The effects of *Nigella sativa* (Ns), *Anthemis hyalina* (Ah) and *Citrus sinensis* (Cs) extracts on the replication of coronavirus and the expression of TRP genes family

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Abstract Extracts of *Anthemis hyalina* (Ah), *Nigella sativa* (Ns) and peels of *Citrus sinensis* (Cs) have been used as folk medicine to fight antimicrobial diseases. To evaluate the effect of extracts of Ah, Ns and Cs on the replication of coronavirus (CoV) and on the expression of TRP genes during coronavirus infection, HeLa-CEACAM1a (HeLa-epithelial carcinoembryonic antigen-related cell adhesion molecule 1a) cells were inoculated with MHV-A59 (mouse hepatitis virus–A59) at moi of 30. 1/50 dilution of the extracts was found to be the safe active dose. ELISA kits were used to detect the human IL-8 levels. Total RNA was isolated from the infected cells and cDNA was synthesized. Fluidigm Dynamic Array nanofluidic chip 96.96 was used to analyze the mRNA expression of 21 TRP genes and two control genes. Data was analyzed using the BioMark digital array software. Determinations of relative gene expression values were carried out by using the $2^{-\Delta\Delta Ct}$ method (normalized threshold cycle (Ct) value of sample minus normalized Ct value of control). $TCID_{50}$/ml (tissue culture infectious dose that will produce cytopathic effect in 50 % of the inoculated tissue culture cells) was found for treatments to determine the viral loads. The inflammatory cytokine IL-8 level was found to increase for both 24 and 48 h time points following Ns extract treatment. TRPA1, TRPC4, TRPM6, TRPM7, TRPM8 and TRPV4 were the genes which expression levels changed significantly after Ah, Ns or Cs extract treatments. The virus load decreased when any of the Ah, Ns or Cs extracts was added to the CoV infected cells with Ah extract treatment leading to undetectable virus load for both 6 and 8hpi. Although all the extract treatments had an effect on IL-8 secretion, TRP gene expression and virus load after CoV infection, it was the Ah extract treatment that showed the biggest difference in virus load. Therefore Ah extract is the best candidate in our hands that contains potential treatment molecule(s).

Keywords Coronavirus (CoV) · TRP channels · *Anthemis hyalina* · *Nigella sativa* · *Citrus sinensis*

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Introduction

Coronaviruses (CoV), members of the *Coronaviridae* family, are enveloped viruses that contain non-segmented, positive-stranded genomic RNA [1–6]. Ultrastructural analysis of CoV have revealed that they form pleomorphic particles that are roughly spherical but show variations in size (80–120 nm in diameter) and shape [7–10]. The entire CoV replication cycle take places in the cytoplasm. In general, enveloped viruses are able to use a variety of cellular membranes at different steps of their life cycles. In fact, almost any membranous subcellular compartment appears to be used by particular viruses to support their replication complex and assembly process [10, 11].

Coronaviridae family causes various diseases including bronchitis, gastroenteritis, hepatitis systemic diseases and etc. on humans, animal and birds [12, 13]. Both host and viral factors affect the coronavirus virulence and the disease severity in animals. The disease is generally most severe in newborns [14].

In spring 2003, a new human CoV has been infamously notorious due to an epidemic outbreak in South East Asia and Canada [15]. At the time, the accused virus was rapidly identified as the SARS-CoV but it did not look like the human CoVs. The SARS-CoV alarmed the world because it sickened more than 8,000 people and killed nearly 800 of them [16]. Recently, a novel coronavirus caused respiratory infectious disease with high mortality. The virus is called Middle East respiratory syndrome coronavirus (MERS-CoV). This novel MERS-CoV was first reported in Saudi Arabia and eight other countries in 2012 [17]. Currently, there are no approved drugs against CoVs but some potential therapies have been proposed.

It has been revealed that nucleocapsid (N) and non-structural protein 3 (nsp3) interactions is important for CoV replication [18]. The nsp3 interacts with N protein through its EF motif site that contains a calcium binding domain, which can mean that this interaction could depend on calcium. It has been shown that intracellular calcium signaling triggers the elevation of reactive oxygen species in mitochondria that effect replication of influenza virus [19]. Also poliovirus infection increases intracellular calcium concentration [20]. These findings suggest that intracellular calcium concentration might be important for viral replication; therefore viruses may be targeting cellular mechanisms that regulate this concentration.

