INVITED REVIEW THE EXPERIMENTAL GRANULOMA. A HYPOTHESIS TO EXPLAIN THE PERSISTENCE OF THE LESION

M. MARIANO

SUMMARY

Granulomatous inflammation is the morphological substrate of a variety of important infectious diseases such as tuberculosis, leprosy, schistosomiasis and others. Nevertheless, although many aspects of this special type of inflammation are known, fundamental questions concerning granuloma formation, persistence, fate and significance for host-parasite relationships still remain to be elucidated. In this brief review, the basic and more relevant literature related to experimental investigations on granuloma physiopathology is presented. Based on recent investigations performed in our laboratory showing that MDF (Macrophage Deactivating Fator) secreted by epithelioid cells and characterized as the calcium-binding protein protein MRP-14 deactivates activated macrophages, a hypothesis to explain the persistence of granulomatous inflammation is put forward.

KEYWORDS: Macrophage; Epithelioid cell; MDF; MRP-14; Granulona; Granulomatous inflammation.

INTRODUCTION

The last hundred years were not enough to draw a comprehensive picture of the mechanisms which govern the natural resistance to infection. Different areas of investigation have focused on the phenomenon and since then the inflammatory response, elicited by the presence of the parasite in a tissue, has been regarded as a major component of the process. With a few exceptions the first interaction between the parasite and the host takes place in an inflammatory *milieu*. The future course of infection depends largely on the effectiveness of phagocytic cells, specific antibodies, activated lymphoid cells and other unknown factors responsible for the killing and degradation of the parasite. Depending on the virulence of the parasite and the natural resistance of the host, the infection may be aborted, the

host killed or, an unstable equilibrium between host and parasite be established with the appearence of signals and symptoms of the disease. If an unstable host-parasite equilibrium prevails, the inflammatory process originating in the infected organ or tissue may have peculiar characteristics which depend on the specific properties of the parasite and on the genetic background of the host. This is the case for microorganisms and certain materials capable of inducing a characteristic response of the affected tissue - the granulomatous reaction - described more than a century ago by Virchow. The granulomatous inflammation is the morphological substrate for many infectious diseases, such as tuberculosis, leprosy, brucellosis, schistosomiasis, sarcoidosis and a large number of visceral mycosis. In

addition, noninfectious irritating agents such as talc, silica, berillum or zirconion are capable of evoking a granulomatous reaction in tissues to which they are applied. The large incidence of the granulomatous response in a variety of pathologic states is sufficient to encourage any effort to understand the unknown mechanisms which determine granuloma.formation. In this review the basic information which has been obtained from experimental studies on granulomatous inflammation will be summarized. Based on experiments showing that epithelioid cells secret a factor which deactivates activated macrophages, a hypothesis is proposed to explain the persistence of this peculiar type of inflammatory response.

Morphology of the granuloma

The typical granuloma induced by the Mycobacterium tuberculosis in man and laboratory animals, is a focal collection of mononuclear cells surrounded by connective tissue which includes fibroblasts, collagen fibers and newly formed vessels. A halo of small round cells, comprising newly arrived monocytes and a small number of lymphocytes is observed between the proliferating connective tissue cells and the focus of the lesion. Large mononuclear cells are arranged in a palisade formation which occupies the centre of the lesion. These cells exhibit a huge cytoplasm and a nucleus in which the chromatin is dispersed. They are referred to as epithelioid cells because of their resemblance to epithelial cells. Among the epithelioid cells, multinucleated cells or macrophage polykarya are observed and a central area of necrosis may occur ^{2, 4}. The unique morphological characteristic of the granulomatous inflammation is the occurrence of epithelioid cells and the concentric appearence of the whole lesion. Some epithelioid granulomas are also heavily invaded by neutrophils 30 or by eosinophils [reviewed in ref. 12].

Granulomatogenic "Irritants"

Two basic categories of etiologic factors are recognized as inducers of granulomatous inflammatory responses. The first comprises inert substances that are unable to mount an immune response. These are artificially termed foreign-body types of agents and although non-immunogenic, their chemical properties render them able to generate granulomas which differ in evolution, dynamics, duration, severity and involution rate from infectious granulomas. There is a spec-

trum of responses to noninfectious "foreign bodies" ranging from a discrete or "quiescent" granuloma as induced by plastic beads ³⁷, bentonite ³⁷, carrageenan ⁷¹ in one pole, to a vigorous, "active" process as induced by silica 77, cord factor 7, streptococcal cell wall 78 on the opposite pole. The second category of granulomatogenic factors can induce immune reactions which drastically interfere with the dynamics and fate of the granulomatous lesion. To this category belong pathogens such as bacteria, fungi, viruses, protozoa and worms. Also metals such as zirconium and beryllium 38, 109 and artificially-made "irritants" composed of "foreignbody" particles, such as bentonite, coupled to an antigenic substance are included ³⁷. Despite their diversified nature, all granulomatogenic factors share one basic property, namely, they are poorly degradable materials 97. 98. However, this property alone can not explain why a granulomatous lesion develops under the influence of such agents rather than a diffuse chronic inflammatory response.

Cells in Granulomatous Inflammation

Cells which comprise the typical tuberculous granuloma are macrophages, epithelioid cells, macrophage polykaria or inflammatory giant cells, and lymphocytes. (Fig. 1) The distribution of these cells within the lesion has been thoroughly described [2,4 reviewed in 1, 12]. However, some granulomatous lesions, as reported by ADAMS 12, do not contain all the cell types mentioned above. Furthermore, the lesions may exhibit infiltration of polymorphonuclear leucocytes as seen in fungal induced lesions 30,72 or of plasma cells as is the case in schistosomotic lesions 12. In our view, the epithelioid cell is the characteristic component of the granulomatous reaction 70. Fibroblasts and newly formed vessels are seen to proliferate, particularly on the peripheral border of the developing tuberculous granuloma 25. The extent to which this occurs, however, depends on both the nature of the causative agent and the animal species under study (BIRMAN & MARIANO, unpublished results).

The macrophage

The macrophage is the basic architectural and functional unit of chronic inflammation. While the morphology and function of resident, stimulated and activated macrophages from different body cavities has been extensively investigated in laboratory animals [reviewed in: 5,58,74,106,114], the characterization of macrophages in chronic lesions, particularly in

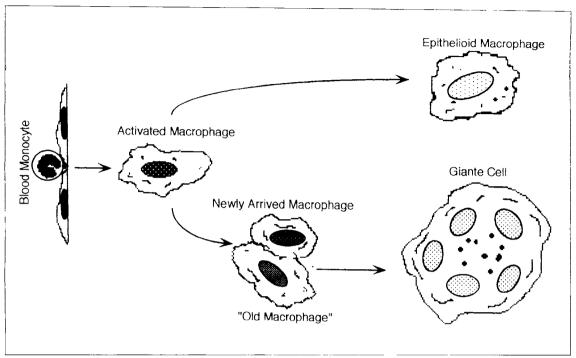


Fig. 1 - Schematic representation of macrophage transformation into epithelioid or giant cells in granulomatous lesions.

granulomas, has been hindered mainly because of the wide morphological variation found among these cells. ADAMS ² made a carefull ultrastructural study of BCG-induced granulomas in guinea pigs and showed that newly arrived monocytes are initially simple cells which progressively increase their nuclear euchromatin content, develop prominent nucleoli, extensive cytoplasm, free ribosomes, abundant Golgi apparatus, mitochondria and large lysosomes, and finally acquire the typical morphology of inflammatory macrophages.

Origin of the macrophage - Cytokinetic studies have unquestionably established that macrophages participating in chronic inflammatory reactions originate from a precursor cell in the bone marrow, viz. the promonocyte which matures into the monocyte and reaches the injured tissue via the circulating blood ^{96,112}. These findings allowed a new interpretation of the concept of "chronic proliferative lesion", according to which the inflammatory granuloma would be generated by the multiplication of tissue histiocytes ⁴. The current view states that once at the inflammed site, macrophages may proliferate, the intensity of proliferation depending chiefly on the nature of the irritant ⁸⁸. Immunity may also play a part in the phenomenon through the enhancement of the rate of macrophage

proliferation ^{8, 26}. Studies on BCG-induced granulomas in rats showed that macrophages divide only twice during their first week of residence in tissues and that local division does not substantialy contribute to the total number of cells in the lesion ^{88, 94}.

Turnover - The development of a granulomatous lesion is a very active cellular process in which migrated monocytes may proliferate 88.94, be actively recruited from blood 96, 112 or become long lived 88, 95, depending on the type of irritant used to induce the lesion. In terms of cell kinetics, granulomata represent a spectrum with turnover reaction at one end (tuberculous lesions and lesions induced by B. pertussis vaccine) and low turnover reaction at the other end (carrageenan-induced lesion) 95. The turnover rate is reflected both in the speed of entry of new cells and the rate of division of existing cells [reviewed in 99]. Another relevant factor modulating cell turnover in a granulomatous lesion is the immune state of the host. An accelerated rate of macrophage turnover was observed in tuberculin hypersensitivity lesions induced in rabbits previously immunized with BCG 26. Little is known, however, about the nature of the chemotactic factors which govern cell kinetic variations in the lesions.

