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# THE EFFECT OF PHYSICAL ACTIVITY ON THE BRAIN DERIVED NEUROTROPHIC FACTOR: FROM ANIMAL TO HUMAN STUDIES

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It is well documented that physical activity can induce a number of various stimuli which are able to enhance the strength and endurance performance of muscles. Moreover, regular physical activity can preserve or delay the appearance of several metabolic disorders in the human body. Physical exercise is also known to enhance the mood and cognitive functions of active people, although the physiological backgrounds of these effects remain unclear. In recent years, since the pioneering study in the past showed that physical activity increases the expression of the brain derived neurothophic factor (BDNF) in the rat brain, a number of studies were undertaken in order to establish the link between that neurothrophin and post-exercise enhancement of mood and cognitive functions in humans. It was recently demonstrated that physical exercise can increase plasma and/or serum BDNF concentration in humans. It was also reported that physical exercise or electrical stimulation can increase the BDNF expression in the skeletal muscles. In the present review, we report the current state of research concerning the effect of a single bout of exercise and training on the BDNF expression in the brain, in both the working muscles as well as on its concentrations in the blood. We have concluded that there may be potential benefits of the exercise-induced enhancement of the BDNF expression and release in the brain as well as in the peripheral tissues, resulting in the improvement of the functioning of the body, although this effect, especially in humans, requires more research.

Key words: brain derived neurothophic factor, cognitive function, exercise, learning, mood, training

#### INTRODUCTION

It is well known that physical activity provides a number of various stimuli which are able to enhance both the metabolic and functional status of the human body. Physical training within a relatively short period of time (weeks or months) is able to increase the expression of a number of genes involved in any enhancement of a physical capacity (1). Perhaps the most spectacular discovery concerning the adaptation of the body to physical training was presented by John Holloszy, who has shown that regular physical activity/training can induce mitochondria biogenesis in skeletal muscles, leading to an improvement of physical performance (2-4). An increase in muscle mitochondria density enhances the metabolic stability of the muscle during exercise and increases muscle performance (5-7).

For a long period of time physical training was almost exclusively associated with athletes and their preparation for top sports events. However, during the last few decades the amount of studies concerning the effect of physical activity/training and its effects on the health status of healthy yet untrained people as well as of patients has substantially increased (8). They were mainly focused on the effect of training on the adaptation of the cardiovascular, hormonal and muscle systems. The new vision of

the benefits of regular physical activity has been presented in a series of experiments showing the anti-inflammatory action of physical exercise (for review see (9, 10)). It was demonstrated that the moderate intensity of physical exercise can be an important factor in the prevention as well as in the healing of several metabolic disturbances of the human body (1, 8, 11).

Physical exercise is also known to enhance the mood and cognitive functions in humans (12-15), although the physiological backgrounds of those effects remain unclear. In recent years there has been a growing interest in research concerning the effect of physical exercise and training on the functioning of the brain, with a special focus on its effect on the brain derived neurotrophic factor (BDNF). Several of these studies showed that exercise induced an up-regulation of the BDNF in the hippocampus and might therefore play an important role in the enhancement of cognitive functions in humans (for review see. (16)).

The BDNF, its isoforms and their receptors

BDNF is most abundant in the growth factor family (17-20). This protein, to begin with, was purified from the brain of a pig (21). The BDNF is widely expressed in the rodent brain, and is especially abundant in the hippocampus, cerebral cortex,

cerebellum, striatum and the amygdala (22, 23). A recent study by Zhang *et al.* (24), showed an intense expression of the BDNF in various parts of an adult monkey brain including: cerebral cortex (layers III and IV), hipocampus (granular cell layer), midbrain (substantia nigra), pons (abducent and facial nucleus), medulla oblongata (hypoglossal nucleus, cuneate and gracile nucleus) and thalamus and hypothalamus nuclei (arcuate) as well as a moderate and mild expression in several other regions of the brain (for an overview, see *Table 1* in (24)). BDNF expression was also reported in various parts of the human brain, including the hippocampus, claustrum, amygdala, bed nucleus of the stria terminalis, septum and the nucleus of the solitary tract (25).

It has been demonstrated that the BDNF plays a critical role in the activity-dependent processes, including synapse development and plasticity (26-28). It was reported that BDNF, by acting via the protein tyrosine kinase receptor (TrkB) (20, 29) regulates a number of processes including neuronal development and its functions (19, 30-32). BDNF is involved in memory formation, including learning and behavior, synaptic plasticity, synaptic efficacy and neuronal connectivity, plus it promotes the development of immature neurons and enhances the survival of adult neurons (19, 33, 34). According to Monteggia et al. (31) the role of BDNF in the adult brain may be different from that in the developing brain. It was demonstrated that the loss of BDNF selectively in the brain of adult mice resulted in impaired hippocampal function, whereas the loss of BDNF during the early stages of development contributed to hyperactivity as well as to the more severe impairments in hippocampal-dependent learning (31).

BDNF, similar to other neurothropins, is initially synthesized as a precursor (pro-BDNF with MW of 32 kDa), which is subsequently cleaved to generate the mature BDNF (mBDNF with MW of 14 kDa) (see, (35, 36). Additionally, a third BDNF isoform with MW of 28 kDa was recognized as well (see *Fig. 1* in (35)). This BDNF isoform, known as truncated BDNF, is not further cleaved (37). It is well documented that the mBDNF and the pro-BDNF are biologically active, whereas the function of the truncated BDNF is still unknown.

BDNF exerts its biological effects in the neural system via two types of receptor: the tyrosine kinase receptor (Trk) B receptor and the pan-neurotropin receptor p75 (p75  $^{\mbox{\scriptsize NTR}})$  (for an overview see (38)). It was demonstrated that pro-BDNF preferentially interacts with the p75NTR, whereas mBDNF selectively binds and activates the TrkB (39, 40). In this way various BDNF isoforms generate sometimes opposite effects. For example, it has been reported that the activation of p75<sup>NTR</sup> by an endogenous pro-BDNF results in long term depression (LTD) in the hippocampus (41, 42) and induces apoptosis in peripheral neurons (43), whereas activation of the TrkB receptors by mBDNF is essential for long term potentiation (LTP) (44, 45) and regulates the neuronal development and its functions (31). Moreover, it has been recently demonstrated that the exogenous pro-BDNF suppresses synaptic transmission and structurally causes axonal retraction by activation presynaptic p75NTR (46). These authors also showed that muscle stimulation induces a secretion of pro-BDNF, which elicits either synaptic potentiation or depression, depending on whether it is proteolitically cleaved (46). For an overview of the major intracellular signaling pathway activated throughout TrkB and p75NTR receptors see Fig. 2 in (38). It is of interest how the two receptors communicates with each other in order to provide optimal cell functioning. This issue has been recently reviewed by Reichardt (38) showing that the signaling pathways initiated thought Trk receptors act at several steps to suppress the major pro-apoptotic-signaling pathway stimulated by p75NTR to maintain a proper cell functioning (see also (47)). For this reason, in order to evaluate

the effect of the BDNF release on physiological functions, it is important to determine both the amount of proBDNF and the mature BDNF release from the neurons. However, relatively little data on the measurements of all the above mentioned BDNF isoforms has been reported so far.

