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Review

The Impact of Exercise on Redox Equilibrium in Cardiovascular Diseases

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Abstract: Cardiovascular diseases constitute the most important public health problem in the world. They are characterized by inflammation and oxidative stress in the heart and blood. Physical activity is recognized as one of the best ways to prevent these diseases, and it has already been applied in treatment. Physical exercise, both aerobic and anaerobic and single and multiple, is linked to the oxidant–antioxidant imbalance; however, this leads to positive adaptive changes in, among others, the increase in antioxidant capacity. The goal of the paper was to discuss the issue of redox equilibrium in the human organism in the course of cardiovascular diseases to systemize updated knowledge in the context of exercise impacts on the organism. Antioxidant supplementation is also an important issue since antioxidant supplements still have great potential regarding their use as drugs in these diseases.

Keywords: redox balance; cardiovascular diseases; physical activity; cardiovascular risk factors; biomarkers



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1. Introduction

Cardiovascular diseases (CVDs) are a group of disorders of the heart and blood vessels. They include coronary heart disease (CHD), cerebrovascular disease, peripheral arterial disease (PAD), rheumatic heart disease (RHD), congenital heart defects (CHDs), deep vein thrombosis (DVT) and pulmonary embolism (PE). Although CVDs are well recognized and, for this reason, effectively treated, they cause the highest number of deaths annually. According to the World Health Organization (WHO), 32% of all global deaths in 2019 were caused by CVDs, mainly heart attack and stroke. Most CVDs can be prevented by addressing behavioral risk factors such as tobacco use, unhealthy diet, physical inactivity and harmful use of alcohol [1]. Physical inactivity, which is a sedentary lifestyle, is a particularly important health problem of modern civilization, as it is the main cause of excess weight and obesity, which are major contributors of morbidity and mortality [2].

One of the most crucial pathogenic factors in the case of CVDs is oxidative stress, which is the disturbed oxidant–antioxidant balance towards an excess of oxidants—reactive oxygen species (ROS) and reactive nitrogen species (RNS). Crucial here are primary endogenous ROS and RNS, superoxide anion radical (${}^{\bullet}O_2^{-}$) and nitric oxide (${}^{\bullet}NO$) since they are the source of subsequent ROS and RNS. The first one is the product of the reduction of the oxygen molecule by one electron (instead of four) in the mitochondrial transport chain and upregulated activities of oxidoreductases (xanthine oxidoreductase, XOR and nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, NOX). The second one, in turn, is most often produced by endothelial nitric oxide synthase (eNOS). Oxidative stress causes damage (oxidative modification, nitration) to proteins, amino acids, lipids and nucleic acids, which may contribute to inflammation [3] (Figure 1).

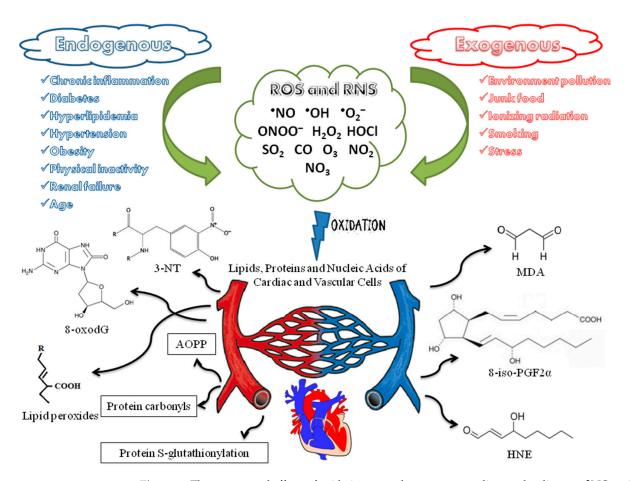


Figure 1. The sources and effects of oxidative stress that promote cardiovascular diseases. $^{\bullet}$ NO—nitric oxide; $^{\bullet}$ OH—hydroxyl radical; $^{\bullet}$ O2 $^{-}$ —superoxide anion radical; ONOO $^{-}$ —peroxynitrite; H₂O₂—hydrogen peroxide; HOCl—hypochlorous acid; SO₂—sulfur dioxide; CO—carbon monoxide; O₃—ozone; NO_{2/3}—nitrogen di/trioxide; 3-NT—3-nitrotyrosine; 8-oxodG—8-oxo-7,8-dihydro-2'-deoxyguanosine; AOPP—advanced oxidation protein products; MDA—malondialdehyde; 8-iso-PGF2 α —8-iso-prostaglandin F2 α ; HNE—4-hydroxy-2-nonenal.

A great tool for the prevention and treatment of CVDs may be physical activity (PA) on account of the fact that it improves the antioxidant response against ROS and RNS. Leaving aside many aspects of exercise-induced impacts on the organism, the impact of PA on redox balance seems especially important in the context of CVDs [4]. During physical exercise, oxygen uptake may be even 20 times greater versus the uptake at rest, whereas the oxygen consumption in muscles may increase even 200 times. Thus, the production of ROS increases significantly (the abovementioned incomplete reduction of O2 in mitochondria). This is also strictly associated with the intensified generation of RNS and is strongly and positively correlated with exercise intensity [5]. The postexercise oxidative stress may induce radical-mediated microinjuries of muscle fibers and connective tissues, which results in muscle pain, the prolongation of recovery and, consequently, a decrease in sports performance. Thus, it seems reasonable to reduce the exercise-induced ROS and RNS formation in order to improve performance, for example, via antioxidant supplementation [6]. However, there are also reports that indicate a completely opposite effect of such a procedure since it is the fact that reactive oxygen and nitrogen species are necessary for proper muscle contraction both at rest and during PA. Therefore, the literature recommends an appropriate supply of antioxidants in the form of a balanced diet as the best method of maintaining the oxidant-antioxidant equilibrium in exercising people [7]. Besides, a key issue is the adaptation for the increased concentration of reactive oxygen and nitrogen species after exercise because the organism can adapt to them by

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enhancing its antioxidant response and thus can adapt to exercise-induced disorders. In professional sportsmen, increased activities of antioxidant enzymes are commonly observed [5]. Similarly, in diseases of the cardiovascular system, oxidative stress is a key issue, and antioxidant supplementation has found no clear benefit in the prevention and treatment of CVDs [8,9]. Therefore, the current, popular direction for scientific research is searching for non-pharmacological agents. This primarily applies to physical training, which is included in the WHO recommendations regarding the prevention of CVDs, and positive impacts, which may result primarily from the adaptation of the organism to increased concentrations of reactive oxygen and nitrogen species [10].

The present paper provides an overview of the role of radical-mediated stress in CVDs, the role of PA in their prevention and treatment and, finally, it discusses the effectiveness of antioxidant supplementation in this context.

2. The Oxidant-Antioxidant Balance in Human Organisms

In the course of numerous disease entities, ROS and RNS are an etiological factor, and the generation of these molecules increases as a result of homeostasis disorders [11,12]. Disease states are not the only source of ROS and RNS. The aerobic metabolic processes necessary for the proper functioning of tissues pose a great challenge to the mechanisms responsible for maintaining redox homeostasis [13,14]. Reactive oxygen and nitrogen species constitute a highly reactive group of radicals and non-radical chemical compounds which contain oxygen or nitrogen in their structure [15]. These molecules are characterized by a short lifetime due to interactions with other biomolecules present in the cell [16]. ${}^{\bullet}O_2^{-}$, hydrogen peroxide (H₂O₂) and hydroxyl radicals (*OH) are ROS generated by endogenous metabolic processes involving NOX, myeloperoxidase (MPO) and lipoxygenase (LOX) [17]. The main endogenous source of RNS is the metabolism of L-arginine with the participation of selected isoforms of nitric oxide synthase (NOS)—eNOS, inducible NOS (iNOS) and neuronal NOS (nNOS). Reactions catalyzed by these enzymes produce •NO [18]. External environmental factors are also sources of reactive oxygen and nitrogen species. Air pollution related to industrial human activity leads to an increase in the level of sulfur dioxide (SO₂), carbon monoxide (CO), ozone (O₃) and nitrogen dioxide (NO₂) [19]. Additionally, exposure to ionizing radiation, smoking and chronic inflammation lead to a significant increase in ROS generation [20] (Figure 1).

Both ROS and RNS fulfill important biological functions and are essential for the maintenance of homeostasis. ROS are involved in the redox communication between cells [21]. This phenomenon enables the control of cell proliferation and apoptosis [22]. The immune system uses ROS to neutralize pathogens in a process called the respiratory burst [23]. NO is essential for the proper functioning of the cardiovascular system. This RNS maintains adequate blood flow by regulating the resistance of blood vessels and also inhibits the aggregation of platelets and leukocytes [24].

Reactive oxygen and nitrogen species generated in excessive amounts lead to unfavorable phenomena. Antioxidants are molecules that reduce the negative impact of ROS and RNS on tissues [25]. One of the lines of antioxidant defense are endogenous biomolecules. Antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPx) have a crucial role in this system [26]. Endogenous antioxidants with a low molecular mass are equally important. This group includes glutathione, uric acid, lipoic acid, bilirubin and melatonin [27]. Compounds with antioxidant potential may be supplied with food. Exogenous antioxidants such as vitamins A, C, E and polyphenols are especially important when endogenous systems cease to function effectively [28].

The condition where the generation of ROS and RNS is particularly intense and the antioxidant mechanisms are insufficient is called oxidative stress. Free radicals are reactive with proteins, lipids and the genetic material of the cell [29]. Oxidative modifications of short peptides and large proteins lead to the impairment of the functions of individual biomolecules, affecting the conformation of the protein and the activity of enzymes [30]. After the interaction with ROS, lipids building cell membranes undergo peroxidation and

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no longer constitute an effective barrier protecting the internal elements of the cell [31]. Mutations and, consequently, carcinogenesis are the result of the influence of ROS on DNA [32]. Considering the negative influence of oxidative stress on biomolecules and, at the same time, the significant role of ROS and RNS, it is necessary to maintain a redox balance.