Specific surface antigens - Mouse peritoneal macrophages exhibit specific antigens on their surface membranes. These antigens are detected by specific antisera and are reliable markers which allow distinguish macrophages from lymphocytes and other mononuclear cells 32, 45, 84. However less clear results are obtained when antisera are tested against macrophages other than those obtained from the peritoneal cavity 66. Macrophages at other sites may be more heterogeneous either because they are derived from different precursors or because they are at different stages of differentiation. Recent experimental evidence favors the later possibility (Discussed in Epithelioid cell - Differentiation antigens). Similar results were obtained with macrophages cultured "in vitro" 107. The possible significance of these findings for the pathobiology of chronic lesions remains to be elucidated.

Surface receptors and phagocytosis - Macrophages are multifunctional cells. They are phagocytic and are capable of secreting a large variety of biologically active substances [reviewed in: 10]; they are involved in the processing and presentation of antigens to lymphoid cells; and in some way they regulate the immune response³². However, the ability to phagocytose and eventually digest foreign materials is its most characteristic activity. Phagocytosis by macrophages has been extensively studied "in vitro" [reviewed in: 102,103]. Cell membrane receptors involved in the phemomenon have been demonstrated on these cells. Whereas phagocytosis by macrophages has been thoroughly studied "in vitro", little is known about the phagocytic activity of macrophages in chronic inflammatory lesions. Subcutaneous implantation of glass coverslips in the mouse induces a foreignbody type of inflammatory reaction which has proved useful for the study of the phagocytic activity of macrophages "in vivo". Adherent cells stick to the glass surface and can be collected for microscopic observation 88, 112. With this technique phagocytosis mediated by Fc, C3b, or other surface receptors, could be demonstrate with cells that were recovered from coverslips implanted in mice for less than 14 days. Later, most macrophages appear to loose surface receptors with the consequent impairment of phagocytic activity 64, 80. Similar findings were observed in chickens 46. Results obtained in guinea-pigs indicate that if M. tuberculosis is subcutaneously inoculated with the coverslip, macrophages on the glass surface are significantly more phagocytic, provided the animal was previously immunized with CFA. This increased phagocytic activity is maximal at day 3 to 6 and then declines to control levels on the fourteenth day post infection ⁶⁵. The increased phagocytic activity of macrophages in delayed hypersensitivity mediated lesions was also observed in cyclophosphamide-treated mice immunized with sheep red blood cells ⁹.

Epithelioid cell

Inflammatory macrophages may develop into epithelioid cells which are the characteristic components of the granulomatous inflammation. The epithelioid cell is recognized under the light microscope as possessing a pale, elongated nucleus and a cloudy eosinophilic cytoplasm whose outline can merge with that of the neighboring cells. Epithelioid cells arrange themselves either in layers or form discrete aggregates. They are found mainly in the central part of a lesion or in surrounding necrotic areas [4, reviewed in 39]. Observed under the electron microscope, epithelioid cells resemble large polygonal macrophages with a large euchromatic nucleus, prominent nucleoli, and an abundant cytoplasm filled with many lysosomes and mitochondria. An extensive endoplasmic reticulum is seen, and bundles of microfilaments may be observed. The cells appear closely associated, and are interlocked by twinning pseudopodes ^{2, 13}. A characteristic type of cytoplasmic granules has been described in epithelioid cells 111.

Origin and differentiation - It is now clear that the epithelioid cell do derive from the macrophage. PAPADIMITRIOU & SPECTOR 79 implanted pieces of cellophane into the subcutaneous tissue of mice and recovered cells morphologically alike to the epithelioid component of granulomas. Epithelioid-like cells were also described in foreign-body lesions induced in mice by subcutaneous implantation of glass coverslips 64, 111. In a detailed morphological study of BCG-induced granulomas in guinea pigs, ADAMS 2 reported the changes undergone by monocytes during their transformation into epithelioid cells. Five distinct stages were then sequentially defined in the course of cell differentiation: monocyte, immature macrophage, macrophage, immature epithelioid cell, and epithelioid cell. Immature epithelioid cells are observed in 21 day old lesions. Epithelioid cells are first recognizable after approximately 30 days. These observations were confirmed in experiments in which lesions were induced by killed tubercle bacilli on CFA-sensitized guinea pigs 13.

Epithelioid cells appear to have a reduced capacity to proliferate as compared to newly arrived macrophages ¹⁰⁸.

Differentiation antigens - SORG and his group 76 cloned two calcium-binding proteins from the supernatant of MIF reaction. These proteins which belongs to the family of the S-100 calcium-bindind proteins ⁵⁶, named MRP-8 and MRP-14 (MIF Related Proteins with molecular weight of 8 and 14), were shown to be present in neutrophils and monocytes 115. Macrophages and neutrophils express these proteins in different types of inflammatory reaction. Nevertheless, as macrophages mature in the inflammatory foci they no longer express these markers 86. The real role played by these proteins in leukocyte physiology and pathology remains to be determined [reviewed in 49]. Nevertheless, DELABIE et al. 31 demonstrated that, contrary to observed in inflammatory macrophages in which the simultaneous expression of MRP-8/14 is a constant, only MRP-14 can be detected in the cytoplasm of epithelioid cells of typical tuberculous granulomas in man. According to these authors, "foreign-body" granulomas do not present this pattern of reaction. Using monospecific antibodies directed to these proteins in immunostained preparations, we have shown that epithelioid macrophages from coverslips implanted into the subcutaneous tissue of mice, a typical foreign-body granuloma, also express MRP-14 but not MRP-8 (MARIANO et al. in preparation). Yet, we have confirmed the preponderant expression of MRP-14 in epithelioid cells from different types of granulomas in man including "foreign-body" granulomas induced by the presence of silk suture thred in the tissues. These finds undoubtedly opens the opportunity to revise the concept of "epithelioid cell" in chronic inflammation.

Phagocytosis - The ultrastructural organization of epithelioid cells suggests that they are poorly phagocytic. Indeed, PAPADEMITRIOU & SPECTOR 79 found that cells developing on implanted cellophane sheets and endowed with morphological characteristics of epithelioid cells were less phagocytic than their macrophage precursors. On the other hand, they exhibit active pinocytosis and exocytosis of colloidal gold particles. Inflammatory macrophages, which adhere to the surface of subcutaneously inserted coverslips, express Fc, C3b and other surface receptors and are actively phagocytic via these binding sites 64. However, about 50% of the mono and multinucleated cells which are recovered from 21 day old coverslips no longer express

these surface receptors and no longer phagocytose test particles. Ultrastructurally, these cells have the classical morphological characteristics of epithelioid cells. Impaired phagocytic activity has also been observed in giant cells. This appears to suggest that the macrophage-derived giant cells may also undergo an epithelioid transformation. One of the unsolved problems regarding the biology of epithelioid cells is whether this type of macrophage transformation is a permanent and stable phenomenon. Experimental evidence shows that the impaired phagocytic capacity of epithelioid cells may be reversed. When epithelioid macrophages and macrophage polykarya are incubated with a mixture of antibody coated sheep red blood cells and levamisole they actively phagocyte the complex. indicating that at least in some circumstances epithelioid cells may regain the ability to phagocytose 60.

Epithelioid cell function - The function(s) of epithelioid cells in granuloma physiopathology is still obscure. Based on the morphology of these cells PAPADIMITRIOU & SPECTOR ⁷⁹ have suggested that secretion of biologically active factors is the main function of these cells. CAMARERO et al. ^{17, 18} demonstrated that "pre-epithelioid" and epithelioid cell populations on the surface of implanted glass coverslips in mice, secret a factor(s) which blocks O₂ release from newly arrived macrophages. Besides this, secretory capacity of epithelioid cells were shown to control the rate of multiplication of Mycobacterium tuberculosis-BCG in granulomas induced by this bacteria in the hamster check-pouch ⁹¹.

"In vitro" culture of epithelioid cells - When mammalian macrophages are cultured "in vitro", the typical epithelioid transformation is not observed. Chicken blood mononuclear cells were cultured "in vitro" and cells with typical epithelioid morphology were found in the culture 105. Subsequently, however, it was observed that more than 90% of the mononuclear cells that adhere to culture dishes were thrombocytes and not monocytes as previously considered. Chicken thrombocytes can exhibit a behaviour analogous to mouse inflammatory macrophages because they are also able to fuse and form giant cells which then loose surface receptors involved in phagocytosis 46. Attempts have been made to maintain an "in vitro" culture of cells adherent to coverslip-implants in subcutaneous tissues of mice. The attempts were unsuccessful: after 3 days in culture, adherent macrophages and macrophage polykarya begin to round up, detach from the glass surface, and start to show morphological signs of cell degeneration (personal observations). Further investigations are needed to understand this phenomenon.

