

GENE FAMILY UPDATE

Open Access

The human protein disulfide isomerase gene family

James J Galligan¹ and Dennis R Petersen^{2*}

Abstract

Enzyme-mediated disulfide bond formation is a highly conserved process affecting over one-third of all eukaryotic proteins. The enzymes primarily responsible for facilitating thiol-disulfide exchange are members of an expanding family of proteins known as protein disulfide isomerases (PDIs). These proteins are part of a larger superfamily of proteins known as the thioredoxin protein family (TRX). As members of the PDI family of proteins, all proteins contain a TRX-like structural domain and are predominantly expressed in the endoplasmic reticulum. Subcellular localization and the presence of a TRX domain, however, comprise the short list of distinguishing features required for gene family classification. To date, the *PDI* gene family contains 21 members, varying in domain composition, molecular weight, tissue expression, and cellular processing. Given their vital role in protein-folding, loss of PDI activity has been associated with the pathogenesis of numerous disease states, most commonly related to the unfolded protein response (UPR). Over the past decade, UPR has become a very attractive therapeutic target for multiple pathologies including Alzheimer disease, Parkinson disease, alcoholic and non-alcoholic liver disease, and type-2 diabetes. Understanding the mechanisms of protein-folding, specifically thiol-disulfide exchange, may lead to development of a novel class of therapeutics that would help alleviate a wide range of diseases by targeting the UPR.

Keywords: Disulfide bond, Thioredoxin, Calsequestrin, UPR, Unfolded protein response, ER stress

Introduction

Increasing evidence supports an important role for misfolded proteins in the pathogenesis of numerous diseases including diabetes, Alzheimer disease, Parkinson disease, and both alcoholic and non-alcoholic liver disease [1]. Accumulation of misfolded proteins (or erred protein load) is generally caused by either decreased disposal of erred protein or a decrease in the correct folding of proteins [1]. Disulfide bond formation represents a fundamentally important post-translational modification that is a critical step in the folding of nascent peptides in the endoplasmic reticulum (ER) [2]. These covalent linkages are formed between the side-chains of cysteine residues and represent a key rate-limiting step in protein maturation [3]. The enzymatic formation, breakage, and subsequent rearrangement of cysteine linkages are crucial to protein structure and function and primarily mediated

by members of the protein disulfide isomerase (PDI) family [4]. All genes in the *PDI* family are part of a superfamily referred to as the thioredoxin (TRX) superfamily, which also includes the glutaredoxins, TRXs, ferredoxins, and peroxidoxins [5].

The *PDI* gene family currently comprises 21 genes, varying in size, expression, localization, and enzymatic function. Although it is implied that all members of the PDI family possess the ability to rearrange disulfide bonds, only a subset is considered orthologous and able to carry out these reactions, with the other members being paralogous and linked to the family through evolution, not function [4]. While these proteins may be functionally different, the unifying feature of all PDI family members is the presence of a TRX-like domain [2]. These may be present as either a catalytically active **a** or **a'** domain (the presence of a CXXC motif) or a catalytically inactive **b** or **b'** domain (for a more detailed review on the precise role of these domains, see the work of Ellgaard et al.) [2,4]. Extensive research has assessed the roles of these domains and revealed the **b'** domain to be

* Correspondence: Dennis.Petersen@UCDenver.edu

²Molecular Toxicology and Environmental Health Sciences Program, Department of Pharmacy and Pharmaceutical Sciences, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, USA
Full list of author information is available at the end of the article

the primary peptide- or protein-binding domain [4]. Previous literature has highlighted the features of a number of *PDI* family members; however, with an increasing amount of cDNA and EST sequence information deposited in the NCBI database, a composite review is required to further characterize and compare all 21 current members of the *PDI* gene family (Table 1).

Domain composition of the *PDI* family proteins

Proteins in the *PDI* family are largely expressed in the ER, although few family members have been detected in other subcellular compartments [6]. Due to their localization, the presence of a short NH₂-terminal signal peptide exists in all members of the family. These peptides are typically 15–30 amino acids (a.a.) in length and are cleaved upon entry into the ER [7]; this has led to some sequence discrepancy among multiple *PDI* proteins. As indicated, the common thread between all members of the *PDI* proteins is the presence of at least one TRX-like domain, whether it is catalytically active (a) or inactive (b) [2]. The active site of the a-type domains also varies greatly, with the “classical” motif being comprised of Cys-Gly-His-Cys. The cysteine

residues in these active sites are considered redox active, undergoing active shuffling of disulfide bonds [2]. The surrounding a.a. largely play a role in the regulation of the pKa of the cysteines, dictating the local redox potential and thus regulating the catalytic ability of these cysteines to actively oxidize or reduce disulfide bonds (for a more comprehensive review on the redox potential of *PDI*, see the work of Hatahet et al.) [3]. Extensive biochemical and biophysical experimentation has taken place analyzing TRX-like domain containing proteins; however, a complete crystal structure is currently not available for most family members. Another common characteristic of the *PDI* family of proteins is the presence of a COOH-terminal ER retention sequence comprised of a Lys-Asp-Glu-Leu-like sequence [2]. Whereas these sequences may differ greatly in a.a. composition, only four *PDI* proteins do not contain this sequence. Figure 1 shows the domain composition of the 21 proteins in the *PDI* family.

Evolutionary divergence of the *PDI* gene family

As mentioned, all genes encompassed in the *PDI* family belong to the *TRX* superfamily of genes [5]. The unifying

Table 1 Human *PDI* genes as listed in the Human Gene Nomenclature Committee (HGNC) database

Gene name	Other aliases	Protein name	Chromosomal location	Amino acids
<i>AGR2</i>	XAG-2, HAG-2, AG2, PDIA17	Anterior gradient protein 2 homolog	7p21.3	175
<i>AGR3</i>	HAG3, hAG-3, BCMP11, PDIA18	Anterior gradient protein 3 homolog	7p21.1	166
<i>CASQ1</i>	PDIB1	Calsequestrin-1	1q21	396
<i>CASQ2</i>	PDIB2	Calsequestrin-2	1p13.3-p11	399
<i>DNAJC10</i>	MTHr, ERdj5	DnaJ (Hsp40) homolog, subfamily C, member2q32.1 10		793
<i>ERP27</i>	FLJ32115, ERp27, PDIA8	Endoplasmic reticulum resident protein 27	12p12.3	273
<i>ERP29</i>	ERp28, ERp31, ERp29, PDI-DB, PDIA9	Endoplasmic reticulum resident protein 29	12q24.13	261
<i>ERP44</i>	KIAA0573, PDIA10	Endoplasmic reticulum resident protein 44	9q22.33	406
<i>P4HB</i>	DIA1, PROHB, DSI, GIT, PDI, PO4HB, P4Hb	Protein disulfide-isomerase	17q25	508
<i>PDIA2</i>	PDA2, PDip	Protein disulfide-isomerase A2	16p13.3	525
<i>PDIA3</i>	P58, ERp61, ERp57, ERp60, GRP57, PI-PLC, HsT17083	Protein disulfide-isomerase A3	15q15	505
<i>PDIA4</i>	ERP70, ERP72	Protein disulfide-isomerase A4	7q35	645
<i>PDIA5</i>	PDIR, FLJ30401	Protein disulfide-isomerase A5	3q21.1	519
<i>PDIA6</i>	P5, ERp5	Protein disulfide-isomerase A6	2p25.1	440
<i>PDILT</i>	PDIA7, ERp65	Protein disulfide-isomerase-like protein of the16p12.3 testis		584
<i>TMX1</i>	TMX, PDIA11	Thioredoxin-related transmembrane protein 1	14q22.1	280
<i>TMX2</i>	PDIA12	Thioredoxin-related transmembrane protein 2	11cen-q22.3	296
<i>TMX3</i>	FLJ20793, KIAA1830, PDIA13	Protein disulfide-isomerase TMX3	18q22	454
<i>TMX4</i>	DJ971N18.2, KIAA1162, PDIA14	Thioredoxin-related transmembrane protein 4	20p12	349
<i>TXNDC5</i>	MGC3178, FLJ21353, FLJ90810, EndoPDI, Hcc-2, ERp46, Thioredoxin domain-containing protein PDIA15	5	6p24.3	432
<i>TXNDC12</i>	TLP19, ERP18, ERP19, HAG-1, AGR1, PDIA16	Thioredoxin domain-containing protein 12	1p32.3	172

DNAJC10 has been included following identification of this gene in the *PDI* gene family. Synonyms of these genes have also been corrected.

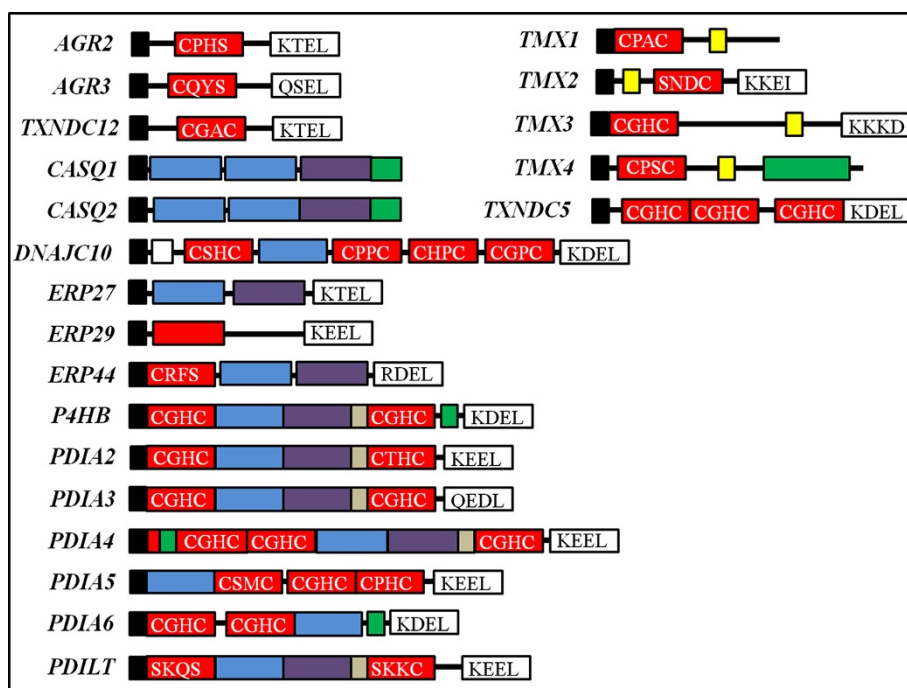


Figure 1 Schematic representation of the domain compositions of the 21 proteins in the *PDI* gene family. All proteins contain a short NH₂-terminal signal sequence designated in black. Catalytically active TRX-like domains (**a** or **a'**) are represented in red with active sites noted; inactive TRX domains in blue (**b**) and purple (**b'**); green represents the Asp/Glu rich Ca²⁺-binding domains; linker regions (gray); transmembrane domains (yellow); COOH-terminal ER-retention sequences in white with a.a. composition denoted. Although ERP29 does contain an **a**-like domain, this is based on homology and not catalytic efficiency. Sequences and domain compositions were acquired and verified from the National Center for Biotechnology Information (NCBI) database. Figure was adapted and modified from [4].