3. Oxidative Stress in Cardiovascular Diseases

Oxidative damage to the cardiovascular system concerns first of all the lipid membranes of the cardiac and vascular myocytes [33] and generates numerous peroxide and aldehyde compounds such as lipid hydroperoxides, malondialdehyde (MDA), F2-isoprostanes and 4-hydroxy-2-nonenal (HNE) [34–40]. All these compounds are known as the classical biomarkers of oxidative stress (Figure 1).

Since 1968, the level of MDA in biological samples has been measured by means of the spectrophotometric method with thiobarbituric acid (TBA) [41]. Unfortunately, under conditions of this commonly used assay, non-lipid compounds, e.g., derivatives of glucose, sucrose, 2-deoxyribose and sialic acid, abundantly contained in biological material, may form MDA in the TBA reaction mixtures. Hence, its specificity has been strongly questioned for over 30 years [42–45]. Nevertheless, more sophisticated high-performance liquid chromatographic (HPLC) methods have been developed for the separation of the MDA-TBA adduct from other possible TBA reactants using spectrofluorimetric detection. Consequently, researchers showed that increased serum MDA levels predicted adverse health outcomes in 634 patients with stable coronary artery disease (CAD) [46] and in 774 patients suffering from chronic heart failure (CHF) [47].

F2-isoprostanes belong to stable prostaglandin-like compounds with distinct sidechain structures compared to cyclooxygenase-derived prostaglandins. They are generated in vivo as products of the free radical-induced peroxidation of arachidonic acid [34,35]. Among the group of sixty-four F2-isoprostanes, 8-iso-prostaglandin F2 α (8-iso-PGF2 α , also known as 8-epi prostaglandin F2 α , 8-epi PGF2 α) was selected as a quantitative biomarker of oxidative stress [48]. For the first time, the association of oxidative stress with a progression of heart failure in humans (ischemic heart disease (IHD) and/or valvular heart disease) was demonstrated in 1998 by Mallat et al. The scale of the oxidative damage in the heart, expressed by the levels of 8-iso-PGF2 α measured in pericardial fluid, was significantly correlated with ventricular dilatation, which was assessed by, among others, the measurement of left ventricular end-diastolic (r = 0.5, p = 0.008) as well as end-systolic (r = 0.46, p = 0.026) diameters [39]. Moreover, during 17 years of the follow-up study, it was observed that the risk of developing fatal CHD (n = 141) and stroke (n = 109) was 80% higher among postmenopausal women with a urinary concentration of 8-iso-PGF2α included in the highest quartile (the odds ratio was 1.8, and a 95% confidence interval (CI) amounted to 1.1–3.1, p = 0.02) [49]. Another case-control study showed that patients with CHD (n = 93) also significantly differed from healthy controls (n = 93) in terms of urinary 8-iso-PGF2α levels (120–193 pmol/mmol creatinine vs. 77–139 pmol/mmol creatinine, respectively, p < 0.001). A multivariate analysis indicated that among the five risk factors of CHD (body mass index (BMI), C-reactive protein, high-density lipoprotein (HDL) cholesterol, systolic blood pressure and 8-iso-PGF2 α) only C-reactive protein (>3 mg/L, p < 0.01) and 8-iso-PGF2 α (\geq 131 pmol/mmol creatinine, p < 0.001) had a predictive value of CHD occurrence [33].

In a clinical study, the expression of HNE-modified proteins was measured immuno-histochemically in endomyocardial biopsy samples derived from 23 patients with dilated cardiomyopathy. A 5.3-fold greater HNE-positive heart area was demonstrated in these samples compared to those collected from control subjects (n = 13, p < 0.0001), which was indicative of the oxidative stress in the myocardium of patients with heart failure [40].

Moreover, other biomarkers of oxidative stress were also found in the course of CVDs, e.g., the products of a nitrosative stress (excessive RNS concentration). They are represented, among others, by 3-nitrotyrosine (3-NT), the nitration product of tyrosine residues in the proteins present in the vessel wall or in the myocardium [3,50,51]. However, the literature

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provides incoherent data on the prognostic value of 3-NT in the context of CVDs. Using an accurate HPLC with the tandem mass spectrometry method, Shishehbor et al. found significantly higher levels of 3-NT (p < 0.001) in the serum proteins of CAD patients (9.1 µmol/mol tyrosine as the median) compared to the group of healthy individuals (5.2 µmol/mol tyrosine) [52]. On the other hand, the ELISA-based measurements indicated a lack of an association between the levels of 3-NT and mortality rates among CAD patients (a follow-up time of 4 years) [53] (Figure 1).

An oxidatively damaged DNA is also observed in people suffering from CVDs. 8-oxo-7,8-dihydro-2'-deoxyguanosine (8-oxodG) was widely assessed as a cellular [54] and urinary [55] marker of DNA oxidative damage. A meta-analytic study performed by Di Minno et al. revealed that 810 patients with CVDs such as CAD, carotid atherosclerosis, PAD and stroke showed markedly higher 8-oxodG levels, both in leukocyte DNA and urine, as compared to 1106 control individuals (standard mean difference (SMD) equal to 1.04, 95% CI: 0.61-1.47, p < 0.00001) [56]. A similar meta-analytic review yielded the conclusion that 446 patients with heart failure showed significantly elevated 8-oxodG levels compared to 140 healthy controls (SMD = 0.89, 95% CI = 0.68-1.10, p < 0.00001) [57] (Figure 1).

4. Exercise Impacts on Redox State

Physical exercise is one of the physiological factors which, through the enhanced generation of ROS and RNS, impacts oxidant–antioxidant balance. The generation of these active derivatives of oxygen and nitrogen is indispensable for the appropriate course of adaptive processes for exercise [58]. However, when ROS are generated in excess, they may lead to the damage of cell components [59–61], including lipids, proteins and DNA [62].

The main source of ROS during exercise has not been ultimately defined so far, which results, among others, from the fact that oxidative stress caused by PA may be evoked not only in skeletal muscle but also in other tissues [63]. Initially, what was considered the main source of ROS during exercise was the mitochondrial respiratory chain; however, more recent research demonstrated that skeletal muscle mitochondria produce more *O₂in the basal condition than during exercise [64,65]. Under the conditions of PA, what also decreases is the generation of H₂O₂ by mitochondria [66]. What is considered as a significant source of ROS during exercise is xanthine oxidase (XO) and NOX. XO is generated under conditions of ischemia from xanthine dehydrogenase. Such a state may be a result of exhaustive physical exercises when blood flow is shunted from many organs and tissues and is redirected to the working muscles. After concluding exercise under the conditions of reperfusion, XO generates ${}^{ullet}O_2^-$ and H_2O_2 as by-products of xanthine and hypoxanthine oxidation to uric acid [67]. The activation of endothelial XO was proved, for instance, 24 h after a single anaerobic exercise in men and women [68]. A significant source of ROS in skeletal muscles after exercise apart from mitochondria and XO is also NOX. The basic function of this family of enzymes is ROS generation. NOX occurs not only in phagocytic cells but also in other types of cells; among others, in endothelial cells, cardiomyocytes and skeletal muscle [69]. It was demonstrated, among others, that highintensity interval training in mice is associated with an increase in NOX2 activity in skeletal muscles [70].

PA is also accompanied by an enhanced generation of ${}^{\bullet}$ NO [71]. The ${}^{\bullet}$ NO generated in low concentrations in various types of cells has the function of cellular or intracellular signaling molecules engaged, among others, in the regulation of vascular wall tension. Generated in excess, they may indicate destructive properties [72]. Activation was proved of both the iNOS and the eNOS after exhaustive exercise [73]. The source of ${}^{\bullet}$ NO is also erythrocytes. It is assumed that this is a source of ${}^{\bullet}$ NO that may play a role in the regulation of local blood flow dynamics during PA [74]. On the other hand, however, the reaction of ${}^{\bullet}$ NO with ${}^{\bullet}$ O₂ leads to the formation of a strong RNS—peroxynitrite (ONOO⁻) [75].

Various types of PA probably elicit different pathways of free radical production [67,76]. The connection between exercise and oxidative stress is extremely complex, depending on the mode, intensity and duration of exercise [77] and the training status of the

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individual [78]. PA can be differentiated into aerobic and anaerobic (depending on the intensity), acute and chronic (in relation to the frequency) and concentric, eccentric and isometric (in relation to muscle contractions) [67]. It was proved that oxidative stress accompanies both aerobic exercise, especially an intense one [79,80], and anaerobic [80,81]. Aerobic exercise as early as at its beginning leads to the enhanced generation of ROS, whereas anaerobic exercise may induce prolonged ROS generation [82].

Single bouts of exercise lead to oxidative stress [83,84]. As a result of physical training, which is accompanied by repetitive exposure to the enhanced generation of reactive oxygen and nitrogen species, adaptation changes occur, which may protect against oxidative stress. This fact remains in accordance with the so called hormesis theory, according to which low-level exposure to ROS elicits beneficial stress adaptation, whereas too high of a generation of ROS leads to oxidative stress [85]. Tolerance increase to exercise-induced oxidative stress was demonstrated, among others, after aerobic training in overweight/obese adolescent girls [86]. An adaptive increase in antioxidant enzyme activity after training was indicated after 3 months of karate training in elite athletes [87] and in the erythrocytes of handball athletes during 6 months of monitoring [88]. Some of the research also proves that a single bout of exercise confers increased resistance to a subsequent non-exercise oxidative challenge, but only in young people [89].