The effect of exercise on the BDNF expression in a rodent's brain

It was demonstrated that physical activity/training can increase BDNF gene expression in the brain (48-53). The interest in this area of research was initiated by Nepher et al. (48), who originally reported a significant positive correlation between the mean distance run on a running wheel and the mRNA for BDNF in the hippocampus and caudal neocortex of the studied rats. The authors concluded that physical activity could increase the availability of BDNF to these cells by upregulating its expression in the hippocampus, and as a result it was proposed that exercise induced the up-regulation of BDNF and could help to increase the brain's resistance to damage and degeneration through BDNF's support of neuronal growth, function and survival (48). Subsequently, Nepper (49) have reported that the 2-7 nights of running resulted in a significant increase of mRNA for BDNF in the rat hippocampus. These original findings by Neppher et al. (48, 49) were confirmed by others, who showed that, indeed, physical activity/training is able to up-regulate the BDNF expression in animal brains. For example, Oliff et al. (54) have found that the brain mRNA expression in rats correlates with the distance run during voluntary activity. Furthermore, the authors have reported that as little as 6 hours of voluntary wheel running resulted in a significant up-regulation of the hippocampal BDNF mRNA expression in rats, which remained elevated after 12 hours of voluntary running Oliff et al. (54). Recently, Rassmussen et al. (55) reported that a single bout of exercise resulted in the significant up-regulation of BDNF mRNA in the hippocampus and cortex of a mouse, with a peak occurring at about 2 hours after finishing the treadmill exercise bout.

An interesting observation concerning the dynamics of the changes in the BDNF protein levels in rats during the time course of training and detraining was published by Berchtold et al. (52). The authors reported that a daily exercise training (voluntary running exercise performed on a running wheel) resulted in a significant increase in the BDNF protein level after 14 days of training, which continued to rise until the end of the training period (i.e. up to 90 days). Moreover, it remained significantly elevated (above the level found in the sedentary rats) up to the 7th day after finishing the daily training (see Fig. 2A and 3A in (52)). The authors have also studied the effect of the same kind of training performed on alternating days. It was found that this training can also significantly increase the BDNF protein level in the hippocampus of the rats, although this increase is slower and it decays much faster when compared to the daily exercise training (see, Fig. 2B and 3B in Berchtold et al. (52)). This data suggests that physical training can upregulate the BDNF protein level in the brain, but the most beneficial seems to be when the daily training is performed for at least several months.

An interesting observation concerning the various types of exercise: treadmill running *vs.* voluntary wheel running on cognitive functions was recently reported by Liu *et al.* (53). These authors have demonstrated that although both moderate treadmill running and wheel running up-regulated the BDNF-TrkB pathway in the hippocampus, both forms of the training protocols exerted varied effects in the different regions of the brain and on its functions. The authors concluded that different forms of exercise induced changes of the neuroplasticity in different brain regions and also exerted diverse effects on

various forms of learning and memory. This effect should be taken into consideration when interpreting the effect of exercise on brain plasticity and its functioning.

#### Exercise-induced BDNF and learning benefits

It was postulated by Figurov et al. (45) that BDNF may regulate LTP in the developing hippocampus as well as the adult hippocampus by enhancing synaptic responses to tetanic stimulation. The authors demonstrated that BDNF promoted the induction of LTP by tetanic stimulation in young hippocampal slices, which with their absence of BDNF showed only shortterm potentiation (STP) (45). The LTP is considered to be a form of synaptic plasticity involved in long-term memory formation (56, 57). The importance of BDNF in LTP was also reported by others (see e.g. (32, 58-61)). Additional evidence for the role of BDNF in the cognitive function of the brain comes from the study by Alonso et al. (62), which shows that an intrahippocampal administration of recombinant human BDNF facilitated longterm memory (LTM) formation in the rat brain (62), whereas bilateral infusion of the function-blocking anti-BDNF antibody in to the CA1 region of the dorsal hippocampus impaired LTM retention scores in rats. Lee et al. (63) have shown that the process of memory consolidation in rats is strictly dependent upon the presence of BDNF in the hypothalamus. It was also recently postulated that plasma BDNF concentration can be considered as a biomarker of memory and general cognitive function in women (64).

Since physical activity is able to up-regulate the BDNF expression in the rat hippocampus (48-50), it was postulated that this increase may play an important role in the cognitive functions, including learning and memory (65). Indeed, the exercise-induced up-regulation of BDNF in the hypothalamus was related to an improvement of cognitive function, including memory, in rodents (51, 66, 67), whereas the inhibition or downregulation of BDNF or TrkB impaired memory formation (67-70). This data suggests that, indeed, regular physical activity might potentiate the cognitive functions via the exercise-induced up-regulation of the hippocampus BDNF (32, 53, 71). It was also recently reported that the training-induced BDNF expression in the peripheral cortex of exercising rats was strongly correlated with object recognition memory (72). This is the first evidence whereby the changes in the BDNF level are associated with the voluntary exercise-induced improvement in non-spatial memory, which is itself mediated by structures outside the hippocampus (72).

#### BDNF polimorphism

Additional evidence for the importance of BDNF in learning and cognitive functions in humans is provided by recent studies (73), initiated by Egan et al. (74), who have found that the presence of the Val<sup>66</sup>Met BDNF polymorphism in humans (a methionine (Met) substitution for valine (Val) at codon 66), found in one or in both alleles in approximately 30% of people (73), was associated with poorer episodic memory and abnormal hippocampal activation. Further studies by this group (75) provided additional evidence for the importance of the BDNF Val<sup>66</sup>Met polymorphism in memory performance. Subsequently, Kleim et al. (76) reported that the subjects lacking the BDNF Val<sup>66</sup>Met polymorphism showed an expansion of the motor map with training, whereas subjects with the BDNF Val<sup>66</sup>Met polymorphism in one or both alleles showed only a little of such plasticity. Cheeran et al. (77) recently found that the response of healthy subjects to three different plasticity-inducing protocols in the motor cortex is associated with the polymorphism of the BDNF gene that they carry. According to Cheeran et al. (77), the

polymorphism in the BDNF gene may be one factor that influences the natural response of the brain to injury and disease. Moreover, it was reported that the Val<sup>66</sup>Met polymorphism may play a key role in the genetic predisposition to anxiety and depressive disorders (Chen *et al.* (78)). These observations were further developed by Soliman *et al.* (79), who demonstrated that variant BDNF alleles may play a role in anxiety disorders, showing an impaired learning of the cues that signal safety versus threat and in the efficacy of treatments that rely on the extinction mechanism, such as exposure therapy. According to Ninan *et al.* (80) the BDNF Val<sup>66</sup>Met polymorphism has a direct effect on NMDA receptor transmission, which may account for the changes in the synaptic plasticity in the hippocampus.

These studies clearly demonstrate that the disturbances in the BDNF structure, such as the Val<sup>66</sup>Met polymorphism, affects memory and learning and that it might also be involved in the origin of several neurological and psychiatric conditions (77).