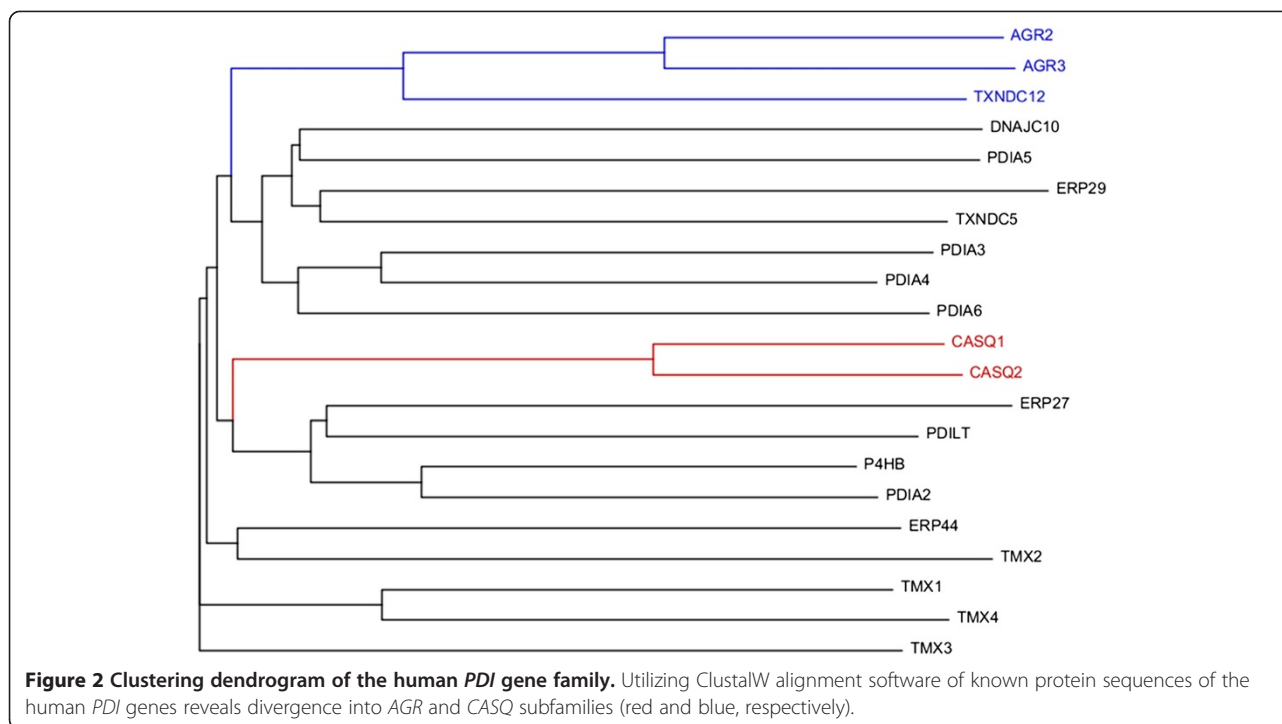
theme between these proteins is the presence of at least one TRX-like domain, whether this be catalytically active (**a** or **a'**) or inactive (**b** or **b'**) [4]. These domains contain a TRX structural fold that has amino acids arranged in a conserved three-dimensional conformation [8]. While the enzymatic function of these domains is not conserved, the current theory proposes that all PDI family members evolved through domain duplications from an ancestral prokaryotic PDI which contained a single TRX domain [9]. Although all human PDIs possess a TRX-like domain, this remains one of the few similarities between these proteins, differing greatly in molecular mass and a.a. composition outside of the TRX fold. Phylogenetic analysis, presented as a dendrogram in Figure 2, outlines the evolutionary divergence of the human PDI proteins. Sequence analysis reveals a subset of genes within this family that are evolutionarily related, as shown by the calnexin (CASQ) and anterior gradient (AGR) branches (in red and blue, respectively). Supporting the hypothesis that these subsets of genes are phylogenetically related, a high degree of similarity was also observed with both domain architecture (Figure 1) and sequence identity (Table 2). Although similarities are evident, the overall homology between the proteins is quite minimal. This is supported by previous attempts

to cluster large sets of both eukaryotic and prokaryotic PDIs where marginal resolution of PDI domains was also observed [9]. Given the broad spectrum of both enzymatic functions and domain compositions, it is not surprising that the proteins share little sequence homology with one another (Table 2).

The human *PDI* gene family

DNAJC10

The *DNAJC10* gene is located at Chr 2q32.1 and encodes the 793-a.a. DNAJC10 protein (also commonly known as ERdj5 or MTHr) [10]. The *DNAJC10* gene consists of 3483 bp; transcription of two splice variants has been identified due to a skipped exon, resulting in a 138-bp (46 a.a.) absence, present between nucleotides 1243 and 1244 [10]. To date, a total of four missense single-nucleotide polymorphisms (SNPs) have been reported for ERdj5, located at a.a. 76 (Asp → Asn), 347 (Leu → Ile), 414 (Tyr → Cys) and 646 (His → Gln); ERdj5 also contains one potential Asn-linked glycosylation site, present at a.a. 530. Like other *PDI* family members, the *DNAJC10* promoter region contains a putative ER stress element (ERSE) box, yielding gene induction following ER stress [11]. Expression patterns of *DNAJC10* revealed ubiquitous expression with varying intensities and high



levels of expression in secretory tissues such as the pancreas and testis [10,11]. In addition to the PDI family, DNAJC10 is also a member of the ERdj family, being comprised of an unverified NH₂-terminal signal peptide (32 a.a.), one DnaJ domain (which plays a major role in protein folding), five TRX domains (one **b** and four **a** type domains, active sites Cys-Ser-His-Cys, Cys-Pro-Pro-Cys, Cys-His-Pro-Cys, and Cys-Gly-Pro-Cys), and a COOH-terminal ER retention sequence (Lys-Asp-Glu-Leu) [10,12]. Despite the high number of TRX domains, DNAJC10 was found to possess roughly one-third the activity of P4HB and displayed no oxidase or isomerase activity [12]. These results were found to reflect an apparent redox equilibrium constant of 190 mM, roughly 100 times more reducing than that of the ER [12].

ERP27

Currently, little is known about the precise role and function of *ERP27*. *ERP27* was discovered in 2006 following a database search for novel human PDI family members [13]. The *ERP27* gene has been mapped to Chr 12p12.3 and encodes the 273-a.a protein. The ERP27 protein contains a cleavable NH₂-terminal signal sequence, leaving the mature protein to be 248 a.a, beginning at Glu-26 [13]. ERP27 does not contain a redox active Cys-X-X-Cys TRX motif, however, the protein does contain one **b** and one **b'** type domains and an ER retention sequence (Lys-Val-Glu-Leu) [13]. ERP27 was also found to directly interact with another PDI family member, PDIA3 [13]. Expression of ERP27 is fairly

ubiquitous, with highest expression found in pancreas [13]. To date, no knockout studies have been conducted and future studies are needed to fully understand the function and role of ERP27.

ERP29

The *ERP29* gene is located at Chr 12q24.13 and may represent a gene-duplication event with *ERP27*, given their close proximity. *ERP29* encodes a protein of 261 a. a. termed ERP29 (also known as ERP28). In a comprehensive study of the genomic organization of *ERP29*, Sargsyan et al. studied a 5'-flanking region consisting of ~2 kb, 3 exons, 2 introns and a 3'-flanking region of 0.31 kb [14]. ERP29 contains an NH₂-terminal signal peptide (32 a.a.), one **a** type TRX domain and a COOH-terminal ER-retention sequence (Lys-Glu-Glu-Leu) [15]. While the presence of one **a** type domain is present, ERP29 is unique in that it does not contain an active-site motif; assignment of the TRX domain is placed strictly on sequence homology with the **a**-type TRX domain, not on the activity of the domain. ERP29 contains two potential phosphorylation sites, both located on tyrosine residues, located at a.a. 64 and 66. ER localization of ERP29 was also confirmed using immunofluorescence and subcellular fractionation [15]. ERP29 is ubiquitously expressed with high levels being found in secretory tissues as well as the prostate, pancreas, and liver [14,16]. Although *ERP29* lacks any identified ERSE in its promoter region, ERP29 is described as an ER stress-inducible protein and has been shown to

Table 2 Amino acid similarity between human PDI proteins, as reported using EMBOSS Water Pairwise Alignment Algorithm (http://www.ebi.ac.uk/Tools/psa/emboss_water/)

