

# The interactions of age, genetics, and disease severity on tacrolimus dosing requirements after pediatric kidney and liver transplantation

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## Abstract

**Purpose** In children, data on the combined impact of age, genotype, and disease severity on tacrolimus (TAC) disposition are scarce. The aim of this study was to evaluate the effect of these covariates on tacrolimus dose requirements in the immediate post-transplant period in pediatric kidney and liver recipients.

**Methods** Data were retrospectively collected describing tacrolimus disposition, age, *CYP3A5* and *ABCB1* genotype, and pediatric risk of mortality (PRISM) scores for up to 14 days post-transplant in children receiving liver and renal transplants. Initial TAC dosing was equal in all patients and adjusted using therapeutic drug monitoring. We determined the relationship between covariates and tacrolimus disposition.

**Results** Forty-eight kidney and 42 liver transplant recipients (median ages 11.5 and 1.5 years, ranges 1.5–17.7 and 0.05–14.8 years, respectively) received TAC post-transplant. In both

transplant groups, younger children (<5 years) needed higher TAC doses than older children [kidney: 0.15 (0.07–0.35) vs. 0.09 (0.02–0.20) mg/kg/12h,  $p=0.046$ , liver: 0.12 (0.04–0.32) vs. 0.09 (0.01–0.18) mg/kg/12h,  $p=0.038$ ]. In kidney but not liver transplants, *CYP3A5* expressors needed significantly higher TAC doses than nonexpressors [0.15 (0.07–0.20) vs. 0.09 (0.02–0.35) mg/kg/12h,  $P=0.001$ ]. In these patients, age and *CYP3A5* genotype were independently associated with TAC dosing requirement. In liver, but not kidney transplant patients, homozygous *ABCB1* T-T-T haplotype carriers needed higher TAC doses than noncarriers [0.26 (0.15–0.32) vs. 0.11 (0.01–0.25) mg/kg/12h,  $p=0.013$ ]. **Conclusion** *CYP3A5* genotype may explain variation in tacrolimus disposition early after transplant in pediatric kidney recipients, independent of age-related variation. In contrast, in pediatric liver recipients, variation in tacrolimus disposition appears related to age and *ABCB1* genotype. These

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findings illustrate the importance of the interplay among age, genotype, and transplant organ on tacrolimus disposition.

**Keywords** Tacrolimus · Liver transplant · Kidney transplant · CYP3A5 · ABCB1 · Pharmacokinetics · Pediatrics

## Introduction

Tacrolimus is a potent immunosuppressive drug, used in most pediatric solid organ transplant recipients to prevent transplant rejection. This drug is characterized by a narrow therapeutic window and high inter- and intraindividual variability in disposition [1]. Drug concentrations can show up to 100-fold variation in individual patients. Transplant rejection rates as well as most of the adverse events associated with tacrolimus (including nephrotoxicity, neurotoxicity, hypertension, infections, and lymphoproliferative disease) may be, at least partially, concentration-related [2]. These serious adverse events associated with tacrolimus therapy provide a compelling rationale for the use of rigorous therapeutic drug monitoring. This is especially true because, with a tacrolimus half-life of up to 40 h, it may take 3–5 days before a tacrolimus dosing adjustment results in a new, adequate blood concentration [2]. All this time the patient may be at an increased risk for transplant rejection or adverse events.

Tacrolimus pharmacokinetics has been extensively investigated in adults. However, data in children are more limited [3]. Young children (1–6 years of age) appear to need higher doses per kilogram body weight of tacrolimus than older children and adults to maintain similar trough concentrations. The reason for this age-related faster clearance rate is unknown.

In addition to age-related differences in tacrolimus pharmacokinetics, large inter- and intraindividual variation is observed due to other factors. For example, genetic variation in CYP3A5 has been shown to explain a large part of the variation in tacrolimus disposition in both adult and small cohorts of pediatric solid organ transplant patients, as recently reviewed by Quteineh et al. [4–12]. The impact of genetic variation in ABCB1 gene activity on tacrolimus disposition remains controversial to date [13].

The impact of age and genetic variation appears to be weakened in the immediate post-transplantation period, while intraindividual variation appears larger [14]. This finding may be explained by other factors affecting tacrolimus disposition postoperatively. For example, CYP3A activity appears to be downregulated by proinflammatory factors, mainly TNF $\alpha$ , IL-1, and IL-6, which may be released after organ transplant surgery [15]. Yet, data regarding the effect of inflammation or disease severity on tacrolimus disposition in the immediate post-transplant period are lacking.

While limited data on the effect of age and CYP3A5 genotype on tacrolimus pharmacokinetics in children exist, the interplay of age, CYP3A5 genotype, and disease severity in relation to tacrolimus pharmacokinetics in pediatric solid transplant patients has not been elucidated, especially not in the immediate post-transplant period, when variation appears to be largest. This is important, as age-related changes in pharmacokinetics may obscure genetic variation. Similarly, inflammation and/or organ failure may reduce drug metabolism in patients who would otherwise have tacrolimus clearance comparable to adults or even faster. Due to this lack of data on the interplay of these possible covariates, the pediatric transplant patient is subjected to a higher risk of adverse events or transplant rejection.

