Ferrell RE** 1994 Preliminary localization of a gene for autosomal dominant hypoparathyroidism to chromosome 3q13. *Pediatr Res* 36:414-417
 151. **Perry YM, Finegold DN, Armitage MM, Ferrell RE** 1994 A missense mutation in the Ca-sensing receptor causes familial autosomal dominant hypoparathyroidism. *Am J Hum Genet* 55S:A17
 152. **Pearce SHS, Coulthard M, Kendall-Taylor P, Thakker RV** 1995 Autosomal dominant hypocalcaemia associated with a mutation in the calcium-sensing receptor. *J Bone Miner Res* 10:[Suppl 1]:S176 (Abstract 149)
 153. **Davies M, Mughal Z, Selby PL, Tymms DJ, Mawer EB** 1995 Familial benign hypocalcemia. *J Bone Miner Res* 10:[Suppl 1]:S507
 154. **Parma J, Van Sande J, Swillens S, Tonacchera M, Dumont J, Vassart G** 1995 Somatic mutations causing constitutive activity of the thyrotropin receptor are a major cause of hyperfunctioning thyroid adenomas: identification of additional mutations activating both the cyclic adenosine 3',5'-monophosphate and inositol phosphate-Ca²⁺ cascades. *Mol Endocrinol* 9:725-733
 155. **Shenker A, Laue L, Kosugi S, Merendino Jr JJ, Minegishi T, Cutler Jr GB** 1993 A constitutively activating mutation of the luteinizing hormone receptor in familial male precocious puberty. *Nature* 65:652-654
 156. **Thompson DB, Samowitz WS, Odelberg S, Davis RK, Szabo J, Heath III H** 1995 Genetic abnormalities in sporadic parathyroid adenomas: loss of heterozygosity for chromosome 3q markers flanking the calcium receptor locus. *J Clin Endocrinol Metab* 80:3377-3380
 157. **Hosokawa Y, Pollak MR, Brown EM, Arnold A** 1995 Mutational analysis of the extracellular Ca²⁺-sensing receptor gene in human parathyroid tumors. *J Clin Endocrinol Metab* 80:3107-3110
 158. **Kifor O, Moore Jr FD, Wang P, Goldstein M, Vassilev P, Kifor I, Hebert SC, Brown EM** 1996 Reduced immunostaining for the extracellular Ca²⁺-sensing receptor in primary, uremic secondary hyperparathyroidism. *J Clin Endocrinol Metab* 81:1598-1606
 159. **Heath III H, Sanguinetti EL, Oglesby S, Marriott TB** 1995 Inhibition of human parathyroid hormone secretion *in vivo* by NPS R-568, a calcimimetic drug that targets the parathyroid-cell surface calcium receptor. *Bone* 16 [Suppl 1]:85S
 160. **Fox J, Petty BA, Nemeth EF** 1993 A first generation calcimimetic compound (NPS R-568) that acts on parathyroid cell calcium receptor: a novel approach for hyperparathyroidism. *J Bone Miner Res* 8:[Suppl]:S181 (Abstract)