

The genetics of glutamate oxaloacetate transaminase (GOT) in *Pinus merkusii* Jungh. et de Vriese

ISKANDAR ZULKARNAEN SIREGAR, TEDI YUNANTO

Department of Silviculture, Faculty of Forestry, Bogor Agricultural University, Kampus IPB Darmaga, P.O. Box 168, Bogor 16680, West Java, Indonesia, Tel. +62-251-8626806, Fax. +62-251-8626886, ✉email: siregar@ipb.ac.id

Manuscript received: 10 November 2009. Revision accepted: 29 December 2009.

ABSTRACT

Siregar IZ, Yunanto T (2010) The genetics of glutamate oxaloacetate transaminase (GOT) in *Pinus merkusii* Jungh. et de Vriese. *Biodiversitas 11*: 5-8. Inheritance and linkage analysis of glutamate oxaloacetate transaminase (GOT) in *P. merkusii* were performed using megagametophyte tissues of single tree seed-lots. One monomorphic locus (GOT-A) and three polymorphic loci (GOT-B, GOT-C and GOT-D) were identified, based on the allelic 1:1 segregation of putatively heterozygous trees. Linkage analysis revealed that GOT-C and GOT-D were closely linked loci with an average recombination rate (R) of about 5%. This result which is in contrast to the GOT genetics in other pine species was discussed with regard to the probably occurring gene duplication of one GOT locus.

Key words: *Pinus merkusii*, GOT, isozymes, inheritance and linkage.

INTRODUCTION

The glutamate oxaloacetate transaminase (GOT) which is identical to aspartate amino transferase (AAT, E.C. 2.6.1.1) is one of the enzyme systems often used in isozyme studies on plant and animal genetics. It catalyzes the reversible reaction of glutamate and oxaloacetate to 2-oxoglutarate and aspartate, thus having an important role in nitrogen metabolism by distributing this nutrient originally assimilated into glutamate to other compounds (Ireland and Joy 1985). In surveys of higher plant isozymes, three to four electrophoretic GOT zones have been reported (for review, see Gottlieb 1982). In maize, three different isozymes associated with the mitochondria (mGOT), with the glyoxysomes (gGOT), and with the soluble fraction of the cytosol (sGOT) were described, however, in later studies it was established that GOT occurs also in plastids (Weeden and Gottlieb 1980).

In genetic studies on forest tree species, GOT has frequently been assayed to determine genetic diversity and differentiation (Aguirre-Planter et al 2000; Ledig 2000; Ledig et al. 2001; Shea et al. 2002), since its controlling gene loci were found to be largely polymorphic. In conifer, GOT is generally encoded by three gene loci, one of which (GOT-C or GOT-3) specifies an isozyme consisting of double- or triple-banded variants near the cathode in zymograms. If the separation buffer in electrophoretic analysis has a pH value of 8.0 or 8.1, one or two bands of GOT-C migrate to the anode and one band migrates to the cathode, but all bands of this isozyme reveal a comigration, so that they were believed to be encoded by the same gene locus. In pine species, GOT has also been found to be encoded by three gene loci, i.e. GOT-A, GOT-B and GOT-C (Mejnartowicz and Bergmann 1985). However, Chung

(1981) reported a fourth locus of GOT in *P. sylvestris* and classified it as GOT-D, although this additional isozyme may correspond to one of the above-mentioned subbands comigrating with bands of GOT-C (Mejnartowicz and Bergmann 1985). Similarly, four isozyme zones were found in seed samples of *Pinus merkusii* from Thailand, however, one of these zones near the origin was invariant and was assigned to GOT-C which controls another variable zone (Changtragoon and Finkeldey 1995). However, new data with *P. merkusii* single tree seed-lots from Indonesia revealed the existence of two separate gene loci, i.e. GOT-C and GOT-D, controlling these multiband configurations in GOT zymograms (Siregar and Yunanto 2008). The studies on inheritance and linkage were carried previously with objective to confirm the number of GOT loci in *P. merkusii*. Output of the studies would be of significant importance in future biodiversity assessment, i.e. genetic diversity, of the genus *Pinus*.