The aim of this study was therefore to retrospectively determine the combined effect of age, CYP3A5 and ABCB1 genotype, and disease severity on tacrolimus dosing requirements and disposition in a cohort of pediatric liver and kidney recipients in the 2 weeks following transplantation. These two patient groups differ in that, in the liver transplant patients, recipient CYP3A5 and ABCB1 genotype may only affect intestinal CYP3A5 and ABCB1 activity, whereas in the kidney transplant patients, CYP3A5/ABCB1 genotype may affect both intestinal and liver CYP3A5/ABCB1 activity.

## Methods and materials

### Study design

Retrospective cohort study in pediatric liver and kidney transplant patients who received tacrolimus in the first 14 days after transplant.

### Patient population

Patients were eligible for study entry if they were 0–18 years of age at the time of liver or kidney transplant and received tacrolimus during the first 14 days post-transplant between 2000 and 2008 at the Hospital for Sick Children, Toronto, Ontario. From December 2006 to March 2008, all patients who visited the Outpatient Transplant Clinic or who were admitted to the hospital for transplant or follow-up were approached for informed consent. Transferred patients (out of the SickKids program) or deceased patients were not included. For deceased patients no DNA was available. The study was approved by the Hospital for Sick Children's Research Ethics Board. Informed consent and, if applicable, children's assent were obtained from all parents/legal representatives and/or patients.

### Immunosuppression protocol

All patients were started by the transplant physician on SickKid's transplant immunosuppression protocol therapy consisting of methylprednisolone, mycophenolate mofetil, and tacrolimus. A minority of patients received basiliximab or thymoglobulin (Table 1). Methylprednisolone (10 mg/kg) was intravenously administered at the time of graft reperfusion and gradually tapered after transplant (Table 1). Tacrolimus starting dose was 0.1 mg/kg orally twice daily in all patients and was subsequently adjusted by routine pre-dose tacrolimus concentrations to reach the preset target tacrolimus  $C_0$  concentration of 10–15 ng/ml for both groups. No predefined dose adaptation algorithm was used. Doses were adapted by the transplant physician in collaboration with the hospital pharmacist.

### Clinical data collection

The following patient characteristics were collected from SickKids electronic patient databases: transplant type, age at transplant, sex, weight, PRISM III score, duration of mechanical ventilation and ICU stay, chemical and hematological test results, and relevant co-medication (CYP3A5/ABCB1 inducers/inhibitors). Initial liver graft failure was defined as ASAT and/or ALAT >1,500 IU/l in the first 72 h after transplant, as defined by Nanashima et al. [16].

The following information was collected on tacrolimus: doses (date/time) and tacrolimus morning pre-dose blood concentrations (date/time).

The PRISM III score is a widely used and validated score in the pediatric intensive care setting assessing the severity of illness and the potential risk of mortality in critically ill children [17]. The PRISM is based on 14 routinely measured clinical and laboratory parameters with age-normalized normal ranges [17].

### Outcome measures

The primary outcome measure was median tacrolimus dosing requirement (normalized for body weight). Secondary outcome measures were median tacrolimus concentrations and concentration/dose ratio (normalized for body weight), as surrogate marker for tacrolimus clearance.

### Independent variables

Independent variables included recipient age, *CYP3A5* and *ABCB1* genotype, and PRISM score.

### DNA collection

For DNA analysis, blood for DNA (0.5 ml) was collected during regular blood work at the Transplant Outpatient Clinic. In cases where DNA collection from blood was not possible, saliva was collected using the Oragene TM DNA

**Table 1** Characteristics of the 90 pediatric liver and kidney transplant recipients

	Kidney			Liver			P-value
	Number	Median	Range	Number	Median	Range	
Male/female	29/19						0.39
Weight (kg)	48	33.7	10.7–80	42	10.9	2.6–64	<0.000
Age (years)	48	11.5	1.5–17.7	42	1.5	0.05–14.8	<0.000
PRISM	40	7	2–16	30	7.5	0–28	0.497
Days in ICU	48	2	1–14	42	6	2–14	<0.000
Days on ventilator	48		0–4	42	1	0–12	<0.000
Mean airway pressure (cm H <sub>2</sub> O)	3	13	11–14	31	11	8–18	0.214
Urinary output (mL/kg/u)	42	5.0	1.5–30.4	42	3	0.8–5.9	<0.000
Creatinin (μmol/L)	48	87.0	19–415	42	26.3	14–138	0.000
Albumin (g/L)	48	31.3	26.7–37	42	31.0	20–37	0.255
Hematocrit (L/L)	48	0.3	0.23–0.48	42	0.3	0.25–0.45	0.017
Methylprednisolone dose							
Day 1 (mg/kg/day)	48	2	0.7–3	42	3.9	1.8–4	
Day 7 (mg/kg/day)	48	1.6	1.5–1.8	42	0.3	0.1–0.3	
Day 14 (mg/kg/day)	48	1.2	1.0–1.5	42	0.28	0.05–0.3	
Basiliximab (yes/no)	0/48			6/36			
Thymoglobulin (yes/no)	2/46			4/38			

self-collection kit following the manufacturer's instructions (DNA Genotek, Kanata, ON).