It was demonstrated that exercise also impacts the existing balance between ROS and RNS in the vascular system. Regular PA improves vascular function, which in part derives from a reduction in cellular ROS and a restoration of *NO bioavailability (a change in the ROS/NO balance to favor *NO) [90].

5. Effects of Physical Exercise on Redox Equilibrium in Cardiovascular Diseases

CVDs constitute disorders manifested by a chronic inflammatory state and alterations of the oxidant–antioxidant balance of the oxidative stress character [91,92]. It is common knowledge, however, that muscle activity during physical exercise leads to a significant increase in ROS concentration and improves the antioxidant defense system in the organism [93,94].

Linke et al. examined 23 patients with CHF, aged 56 ± 4 , and 12 controls (healthy subjects at an approximate age to the patients). The patients were randomly divided into two groups—the group of people exercising every day for 6 months and the group not exercising during this period of time (sedentary lifestyle). The PA of the patients was diversified; however, it was mainly effort on the cycloergometer for 20 min daily with a heart rate corresponding to 70% of the maximal oxygen uptake. What constituted the test material were the bioptates of the skeletal muscle. SOD, CAT and GPx activity were determined in homogenized muscle samples as well as with nitrotyrosine concentrations. What was observed in patients without division into groups was a significantly lower activity of enzymes and higher concentrations of nitrotyrosine than in the controls with regard to baseline levels. The training, however, caused a significant increase in CAT and GPx activity in the patients, and these values were higher than in the non-exercising subjects, as well as a decrease in the concentration of nitrotyrosine (p < 0.05). Thus, it was not proved that PA altered SOD activity in the skeletal muscles of subjects with CHF [95] (Table 1). A lack of an impact of aerobic exercise on SOD was also proved by Fenty-Stewart et al. in the plasma of venous blood in 100 hypertensive patients (50-75 years). However, they observed a positive correlation between SOD activity and 8-iso-PGF2 α concentration in urine [96], whereas, Kostić et al. observed a statistically significant decrease in erythrocyte GPx activity after an acute exercise test on the cycloergometer in peripheral blood in 40 hypertensive patients (51.19 \pm 8.37 years) [97]. Yu et al. examined dependence between the type and frequency of physical exercise, and redox biomarkers in the plasma of venous blood in persons between 45 and 79 years old with idiopathic arterial hypertension (n = 402), with the use of a control group (n = 1047). The study considered five types of PA, from low intensity to high intensity exercises such as walking/dancing, taiji/yoga, running/biking/climbing, ball sports and gym workouts including swimming. Each type of PA was divided into

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3 degrees of performance frequency. The PA of all subjects was therefore rare (0 times/week), moderately frequent (1–3 times/week) or frequent (4–7 times/week). The authors noticed that even the least intensive physical exercise may have had a favorable impact on redox equilibrium (a decrease in MDA and HNE concentration, an increase in SOD activity) in the hypertensive patients, which is also accompanied by improvement with regard to clinical parameters (arterial blood pressure, BMI, glycosylated hemoglobin, triglycerides) if exercises are performed adequately frequently and regularly [98]. On the other hand, in the study on the oxidant-antioxidant balance in peripheral blood in 43 participants with arterial hypertension, aged 60-80, the impact of regular walking performed in a continuous manner (30 min walking at the pace of one step per second, three times a week) and at intervals (3 \times 10 min march at the pace of one step per second + 1 min break after each repetition, three times a week) was determined. The group walking in a continuous manner was constituted by 8 males and 14 females, whereas the interval group consisted of 2 males and 19 females. After 12 weeks, the concentration of reduced glutathione (GSH) in both groups was increased, and the concentration of oxidized glutathione (GSSG) was decreased (p < 0.05) (both parameters were examined in the whole blood). Additionally, MDA concentrations in plasma in both groups were subject to a statistically significant decrease. The differences between the groups, however, were statistically insignificant with the exception of some clinical parameters (the triglycerides concentration, the ratio of the total cholesterol to HDL and the atherosclerogenic index were lower in the group walking continuously; p < 0.05). Thus, the study confirmed the advantageous influence of long-term and regular aerobic exercise of a low intensity on redox equilibrium in the venous blood of hypertensive subjects [99]. Tsarouhas et al., however, determined the total antioxidant capacity (TAC) in the serum of 20 males and 7 females (66.8 ± 13.1 years) with CHF subjected to effort, and 9 males and 3 females (67.0 \pm 5.6 years) with the same disease without introducing a training program. All the ill persons were treated with statins, and apart from the TAC, the lipid and glycemic profiles and the TNF- α concentrations in the serum were examined in these patients. The control group constituted 12 healthy males and 5 healthy females (65.7 \pm 9.4 years). After 12 weeks of training, an increase in the TAC was observed in the training group in relation to the non-training one. Additionally, the TAC was higher in both groups of persons with cardiac failure in comparison with the control group. The researchers stated that in patients with CHF, daily, moderate, unsupervised physical activity such as walking was able to improve their lipid and glycemic profile with the simultaneous alleviation of the inflammatory state and oxidative stress [100]. In 18 patients with cardiac failure (13 males and 5 females), aged 28-59, venous blood tests were performed after the application of a 30 min and a 45 min effort of low intensity and a 30 min effort of moderate intensity. The MDA concentration in the plasma was determined as well as CAT and SOD activity in the erythrocytes. Moderate physical effort caused the most significant changes; among others, a significant increase in the MDA concentration and CAT activity. Thus, the intensity of the exercises was in this case a more significant factor with respect to evoking physiological effects than the duration of the effort [101]. A long-term and extremely complex study was performed by Raberin et al., who examined the relation between PA and sex, as well as cardiovascular risk factors and redox parameters in peripheral blood in the elderly. Altogether, 1011 healthy subjects were qualified for the study, who were at least 65 years old on recruitment day. The study lasted 10 years in total. During that time, the persons recruited were observed and tested clinically with respect to cardiovascular and cerebrovascular events. In the final year, among 545 persons (318 females aged 75.8 \pm 1.2 and 227 males aged 75.8 \pm 1.1), redox parameters in the plasma were determined and they were the following: uric acid (UA), *NO (as the sum of nitrite and nitrate (NOx) concentrations), ferric-reducing antioxidant power (FRAP), advanced oxidation protein products (AOPP), MDA, GPx and SOD. The authors concluded in general that biochemical cardiovascular risk factors, including redox parameters, occur in females at a lower level than in males, which may explain the lower percentage of CVDs in older women. It was not observed, however, that PA correlated negatively with the level of cardiovascular risk

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factors tested both in females and in males [102]. There also exists a study in which redox parameters were determined in saliva. The study encompassed two groups of patients after myocardial infarction, i.e., 21 patients aged 42-71 (67% males, group I) and 21 patients aged 47–79 (81% males, group II). Group I was subjected to physical effort on a cycloergometer, and group II performed breathing and balance exercises. The MDA and TAC concentrations were measured, which were expressed in the form of the 2.2-diphenyl-1-picryl-hydrazyl (DPPH) concentration. The MDA concentration was decreased and the TAC increased in group II (p < 0.05). In group I, no statistically significant differences were observed. Thus, it was demonstrated that the form of PA may have a different impact on antioxidant capabilities or on ROS concentration in saliva in patients after myocardial infarction [103]. Lately, comprehensive data on redox balance in individuals with CVDs were provided by Tofas et al., who studied patients suffering from CAD. The patients were submitted to an 8-month training with division into three groups depending on exercise type, i.e., aerobic (n = 15, 61 \pm 7 years), anaerobic (weight) (n = 11, 62 \pm 8 years) and mixed (aerobic and weight exercises) (n = 15, 64 ± 6 years). The fourth group of patients was the control, not exercising group (n = 15, 64 \pm 8 years). The authors revealed that all types of training in CAD patients induced positive changes in the redox equilibrium in their peripheral blood; however, the most remarkable and pronounced alterations were found after aerobic training (compared to pre-training, baseline values). The authors determined the concentrations of thiobarbituric acid reactive substances (TBARS) and protein carbonyls (PCs) in plasma, GSH and GSSG in erythrocytes, as well as the TAC in serum and the CAT activity in erythrocytes. In all patients (all training groups), a statistically significant decrease in the systolic blood pressure was also observed, whereas the diastolic blood pressure decreased in the aerobic and anaerobic groups compared to the control individuals (health and anthropometric measures similar to the non-training patients in the study period; n = 15, 64 ± 8 years). Interestingly, most of the redox status parameters restored in all the CAD subjects close to the pre-exercise values at the end of the 3-month detraining period that was followed immediately by an 8-month training session [104]. The examination of 94 patients (60 males and 34 females, 68.0 \pm 14.5 years) hospitalized due to acute heart failure and subjected to an exercise test proved, however, that ROS generation in response to physical effort increases the number of cardiac events. The authors concluded that if antioxidant capacity increases as a result of PA, then we can expect beneficial effects for cardiac failure patients; however, if it does not happen and the ROS concentration significantly increases, the effects are negative [105] (Table 1).

Table 1. Exercise impacts on redox balance in cardiovascular diseases.

Cardiovascular Disorder	Study Group	Exercise	Material	Outcome	Ref.
Chronic heart failure	Patients exercising and not exercising (n = 23) Controls (n = 12)	aerobic (bpm of 70% VO _{2max}), 20 min/day, 6 months	Skeletal muscle	↑CAT ↑GPx ↓NT	[95]
	Patients exercising (n = 27) Patients not exercising (n = 12) Controls (n = 17)	aerobic (moderate), every day, 12 weeks	Peripheral blood	↑serum TAC	[100]
	Patients (n = 18)	a single 30 min bout and 45 min bouts of low intensity, and a single 30 min bout of moderate intensity	Peripheral blood	↑plasma MDA ↑erythrocytic CAT (moderate exercise)	[101]
Acute heart failure	Patients (n = 94)	aerobic and anaerobic, 30 min/day (cardiac rehabilitation), 5 days/week during hospitalization	Peripheral blood	†serum dROM = †no. of cardiac events (poor prognosis)	[105]

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Table 1. Cont.