MATERIALS AND METHODS

Seeds

Bulk seed-lots of *P. merkusii* from Aceh and Java (Indonesia) were primarily used to optimise the experimental methods for GOT resolution. In the second step haploid megagametophytes (endosperms) of 18 single tree seed-lots were analysed according to the procedure largely used by Changtragoon and Finkeldey (1995). Seeds were immersed overnight in water, dissected and the embryo carefully separated from the megagametophytes. The megagametophytes were ground in two drops of homogenising buffer (0.1 M Tris-HCl pH 7.3, containing 0.03% DTT and 2.5 % PVP).

Electrophoresis

These crude homogenates were then subjected to horizontal starch gel electrophoresis, using the buffer system of Ashton and Bradon, pH 8.6. Further details of electrophoretic and staining procedures were given by Liengsiri et al. (1990).

Segregation analysis

Inheritance of isozyme variants was confirmed through analysis of segregation ratios of haploid megagametophytes from putatively heterozygous mother trees. A test for goodness of fit to the 1:1 ratio, chi-square test (χ^2), was performed separately for each parent tree. In this analysis, all statistical tests were set up at 5% level of significance.

Linkage analysis

Chi-square tests (χ^2 test) of linkage were performed for each pair of segregating loci (Weir 1990). Two chi-square tests (χ^2) were calculated: the first one tests Mendelian segregation of alleles at each individual locus in the pair, and the last one tests for linkage between loci. A linkage analysis was performed for each individual parent tree when its progeny was found to be heterogenous at a given pair of loci. When linkage between loci was detected, the estimated frequency of recombination (r) was calculated using the formula $r = l/n$, where l is the number of recombinants and n is the total number of megagametophytes.

RESULTS AND DISCUSSION

The phenotypic variation and inheritance of GOT

After electrophoretic separation of extracts from seed megagametophytes, the gels stained for GOT showed four zones where one zone migrated towards the cathode. Figure 1 shows the four GOT zones in zymograms as well as variations in zone B and zone D. Contrary to this study, Zhelev et al. (2002) reported a monomorphic GOT enzyme system on *Pinus peuce*.

Estimates of the statistics used to test the conformity to the Mendelian mode of inheritance for each polymorphic locus of GOT zone are presented in Table 1. The zone GOT-A was invariant in the material used and supposed to be controlled by a single gene locus. Contrary to *P. merkusii* in Thailand, three allozymes were observed in the GOT-B zone and genetic analysis conducted with 10 single tree seed-lots showed that this zone is controlled by one single gene locus (Table 1).