#### Plasma and/or serum BDNF level in healthy humans

It was originally demonstrated by Rosenfeld et al. (81), that the BDNF can be detected in both the human serum and in the plasma, besides its concentration in the serum is more than 200fold higher than in the plasma. Currently, based on a larger sample of subjects, it is well established that the serum BDNF concentration [BDNF]<sub>s</sub> in healthy humans is higher by about 100-fold than in plasma [BDNF]<sub>p</sub> (82, 83). Most of the BDNF circulating with the blood is stored in the platelets (84, 85). Therefore, a close correlation exists between the count of the platelets and the serum BDNF concentration in humans (82). The circulating BDNF is produced by a number of peripheral non-neuronal tissues, including vascular human endothelial cells (86-89), T cells, B cells and monocytes (90). Additionally, it was shown that BDNF mRNA in the skeletal muscles of rodents increases in response to contraction (65, 91). The recent study by Matthews et al. (92), confirmed the previous observations showing that skeletal muscles can indeed produce BDNF, although in the light of their study the muscle-produced BDNF is not released into the circulation and cannot therefore account for its changes in serum or plasma.

According to Fujimura et al. (85), the platelets can bind, store and release BDNF upon activation at the site of traumatic injury in order to facilitate the repair of peripheral nerves or other tissues that contain TrkB. Moreover, these authors (85) question the role of platelets produced by the circulating BDNF. Rojas Vega et al. (93), and have recently postulated that the elevated serum BDNF concentrations might be beneficial for improving recovery after spinal cord injury. However, the contribution of varied sites where BDNF is periphally produced is not established yet. According to Nakahashi et al. (87) the endothelial cells may significantly contribiute to circulating BDNF. It was demonstrated that BDNF can cross the bloodbrain barrier (94, 95) in both directions, i.e. from the brain to the periphery and from the periphery to the brain (95), via the high capacity saturable transporter system (95). Furthermore, it was reported that the BDNF level in the brain correlates with the serum BDNF concentration (96), therefore it has been suggested that the blood level of the BDNF may reflect the brain level and vice-versa. According to Lommatzch et al. (82), the changes in [BDNF]<sub>p</sub> are reflecting its changes in the brain. It should be mentioned, however, that some authors (42, 97) challenged the finding of Poduslao and Curran (94) and of Pan et al. (95) where the exchange of BDNF between the brain tissue and blood is concerned. It was recently reported that during physical exercise the increase in the [BDNF]<sub>p</sub> concentration in humans was due to an enhanced release of BDNF from the brain (55). These authors have shown that in humans, both at rest and during exercise, the

brain contributed to 70-80% of the circulating BDNF, while the other contribution decreased following 1 hour of recovery. According to Rasmussen *et al.* (55), the brain is a major but not sole contributor to the circulation of BDNF both at rest and during exercise. This is an important new observation, although the magnitude of the contribution of the brain to peripheral BDNF concentration, as reported by Rasmussen *et al.* (55), should be considered with caution, since the contribution from other peripheral sources of BDNF release during exercise, such as the platelets, vascular endothelial cells, smooth muscle cells and other cells, requires more research.

#### Plasma and/or serum BDNF level in mental disorders

Since BDNF plays a critical role in the functioning of the brain (19, 28, 31), some studies have attempted to relate the BDNF with major mental disorders: depression (98, 99) and schizophrenia (100). A number of studies were undertaken to assess the levels of BDNF in plasma [BDNF]<sub>p</sub> and/or in serum [BDNF]<sub>s</sub> in patients with depression or schizophrenia. Moreover, Komulainen *et al.* (64) have recently reported that plasma BDNF is a biomarker of impaired memory and general cognitive function in ageing women.

#### Depression

It was reported that the basal [BDNF]<sub>s</sub> in untreated patients suffering from major depressive disorders is significantly lower than in control subjects (101-105). Additionally, low plasma BDNF was reported to be associated with suicidal behavior in depressed patients (Kim *et al.* (104)). Interestingly, it was reported that 8-12 weeks of treatment with antidepressant drugs resulted in a significant increase in serum BDNF concentration in the studied patients (102, 103, 105). It was also recently shown that the basal [BDNF]<sub>p</sub> in patients suffering from post-traumatic stress disorder was significantly lower than in the healthy subjects (106).

## Schizoprenia

A large body of data concerning the serum [BDNF]<sub>s</sub> was collected from schizophrenic patients, although the emerging picture is not clear. While most of the studies showed a lower [BDNF]<sub>p</sub> or [BDNF]<sub>s</sub> concentrations in patients with schizophrenia, when compared to healthy controls (107-110), but some research (111, 112) reported similar [BDNF]<sub>p</sub> or [BDNF]<sub>s</sub> concentrations in both groups. Occasionally, even higher than normal levels of [BDNF]<sub>s</sub> where observed in schizophrenic patients (113, 114). Some of the differences in the [BDNF]<sub>s</sub> levels in schizophrenic patients found in different studies might be due to the various stages and the level of severity of the illness besides the treatment methodology.

Some interesting observations concerning the [BDNF]<sub>s</sub> levels in schizophrenic patients were recently reported by Carlino *et al.* (115). These authors measured both the total BDNF levels (as in most of the above mentioned studies) as well as three BDNF isoforms (pro-BDNF, truncated BDNF and mat-BDNF) levels in serum. The total BDNF concentration in the studied schizophrenic patients was slightly lower than in the control group, although the most interesting findings of this study were the clear differences in the pro-BDNF, truncated BDNF and the mat-BDNF isoforms in patients *vs.* the control group. Namely, the serum pro-BDNF and mat-BDNF concentrations were significantly higher in schizophrenic patients, whereas the level of the truncated BDNF isoform was significantly lower in the schizophrenic patients. Moreover, the reduced levels of the serum truncated BDNF total BDNF ratio

correlated with the worst PANSS negative and positive symptoms and with the poorer neurocognitive performance (see Carlino *et al.* (115)).

## BDNF and type 2 diabetes mellitus

It was shown that serum BDNF concentrations in type 2 diabetes mellitus (T2DM) patients are significantly lower than in non-diabetic controls (116, 117). Opposite results were reported by Suwa et al. (118), showing that serum BDNF levels in newly diagnosed female patients with T2DM were significantly higher than those in the control subjects. According to Hristowa et al. (119), elevated plasma and/or serum BDNF concentration may be an early marker of pathological metabolic changes in the body. On the other hand, it was recently reported that 6 weeks of endurance training has induced an improvement in the physical capacity of humans, involving an increase in maximal oxygen uptake (VO<sub>2max</sub>), an increase in the power generating capability at the VO<sub>2max</sub>, a decrease in lipid peroxidation, and that a slight decrease in insulin resistance was accompanied by a significant increase in basal as well as the exercise induced plasma BDNF concentrations in young healthy men (120). Furthermore, it was reported that the administration of BDNF to diabetic mice improved glucose (121-123) and lipid metabolism (124). These recent findings suggest that an elevated serum and/or plasma BDNF concentration in healthy individuals may have opposite meaning than in patients with metabolic disorders (see (118, 120, 125, 126). Additionally, the systematic study of the changes in serum BDNF in patients at various stages of the T2DM are needed in order to establish its role in the origin of this metabolic disorder.