One of the possible targets of viruses to change the intracellular calcium concentration are the ion channels. The transient receptor potential proteins (TRPs) is group of ion channels family that are responsible for a wide range of cellular functions including adjusting intracellular Ca^{2+} concentration [21]. The TRP superfamily can be divided into seven families: TRPC (canonical), TRPV (vanilloid), TRPM (melastatin), TRPN and TRPA (ankyrin), TRPML (mucolipin) and TRPP (polycystin) [22]. TRP channels are triggered by a many different stimuli such as; growth factors, chemicals, temperature, depletion of intracellular calcium level and etc. Triggering mechanism of channels and selectivity can be different between the TRPs [23]. The expression levels of TRPM and TRPV families have been associated with tumor growth and progression [23–25].

Herbal remedies have been used as folk medicine to treat many diseases. Some herb extracts were shown to inhibit virus replication [26, 27]. One of these herbs is a chamomile plant called *Anthemis hyalina* (Ah). *A. hyalina* belongs to plant family of Asreraceae genus; Anthemis, species; *A. hyalina DC*. Chamomile is regularly used in tea making and

to relieve stomachache and digestive system problems. Chamomile was shown to have anti-inflammatory effects and to decrease wound healing time [28]. Antimicrobial activity of different species in the genus of *Anthemis* have been documented [29]. Twenty-four different components were characterized in Ah extract. The major components were carvacrol (38.4 %) and α-pinene (30.9 %) [30].

Citrus sinensis (Cs) belongs to plant family of Rutaceae, genus citrus, species *Citrus sinensis (L.) Osbeck*. Another herb used for various treatments is Cs, known as sweet orange. In Turkey, people consume Cs by making jam and eating especially during flu seasons in order to get more vitamin C, which is believed to help recover from flu. Cs peels extract contain high amount of flavonoids, limonene and linalool [31]. Cs peel extract has antioxidant and antimicrobial activities [31, 32]. Limonene is an insecticidal compound with low oral or contact toxicity to mammals [33]. Flavonoids have also been shown to have antiviral activity [34] [35].

Nigella Sativa belongs to the plant family of Ranunculaceae, genus; Nigella, species; *Nigella sativa L.* Ns seeds that are known as black cumin is another plant that is used for various treatments. In Turkey, Ns seeds are commonly used on breads and cookies. In addition, some people mix Ns seeds and honey to use for upper respiratory tract infections and some stomach diseases. Ns extract has antiviral, antitumor and antimicrobial activities [36–38]. Ns extract oil contains thymoquinone (27.8–57.0 %), ρ-simen (7.1–15.5 %), karvakrol (5.8–11.6 %), t-anetol (0.25–2.3 %), 4-terpineol (2.0–6.6 %) and longifoline (1.0–8.0 %) [39, 40]. It has been revealed that thymoquinone ve dithymoquinone are active components in Ns extract [41].

In this study we set to find the effects of Ah, Cs and Ns extracts on the replication of CoV and the possible involvement of TRP genes in the replication of CoV.

Materials and methods

Cells, virus and time-course analysis of MHV infection

HeLa-CEACAM1a (the epithelial carcinoembryonic antigen-related cell adhesion molecule 1) and murine fibroblast LR7 cells [42] that were used to propagate and titrate MHV-A59 (mouse hepatitis virus–A59) were maintained in Dulbecco's Modified Eagle Medium (DMEM; Sigma, St. Louis, MO) containing 10 % fetal calf serum (Thermo, Waltham, MA), 100 IU of penicillin/ml and 100 µg/ml of streptomycin (both from Life Technologies, Rochester, NY). HeLa-CEACAM1a cells were inoculated with MHV-A59 at a moi of 30 [43, 44]. After 30 min, the infected cells were washed and maintained in complete medium. Subsequently, the infected cells and culture supernatants were

collected for analysis at 0, 6, 8 h, p.i. When plant extracts were added to the HeLa-CEACAM1a cells, they were added after viral infection and were washed away 1 h later.

Preparation of extracts

Air-dried plant powdered; seeds of Ns, flowers and buds of Ah and peels of Cs were used for extraction (100 g of material of each separately). All extraction experiments were performed by 200 ml ethanol addition onto powders and incubation at room temperature for a 24 h. The extracts were filtered using Whatman filter paper and ethanol was eliminated by rotary vacuum evaporator at 55 °C. The plant extracts were dissolved in 10 ml distilled water.