	AGR2	AGR3	CASQ1	CASQ2	DNAJC10	ERP27	ERP29	ERP44	P4HB	PDIA2	PDIA3	PDIA4	PDIA5	PDIA6	PDILT	TMX1	TMX2	TMX3	TMX4	TXNDC5	
AGR2																					
AGR3	65.0% (81.4%)																				
CASQ1	18.8% (34.8%)	17.4% (40.2%)																			
CASQ2	15.3% (37.4%)	17.40% (34.0%)	68.4% (86.1%)																		
DNAJC10	16.2% (37.2%)	19.0% (35.8%)	16.8% (33.3%)	18.2% (33.3%)																	
ERP27	28.2% (41.0%)	20.5% (40.2%)	17.2% (34.4%)	19.3% (37.6%)	18.6% (35.8%)																
ERP29	20.6% (40.2%)	23.3% (39.5%)	18.2% (31.1%)	20.4% (33.0%)	18.1% (30.9%)	15.7% (38.9%)															
ERP44	22.2% (39.4%)	26.5% (47.0%)	19.2% (42.9%)	20.0% (40.8%)	20.4% (30.6%)	24.3% (40.5%)	25.3% (42.0%)														
P4HB	20.1% (34.3%)	21.7% (29.2%)	21.6% (39.3%)	22.9% (38.3%)	20.2% (36.0%)	32.5% (54.1%)	24.50% (39.9%)	23.5% (42.9%)													
PDIA2	23.5% (35.5%)	20.8% (38.7%)	21.2% (31.8%)	20.6% (38.7%)	20.8% (32.1%)	28.6% (49.0%)	23.3% (37.2%)	23.3% (37.7%)	47.3% (64.5%)												
PDIA3	21.7% (37.5%)	16.9% (37.9%)	21.5% (33.3%)	18.8% (31.3%)	21.4% (37.5%)	21.8% (38.9%)	22.3% (34.1%)	23.1% (40.6%)	33.6% (51.0%)	31.1% (49.5%)											
PDIA4	19.80% (31.6%)	19.7% (38.6%)	18.8% (38.2%)	21.4% (38.5%)	17.5% (30.8%)	18.3% (35.0%)	29.6% (48.1%)	24.0% (40.9%)	36.8% (55.1%)	29.1% (46.2%)	41.9% (60.2%)										
PDIA5	24.2% (45.1%)	22.1% (44.1%)	21.3% (35.1%)	21.7% (36.9%)	21.4% (40.0%)	19.6% (33.5%)	19.7% (33.9%)	22.6% (40.6%)	24.3% (38.3%)	23.1% (35.0%)	22.9% (37.1%)	21.2% (31.5%)									
PDIA6	27.6% (35.2%)	15.2% (57.6%)	24.0% (40.6%)	19.4% (33.1%)	32.7% (48.0%)	20.6% (34.8%)	17.0% (32.2%)	20.2% (38.4%)	23.5% (31.4%)	21.8% (28.6%)	24.2% (33.4%)	25.8% (40.0%)	28.1% (43.1%)								
PDILT	23.9% (45.7%)	16.2% (35.9%)	20.1% (43.4%)	21.1% (41.8%)	20.4% (36.1%)	29.7% (54.1%)	19.1% (38.5%)	21.9% (40.9%)	31.8% (52.4%)	32.1% (51.3%)	22.1% (37.1%)	26.0% (46.7%)	19.9% (35.3%)	23.5% (36.3%)							
TMX1	18.6% (36.1%)	24.8% (36.0%)	23.7% (37.8%)	17.2% (29.9%)	21.1% (38.2%)	23.9% (39.4%)	15.5% (32.3%)	25.8% (49.0%)	29.9% (46.9%)	25.7% (40.0%)	30.0% (50.0%)	25.1% (41.3%)	35.0% (56.4%)	36.2% (59.0%)	27.0% (39.3%)						
TMX2	23.0%	18.6%	20.2%	28.6%	23.3%	22.0%	25.6%	23.10%	25.0%	28.3%	25.8%	25.7%	16.4%	24.7%	24.7%	24.6%					

Table 2 Amino acid similarity between human PDI proteins, as reported using EMBOSS Water Pairwise Alignment Algorithm (http://www.ebi.ac.uk/Tools/psa/emboss_water/) (Continued)

	(36.1%)	(33.1%)	(39.9%)	(44.4%)	(44.8%)	(41.5%)	(43.6%)	(38.0%)	(40.9)	(44.6%)	(36.9%)	(43.7%)	(34.9%)	(38.9%)	(42.9%)	(43.1%)				
TMX3	20.0%	19.7%	20.3%	21.8%	22.4%	18.1%	20.5%	21.4%	27.8%	21.6%	23.1%	27.1%	23.5%	45.3%	19.2%	34.2%	18.7%			
	(37.4%)	(36.5%)	(43.0%)	(43.6%)	(44.1%)	(33.3%)	(35.7%)	(34.7%)	(47.9%)	(40.3%)	(41.1%)	(42.7%)	(41.2%)	(64.2%)	(32.2%)	(50.5%)	(38.4%)			
TMX4	23.5%	25.7%	27.0%	23.1%	23.7%	28.6%	19.5%	20.1%	21.8%	24.9%	24.3%	20.5%	27.2%	24.0%	25.0%	43.6%	24.4%	22.4%		
	(39.5%)	(36.6%)	(62.2%)	(47.3%)	(39.4%)	(42.9%)	(36.0%)	(37.0%)	(34.4%)	(37.0%)	(45.7%)	(32.1%)	(41.6%)	(37.1%)	(42.5%)	(65.0%)	(42.2%)	(34.2%)		
TXNDC5	21.6%	20.9%	22.9%	20.7%	25.4%	22.0%	20.8%	33.3%	26.2%	26.9%	24.4%	23.5%	27.3%	33.0%	23.3%	24.6%	29.5%	51.4%	21.6%	
	(28.4%)	(39.5%)	(37.9%)	(34.0%)	(41.5%)	(37.9%)	(32.9%)	(50.0%)	(37.2%)	(37.5%)	(38.5%)	(34.1%)	(43.8%)	(53.0%)	(34.9%)	(33.8%)	(50.0%)	(71.6%)	(38.0%)	
TXNDC12	34.2%	38.3%	20.0%	41.2%	18.5%	20.2%	27.5%	21.1%	26.2%	26.9%	24.1%	22.9%	22.7%	26.9%	23.1%	27.7%	21.6%	31.9%	18.7%	20.1%
	(50.6%)	(53.9%)	(30.8%)	(70.6%)	(36.3%)	(30.2%)	(40.0%)	(32.2%)	(37.2%)	(37.5%)	(39.7%)	(41.2%)	(43.8%)	(37.0%)	(38.2%)	(39.8%)	(37.4%)	(48.9%)	(32.1%)	(31.5%)

Sequence identity and similarity (in parentheses) are noted.

co-localize with other ER stress-associated chaperones, glucose-regulated protein 78 (GRP78 or BiP) [16]. ERP29 has been postulated to play a role in the progression of tumorigenesis in mice; following implantation of both knockdown and over-expressed null ERP29 MCF-7 cells, a significant decrease in tumor size and altered morphogenesis was observed in mice [17]. Currently, no knockout mouse is available for *Erp29*.

ERP44

The *ERP44* gene is located at Chr 9q22.33 and encodes the 406-a.a. ERP44 protein [18]. ERP44 was discovered following immunoprecipitation experiments with human endoplasmic reticulum oxidoreductase-1 α (ERO1-L α) and was originally identified as KIAA0573 [18]. The coding sequence of *ERP44* contains 12 exons and there is an ERSE in the promoter region of the gene [18]. The human ERP44 protein contains an unverified NH₂-terminal signal peptide (29 a.a.), three TRX-like domains (one being the catalytically active a domain with a Cys-Arg-Phe-Ser active site), and the ER-retention sequence (Arg-Asp-Glu-Leu) [18]. The precise physiological function of ERP44 has yet to be determined; however, oxidation of ERO1 α has been observed, suggesting that ERP44 may control the function of the ERO1 proteins, thus controlling the redox state of the ER [18]. ERP44 is up-regulated during ER stress response and has also been proposed to play a role in adiponectin secretion which influences glucose regulation and fatty acid catabolism [18,19].

P4HB

P4HB is the first described member of the *PDI* gene family and was originally identified as the β -subunit of human prolyl-4-hydroxylase (P4H) [20,21]. The *P4HB* gene is located at Chr 17q25 and consists of 11 exons [20,22]. The promoter region of *P4HB* contains 11 protein binding sites, including an ERSE, underscoring the dynamic nature of *P4HB* transcriptional regulation [23]. Although the presence of an ERSE has been confirmed in the promoter region of the gene, P4HB is considered to be a weakly-induced ER stress protein, likely due to its high abundance. The promoter region also contains six CCAAT elements in the first 378 nt of the gene, and mutations introduced into any of these elements was found to reduce promoter activity by up to 50% [23].

The *P4HB* gene encodes a 508-a.a. protein containing a 17-a.a. signal peptide, four TRX domains with two **a** type (Cys-Gly-His-Cys, Cys-Gly-His-Cys), an Asp/Glu rich domain (a.a. 480 to 500), and a COOH-terminal ER retention sequence (Lys-Glu-Asp-Leu) [2]. P4HB is ubiquitously expressed in nearly all tissues and is very highly abundant; estimations predict P4HB to account for up to 0.8% of total cellular protein [24]. Currently, a

crystal structure has yet to be resolved for the full, intact protein, although multiple domains have been solved. Despite this, P4HB remains the most widely studied and understood protein in the family. P4HB is effective at oxidizing, reducing, and isomerizing disulfide bonds both *in vitro* and *in vivo* and exists as a homodimer [25,26]. Although its role in disulfide bond generation remains the most widely studied enzymatic action, P4HB has also been shown to exhibit chaperone-like activity, demonstrating an additional role in maturation of nascent proteins regardless of the presence of disulfide bonds [27,28]. P4HB has also been shown to be an essential subunit for microsomal triglyceride transfer protein and P4H [21,29]. To date, no viable knockout mouse strain for *P4hb* has been reported, likely due to its critical role in disulfide bond generation [3].

PDIA2

PDIA2 was identified in 1996 as a pancreas-specific member of the PDI family, resulting in its common name, PDIp [30]. Located on Chr 16p13.3, the initial characterization of the *PDIA2* gene revealed that it encodes a protein with an ORF of 511 a.a.; these studies were unable to validate an in-frame stop codon located 5' upstream of the ATG start site [30]. Due to this discrepancy, the *PDIA2* sequence was verified in 2006 to have an NH₂-terminal signal sequence, generating a protein of 525 a.a. in length. The *PDIA2* gene also yields two splice variants encoding two isoforms of the mature *PDIA2* protein, varying by three amino acids in length (isoform 2 does not contain a.a. 181 to 183). Multiple SNPs have been detected in the *PDIA2* gene, resulting in mutations at a.a. 39 (Pro \rightarrow Ser), 119 (Thr \rightarrow Arg), 185 (Glu \rightarrow Lys), 286 (Thr \rightarrow Met), 382 (Pro \rightarrow Ala), 388 (Arg \rightarrow Gln), and 502 (Pro \rightarrow Ser).

The *PDIA2* protein contains two **a** type (Cys-Gly-His-Cys and Cys-Thr-His-Cys active sites) and one **b** and one **b'** type domains; although redox active, *PDIA2* was found to be less effective than P4HB in assays for both reduction and oxidation of disulfides [30]. In addition to its role as a folding catalyst, *PDIA2* has been proposed to play a role in the production and secretion of digestive enzymes *in vivo* [31]. Evidence has also suggested a role for *PDIA2* in the binding and regulation of intracellular 17 β -estradiol levels, thus regulating estrogen synthesis [32]. *PDIA2* contains three sites for Asn-linked glycosylation, located at residues 127, 284, and 516 [30]. To date, no knockout studies have been conducted with *Pdia2*.