The effect of a single bout of physical exercise and training on the plasma or serum BDNF concentrations in humans

It was reported that a single bout of physical exercise can increase the plasma or serum BDNF concentrations in healthy humans. It was originally demonstrated by Gold et al. (127) that a single session of prolonged exercise (30 minutes cycling at 60% VO<sub>2</sub>max), resulted in a significant increase in the serum BDNF concentration both in healthy individuals as well as in multiple sclerosis patients. Subsequently, in the study by Rojas Vega et al. (128), it was shown that a single bout of maximal incremental exercise resulted in a significant increase in the serum BDNF in recreational athletes, whereas 10 minutes of moderate aerobic cycling was not sufficient to increase the serum BDNF concentration above the pre-exercise level. This finding is in accordance with the study by Ferris et al. (129), which shows that the magnitude of the increase in the serum BDNF concentration during exercise is dependent on the intensity of the exercise. A significant increase in the serum BDNF concentration in young healthy men has also been reported by Winter et al. (130), after very high intensity running for a small duration (2 runs until exhaustion, lasting 3 minutes each, with 2 minutes break). Similarly, Tang et al. (131) reported a significant increase in the serum BDNF concentration in young and healthy men after short term high intensity of 15 minutes step-exercise. No effect of a single bout of maximal incremental cycling exercise - (up to  $VO_{2\text{max}}$ , lasting about 30 minutes), on the plasma BDNF concentration was found in young healthy men (120). Recently it was reported that a prolonged - 4 hours rowing exercise resulted in a significant increase in the plasma BDNF concentration in humans (55). Additionally, Yarrow et al. (132) have reported that a single session of resistance exercise resulted in a significant increase in the serum BDNF concentration in young and healthy men.

Regarding the effect of physical training on the basal and exercise-induced changes of plasma or serum BDNF

concentrations, the picture is more complex. Both the effects of the endurance as well as the strength training program in a varied groups of subjects on the basal and exercise induced plasma or serum BDNF concentrations were evaluated. Firstly considering the effects of endurance training on the basal and the exerciseinduced plasma or serum BDNF concentrations, Schulz et al. (133), who studied the effect of 8-week aerobic bicycle training on the serum BDNF concentration in patients with multiple sclerosis, found no effect of the training on the basal as well as the exercise-induced serum BDNF concentration. Similarly, Castellano and White (134), who studied the effect of endurance cycling training on the serum BDNF concentration in multiple sclerosis patients, found a temporary (after 4 weeks of training) increase in the serum BDNF concentration, which returned to the pre-training level after 8 weeks of training. Zoladz et al. (120), who studied the effect of 5 weeks of moderate intensity training, have found a significant increase in the basal plasma BDNF concentration as well as a significant exercise-induced increase in the plasma BDNF concentrations in young and healthy men. Recently, Seifert et al. (135) reported that 3 months of endurance training in young and healthy men enhanced the resting release of BDNF from the brain but had no effect on the magnitude of the exercise-induced increase in the plasma BDNF concentration.

Where the effect of the strength training on the on basal and the exercise-induced plasma or serum BDNF concentrations is concerned, the reported results are not consistent. Levinger *et al.* (126) have shown that 10-week resistance training did not affect the plasma baseline BDNF concentration in middle aged subjects. Likewise, Goekint *et al.* (136) have found no effect from 10-week strength training on the basal and post exercise serum BDNF concentration in young and healthy men. On the other hand, Yarrow *et al.* (132) have recently reported that a 5-week resistance training augments the exercise-induced transient increase in serum BDNF concentration in young and healthy yet previously untrained males.

The training-induced an increase in the [BDNF]<sub>p</sub> and [BDNF]<sub>s</sub> might play a role in the exercise-induced improvement of mood (137), as well as in the protection and regeneration of various tissues (93, 138). The training-induced enhancement of the circulating BDNF might be involved in the neoangiogenesis of the cardiac and skeletal muscles after training (86, 88). Furthermore, the training-induced elevation of BDNF may be beneficial to the efficacy of pharmacological antidepressant treatment (139, 140). It was also postulated (120) that the training-induced increase in [BDNF]<sub>p</sub> *via* its action in the central nervous system may also enhance motor learning ability in athletes. In the light of these recent studies (141), the training-induced up-regulation of the skeletal muscle BDNF, as shown by Matthews *et al.* (92) might increase fat oxidation in skeletal muscles in an AMPK-dependent fashion.

Further studies in humans are needed to establish first of all the effect of the single bout of exercise and training on the changes in the serum levels of the pro-BDNF, truncated BDNF and the mat-BDNF isoforms in humans and to relate their levels to the mood state and the cognitive performance. More data is needed to establish the role of BDNF in the enhancement of the metabolic status of skeletal muscles.

### CONCLUSIONS

We have concluded that in the light of the available data collected mainly from rodents, physical training is able to upregulate the BDNF expression in some regions of the brain. The existing evidence indicates that the training-induced upregulation of BDNF expression in the brain may play a role in the improvement of mood as well as in an enhancement of cognitive functions. Moreover, physical exercise, similar to

pharmacological treatment with antidepressant drugs, is able to enhance the level of plasma and/or serum BDNF concentrations in humans. However, the physiological importance of the training-induced changes in plasma and/or serum BDNF concentrations in humans remains to be established. Further studies are required to determine the effect of a single bout of exercise and training on the three BDNF isoforms (pro-BDNF, truncated BDNF and mat-BDNF) levels in serum, especially in relation to the mental state and cognitive performances in humans.

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#### REFERENCES

- Booth FW, Laye MJ. The future: genes, physical activity and health. Acta Physiol (Oxf) 2010; 199: 549-556.
- Holloszy JO, Booth FW. Biochemical adaptations to endurance exercise in muscle. *Annu Rev Physiol* 1976; 38: 273-291.
- Holloszy JO. Biochemical adaptations in muscle. Effects of exercise on mitochondrial oxygen uptake and respiratory enzyme activity in skeletal muscle. *J Biol Chem* 1967; 242: 2278-2282.
- Holloszy JO. Regulation by exercise of skeletal muscle content of mitochondria and GLUT4. *J Physiol Pharmacol* 2008; 59(Suppl 7): 5-18.
- Dudley GA, Tullson PC, Terjung RL. Influence of mitochondrial content on the sensitivity of respiratory control. *J Biol Chem* 1987; 262: 9109-9114.
- Constable SH, Favier RJ, McLane JA, Fell RD, Chen M, Holloszy JO. Energy metabolism in contracting rat skeletal muscle: adaptation to exercise training. *Am J Physiol* 1987; 253: C316-C322.
- Zoladz JA, Korzeniewski B, Grassi B. Training-induced acceleration of oxygen uptake kinetics in skeletal muscle: the underlying mechanisms. *J Physiol Pharmacol* 2006; 57(Suppl 10): 67-84.
- Pedersen BK, Saltin B. Evidence for prescribing exercise as therapy in chronic disease. Scand J Med Sci Sports 2006; 16(Suppl 1): 3-63.
- Pedersen BK. The diseasome of physical inactivity-and the role of myokines in muscle-fat cross talk. *J Physiol* 2009; 587: 5559-5568.
- Pedersen BK. Edward F. Adolph distinguished lecture: muscle as an endocrine organ: IL-6 and other myokines. J Appl Physiol 2009; 107: 1006-1014.
- Laughlin MH, Roseguini B. Mechanisms for exercise training-induced increases in skeletal muscle blood flow capacity: differences with interval sprint training versus aerobic endurance training. *J Physiol Pharmacol* 2008; 59(Suppl 7): 71-88.
- 12. Kramer AF, Hahn S, Cohen NJ, et al. Ageing, fitness and neurocognitive function. *Nature* 1999; 400: 418-419.
- Colcombe S, Kramer AF. Fitness effects on the cognitive function of older adults: a meta-analytic study. *Psychol Sci* 2003; 14: 125-130.
- 14. Ruscheweyh R, Willemer C, Kruger K, et al. Physical activity and memory functions: an interventional study.