Giant cell (Macrophage polykarion)

The presence of multinucleated cells is a constant feature in certain types of chronic inflammatory lesions. These cells are seen in tuberculous lesions in man and bovine cattle. At least two cell categories can be distinguished by their morphological characteristics: the Langhans giant cells which have nuclei disposed peripherally so as to form a crown-like figure, and the foreign-body giant cells with nuclei dispersed throughout the cytoplasm in a somewhat less organized manner. The Langhans cell type probably derives from the foreign-body type cell and represents a more advanced/mature cell stage 61. The significance of cell multinucleation in these lesions is still obscure. In tuberculous lesions, its occurrence appears to be speciesdependent since multinucleated cells are rarely observed in dogs and cats ³⁶. In experimentally induced tuberculous lesions, multinucleated cells are seen in guinea pigs and in the rabbits, but are less frequent in mice, rats and hamsters (BIRMAN & MARIANO, unpublished results).

Origin, differentiation and fate - The macrophage origin of multinucleated cells in chronic inflammatory lesions is now well established. When bonemarrow macrophage precursors are labelled with tritiated thymidine (3HT) and a chronic lesion is then induced, the isotopic marker can be detected in the nuclei of giant cells which appeared during the development of the granuloma 44, 61, 88. Macrophage fusion is thought to be the basis for the origin of multinucleated cells. Cytoplasmic contact and fusion between macrophages is observed in electronmicrographs of the lesions. Since fusion processes appear to be a rapid event, they are not easily observed under the microscope. Giant cells may also be formed through repeated nuclear division without cytokinesis. However, labelling experiments have shown that synchronous and asynchronous DNA synthesis occurs in only a small number of multinucleated cells. These nuclei may enter the mitotic phase of division, with their chromosomes presenting typical structural aberrations. At the end of the mitotic phase abnormal poliploid nuclei result 61. Once multinucleated cells are formed, newly arrived macrophages may be incorporated into the syncytium, thereby increasing the total number of actual nuclei. Since pretreatment of

experimental animals with colchicine induces the transformation of Langhans cells into foreign-body cell types, probably by disorganizing microtubular structures in the cytoplasm, it was suggested that foreign-body giant cells really mature into the Langhans type. This proposed sequence of events is further supported by the fact that foreign-body giant cells are the first to appear on the surface of subcutaneously implanted coverslips while the typical Langhans cells only appear at a later stage. Ultrastructural observations are also consistent with this interpretation 61 despite some conflicting results 111. Although two basic types of giant cells have been distinguished, the giant cell population as a whole exhibits considerable morphological variation. It has been shown that when glass coverslips are left in the subcutaneous tissue of mice for more than 21 days, peculiar Langhans giant cells appear on the glass surface. When observed by phase contrast microscopy they look globous and supported by a cytoplasmic holdfast, resembling a "crystal ball". These cells share some ultrastructural and functional characteristics with epithelioid macrophages 64. The fate of multinucleated cells in chronic inflammation has not been elucidated to date, although it is known that when they are inserted into the subcutaneous tissue of mice inside millipore diffusion chambers they have a life-span of about 6 days 61.

Giant cell formation - One of the models used for studying giant cells in chronic inflammation is that of coverslip-induced lesions. Using mice and rats as experimental animals it was found that multinucleated cells appear on the glass surface four days after coverslip implantation. The determining factor(s) for cell fusion is not known. At last in this model in which activated lymphocytes do not participate in the turnover and differentiation of the lesion, IL-4 69 and other lymphokines can be ruled out as factors which induce macrophage fusion. One hypothesis is that giant cells might result from fusions of senescent macrophages. To test this hypothesis, 48-hour coverslips were put inside millipore chambers, and these then inserted back into "donors" from which the coverslips had been recovered. Removal of the chamber at different time intervals showed that giant cell formation did not occur. However, when holes were made in the chambers to allow newly arrived macrophages to enter the chamber, then macrophage polykarya were formed. This finding indicates that fusion might have occurred between both, the "old" and the "young" populations of

macrophages 61. The "old" population might consist of epithelioid cells. Nevertheless, subsequent data showed that epithelioid cells only appear on the glass surface after 14 to 21 days of inflammation 64, whereas multinucleation is first observed on the fourth day after coverslip implantation. Because macrophages recovered from 48 hour coverslips have chromosomes with typical aberrations it was suggested that cell fusion occurs when newly arrived macrophages recognize these changed cells 61. CHAMBERS 19.20, however, challenged this hypothesis and presented evidence that giant cell formation results from the attempt of two or more macrophages to phagocytose the same particle, with the consequent fusion of their plasma membranes. Another intriguing problem is why giant cells are formed in some chronic lesions. It was proposed that the turnover of the lesion might be the real factor 61,99. Giant cell formation was almost abolished when BCG was simultaneously inoculated into the subcutaneous pocket where glass coverslips were inserted 9. Taking this experiment further, the same authors compared the effects of inoculating BCG, heat-killed BCG and carrageenan; the previous observation that alive BCG blocks giant cell formation was confirmed, heat-killed BCG was found not to interfere with the process, and carrageen greatly increased both the number and size of multinucleated cells, a set of results which indirectly suggests that the cell turnover of a granuloma may indeed be a determining factor for the formation of macrophage polykarya 8.

Surface receptors, surface antigens and phagocytosis - In granulomas induced by mycobacteria, fungi or even inert particles, the causative agents are frequently seen within the cytoplasm of multinucleated cells. In the past, the capacity of these cells to phagocytose was a matter of debate, since the material found within their cytoplasm could have been ingested by parent macrophages before cell fusion had occurred. In recent years, experimental evidence has been accumulated which supports the view that macrophage polykarya are one of the cellular components of the phagocytic system; giant cells have been shown to phagocytose heat-killed mycobacteria both "in vivo" and "in vitro" 61 and they also express Fc and C3b surface receptors found in macrophages. PAPA-DIMITRIU et al., 80 showed that the phagocytic ability of giant cells was inversely related to the size of the polykaryon i.e., as the number of nuclei increased, the capacity to phagocytose decreased. These observations suggest that the incorporation of macrophages during giant cell formation is accompanied by a progressive loss of surface receptors involved in phagocytosis. Indeed, it was found that, the great majority of the giant cell population that was recovered from coverslips subcutaneously implanted in mice for more than 21 days, did not express surface receptors for the Fc region of heterologous IgG or for the C3b component of the complement system. The loss of these receptors, together with evidence obtained from ultrastructural studies, led to the suggestion that macrophage polykarya are transformed into "epithelioid polykarya", a phenomenon analogous to the epithelioid transformation of mononuclear phagocytes 64. However, "in vitro" it was found that when levamisole is added to the culture medium, the "paralyzed" multinucleated cells do reexpress Fc receptors 60. Although the above data show that inflammatory macrophage polykarya are phagocytic cells, the primordial function of these cells remains to be elucidated.

Lymphocytes

It is now unquestionable that lymphocytes play a central role in a large variety of inflammatory conditions by mediating local or systemic immune responses. There is also evidence to show that these cells may modulate non-immune inflammatory lesions 42. Ultrastructural observations, and studies using immunocytochemical methods, have shown that lymphocytes and plasma cells are found in different types of granulomas 110. The presence of Migration Inhibition Factor (MIF) and eosinophil chemotactic lymphokines in liver granulomas isolated from schistosome-infected mice indirectly suggests the existence of T-cells in these lesions 11. Using immunofluorescent methods it was shown that the T and B lymphocytes comprise less than O.1% of cells in coverslip-induced granulomas in mice. Similar results were observed when coverslips and PPD were inserted into animals previously sensitized with CFA (MARIANO & RIBEIRO DOS SANTOS, unpublished results). These observations reinforce the concept that macrophages are the prevalent mononuclear cell population which migrate into chronic lesions, and that a small number of sensitized lymphocytes is enough to change the turnover and functional activities of macrophages in inflammed tissues 67.

Granulocytes

Neutrophils come into play during the early stages of a forming tubercle or tuberculous granuloma ⁴. Large

numbers of these cells are also present in early lesions evoked by BCG in different animals species (BIRMAN & MARIANO, unpublished results). With the development of the granuloma, neutrophils tend to disappear. Despite the quantitative reduction, neutrophils play a significant role in liquefying tuberculous granulomas, leading to the formation of cavities and dissemination of tuberculous bacilli 41. These cells are also present in P. brasiliensis granulomas. The role of neutrophils in granuloma formation has not been fully established. Some results show that their migration to the site of inflammation by chemotactic attraction is not dependent on the complement system 15 but on factor(s) released by macrophages 14. Eosinophils are moderately phagocytic cells 53 which may have an important influence on the modulation of histamine release by mast cells and basophils 51. These cells have been traditionally linked with allergic reactions and parasitic infection. Eosinophilia and accumulation of lesion-associated eosinophils in hepatic schistosoma granulomas seem to be mediated by T-cell derived lymphokines 50. Eosinophils are present in different types of granulomatous inflammation, and are a constant feature of the early stages of experimental hepatic granulomas in schistosoma-infected mice 92. The migration of these cells to such lesions is a T cell-dependent phenomenon, but their role is still obscure. The idea that eosinophils might have some parasiticidal action comes from "in vitro" experiments, which show that they exhibit an antibody-dependent cytotoxic effect on schistosome eggs 52. Despite the occurrence of large numbers of basophils in Crohn's disease granulomas, little is known about the relevance of these cells in other types of hypersensitivity granulomatous lesions 12.