Quantitative analysis of IL-8 by enzyme-linked immunosorbent assay (ELISA)

ELISA kits were used to detect the human IL-8 according to the user's manual (Quantikine, ELISA, Human CXCL8/IL-8 Immunoassay, DY208, R&D Systems, Minneapolis, MN). HeLa CEACAM1 cells were treated with 1/50 and 1/100 diluted concentrations of Ns, Ah and Cs extracts. Additionally HeLa CEACAM1 cells were infected with MHV-A59. Infected cells were treated with 1/50 and 1/100 diluted concentrations of Ns, Ah and Cs extracts. Subsequently, culture supernatants were collected at 0, 6, and 8 h post infection to measure IL-8 levels by using enzyme-linked immunosorbent assay (ELISA).

Monitoring the extracellular release of the virus

Infected HeLa CEACAM1a supernatants were collected and end point dilutions were made for $TCID_{50}$ analysis at 6 and 8 h post infection. The amount of virus present in the culture supernatants were evaluated by using LR7 cells and $TCID_{50}$ values were calculated.

Isolation of total RNA and TaqMan analysis

Total RNA was isolated from the infected cells using RNeasy mini-kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions with subsequent DNaseI treatment on the column. The amount of RNA concentration was determined by spectrometry using a Nanodrop 100 (Thermo Scientific, DE, USA).

cDNA synthesis

Complementary DNA (cDNA) was synthesized by using Qiagen miScript Reverse Transcription Kit (Qiagen Sample and Assay Technologies, Hilden, Germany) according to manufacturer's instructions. Mixture of 5× miScript RT

buffer and miScript Reverse Transcription mix was prepared at given ratios and 1.25 µl of it was distributed for each reaction. 0.2 µg of RNA samples was added on reaction mixtures and they were incubated at 37 °C for 60 min then at 95 °C for 5 min. Synthesized cDNAs were then diluted 1:5 with low EDTA TE buffer and subsequently transferred on ice block and kept at −20 °C till use.

Fluidigm real-time PCR quantifications

Fluidigm Dynamic Array nanofluidic chip 96.96 was used for mRNA expression assay (Fluidigm, South San Francisco, CA USA). mRNA expression levels of 23 genes were analyzed by BioMark^TM HD System real-time PCR (Fluidigm) (Table 1). Scrutinizing of the data and counting of the generated signals in reaction chambers was achieved by fluidigm real-time PCR Analysis software (Fluidigm).

Intracellular calcium level determination using atomic absorption spectrometer

HeLa CEACAM1a cells were infected with MHV-A59 cells. After infection, HeLa CEACAM1 cells were incubated with Ns, Ah and Cs extracts for 6 and 8 h. Infected cells were collected for quantification of total calcium ions. Concentration of calcium ions (µg/l) was determined by using an AAS-400 Perkin-Elmer (USA) atomic absorption spectrometer (FAAS) in an air-acetylene flame, according to the user's manual provided by the manufacturer. A calcium hallow cathode lamp was used as the radiation source at 423 nm.

Statistical analysis

Data was analyzed using the BioMark digital array software and the numbers of positive chambers were corrected to estimate the true number of copies. This number was used to determine the number of copies in the original sample. Determinations of relative gene expression were carried out by using the $2^{-\Delta\Delta Ct}$ method (comparative Ct method) where Ct values of the samples of interest are compared with a control (uninfected cells) after the Ct values of the control and the samples are normalized to an endogenous housekeeping gene. Expression levels were shown relative to the uninfected cells. GAPDH was used as the housekeeping gene for normalization of the expressions.