- Neurobiol Aging 2009; doi:10.1016/j.neurobiolaging. 2009 08 001
- Berchtold NC, Castello N, Cotman CW. Exercise and timedependent benefits tolearning and memory. *Neuroscience* 2010; 167: 588-597.
- Cotman CW, Berchtold NC, Adlard PA, Perreau VM. Exercise and the brain. In: Molecural and Cellular Exercise Physiology. FC Mooren, K Völker (eds). Campaign, USA, Human Kinetics, 2005, pp. 331-341.
- Lindsay RM, Wiegand SJ, Altar CA, DiStefano PS. Neurotrophic factors: from molecule to man. *Trends Neurosci* 1994; 17: 182-190.
- 18. Huang EJ, Reichardt LF. Trk receptors: roles in neuronal signal transduction. *Annu Rev Biochem* 2003; 72: 609-642.
- 19. Binder DK, Scharfman HE. Brain-derived neurotrophic factor. *Growth Factors* 2004; 22: 123-131.
- 20. Kozisek ME, Middlemas D, Bylund DB. Brain-derived neurotrophic factor and its receptor tropomyosin-related kinase B in the mechanism of action of antidepressant therapies. *Pharmacol Ther* 2008; 117: 30-51.
- Barde YA, Edgar D, Thoenen H. Purification of a new neurotrophic factor from mammalian brain. *EMBO J* 1982; 1: 549-553.
- 22. Dugich-Djordjevic MM, Peterson C, Isono F, *et al.* Immunohistochemical visualization of brain-derived neurotrophic factor in the rat brain. *Eur J Neurosci* 1995; 7: 1831-1839.
- Kawamoto Y, Nakamura S, Nakano S, Oka N, Akiguchi I, Kimura J. Immunohistochemical localization of brainderived neurotrophic factor in adult rat brain. *Neuroscience* 1996; 74: 1209-1226.
- 24. Zhang HT, Li LY, Zou XL, et al. Immunohistochemical distribution of NGF, BDNF, NT-3, and NT-4 in adult rhesus monkey brains. J Histochem Cytochem 2007; 55: 1-19.
- 25. Murer MG, Boissiere F, Yan Q, et al. An immunohistochemical study of the distribution of brain-derived neurotrophic factor in the adult human brain, with particular reference to Alzheimer's disease Neuroscience 1999; 88: 1015-1032.
- 26. Lu B, Figurov A. Role of neurotrophins in synapse development and plasticity. *Rev Neurosci* 1997; 8: 1-12.
- Hofer MM, Barde YA. Brain-derived neurotrophic factor prevents neuronal death in vivo. *Nature* 1988; 331: 261-262.
- McAllister AK, Katz LC, Lo DC. Neurotrophins and synaptic plasticity. Annu Rev Neurosci 1999; 22: 295-318.
- 29. Soppet D, Escandon E, Maragos J, et al. The neurotrophic factors brain-derived neurotrophic factor and neurotrophin-3 are ligands for the trkB tyrosine kinase receptor. Cell 1991; 65: 895-903.
- 30. Blum R, Kafitz KW, Konnerth A. Neurotrophin-evoked depolarization requires the sodium channel Na(V)1.9. *Nature* 2002; 419: 687-693.
- Monteggia LM, Barrot M, Powell CM, et al. Essential role of brain-derived neurotrophic factor in adult hippocampal function. Proc Natl Acad Sci USA 2004; 101: 10827-10832.
- 32. Cunha C, Brambilla R, Thomas KL. A simple role for BDNF in learning and memory? Front Mol Neurosci 2010; 3: 1-14.
- 33. Ebadi M, Bashir RM, Heidrick ML, *et al.* Neurotrophins and their receptors in nerve injury and repair. *Neurochem Int* 1997; 30: 347-374.
- 34. Tyler WJ, Alonso M, Bramham CR, Pozzo-Miller LD. From acquisition to consolidation: on the role of brain-derived neurotrophic factor signaling in hippocampal-dependent learning. *Learn Mem* 2002; 9: 224-237.
- Mowla SJ, Farhadi HF, Pareek S, et al. Biosynthesis and posttranslational processing of the precursor to brain-derived neurotrophic factor. J Biol Chem 2001; 276: 12660-12666.