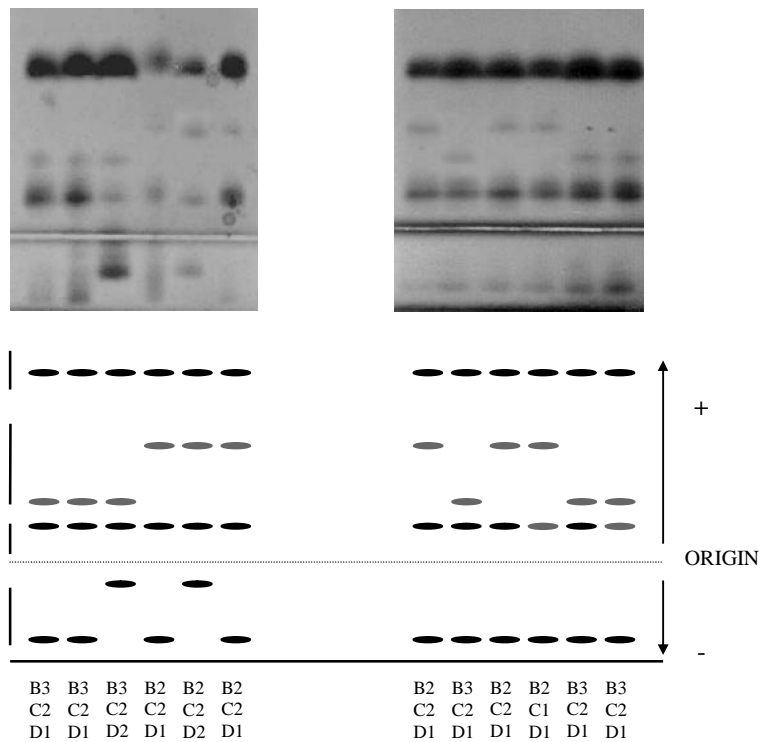


Figure 1. The GOT banding patterns of megagametophytes (*In*) from two trees. The indication of independent GOT-C and GOT-D zones is shown on the left zymogram.

Four allozymes were identified in the GOT-C zone. The same number was also observed by Na'iem and Indrioko (1996). The segregation at GOT-C was regular for several conifers species (Sousa et al. 2001). Nine out of twelve seed lots tested for segregation showed Mendelian 1:1 ratio, confirming that GOT-C is controlled by a single gene locus with four alleles (Table 1). Two heterozygous trees (J54 and J47) with genotype C_2C_3 and C_2C_4 , respectively, indicated a very strong segregation distortion.

The GOT-D zone was highly polymorphic, possessing two allozymes and genetic analysis conducted with 18 single tree seed-lots showed that this zone is controlled by a separate gene locus. Two seed lots showed segregation distortion and the allele D_2 seemed to be in favour.

Linkage relationships

As was shown in Figure 1, the zones GOT-C and GOT-D in zymograms appear to be independent isozymes which are encoded by two separate gene loci, however, these loci may be linked. In the following analysis, linkage among the three variable GOT loci will be assessed. Three pairs of heterozygous GOT loci were tested in different tree samples and the results of linkage analysis and statistical tests are presented in Table 2. According to the observed recombination values for significantly linked loci, only one linkage group can be constructed. GOT-C/GOT-D was closely linked with an average recombination value of 0.046. Other possible linkages were GOT-B/GOT-D ($R=0.167$) based on only one significant case of linkage, however, the allelic distribution in the other three samples

also tends to some loose linkage. Significant linkage in one but not all trees has also been reported for other conifer species (Fallour et al. 2001; Pastorino and Gallo 2001).

Table 1. Observed segregation of allozymes from megagametophytes trees of *P. merkusii* at gene loci of GOT and goodness of fit to the 1: 1 expected ratio.