#### Laboratory analysis

**Tacrolimus concentrations** Tacrolimus blood concentrations were determined in EDTA whole blood (0.25 ml), using LC-MS-MS as described previously [18].

**Genotyping** Genomic DNA was isolated from 0.2 ml ethylenediaminetetraacetic acid-treated whole blood using a MagnaPure LC (Roche Diagnostics, Mannheim, Germany). Genotyping was done using Taqman allelic discrimination assays on the ABI Prism 7900 HT sequence detection system (Applied Biosystems, CA, USA) on 1 ng of genomic DNA, as previously reported [19, 20]. Patients not carrying the *CYP3A5*\*3 allele were assigned the *CYP3A5*\*1/\*1 genotype by default. For the allelic variant G2677A/T, two separate assays were designed, one detecting G2677T, one detecting G2677A.

#### Statistical analyses

Data are presented as mean±SD or median (range) when the data were not normally distributed.

*CYP3A5* and *ABCB1* genotypes were tested for Hardy-Weinberg equilibrium. For the results, we compared mean differences using *t*-test or the rank-sum test when the data were not normally distributed. As the *ABCB1* polymorphisms are expected to be in linkage disequilibrium, multiple testing does not apply to the three different genotypes and one haplotype [21]. For comparison of categorical variables, chi-squared or Fisher's exact test was used, as appropriate. We studied correlations using Spearman's correlation test. With an expected cohort of 100 patients to be included (50 liver, 50 kidney), we a priori restricted our analysis to a maximum of four variables (age, *CYP3A5* and *ABCB1* genotype, and PRISM score), based on the general rule of thumb for multiple regression analysis that 10–20 patients are needed per variable.

To test the relative contribution and interaction of significant covariates from univariate analysis, multivariate analysis was performed using a general linear model. Data analysis was performed using SPSS statistical software (SPSS 16.0, Chicago, IL)

## Results

#### Patient inclusion and clinical characteristics

The total transplanted patient cohort at the Hospital for Sick Children transplant unit as of February 2008 was 124

kidney and 91 liver transplant patients. Of these patients, approximately 80% received tacrolimus in the first 2 weeks of transplant. All patients who received tacrolimus in the first 2 weeks after transplant were approached for informed consent when they visited the SickKids Transplant Outpatient clinic between November 2006 and February 2008. In addition, all patients who received a new transplant during this period were asked for informed consent. Informed consent rate was >95%. Patients that were missed were patients that moved out of the Toronto area or that were transferred to the adult transplant clinic for follow-up. In addition, patients that passed away could not be included, as we could not obtain DNA from these patients.

Between December 2006 and March 2008, 42 pediatric liver recipients and 48 kidney recipients were enrolled (Table 1). The liver recipients were much younger than the kidney recipients [median (range)]: 1.5 years (0.05–14.8) and 11.5 years (1.5–17.7),  $p<0.001$ , respectively. Median durations of mechanical ventilation and stay at the ICU were longer in the liver than in the kidney patients (Table 1).

#### Tacrolimus dosing requirements and disposition

Median tacrolimus doses were similar in both transplant groups, with large interindividual variation in tacrolimus doses over the first 14 days post-transplant: 0.1 mg/kg (range 0.01–0.35, IQR 0.08–0.15) for kidney and 0.1 mg/kg (range 0.01–0.31, IQR 0.08–0.16) for liver recipients (Table 2). On average 9 tacrolimus concentrations (range 4–12) per patient were available for the first 14 days post-transplant. Median concentration/dose ratios (as surrogate measure of clearance rate) also showed large interindividual variation: 78 (range 25–280, IQR 45–108) ng/ml\*mg/kg/12h for kidney and 87 ng/ml\*mg/kg/12h (range 25–280, IQR 58–139) for liver recipients.

Less than 40% of all tacrolimus concentrations were in the predefined therapeutic target range (10–15 ng/ml), as shown in Fig. 1. In the kidney recipients, more than 50% of all concentrations were below the therapeutic target range. In the liver recipients, 33% of concentrations were below and 25% of all concentrations above this range. In this group, very high concentrations were observed.

