Zootaxa 4363 (4): 544–552
http://www.mapress.com/j/zt/

Article

ISSN 1175-5326 (print edition)
ZOOTAXA
ISSN 1175-5334 (online edition)

https://doi.org/10.11646/zootaxa.4363.4.6
http://zoobank.org/urn:lsid:zoobank.org:pub:179512DA-1943-4F8E-931B-4D14D2EF91D2

The marbled crayfish (Decapoda: Cambaridae) represents an independent new species

FRANK LYKO

[1]*Division of Epigenetics, DKFZ-ZMBH Alliance, German Cancer Research Center, 69120 Heidelberg, Germany*
Correspondence: Deutsches Krebsforschungszentrum Im Neuenheimer Feld 580
69120 Heidelberg, Germany phone: +49-6221-423800 fax: +49-6221-423802
E-mail: f.lyko@dkfz.de

Abstract

Marbled crayfish are a globally expanding population of parthenogenetically reproducing freshwater decapods. They are closely related to the sexually reproducing slough crayfish, *Procambarus fallax*, which is native to the southeastern United States. Previous studies have shown that marbled crayfish are morphologically very similar to *P. fallax*. However, different fitness traits, reproductive incompatibility and substantial genetic differences suggest that the marbled crayfish should be considered an independent species. This article provides its formal description and scientific name, *Procambarus virginalis* **sp. nov.**

Key words: parthenogenesis, annulus ventralis, genetic analysis, mitochondrial DNA

Introduction

Marbled crayfish were first described in 2001 as the only known obligatory parthenogen among the approximately 15,000 decapod crustaceans (Scholtz *et al.*, 2003). The animals were first described in the German aquarium trade in the late 1990s (Scholtz *et al.*, 2003) and became widely distributed in subsequent years under their German name "Marmorkrebs". Stable populations have developed from anthropogenic releases in various countries including Madagascar, Germany, Czech Republic, Hungary, Croatia and Ukraine (Chucholl *et al.*, 2012; Jones *et al.*, 2009; Kawai *et al.*, 2009; Liptak *et al.*, 2016; Lokkos *et al.*, 2016; Novitsky & Son, 2016; Patoka *et al.*, 2016). On Madagascar, marbled crayfish form an aggressively expanding population that has invaded a large variety of habitats (Jones *et al.*, 2009). Due to their predicted ability to found large populations from single animals (Feria & Faulkes, 2011), marbled crayfish introductions have a significant potential to endanger indigenous crayfish species through competition or pathogen transmission (Chucholl & Pfeiffer, 2010; Jimenez & Faulkes, 2011; Keller *et al.*, 2014).

Several important characteristics of marbled crayfish have been described in detail, including morphology (Kawai *et al.*, 2009), embryonic development (Alwes & Scholtz, 2006; Seitz *et al.*, 2005), life history (Seitz *et al.*, 2005; Vogt, 2008; Vogt *et al.*, 2004), parthenogenetic reproduction (Martin *et al.*, 2007; Scholtz *et al.*, 2003; Vogt *et al.*, 2008) and a triploid karyotype (Martin *et al.*, 2016). Using morphological characters and molecular markers, Martin et al. identified the sexually reproducing slough crayfish *Procambarus fallax* from Florida as the closest relative of marbled crayfish and suggested the provisional name *Procambarus fallax* forma *virginalis* (Martin *et al.*, 2010). However, the International Code of Zoological Nomenclature (ICZN, 1999) excludes "forma" for names published after 1960 (Article 15.2). Consequently, the provisional name for marbled crayfish was also considered "unavailable" in a recent taxonomic summary of freshwater crayfish (Crandall & De Grave, 2017).

Importantly, recent findings have supported the notion that marbled crayfish should be considered as an independent species, for which the scientific name *Procambarus virginalis* was suggested (Vogt *et al.*, 2015). These findings are summarized below and complemented by a formal new species description.