Cardiovascular Disorder	Study Group	Exercise	Material	Outcome	Ref.
Hypertension	Patients (n = 100)	aerobic (50 \rightarrow 70% VO _{2max}), 20 \rightarrow 40 min/day, 3 days/week, 6 months	Peripheral blood, urine	plasma SOD = urine 8-iso-PGF2α (positive correlation)	[96]
	Patients $(n = 40)$ Controls $(n = 20)$	maximal (a single acute bout)	Peripheral blood	↓erythrocytic GPx	[97]
	Patients (n = 402) Controls (n = 1047)	aerobic (low-high intensity and frequency) aerobic (low intensity,	Plasma of peripheral blood	↓MDA ↓HNE ↑SOD	[98]
	Patients (n = 43)	continuous and intermittent), 30 min/day, 3 days/week, 12 months	Peripheral blood	whole blood ↑GSH ↓GSSG ↓MDA	[99]
Cardiovascular and cerebrovascular events	Patients (n = 545)	Approx. 5 h/day of low-intensity physical activity (<3 METs)	Plasma of peripheral blood	↓redox parameters in females vs. males	[102]
Myocardial infarction	Patients (n = 42)	single aerobic, 30 min (low intensity) (cardiac rehabilitation)— cycloergometer or breathing and balance	Saliva	↓MDA ↑TAC after breathing and balance exercises	[103]
Coronary artery disease	Patients (n = 56)	aerobic, anaerobic or mixed, 50–60 min/day, 3 days/week, 8 months	Peripheral blood	↑GSH ↓GSSG ↓TBARS (aerobic exercises)↓PCs ↑TAC ↑CAT (all types of exercises)	[104]

Abbreviations and symbols: bpm—beats per minute (heart rate); VO_{2max} —maximal oxygen uptake; \uparrow —increased level; \downarrow —decreased level; CAT—catalase; GPx—glutathione peroxidase; NT—nitrotyrosine; TAC—total antioxidant capacity; MDA—malondialdehyde; d-ROM—diacron reactive oxygen metabolites (mainly hydroperoxides); SOD—superoxide dismutase; iso-PGF2 α —8-iso-prostaglandin F2 α ; METs—metabolic equivalents; HNE—4-hydroxy-2-nonenal; GSH—reduced glutathione; GSSG—oxidized glutathione; TBARS—thiobarbituric acid reactive substances; PCs—protein carbonyls.

6. Pathological Consequences of Oxidative Stress in Cardiovascular System and Relation to Exercise

As mentioned, CVDs are disorders characterized by oxidative stress and inflammation in the myocardium and blood vessels [91,92]. Oxidative stress may be both a primary and a secondary factor of many CVDs [106].

In myocardium, the overproduction of ROS is manifested by the hypertrophy and fibrosis initiated by angiotensin II. This peptide hormone leads to NOX activation. As a result, formed ${}^{\bullet}O_2^-$ induces redox signaling pathways, the final effect of which is increased protein synthesis, including procollagen I and III [107,108]. Heart disorders induced by oxidative stress may also result from impacts of ROS directly on calcium channels in the sarcolemma (ryanodine receptors) and sarcoplasmic reticulum [109].

Oxidative stress principally triggers specific arterial dysfunction manifested by arterial stiffness—atherosclerosis [106]. ROS, RNS and oxidatively injured molecules induce the expression of inflammatory cytokines and adhesion molecules in endothelial cells and smooth myocytes, leading to the atherogenic remodeling of the vascular wall. Significantly higher concentrations of ROS were reported in acute CAD compared to chronic coronary syndromes (i.e., stable angina pectoris), which also suggests the impact of ROS on atheroma plaque stability [110]. In this case, it is also caused by NOX, which was proven by several clinical studies in human subjects [111]. NOX is activated and induces oxidative stress in the endothelium by angiotensin II-dependent signaling, but also through platelet-derived growth factor (PDGF) and tumor necrosis factor alpha (TNF- α). Hemodynamic param-

eters influence the NOX activity, as well. Laminar flow decreases NOX activity while a turbulent flow favors the enzyme activity; therefore, arterial hypertension itself may induce atherosclerosis, whereas oxidative stress can be the source of hypertension. ${}^{\bullet}O_2{}^{-}$ decreases ${}^{\bullet}NO$ concentration to form ONOO $^{-}$, and together with H_2O_2 can increase endothelin-1 synthesis and induce endothelial cell apoptosis, leading to vasoconstriction [106]. It was also reported that ONOO $^{-}$ is a significant factor of pathological mechanisms in brain stroke, as it leads to the contraction of the smooth myocytes of the cerebral arterioles [112]. Moreover, increased ONOO $^{-}$ concentrations were found in hyperhomocysteinemia, another important risk factor for endothelial dysfunction, peripheral arterial disease and IHD [113]. Other source of ROS, oxidative stress, inflammation and consequently atherosclerosis may also be XO and LOX. In addition to ${}^{\bullet}NO$, oxidatively injured endothelial cells also release significantly lower amounts of other vasodilators (prostacyclin, adrenomedullin, hyperpolarizing endothelial factor) and become too permeable to the oxidized form of low-density lipoproteins (ox-LDL). Oxidative stress disrupts vascular wall function as well as directly acting vascular smooth muscle cells, in which it damages contractile proteins [106].

It should also be mentioned in this section that all the aforementioned molecular mechanisms that lead to atherosclerosis accompany diabetes [106]. Diabetes mellitus (DM) is therefore a pathological condition of the organism that particularly predisposes them to CVDs. Hence, individuals suffering from diabetes are accompanied by hypertension, dyslipidemia, hyperglycemia and insulin resistance [114]. Hyperglycemia, the primary symptom of DM, is also combined with ROS production. The autooxidation of glucose and the non-enzymatic glycation of proteins generates ${}^{\bullet}O_2{}^{-}$. For instance, the glycation may occur directly between glucose and LDL or apolipoprotein B to form advanced glycation end products (AGEs), which induce lipid peroxidation. ROS facilitate that process. Thus, AGEs are pathological changes in the structure of plasma proteins; however, they have their own specific receptors (receptors of AGE, RAGE). The binding of AGEs to RAGE in various cell types (i.e., endothelial cells, smooth muscle cells, macrophages, monocytes and lymphocytes) contributes to ROS production and inflammation dependent on the activation of NOX and nuclear factor kappa B (NF-kB) [106].

The impact of PA on patients with CVDs seems to be unambiguously positive if applied at therapeutic doses. This was confirmed by at least several studies. One of them is the mentioned study of Tsarouhas et al., who reported that in patients with CHF, daily, moderate and unsupervised physical exercise for 12 weeks, such as walking, improved their lipid and glycemic profile with a simultaneous alleviation of the inflammatory state and oxidative stress [100]. In another one, systolic and diastolic blood pressure as well as flow-mediated vasodilation (endothelial function assessment) meaningfully improved after 6 weeks of strength training (knee extensor exercise, three times a week) in six mildly hypertensive men (Stage 1) (71 \pm 2 years) [115]. Another study showed that, according to the authors, physical effort improved systolic blood pressure, pulse pressure, central aortic systolic blood pressure and central aortic pulse pressure, and it may delay arterial aging in hypertensive patients (n = 63; age of 40–70 years). The training program included 50 min of exercise, divided into 30 min aerobic exercises and 20 min resistance exercises per session, four times a week for 4 months [116].

7. Antioxidant Supplementation—Unconventional Treatment of Cardiovascular Diseases

The primary care level in cardiovascular disease reduction and treatment should cover the following drugs: aspirin, beta-blockers, angiotensin-converting enzyme inhibitors and statins [1]. An especially wide range of action and effectiveness have statins [117]. The anti-inflammatory response of high-dose statins and their beneficial effects on vascular functionality were even proven, which were also accompanied by a decrease in ROS concentration [106]. In this context, antioxidant supplementation also seems to be important since the cardioprotective effects of dietary antioxidants are well known. Indeed, a diet rich in fruits and vegetables results in an increase in serum antioxidant capacity and a decrease

in oxidative stress. In contrast, studies on antioxidant supplementation, even those that are numerically significant, have revealed no clear benefit in the prevention and therapy of CVDs. Both short- and long-term clinical trials have failed to consistently support the cardioprotective effects of supplemental antioxidant intake. For these reasons, and also to limit the immoderate consumption of antioxidant supplements, both the US Food and Drug Administration and the European Food Safety Authority have excluded from the package labels any information that could imply the potential health benefits of products with antioxidants [9]. For instance, Ardalani et al. demonstrated that the consumption of Cynara scolymus (artichoke) powder as a rich source of phenolic and antioxidant compounds could potentially improve BMI and systolic blood pressure in hypertensive patients [118]. In turn, in the mentioned six men with mild hypertension (Stage 1) (71 \pm 2 years), the benefits resulting from exercise (an improved systolic and diastolic blood pressure and flowmediated vasodilation after a 6-week knee extensor exercise, 3 times/week) deteriorated because of antioxidant supplementation (vitamins C and E, and alpha-lipoic acid). The levels of the parameters returned to the baseline state in the (hypertensive) subjects as a result of this supplementation. Moreover, a single intake of the supplement in a high dose (the documented antioxidant effect in the peripheral blood plasma) before starting the exercise program did not change the levels of the parameters studied [115].