Results and discussion
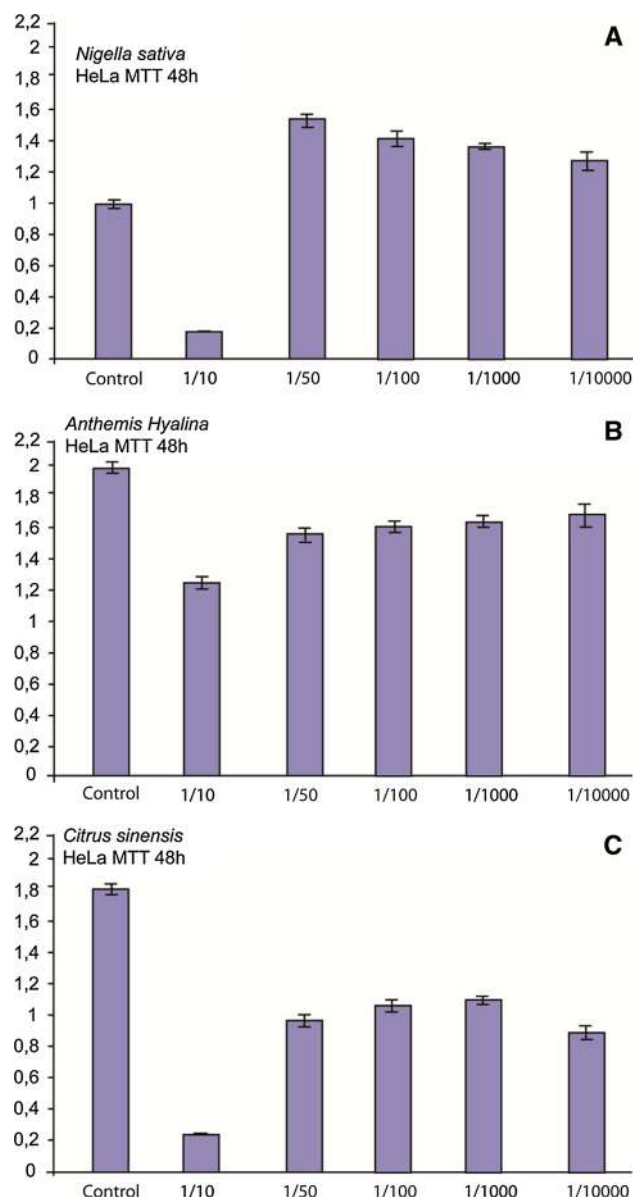
Determination of maximum non-toxic doses

In order to assess any possible toxic effects of Ns, Ah and Cs extracts on HeLa CEACAM1a cells, serial dilutions

Table 1 The list of genes whose expression levels were measured by BioMark™ HD System real-time PCR

TRPA1	TRPC4	TRPC7	TRPM3	TRPM6	TRPV1	TRPV4	GAPDH
TRPC1	TRPC5	TRPM1	TRPM4	TRPM7	TRPV2	TRPV5	
TRPC3	TRPC6	TRPM2	TRPM5	TRPM8	TRPV3	TRPV6	
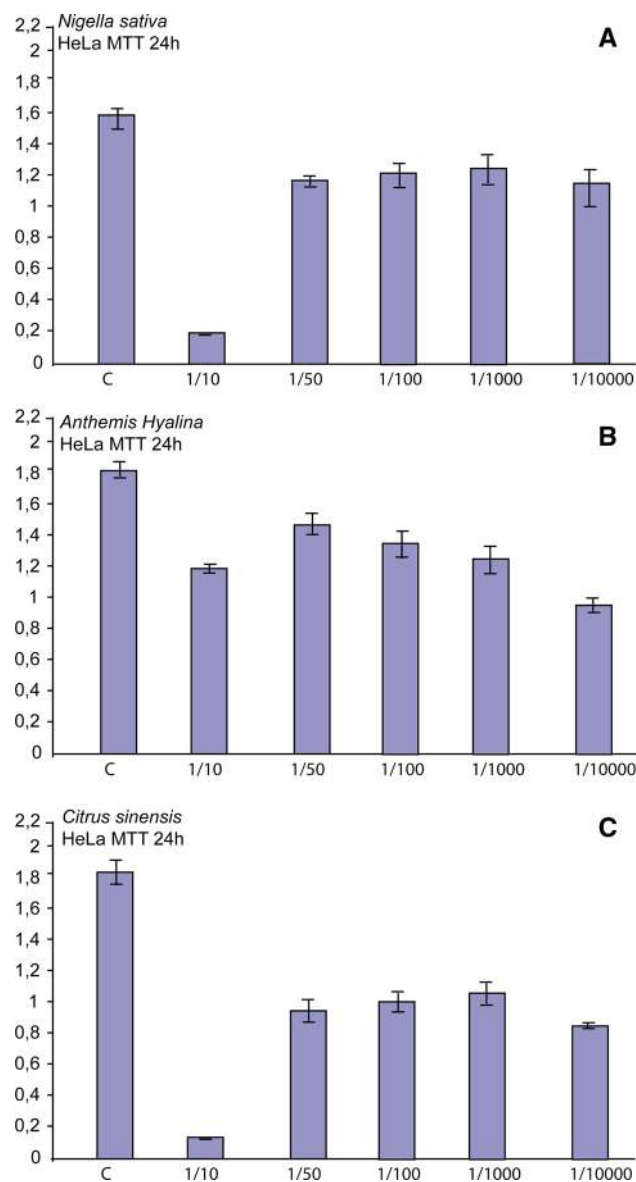
Fig. 1 Effects of Ns, Ah and Cs extracts on the survival of HeLa CEACAM1a cells. HeLa CEACAM1a cells were incubated with the diluted extracts for 24 h and cell toxicity was measured by MTT assay. The *bars* show the averages from 2 independent experiments with 4 repeats for each treatment. *Error bars* show standard error. *C* negative control