Tree	Genotype	Sample size (N _{ij})	Observed segregation		t ² test
			(N _i)	(N _j)	
GOT-B					
J04	B ₂ B ₃	12	7	5	0.333
J09	B ₂ B ₃	12	6	6	0.000
J14	B ₂ B ₃	15	10	5	1.667
J29	B ₂ B ₃	22	13	9	0.727
J33	B ₂ B ₃	16	4	12	4.000*
J34	B ₂ B ₃	29	15	14	0.034
J37	B ₂ B ₃	16	9	7	0.250
J54	B ₂ B ₃	43	24	19	0.581
AI40	B ₂ B ₃	18	11	7	0.889
A04	B ₂ B ₃	20	8	12	0.800
Pooled	B ₂ B ₃	203	107	96	0,596
GOT-C					
J01	C ₁ C ₂	27	12	15	0.333
J02	C ₁ C ₂	34	21	13	1.882
J36	C ₁ C ₂	40	25	15	2.500
Pooled	C ₁ C ₂	101	58	43	4.715
J16	C ₂ C ₃	36	16	20	0.444
J26	C ₂ C ₃	62	39	23	4.129*
J54	C ₂ C ₃	43	39	4	28.488***
Pooled	C ₂ C ₃	141	94	47	282***
J27	C ₂ C ₄	57	31	26	0.439
J38	C ₂ C ₄	16	9	7	0.250
J47	C ₂ C ₄	36	26	10	7.111**
J48	C ₂ C ₄	20	12	8	0.800
J52	C ₂ C ₄	43	18	25	1.140
AI41	C ₂ C ₄	25	12	13	0.040
Pooled	C ₁ C ₂	197	108	89	1.980
GOT-D					
J01	D ₁ D ₂	27	16	11	0.926
J02	D ₁ D ₂	34	14	20	1.059
J04	D ₁ D ₂	12	7	5	0.333
J06	D ₁ D ₂	12	6	6	0.000
J09	D ₁ D ₂	12	6	6	0.000
J11	D ₁ D ₂	22	9	13	0.727
J12	D ₁ D ₂	39	6	33	18.692***
J27	D ₁ D ₂	57	48	9	26.684***
J29	D ₁ D ₂	21	10	11	0.048
J31	D ₁ D ₂	22	12	10	0.182
J34	D ₁ D ₂	29	14	15	0.034
J36	D ₁ D ₂	40	14	26	3.600
J47	D ₁ D ₂	36	14	22	1.778
J51	D ₁ D ₂	24	10	14	0.667
J52	D ₁ D ₂	40	20	20	0.000
J54	D ₁ D ₂	43	20	23	0.209
AI41	D ₁ D ₂	25	13	12	0.040
A39	D ₁ D ₂	17	11	6	1.471
Pooled	D ₁ D ₂	512	250	262	0.281

Note: Significant levels are 0.05 (*), 0.01 (**), 0.001 (***)

Table 2. Pairs of loci for which significant levels of linkage were detected in at least one tree.

Tree no.	Observed number by allelic combination				locus A	locus B	Linkage	Recombination Fraction (R)
	A _i B _i	A _i B _j	A _j B _i	A _j B _j				
GOT-B/GOT-C								
J54	20	4	19	0	0.581	28.488***	0.209	0.535
GOT-B/GOT-D								
J54	14	10	6	13	0.581	0.209	2.814	0.372
J04	6	1	1	4	0.333	0.333	5.333*	0.167
J12	8	7	6	8	0.034	0.034	0.310	0.448
J22	14	10	6	13	0.581	0.209	2.814	0.372
GOT-C/GOT-D								
AI41	2	10	11	2	0.040	0.040	11.560***	0.160
J01	0	13	12	0	0.040	0.040	25.000***	0.000
J02	0	11	7	0	0.889	0.889	18.000***	0.000
J36	0	25	14	1	2.500	3.600	36.100***	0.025

Note: Significant levels are 0.05(*), 0.001(***)

Discussion

The genetic analysis conducted in this study has been based on megagametophytes of seeds from putatively heterozygous trees, since progenies from controlled crosses were not available. In coniferous tree species, isozyme analysis of haploid megagametophytes, which genetically represent the female gametes, allows observation of regular meiotic segregation at individual loci (Bergmann 1973). This method therefore allows for both, the identification of the number of loci controlling an enzyme system and the independent transmission or linkage of alleles among these different loci. The study on the genetic control of observed isozyme phenotypes in *P. merkusii* was mainly carried out to confirm the results of previous works (Changtragoon and Finkeldey 1995; Szmidi et al. 1996). However, in contrast to these studies it was noticed that the enzyme system GOT turned out to be controlled by four gene loci. This result is surprising because it is much in contrast to all other pine species where only three GOT loci were identified. Furthermore, it was established that the allelic segregation at two loci did not occur independently. However, Mejnartowicz and Bergmann (2002) and Lewandowski et al. (2002) reported that four AAT or GOT activity zones were found in *Abies alba* and *Picea abies*, respectively.