The confusing effects of antioxidant supplementation can be explained in several ways. First, the term "antioxidant" applies to a very wide variety of chemical entities that share only the capability of chemical reduction (the donation of electrons). Dietary antioxidants are substances both with lipophilic and hydrophilic properties; moreover, the size and complexity of antioxidants vary widely. They can be small and simple (e.g., salicylates, ascorbic acid) or large and very complex, like polyphenols (e.g., tannic acid). This heavily impacts their bioavailability, which also depends on metabolism and the distribution of antioxidants in the organism. Bioavailability, which is an active form of the antioxidant in a target place, is a highly complex issue that depends on resistance to digestion and metabolic conversion by the gut microbiome, absorption, metabolism and clearance. Finally, antioxidants are, by definition, rapidly oxidized. Oxidation before or during ingestion might not only abolish antioxidant properties but could also actively promote oxidative stress, depending on the nature of the product formed. Hence, the donation of the electron(s) by the antioxidant is linked to its oxidation, which may lead to the formation of its reactive entity, including radical ones [8]. Moreover, it should be remembered that the effects of antioxidants in the healthy organism, in physiological conditions, may differ compared to pathological conditions, in the course of the disease, especially those characterized by redox disturbances.

8. Conclusions

It is widely known that PA positively influences quality of life, improving physical and mental fitness and, consequently, may also reduce the emergence and development of CVDs [119]. CVDs are commonly characterized by glucose metabolic syndrome, oxidative stress and inflammation, whilst PA is known to improve redox status, insulin sensitivity and endothelial function [100]. The effects of PA on oxidant–antioxidant equilibrium can be ambiguous and depend on the duration and intensity of exercise; however, in general, they are positive [95–105] (Table 1). Physical exercise of different intensities (usually of moderate or of low intensity, and long term) is already widely recommended as an aid in the prevention and treatment of CVDs. Exercise is recommended, for example, in the prevention of CAD, and as part of the treatment process in myocardial infarction, coronary bypass graft, heart transplantation and CHDs [93]. With regard to antioxidant supplementation as a potentially new approach to the treatment, it is not recommended due to inconsistent data coming from clinical trials. Probably, the best way to keep the oxidant–antioxidant equilibrium is a supply of antioxidants in a balanced diet, as the literature recommends in the case of exercising people.

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Abbreviations

coronary artery disease, also called coronary heart disease (CHD) or ischemic

heart disease (IHD)

CAT catalase

CHDs congenital heart defects CHF chronic heart failure CVDs cardiovascular diseases GPx glutathione peroxidases HNE 4-hydroxy-2-nonenal H_2O_2 hydrogen peroxide 8-iso-PGF2 α 8-iso-prostaglandin F2 α

LOX lipoxygenase MDA malondialdehyde

NOX nicotinamide adenine dinucleotide phosphate (NADPH) oxidase

NO nitric oxide

O₂ superoxide anion radical

OH hydroxyl radical PA physical activity

RNS reactive nitrogen species
ROS reactive oxygen species
SOD superoxide dismutase
XO xanthine oxidase

References

- 1. World Health Organization. Cardiovascular Diseases (CVDs). Available online: https://www.who.int/news-room/fact-sheets/detail/cardiovascular-diseases-(cvds) (accessed on 28 July 2022).
- Popa, A.R.; Fratila, O.; Rus, M.; Aron, R.A.; Vesa, C.M.; Pantis, C.; Diaconu, C.C.; Bratu, O.; Bungau, S.; Nemeth, S. Risk factors for adiposity in the urban population and influence on the prevalence of overweight and obesity. *Exp. Ther. Med.* 2020, 20, 129–133. [CrossRef] [PubMed]
- 3. Galougahi, K.K.; Antoniades, C.; Nicholls, S.J.; Channon, K.M.; Figtree, G.A. Redox biomarkers in cardiovascular medicine. *Eur. Heart J.* **2015**, *36*, 1576–1582. [CrossRef] [PubMed]
- 4. Pinckard, K.; Baskin, K.K.; Stanford, K.I. Effects of exercise to improve cardiovascular health. *Front. Cardiovasc. Med.* **2019**, *6*, 69. [CrossRef]
- 5. Sutkowy, P.; Woźniak, A.; Boraczyński, T.; Boraczyński, M.; Mila-Kierzenkowska, C. The oxidant-antioxidant equilibrium, activities of selected lysosomal enzymes and activity of acute phase protein in peripheral blood of 18-year-old football players after aerobic cycle ergometer test combined with ice-water immersion or recovery at room temperature. *Cryobiology* **2017**, 74, 126–131. [CrossRef]
- 6. Sutkowy, P.; Woźniak, A.; Boraczyński, T.; Mila-Kierzenkowska, C.; Boraczyński, M. Postexercise impact of ice-cold water bath on the oxidant-antioxidant balance in healthy men. *BioMed Res. Int.* **2015**, 2015, 706141. [CrossRef]
- 7. Peternelj, T.T.; Coombes, J.S. Antioxidant supplementation during exercise training: Beneficial or detrimental? *Sports Med.* **2011**, 41, 1043–1069. [CrossRef] [PubMed]
- 8. Goszcz, K.; Deakin, S.J.; Duthie, G.G.; Stewart, D.; Leslie, S.J.; Megson, I.L. Antioxidants in cardiovascular therapy: Panacea or false hope? *Front. Cardiovasc. Med.* **2015**, *2*, 29. [CrossRef]
- 9. Pellegrino, D. Antioxidants and cardiovascular risk factors. Diseases 2016, 4, 11. [CrossRef]
- 10. Tian, D.; Meng, J. Exercise for prevention and relief of cardiovascular disease: Prognoses, mechanisms, and approaches. *Oxid. Med. Cell Longev.* **2019**, 2019, 3756750. [CrossRef]
- 11. Canton, M.; Sánchez-Rodríguez, R.; Spera, I.; Venegas, F.C.; Favia, M.; Viola, A.; Castegna, A. Reactive oxygen species in macrophages: Sources and targets. *Front. Immunol.* **2021**, *12*, 734229. [CrossRef]
- 12. Forrester, S.J.; Kikuchi, D.S.; Hernandes, M.S.; Xu, Q.; Griendling, K.K. Reactive oxygen species in metabolic and inflammatory signaling. *Circ. Res.* **2018**, *122*, 877–902. [CrossRef] [PubMed]

13. Tretter, V.; Hochreiter, B.; Zach, M.L.; Krenn, K.; Klein, K.U. Understanding cellular redox homeostasis: A challenge for precision medicine. *Int. J. Mol. Sci.* **2021**, 23, 106. [CrossRef] [PubMed]