Fig. 2 Effects of Ns, Ah and Cs extracts on the survival of HeLa CEACAM1a cells. HeLa CEACAM1a cells were incubated with the extracts for 48 h and cell toxicity was measured by MTT assay. The *bars* show the averages from two independent experiments with four repeats for each treatment. *Error bars* show standard error. *C* negative control

(1/10, 1/50, 1/100, 1/1000, 1/10.000) of Ns, Ah and Cs extracts were incubated with HeLa CEACAM1a cells for 24 and 48 h. The cell viability was assessed by the tetrazolium salt 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium

bromide (MTT) assay. These three plant extracts showed toxic effect at 1/10 dilution, however toxicity of the extracts clearly decreased at 1/50 dilution for 24 and 48 h time points (Figs. 1, 2). These results suggest that the cytotoxic effect of
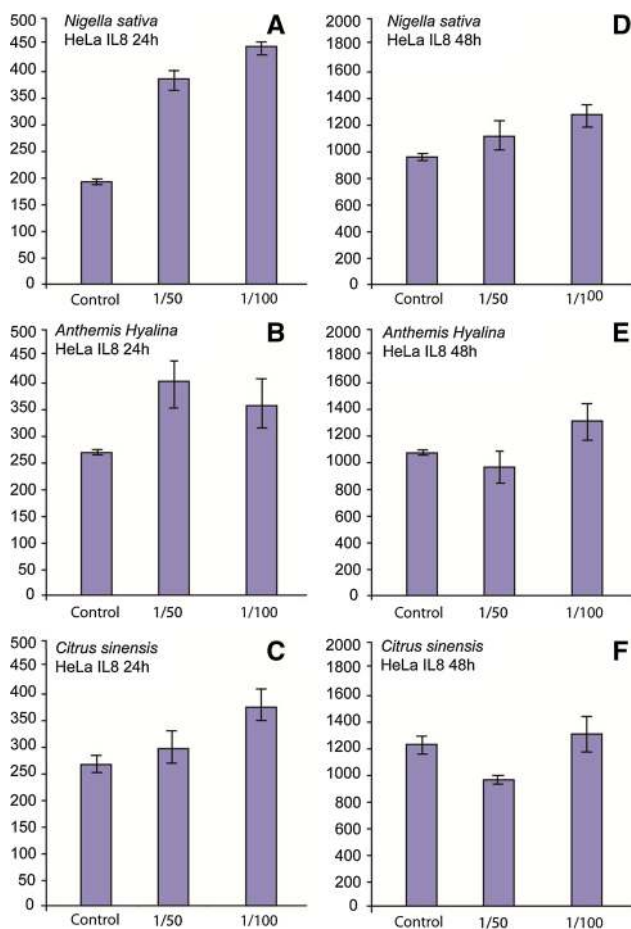
Fig. 3 Effects of Ns, Ah and Cs extracts on the secretion of IL-8 from HeLa CEACAM1a cells after 24 and 48 h. The *bars* show the averages from two independent experiments with four repeats for each treatment. *Error bars* show standard error. *C* negative control

these plants extracts were concentration dependent but not time-dependent in that less cells were viable at 1/10 dilution compared to 1/50 dilution. Based on these results, the plant extract dilution of 1/50 was accepted as an active dose.

IL-8 levels following extract treatments of CoV infected cells

The levels of the inflammatory cytokine IL-8 in the supernatants of cells that were treated with the plant extracts were determined by ELISA (Fig. 3). IL-8 was chosen for its possible role in preventing DC cells from priming T-cells during CoV infection and inducing calcium signaling that might be related to TRP genes [45]. IL-8 levels increased following treatment by Ns, Ah and Cs extracts for both 1/50 and 1/100 dilutions at 24 h. However IL-8 levels had decreased at 48 h for Ah and Cs treated cells. This suggests that Ns extract was better at stimulating

IL-8 secretion from infected cells to lead to a bigger inflammatory response. When IL-8 levels were measured following plant extract treatment of infected cells 6 and 8 h post-infection it was found that IL-8 levels were below the lowest standard and very low (Supplementary figure). Therefore we cannot comment on the effect of plant extracts on IL-8 secretion from CoV infected cells.