PDIA3

Originally identified as phospholipase C alpha, the *PDIA3* gene, located at Chr 15q15, encodes the 505-a.a. *PDIA3* protein (also commonly known as ERp57, ERp60,

P58) [33]. Like other PDI family members, PDIA3 contains a signal peptide, corresponding to the first 24 a.a., yielding a mature protein of 481 a.a. in length [34]. PDIA3 contains two a type (Cys-Gly-His-Cys and Cys-Gly-His-Cys active sites), one b and one b' type domain and an ER retention sequence, Gln-Glu-Asp-Leu [35,36]. PDIA3 expression has been detected in liver, placenta, lung, pancreas, kidney, heart, skeletal muscle, brain, and spermatozoa [33,37]. Presently, only one missense SNP has been reported for *PDIA3*, resulting in a mutation of Lys → Arg at a.a. 415.

The precise physiological role of PDIA3 has come under much scrutiny. Following the initial characterization of the protein *in vitro*, PDIA3 (termed ERp60 or P58 at the time) was identified as a cysteine protease despite little sequence homology with other heavily studied cysteine proteases [35]. In 1995, however, PDIA3 was determined to be redox active, showing the ability to reduce insulin disulfides. Bourdi et al. were also able to definitively prove that PDIA3 possessed no protease activity [36]. PDIA3 has also been shown to play a role in the correct folding of glycoproteins when in a complex containing both calnexin and calreticulin [38]. The physiological role of PDIA3 has also been investigated in rodent models, utilizing *Pdia3*(-/-) knockout mice. Although ubiquitous deletion of *Pdia3* was found to be embryolethal, heterozygous knockouts were generated, revealing multiple bone abnormalities, most notably in the femur [39]. Ablation of *Pdia3* was found to abolish signaling induced by 1,25-dihydroxyvitamin D₃, a crucial regulator of bone and cartilage development, by eliminating signaling through protein kinase C [39]. Additional knockout studies in murine B cells revealed a critical role for PDIA3 in the presentation of antigens by major histocompatibility complex I molecules [40]. Although additional work is needed, these studies suggest a wide array of physiological roles for PDIA3.

PDIA4

PDIA4 is located at Chr 7q35 and encodes the 645-a.a. PDIA4 protein (commonly known as ERp72). Although not much is known about the physiological role of *PDIA4*, studies indicate that this gene is induced following ER stress; these results have been found to be the result of a putative ERSE in the promoter region of the gene [41,42]. Like other PDI family members, PDIA4 contains an NH₂-terminal signal peptide of 21 a.a., yielding a mature protein of 625 a.a. [43]. PDIA4 contains five TRX domains, three a type (with all three active sites being comprised of Cys-Gly-His-Cys), one b and one b' type; PDIA4 also contains the ER retention sequence (Lys-Glu-Glu-Leu) [43,44]. Mutagenesis studies to the active-site cysteines revealed varying degrees of decreased enzymatic activity, whereas mutagenesis to multiple domains revealed a more pronounced reduction

in enzymatic activity of the protein [44]. PDIA4 also contains a string of highly acidic residues near the NH₂-terminus of the protein; while the precise role of these residues remains unknown, they have been proposed to play a role in regulation of Ca²⁺, yielding its rat homolog name calcium-binding protein-2 (CaBP2) [45]. PDIA4 is a fairly ubiquitously expressed protein, although less abundant than PDI, expression patterns are similar to those of PDIA3 [46]. One missense SNP has been reported in the *PDIA4* gene, resulting in a mutation located at residue 173 (Thr → Met). Studies analyzing *Pdia3* knockdown revealed partial functional restoration by the PDIA4 protein, although, to date, no *Pdia4* knockout mouse has been generated [47].

PDIA5

Although discovered in 1995, little is known about the precise role of the *PDIA5* gene. Located on Chr 3q21.1, *PDIA5* encodes the 519-a.a. PDIA5 (or PDI-related protein). PDIA5 contains four TRX domains (three a and one b-like domain), made up of active sites Cys-Ser-Met-Cys, Cys-Gly-His-Cys and Cys-Pro-His-Cys, a COOH-terminal ER retention sequence (Lys-Glu-Glu-Leu), and an unverified signal sequence comprising the first 21 a.a. [48]. Despite an additional Cys-X-X-Cys motif, Horibe et al. revealed that PDIA5 has significantly less enzymatic activity than that of P4HB [49]. The contributions of each Cys-X-X-Cys active site were also investigated, revealing varying degrees of altered activity following mutations to each, or multiple, active sites [49]. It was concluded that the second active site (Cys-Gly-His-Cys) was the most critical for isomerase activity and that all three motifs are not required for activity [49]. Much like P4HB, PDIA5 was also shown to exhibit chaperone-like activity by refolding denatured rhodanase, which does not contain any disulfide bonds [49]. PDIA5 mRNA has been detected in liver, kidney, lung, and brain—with the highest level of secretion being noted in the liver [48]. Although to date no ERSE has been identified, *PDIA5* has been shown to be moderately up-regulated following induction of the ER-stress response in cultured cells [48]. In a 2011 study by Carbone et al., a significant association was found between the SNP, rs11720822, and primary open-angle glaucoma in two separate populations [50]. No viable knockout mouse has been generated for *Pdia5*.

PDIA6

Much like *PDIA5*, little is known about the role of *PDIA6* both *in vitro* and *in vivo*. The *PDIA6* gene is located at Chr 2p25.1 and encodes the 440-a.a. PDIA6 protein (commonly reported as P5 or ERP5). PDIA6 contains an NH₂-terminal signal sequence of 19 a.a., three TRX domains (two a type and one b) consisting of

two Cys-Gly-His-Cys active sites, an Asp/Glu rich domain and a COOH-terminal Lys-Asp-Glu-Leu ER-retention sequence [51,52]. Recombinant PDIA6 demonstrates both isomerase and chaperone activities, although approximately 45% and 50% to 60% to that of P4HB, respectively [53]. Point mutations have also been conducted to the active-site cysteines, revealing that NH₂-terminal cysteines in each active site exhibit the majority of isomerase activity [53]. *PDIA6* contains an ERSE in its promoter region, which was recently validated *in vitro* utilizing over-expression of the ER-stress transcription factor, X-box protein-1 (XBP-1); *PDIA6* was found to be significantly increased, demonstrating inducibility by the unfolded protein response [54,55]. A complete expression profile for *PDIA6* has yet to be conducted; however, high levels of the protein were detected in platelets. Cell-surface expression of *PDIA6* was found to be necessary for the proper development and function of platelets, whereas inhibition of the protein using anti-*PDIA6* antibodies revealed inhibition of platelet aggregation [51]. *PDIA6* has also been shown to directly interact with GRP78 (or BiP) suggesting a role for *PDIA6* in the refolding of substrates that have been targeted to BiP [56]. Presently, one missense SNP has been identified (rs4807) resulting in a point mutation at a.a. 214 (Lys → Arg). At present, no viable *Pdia6* knockout mouse is available.

PDILT

Expression of the *PDILT* gene has been reported to be exclusively limited to the testis. *PDILT* is located on Chr 16p12.3, encoding the 584-a.a. *PDILT* protein [57]. Despite the presence of two a type TRX domains (with non-classical Ser-Lys-Gln-Ser and Ser-Lys-Lys-Cys motifs), *PDILT* does not exhibit the ability to oxidize or reduce disulfide bonds, although evidence has supported *PDILT* to engage in disulfide-bonded complexes *in vitro* [57,58]. *PDILT* contains a predicted NH₂-terminal signal peptide of 20 a.a. in length, a COOH-terminal ER-retention sequence (Lys-Glu-Glu-Leu) and is heavily glycosylated through nine potential Asn-linked glycosylation sites [57]. Much like P4HB, *PDILT* also interacts with the oxidoreductase ERO1 α in cultured cells, suggesting a role in the shuffling of electrons in the ER lumen [57]. No knockout mouse is currently available and considerable research is needed to fully elucidate the precise role of *PDILT*.

TXNDC5

TXNDC5 is located at Chr 6p24.3 and encodes the 432-a.a. endothelial PDI, *TXNDC5* (or EndoPDI) protein [59]. Despite its discovery in 2003, little research has been conducted on the role of *TXNDC5* *in vivo*. *TXNDC5* contains a signal peptide of 32 a.a., three a-type TRX domains all with Cys-Gly-His-Cys active sites, and a

COOH-terminal ER retention sequence, KDEL [59]. *TXNDC5* was originally identified in a screen for proteins highly expressed in endothelial cells, leading to its common name, EndoPDI. Subsequent studies revealed *TXNDC5* expression in a number of tissues with the highest expression being found in lymph nodes, stomach, pancreatic islets, and heart [59,60]. *TXNDC5* is induced under conditions of hypoxia, and loss of *TXNDC5* leads to an increase in apoptotic cell death in microvascular endothelial cells during hypoxia, but not normoxia [59]. Preliminary studies have also implicated a role for *TXNDC5* in diabetes, noting a decrease in the expression of the protein in pancreatic islets in animals with consistently elevated glucose levels [60]. In a 2010 study, Jeong et al. investigated the role of *TXNDC5* in development of the skin disorder vitiligo [61]. A total of 230 Korean patients with non-segmental vitiligo were investigated for SNPs in the *TXNDC5* gene; in total, seven SNPs were identified in the *TXNDC5* gene, three of which (rs1043784, rs7764128, and rs8643) demonstrated an association with the vitiligo phenotype [61]. Although relevant *in vivo* studies have been conducted on the role of *TXNDC5* no biochemical parameters have been evaluated, with regard to its role in disulfide bond oxidation and reduction. A viable knockout mouse has not been generated for the study of *Txndc5*.

The anterior gradient homolog genes

AGR2

The *AGR2* gene is located at Chr 7p21.3 and encodes the 175-a.a. anterior gradient protein 2 homolog (*AGR2*) [62,63]. The *AGR2* protein has an NH₂-terminal signal sequence of 20 a.a., one TRX domain (with active site Cys-Pro-His-Ser), and a COOH-terminal ER-retention sequence (Lys-Thr-Glu-Leu) [63]. Human *AGR2* was originally identified in estrogen receptor-positive MCF7 cells using suppression subtractive hybridization [64]. Expression of *AGR2* transcripts has been detected in lung, pancreas, trachea, stomach, colon, prostate, and small intestine. *AGR2* has also been investigated as a potential biomarker for hormone-responsive breast cancer in estrogen receptor- α -positive breast cancer cell lines [64].