- 14. Sies, H. Oxidative eustress: On constant alert for redox homeostasis. Redox Biol. 2021, 41, 101867. [CrossRef]
- 15. Al-Shehri, S.S. Reactive oxygen and nitrogen species and innate immune response. Biochimie 2021, 181, 52–64. [CrossRef]
- 16. Luo, T.; Wang, D.; Liu, L.; Zhang, Y.; Han, C.; Xie, Y.; Liu, Y.; Liang, J.; Qiu, G.; Li, H.; et al. Switching reactive oxygen species into reactive nitrogen species by photocleaved O₂-released nanoplatforms favors hypoxic tumor repression. *Adv. Sci.* **2021**, *8*, 2101065. [CrossRef] [PubMed]
- 17. Liguori, I.; Russo, G.; Curcio, F.; Bulli, G.; Aran, L.; Della-Morte, D.; Gargiulo, G.; Testa, G.; Cacciatore, F.; Bonaduce, D.; et al. Oxidative stress, aging, and diseases. *Clin. Interv. Aging* **2018**, *13*, 757–772. [CrossRef] [PubMed]
- 18. Ghimire, K.; Altmann, H.M.; Straub, A.C.; Isenberg, J.S. Nitric oxide: What's new to NO? *Am. J. Physiol.-Cell Physiol.* **2017**, 312, C254–C262. [CrossRef]
- 19. Niemann, B.; Rohrbach, S.; Miller, M.R.; Newby, D.E.; Fuster, V.; Kovacic, J.C. Oxidative stress and cardiovascular risk: Obesity, diabetes, smoking, and pollution: Part 3 of a 3-part series. *J. Am. Coll. Cardiol.* **2017**, 70, 230–251. [CrossRef]
- 20. Takahashi, K.; Okumura, H.; Guo, R.; Naruse, K. Effect of oxidative stress on cardiovascular system in response to gravity. *Int. J. Mol. Sci.* 2017, *18*, 1426. [CrossRef]
- 21. Sies, H.; Jones, D.P. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat. Rev. Mol. Cell Biol.* **2020**, 21, 363–383. [CrossRef]
- 22. Aggarwal, V.; Tuli, H.; Varol, A.; Thakral, F.; Yerer, M.; Sak, K.; Varol, M.; Jain, A.; Khan, M.; Sethi, G. Role of reactive oxygen species in cancer progression: Molecular mechanisms and recent advancements. *Biomolecules* **2019**, *9*, 735. [CrossRef] [PubMed]
- 23. El-Benna, J.; Hurtado-Nedelec, M.; Marzaioli, V.; Marie, J.-C.; Gougerot-Pocidalo, M.-A.; Dang, P.M.-C. Priming of the neutrophil respiratory burst: Role in host defense and inflammation. *Immunol. Rev.* **2016**, *273*, 180–193. [CrossRef] [PubMed]
- 24. Tejero, J.; Shiva, S.; Gladwin, M.T. Sources of vascular nitric oxide and reactive oxygen species and their regulation. *Physiol. Rev.* **2019**, *99*, 311–379. [CrossRef]
- 25. Sinha, N.; Dabla, P. Oxidative stress and antioxidants in hypertension—A current review. *Curr. Hypertens. Rev.* **2015**, *11*, 132–142. [CrossRef]
- 26. Sies, H. Oxidative stress: A concept in redox biology and medicine. Redox Biol. 2015, 4, 180–183. [CrossRef] [PubMed]
- 27. Neha, K.; Haider, M.R.; Pathak, A.; Yar, M.S. Medicinal prospects of antioxidants: A review. Eur. J. Med. Chem. 2019, 178, 687–704. [CrossRef] [PubMed]
- 28. Pisoschi, A.M.; Pop, A. The role of antioxidants in the chemistry of oxidative stress: A review. *Eur. J. Med. Chem.* **2015**, 97, 55–74. [CrossRef]
- 29. Nakai, K.; Tsuruta, D. What are reactive oxygen species, free radicals, and oxidative stress in skin diseases? *Int. J. Mol. Sci.* **2021**, 22, 10799. [CrossRef]
- 30. O'Flaherty, C.; Matsushita-Fournier, D. Reactive oxygen species and protein modifications in spermatozoa. *Biol. Reprod.* **2017**, 97, 577–585. [CrossRef]
- 31. Su, L.-J.; Zhang, J.-H.; Gomez, H.; Murugan, R.; Hong, X.; Xu, D.; Jiang, F.; Peng, Z.-Y. Reactive oxygen species-induced lipid peroxidation in apoptosis, autophagy, and ferroptosis. *Oxid. Med. Cell. Longev.* **2019**, 2019, 5080843. [CrossRef]
- 32. Kirtonia, A.; Sethi, G.; Garg, M. The multifaceted role of reactive oxygen species in tumorigenesis. *Cell. Mol. Life Sci.* **2020**, 77, 4459–4483. [CrossRef] [PubMed]
- 33. Schwedhelm, E.; Bartling, A.; Lenzen, H.; Tsikas, D.; Maas, R.; Brümmer, J.; Gutzki, F.-M.; Berger, J.; Frölich, J.C.; Böger, R.H. Urinary 8-iso-prostaglandin F2alpha as a risk marker in patients with coronary heart disease: A matched case-control study. *Circulation* **2004**, *109*, 843–848. [CrossRef] [PubMed]
- 34. Morrow, J.D.; Harris, T.M.; Roberts, L.J. Noncyclooxygenase oxidative formation of a series of novel prostaglandins: Analytical ramifications for measurement of eicosanoids. *Anal. Biochem.* **1990**, *184*, 1–10. [CrossRef]
- 35. Morrow, J.D.; Hill, K.E.; Burk, R.F.; Nammour, T.M.; Badr, K.F.; Roberts, L.J. A series of prostaglandin F2-like compounds are produced in vivo in humans by a non-cyclooxygenase, free radical-catalyzed mechanism. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 9383–9387. [CrossRef]
- 36. Belch, J.J.; Bridges, A.B.; Scott, N.; Chopra, M. Oxygen free radicals and congestive heart failure. *Br. Heart. J.* **1991**, *65*, 245–248. [CrossRef]
- 37. McMurray, J.; Chopra, M.; Abdullah, I.; Smith, W.E.; Dargie, H.J. Evidence of oxidative stress in chronic heart failure in humans. *Eur. Heart J.* **1993**, *14*, 1493–1498. [CrossRef]
- 38. Keith, M.; Geranmayegan, A.; Sole, M.J.; Kurian, R.; Robinson, A.; Omran, A.S.; Jeejeebhoy, K.N. Increased oxidative stress in patients with congestive heart failure. *J. Am. Coll. Cardiol.* **1998**, *31*, 1352–1356. [CrossRef]
- 39. Mallat, Z.; Philip, I.; Lebret, M.; Chatel, D.; Maclouf, J.; Tedgui, A. Elevated levels of 8-iso-prostaglandin F2α in pericardial fluid of patients with heart failure. *Circulation* **1998**, *97*, 1536–1539. [CrossRef]
- 40. Nakamura, K.; Kusano, K.; Nakamura, Y.; Kakishita, M.; Ohta, K.; Nagase, S.; Yamamoto, M.; Miyaji, K.; Saito, H.; Morita, H.; et al. Carvedilol decreases elevated oxidative stress in human failing myocardium. *Circulation* **2002**, *105*, 2867–2871. [CrossRef]
- 41. Yagi, K.; Nishigaki, I.; Ohama, H. Measurement of serum TBA-value. Vitamins 1968, 37, 105–112.

42. Wade, C.R.; van Rij, A.M. Plasma thiobarbituric acid reactivity: Reaction conditions and the role of iron, antioxidants and lipid peroxy radicals on the quantitation of plasma lipid peroxides. *Life Sci.* **1988**, *43*, 1085–1093. [CrossRef]

- 43. Pryor, W.A.; Stanley, J.P.; Blair, E. Autoxidation of polyunsaturated fatty acids: II. A suggested mechanism for the formation of TBA-reactive materials from prostaglandin-like endoperoxides. *Lipids* **1976**, *11*, 370–379. [CrossRef] [PubMed]
- 44. Ohkawa, H.; Ohishi, N.; Yagi, K. Reaction of linoleic acid hydroperoxide with thiobarbituric acid. *J. Lipid Res.* **1978**, *19*, 1053–1057. [CrossRef]
- 45. Wade, C.R.; Jackson, P.G.; Van Rij, A.M. Quantitation of malondialdehyde (MDA) in plasma, by ion-pairing reverse phase high performance liquid chromatography. *Biochem. Med.* **1985**, *33*, 291–296. [CrossRef]
- 46. Walter, M.F.; Jacob, R.F.; Jeffers, B.; Ghadanfar, M.M.; Preston, G.M.; Buch, J.; Mason, R.P. Serum levels of thiobarbituric acid reactive substances predict cardiovascular events in patients with stable coronary artery disease: A longitudinal analysis of the prevent study. *J. Am. Coll. Cardiol.* **2004**, *44*, 1996–2002. [CrossRef]
- 47. Romuk, E.; Wojciechowska, C.; Jacheć, W.; Zemła-Woszek, A.; Momot, A.; Buczkowska, M.; Rozentryt, P. Malondialdehyde and uric acid as predictors of adverse outcome in patients with chronic heart failure. *Oxid. Med. Cell. Longev.* 2019, 2019, e9246138. [CrossRef]
- 48. Delanty, N.; Reilly, M.P.; Pratico, D.; Lawson, J.A.; McCarthy, J.F.; Wood, A.E.; Ohnishi, S.T.; Fitzgerald, D.J.; Fitzgerald, G.A. 8-epi PGF2α generation during coronary reperfusion. *Circulation* **1997**, *95*, 2492–2499. [CrossRef]
- 49. Roest, M.; Voorbij, H.A.M.; Van der Schouw, Y.T.; Peeters, P.H.M.; Teerlink, T.; Scheffer, P.G. High levels of urinary F2-isoprostanes predict cardiovascular mortality in postmenopausal women. *J. Clin. Lipidol.* **2008**, *2*, 298–303. [CrossRef]
- 50. Peluffo, G.; Radi, R. Biochemistry of protein tyrosine nitration in cardiovascular pathology. *Cardiovasc. Res.* **2007**, 75, 291–302. [CrossRef]
- 51. Daiber, A.; Hahad, O.; Andreadou, I.; Steven, S.; Daub, S.; Münzel, T. Redox-related biomarkers in human cardiovascular disease—Classical footprints and beyond. *Redox Biol.* **2021**, 42, 101875. [CrossRef]
- 52. Shishehbor, M.H.; Aviles, R.J.; Brennan, M.-L.; Fu, X.; Goormastic, M.; Pearce, G.L.; Gokce, N.; Keaney, J.F.; Penn, M.S.; Sprecher, D.L.; et al. Association of nitrotyrosine levels with cardiovascular disease and modulation by statin therapy. *JAMA* 2003, 289, 1675–1680. [CrossRef] [PubMed]
- 53. Quidim, A.V.L.; Bruno, T.; Leocádio, P.C.L.; Santos, I.S.; Alvarez-Leite, J.I.; dos Reis Menta, P.L.; Lotufo, P.A.; Bensenor, I.M.; Goulart, A.C. The prognostic value of nitrotyrosine levels in coronary heart disease: Long-term evaluation in the acute coronary syndrome registry strategy (erico study). *Clin. Biochem.* **2019**, *66*, 37–43. [CrossRef] [PubMed]
- 54. Collins, A. Comparison of different methods of measuring 8-oxoguanine as a marker of oxidative DNA damage. *Free Rad. Res.* **2000**, *32*, 333–341. [CrossRef] [PubMed]
- 55. Olinski, R.; Rozalski, R.; Gackowski, D.; Foksinski, M.; Siomek, A.; Cooke, M.S. Urinary measurement of 8-oxodG, 8-oxoGua, and 5HMUra: A noninvasive assessment of oxidative damage to DNA. *Antioxid. Redox Signal.* **2006**, *8*, 1011–1019. [CrossRef]
- 56. Di Minno, A.; Turnu, L.; Porro, B.; Squellerio, I.; Cavalca, V.; Tremoli, E.; Di Minno, M.N.D. 8-hydroxy-2-deoxyguanosine levels and cardiovascular disease: A systematic review and meta-analysis of the literature. *Antioxid. Redox Signal.* **2016**, 24, 548–555. [CrossRef]
- 57. Di Minno, A.; Turnu, L.; Porro, B.; Squellerio, I.; Cavalca, V.; Tremoli, E.; Di Minno, M.N.D. 8-hydroxy-2-deoxyguanosine levels and heart failure: A systematic review and meta-analysis of the literature. *Nutr. Metab. Cardiovasc. Dis.* **2017**, 27, 201–208. [CrossRef]
- 58. Jitcă, G.; Ösz, B.E.; Tero-Vescan, A.; Miklos, A.P.; Rusz, C.M.; Bătrinu, M.G.; Vari, C.E. Positive aspects of oxidative stress at different levels of the human body: A review. *Antioxidants* **2022**, *11*, 572. [CrossRef]
- 59. Wozniak, A.; Mila-Kierzenkowska, C.; Szpinda, M.; Chwalbinska-Moneta, J.; Augustynska, B.; Jurecka, A. Whole-body cryostimulation and oxidative stress in rowers: The preliminary results. *Arch. Med. Sci.* **2013**, *9*, 303–308. [CrossRef]
- 60. Sutkowy, P.; Woźniak, A.; Boraczyński, T.; Mila-Kierzenkowska, C.; Boraczyński, M. The effect of a single Finnish sauna bath after aerobic exercise on the oxidative status in healthy men. Scand. *J. Clin. Lab. Investig.* **2014**, 74, 89–94. [CrossRef]
- 61. Paprocki, J.; Sutkowy, P.; Piechocki, J.; Woźniak, A. Markers of oxidant-antioxidant equilibrium in patients with sudden sensorineural hearing loss treated with hyperbaric oxygen therapy. *Oxid. Med. Cell. Longev.* **2019**, 2019, 8472346. [CrossRef]
- 62. Sun, M.S.; Jin, H.; Sun, X.; Huang, S.; Zhang, F.L.; Guo, Z.N.; Yang, Y. Free radical damage in ischemia-reperfusion injury: An obstacle in acute ischemic stroke after revascularization therapy. *Oxid. Med. Cell. Longev.* **2018**, 2018, 3804979. [CrossRef] [PubMed]
- 63. Kawamura, T.; Muraoka, I. Exercise-induced oxidative stress and the effects of antioxidant intake from a physiological viewpoint. *Antioxidants* **2018**, 7, 119. [CrossRef] [PubMed]
- 64. Goncalves, R.L.S.; Quinlan, C.L.; Perevoshchikova, I.V.; Hey-Mogensen, M.; Brand, M.D. Sites of superoxide and hydrogen peroxide production by muscle mitochondria assessed ex vivo under conditions mimicking rest and exercise. *J. Biol. Chem.* 2015, 290, 209–227. [CrossRef] [PubMed]
- 65. Bouviere, J.; Fortunato, R.S.; Dupuy, C.; Werneck-de-Castro, J.P.; Carvalho, D.P.; Louzada, R.A. Exercise-stimulated ROS sensitive signaling pathways in skeletal muscle. *Antioxidants* **2021**, *10*, 537. [CrossRef] [PubMed]
- Wong, H.S.; Dighe, P.A.; Mezera, V.; Monternier, P.A.; Brand, M.D. Production of superoxide and hydrogen peroxide from specific mitochondrial sites under different bioenergetic conditions. J. Biol. Chem. 2017, 292, 16804–16809. [CrossRef]

