

The endocrine activities of 8-prenylnaringenin and related hop (*Humulus lupulus* L.) flavonoids

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ABSTRACT The female flowers of the hop plant have long been used as a preservative and a flavoring agent in beer, but they are now being included in some herbal preparations for women for "breast enhancement". This study investigated the relative estrogenic, androgenic and progestogenic activities of the known phytoestrogen, 8-prenylnaringenin, and structurally related hop flavonoids. 6-Prenylnaringenin, 6,8-diprenylnaringenin and 8-geranylnaringenin exhibited some estrogenicity, but their potency was less than 1% of that of 8-prenylnaringenin. 8-Prenylnaringenin alone competed strongly with 17β -estradiol for binding to both the α - and β -estrogen receptors. None of the compounds (xanthohumol, isoxanthohumol, 8-prenylnaringenin, 6-prenylnaringenin, 3'-geranylchalconaringenin, 6-geranylnaringenin, 8-geranylnaringenin, 4'-O-methyl-3'-prenylchalconaringenin and 6,8-diprenylnaringenin) nor polyphenolic hop extracts showed progestogenic or androgenic bioactivity. These results indicate that the endocrine properties of hops and hop products are due to the very high estrogenic activity of 8-prenylnaringenin and concern must be expressed about the unrestricted use of hops in herbal preparations for women.

The female flowers ("cones") of hops (*Humulus lupulus* L.) are used primarily as a preservative and as a flavoring agent in beer. A recurring suggestion over the years has been that hops have a powerful estrogenic activity. When hops were picked by hand, menstrual disturbances amongst female pickers were common (1). In Germany, hop baths were used for the treatment of gynaecological disorders and hop extracts have been reported to reduce hot flushes in menopausal women (2). Currently hops are being incorporated into a number of commercial preparations for women with claims of "breast enhancement".

We recently identified 8-prenylnaringenin as a very potent phytoestrogen in hops, with an activity equal to or greater than other established plant estrogens (3). The lupulin glands of the hop flowers contain 8-prenylnaringenin along with other prenylflavonoids and the hop acids essential in brewing. The prenylated chalcone, xanthohumol (Fig.1) is the major flavonoid component of the lupulin secretion, reaching a content of about 1% of the dry weight of the hop cone (1).

Other prenylflavonoids including isoxanthohumol, 6-prenylnaringenin and 8-prenylnaringenin as well as geranylated flavonoids are minor constituents and some of these may be derived from non-enzymatic mechanisms (4). The relative proportions of the individual flavonoids can change considerably due to conversions during drying, storage and processing of hops and, furthermore, during brewing, hence various amounts of these compounds are found in hop products and beers (5, 6).

In view of the potent estrogenic activity of 8-prenylnaringenin in hop cones and the variable amounts of the individual prenylflavonoids in hop products, the present study was undertaken to characterize the endocrine activities of the various hop flavonoids to which humans may be exposed. The estrogenic, androgenic and progestogenic actions of the compounds were investigated using sensitive *in-vitro* bioassays (7, 8, 9). In addition, following the reported high affinity of some phytoestrogens for the β -estrogen receptor (ER β) (10), competitive binding studies between 8-prenylnaringenin and 17β -estradiol were performed with purified α - and β -estrogen receptors.

Materials and methods

Polyphenolic extracts of hops. Polyphenolic hop extracts and prenylated/geranylated compounds (xanthohumol, isoxanthohumol, 8-prenylnaringenin, 6-prenylnaringenin, 3'-geranylchalconaringenin, 6-geranylaringenin, 8-geranylaringenin, 4'-*O*-methyl-3'-prenylchalconaringenin and 6,8-diprenylnaringenin) were isolated and identified from hops, as previously described (3, 4). Hop extracts and pure compounds were diluted in ethanol to prepare 1 mM stock solutions. For use in the yeast screens, 20 μ l aliquots were added to individual wells in a 96-well plate and the ethanol was evaporated before adding the hormone-inducible yeasts.