- Yang J, Siao CJ, Nagappan G, et al. Neuronal release of proBDNF. Nat Neurosci 2009; 12: 113-115
- 37. Seidah NG, Mowla SJ, Hamelin J, *et al.* Mammalian subtilisin/kexin isozyme SKI-1: a widely expressed proprotein convertase with a unique cleavage specificity and cellular localization. *Proc Natl Acad Sci USA* 1999; 96: 1321-1326.
- 38. Reichardt LF. Neurotrophin-regulated signalling pathways. *Philos Trans R Soc Lond B Biol Sci* 2006; 361: 1545-1564.
- 39. Chao MV, Bothwell M. Neurotrophins: to cleave or not to cleave. *Neuron* 2002; 33: 9-12.
- Ibanez CF. Jekyll-Hyde neurotrophins: the story of proNGF. *Trends Neurosci* 2002; 25: 284-286. Erratum: *Trends Neurosci* 2002; 25: 378.
- 41. Rosch H, Schweigreiter R, Bonhoeffer T, Barde YA, Korte M. The neurotrophin receptor p75<sup>NTR</sup> modulates long-term depression and regulates the expression of AMPA receptor subunits in the hippocampus. *Proc Natl Acad Sci USA* 2005; 102: 7362-7367.
- 42. Wu D. Neuroprotection in experimental stroke with targeted neurotrophins. *NeuroRx* 2005; 2: 120-128.
- 43. Teng HK, Teng KK, Lee R, et al. ProBDNF induces neuronal apoptosis via activation of a receptor complex of p75<sup>NTR</sup> and sortilin. J Neurosci 2005; 25: 5455-5463.
- 44. Korte M, Carroll P, Wolf E, Brem G, Thoenen H, Bonhoeffer T. Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc Natl Acad Sci USA* 1995; 92: 8856-8860.
- 45. Figurov A, Pozzo-Miller LD, Olafsson P, Wang T, Lu B. Regulation of synaptic responses to high-frequency stimulation and LTP by neurotrophins in the hippocampus. *Nature* 1996; 381: 706-709.
- Yang F, Je HS, Ji Y, Nagappan G, Hempstead B, Lu B. Pro-BDNF-induced synaptic depression and retraction at developing neuromuscular synapses. *J Cell Biol* 2009; 185: 727-741.
- Yoon SO, Casaccia-Bonnefil P, Carter B, Chao MV. Competitive signaling between TrkA and p75 nerve growth factor receptors determines cell survival. *J Neurosci* 1998; 18: 3273-3281.
- Neeper SA, Gomez-Pinilla F, Choi J, Cotman CW. Exercise and brain neurotrophins. *Nature* 1995; 373: 109.
- Neeper SA, Gomez-Pinilla F, Choi J, Cotman CW. Physical activity increases mRNA for brain-derived neurotrophic factor and nerve growth factor in rat brain. *Brain Res* 1996; 726: 49-56.
- van Praag H, Christie BR, Sejnowski TJ, Gage FH. Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc Natl Acad Sci USA* 1999; 96: 13427-13431.
- 51. Adlard PA, Perreau VM, Engesser-Cesar C, Cotman CW. The timecourse of induction of brain-derived neurotrophic factor mRNA and protein in the rat hippocampus following voluntary exercise. *Neurosci Lett* 2004; 363: 43-48.
- 52. Berchtold NC, Chinn G, Chou M, Kesslak JP, Cotman CW. Exercise primes a molecular memory for brain-derived neurotrophic factor protein induction in the rat hippocampus. *Neuroscience* 2005; 133: 853-861.
- Liu YF, Chen HI, Wu CL, et al. Differential effects of treadmill running and wheel running on spatial or aversive learning and memory: roles of amygdalar brain-derived neurotrophic factor and synaptotagmin I. J Physiol 2009; 587: 3221-3231.
- 54. Oliff HS, Berchtold NC, Isackson P, Cotman CW. Exercise-induced regulation of brain-derived neurotrophic factor (BDNF) transcripts in the rat hippocampus. *Brain Res Mol Brain Res* 1998; 61: 147-153.
- 55. Rasmussen P, Brassard P, Adser H, *et al.* Evidence for a release of brain-derived neurotrophic factor from the brain during exercise. *Exp Physiol* 2009; 94: 1062-1069.

- 56. Bliss TV, Collingridge GL. A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 1993; 361: 31-39.
- Martin SJ, Grimwood PD, Morris RG. Synaptic plasticity and memory: an evaluation of the hypothesis. *Annu Rev Neurosci* 2000; 23: 649-711.
- Akaneya Y, Tsumoto T, Kinoshita S, Hatanaka H. Brainderived neurotrophic factor enhances long-term potentiation in rat visual cortex. *J Neurosci* 1997; 17: 6707-6716.
- Kafitz KW, Rose CR, Thoenen H, Konnerth A. Neurotrophin-evoked rapid excitation through TrkB receptors. *Nature* 1999; 401: 918-921.
- 60. Kossel AH, Cambridge SB, Wagner U, Bonhoeffer T. A caged Ab reveals an immediate/instructive effect of BDNF during hippocampal synaptic potentiation. *Proc Natl Acad Sci USA* 2001; 98: 14702-14707. Erratum: *Proc Natl Acad Sci USA* 2002; 99: 541.
- 61. Hennigan A, Callaghan CK, Kealy J, Rouine J, Kelly AM. Deficits in LTP and recognition memory in the genetically hypertensive rat are associated with decreased expression of neurotrophic factors and their receptors in the dentate gyrus. *Behav Brain Res* 2009; 197: 371-377.
- 62. Alonso M, Vianna MR, Izquierdo I, Medina JH. Signaling mechanisms mediating BDNF modulation of memory formation in vivo in the hippocampus. *Cell Mol Neurobiol* 2002; 22: 663-674.
- 63. Lee JL, Everitt BJ, Thomas KL. Independent cellular processes for hippocampal memory consolidation and reconsolidation. *Science* 2004; 304: 839-843.
- 64. Komulainen P, Pedersen M, Hanninen T, et al. BDNF is a novel marker of cognitive function in ageing women: the DR's EXTRA Study. Neurobiol Learn Mem 2008; 90: 596-603.
- 65. Gomez-Pinilla F, Ying Z, Opazo P, Roy RR, Edgerton VR. Differential regulation by exercise of BDNF and NT-3 in rat spinal cord and skeletal muscle. *Eur J Neurosci* 2001; 13: 1078-1084.
- van Praag H, Christie BR, Sejnowski TJ, Gage FH. Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc Natl Acad Sci USA* 1999; 96: 13427-13431.
- 67. Gomez-Pinilla F, Vaynman S, Ying Z. Brain-derived neurotrophic factor functions as a metabotrophin to mediate the effects of exercise on cognition. *Eur J Neurosci* 2008; 28: 2278-2287.
- 68. Ma YL, Wang HL, Wu HC, Wei CL, Lee EH. Brain-derived neurotrophic factor antisense oligonucleotide impairs memory retention and inhibits long-term potentiation in rats. *Neuroscience* 1998; 82: 957-967.
- 69. Mu JS, Li WP, Yao ZB, Zhou XF. Deprivation of endogenous brain-derived neurotrophic factor results in impairment of spatial learning and memory in adult rats. *Brain Res* 1999; 835: 259-265.
- Rattiner LM, Davis M, French CT, Ressler KJ. Brainderived neurotrophic factor and tyrosine kinase receptor B involvement in amygdala-dependent fear conditioning. J Neurosci 2004; 24: 4796-4806.
- Cotman CW, Berchtold NC. Exercise: a behavioral intervention to enhance brain health and plasticity. *Trends Neurosci* 2002; 25: 295-301.
- Hopkins ME, Bucci DJ. BDNF expression in perirhinal cortex is associated with exercise-induced improvement in object recognition memory. *Neurobiol Learn Mem* 2010; 94: 278-284.
- 73. Cramer SC. A window into the molecular basis of human brain plasticity. *J Physiol* 2008; 586: 5601.
- 74. Egan MF, Kojima M, Callicott JH, et al. The BDNF Val<sup>66</sup>Met polymorphism affects activity-dependent secretion of BDNF and human memory and hippocampal function. Cell 2003; 112: 257-269.