67. Gomes, E.C.; Silva, A.N.; de Oliveira, M.R. Oxidants, antioxidants, and the beneficial roles of exercise-induced production of reactive species. *Oxid. Med. Cell. Longev.* **2012**, 2012, 756132. [CrossRef]

- 68. Wiecek, M.; Maciejczyk, M.; Szymura, J.; Kantorowicz, M.; Szygula, Z. Impact of single anaerobic exercise on delayed activation of endothelial xanthine oxidase in men and women. *Redox Rep.* **2017**, 22, 367–376. [CrossRef]
- 69. Bedard, K.; Krause, K.H. The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. *Physiol. Rev.* **2007**, *87*, 245–313. [CrossRef]
- Henriquez-Olguin, C.; Renani, L.B.; Arab-Ceschia, L.; Raun, S.H.; Bhatia, A.; Li, Z.; Knudsen, J.R.; Holmdahl, R.; Jensen, T.E. Adaptations to high-intensity interval training in skeletal muscle require NADPH oxidase 2. *Redox Biol.* 2019, 24, 101188. [CrossRef]
- 71. Oral, O. Nitric oxide and its role in exercise physiology. J. Sports Med. Phys. Fitness 2021, 61, 1208–1211. [CrossRef]
- 72. Bove, P.F.; van der Vliet, A. Nitric oxide and reactive nitrogen species in airway epithelial signaling and inflammation. *Free Radic. Biol. Med.* **2006**, *41*, 515–527. [CrossRef] [PubMed]
- 73. Gomez-Cabrera, M.-C.; Borrás, C.; Pallardó, F.V.; Sastre, J.; Ji, L.L.; Viña, J. Decreasing xanthine oxidase-mediated oxidative stress prevents useful cellular adaptations to exercise in rats. *J. Physiol.* **2005**, *567*, 113–120. [CrossRef] [PubMed]
- 74. Baskurt, O.K.; Ulker, P.; Meiselman, H.J. Nitric oxide, erythrocytes and exercise. *Clin. Hemorheol. Microcirc.* **2011**, 49, 175–181. [CrossRef] [PubMed]
- 75. Khan, M.A.; Akram, M.F.; Alam, K.; Ahsan, H.; Rizvi, M.A. Peroxynitrite-mediated structural changes in histone H2A: Biochemical and biophysical analysis. *Protein Pept. Lett.* **2020**, 27, 989–998. [CrossRef]
- 76. Vollaard, N.B.J.; Shearman, J.P.; Cooper, C.E. Exercise-induced oxidative stress: Myths, realities and physiological relevance. *Sports Med.* **2005**, *35*, 1045–1062. [CrossRef]
- 77. Pingitore, A.; Lima, G.P.; Mastorci, F.; Quinones, A.; Iervasi, G.; Vassalle, C. Exercise and oxidative stress: Potential effects of antioxidant dietary strategies in sports. *Nutrition* **2015**, *31*, 916–922. [CrossRef]
- 78. Thirupathi, A.; Wang, M.; Lin, J.K.; Fekete, G.; István, B.; Baker, J.S.; Gu, Y. Effect of different exercise modalities on oxidative stress: A systematic review. *BioMed Res. Int.* **2021**, 2021, 1947928. [CrossRef]
- 79. Moflehi, D.; Kok, L.Y.; Tengku-Kamalden, T.F.; Amri, S. Effect of single-session aerobic exercise with varying intensities on lipid peroxidation and muscle-damage markers in sedentary males. *Glob. J. Health Sci.* **2012**, *4*, 48–54. [CrossRef]
- 80. Ammar, A.; Trabelsi, K.; Boukhris, O.; Glenn, J.M.; Bott, N.; Masmoudi, L.; Hakim, A.; Chtourou, H.; Driss, T.; Hoekelmann, A.; et al. Effects of aerobic-, anaerobic- and combined-based exercises on plasma oxidative stress biomarkers in healthy untrained young adults. *Int. J. Environ. Res. Public Health* **2020**, 17, 2601. [CrossRef]
- 81. Wiecek, M.; Maciejczyk, M.; Szymura, J.; Szygula, Z.; Kantorowicz, M. Changes in non-enzymatic antioxidants in the blood following anaerobic exercise in men and women. *PLoS ONE* **2015**, *10*, e0143499. [CrossRef]
- 82. Shi, M.; Wang, X.; Yamanaka, T.; Ogita, F.; Nakatani, K.; Takeuchi, T. Effects of anaerobic exercise and aerobic exercise on biomarkers of oxidative stress. *Environ. Health Prev. Med.* **2007**, *12*, 202–208. [CrossRef] [PubMed]
- 83. Brinkmann, C.; Blossfeld, J.; Pesch, M.; Krone, B.; Wiesiollek, K.; Capin, D.; Montiel, G.; Hellmich, M.; Bloch, W.; Brixius, K. Lipid-peroxidation and peroxiredoxin-overoxidation in the erythrocytes of non-insulin-dependent type 2 diabetic men during acute exercise. *Eur. J. Appl. Physiol.* **2012**, *112*, 2277–2287. [CrossRef] [PubMed]
- 84. Otocka-Kmiecik, A.; Lewandowski, M.; Stolarek, R.; Szkudlarek, U.; Nowak, D.; Orlowska-Majdak, M. Effect of single bout of maximal exercise on plasma antioxidant status and paraoxonase activity in young sportsmen. *Redox Rep.* **2010**, *15*, 275–281. [CrossRef] [PubMed]
- 85. Henriquez-Olguin, C.; Boronat, S.; Cabello-Verrugio, C.; Jaimovich, E.; Hidalgo, E.; Jensen, T.E. The emerging roles of nicotinamide adenine dinucleotide phosphate oxidase 2 in skeletal muscle redox signaling and metabolism. *Antioxid. Redox Signal.* **2019**, 31, 1371–1410. [CrossRef]
- 86. Youssef, H.; Groussard, C.; Lemoine-Morel, S.; Pincemail, J.; Jacob, C.; Moussa, E.; Fazah, A.; Cillard, J.; Pineau, J.C.; Delamarche, A. Aerobic training suppresses exercise-induced lipid peroxidation and inflammation in overweight/obese adolescent girls. *Pediatr. Exerc. Sci.* 2015, 27, 67–76. [CrossRef]
- 87. Jemili, H.; Mejri, M.A.; Bouhlel, E.; Amri, M. Biochemical status, oxidative and antioxidant responses after 3-month specific training in elite karate athletes. *Physiol. Int.* **2017**, *104*, 344–354. [CrossRef]
- 88. Marin, D.P.; Bolin, A.P.; Campoio, T.R.; Guerra, B.A.; Otton, R. Oxidative stress and antioxidant status response of handball athletes: Implications for sport training monitoring. *Int. Immunopharmacol.* **2013**, *17*, 462–470. [CrossRef]
- 89. Nordin, T.C.; Done, A.J.; Traustadóttir, T. Acute exercise increases resistance to oxidative stress in young but not older adults. *Age* **2014**, *36*, 9727. [CrossRef]
- 90. Durand, M.J.; Gutterman, D.D. Exercise and vascular function: How much is too much? *Can. J. Physiol. Pharmacol.* **2014**, 92, 551–557. [CrossRef]
- 91. Mendes, F.; Martins, D. The impact of exercise on inflammation, oxidative stress and remodelling in cardiac muscle. *Arq. Bras. Cardiol.* **2020**, *114*, 106–108. [CrossRef]
- 92. Valaei, K.; Taherkhani, S.; Arazi, H.; Suzuki, K. Cardiac oxidative stress and the therapeutic approaches to the intake of antioxidant supplements and physical activity. *Nutrients* **2021**, *13*, 3483. [CrossRef] [PubMed]
- 93. Shephard, R.J.; Balady, G.J. Exercise as cardiovascular therapy. Circulation 1999, 99, 963–972. [CrossRef] [PubMed]
- 94. Senoner, T.; Dichtl, W. Oxidative stress in cardiovascular diseases: Still a therapeutic target? Nutrients 2019, 11, 2090. [CrossRef]