Determination of estrogenic, androgenic progestogenic activity. Hormonal activity was investigated using three separate recombinant yeast screens. Estrogenic activity was determined using an estrogen-inducible yeast screen (*Saccharomyces cerevisiae*) expressing the human estrogen receptor and containing expression plasmids carrying estrogen-responsive sequences controlling the reporter gene lac-Z (encoding the enzyme β -galactosidase) (7). Estrogenic activity was determined from the metabolism of chlorophenol red β -D-galactopyranoside by monitoring the absorbance at 540 nm. The principle of the androgen yeast screen was similar to that of the estrogen screen, with the yeast strain PGKhAR expressing the human androgen receptor (8). The androgen screen responded to testosterone and 5 α -dihydrotestosterone at levels of 10^{-9} M. Both the estrogenic and androgenic yeast screen were originally developed in the Genetics Department of Glaxo Wellcome plc (Stevenage, Herts, UK) and were a gift from Professor J. Sumpter, Brunel University, UK. The progestogenic yeast screen was based on a yeast strain (DY150) containing the DNA sequence of the human progesterone receptor and expression plasmids carrying progesterone-responsive sequences expressing the enzyme β -galactosidase (9). In this assay, the stimulation of β -galactosidase-activity was detected after lysing the cells with 5% CHAPS (11). The progestogen screen responded to progesterone at levels of $>10^{-9}$ M.

Receptor binding. Human recombinant estrogen receptors α and β (ER α and ER β) were obtained from PanVera Corporation (Madison, USA). Dilutions of the compounds were incubated in 100 μ l buffer (10 mM Trizma preset crystals (pH 7.5), 10% glycerol, 2 mM DDT, 1 mg/ml bovine serum albumin) with 15 nM [3 H] 17 β -estradiol (84.0 Ci.mmol $^{-1}$; Amersham Life Science, Amersham UK) and ER (1.5 nM). The mixture was incubated overnight at 4 $^{\circ}$ C and the free and bound hormone were separated using 100 μ l 50% hydroxylapatite slurry (in 50 mM Tris-CL, pH 7.4, 1 mM EDTA). After three washes in buffer (ER α : 40 mM Tris pH 7.5, 100 mM KCl, 1 mM EDTA, 1 mM EGTA; ER β : 40 mM Tris, pH 7.5), the slurry was extracted with 2 washes of 200 μ l ethanol.

Chemicals. All chemicals were obtained from Sigma Chemical, Dorset, UK.

Results

Estrogenic activity was detected in four hop flavonoids (Fig 2), with their activity in the order: 8-prenylnaringenin \gg 6-prenylnaringenin, 8-geranylaringenin and 6,8-diprenylnaringenin. No estrogenicity was found in xanthohumol, isoxanthohumol, 3'-geranylchalconaringenin or 4'-*O*-methyl-3'-prenylchalconaringenin. Also, 6-geranylaringenin was inactive. None of the compounds, nor polyphenolic extracts of hops, showed any activity in either the androgenic or the progestogenic yeast screens (data not shown).

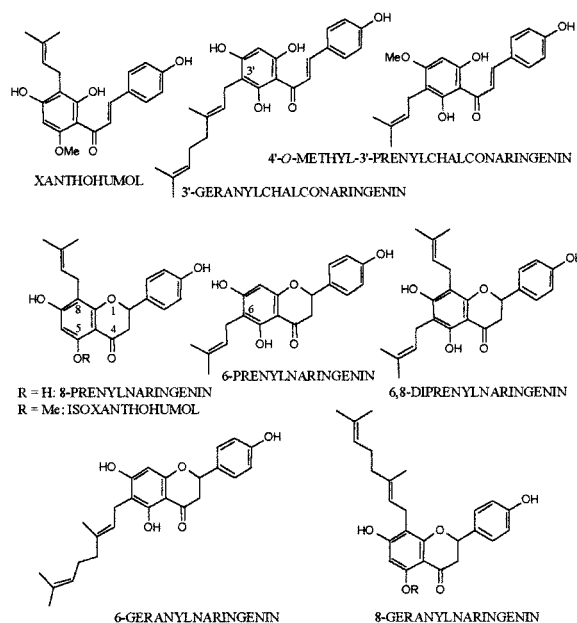


Fig. 1: Structures of hop flavonoids

The estrogenic activity of 8-prenylnaringenin was confirmed in competitive binding assays with purified ER α and ER β . 8-Prenylnaringenin competed strongly with 17 β -estradiol for binding to both receptors with a relative binding affinity of about 0.1 (17 β -estradiol = 1). 8-Geranylaringenin showed some competition (Relative Binding Affinity > 0.001). None of the other compounds was able to displace 3 H-17 β -estradiol from either receptor (Relative Binding Affinity > 0.0001).