Vascularization of granulomas

The limits of a granulomatous lesion are not always sharply defined. Granulomas are surrounded by a capsule of proliferating connective tissue composed of fibroblasts, collagen fibers, amorphous substances and newly formed vessels. Connective tissue proliferation and the formation of new vessels are important events in granuloma formation. The connective tissue is the source of cells recruited to the lesion, and the newly formed vessels provide oxygen and nutrient supply to cells present in the granuloma. Spontaneous tuberculous granulomata in man are avascular, i.e. no vascularization occurs within the mass of macrophages and epithelioid cells, which make up the central part of the lesion 4. This observation suggests that either cells

within the granuloma (epithelioid cells) inhibit neovascularization or stimulatory substances to endothelial cell multiplication are not secreted, for instance, by macrophages. 104. However, little is known about vascularization of other types of granulomas. COURTADE et al., 25 studied the vascularization of BCG induced granulomas in rabbits. They showed a 60% increase in capillary density in non necrotic lesions. This increase was constant in necrotic lesions and in lesions induced by tuberculin. Little information is available concerning the possible influence of the connective tissue on granuloma formation, MELRO & MARIANO 70 described a typical granulomatous response around Schistosoma mansoni eggs, which were "freely floating" in the peritoneal cavity of experimentally infected mice. These observations suggest that the connective tissue does not play a fundamental role in granuloma formation and organization although an extracellular matrix, not yet fully characterized, was observed in these "free floating" lesions. A similar phenomenon as that described by MELRO and MARIANO 70 is observed in lower animals. When crustaceans, for instance, are invaded by living or inert foreign bodies, the hemocytes react against them by means of phagocytosis and multicellular reaction resulting in "encapsulation" or "nodule formation" 85. These lesions are structurally similar to granulomas observed in mammals suggesting that this type of cellular response to foreign bodies is a very old phenomenon in phylogenesis.

Granuloma formation

Foreign-body versus Immune Granuloma

Little is known about the mechanisms which determine macrophages to organize into a focal and compact lesion to form a granuloma and not to diffuse in chronic inflammatory process. One possible explanation is that granulomatogenic agents behave as chemoattractants or induce the generation of chemotatic factors from tissues, thereby centripedally directing the cells to surround and to envelop the irritant. If the causative agent is particulate and larger than the phagocytes as is the case for bentonite, plastic beads or schistosome eggs this behaviour of the cells seems obvious. However, this does not appear to be the case in other circumstances. It has been long known that when virulent tuberculous bacilli are inoculated into the subcutaneous tissue of a non-sensitized guinea pig, the bacteria is detected in regional lymph nodes within a short interval of time 16. At the point of inoculation a mild acute inflammatory reaction will develop followed by migration of polymorphonuclear leukocytes and monocytes which phagocytose the inoculated bacteria. The neutrophilic migration subsides and macrophages which phagocytosed the bacilli start to aggregate, resulting in a focal lesion. In about one week a typical granulomatous lesion develops. The mycobacteria are then limited to the necrotic area of the granuloma and are present in small numbers within epithelioid cells. They are not detected at the halo of newly migrated cells which delimitates the edge of the lesion. Inert particles (silica, glass, talc, cotton) if injected into the connective tissue of experimental animals will also evoke a transient acute inflammatory response that is followed by an intense macrophage accumulation and granuloma formation. Macrophages, epithelioid cells and giant cells are found in these foreign-body granulomas [reviewed in 12]. In a series of elegant experiments WARREN 113 showed the influence of the immune system, on the evolution of schistosome egg granulomas in mice. Based on these data he classified granulomas in immunological and non-immunological granulomas. This classification suggests that the formation of the immunological granulomata is entirely dependent upon sensitized and activated T cells. In support of this hypothesis BOROS & WARREN 11 showed that the injection of bentonite particles covered with different antigens into the circulation of sensitized animals induces, in the lungs, a mononuclear cell collection around the particles only when the animals were previously sensitized to the same antigen. More recently it has been shown that "granuloma" develops "in vitro" when sensitized T cells are put in mixed cultures of macrophages and schistosome eggs 6, 33, 34, 57. These experiments suggest that antigen activated T lymphocytes secret lymphokines which attract macrophages and hold these cells together resulting in the granuloma formation. This mechanism of "immune-granuloma" formation can be questioned. In the experiments of BOROS & WAR-REN 11 and in those of DOUGHTY & PHILLIPS 33.34 the morphology of the lung lesions or of the cell collection around the eggs "in vitro" can not be classified as granulomatous lesions. The presence of epithelioid cells in these experimental situations has not been demonstrated, and a simple focal mononuclear cell collection should not be regarded as a granuloma. The same argument applies for focal hepatic lesions induced in mice by L. donovani 68. Even considering the limitations of these models a series of experiments

demonstrate that interleukin-1 but not interleukin-2 plays a role in the initiation and development of pulmonary granulomas in mice 54 . Evidences that interleukin-1 and TNF- α are envolved with the phenomenon of macrophage aggregation around inert particles in vitro has been recently demonstrated 90 . The participation of TNF- α in the genesis of BCG-induced granuloma in mice has been clearly demonstrated by KINDLER et al. 55 .

Another set of experiments also rules out the participation of T-cells in granuloma formation. Nude mice develop granulomatous lesions when injected with BCG 3 and when inoculated with schistosome these mice are able to mount a granulomatous response around the eggs 40. CHEHL et al. 21 have also shown that the inoculation of M. leprae in nude mice induces typical granulomatous responses in these animals. Interestingly, about two months after inoculation of the bacteria, the lesions are no more granulomatous but resemble those observed in lepromatous patients. Another evidence that granuloma formation does not depend on activated T cells comes from the work of MELRO & MARIANO 70 who showed a typical granulomatous arrangement of mononuclear cells around viable schistosome eggs inoculated into the peritoneal cavity of noninfected mice. The "free floating" granulomas were observed 48 hrs after the inoculation of the eggs. Finally, the inoculation of BCG in the cheek pouch of hamsters which lacks lymphatic drainage, induces typical granulomas despite the fact that the animals do not react to the intradermal injection of PPD 91. More recently, NORTH et al. 75 demonstrated that typical granulomatous response is observed in the liver of SCID mice inoculated with BCG. These experiments clearly show that neither T nor B lymphocytes play any role in the genesis of granulomas. These observations, added to the fact that a foreign body, such as a glass coverslip can induce epithelioid transformation of macrophages 64 and giant cell formation 22.87, the two basic cellular components of the granulomatous response, strongly suggest that granuloma formation is a macrophage phenomenon influenced by but not dependent on lymphocytes. However, the chemical characteristics of the granulomatogenic "irritant" which induces macrophage aggregation and transformation into epithelioid cells, remains to be elucidated.

Immune modulation of the granuloma

While granuloma formation largely depends on

the nature of the "irritant" which determines macrophage aggregation and modulation into giant cells and epithelioid cells, there is no doubt that the immune responses do modulate the expansion and the morphology of lesions. The pioneer work of WARREN 113 on the influence of the immune system on the schistosome egg granuloma indicate that the volume of lung granulomas, induced by intravenous injections of schistosome eggs, is increased when sensitized lymphoid cells are transferred to non sensitized recipients. Later, BOROS and colleagues [reviewed in 12] reported that transfer of lymphoid cells from animals in the late stages of infection, reduces the volume of schistosome egg granulomas. The phenomenon has been analyzed in more detail to show that the inflammatory response to schistosome eggs involves a dynamic interaction of lymphokine-secreting Lyt 1+ effector and Lyt 2+ suppressor T cells. Some evidences suggest that the suppressor T cells are recruited in a feedback manner by Lyt 1-, Oa+ helper cells and exerts their effects through soluble suppressor factors which can regulate lymphokine production ^{22,24}. PERAÇOLI et al. 81 observed a clear correlation between the humoral or cellular response to Paracoccidioides brasiliensis in hamsters and granuloma morphology. When cell mediated immunity, as evaluated by lymphoid blast transformation "in vitro" is high, a typical granulomatous response to the fungi is observed, i.e., the granulomas are dense and well organized. Conversely, when cell mediated immunity decreases, with the concomitant increase of specific antibody titres, the granulomas are loose and not well defined. This model closely ressembles the morphological and immunological characteristics of leprosy in man as far as cell mediated immunity, humoral response and granuloma formation are concerned 35, de BRITO et al. 27,28,29 have brought evidences that antibody-mediated immune response might participate in the pathogenesis of schistosomal granulomas by permeating the mantel of epithelioid cells and neutralizing small doses of egg antigens shaded by living eggs. For these authors, this might prevent large amounts of immune complexes formation and consequent increase in tissue destruction.