95. Linke, A.; Adams, V.; Schulze, P.C.; Erbs, S.; Gielen, S.; Fiehn, E.; Möbius-Winkler, S.; Schubert, A.; Schuler, G.; Hambrecht, R. Antioxidative effects of exercise training in patients with chronic heart failure: Increase in radical scavenger enzyme activity in skeletal muscle. *Circulation* 2005, 111, 1763–1770. [CrossRef] [PubMed]

- 96. Fenty-Stewart, N.; Park, J.Y.; Roth, S.M.; Hagberg, J.M.; Basu, S.; Ferrell, R.E.; Brown, M.D. Independent and combined influence of AGTR1 variants and aerobic exercise on oxidative stress in hypertensives. *Blood Press.* **2009**, *18*, 204–212. [CrossRef]
- 97. Kostić, N.; Čaparević, Z.; Marina, D.; Ilić, S.; Radojković, J.; Čosić, Z.; Ćelić, V.; Penčć, B.; Radojković, I. Impact of acute exercise on antioxidant enzymes activity and lipid status in blood of patients with hypertension. *Vojnosanit. Pregl.* **2009**, *66*, 695–699. [CrossRef]
- 98. Yu, Y.; Gao, Q.; Xia, W.; Zhang, L.; Hu, Z.; Wu, X.; Jia, X. Association between physical exercise and biomarkers of oxidative stress among middle-aged and elderly community residents with essential hypertension in China. *BioMed Res. Int.* **2018**, 2018, 4135104. [CrossRef]
- 99. Prasertsri, P.; Phoemsapthawee, J.; Kuamsub, S.; Poolpol, K.; Boonla, O. Effects of long-term regular continuous and intermittent walking on oxidative stress, metabolic profile, heart rate variability, and blood pressure in older adults with hypertension. *J. Environ. Public Health* **2022**, 2022, 5942947. [CrossRef]
- 100. Tsarouhas, K.; Tsitsimpikou, C.; Haliassos, A.; Georgoulias, P.; Koutsioras, I.; Kouretas, D.; Kogias, J.; Liosis, I.; Rentoukas, E.; Kyriakides, Z. Study of insulin resistance, TNF-α, total antioxidant capacity and lipid profile in patients with chronic heart failure under exercise. *In Vivo* **2011**, *25*, 1031–1037.
- 101. Ribeiro-Samora, G.A.; Rabelo, L.A.; Ferreira, A.C.C.; Favero, M.; Guedes, G.S.; Pereira, L.S.M.; Parreira, V.F.; Britto, R.R. Inflammation and oxidative stress in heart failure: Effects of exercise intensity and duration. *Braz. J. Med. Biol. Res.* **2017**, *50*, 1–8. [CrossRef]
- 102. Raberin, A.; Connes, P.; Barthélémy, J.C.; Robert, P.; Celle, S.; Hupin, D.; Faes, C.; Rytz, C.; Roche, F.; Pialoux, V. Role of gender and physical activity level on cardiovascular risk factors and biomarkers of oxidative stress in the elderly. *Oxid. Med. Cell. Longev.* **2020**, 2020, 1315471. [CrossRef] [PubMed]
- 103. Gawron-Skarbek, A.; Chrzczanowicz, J.; Nowak, D.; Gawor, R.; Kostka, T. Effects of two different types of single exercise modes on salivary C-reactive protein concentration, oxidative stress and antioxidant capacity in post-myocardial infarction patients. *Redox Rep.* **2021**, *26*, 29–34. [CrossRef] [PubMed]
- 104. Tofas, T.; Fatouros, I.G.; Draganidis, D.; Deli, C.K.; Chatzinikolaou, A.; Tziortzis, C.; Panayiotou, G.; Koutedakis, Y.; Jamurtas, A.Z. Effects of cardiovascular, resistance and combined exercise training on cardiovascular, performance and blood redox parameters in coronary artery disease patients: An 8-month training-detraining randomized intervention. *Antioxidants* 2021, 10, 409. [CrossRef] [PubMed]
- 105. Shibata, A.; Izumiya, Y.; Yamaguchi, Y.; Kitada, R.; Iwata, S.; Ehara, S.; Izumi, Y.; Hanatani, A.; Yoshiyama, M. Increased oxidative stress during exercise predicts poor prognosis in patients with acute decompensated heart failure. *ESC Heart Fail.* **2021**, *8*, 3885–3893. [CrossRef] [PubMed]
- 106. Rotariu, D.; Babes, E.E.; Tit, D.M.; Moisi, M.; Bustea, C.; Stoicescu, M.; Radu, A.-F.; Vesa, C.M.; Behl, T.; Bungau, A.F.; et al. Oxidative stress—Complex pathological issues concerning the hallmark of cardiovascular and metabolic disorders. *Biomed. Pharmacother.* 2022, 152, 113238. [CrossRef] [PubMed]
- 107. Shih, N.L.; Cheng, T.H.; Loh, S.H.; Cheng, P.Y.; Wang, D.L.; Chen, Y.S.; Liu, S.H.; Liew, C.C.; Chen, J.J. Reactive oxygen species modulate angiotensin II-induced beta-myosin heavy chain gene expression via Ras/Raf/extracellular signal-regulated kinase pathway in neonatal rat cardiomyocytes. *Biochem. Biophys. Res. Commun.* 2001, 283, 143–148. [CrossRef]
- 108. Liu, Q.; Sargent, M.A.; York, A.J.; Molkentin, J.D. ASK1 regulates cardiomyocyte death but not hypertrophy in transgenic mice. *Circ. Res.* **2009**, *105*, 1110–1117. [CrossRef]
- 109. Zima, A.V.; Blatter, L.A. Redox regulation of cardiac calcium channels and transporters. *Cardiovasc. Res.* **2006**, 71, 310–321. [CrossRef]
- 110. Doughan, A.K.; Harrison, D.G.; Dikalov, S.I. Molecular mechanisms of angiotensin II-mediated mitochondrial dysfunction: Linking mitochondrial oxidative damage and vascular endothelial dysfunction. *Circ. Res.* **2008**, *102*, 488–496. [CrossRef]
- 111. Pollock, D.M. Endothelin, angiotensin, and oxidative stress in hypertension. Hypertension 2005, 45, 477–480. [CrossRef]
- 112. Seet, R.C.; Lee, C.-Y.J.; Chan, B.P.; Sharma, V.K.; Teoh, H.-L.; Venketasubramanian, N.; Lim, E.C.; Chong, W.-L.; Looi, W.-F.; Huang, S.-H. Oxidative damage in ischemic stroke revealed using multiple biomarkers. *Stroke* **2011**, *42*, 2326–2329. [CrossRef] [PubMed]
- 113. Chakravarti, B.; Chakravarti, D.N. Oxidative modification of proteins: Age-related changes. *Gerontology* **2007**, *53*, 128–139. [CrossRef] [PubMed]
- 114. Vesa, C.M.; Popa, L.; Popa, A.R.; Rus, M.; Zaha, A.A.; Bungau, S.; Tit, D.M.; Corb Aron, R.A.; Zaha, D.C. Current data regarding the relationship between type 2 diabetes mellitus and cardiovascular risk factors. *Diagnostics* **2020**, *10*, 314. [CrossRef] [PubMed]
- 115. Wray, D.W.; Uberoi, A.; Lawrenson, L.; Bailey, D.M.; Richardson, M.S. Oral antioxidants and cardiovascular health in the exercise-trained and untrained elderly: A radically different outcome. *Clin. Sci.* 2009, 116, 433–441. [CrossRef] [PubMed]
- 116. Iurciuc, S.; Avram, C.; Turi, V.; Militaru, A.; Avram, A.; Cimpean, A.M.; Iurciuc, M. Physical training, hemodynamic parameters and arterial stiffness: Friends or foes of the hypertensive patient? *In Vivo* **2016**, *30*, 521–528.
- 117. Gheorghe, G.; Toth, P.P.; Bungau, S.; Behl, T.; Ilie, M.; Pantea Stoian, A.; Bratu, O.G.; Bacalbasa, N.; Rus, M.; Diaconu, C.C. Cardiovascular risk and statin therapy considerations in women. *Diagnostics* **2020**, *10*, 483. [CrossRef]

118. Ardalani, H.; Jandaghi, P.; Meraji, A.; Moghadam, M.H. The effect of Cynara scolymus on blood pressure and BMI in hypertensive patients: A randomized, double-blind, placebo-controlled, clinical trial. Complement. *Med. Res.* **2020**, 27, 40–46. [CrossRef]

119. Ciumărnean, L.; Milaciu, M.V.; Negrean, V.; Orășan, O.H.; Vesa, S.C.; Sălăgean, O.; Iluţ, S.; Vlaicu, S.I. Cardiovascular risk factors and physical activity for the prevention of cardiovascular diseases in the elderly. *Int. J. Environ. Res. Public Health* **2022**, *19*, 207. [CrossRef]