Utilizing knockout studies in mice, the *Agr2* gene was found to result in the inability to produce mucin, leading in an increased susceptibility to experimentally induced colitis and intestinal disease [65]. Due to its role in disulfide bond generation, it was hypothesized that *AGR2* was responsible for the processing of MUC2, the major intestinal mucin. This protein contains >200 cysteine residues involved in various inter- and intra-protein disulfide bonds and has been found to directly associate with *AGR2* [65]. Following this report, Zheng et al. investigated *AGR2* and *AGR3* as potential candidate genes for inflammatory bowel disease in humans [66]. A cohort of

2,540 patients having either ulcerative colitis or Chron's disease was investigated for SNPs in *AGR2* and *AGR3*; in total, 30 SNPs were identified, 25 were located in the *AGR2* gene, while 5 were located in the *AGR3* gene [66]. The promoter region of the *AGR2* gene was also found to contain binding sites for hepatic nuclear factor-1, hepatocyte nuclear factor 3- α (FOXA1) and hepatocyte nuclear factor 3- β (FOXA2)—transcription factors that have been reported to play a role in the morphogenesis of goblet cell differentiation during formation. In summary, two total SNPs in the 5' promoter region of the *AGR2* gene were found to be associated with the risk haplotype of ulcerative colitis in two independent populations, providing further evidence for a role for *AGR2* in disease pathogenesis [66].

AGR3

The *AGR3* gene is located at Chr 7p21.1 and encodes the 166-a.a. anterior gradient protein-3 (*AGR3*) homolog [63]. *AGR3* transcripts have been detected in lung and pancreas and, resembling *AGR2*, *AGR3* has been reported to be co-expressed with estrogen receptor- α -containing breast cancer cell lines, suggesting it to be a marker for hormone-responsive breast cancer [63,64]. Unlike *AGR2*, however, little is known about the precise physiological role of *AGR3*. *AGR3* protein was originally identified as breast cancer membrane protein 11 (BCMP11), following a proteomic screen of membrane proteins in breast cancer cell lines, and was later named *AGR3* due to the high degree of sequence homology with *AGR2* (see Table 2) [67]. Like *AGR2*, *AGR3* contains one redox-active center, comprised of amino acids Cys-Gln-Tyr-Ser, an NH₂-terminal signal peptide composed of 21 a.a., and a COOH-terminal ER-retention sequence (Gln-Ser-Glu-Leu) [63]. Recently, an increase in *AGR3* expression was observed in serous border-line ovarian tumors and low-grade serous ovarian carcinoma [68]. Utilizing Kaplan-Meier survival curves, King et al. also established that patients with *AGR3*-expressing tumors survived significantly longer than those patients lacking *AGR3*-expressing tumors [68].

TXNDC12

TXNDC12, also known as *AGRI*, *TLP19*, and *ERP18/19*, contains conserved intron positions with respect to amino acid sequence with the *AGR2* and *AGR3* genes [63]. Persson et al. also reported that several of the individual exon lengths are identical (or altered with one codon) to *AGR2* and *AGR3* [63]. *TXNDC12* has been mapped to Chr 1p32.3 and contains seven exons spanning more than 35 kb [69]. *TXNDC12* is ubiquitously expressed in all tissues, with the highest expression being found in the liver and placenta. *TXNDC12* in the placenta was found to express an additional transcript of 1.2 kb which is

associated with two poly(A) addition signals in its 3'-UTR [69]. The *TXNDC12* protein contains 172 a.a. with 149 a.a. comprising the mature form of the protein (Ser²⁴ – Leu¹⁷²); *TXNDC12* has one active site comprised of Cys-Gly-Ala-Cys and an ER-retention sequence (Glu-Asp-Glu-Leu) [70]. Unlike the other members of the *AGR* subfamily, extensive work has been conducted on the biochemical and physiochemical actions of *TXNDC12*. The enzymatic activity of *TXNDC12* has been found to be limited strictly to disulfide bond generation and not reduction; these studies were also confirmed with the use of point mutations to the active-site cysteines (Cys-Gly-Ala-Cys), after which no detectable activity was found [70]. Chemical denaturation curves were also found to favor greater protein stability in the reduced form over the oxidized form, a property consistent with other PDI family members [70].

The CASQ genes

The *CASQ* genes (1 and 2) are interesting members of the *PDI* family, possessing no cysteine containing redox-active sites and therefore playing no role in the formation or reduction of disulfide bonds. As indicated, many of the *PDI* family proteins bind Ca²⁺ with relatively high capacity and low affinity [71]. Following studies on *CASQ*, Shin et al. found that the COOH-terminal Asp-rich domain played a major role in storage of Ca²⁺ through interaction with ryanodine receptor (RYR), a protein involved in Ca²⁺ release from the sarcoplasmic reticulum (SR) [72]. These proteins, therefore, possess unique functions relating to the *PDI* family of proteins, despite limited sequence homology.

CASQ1

The *CASQ1* gene is located on Chr 1q21 and encodes the 396-a.a. *CASQ1* protein [73]. Expression of the mature *CASQ1* protein is primarily limited to the SR of fast-twitch skeletal muscles [74]. Studies in rabbit have revealed *CASQ1* to be a high-capacity (40 to 50 Ca²⁺ per molecule of *CASQ1*), moderate-affinity (K_d = 1 mM) Ca²⁺-binding protein that does not contain an ER-retention sequence [74-76]. Although experimentally unverified, the first 34 a.a. of *CASQ1* encode the signal peptide, leaving the mature protein at 362 a.a. in length—which contains three TRX domains (two **b** and one **b'**) and a string of highly acidic residues from a.a. 353 – 396. Like other members of the *PDI* family, these Asp/Glu-rich stretches of a.a. are thought to be the primary binding regions for Ca²⁺; *CASQ1* plays a major role in Ca²⁺ flux through the regulation of Ca²⁺ channel activity and interaction with Ca²⁺ directly [72]. Systemic knockout studies in mice revealed hypersensitivity to heat and volatile anesthetics, along with a phenotypic resemblance to malignant hyperthermia [77]; these effects were found to be due to increased Ca²⁺ following an

increased exposure to heat [78]. *CASQ1* also contains one potential Asn-linked glycosylation site, found at a.a. 350.

SNPs in the *CASQ1* gene have been reported in cases of diabetes in both Old-Order Amish and Northern European Caucasians [79,80]. Out of 26 identified SNPs, SNP *CASQ-1404* (rs1186694) in the 5' flanking region was found to have a statistically significant association with type-2 diabetes in Northern European Caucasians [79]. In a similar study analyzing type-2 diabetes susceptibility in Old-Order Amish populations, SNPs rs2275703 and rs617698 were defined as the 'at-risk alleles' [80]. Although mechanistically these correlations have not been confirmed, previous work has identified a putative role for Ca^{2+} release from the SR into the cytosol in regulating glucose transporter-4 (GLUT4), a key enzyme in regulation of glucose transport by insulin [80,81].

CASQ2

The *CASQ2* gene is located on Chr. 1p13.3-p11, and likely is the result of gene duplication with *CASQ1*—given their chromosomal location. *CASQ2* encodes the 399-a.a. *CASQ2*, which shares 91% identity with its homologue, *CASQ1* (see Table 2) [82,83]. The *CASQ2* protein is expressed exclusively in cardiac muscle and serves as the major Ca^{2+} reservoir in the SR of myocardium; *CASQ2* also interacts with the RYR2 channel, regulating Ca^{2+} flux from the SR [76,84]. Much like *CASQ1*, *CASQ2* contains an unverified NH_2 -terminal signal peptide (19 a.a.), three **b**-like TRX domains (two **b** and one **b'**), no ER-retention sequence, and a string of highly acidic residues (a.a. 356 to 399); *CASQ2* does, however, contain one potential Asn-linked glycosylation site at a.a. 355.

The *CASQ2* gene has become a heavily researched target for diseases associated with arrhythmic heartbeats. In 2001, Lahat et al. investigated missense mutations found in the coding region of the gene [83]. One SNP (G → C) was found to result in an aspartic acid changed to a histidine at a.a. 307 of the mature protein, potentially altering the Ca^{2+} -chelating function of that region [83]. This SNP was found to be associated with Bedouin families from Israel susceptible to catecholamine-induced polymorphic ventricular tachycardia [83]. These studies were later confirmed in *Casq2*(-/-) knockout mice, revealing susceptibility to polymorphic ventricular tachycardia following exposure to catecholamines [85].

Thioredoxin-related transmembrane proteins

The thioredoxin-related transmembrane (*TMX*) genes are newly discovered members of the *PDI* gene family. To date, little is known about the precise function of these genes; however, all four members in the *PDI* family consist of one TRX domain, one transmembrane domain, and non-conventional ER-retention sequences.

TMX1

Discovered in 2001, *TMX1* is located at Chr 14q22.1 and encodes the 280-a.a. *TMX1* protein [86]. *TMX1* contains an NH_2 -terminal signal sequence of 26 a.a., one **a**-type TRX domain with active site Cys-Pro-Ala-Cys, one transmembrane domain (a.a. 183 to 203), and lacks an ER-retention sequence [86]. Expression of *TMX1* is fairly ubiquitous, with highest levels detected in liver, kidney, placenta, and lung [86]. Mature *TMX1* possesses the ability to both oxidize and reduce disulfide bonds, although chaperone-like activity has yet to be investigated [86,87]. The *TMX1* gene does not contain a putative ERSE in the promoter region, supporting evidence that *TMX1* is not induced by numerous ER-stress-inducing agents; over-expression of the protein in cultured cells, however, has revealed amelioration of both Brefeldin A-induced apoptosis and tunicamycin-induced ER stress [86,88]. No knockout mouse has been generated for the study of *Tmx1*.