The special case of leprosy granuloma

Since the discovery of the etiologic agent of lepra in man by HANSEN ⁴⁸ many attempts to reproduce the disease in experimental animals have failed. Some progress has been achieved after the work of SHEPARD ⁸⁹ who succeeded in growing the bacilli in

the foot pad of Balb/c mice. In addition M. leprae was found to grow and disseminates in athymic nude mice much more vigorously than it does in the heterozygote counterpart 21. In both experimental models, the bacilli induce a granulomatous response which, however, does not keep a close morphological correlation to those observed in tuberculoid leprosy in man. The typical peripheral nerve damage observed in tuberculoid leprosy in man is not observed in these models. In contrast to what is observed after the inoculation of other mycobacteria in the subcutaneous tissue of experimental animals, the injection of dead M. leprae does not induce a significant inflammatory response. When live bacilli are inoculated even in Balb/c or Nu/Nu mice, about two months have to elapse before the bacilli start multiplication, followed by an intense inflammatory response. Although the mechanisms involved in the initiation of the inflammatory response to live $M_{\underline{\cdot}}$ leprae are not known, it appears to be a T-independent phenomenon since it does occur in athymic mice 21. Experiments from this laboratory demonstrate that dead M. leprae and particularly its lipidic components blocks phagocytosis by macrophages "in vivo" suggesting that the bacteria induces immune-repulsion in the host 73. We have also shown that the inoculation of dead M. leprae from human origin (Lepromin) in the foot pad of hamsters previously immunized with BCG, induces, after 16 days of inoculation, a typical granulomatous response which surrounds myelinated peripheral nerves 59. Although nerve damage could not yet be demonstrated, this model seems useful for the study of the basic mechanisms by which granuloma formation is induced by M. leprae antigens.

Concomitant immunity and a hypothesis for granuloma persistence

The classical Koch's phenomenon is a biological paradox not yet solved. If a guinea pig is inoculated with live *M. tuberculosis* the animals will die in three to four weeks. Nevertheless, if an inoculated animal is subcutaneously reinoculated with the same microorganism, a flourishing inflammatory response will appear at the site of inoculation which may become necrotic in a few days and heals in about two weeks. No bacteria are found in the regional lymph nodes. Therefore, the animal is competent to kill the microorganism after the second but not after the first inoculation. The basic difference between both circumstances is that following the first inoculum multiple granulomas develop whereas the second inoculation induces a chronic

inflammatory reaction with no granuloma formation. This phenomenon, named concomitant immunity, occurs not only in infectious diseases 82 but also in some types of transplantable tumors 43. While the second inoculation could be interpreted a classical T cell mediated delayed type of hypersensitivity, this latter phenomenon is not always effective in eliminating the bacilli within the granuloma. Helper and suppressor T cells have been shown to populate the mantel surrounding infectious granulomas but not the centre of the lesion and it has been suggested that these cells might modulate macrophage functions and turnover 11, 100, 101, The reason why newly arrived macrophages and lymphocytes do not penetrate into the mass of epithelioid cells which have taken up the bacilli in their cytoplasm is not known. A plausible explanation for the failure of immune effector mechanisms to eliminate the bacilli within a granuloma is that the cells inside the lesion might deactivate lymphoid cells and/or newly arrived macrophages. Evidence for this hypothesis has recently emerged 17, 18. Macrophages on the surface of glass coverslips subcutaneously implanted in mice for 7 days, spontaneously release superoxide anion in appreciable amounts. If the coverslips are removed after 14 and 21 days of implantation, the cells on their surface no longer release detectable amounts of this radical, even following stimulation with phorbol mirystate acetate. When a delayed type of hypersensitivity reaction was induced around the 14 days old coverslips, to recruit macrophages, the histological analysis of the coverslips showed the presence of newly arrived cells but no O, release has been detected. When 5 days old lesions were tested, the induction of a delayed type of hypersensitivity reaction, around the coverlips, induced a marked increase in superoxide liberation by the cells. These results suggest that the "old" cells in the lesion might be responsible for deactivating the newly arrived cells. To analyze this hypothesis, 5 days old coverslips were co-cultured with 14 day old coverslips and the release of O, was measured. Under these circumstances, superoxide anion release by the cells on coverslips removed after 5 days was completely inhibited suggesting that the "old" cells secrete a factor or factors which deactivate inflammatory macrophages despite the activation of these cells by lymphokines secreted by activated T lymphocytes. This factor was named "Macrophage Deactivating Factor, MDF". MDF was partially characterized as a low molecular weight protein (12 kDa), termo resistant and sensitive to tripsin and pronase 17. We have recently produced a policional antiserum directed to antigens secreted by murine epithelioid macrophages 63. These antibodies immunostained epithelioid cells from coverslips implanted for more than 14 days in the subcutaneous tissue of mice as well as epithelioid cells from BCGinduced granulomas in the same species. They also blocked the MDF deactivating activity "in vitro" and decreased the volume of BCG-induced granulomas in the foot pad of mice when injected intraperitonealy in these animals. Following the observations of DELABIE et al. 31 who showed that epithelioid cells from tuberculous but not from foreign-body granulomata in man selectively express MRP-14 but not MRP-8 calcium-binding proteins 76 in their cytoplasm, we investigated by immunocytochemical techniques, the expression of these proteins in cells of the coverslip model. It was observed that cells on coverslips removed after 3, 5 and 7 days of implantation do not express these antigens. Nevertheless, cells previously characterized as epithelioid cells 64 on the surface of coverslips removed after 14 days of coverslip implantation are MRP-14⁻/8⁻ cells. We have also observed that epithelioid cells from typical (thread suture) granulomas also express MRP-14 but not MRP-8 antigens. These data show, contrary to the observations of DELABIE et al. 31, that epithelioid cells from foreign-body granulomata in man and mice express in their cytoplasm MRP-14 but not MRP-8 calcium-binding proteins. Taking these investigations further, we showed that MRP-14 calcium binding protein can be detected in the supernatant of epithelioid cell cultures and, that monoclonal antibodies anti-MRP-14 protein, blocks the MDF activity of this supernatant. Finally, highly purified MRP-14 but not MRP-8 blocks superoxide anion release by activated macrophages "in vitro". These results demonstrate a biochemical identity between MDF and the MRP-14 calcium-binding protein (MARIANO & SORG, in preparation). The mechanism of macrophage deactivation by MRP-14 is not yet determined. Nevertheless, activation of macrophages by lipid A immediately (30 sec.) causes a rapid and extensive intracellular fluxes of Ca i.e., the ion concentration rises from basal levels of 55 nM to \sim 600 nM 83. Yet, inhibitors of calcium fluxes and protein synthesis respectively block macrophage activation by INF γ and LPS ^{47, 93}. Based on these observations, it is licit to speculate that MRP-14, a calcium binding protein of unknown function, in some way might down-regulate Ca**

fluxes in activated macrophages thus deactivating these cells.

These results led us to hypothesize ⁶² that epithelioid cells secret the MRP-14 calcium-binding protein (MDF) which by deactivating the newly arrived cells impair antigen elimination with consequent persistence of the lesion.(Fig. 2)

If this hypothesis is true, it could be a plausible explanation for the persistence of tuberculous granulomas for instance, despite the existence of an effective and sterilizing peripheral immunity (Koch's phenomenon).

FINAL COMMENTS

Studies on the granulomatous inflammation with a variety of methods have provided a large amount of data which helped to elucidate intricate aspects of the phenomenon: first the granuloma is mainly composed of cells of the Mononuclear Phagocytic System, the epithelioid cells being the landmark of the process; second the granulomatous reaction is primarily a response of macrophages that is induced by certain types of "irritants"; third the lesion is maintained by a con-

stant recruitment of cells from the circulation, mainly monocytes, which can be transformed into activated macrophages or epithelioid cells and which may occasionally fuse to form giant cells. Finally, lymphocyte responses may influence the lesion either to expand or to shrink. Nevertheless, many aspects of the granulomatous response remain to be elucidated. One of the most important questions to be solved concerns the characteristics of the granulomatogenic agents. Some but not all pathogens are kept alive within macrophages and induces these cells to aggregate which results in the typical morphology of the lesion. The epithelioid transformation of macrophages should be studied more in detail since this type of macrophage modulation is a unique manifestation of the process. A major limitation of these studies is the fact that the epithelioid modulation of macrophages has never been observed in vitro. The understanding of this phenomenon would certainly help to elucidate the role these cells play in granuloma formation, modulation and fate. Finally, once the interactions between the immune system and granuloma maintenance are better understood, prophylactic and chemotherapeutic agents shall be more efficiently developed to control a large spectrum of granulomatous diseases of man and animals.