#### Effect of *CYP3A5* and *ABCB1* genotype on tacrolimus dosing and concentrations

DNA for both *CYP3A5* and *ABCB1* genotyping was available for 39 kidney and 32 liver recipients. Missing results were due to limitations in sample collection logistics and DNA analysis. The genotyped and nongenotyped groups did not differ in age, gender, weight, or tacrolimus exposure. DNA for *CYP3A5* genotyping was available for

**Table 2** Tacrolimus (tac) dosing characteristics of the 90 subjects by transplant type

	Kidney			Liver			P-value
	Number	Median	Range	Number	Median	Range	
Tac samples per patient	48	9	2–11	42	9	5–12	0.221
Median tac trough level (ng/mL)	48	9	2.8–15.5	42	11.5	5–15	0.000
Median tac dose (mg/kg)	48	0.1	0.02–0.35	42	0.1	0.01–0.32	0.389
Conc/dose (ng/ml per mg/kg)	48	78	25–280	42	87	25–1,939	0.162
Tac concentrations <5 ng/mL	48	5	0–11	42	3	0–9	0.003
Tac concentrations 10–15 ng/mL	48	2	0–9	42	3	0–7	0.033
Tac concentrations >15 ng/mL	48	0	0–3	42	2	0–5	0.000
Upward dose changes	48	4	0–8	42	4	1–8	0.889
Downward dose changes	48	2	0–5	42	3	0–5	0.010

44 of 48 kidney recipients and for 35 of 42 liver recipients. Genotype distribution for kidney recipients was  $n=1$ , 15, and 28, respectively, for *CYP3A5*\*1/\*1, \*1/\*3, and \*3/\*3. Similarly, distribution in liver recipients was  $n=1$ , 12, and 22. Genotype distribution was in Hardy-Weinberg equilibrium for both groups ( $p=0.53$  and  $p=0.67$ , respectively).

In kidney recipients, *CYP3A5* genotype was significantly associated with median tacrolimus dosing requirements, number of dose changes, and concentration/dose ratio (Table 3). *CYP3A5* expressors needed higher tacrolimus doses [median 0.14 (range 0.07–0.21) vs. 0.09 (0.01–0.35)

mg/kg/12h,  $p=0.001$ ] than nonexpressors (Fig. 2). *CYP3A5* expressors also needed more upward dose changes, had lower median tacrolimus concentrations, and had lower C/D ratios (Table 3). In contrast, in liver recipients, we did not observe a relationship between recipient *CYP3A5* and tacrolimus dosing requirements (Fig. 2), concentrations, or C/D ratios (Table 3).

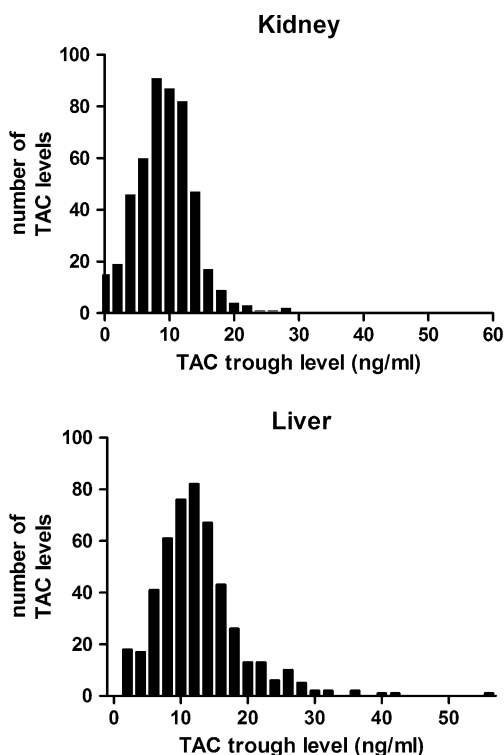
DNA for *ABCB1* genotyping was available for 45 kidney and 32 liver recipients. Genotype distribution is shown in Table 4. Genotype distribution was in Hardy-Weinberg equilibrium for all three polymorphisms tested in both transplant groups.

Upon inspection of the individual boxplots of dosing requirements vs. individual *ABCB1* SNPs, in liver, but not kidney, recipients homozygous for all three T alleles of polymorphisms, we noticed a trend towards higher dosing requirements. We then decided to analyze the relationship of the TT-TT-TT haplotype and found that liver recipients with this haplotype ( $n=3$ ) needed higher tacrolimus doses than the other patients ( $n=29$ ) [0.26 (0.15–0.32) vs. 0.11 (0.01–0.25);  $p=0.013$ ].

#### Effect of age and PRISM on tacrolimus dosing and concentrations

Children younger than <5 years of age needed higher tacrolimus doses per kilogram of body weight than older children after both kidney and liver transplant (Fig. 3) [kidney: median 0.15 (range: 0.07–0.35) vs. 0.09 (0.02–0.20) mg/kg/12h,  $p=0.046$ , liver: 0.12 (0.04–0.32) vs. 0.09 (0.01–0.18) mg/kg/12h,  $p=0.038$ ].

The PRISM mortality scores were not correlated with tacrolimus dosing requirements, tacrolimus trough concentrations, or concentration/dose ratios in both transplant groups. Tacrolimus disposition did not differ between pediatric liver transplant recipients with ( $n=7$ ) or without ( $n=35$ ) initial graft liver failure.