TMX2

Perhaps the least researched gene in the family, *TMX2* was discovered in 2003 [89]. Located on Chr 11cen-q22.3, *TMX2* encodes the 296-a.a. *TMX2* protein [89]. Like *TMX1*, *TMX2* contains a COOH-terminal signal peptide (48 a.a.), one **a**-type TRX domain (Ser-Asn-Asp-Cys active site), one transmembrane domain (a.a. 104 – 126), and an ER-retention sequence comprised of Lys-Lys-Glu-Ile [89]. Expression of *TMX2* is fairly ubiquitous—with high levels detected in heart, brain, liver, kidney, and pancreas [89]. Although Meng et al. provided the initial characterization of the protein, sequence discrepancies have been found. The official NCBI sequence of the *TMX2* protein reveals a protein of 296 a.a and a second isoform, lacking an in-frame exon in the central coding region, encoding a protein of 258 a.a (isoform 2 differs between a.a. 84 to 122). Future studies are required to fully elucidate the role of *TMX2* *in vivo*. The availability of a *TMX2* knockout mouse has not been reported.

TMX3

The human *TMX3* gene is located at Chr 18q22 and encodes the 454-a.a. *TMX3* protein [90]. Following cleavage of the 24-a.a. signal peptide, the 430-a.a. mature protein consists of one **a**-type TRX domain, with the active site being comprised of Cys-Gly-His-Cys, a transmembrane domain (located at a.a. 375 to 397), and the ER-retention sequence (Lys-Lys-Lys-Asp) [90]. Uncharacteristic to most *PDI* family members, *TMX3* contains a luminal domain with weak sequence similar to that of the *CASQ* proteins [90]. Although no research has been conducted on the role of this domain in activity of the protein, it has been postulated to regulate Ca^{2+} in a manner similar to that of other *CASQ* proteins. NCBI

reports two isoforms for *TMX3* (one encoding a 195-a.a. protein), though experimentally this has not been validated [90]. *TMX3* has been detected in brain, testis, lung, skin, kidney, uterus, bone, stomach, liver, prostate, placenta, eye, and muscle, with highest levels detected in heart and skeletal muscle [90]. *TMX3* contains two sites of Asn-linked glycosylation (a.a. 258 and 313), which have been validated *in vitro* and is not induced under conditions of ER stress [90]. Although far less efficient than P4HB, *TMX3* does display the ability to oxidize disulfide bonds; this is likely due to the presence of only one Cys-Gly-His-Cys active site [90].

Although no knockout mouse has been generated for *TMX3*, studies have been conducted in mice, targeting *TMX3* transcripts using morpholinos [91]. Investigating the mechanisms behind microphthalmia in humans, a genetic disease associated with retarded growth of the eye, a 2.7-Mb deletion was found at Chr 18q22.1, leading to deletion of the *TMX3* gene [91]. Studies in mice using a targeted approach to delete the *TMX3* gene revealed a similar phenotype, which was rescued following injection of human *TMX3* mRNA [91]. Sequencing of 162 patients with anophthalmia or microphthalmia revealed two missense mutations, leading to the missense SNPs, R39N, and D108N [91]. Future studies are required to fully elucidate the precise role of *TMX3*, although preliminary studies reveal exciting areas for research.

TMX4

The *TMX4* gene is located at Chr 20p12 and encodes the 349-a.a. *TMX4* protein [92]. *TMX4* consists of an NH₂-terminal signal sequence (23 a.a.), one α -type TRX domain (a.a. 39 – 136 with the active site comprised of Cys-Pro-Ser-Cys), a transmembrane domain (a.a. 188 to 210), a string of highly acidic a.a. (224 to 334), and (like the other *TMX* proteins) lacks an ER-retention sequence [93]. *TMX4* is ubiquitously expressed, with highest levels detected in heart [92]. Preliminary studies show that *TMX4* is not induced following conditions of ER stress and does not contain a putative ERSE in the promoter region of the gene [92]. *TMX4* contains one site of Asn-linked glycosylation and two sites of Ser phosphorylation, which have all been experimentally validated [93]. Enzymatic activity was confirmed by observing reduction of insulin disulfides; a dominant-negative mutant, with the active-site cysteines mutated to serine, displayed no enzymatic activity [92]. No knockout mouse has been generated and substantial work will be required to understand the role of *TMX4* *in vivo*.

Conclusions

The PDI family of proteins consists of 21 members varying greatly in enzymatic activity, domain architecture, and tissue specificity. Although the predominant role of the PDI proteins is the regulation of protein folding

in vivo—through the oxidation, reduction and isomerization of disulfide bonds—these proteins have also been shown to regulate calcium homeostasis in the ER and induction of the unfolded protein response (UPR). Since its discovery over 40 years ago, PDI has become one of the most highly studied proteins and, despite these advances, extensive research is still needed to fully understand the role of PDI *in vivo*. The more recently characterized *TMX* genes have displayed promise in novel therapeutics, ranging from disorders of the eye to regulation of the ER-stress response. Many of the genes in the *PDI* family contain a putative ERSE sequence in the promoter region of the gene, suggesting a role in the UPR. Further research on the role of these proteins in the UPR is required before effective therapeutics can be generated for a plethora of disease states associated with the ER-stress response. The 21 members of the PDI family of proteins encompass many physiological responses, and these proteins will likely provide compelling avenues for future research.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

JJG carried out the sequence alignment and drafted the manuscript. DRP designed and funded the study. Both authors read and approved the final manuscript.

Acknowledgments

Studies were supported by the National Institutes of Health/National Institutes of Alcoholism and Alcohol Abuse under grant numbers R37AA09300 (DRP), R01DK074487-01 (DRP) and F31AA018606-01 (JJG).

Author details

¹Department of Pharmacology, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, USA. ²Molecular Toxicology and Environmental Health Sciences Program, Department of Pharmacy and Pharmaceutical Sciences, University of Colorado Anschutz Medical Campus, Aurora, CO 80045, USA.

Received: 18 October 2011 Accepted: 14 May 2012

Published: 5 July 2012

References

1. Malhotra JD, Kaufman RJ: **The endoplasmic reticulum and the unfolded protein response.** *Semin Cell Dev Biol* 2007, **18**:716–731.
2. Appenzeller-Herzog C, Ellgaard L: **The human PDI family: versatility packed into a single fold.** *Biochim Biophys Acta* 2008, **1783**:535–548.
3. Hatahet F, Ruddock LW: **Protein disulfide isomerase: a critical evaluation of its function in disulfide bond formation.** *Antioxid Redox Signal* 2009, **11**:2807–2850.
4. Ellgaard L, Ruddock LW: **The human protein disulphide isomerase family: substrate interactions and functional properties.** *EMBO Rep* 2005, **6**:28–32.
5. Jacquot JP, Gelhaye E, Rouhier N, Corbier C, Didierjean C, Aubry A: **Thioredoxins and related proteins in photosynthetic organisms: molecular basis for thiol dependent regulation.** *Biochem Pharmacol* 2002, **64**:1065–1069.
6. Turano C, Coppari S, Altieri F, Ferraro A: **Proteins of the PDI family: unpredicted non-ER locations and functions.** *J Cell Physiol* 2002, **193**: 154–163.
7. Rapoport TA: **Protein transport across the endoplasmic reticulum membrane: facts, models, mysteries.** *FASEB J* 1991, **5**:2792–2798.
8. Houston NL, Fan C, Xiang JQ, Schulze JM, Jung R, Boston RS: **Phylogenetic analyses identify 10 classes of the protein disulfide isomerase family in**