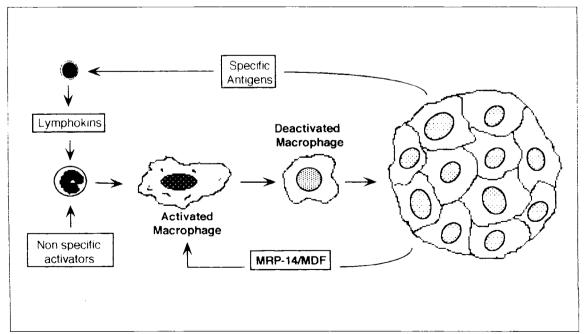


Fig. 2 - Schematic and hypothetical representation of the role played by MRP-14/MDF monokine secreted by epithelioid cells, on macrophage deactivation and consequent persistence of granulomatous lesions.

RESUMO

O granuloma experimental. Uma hipótese para explicar a persistência da lesão.

A inflamação glanulomatosa é o substrato morfológico de uma variedade de doenças infecciosas importantes tais como tuberculose, lepra, esquistossomose e outras. Embora muitos aspectos deste tipo especial de inflamação sejam conhecidos, questões fundamentais referentes à formação do granulona, persistência, destino e significado da relação hospedeiro-parasita permanecem para serem elucidados. Nesta revisão é apresentada a literatura básica e mais relevante relacionada à investigação experimental sobre a fisiopatologia do granuloma. Baseados em investigações recentes realizadas, em nosso laboratório mostrando que o fator de desativação do macrófago (MDF) secretado pelas células epitelióides e caracterizado pela proteina MRP-14, uma proteina ligante de cálcio, foi levantada hipótese para explicar a persistência da inflamação granulomatosa.

ACKNOWLEDGMENTS

The constructive criticisms of this review by Werner Haas, Cord Sunderkötter, Clemens Sorg and Sonia Jancar are gratefully acknowledged.

REFERENCES

- ADAMS, D. O. The granulomatous inflammatory response. Amer. J. Path., 84: 164-191, 1976.
- ADAMS, D. O. The structure of mononuclear phagocytes differentiation "in vivo".
 Sequencial fine and histologic studies of the effect of Bacillus Calmette-Guerin (BCG). Amer. J. Path., 76: 17-48, 1974.
- ALLRED, C.; KOBAYASHI, K. & YOSHIDA, T. Anergy-like immunosuppression in mice bearing pulmonary foreign-body granulomatous inflammation. Amer. J. Path. 121: 466-473, 1989.
- ASCHOFF, L. Tratado de anatomia patologica. Barcelona, Ed. Labor, 1950. p. 546-555.
- AUGER, M. J. & ROSS, J. A. The biology of macrophage. In: LEWIS, C. E. & O'D.McGEE, J., ed -. The natural immune system: the macrophage. Oxford, Oxford University Press, 1992. p. 1-57.
- BENTLEY, A. G; DOUGHTY, B. L. & PHILIPS, S. M. -Ultraestructral analysis of the cellular response to *Schistosoma mansoni*. III. The "in vitro" granuloma. Amer. J. trop. Med. Hyg., 3: 1168-1180, 1982.
- BIKIERKUNST, A.; LEVY, I. S.; YARKONI, E, ADAM, A. & LEDERER, E. - Granuloma formation induction in mice by chemically defined mycobacterial fractions. J. Bact., 100: 95-102, 1969.

- BIRMAN, E. G. & MARIANO, M. The influence of inflammatory agents on giant cell formation. Braz. J. med. biol. Res., 18:507-512, 1985.
- BIRMAN, E. G. & MARIANO, M. The modification of macrophage activity in experimental granulomas by cyclophosphamide. J. Path., 134: 133-146, 1981.
- BONNEY, R. J. & DAVIES, P. Possible autoregulatory functions of the secretory products of mononuclear phagocytes. In: ADAMS, D.O. & HANNA Jr., M.G. - Macrophage activation. Contemp. Top. Immunobiol., 13: 199-218, 1984.
- BOROS, D. L. & WARREN, K. S. The bentonite granuloma: characterization of a model system for infectious and foreign body granulomatous inflammation using soluble mycobacterial, histoplasma and schistosoma antigens. Immunology, 24: 511-529, 1973
- BOROS, D. L. -. Granulomatous inflammation. Progr. Allergy, 24: 183-267, 1978.
- BROWETT, P. J.; SIMPSPON, L. O. & BLENNERHASSETT, B. -Experimental granulomatous inflammation: the ultraestructure of the granuloma induced by injection of tubercle bacilli into Freund adjuvant-sensitized guinea-pigs. J. Path. 129: 191-201, 1979.
- CALICH, V. L. G.; COPPI VAZ, C. A. & BURGER, E. PMN chemotactic factor produced by glass-adherent cells in the acute inflammation caused by *Paracoccidioides brasiliensis*. Brit. J. exp. Path., 66: 57-65, 1985.
- CALICH, V. L. G.; KIPNIS, T. L.; MARIANO, M.; FAVA NETO, C.
 DIAS DA SILVA, W. The activation of the complement system by *Paracoccidioides brasiliensis* "in vitro": its opsonic effect and possible significance for an "in vivo" model of infection. Clin. Immunol. Immunopath., 12: 20-30, 1979.
- CALMETTE, A. L' infection bacillaire et la Tuberculose chez l'homme et chez les animaux. 4. ed. (Boquet, L.) Negre Paris, Masson, 1936, p.422-740.
- CAMARERO, V. C. P. C.; COLEPICOLO, P.; RIBEIRO, J.M.C. & KARNOVSKY, M. L. - Leukocyte-deactivating factor from macrophages: partial purification and biochemical characterization. A novel cytokine. J. cell. Physiol., 157: 84-89, 1993.
- CAMARERO, V. C. P. C; JUNQUEIRA, V.; COLEPICOLO, P.; KARNOVSKY, M. L. & MARIANO, M. - Epithelioid macrophages secrete a deactivating factor for superoxide release. J. cell. Physiol., 145: 481-487, 1990.
- CHAMBERS, T.J. Failure of altered macrophage surface to lead to formation of polykaryons. J. Path., 122: 185-189, 1977.
- CHAMBERS, T.J. Multinucleate giant cells. J. Path., 126: 125-148, 1978.
- CHEHL, S.; RUBY, J.; JOB, C. K. & HASTINGS, R. C. The growth of *Mycohacterium leprae* in nude mice. Lepr. Rev., 54: 283-304, 1983
- CHENSUE, S. W. & BOROS, D. L. Modulation of granulomatous hypersensitivity. I. Characterization of T-lymphocytes involved in the adoptive supression in *Schistosoma mansoni*-infected mice. J. Immunol., 123: 1409-1414, 1979.

- CHENSUE, S. W.; BOROS, D. L. & DAVID, C. S. Regulation of granulomatous inflammation in murine schistosomiasis. "In vitro" characterization of T lymphocyte subsets involved in the production and suppression of migration inhibitor factor. J. exp. Med., 151: 1398-1412, 1980.
- CHENSUE, S. W.; WELLHAUSEN, S. R. & BOROS, D. L. -Modulation of granulomatous hypersensitivity. II. Participation of Ly1+ and Ly2+t lymphocytes in the suppression of granuloma formation and lymphokine production in *Schistosoma mansoni* infected mice. J. Immunol., 127: 363-367, 1981.
- COURTADE, E. T.; TSUDA, T.; THOMAS, C. R. & DANNEMBERG Jr., A. M. . Capillary density in developing and healing tuberculous lesions produced by BCG in rabbits: a quantitative study. Amer. J. Path., 78: 243-260, 1975.
- DANNEMBERG Jr., A. M.; ANDO, M.; SHIMA, K. & TSUDA, T.

 Macrophage turnover and activation in tuberculous granulomata. In:
 FURTH, R. Van, ed. Mononuclear phagocytes in immunity infection and pathology. Oxford, Blackwell, 1975. p. 959-967
- DE BRITO, T. & FRANCO, M.F. Granulomatous inflammation. Rev. Inst. Med. trop. S. Paulo, 36: 185-192, 1994.
- DE BRITO, T.; HOSHINO-SHIMIZU, S. & SILVA, L.C. -Immunopathology of experimental schistosomiasis (S. mansoni) egg granulomas in mice: possible defense mechanisms mediated by local immune complexes. J. Path., 140: 1-28, 1983.
- DE BRITO, T.; HOSHINO-SHIMIZU, S.; SILVA, L.C. et al. -Chronic human schistosomiasis mansoni: schistosomal antigens, immunoglobulins and complement C3 detection in the liver. Liver, 5: 64-70, 1985.
- DEFAVERI, J.; REZKALLAH, M. T. & FRANCO, M. F. -. Experimental paracoccidiodomycosis in mice: morphology and correlation of lesions with humoral and immune response. Mycopathologia (Den Haug), 77: 3-11, 1982.
- DELABIE, J.; DE WOLF-PEETERS, C.; VAN DEN OORD, J. J. & DESMET, V. J. - Differential expression of the calcium-binding proteins MRP-8 and MRP-14 in granulomatous conditions: an immunohistochemical study. Clin. exp. Immunol., 81: 123-126, 1990.
- DELOVITCH, T. L.; PHILLIPS, L. M.; NAQUET, P. et al. Antigen internalization, processing, and recycling by antigen-presenting cells. Progr. Immunol., 6: 238-253, 1986.
- DOUGHTY, B. L. & PHILLIPS, S. M. Delayed hypersensitivity granuloma formation around *Schistosoma mansoni* eggs "in vitro". 1 Definition of the model. J. Immunol., 128:30-36, 1982.
- DOUGHTY, B. L. & PHILLIPS, S. M. Delayed hypersensitivity granuloma formation and modulation around *Schistosoma mansoni* eggs "in vitro". II. Regulatory T cell subsets. J. Immunol., 128: 37-42, 1982.
- DUNCAN, E. S. & STEWARD-TULL. Mycobacterium leprae. The bacteriologist's enigma. In: RATLEDGE, C., ed. - The biology of the mycobacteria. Stanford, Academic Press, 1982. p. 273-298.
- DUNGWORTH, D. L. The respiratory system. In: JUBB, K.V.F.; KENNEDY, P.C. & PALMER, N., ed. - Pathology of domestic animals. London, Academic Press, 1985. v.2, p 413-556.