- Hariri AR, Goldberg TE, Mattay VS, et al. Brain-derived neurotrophic factor val66met polymorphism affects human memory-related hippocampal activity and predicts memory performance. J Neurosci 2003; 23: 6690-6694.
- Kleim JA, Chan S, Pringle E, et al. BDNF val66met polymorphism is associated with modified experiencedependent plasticity in human motor cortex. Nat Neurosci 2006; 9: 735-737.
- 77. Cheeran B, Talelli P, Mori F, *et al.* A common polymorphism in the brain-derived neurotrophic factor gene (BDNF) modulates human cortical plasticity and the response to rTMS. *J Physiol* 2008; 586: 5717-5725.
- Chen ZY, Jing D, Bath KG, et al. Genetic variant BDNF (Val66Met) polymorphism alters anxiety-related behavior. Science 2006; 314: 140-143.
- Soliman F, Glatt CE, Bath KG, et al. A genetic variant BDNF polymorphism alters extinction learning in both mouse and human. Science 2010; 327: 863-866.
- Ninan I, Bath KG, Dagar K, et al. The BDNF Val66Met polymorphism impairs NMDA receptor-dependent synaptic plasticity in the hippocampus. J Neurosci 2010; 30: 8866-8870.
- 81. Rosenfeld RD, Zeni L, Haniu M, *et al.* Purification and identification of brain-derived neurotrophic factor from human serum. *Protein Expr Purif* 1995; 6: 465-471.
- 82. Lommatzsch M, Zingler D, Schuhbaeck K, *et al.* The impact of age, weight and gender on BDNF levels in human platelets and plasma. *Neurobiol Aging* 2005; 26: 115-123.
- 83. Trajkovska V, Marcussen AB, Vinberg M, Hartvig P, Aznar S, Knudsen GM. Measurements of brain-derived neurotrophic factor: methodological aspects and demographical data. *Brain Res Bull* 2007; 73: 143-149.
- Yamamoto H, Gurney ME. Human platelets contain brainderived neurotrophic factor. *J Neurosci* 1990; 10: 3469-3478
- 85. Fujimura H, Altar CA, Chen R, *et al.* Brain-derived neurotrophic factor is stored in human platelets and released by agonist stimulation. *Thromb Haemost* 2002; 87: 728-734.
- Donovan MJ, Lin MI, Wiegn P, et al. Brain derived neurotrophic factor is an endothelial cell survival factor required for intramyocardial vessel stabilization. Development 2000; 127: 4531-4540.
- 87. Nakahashi T, Fujimura H, Altar CA, *et al.* Vascular endothelial cells synthesize and secrete brain-derived neurotrophic factor. *FEBS Lett* 2000; 470: 113-137.
- 88. Kermani P, Hempstead B. Brain-derived neurotrophic factor: a newly described mediator of angiogenesis. *Trends Cardiovasc Med* 2007; 17: 140-143.
- 89. Madri JA. Modeling the neurovascular niche: implications for recovery from CNS injury. *J Physiol Pharmacol* 2009; 60(Suppl 4): 95-104.
- Kerschensteiner M, Gallmeier E, Behrens L, et al. Activated human T cells, B cells, and monocytes produce brainderived neurotrophic factor in vitro and in inflammatory brain lesions: aneuroprotective role of inflammation? J Exp Med 1999; 189: 865-870.
- 91. Dupont-Versteegden EE, Houle JD, Dennis RA *et al.* Exercise-induced gene expression in soleus muscle is dependent on time after spinal cord injury in rats. *Muscle Nerve* 2004; 29: 73-81.
- 92. Matthews VB, Astrom MB, Chan MH, et al. Brain-derived neurotrophic factor is produced by skeletal muscle cells in response to contraction and enhances fat oxidation via activation of AMP-activated protein kinase. *Diabetologia* 2009; 52: 1409-1418.
- 93. Rojas Vega S, Abel T, Lindschulten R, Hollmann W, Bloch W, Struder HK. Impact of exercise on neuroplasticity-related

- proteins in spinal cord injured humans. *Neuroscience* 2008; 153: 1064-1070.
- Poduslo JF, Curran GL. Permeability at the blood-brain and blood-nerve barriers of the neurotrophic factors: NGF, CNTF, NT-3, BDNF. *Brain Res Mol Brain Res* 1996; 36: 280-286
- 95. Pan W, Banks WA, Fasold MB, Bluth J, Kastin AJ. Transport of brain-derived neurotrophic factor across the blood-brain barrier. *Neuropharmacology* 1998; 37: 1553-1561.
- 96. Karege F, Schwald M, Cisse M. Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. *Neurosci Lett* 2002; 328: 261-264.
- 97. Sakane T, Pardridge WM. Carboxyl-directed pegylation of brain-derived neurotrophic factor markedly reduces systemic clearance with minimal loss of biologic activity. *Pharm Res* 1997; 14: 1085-1091.
- 98. Duman RS, Heninger GR, Nestler EJ. A molecular and cellular theory of depression. *Arch Gen Psychiatry* 1997; 54: 597-606.
- 99. Duman RS, Malberg J, Nakagawa S, D'Sa C. Neuronal plasticity and survival in mood disorders. *Biol Psychiatry* 2000; 48: 732-739.
- 100. Shoval G, Weizman A. The possible role of neurotrophins in the pathogenesis and therapy of schizophrenia. Eur Neuropsychopharmacol 2005; 15: 319-329.
- 101. Karege F, Schwald M, Cisse M. Postnatal developmental profile of brain-derived neurotrophic factor in rat brain and platelets. *Neurosci Lett* 2002; 328: 261-264.
- 102. Gonul AS, Akdeniz F, Taneli F, Donat O, Eker C, Vahip S. Effect of treatment on serum brain-derived neurotrophic factor levels in depressed patients. *Eur Arch Psychiatry Clin Neurosci* 2005; 255: 381-386.
- 103. Aydemir O, Deveci A, Taskin OE, Taneli F, Esen-Danaci A. Serum brain-derived neurotrophic factor level in dysthymia: a comparative study with major depressive disorder. *Prog Neuropsychopharmacol Biol Psychiatry* 2007; 31: 1023-1026.
- 104. Kim YK, Lee HP, Won SD, et al. Low plasma BDNF is associated with suicidal behavior in major depression. Prog Neuropsychopharmacol Biol Psychiatry 2007; 31: 78-85.
- 105. Yoshimura R, Mitoma M, Sugita A, et al. Effects of paroxetine or milnacipran on serum brain-derived neurotrophic factor in depressed patients. Prog Neuropsychopharmacol Biol Psychiatry 2007; 31: 1034-1037.
- 106. Dell'Osso L, Carmassi C, Del Debbio A, et al. Brainderived neurotrophic factor plasma levels in patients suffering from post-traumatic stress disorder. Prog Neuropsychopharmacol Biol Psychiatry 2009; 33: 899-902.
- 107. Buckley PF, Pillai A, Evans D, Stirewalt E, Mahadik S. Brain derived neurotropic factor in first-episode psychosis. *Schizophr Res* 2007; 91: 1-5.
- 108. Toyooka K, Asama K, Watanabe Y, et al. Decreased levels of brain-derived neurotrophic factor in serum of chronic schizophrenic patients. Psychiatry Res 2002; 110: 249-257.
- 109. Tan YL, Zhou DF, Cao LY, Zou YZ, Zhang XY. Decreased BDNF in serum of patients with chronic schizophrenia on long-term treatment with antipsychotics. *Neurosci Lett* 2005; 382: 27-32.
- 110. Grillo RW, Ottoni GL, Leke R, Souza DO, Portela LV, Lara DR. Reduced serum BDNF levels in schizophrenic patients on clozapine or typical antipsychotics. *J Psychiatr Res* 2007; 41: 31-35.
- 111. Shimizu E, Hashimoto K, Watanabe H, *et al.* Serum brainderived neurotrophic factor (BDNF) levels in