Expressions of TRP genes following extract treatment of CoV infected cells

To show effects of the Ns, Ah and Cs extracts on expression of TRP genes during CoV infection, active doses of the Ns, Ah and Cs extracts were added to HeLa CEA-CAM1a cells one hour before CoV infection. Subsequently, the cells were infected with CoV for 6 h post-infection and 8 h. mRNA expression levels for TRP channel families were investigated by using fluidigm RT PCR. GAPDH expression was used for normalization of gene expression. Uninfected cells were used as a negative control. The infected but not plant extract exposed cells were used as positive control.

The expressions of TRP genes were evaluated 6 and 8 h post-infection. These time points were chosen because CoV shows its cytopathic effects by 8 h post-infection [43]. The changes in the expression levels were determined by calculating fold-changes ($2^{-\Delta\Delta Ct}$) for the TRP genes and normalizing them by dividing with the fold-change for the control gene GAPDH.

Fold-change values greater than 1 indicate an up-regulation and fold-change values less than 1 indicate down-regulation. However fold-change values more than 2 and less than 0.5 are considered significant.

Based on the TRP genes expression analysis results, the expression levels were mainly down (Fig. 4). The TRP channels affected by extracts were TRPA1, TRPC4, TRPM6, TRPM7, TRPM8 and TRPV4. Treatment with Ns, Ah and Cs extracts during CoV infection resulted in the down-regulation of TRPM6 and TRPA1 for both 6 and 8 h time points. TRPC4 was down regulated following Ns and Ah treatments for both 6 and 8 h time points. TRPM7 was down regulated only after Ns treatment for the two time points. Ah treatment resulted in down-regulation of TRPV4 for the two time points. Cs treatment resulted in down-regulation of TRPM8 for the two time points.

Monitoring the extracellular release of the virus

In order to determine the number of viruses following extracts treatment and viral infection, TCID50/ml was determined for the conditions and it was found that compared to the control group, extract treatments lowered the