**Fig. 1** Distribution of tacrolimus concentrations in relation to therapeutic target level by transplant type

**Table 3** Comparison of tacrolimus (tac) dosing characteristics by *CYP3A5* genotype

	Kidney recipients		<i>P</i>	Liver recipients		<i>P</i>
	<i>CYP3A5</i> expressor ( <i>n</i> =16) Median (range)	<i>CYP3A5</i> nonexpressor ( <i>n</i> =28)		<i>CYP3A5</i> expressor ( <i>n</i> =13) Median (range)	<i>CYP3A5</i> nonexpressor ( <i>n</i> =22)	
Median tac trough level (ng/mL)	7.3 (4.7–11.1)	10.3 (2.8–15.5)	0.01	10.8 (8.1–14.8)	11.0 (5.0–15.4)	0.80
Median tac dose (mg/kg)	0.14 (0.07–0.20)	0.09 (0.02–0.35)	0.00	0.14 (0.06–0.32)	0.1 (0–0.26)	0.22
Conc/dose (ng/mL per mg/kg)	50.1 (25.5–92.5)	103.1 (31.3–280.3)	0.00	75.8 (38–136)	105 (24.7–1,939)	0.24
Tac concentrations <10 ng/mL ( <i>n</i> )	7 (2–11)	4 (0–7)	0.00	3 (1–9)	3 (0–7)	0.49
Tac concentrations 10–15 ng/mL ( <i>n</i> )	2 (0–5)	3 (0–9)	0.11	3 (1–7)	3 (0–7)	0.99
Tac concentrations >15 ng/mL ( <i>n</i> )	0 (0–2)	0 (0–3)	0.05	1 (0–5)	2 (0–5)	0.43
Upward dose changes ( <i>n</i> )	4 (3–8)	3 (0–7)	0.00	4 (1–8)	4 (1–6)	0.60
Downward dose changes ( <i>n</i> )	1 (0–4)	1 (0–4)	0.56	3 (0–5)	3 (0–5)	0.85

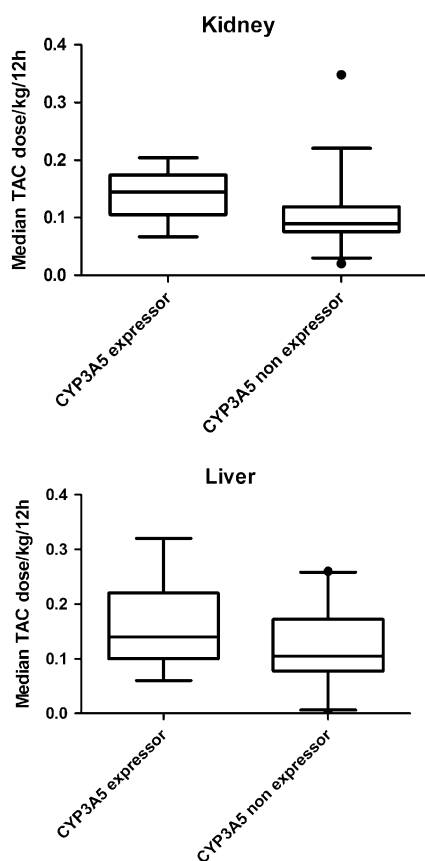
\* $P < 0.05$  *CYP3A5* expressors vs. nonexpressors

### Interplay of covariates on tacrolimus dosing requirements

In kidney recipients, in multivariate analysis, both younger age and *CYP3A5* expressor genotype were independently

associated with higher tacrolimus dosing requirements ( $r^2=0.362$ , corrected model  $p=0.0001$ , age F-value 4.6,  $p=0.001$ , *CYP3A5* genotype F-value 1.3,  $p=0.005$ ).

Children older than 5 years of age who were *CYP3A5* nonexpressors needed 0.09 (0.02–0.15) mg/kg/12h, whereas children younger than 5 years who were *CYP3A5* expressors needed 0.19 (0.18–0.2) mg/kg/12h tacrolimus. Patients younger than 5 who were nonexpressors as well as patients older than 5 who were expressors needed on average 0.13 mg/kg/12h (range 0.07–0.35 mg/kg/12h).



**Fig. 2** Relationship between recipient *CYP3A5* genotype and tacrolimus dosing requirements. Median (horizontal line), 5th and 95th percentiles (box), and range of tacrolimus dosing requirements by *CYP3A5* expressor genotype of the subjects. The denominators for each group are indicated. Of the kidney recipients, *CYP3A5* expressors needed more tacrolimus than nonexpressors ( $p < 0.05$ )