- plants, including single-domain protein disulfide isomerase-related proteins. *Plant Physiol* 2005, **137**:762–778.
9. McArthur AG, Knodler LA, Silberman JD, Davids BJ, Gillin Fd, Sogin ML: **The evolutionary origins of eukaryotic protein disulfide isomerase domains: new evidence from the Amitochondriate protist *Giardia lamblia*.** *Mol Biol Evol* 2001, **18**:1455–1463.
 10. Gu SH, Chen JZ, Ying K, Wang S, Jin W, Qian J, Zhao EP, Xie Y, Mao YM: **Cloning and identification of a novel cDNA which encodes a putative protein with a DnaJ domain and a thioredoxin active motif, human macrothioredoxin.** *Biochem Genet* 2003, **41**:245–253.
 11. Cunnea PM, Miranda-Vizuete A, Bertoli G, Simmen T, Damdimopoulos AE, Hermann S, Leinonen S, Huikko MP, Gustafsson JA, Sitia R, Spyrou G: **ERdj5, an endoplasmic reticulum (ER)-resident protein containing DnaJ and thioredoxin domains, is expressed in secretory cells or following ER stress.** *J Biol Chem* 2003, **278**:1059–1066.
 12. Ushioda R, Hoseki J, Araki K, Jansen G, Thomas DY, Nagata K: **ERdj5 is required as a disulfide reductase for degradation of misfolded proteins in the ER.** *Science* 2008, **321**:569–572.
 13. Alanen HI, Williamson RA, Howard MJ, Hatahet FS, Salo KE, Kaupilla A, Kellokumpu S, Ruddock LW: **ERP27, a new non-catalytic endoplasmic reticulum-located human protein disulfide isomerase family member, interacts with ERp57.** *J Biol Chem* 2006, **281**:33727–33738.
 14. Sargsyan E, Baryshev M, Backlund M, Sharipo A, Mkrtchian S: **Genomic organization and promoter characterization of the gene encoding a putative endoplasmic reticulum chaperone, ERp29.** *Gene* 2002, **285**:127–139.
 15. Ferrari DM, Van Nguyen P, Kratzin HD, Soling HD: **ERp28, a human endoplasmic-reticulum-luminal protein, is a member of the protein disulfide isomerase family but lacks a CXXC thioredoxin-box motif.** *Eur J Biochem* 1998, **255**:570–579.
 16. Mkrtchian S, Fang C, Hellman U, Ingelman-Sundberg M: **A stress-inducible rat liver endoplasmic reticulum protein, ERp29.** *Eur J Biochem* 1998, **251**:304–313.
 17. Mkrtchian S, Baryshev M, Sargsyan E, Chatzistamou I, Volakaki AA, Chaviaras N, Pafiti A, Triantafyllou A, Kiaris H: **ERp29, an endoplasmic reticulum secretion factor is involved in the growth of breast tumor xenografts.** *Mol Carcinog* 2008, **47**:886–892.
 18. Anelli T, Alessio M, Mezghrani A, Simmen T, Talamo F, Bachi A, Sitia R: **ERp44, a novel endoplasmic reticulum folding assistant of the thioredoxin family.** *EMBO J* 2002, **21**:835–844.
 19. Wang ZV, Schraw TD, Kim JY, Khan T, Rajala MW, Follenzi A, Scherer PE: **Secretion of the adipocyte-specific secretory protein adiponectin critically depends on thiol-mediated protein retention.** *Mol Cell Biol* 2007, **27**:3716–3731.
 20. Tasanen K, Parkkonen T, Chow LT, Kivirikko KI, Pihlajaniemi T: **Characterization of the human gene for a polypeptide that acts both as the beta subunit of prolyl 4-hydroxylase and as protein disulfide isomerase.** *J Biol Chem* 1988, **263**:16218–16224.
 21. Pihlajaniemi T, Helaakoski T, Tasanen K, Myllyla R, Huhtala ML, Koivu J, Kivirikko KI: **Molecular cloning of the beta-subunit of human prolyl 4-hydroxylase. This subunit and protein disulfide isomerase are products of the same gene.** *EMBO J* 1987, **6**:643–649.
 22. Pajunen L, Jones TA, Goddard A, Sheer D: **Regional assignment of the human gene coding for a multifunctional polypeptide (P4HB) acting as the beta-subunit of prolyl 4-hydroxylase and the enzyme protein disulfide isomerase to 17q25.** *Cytogenet Cell Genet* 1991, **56**:165–168.
 23. Tasanen K, Oikarinen J, Kivirikko KI, Pihlajaniemi T: **Promoter of the gene for the multifunctional protein disulfide isomerase polypeptide. Functional significance of the six CCAAT boxes and other promoter elements.** *J Biol Chem* 1992, **267**:11513–11519.
 24. Ferrari DM, Soling HD: **The protein disulfide-isomerase family: unravelling a string of folds.** *Biochem J* 1999, **339**(Pt 1):1–10.
 25. Freedman RB, Brockway BE, Lambert N: **Protein disulfide-isomerase and the formation of native disulfide bonds.** *Biochem Soc Trans* 1984, **12**:929–932.
 26. Freedman RB, Hirst TR, Tuite MF: **Protein disulfide isomerase: building bridges in protein folding.** *Trends Biochem Sci* 1994, **19**:331–336.
 27. Song JL, Wang CC: **Chaperone-like activity of protein disulfide-isomerase in the refolding of rhodanese.** *Eur J Biochem* 1995, **231**:312–316.
 28. Wang CC, Tsou CL: **Protein disulfide isomerase is both an enzyme and a chaperone.** *FASEB J* 1993, **7**:1515–1517.
 29. Wetterau JR, Combs KA, Spinner SN, Joiner BJ: **Protein disulfide isomerase is a component of the microsomal triglyceride transfer protein complex.** *J Biol Chem* 1990, **265**:9800–9807.
 30. Desiva MG, Lu J, Donadel G, Modi WS, Xie H, Notkins AL, Lan MS: **Characterization and chromosomal localization of a new protein disulfide isomerase, PDlp, highly expressed in human pancreas.** *DNA Cell Biol* 1996, **15**:9–16.
 31. Klappa P, Stromer T, Zimmermann R, Ruddock LW, Freedman RB: **A pancreas-specific glycosylated protein disulfide-isomerase binds to misfolded proteins and peptides with an interaction inhibited by oestrogens.** *Eur J Biochem* 1998, **254**:63–69.
 32. Fu XM, Zhu BT: **Human pancreas-specific protein disulfide isomerase homolog (PDlp) is an intracellular estrogen-binding protein that modulates estrogen levels and actions in target cells.** *J Steroid Biochem Mol Biol* 2009, **115**:20–29.
 33. Koivunen P, Horelli-Kuitunen N, Helaakoski T, Karvonen P, Jaakkola M, Palotie A, Kivirikko KI: **Structures of the human gene for the protein disulfide isomerase-related polypeptide ERp60 and a processed gene and assignment of these genes to 15q15 and 1q21.** *Genomics* 1997, **42**:397–404.
 34. Charnock-Jones DS, Day K, Smith SK: **Cloning, expression and genomic organization of human placental protein disulfide isomerase (previously identified as phospholipase C alpha).** *Int J Biochem Cell Biol* 1996, **28**:81–89.
 35. Urade R, Oda T, Ito H, Moriyama T, Utsumi S, Kito M: **Functions of characteristic Cys-Gly-His-Cys (CGHC) and Gln-Glu-Asp-Leu (QEDL) motifs of microsomal ER-60 protease.** *J Biochem* 1997, **122**:834–842.
 36. Bourdi M, Demady D, Martin JL, Jabbour SK, Martin BM, George JW, Pohl LR: **cDNA cloning and baculovirus expression of the human liver endoplasmic reticulum P58: characterization as a protein disulfide isomerase isoform, but not as a protease or a carnitine acyltransferase.** *Arch Biochem Biophys* 1995, **323**:397–403.
 37. Kameshwari DB, Bhande S, Sundaram CS, Kota V, Siva AB, Shivaji S: **Glucose-regulated protein precursor (GRP78) and tumor rejection antigen (GP96) are unique to hamster caput epididymal spermatozoa.** *Asian J Androl* 2010, **12**:344–355.
 38. Oliver JD, van der Wal FJ, Bulleid NJ, High S: **Interaction of the thiol-dependent reductase ERp57 with nascent glycoproteins.** *Science* 1997, **275**:86–88.
 39. Wang Y, Chen J, Lee CS, Nizkorodov A, Riemenschneider K, Martin D, Hyzy S, Schwartz Z, Boyan BD: **Disruption of Pdia3 gene results in bone abnormality and affects 1alpha,25-dihydroxy-vitamin D3-induced rapid activation of PKC.** *J Steroid Biochem Mol Biol* 2010, **121**:257–260.
 40. Garbi N, Tanaka S, Momburg F, Hammerling GJ: **Impaired assembly of the major histocompatibility complex class I peptide-loading complex in mice deficient in the oxidoreductase ERp57.** *Nat Immunol* 2006, **7**:93–102.
 41. Paschen W, Gissel C, Linden T, Douthell J: **Erp72 expression activated by transient cerebral ischemia or disturbance of neuronal endoplasmic reticulum calcium stores.** *Metab Brain Dis* 1998, **13**:55–68.
 42. Roy B, Lee AS: **The mammalian endoplasmic reticulum stress response element consists of an evolutionarily conserved tripartite structure and interacts with a novel stress-inducible complex.** *Nucleic Acids Res* 1999, **27**:1437–1443.
 43. Mazzarella RA, Srinivasan M, Haugejorden SM, Green M: **ERp72, an abundant luminal endoplasmic reticulum protein, contains three copies of the active site sequences of protein disulfide isomerase.** *J Biol Chem* 1990, **265**:1094–1101.
 44. Satoh M, Shimada A, Keino H, Kashiwai A, Nagai N, Saga S, Hosokawa M: **Functional characterization of 3 thioredoxin homology domains of ERp72.** *Cell Stress Chaperones* 2005, **10**:278–284.
 45. Nigam SK, Goldberg AL, Ho S, Rohde MF, Bush KT, Sherman MY: **A set of endoplasmic reticulum proteins possessing properties of molecular chaperones includes Ca(2+)-binding proteins and members of the thioredoxin superfamily.** *J Biol Chem* 1994, **269**:1744–1749.
 46. Marcus N, Shaffer D, Farrar P, Green M: **Tissue distribution of three members of the murine protein disulfide isomerase (PDI) family.** *Biochim Biophys Acta* 1996, **1309**:253–260.
 47. Solda T, Garbi N, Hammerling GJ, Molinari M: **Consequences of ERp57 deletion on oxidative folding of obligate and facultative clients of the calnexin cycle.** *J Biol Chem* 2006, **281**:6219–6226.
 48. Hayano T, Kikuchi M: **Molecular cloning of the cDNA encoding a novel protein disulfide isomerase-related protein (PDIR).** *FEBS Lett* 1995, **372**:210–214.