The close linkage between the loci GOT-C and GOT-D has not been reported previously because the two loci are often regarded as one locus. In many conifers, the GOT-C and GOT-D are actually two widely separated, always co-migrating bands and one of these bands is often moving towards the cathode. This band configuration has led to the one-locus hypothesis. This interpretation may be correct if we assume a gene duplication which has occurred just very recently. In this case, recombinants between adjacent gene loci can only be detected among thousands of seed megagametophytes, which were not included in the former inheritance studies. Later on, the distance between the duplicated GOT loci may have become larger (e.g. by some insertions) and, hence, recombinants between the two loci can be detected with smaller seed samples, as is the case with *P. merkusii*. The various recombination frequencies between GOT-C and GOT-D observed in four trees, i.e.

from 0% to 16%, might be attributed to the genotype at the GOT-C locus. The highest recombination rate ($R=16\%$) was observed in tree AI41 which is C_2C_4 , while the other three trees were C_1C_2 . Indication of possible influence of temperature during meiosis has been mentioned by Rudin and Ekberg (1978) to explain the different levels of recombination frequency observed among trees of *P. sylvestris*. The later reason should be excluded from our results because of the absence of such high temperature fluctuation in the tropics.

Additionally, it must be emphasized that the linkage analysis was only based on female gametes. In spite of the general observation that gamete selection is less pronounced on the female than on the male side in flowering plants (Rudin and Ekberg 1978), female gamete selection may have influenced recombination data. In *P. radiata*, it was found that the rate of recombination is 43% greater in male gametes than in female gametes. In addition, significant linkage in one but not in all trees has also been reported for other conifer species. The possible reason for different recombination values among trees is related to environmental effects or modification of chromosome structure, i.e. insertion, translocation, deletion, reversion, etc. A high frequency of events leading to such meiotic irregularities has been observed in conifers (Andersson et al. 1969). Furthermore, the history of colonization of a species might contribute also to slightly different chromosomal arrangement resulting in a deviating linkage pattern as shown in *P. sylvestris* from southern and northern Sweden (Rudin and Ekberg 1978).

The gene duplication followed by mutation is a major mechanism of evolution. Duplication results in two identical copies, each of which can retain original function, becomes functionless (formation of pseudo-gene) or develops a new function. The GOT loci has often been reported to be tightly linked with PGI loci in pines and other coniferous species and this linkage has become an indication of a striking conservatism of chromosomal structure in this species (Ledig 1998). Using *P. merkusii* samples from Thailand and Vietnam, Szmids et al. (1996) have found polymorphism in PGI and this opens up the possibility for future work to confirm the finding of such conservatism of linkage groups.

CONCLUSION

In *P. merkusii*, GOT was identified to have one monomorphic locus (GOT-A) and three polymorphic loci (GOT-B, GOT-C and GOT-D). Linkage analysis confirmed that GOT-C and GOT-D were closely linked loci with an average recombination rate (R) of about 5%.

ACKNOWLEDGEMENT

The authors would like to thank Perum Perhutani for providing sample materials.