### Discussion

We aimed to study the interplay of age, genotype, and disease severity on tacrolimus disposition in the direct post-transplant period in 48 pediatric kidney and 42 pediatric liver transplant recipients. In a large proportion of our patients, tacrolimus concentrations were outside the therapeutic window in the first 14 days after transplant. Our results show that in pediatric kidney recipients, the patient's age and *CYP3A5* genotype correlate with median tacrolimus dosing requirements and concentration/dose ratio in the first 14 days after transplant. In contrast, in pediatric liver recipients, age at transplant and *ABCB1* genotype seem to correlate with tacrolimus dosing requirements. We did not find a relation between disease severity, as determined by the PRISM score, and dosing requirements for either transplant group. Interestingly, in the kidney recipients, both younger age and *CYP3A5* expressor genotype were independently associated with higher dosing requirements. This observation is important as it emphasizes the combined effect of genetic variation and ontogeny in relation to drug disposition. It has been shown in very young children that immaturity of drug metabolizing

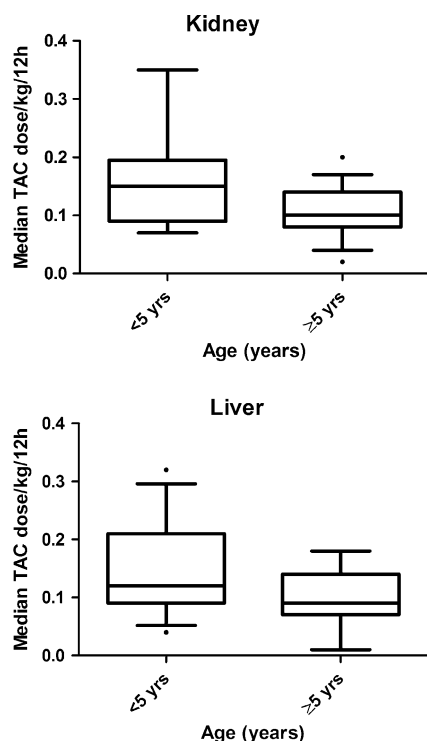
**Table 4** Relationship of *ABCB1* genotype and tacrolimus (tac) dosing and disposition

		Transplant type							
		Kidney				Liver			
		Median	Minimum	Maximum	Number	Median	Minimum	Maximum	Number
<b>ABCB1C3435T</b>									
Median tac dose (mg/kg)	CC	0.12	0.07	0.19	7	0.09	0.04	0.22	10
	CT	0.09	0.02	0.20	16	0.11	0.00	0.25	15
	TT	0.11	0.07	0.35	16	0.15	0.09	0.32	7
Conc/dose (ng/ml per mg/kg)	CC	61.26	38.68	90.72	7	85.46	24.73	364.00	10
	CT	101.16	25.48	280.31	16	96.95	33.20	1,939.80	15
	TT	50.07	31.34	156.40	16	72.14	41.13	209.89	7
Median tac trough level (ng/mL)	CC	9.10	4.70	11.05	7	11.18	5.00	14.40	10
	CT	9.38	4.45	15.50	16	12.05	8.10	15.45	15
	TT	8.43	2.80	13.05	16	10.40	8.10	15.10	7
<b>ABCB1C1236T</b>									
Median tac dose (mg/kg)	CC	0.11	0.02	0.19	11	0.11	0.07	0.22	11
	CT	0.09	0.04	0.20	16	0.10	0.00	0.25	15
	TT	0.11	0.07	0.35	12	0.15	0.06	0.32	6
Conc/dose (ng/ml per mg/kg)	CC	0.10	38.68	280.31	11	0.11	24.73	190.95	11
	CT	65.83	25.48	267.59	16	81.82	33.20	1,939.80	15
	TT	87.04	31.34	156.40	12	121.22	41.13	281.75	6
Median tac trough level (ng/mL)	CC	72.63	4.45	10.60	11	65.17	5.00	15.45	11
	CT	77.40	2.80	15.50	16	87.25	8.25	15.10	15
	TT	8.25	3.15	13.05	12	10.65	8.10	14.80	6
<b>ABCB1G2677AT</b>									
Median tac dose (mg/kg)	GG	0.11	0.02	0.17	10	0.11	0.06	0.22	12
	GT	0.09	0.04	0.20	17	0.10	0.00	0.25	13
	AT	0.15	0.15	0.15	1	0.08	0.08	0.08	1
	GA	0.19	0.18	0.19	2				
	TT	0.09	0.07	0.35	9	0.15	0.12	0.32	6
Conc/dose (ng/ml per mg/kg)	GG	71.61	38.92	280.31	10	83.06	24.73	281.75	12
	GT	90.72	25.48	267.59	17	136.14	33.20	1,939.80	13
	AT	67.77	67.77	67.77	1	133.77	133.77	133.77	1
	GA	49.97	38.68	61.26	2				
	TT	94.77	31.34	156.40	9	65.17	41.13	121.22	6
Median tac trough level (ng/mL)	GG	8.48	4.45	10.60	10	10.33	5.00	13.60	12
	GT	9.40	2.80	15.50	17	12.40	8.70	15.45	13
	AT	10.80	10.80	10.80	1	8.25	8.25	8.25	1
	GA	9.00	6.95	11.05	2				
	TT	8.90	3.15	13.05	9	10.60	8.10	14.80	6

enzyme activity may obscure a genotype effect on drug disposition [22, 23]. In our study, patients were well above the neonatal age, therefore such an effect of ontogeny on gene-related variation in drug metabolism is less likely. Our results suggest that tacrolimus starting dosing guidelines in children should reflect both age and *CYP3A5* genotype to reduce the time needed to reach therapeutic concentrations directly after transplant.