- DUNSFORD, H. A.; LUCIA, H. L.; DOUGHTY, B. L. & LICH-TENBERG, F. VON - Artificial granulomas from bentonite and latex carrier particles. Amer. J. trop. Med. Hyg., 23: 203-217, 1974.
- EPSTEIN W.L.; SKAHEN, J.R. & KLEBANOFF., S. J. -.
 Granulomatous hypersensitivity to zirconium: localization of allergen
 in tissue and its role in formation of epithelioid cells. J. invest.
 Derm., 38: 223-232, 1962.
- EPSTEIN, W. L. Granulomatous hypersensitivity. Progr. Allergy, 11: 36-88, 1967.
- EPSTEIN, W. L.; OKAMOTO, M.; SUYA, H. & FUKUYAMA, K.
 T-cell independent transfer of organized granuloma formation.
 Immunol. Lett., 14: 59-63, 1987.
- GALINDO, B. & MYRVIK, Q. N. Migratory response of granulomatous alveolar cells from sensitized rabbits. J. Immunol., 105: 227-237, 1970.
- GARCIA-LEME, J. Role of lymphocytes in non-immune experimental inflammation. Agents and Actions, 11: 597-602, 1981.
- GERSHON, R. K.; CARTER, R. L. & KONDO, K. On concomitant immunity in tumor bearing hamsters. Nature (Lond.), 213: 674-676, 1967.
- GILLMAN, T. & WRIGHT, L. J. Probable "in vivo" origin of multinucleated giant cells from circulating mononuclears. Nature (Lond.), 209: 263-265, 1966.
- GORDON, S.; LAWSON, L.; RABINOWITZ, P. R. et al. Antigen markers of macrophage differentiation in murine tissue. Curr. Top. Microbiol. Immunol., 181:1-37, 1992.
- GRECCHI, R.; SALIBA, A. M. & MARIANO, M. Morphological changes, surface receptors and phagocytic potencial of fowl mononuclear phagocytes and thrombocytes "in vivo" and "in vitro". J. Path., 130: 23-31, 1980.
- HAMILTON, T. A.; RIGSBEE, J. A.; SCOTT, W. A. & ADAMS, D.
 O. Gamma interferon enhances the secretion of arachidonic acid metabolites from murine peritoneal macrophages stimulated with phorbol derivatives. J. Immunol., 134: 2631-2637, 1985.
- HANSEN, G. H. A. On the ethiology of leprosy. Chirurg. Rev., 55: 459-489, 1875.
- HESSIAN, P. A.; EDGEWORTH, J. & HOGG, N. MRP-8 and MRP-14, two abundant Ca2'-binding proteins of neutrophils and monocytes. J. Leuk. Biol., 53: 197-204, 1993.
- HIRASHIMA, M.; TASHINO, K.; SAKATA, K. & HIRSHIMA, M.

 Isolation of an eosinophil chemotactic lymphokine as a natural mediator for eosinophil chemotaxis from concanavalin A-induced skin reaction sites in guinea pigs. Clin. exp. Immunol., 57: 211-217, 1984.
- HUBSCHER, T. & ELSEN, A. M. An eosinophil derived inhibitor (EDI) of histamine release. J. Allergy clin. Immunol., 51:671-676, 1975.
- JAMES, S. L. & COLLEY, D. G. Eosinophil-mediated destruction of *Schistosoma mansoni* eggs in vitro. II. Role of cytophilic antibody. Cell Immunol., 38: 35-47, 1978.
- 53. JONG, E. C.; HENDERSON, W. J. & KLEBANOFF, S.J. Bac-

- tericidal activity of eosinophil peroxidase. J. Immunol., 124: 1378-1382, 1980.
- 54. KASAHARA, K.; KOBAYASHI, K.; SHIKAMA, Y. et al. Direct evidence for granuloma-inducing activity of interleukin-1. Induction of experimental pulmonary granuloma formation in mice by interleukin-1 coupled beads. Amer. J. Path., 130: 629-638, 1988.
- KINDLER, V.; SAPPINO, A. P.; GRAU, G. E.; PIGUET, P. F. & VASSALLI, P. - The inducing role of tumor necrosis factor in the development of bacterial granulomas during BCG infection. Cell, 56: 731-740, 1989.
- KLIGMAN, D. & HILT, D. C. The S-100 protein famili. Trends Biochem. Sci., 13: 437-443, 1988.
- LAMMIE, P. J.; LINETTE, G. P. & PHILLIPS, M. Charac-terization of Schistosoma mansoni antigen-reactive T cell clones that form granulomas in vitro. J. Immunol., 134: 4170-4175, 1985.
- LASSER, A. The mononuclear phagocytic system: a review. Hum. Path., 14: 108-126, 1983.
- MARIANO, M. & FERREIRA, S. A. BCG infection induces responsiveness to dead Mycohacterium leprae in hamsters. A Mitsuda like reaction. Braz. J. med. biol. Res., 22: 81, 1989.
- MARIANO, M. & MALUCELLI, B. E. Defective phagocytic ability of epithelioid cells reversed by levamisole. J. Path., 130: 33-36, 1980.
- MARIANO, M. & SPECTOR, W. G. The formation and properties of macrophage polykarions (Inflammatory giant cells). J. Path., 113: 1-19, 1974.
- MARIANO, M. Does macrophage deactivating factor play a role in the maintenance and fate of infectious granulomata? Mem. Inst. Oswaldo Cruz, 86; 485, 1991.
- MARIANO, M.; AGUIAR, T.; CARNEIRO, C. R. W. et al. -Antibodies directed to antigens secreted by murine epithelioid macrophages modulate BCG-induced granulomata. Mediators Inflammation, 2: 229-233, 1993.
- 64. MARIANO, M.; NIKITIN, T. & MALUCELLI, B. E. -. Immunological and non-immunological phagocytosis by inflammatory macrophages, epithelioid cells and macrophage polykarions from foreign body granulomata. J. Path., 120: 151-159, 1976.
- MARIANO, M.; NIKITIN, T. & MALUCELLI, B. E. Phagocytic potencial of macrophages from within delayed hypersensitivitymediated granulomata. J. Path., 3: 27-33, 1977.
- MATHEW, R. C.; KATAYAMA, I.; GUPTA, S. K.; CURTIS, J. & TURK, J. L. - Analysis of cells of mononuclear phagocyte series in experimental mycobacterial granulomas by monoclonal antibodies. Infect. Immun., 39: 344-352, 1993.
- McCLUSKEY, R. T.; BENACERRAF, B. & McCLUSKEY, J. W. -Studies on the specificity of the cellular infiltrate in delayed-type hypersensitivity reactions. J. Immunol., 90: 466-472, 1963.
- McELRATH, M. J.: MURRAY, H. W. & COHN, Z. A. The dynamics of granuloma formation in experimental visceral leishmaniasis. J. exp. Med., 164: 1927-1937, 1988.
- 69. McINNES, A. & RENNICK, D. M. Interleukin 4 induces cultured