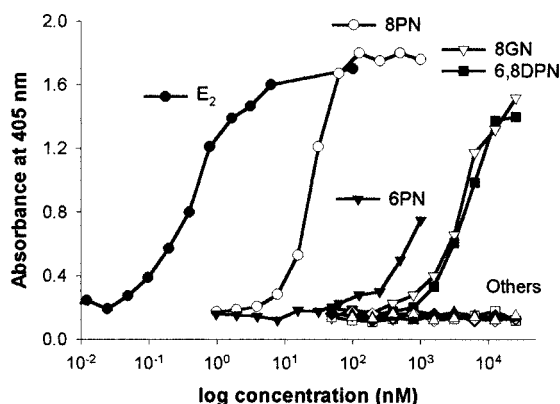


Fig. 2: Assessment of the estrogenic activity of hop-derived prenylflavonoids determined by a yeast screen containing the human estrogen receptor. All points are means \pm s.e.m. of at least 4 determinations. E₂ = 17 β -estradiol; 8PN = 8-prenylnaringenin; 8GN = 8-geranylnaringenin; 6,8DPN = 6,8-diprenylnaringenin; 6PN = 6-geranylnaringenin; Others = xanthohumol, isoxanthohumol, 3'-geranylchalconaringenin and 4'-O-methyl-3'-prenylchalconaringenin.

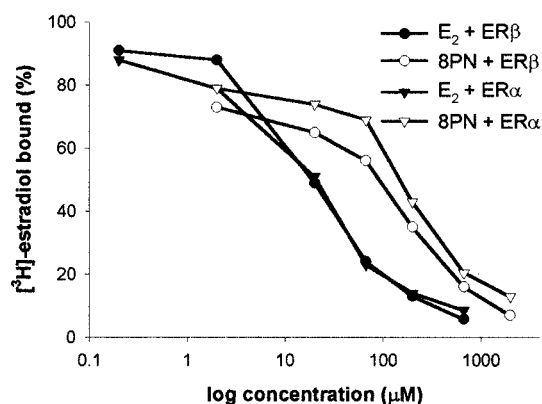


Fig 3: Competitive displacement of [2,4,6,7-³H] 17 β -estradiol from isolated ER α and ER β by 17 β -estradiol and 8-prenylnaringenin. All points are means \pm s.e.m. of 3 determinations. E₂ = 17 β -estradiol; ER = estrogen receptor; 8PN = 8-prenylnaringenin.

Discussion

The results indicate that the endocrine activity of hops and hop products is mainly due to the high estrogenic activity of 8-prenylnaringenin. Some estrogenicity is present in a few other hop-related prenylflavonoids, but the activity is very weak and these compounds are only minor constituents of

hops (5). There was no evidence of either androgenic or progestogenic activity in either polyphenolic hop extracts or pure prenylflavonoids. Like some other phytoestrogens, 8-prenylnaringenin also shows strong affinity for ER β as well as for ER α (10), suggesting that 8-prenylnaringenin may have many potential sites of action within the body.

In addition to the estrogenic activity of 8-prenylnaringenin, a number of other biological activities have been ascribed to individual prenylflavonoids, including antiproliferative effects on breast and colon cancer cell lines, inhibition of cytochrome P450-mediated activation of procarcinogens, inhibition of bone resorption and inhibition of diacylglycerol acyltransferase activity (4, 12-17). This range of potential bioactivities raises the question of whether the exposure to the prenylflavonoids in hops has any physiological significance to humans. The main use of hops is in the flavoring of beer and the content of prenylflavonoids in beers can be up to 4 mg/L. Isoxanthohumol, xanthohumol and 6-prenylnaringenin constitute more than 90% of the total amount of prenylflavonoids in beer, with 8-prenylnaringenin and 6-geranylnaringenin being minor components (\leq 0.24 and 0.07 mg/L, respectively) (5, 6). Whether any of the reported health-beneficial effects of moderate beer consumption (18, 19) can be attributed to prenylflavonoids remains to be determined.

The current-day incorporation of hops into a number of herbal preparations for women, including those claiming "breast enlargement", is of more immediate concern because it may result in much higher levels of human exposure to prenylflavonoids and other hop compounds. Controlled clinical trials of hop constituents have not been done so far. It is conceivable that the claimed efficacy of 8-prenylnaringenin reflects the action as an estrogen either centrally (hot flushes) or peripherally (breast tissue). If this were the case, the potential for adverse effects (e.g. in relation to fertility and hormone-related cancer) must also be considered. The biological effects of prenylated flavonoids within the body are poorly understood and scientific evaluation of the safety of high exposure levels is essential.

Acknowledgements

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References

- 1 Verzele M 1986 100 Years of hop chemistry and its relevance to brewing. *J. Inst. Brew.*, 92: 32-48.
- 2 Goetz P 1990 Traitement des bouffées de chaleur par insuffisance ovarienne par l'extrait de houblon (*Humulus lupulus*). *Rev. Phytothérapie Pratique*, 4: 13-15.
- 3 Milligan SR, Kalita JC, Heyerick A, Rong H, De Cooman L, De Keukeleire D 1999 Identification of a potent phyto-oestrogen in hops (*Humulus lupulus* L.) and beer. *J. Clin. Endocrinol. Metab.*, 83, 2249-2252.