#### Effect of genotype

We were surprised to find a *CYP3A5* genotype effect so early after transplant, as other covariates such as post-surgical inflammation, renal impairment, and changes in albumin and hematocrit concentrations could have obscured pharmacogenetic variation in tacrolimus exposure and dosing. In another study in pediatric kidney patients with



**Fig. 3** Relationship between age and tacrolimus dosing requirements. Median (horizontal line), 5th and 95th percentiles (box), and range of tacrolimus dosing requirements by age of the subjects. Children less than 5 years of age needed more tacrolimus than older children in both transplant groups ( $p < 0.05$ )

similar results [24], patients were studied slightly later after transplant. A single tacrolimus AUC was determined 20  $\pm$  12 days after transplant. In contrast, tacrolimus dosing requirements were not different according to *CYP3A5* genotype in the first 3 months after transplant in adolescent kidney transplant patients ( $n=27$ ), but this lack of association may be due to a lack of power due to the small sample size studied [10].

In pediatric liver recipients, we could not identify a relationship between recipient *CYP3A5* genotype and tacrolimus dosing. The main reason for this lack of association is probably that variations in tacrolimus exposure are largely dependent on variation in hepatic metabolism and to a lesser extent on intestinal metabolism. Hence, donor *CYP3A5* genotype may be more important to explain variability in tacrolimus dosing requirements than the recipient's genotype. Unfortunately, we were not able to collect donor DNA to test this assumption. Our hypothesis is confirmed by data from Fukudo et al., who estimated tacrolimus pharmacokinetics, using trough concentrations, in a cohort of 65 Japanese pediatric living-related liver recipients up to 50 days post-transplant [25]. Donor *CYP3A5* genotype and recipient intestinal *ABCB1* expression but not recipient *CYP3A5* genotype were associated with (recovery of) tacrolimus oral clearance immediately after transplant.

In liver recipients, we found higher dosing requirements in children with the TT-TT-TT haplotype. Our results are in contrast with other studies in adults and children [26]. Most studies did not find a relationship between tacrolimus dosing requirements or disposition and *ABCB1* genotype in liver transplant patients. In one study in 51 pediatric liver recipients, carriers of different variant alleles (e.g., G2677T/A, C3425T, T-T-T haplotype) had higher C:D ratios (possibly reflecting lower clearance) < 3 years after transplant [13]. This result seems to contrast our results. We found a trend (not significant) towards lower C:D ratios in patients with the TT-TT-TT haplotype. A possible explanation for these contradicting results is that, early after liver transplant, intestinal *ABCB1* genotype in concert with intestinal *CYP3A4* largely determines variation in tacrolimus disposition. Lower intestinal *ABCB1* expression may result in more tacrolimus available for intestinal *CYP3A4* metabolism, leading to lower systemic tacrolimus concentrations. Later after liver transplant, when the liver is fully regenerated, hepatic *CYP3A5* may be more important. This is also supported by the lack of a relationship between *ABCB1* genotype and tacrolimus dosing requirements in our kidney recipients. This hypothesis stems from observations of a gender effect on the interaction between *ABCB1* 2677 genotype and *CYP3A4* expression. [27, 28]. There is a significant interaction between this variant (2677 SNP) and a 6 bp deletion variant of *NR1I2* (PXR) influencing *CYP3A4* expression [28]. To our knowledge, none of the other adult or pediatric studies specifically studied a gender effect of *ABCB1* genotype. Our sample size was too small to study the effect of gender in relation to the *ABCB1* genotype and tacrolimus disposition. In addition, a gender effect on *CYP3A4* expression is likely absent in prepubertal children. Another explanation for our finding of an *ABCB1* effect in liver recipients may be a type I error due to the small size of our patient population. Only 3 (TT-TT-TT haplotypes) patients had the variant genotype in a total genotyped population of 32 patients. For this reason, we refrained from studying the interaction between *ABCB1* genotype and age in the liver patients.

The lack of a relationship between *ABCB1* polymorphisms and tacrolimus dosing and disposition in our pediatric kidney recipients is consistent with studies from adult and pediatric kidney recipients [24, 29, 30]. In contrast, others did find a relationship between *ABCB1* genotype and tacrolimus disposition in adult kidney recipients [31–33].

#### Effect of age

In both kidney and liver patients, children younger than 5 years of age needed more tacrolimus than older children. Our findings are in line with previous studies in pediatric



liver and kidney recipients [3, 34, 35, 36, 37]. The reason why younger children need more tacrolimus than older children to reach the same tacrolimus target concentration is unknown. A wide range of age-related differences in drug disposition such as CYP3A4/5 metabolism and p-gp transport, volume of distribution, protein and erythrocyte binding, or renal function has been suggested [3]. To our knowledge, definite data are lacking on the impact of these possible mechanisms on tacrolimus disposition in children.