- schizophrenia are indistinguishable from controls. *Neurosci Lett* 2003; 351: 111-114.
- 112. Huang TL, Lee CT. Associations between serum brainderived neurotrophic factor levels and clinical phenotypes in schizophrenia patients. *J Psychiatr Res* 2006; 40: 664-668.
- 113. Gama CS, Andreazza AC, Kunz M, Berk M, Belmonte-de-Abreu PS, Kapczinski F. Serum levels of brain-derived neurotrophic factor in patients with schizophrenia and bipolar disorder. *Neurosci Lett* 2007; 420: 45-48.
- 114. Reis HJ, Nicolato R, Barbosa IG, Teixeira do Prado PH, Romano-Silva MA, Teixeira AL. Increased serum levels of brain-derived neurotrophic factor in chronic institutionalized patients with schizophrenia. *Neurosci Lett* 2008: 439: 157-159.
- 115. Carlino D, Leone E, Di Cola F, et al. Low serum truncated-BDNF isoform correlates with higher cognitive impairment in schizophrenia. J Psychiatr Res 2010; doi:10.1016/j.jpsychires.2010.06.012
- 116. Krabbe KS, Nielsen AR, Krogh-Madsen R, *et al.* Brainderived neurotrophic factor (BDNF) and type 2 diabetes. *Diabetologia* 2007; 50: 431-438.
- 117. Fujinami A, Ohta K, Obayashi H, et al. Serum brain-derived neurotrophic factor in patients with type 2 diabetes mellitus: relationship to glucose metabolism and biomarkers of insulin resistance. Clin Biochem 2008; 41: 812-817.
- 118. Suwa M, Kishimoto H, Nofuji Y, et al. Serum brain-derived neurotrophic factor level is increased and associated with obesity in newly diagnosed female patients with type 2 diabetes mellitus. Metabolism 2006; 55: 852-857.
- Hristova M, Aloe L. Metabolic syndrome-neurotrophic hypothesis. *Med Hypotheses* 2006; 66: 545-549.
- 120. Zoladz JA, Pilc A, Majerczak J, Grandys M, Zapart-Bukowska J, Duda K. Endurance training increases plasma brain-derived neurotrophic factor concentration in young healthy men. *J Physiol Pharmacol* 2008; 59(Suppl 7): 119-132.
- 121. Ono M, Ichihara J, Nonomura T, *et al.* Brain-derived neurotrophic factor reduces blood glucose level in obese diabetic mice but not in normal mice. *Biochem Biophys Res Commun* 1997; 238: 633-637.
- 122. Ono M, Itakura Y, Nonomura T, Nakagawa T, Nakayama C, Taiji M, Noguchi H. Intermittent administration of brainderived neurotrophic factor ameliorates glucose metabolism in obese diabetic mice. *Metabolism* 2000; 49: 129-133.
- 123. Yamanaka M, Tsuchida A, Nakagawa T, *et al.* Brainderived neurotrophic factor enhances glucose utilization in peripheral tissues of diabetic mice. *Diabetes Obes Metab* 2007; 9: 59-64.
- 124. Tsuchida A, Nonomura T, Nakagawa T, *et al.* Brain-derived neurotrophic factor ameliorates lipid metabolism in diabetic mice. *Diabetes Obes Metab* 2002; 4: 262-269.
- 125. Arentoft A, Sweat V, Starr V, *et al.* Plasma BDNF is reduced among middle-aged and elderly women with impaired insulin function: evidence of a compensatory mechanism. *Brain Cogn* 2009; 71: 147-152.
- 126. Levinger I, Goodman C, Matthews V, et al. BDNF, metabolic risk factors, and resistance training in middleaged individuals. Med Sci Sports Exerc 2008; 40: 535-541.
- 127. Gold SM, Schulz KH, Hartmann S, et al. Basal serum levels and reactivity of nerve growth factor and brainderived neurotrophic factor to standardized acute exercise in multiple sclerosis and controls. J Neuroimmunol 2003; 138: 99-105.
- 128. Rojas Vega S, Struder HK, Vera Wahrmann B, Schmidt A, Bloch W, Hollmann W. Acute BDNF and cortisol response to low intensity exercise and following ramp incremental

- exercise to exhaustion in humans. *Brain Res* 2006; 1121: 59-65.
- 129. Ferris LT, Williams JS, Shen CL. The effect of acute exercise on serum brain-derived neurotrophic factor levels and cognitive function. *Med Sci Sports Exerc* 2007; 39: 728-734
- Winter B, Breitenstein C, Mooren FC, et al. High impact running improves learning. Neurobiol Learn Mem 2007; 87: 597-609.
- 131. Tang SW, Chu E, Hui T, Helmeste D, Law C. Influence of exercise on serum brain-derived neurotrophic factor concentrations in healthy human subjects. *Neurosci Lett* 2008; 431: 62-65.
- 132. Yarrow JF, White LJ, McCoy SC, Borst SE. Training augments resistance exercise induced elevation of circulating brain derived neurotrophic factor (BDNF). *Neurosci Lett* 2010; 479: 161-165.
- 133. Schulz KH, Gold SM, Witte J, *et al.* Impact of aerobic training on immune-endocrine parameters, neurotrophic factors, quality of life and coordinative function in multiple sclerosis. *J Neurol Sci* 2004; 225: 11-18.
- 134. Castellano V, White LJ. Serum brain-derived neurotrophic factor response to aerobic exercise in multiple sclerosis. J Neurol Sci 2008; 269: 85-91.
- Seifert T, Brassard P, Wissenberg M, et al. Endurance training enhances BDNF release from the human brain. Am J Physiol Regul Integr Comp Physiol 2010; 298: R372-R377.
- 136. Goekint M, De Pauw K, Roelands B, et al. Strength training does not influence serum brain-derived neurotrophic factor. Eur J Appl Physiol 2010; 110: 285-293.

- Duman CH, Schlesinger L, Russell DS, Duman RS.
  Voluntary exercise produces antidepressant and anxiolytic behavioral effects in mice. *Brain Res* 2008; 1199: 148-158.
- Cotman CW, Berchtold NC. Exercise: a behavioral intervention to enhance brain health and plasticity. *Trends Neurosci* 2002; 25: 295-301.
- 139. Russo-Neustadt AA, Alejandre H, Garca C, Ivy AS, Chen MJ. Hipocampal brain-derived neurotrophic factor expression following treatment with reboxetine, citalopram, and physical exercise. *Neuropharmacology*, 2004; 29: 2189-2199.
- 140. Rogoz Z, Skuza G, Legutko B. Repeated co-treatment with imipramine and amantadine induces hippocampal brainderived neurotrophic factor gene expression in rats. J Physiol Pharmacol 2007; 58: 219-234.
- 141. Pedersen BK, Pedersen M, Krabbe KS, Bruunsgaard H, Matthews VB, Febbraio MA. Role of exercise-induced brain-derived neurotrophic factor production in the regulation of energy homeostasis in mammals. *Exp Physiol* 2009; 94: 1153-1160.

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