REFERENCES

- Aguirre-Planter E, Furnier GR, Eguiarte LE (2000) Low levels of genetic variation within and high levels of genetic differentiation among populations of species of *Abies* from southern Mexico and Guatemala. *Am J Bot* 87: 362-371.
- Andersson E, Ekberg I, Eriksson G (1969) A summary of meiotic investigations in conifers. *Stud For Suec* 70: 1-20
- Bergmann F (1973) Genetische untersuchungen bei *Picea abies* mit hilfe der isoenzym-identifizierung. II. Genetische kontrolle von esterase- und leucinamino-peptidase-isoenzymen im haploiden endosperm ruhender samen. *Theor Appl Genet* 43: 222-225.
- Changtragoon S, Finkeldey R (1995) Inheritance of isozyme phenotypes of *Pinus merkusii*. *J Trop For Sci* 8 (2): 167-177.
- Chung MS (1981) Biochemical methods for determining population structure in *Pinus sylvestris* L. *Act For Fenn* 173: 28.
- Fallour D, Fady B, Lefevre F (2001) Evidence of variation in segregation patterns within a *Cedrus* population. *J Hered* 92: 260-266
- Gottlieb LD (1982) Conservation and duplication of isozymes in plants. *Science* 216: 373-380.
- Ireland RJ, Joy KW (1985) Plant transaminases. In Christen P, Metzler DE (eds). *Transaminases*. John Wiley & Son, New York.
- Ledig FT (1998) Genetic Variation in *Pinus*. In Richardson DM (ed) *Ecology and biogeography of Pinus*. Cambridge University Press, Cambridge, UK.
- Ledig FT (2000) Founder Effect and the Genetic Structure of Coulter Pine. *American Genetic Association* 91: 307-315.
- Ledig FT, Capó-Arteaga MA, Hodgskiss PD, Sbay H, Flores-López C, Conkle MT, Bermejo-Velázquez B (2001) Genetic diversity and the mating system of a rare Mexican piñon, *Pinus pinceana*, and a comparison with *Pinus maximartinezii* (Pinaceae). *Am J Bot* 88: 1977-1987.
- Lewandowski A, Samocko J, Burczyk J (2002) Inheritance of AAT in *Picea abies* – some old and new facts. *Silvae Genetica* 51: 161-163
- Liengsiri C, Piewluang C, Boyle TJB (1990) Starch gel electrophoresis of tropical trees. A Manual. ASEAN-Canada Forest Tree Seed Centre and Petawawa National Forestry Institute, Muak Lek, Saraburi, Thailand.
- Mejnartowicz L, Bergmann F (1985) Genetic differentiation among Scots pine populations from the lowlands and the mountains in Poland. In: Gregorius, H.-R. (ed.) *Population genetics in forestry*. Springer, Berlin.
- Mejnartowicz L, Bergmann F (2002) Mode of Inheritance of Aspartate Aminotransferase in Silver-Fir (*Abies alba* Mill.). *Silvae Genetica* 52: 15-17
- Na'iem M, Indrioko S (1996) Inheritance of isozyme variants of Tusam (*Pinus merkusii*) artificial stands in Java. In: Rimbawanto A, Widayatmoko AYPBC, Shuaendi H, Furukoshi T (eds). *Proceeding of International Seminar on Tropical Plantation Establishment: improving productivity through genetic practices*. FTIRDI and JICA Yogyakarta.
- Pastorino MJ, Gallo LA (2001) Linkage relationships as a useful tool to state interspecific gene homology: case study with isozyme loci in *Austrocedrus chilensis* (Cupressaceae). *Silvae Genetica* 50: 233-239.
- Rudin D, Ekberg I (1978) Linkage studies in *Pinus sylvestris* L. using macro-gametophyte allozymes. *Silvae Genetica* 27: 1-12.
- Shea KL, Furnier GR (2002) Genetic variation and population structure in central and isolated populations of balsam fir, *Abies balsamea* (Pinaceae). *Am J Bot* 89: 783-791.
- Siregar IZ, Yunanto T (2008) Inference on the possible causes of segregation distortion from open pollination progenies of merkus pine (*Pinus merkusii*). *Hayati J Biosci* 2008 15: 155-158.
- Sousa VA, Hattermer HH, Robinson IP (2001) Inheritance and linkage relationships of isozyme variants of *Araucaria angustifolia* (BERT.) O. KTZE. *Silvae Genetica* 51: 5-6.
- Szmids AE, Wang XR, Changtragoon S (1996) Contrasting patterns of genetic diversity in two tropical pines: *Pinus kesiya* (Royle ex Gordon) and *P. merkusii* (Jungh. et De Vriese). *Theor Appl Genet* 92: 436-441.
- Weeden NF, Gottlieb LD (1980) The genetics of chloroplast enzymes. *J Hered* 71: 392-396.
- Weir BS (1990) *Genetic data analysis*. Sinauer, Sunderland, M.A.
- Zhelev P, Gomory D, Paule L (2002) Inheritance and linkage of allozymes in a Balkan endemic *Pinus peuce* Griseb. *J Hered* 93: 60-63.