Fig. 4 TRP gene expression levels following Ns, Ah and Cs extract treatment of infected HeLa CEACAM1a cells after 6 (**a**) and 8 h post infections (**b**) compared to uninfected cells. The *bars* show the averages from two independent experiments. *Error bars* show standard error
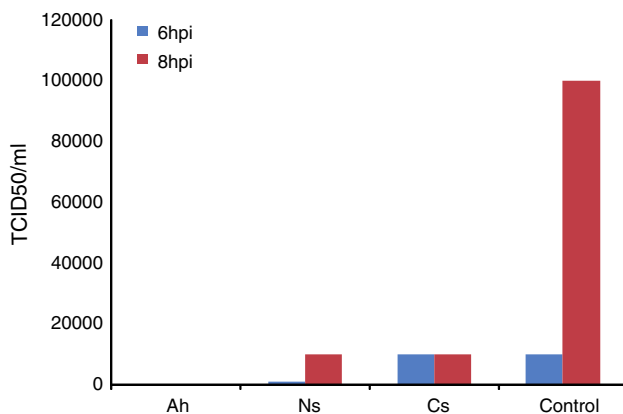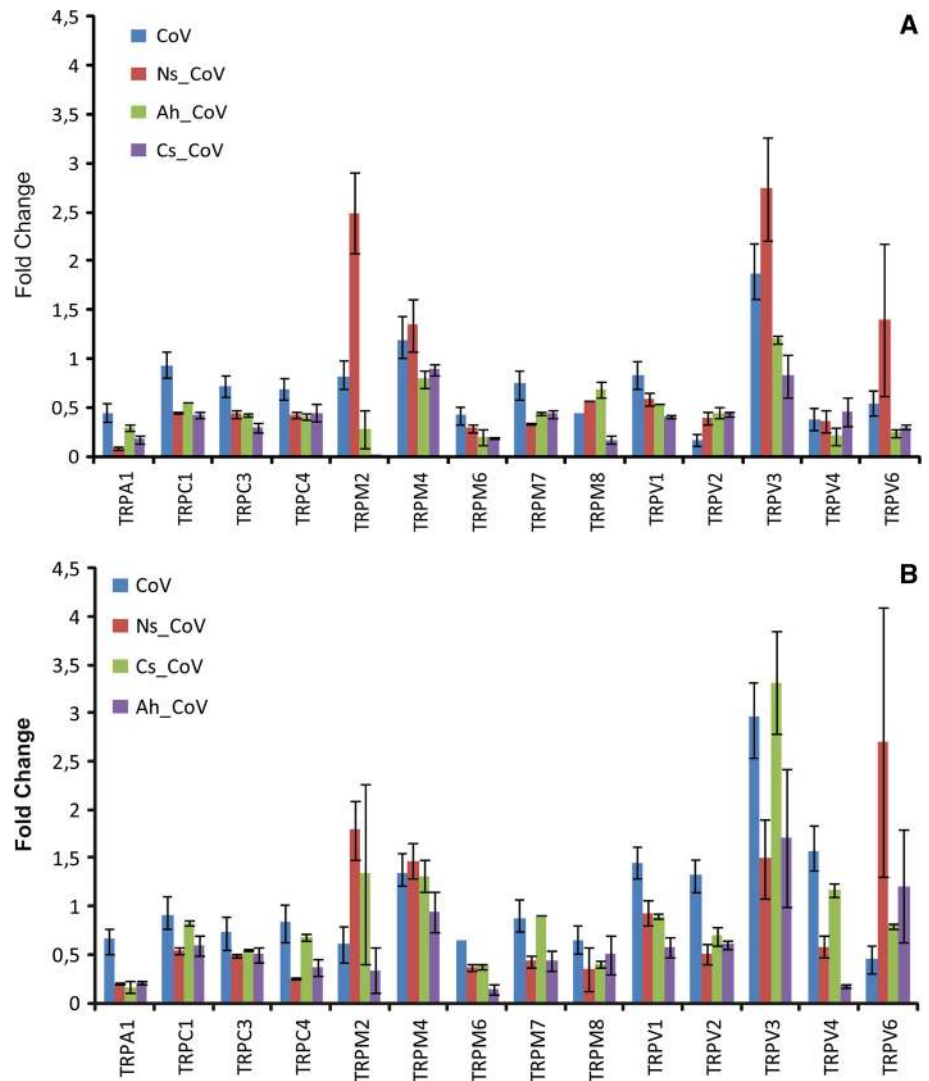

Fig. 5 Virus loads following Ns, Ah and Cs extract treatment of infected HeLa CEACAM1a cells after 6 and 8 h post infections. The amount of virus present in the culture supernatants were evaluated by using LR7 cells and $TCID_{50}$ values were calculated. The *bars* show the averages from two independent experiments with two repeats

virus loads (Fig. 5). In the case of Ah treatments there was no detectable virus. Following Ns treatment the number of viruses was very low at 6 h post infections and it was 1/10th of the control amount by 8 h post infections. Cs treatment resulted in the same number of viruses at 6 and 8 h post infections, which were 1/10th of the positive control numbers by 8 h.