- monocytes/macrophages to form giant multinucleated cells. J. exp. Med., 167: 598, 1988.
- MELRO, M. C. F. & MARIANO, M. -. Extra-tissular S. mansoni egg granuloma in the peritoneal cavity of mice. Mem. Inst. Oswaldo Cruz, 82 (suppl. 4): 245, 1987.
- MONIS, B.; WEINEBERG, T. & SPECTOR, G. J. The carrageenan granuloma in the rat. A model for the study of the structure and function of macrophages. Brit. J. exp. Path., 49: 302-310, 1968.
- MONTENEGRO, M. R. & FRANCO, M. Pathology. In: FRANCO, M.; LACAZ, C. da S.; RESTREPO-MORENO, A & DEL NEGRO, G., ed - *Paracoccidioidomicosis*. Boca Raton, CCR Press, 1993. p. 131-147.
- MOURA, A. C. N. & MARIANO, M. Dead Mycobaterium leprae inhibits phagocytosis by inflammatory macrophages in vivo. Participation of the bacterial cell lipids in the phenomenon. Mem. Inst. Oswaldo Cruz, 85: 381-382, 1990.
- NATHAN, C. F. Secretory products of macrophages. J. clin. invest., 79: 319-326, 1987.
- NORTH, R. J. & IZZO, A. A. Granuloma formation in severe combined immunodeficient (SCID) mice in response to progressive BCG infection. Tendency not to form granulomas in the lung is associated with faster bacterial growth in this organ. Amer. J. Path., 142: 1959-1966, 1993.
- ODINK, K.; CERLETTI, N.; BRÜGGEN, J. et al. Two calciumbinding proteins in infiltrate macrophages of rheumatoid arthritis. Nature, (Lond.) 330: 80-82, 1987.
- OTTOWICZ, J.; ZAJUSZ, K.; NOWAK, B. & PARADOWSKI, Z. The influence of immunologic factors on the development of experimental silicosis in guinea pigs. Arch. Immunol. Ther. exp., 15: 191-203, 1967.
- PAGE, R. C.; DAVIES, P. & ALLISON, A. C. Pathogenesis of the chronic inflammatory lesions induced by Group A streptococcal cell wall. Lab. Invest., 30: 568-581, 1974.
- PAPADIMITRIOU, J. M. & SPECTOR, W. G. The origin, properties and fate of epithelioid cells. J. Path., 105: 187-203, 1971.
- PAPADIMITRIOU, J. M.; ROBERTSON, T. A. & WALTERS, N. I.

 An analysis of the phagocytic potencial of multinucleated foreign body giant cells.
 Amer. J. Path., 78: 343-358, 1975.
- PERAÇOLI, M. T. S.; MOTA, N. G. S. & MONTENEGRO, M. R. -Experimental paracoccidioidomycosis in Syrian hamster: morphology and correlation of lesions with humoral and cell-mediated immunity.
 Mycopathologia (Den Haag), 76: 7-12, 1982.
- PHILLIPS, S. M. & COLLEY, G. D. Immunologic aspects of host response to schistosomiasis: resistance, immunopathology, and eosinophil involvement. Progr. Allergy, 24: 49-57, 1978.
- PRPIC, V.; WEIEL, J. E.; SOMERS, S. D. et al. Effects of bacterial lipopolysaccharide on the hydrolysis of phosphatidylinositol 4.5-bis phosphate in murine peritoneal macrophages. J. Immunol., 139: 526-531, 1987.
- RALPH, P.; HO, M.; LITCOFSKY, P. B. & SPRINGER, T.A. -Expression and induction in vitro of macrophages differentiation antigens on murine cell lines. J. Immunol., 130: 108-114, 1983.

- RATNER, S. & VISON, S.B. Phagocytosis and encapsulation. Cellular immune response in Arthropoda. Amer. Zool., 23: 185-194, 1983.
- ROTH, J.; SUNDERKÖTTER, C.; GOEBELER, J. & SORG, C. Expression of the calcium-binding proteins MRP-8 and MRP-14 by early infiltrating cells in experimental contact dermatitis. Int. Arch. Allergy, 98: 140-145, 1992.
- RYAN, G. B. & SPECTOR, W. G. Macrophage turnover in inflammed connective tissue.
 Proc. roy. Soc. B., 175: 269-292, 1970.
- RYAN, G. B. & SPECTOR, W. G. Natural selection of long-lived macrophages in experimental granuloma. J. Path., 99: 139-136, 1969.
- SHEPARD, C.C. The experimental disease that follows the injection of human leprosy bacilli into mouse foot-pad of mice. J. exp. Med., 112: 445-454, 1960.
- SHIKAMA, Y.; KOBAYASHI, K.; KASAHARA, K. et al. -Granuloma formation by artificial microparticles in vitro. Macrophages and monokines play a critical role in granuloma formation. Amer. J. Path., 134: 1189-1199, 1989.
- SINHORINI, I. L.; MERUSSE, J. L. B. & MARIANO, M. Role of lymphatic drainage on the development of Calmette-Guérin Bacillusinduced granulomas in the hamster. Int. Arch. Allergy., 103: 166-174, 1994.
- SMITH, S. L. & COLLEY, D. G. The ultraestructural development of schistosoma egg granuloma in mice. Parasitology, 75: 119-123, 1977.
- SOMERS, S. D.; WEIEL, J. E.; HAMILTON, T. A. & ADAMS, D. O. Phorbol esters and calcium ionophore can prime murine peritoneal macrophages for tumor cell destruction. J. Immunol., 136: 4199-4204, 1986.
- SPECTOR, W. G. & LYKKE, A. W. J. The cellular evolution of inflammatory granulomata. J. Path. Bact., 92:163-167, 1966.
- SPECTOR, W. G. & RYAN, G. B. New evidence for the existence of long lived macrophages. Nature (Lond.), 221: 860, 1969.
- SPECTOR, W. G. & WILLOUGHBY, D. A. The origin of mononuclear cells in chronic inflammation and tuberculin reactions in the rat. J. Path. Bact., 96: 389-399, 1968.
- SPECTOR, W. G.; HEESOM, N. & STEVENS, J. E. Factors influencing chronicity in inflammation of rat skin. J. Path. Bact., 96: 203-213, 1968.
- SPECTOR, W. G.; REICHOLD, N. & RYAN, G. B. Degradation of granuloma-inducing microorganisms by macrophages. J. Path., 101: 339-354, 1970.
- SPECTOR, W.G. & MARIANO, M. Macrophage behaviour in experimental granulomas. In: FURTH, R. van, ed. - Mononuclear phagocytes in immunity infection and pathology. Oxford, Blackwell, 1975. p. 959-968.
- 100. STADECKER, M. J. & WRIGHT, J. A. Distribution and kinetics of mononuclear phagocytes in granulomas elicited by eggs of Schistosoma mansoni. Amer. J. Path., 116: 245-252, 1984.

- 101. STADECKER, M. J.; WYLER, D. J. & WRIGHT, J. A. Ia antigen expression and antigen presenting function by macrophages isolated from hypersensitivity granulomas. J. Immunol., 128: 2739-2744, 1981.
- STOSSEL, T. P. Phagocytosis. New. Engl. J. Med., 290: 717-723, 1974.
- STOSSEL, T. P. Phagocytosis: recognition and ingestion. Semin. Hemat., 12: 83-116, 1975.
- 104. SUNDERKÖTTER, C.; BEIL, W.; ROTH, J. & SORG, C. Cellular events associated with inflammatory angiogenesis in the mouse. Amer. J. Path., 138: 1-5, 1991.
- 105. SUTTON, J. S. & WEISS, L. Transformation of monocytes in tissue culture into macrophages, epithelioid cells and multinucleated giant cells. An electron microscope study. J. Cell Biol., 28: 303-332, 1966.
- 106. TAKEMURA, A. I. & WERB, Z. Secretory products of macrophages and their physiological function. Amer. J. Physiol., 246: C1-C9, 1984.
- 107. THOMAS, M. A. & MACSWEEN, R. N. M. Heterogeneity of rat alveolar macrophage populations: characterization of their surface antigens by antisera. Brit. J. exp. Path., 62: 65-73, 1981.
- 108. TSUDA, T.; DANNEMBERG, A. M.; ANDO, M.; ABBEY, H. & CORRIN, A. R. Mononuclear cell turnover in chronic inflammation. Amer. J. Path., 83: 255-268, 1976.
- VACHER, J. Immunological responses of guinea pigs to beryllium salts. J. med. Microbiol., 5: 91-108, 1972.
- 110. VAN DEN OORD, J. J.; WOLF-PEETERS, C.; FACCHETTI, F. & DESMET, V. J. Cellular composition of hypersensitivity-type granulomas: immunohistochemical analysis of tuberculous and sarcoidal lymphadenitis. Hum. Path., 15: 559-565, 1984.
- 111. VAN DER RHEE, H. J.; VAN DER BURGH WINTER, C. P. M. & DAEMS, W. T. The differentiation of monocytes into macrophages, epithelioid cells and multinucleated giant cells in subcutaneous granulomas. Cell Tiss. Res., 197: 355-378, 1979.
- 112. VOLKMAN, A. & GOWANS, J. L. The production of macrophages in the rat. Brit. J. exp. Path., 46: 50-61, 1965.
- 113. WARREN, K. S. Functional classification of granulomatous inflammation. Ann. N. Y. Acad. Sci., 278: 7-18, 1976.
- 114. WERB, Z. How the macrophages regulate its extracelullar environment. Amer. J. Anat., 116: 237-256, 1983.
- 115. ZWALDO, G.; BRUGGEN, J.; GERHARDS, G.; SCHELEGGEL, R. & SORG, C. - Two calcium-binding proteins associated with specific stages of myeloid cell differentiation are expressed by subsets of macrophages in inflammatory tissues. Clin. exp. Immunol., 72: 510-515, 1988.

Recebido para publicação em 13/07/1994 Aceito para publicação em 08/09/1994