- 4 **Stevens JF, Taylor AW, Nickerson GB, Ivancic M, Henning J, Haunold A, Deinzer ML** 1998 Prenylflavonoid variation in *Humulus lupulus*: distribution and taxonomic significance of xanthohalenol and 4'-*O*-methylxanthohumol. *Phytochemistry*, 53: 759-775.
- 5 **Stevens JF, Taylor AW, Deinzer ML** 1999 Quantitative analysis of xanthohumol and related prenylflavonoids in hops and beer by liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A*, 832: 97-107.
- 6 **Rong H, Zhao Y, Lazou K, De Keukeleire D, Milligan SR, Sandra P** 2000 Quantitation of 8-prenylnaringenin, a novel phytoestrogen in hops (*Humulus lupulus* L.), hop products and beers, by benchtop HPLC using electrospray ionization. *Chromatographia*, 51: 545-552.
- 7 **Routledge EJ, Sumpter J P** 1996 Estrogenic activity of surfactants and some of their degradation products assessed using a recombinant yeast screen. *Environ. Toxicol. Chem.*, 15: 241-258.
- 8 **Sohoni P, Sumpter J P** 1998 Several environmental oestrogens are also anti-androgens. *J. Endocrinol.*, 158: 327-339.
- 9 **Jin L, Tran DQ, Ide CF, McLachlan JA, Arnold SF** 1997 Several synthetic chemicals inhibit progesterone receptor-mediated transactivation in yeast. *Biochem. Biophys. Res. Commun.*, 233: 139-146.
- 10 **Kuiper GGJM, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, Gustafsson JA** 1997 Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinol.*, 138: 863-870.
- 11 **Iñiguez-Lluhi JA, Lou DY, Yamamoto KR** 1997 Three amino acid substitutions selectively disrupt the activation but not the repression function of the glucocorticoid receptor N-terminus. *J. Biol. Chem.*, 272: 4149-4156.
- 12 **Tobe H, Muraki Y, Kitamura K, Komiyama O, Sato Y, Sugioka T, Maruyama HB** 1997 Bone resorption inhibitors from hop extract. *Biosci. Biotechnol. Biochem.*, 61: 158-159.
- 13 **Miyamoto M, Matsushita Y, Kiyokawa A, Fukuda C, Iijima Y, Sugano M, Akiyama T** 1998 Prenylflavonoids: a new class of non-steroidal phytoestrogens (Part 2). Estrogenic effects of 8-isopentenylnaringenin on bone metabolism. *Planta Med.*, 64: 516-519.
- 14 **De Keukeleire D, De Cooman L, Rong H, Heyerick A, Kalita JC, Milligan SR** 1999 Functional properties of hop polyphenols. In *Plant Polyphenols 2: Chemistry, Biology, Pharmacology, Ecology*. Gross GG, Hemingway RW, Yoshida T (Eds.). Kluwer Academic/Plenum Publishers, New York, USA, pp. 739-760.
- 15 **Henderson MC, Miranda CL, Stevens JF, Deinzer ML, Buhler DR** 2000 *In-vitro* inhibition of human P450 enzymes by prenylated flavonoids from hops, *Humulus lupulus*. *Xenobiotica*, 30: 235-251.
- 16 **Miranda CL, Aponso GL, Stevens JF, Deinzer ML, Buhler DR** 2000 Prenylated chalcones and flavanones as inducers of quinone reductase in mouse Hepa 1c1c7 cells. *Cancer Lett.*, 149: 21-29.
- 17 **Miranda CL, Stevens JF, Helmrich A, Henderson MC, Rodriguez RJ, Yang YH, Deinzer ML, Barnes DW, Buhler DR** 1999 Antiproliferative and cytotoxic effects of prenylated flavonoids from hops (*Humulus lupulus*) in human cancer cell lines. *Food Chem. Toxicol.*, 37: 271-285.
- 18 **Abu-Amsha R, Croft KD, Puddey IB, Proudfoot JM, Beilin LJ** 1996 Phenolic content of various beverages determines the extent of inhibition of human serum and low-density lipoprotein oxidation *in vitro*: identification and mechanism of action of some cinnamic acid derivatives from red wine. *Clin. Sci.*, 9: 449-458.
- 19 **Klatsky AL, Armstrong M A, Friedman GD** 1997 Red wine, white wine, liquor, beer, and risk for coronary artery disease hospitalization. *Am. J. Cardiol.*, 80: 416-420.