#### Variation in concentrations

In our patient population rigorous therapeutic drug monitoring was practiced, which is apparent from a median (range) of 9 (2–12) tacrolimus concentrations and 6 (1–11) dose changes in the first 14 days post-transplant. Despite this rigorous approach, less than 40% of all tacrolimus concentrations were within the predefined target range (10–15 ng/ml). Also, 33% of kidney and 76% of liver recipients had at least one supratherapeutic tacrolimus concentration. For the kidney recipients, this percentage is lower than reported in a UK pediatric renal transplant cohort (65 vs. 33% in our patients) [38]. This difference may be explained by lower initial tacrolimus doses in our patients than in UK patients (0.1 vs. 0.15 mg/kg twice daily). In pediatric liver transplant patients, only 27% of all concentrations were in the therapeutic range, in a study comparing buccal versus nasogastric administered tacrolimus [39]. As has been illustrated in adults, even in the context of a clinical trial, achieving target drug concentrations is challenging, especially early after transplant [40]. It is to be expected that incorporation of known covariates, such as *CYP3A5* genotype and age, in tacrolimus dosing regimens may reduce suboptimal tacrolimus dosing, as we found a relationship between genotype and nontherapeutic tacrolimus concentrations.

We hypothesized that part of the variation in tacrolimus disposition early after transplant may be due to variation in disease severity. We used a compound disease score (PRISM) to evaluate the role of disease severity on tacrolimus disposition but could not identify an effect. One limitation of the PRISM score is that it only takes into account the worst clinical and lab results within the first 24 h after ICU admission. Hence, variation in disease state after the first 24 h is not taken into account. In addition, as this is a compounded score, variations only in liver or kidney graft function that may impact tacrolimus metabolism may be missed using this score. We did, however, not find a difference in tacrolimus disposition when we compared patients with and without initial graft failure. In adult liver recipients, a more sensitive test of liver function, the LiMAX test, could predict tacrolimus trough levels [41].

#### Limitations

A limitation of our study is that tacrolimus dosing information was collected retrospectively from hospital databases. This may contribute to unexplained variation in dosing requirements, for example, when a child vomits and the drug needs to be re-administered. Also, it was not reported clearly how the drug was administered, by mouth, gastric, or duodenal tube, nor was it clear if the drug was given as suspension or as tablets. Furthermore, it was not known if the patient was receiving oral feeds or parenteral nutrition, which may also have contributed to unexplained variation in oral absorption. Previous studies have shown that different administration modes and fasting may also contribute to variation in oral tacrolimus pharmacokinetics [2, 42]. Further studies need to show if tacrolimus oral granules formulation could improve early GI absorption of the drug in children with consequently more stable exposure. Another limitation is that we did not include deceased patients, as it was not possible to collect DNA. Deceased patients may have suffered more frequently from transplant rejection and serious adverse events, resulting in death. It would have been interesting to include these patients, to determine if they were frequent outliers with respect to dosing requirements in relation to their *CYP3A5* genotype. In addition, the lack of impact of PRISM score on tacrolimus disposition in this cohort may have been due to this lack of inclusion of deceased patients, as this might have contributed to a larger variation in PRISM scores.

Based on our results, tacrolimus dosing in pediatric kidney transplant patients could be based on genotype and age. Starting doses of tacrolimus as high as 0.2 mg/kg/12h could be considered in young (<5 years of age) *CYP3A5* expressors (double the frequently used start dose). For the other age-genotype combinations, the effect was less pronounced (0.13 mg/kg/12h for older, *CYP3A5* expressors and younger, nonexpressors vs. 0.9 mg/kg/12h for older, nonexpressors).

In adults, a recent genotype-based dosing trial showed indeed that genotype-based dosing resulted in a shorter time to reach therapeutic levels (8 vs. 25 days in 75% of patients). However, no difference in rejection rate was found [43]. Additionally, adult *CYP3A5*\*3 expressors seem at higher risk to develop tacrolimus-induced nephrotoxicity, despite similar or lower tacrolimus exposure [44]. Hence, at this time genotype-based dosing in children might need to be restricted to trials evaluating its short- and long-term effect.

#### Conclusion

In conclusion, our results indicate that recipient *CYP3A5* genotype and age independently contribute to the variation

in tacrolimus dosing requirements and successful achievement of therapeutic tacrolimus concentrations in pediatric kidney recipients. In this population, individualized dosing schedules, incorporating both age and recipient *CYP3A5* genotype may be needed to optimize tacrolimus dosing. In pediatric liver recipients, our results suggest that recipient *ABCB1* genotype and age, but not *CYP3A5* genotype contribute to tacrolimus dosing requirements. Further studies should focus on the design and validation of genotype (recipient and donor) and age-based dosing regimens of tacrolimus, as well as on the effect of individualized dosing on outcome, i.e., transplant rejection rates and adverse events, such as nephrotoxicity.

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