49. Horibe T, Gomi M, Iguchi D, Ito H, Kitamura Y, Masuoka T, Tsujimoto I, Kimura T, Kikuchi M: **Different contributions of the three CXXC motifs of human protein-disulfide isomerase-related protein to isomerase activity and oxidative refolding.** *J Biol Chem* 2004, **279**:4604–4611.
50. Carbone MA, Chen Y, Hughes GA, Weinreb RN, Zabriskie NA, Zhang K, Anholt RRH: **Genes of the unfolded protein response pathway harbor risk alleles for primary open angle glaucoma.** *PLoS One* 2011, **6**:e20649.
51. Jordan PA, Stevens JM, Hubbard GP, Barrett NE, Sage T, Authi KS, Gibbins JM: **A role for the thiol isomerase protein ERP5 in platelet function.** *Blood* 2005, **105**:1500–1507.
52. Hayano T, Kikuchi M: **Cloning and sequencing of the cDNA encoding human P5.** *Gene* 1995, **164**:377–378.
53. Kikuchi M, Doi E, Tsujimoto I, Horibe T, Tsujimoto Y: **Functional analysis of human P5, a protein disulfide isomerase homologue.** *J Biochem* 2002, **132**:451–455.
54. Belmont PJ, Chen WJ, San Pedro MN, Therauf DJ, Gellings Lowe N, Gude N, Hilton B, Wolkowicz R, Sussman MA, Glembotski CC: **Roles for endoplasmic reticulum-associated degradation and the novel endoplasmic reticulum stress response gene Derlin-3 in the ischemic heart.** *Circ Res* 2009, **106**:307–316.
55. Lee AH, Iwakoshi NN, Glimcher LH: **XBP-1 regulates a subset of endoplasmic reticulum resident chaperone genes in the unfolded protein response.** *Mol Cell Biol* 2003, **23**:7448–7459.
56. Jessop CE, Watkins RH, Simmons JJ, Tasab M, Bulleid NJ: **Protein disulfide isomerase family members show distinct substrate specificity: P5 is targeted to BiP client proteins.** *J Cell Sci* 2009, **122**:4287–4295.
57. van Lith M, Hartigan N, Hatch J, Benham AM: **PDILT, a divergent testis-specific protein disulfide isomerase with a non-classical SXXC motif that engages in disulfide-dependent interactions in the endoplasmic reticulum.** *J Biol Chem* 2005, **280**:1376–1383.
58. van Lith M, Karala AR, Bown D, Gatehouse JA, Ruddock LW, Saunders PT, Benham AM: **A developmentally regulated chaperone complex for the endoplasmic reticulum of male haploid germ cells.** *Mol Biol Cell* 2007, **18**:2795–2804.
59. Sullivan DC, Huminiecki L, Moore JW, Boyle JJ, Poulosom R, Creamer D, Barker J, Bicknell R: **EndoPDI, a novel protein-disulfide isomerase-like protein that is preferentially expressed in endothelial cells acts as a stress survival factor.** *J Biol Chem* 2003, **278**:47079–47088.
60. Alberti A, Karamessinis P, Peroulis M, Kypreou K, Kavvas P, Pagakis S, Politis PK, Charonis A: **ERP46 is reduced by high glucose and regulates insulin content in pancreatic beta-cells.** *Am J Physiol Endocrinol Metab* 2009, **297**:E812–E821.
61. Jeong KH, Shin MK, Uhm YK, Kim HJ, Chung JH, Lee MH: **Association of TXNDC5 gene polymorphisms and susceptibility to nonsegmental vitiligo in the Korean population.** *Br J Dermatol* 2010, **162**:759–764.
62. Petek E, Windpassinger M, Egger H, Krosel PM, Wagner K: **Localization of the human anterior gradient-2 gene (AGR2) to chromosome band 7p21.3 by radiation hybrid mapping and fluorescence in situ hybridisation.** *Cytogenet Cell Genet* 2000, **89**:141–142.
63. Persson S, Rosenquist M, Knoblach B, Khosravi-Far R, Sommarin M, Michalak M: **Diversity of the protein disulfide isomerase family: identification of breast tumor induced Hag2 and Hag3 as novel members of the protein family.** *Mol Phylogenet Evol* 2005, **36**:734–740.
64. Thompson DA, Weigel RJ: **hAG-2, the human homologue of the *Xenopus laevis* cement gland gene XAG-2, is coexpressed with estrogen receptor in breast cancer cell lines.** *Biochem Biophys Res Commun* 1998, **251**:111–116.
65. Park SW, Zhen G, Verhaeghe C, Nakagami Y, Nguyenvu LT, Barczak AJ, Killeen N, Erle D: **The protein disulfide isomerase AGR2 is essential for production of intestinal mucus.** *Proc Natl Acad Sci U S A* 2009, **106**:6950–6955.
66. Zheng W, Rosenstiel P, Huse K, Sina C, Valentonyte R, Mah N, Zeitlmann L, Grosse J, Ruf N, Nürnberg P, Costello CM, Onnie C, Mathew C, Platzer M, Schreiber S, Hampe J: **Evaluation of AGR2 and AGR3 as candidate genes for inflammatory bowel disease.** *Genes Immun* 2006, **7**:11–18.
67. Adam PJ, Boyd R, Tyson KL, Fletcher GC, Stamps A, Hudson L, Poyser HR, Redpath N, Griffiths M, Steers C, Harris AL, Patel S, Berry J, Loader JA, Townsend RR, Daviet L, Legrain P, Parekh R, Terrett JA: **Comprehensive proteomic analysis of breast cancer cell membranes reveals unique proteins with potential roles in clinical cancer.** *J Biol Chem* 2003, **278**:6482–6489.
68. King ER, Tung CS, Tsang YT, Zu Z, Lok GT, Deavers MT, Malpica A, Wolf JK, Lu KH, Birrer MJ, Mok SC, Gershenson DM, Wong KK: **The anterior gradient homolog 3 (AGR3) gene is associated with differentiation and survival in ovarian cancer.** *Am J Surg Pathol* 2011, **35**:904–912.
69. Liu F, Rong YP, Zeng LC, Zhang X, Han ZG: **Isolation and characterization of a novel human thioredoxin-like gene hTLP19 encoding a secretory protein.** *Gene* 2003, **315**:71–78.
70. Alanen HI, Williamson RA, Howard MJ, Lappi AK, Jäntti HP, Rautio SM, Kellokumpu S, Ruddock LW: **Functional characterization of ERp18, a new endoplasmic reticulum-located thioredoxin superfamily member.** *J Biol Chem* 2003, **278**:28912–28920.
71. Coe H, Michalak M: **Calcium binding chaperones of the endoplasmic reticulum.** *Gen Physiol Biophys* 2009, **28**:F96–F103. Spec No Focus.
72. Shin DW, Pan Z, Kim EK, Lee JM, Bhat MB, Parness J, Kim DH, Ma J: **A retrograde signal from calsequestrin for the regulation of store-operated Ca²⁺ entry in skeletal muscle.** *J Biol Chem* 2003, **278**:3286–3292.
73. Fujii J, Willard HF, MacLennan DH: **Characterization and localization to human chromosome 1 of human fast-twitch skeletal muscle calsequestrin gene.** *Somat Cell Mol Genet* 1990, **16**:185–189.
74. MacLennan DH, Wong PT: **Isolation of a calcium-sequestering protein from sarcoplasmic reticulum.** *Proc Natl Acad Sci U S A* 1971, **68**:1231–1235.
75. Fliegel L, Ohnishi M, Carpenter MR, Khanna VK, Reithmeier RA, MacLennan DH: **Amino acid sequence of rabbit fast-twitch skeletal muscle calsequestrin deduced from cDNA and peptide sequencing.** *Proc Natl Acad Sci U S A* 1987, **84**:1167–1171.
76. Yano K, Zarain-Herzberg A: **Sarcoplasmic reticulum calsequestrins: structural and functional properties.** *Mol Cell Biochem* 1994, **135**:61–70.
77. Dainese M, Quarta M, Lyfenko AD, Paolini C, Canato M, Reggiani C, Dirksen RT, Protasi F: **Anesthetic- and heat-induced sudden death in calsequestrin-1-knockout mice.** *FASEB J* 2009, **23**:1710–1720.
78. Zhao X, Min CK, Ko JK, Parness J, do Kim H, Weisleder N, Ma J: **Increased store-operated Ca²⁺ entry in skeletal muscle with reduced calsequestrin-1 expression.** *Biophys J* 2010, **99**:1556–1564.
79. Das SK, Chu W, Zhang Z, Hasstedt SJ, Elbein SC: **Calsquestrin 1 (CASQ1) gene polymorphisms under chromosome 1q21 linkage peak are associated with type 2 diabetes in Northern European Caucasians.** *Diabetes* 2004, **53**:3300–3306.
80. Fu M, Damcott CM, Sabra M, Pollin TI, Ott SH, Wang J, Garant MJ, O'Connell JR, Mitchell BD, Shuldiner AR: **Polymorphism in the calsequestrin 1 (CASQ1) gene on chromosome 1q21 is associated with type 2 diabetes in the old order Amish.** *Diabetes* 2004, **53**:3292–3299.
81. Youn JH, Gulve EA, Holloszy JO: **Calcium stimulates glucose transport in skeletal muscle by a pathway independent of contraction.** *Am J Physiol* 1991, **260**:C555–C561.
82. Otsu K, Fujii J, Periasamy M, Difilippantonio M, Uppender M, Ward DC, MacLennan DH: **Chromosome mapping of five human cardiac and skeletal muscle sarcoplasmic reticulum protein genes.** *Genomics* 1993, **17**:507–509.
83. Lahat H, Pras E, Olender T, Avidan N, Ben-Asher E, Man O, Levy-Nissenbaum E, Khoury A, Lorber A, Goldman B, Lancet D, Eldar M: **A missense mutation in a highly conserved region of CASQ2 is associated with autosomal recessive catecholamine-induced polymorphic ventricular tachycardia in Bedouin families from Israel.** *Am J Hum Genet* 2001, **69**:1378–1384.
84. Gyorke I, Hester N, Jones LR, Gyorke S: **The role of calsequestrin, triadin, and junctin in conferring cardiac ryanodine receptor responsiveness to luminal calcium.** *Biophys J* 2004, **86**:2121–2128.
85. Knollmann BC, Chopra N, Hlaing T, Akin B, Yang T, Ettensohn K, Knollmann BE, Horton KD, Weissman NJ, Holinstat I, Zhang W, Roden DM, Jones LR, Franzini-Armstrong C, Pfeifer K: **Casq2 deletion causes sarcoplasmic reticulum volume increase, premature Ca²⁺ release, and catecholaminergic polymorphic ventricular tachycardia.** *J Clin Invest* 2006, **116**:2510–2520.
86. Matsuo Y, Akiyama N, Nakamura H, Yodoi J, Noda M, Kizaka-Kondoh S: **Identification of a novel thioredoxin-related transmembrane protein.** *J Biol Chem* 2001, **276**:10032–10038.
87. Matsuo Y, Nishinaka Y, Suzuki S, Kojima M, Kizaka-Kondoh S, Kondo N, Son A, Sakakura-Nishiyama J, Yamaguchi Y, Masutani H, Ishii Y, Yodoi J: **TMX, a human transmembrane oxidoreductase of the thioredoxin family: the possible role in disulfide-linked protein folding in the endoplasmic reticulum.** *Arch Biochem Biophys* 2004, **423**:81–87.
88. Matsuo Y, Masutani H, Son A, Kizaka-Kondoh S, Yodoi J: **Physical and functional interaction of transmembrane thioredoxin-related protein with major histocompatibility complex class I heavy chain: redox-based protein quality control and its potential relevance to immune responses.** *Mol Biol Cell* 2009, **20**:4552–4562.
89. Meng X, Zhang C, Chen J, Peng S, Cao Y, Ying K, Xie Y, Mao Y: **Cloning and identification of a novel cDNA coding thioredoxin-related transmembrane protein 2.** *Biochem Genet* 2003, **41**:99–106.

90. Haugstetter J, Blicher T, Ellgaard L: **Identification and characterization of a novel thioredoxin-related transmembrane protein of the endoplasmic reticulum.** *J Biol Chem* 2005, **280**:8371–8380.
91. Chao R, Nevin L, Agarwal P, Riemer J, Bai X, Delaney A, Akana M, JimenezLopez N, Bardakjian T, Schneider A, Chassaing N, Schorderet DF, FitzPatrick D, Kwok P, Ellgaard L, Gould DB, Zhang Y, Malicki J, Baier H, Slavotinek A: **A male with unilateral microphthalmia reveals a role for TMX3 in eye development.** *PLoS One* 2010, **5**:e10565.
92. Sugiura Y, Araki K, Iemura S, Natsume T, Jun Hoseki J, Nagata K: **Novel thioredoxin-related transmembrane protein TMX4 has reductase activity.** *J Biol Chem* 2010, **285**:7135–7142.
93. Roth D, Lynes E, Riemer J, Hansen HG, Althaus N, Simmen T, Ellgaard L: **A di-arginine motif contributes to the ER localization of the type I transmembrane ER oxidoreductase TMX4.** *Biochem J* 2010, **425**:195–205.

doi:10.1186/1479-7364-6-6

Cite this article as: Galligan and Petersen: The human protein disulfide isomerase gene family. *Human Genomics* 2012 **6**:6.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