Intracellular calcium levels

Following the determination of the expression of TRP genes, we sought to find the intracellular calcium levels (Fig. 6). Intracellular calcium levels did not change after addition of plant extracts to uninfected cells. However addition of plant extracts to CoV infected cells increased intracellular calcium levels.
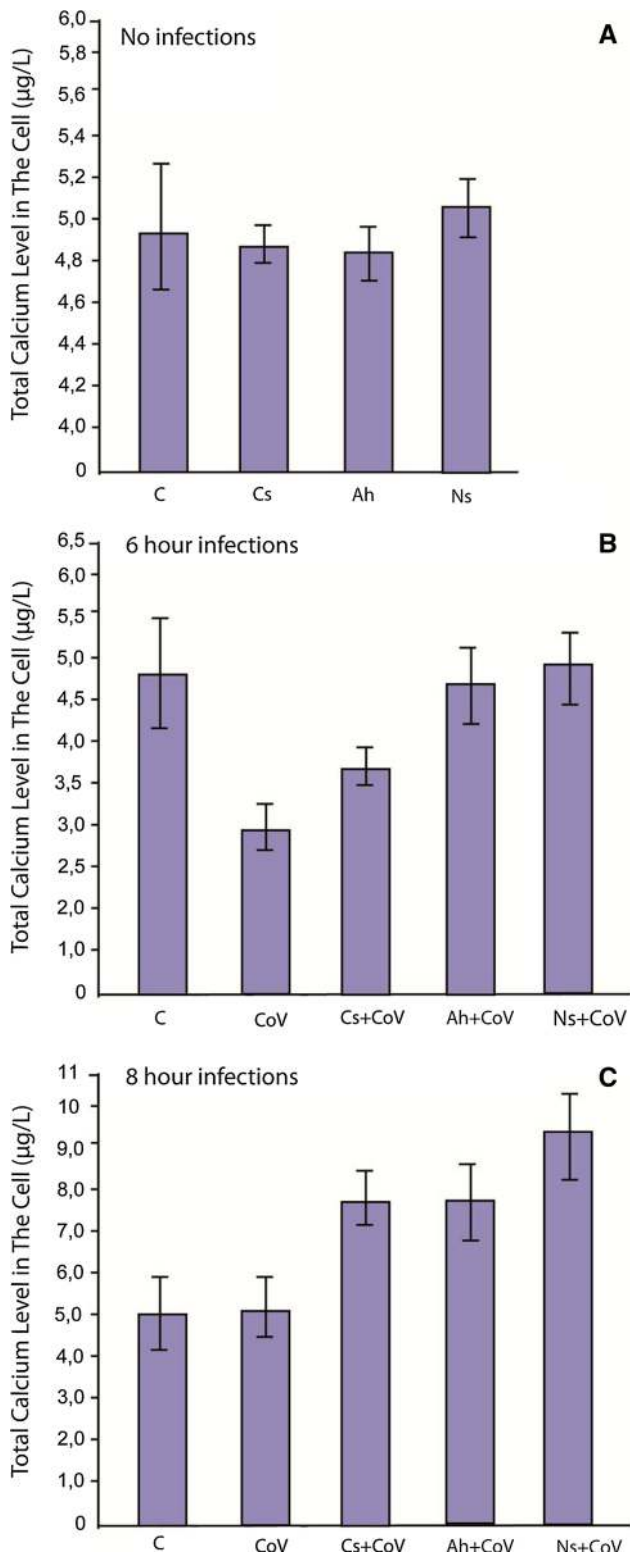
Fig. 6 Intracellular calcium levels in uninfected (**a**) or infected cells 6hpi (**b**) and 8 h post infections (**c**) with and without plant extract treatment

Conclusion

The results presented here suggest that treatment of cells with Ns, Ah or Cs extracts prior to infection with CoV decreases the replication of the virus. IL-8 secretion was increased at 24 h for Ns and Ah extract treatments but Cs treatment did not show a significant increase in IL-8 secretion at 24 h. However, IL-8 secretion was very low at early time points after infection (Fig. 3). Therefore it is not known if IL-8 levels are related to a decreased virus load.

Following gene expression analysis the TRP genes with changed expression levels were identified as follows: TRPM6 and TRPA1 were the two genes that were down regulated for both 6 and 8 h time points in CoV infected and extract treated cells, TRPC4 gene was down regulated following Ns and Ah treatments for both 6 and 8 h time points, TRPM7 was down regulated only after Ns treatment for the two time points, TRPV4 gene was down regulated following Ah treatment for the two time points and TRPM8 was down regulated following Cs treatment for the two time points.

A general down regulation trend for TRP genes was observed in the CoV infected and extract treated cells. Together with the data showing that virus loads were decreased upon extract treatments we predicted that TRP genes might be involved in CoV survival inside cells. Considering that replication of other viruses were related to intracellular calcium concentration and TRPs modulate ion concentrations, including calcium we measured intracellular calcium concentrations and found that they increased following plant extract treatments of infected cells. The controversial TRP gene expression and calcium concentration results suggest that they may not be related to decreased viral loads in plant extract treated infected cells or decreased TRP gene expression and/or increased calcium concentration decrease viral loads through unknown mechanisms. Undetectable levels of the virus after Ah treatment can mean a specific factor being involved following that treatment and this factor can be the TRPV4 gene, whose expression went down only in the case of Ah treatment for both 6 and 8 h time points. Currently there is no literature tying TRP genes to viral survival.

The reason for decreased virus loads following extract treatments is not known at this point. Further research is required in order to identify the specific ingredient(s) in the extracts that led to this phenotype and also to identify the mechanism by which it prevents virus replication or leads to more efficient virus killing. Undetectable virus loads after Ah extract treatment can mean future research can identify certain molecule(s) in Ah extract that can be used

as highly effective drugs against not only CoV but also other viruses as well.

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