 Accepted by J. Goy: 27 Nov. 2017; published: 13 Dec. 2017

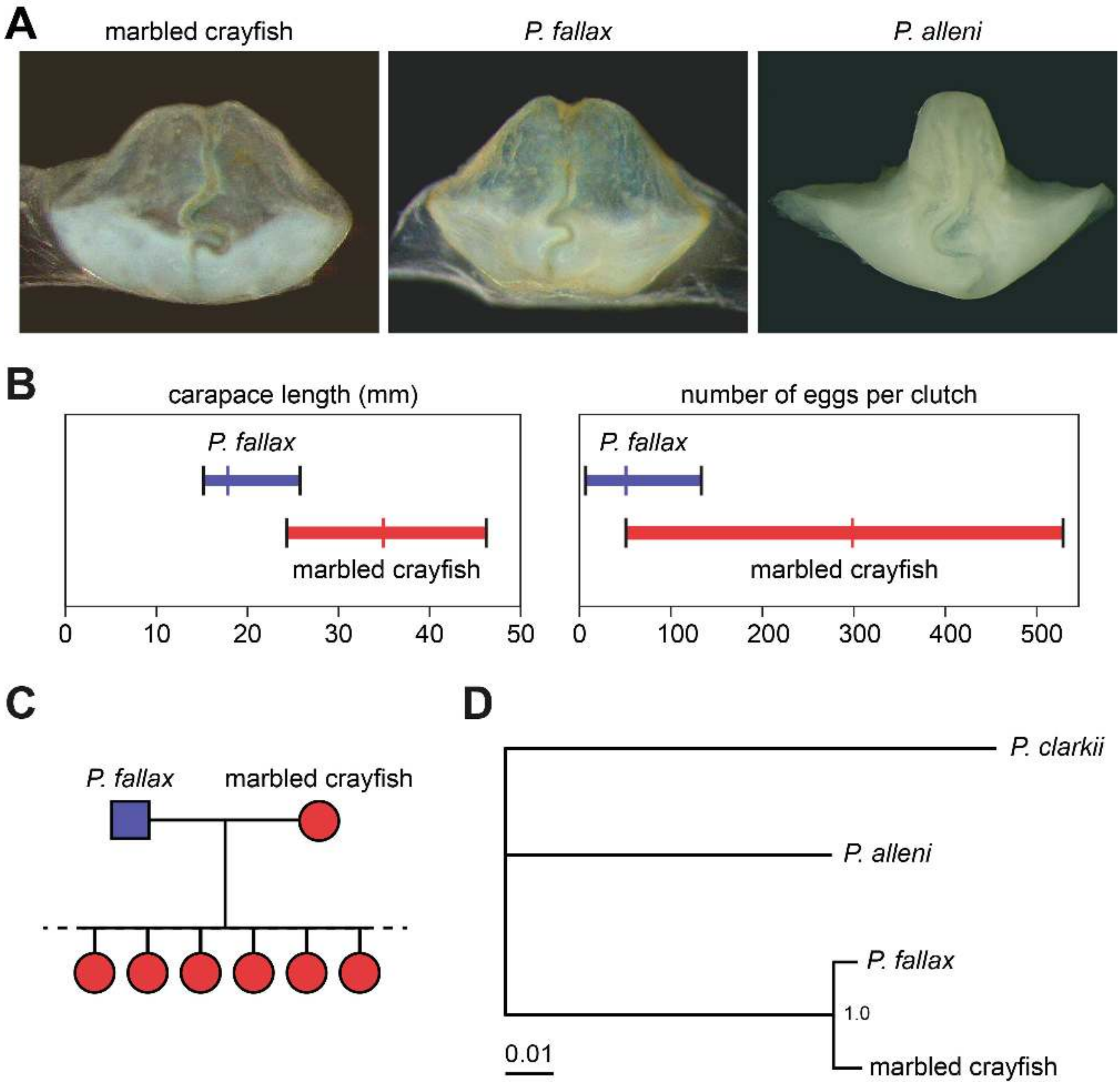

FIGURE 1. Comparison of morphological characters between marbled crayfish and *P. fallax*. (A) Annulus ventralis from exuvia of marbled crayfish, *P. fallax* and *P. alleni*. (B) Different life history traits in populations of ovigerous marbled crayfish (red) and *P. fallax* females (blue). Horizontal bars indicate ranges and vertical lines indicate mean values (m) and lower and upper range limits. (C) Parentage analysis in crossbreeding experiments of marbled crayfish and *P. fallax*. Colors indicate specific genotypes, as determined by microsatellite analysis. (D) Phylogenetic tree representing genetic relationships between marbled crayfish, *P. fallax*, *P. alleni* and *P. clarkii*. The tree is based on complete mitochondrial genome sequences, numbers indicate bootstrap values. All data adapted from (Vogt *et al.*, 2015) with technical details provided in the reference.

Results

Morphological characters and life history traits. Marbled crayfish are morphologically similar to *P. fallax* and to *P. alleni* (Kawai *et al.*, 2009; Martin *et al.*, 2010). A discrimination according to the taxonomically highly relevant first pleopods of breeding (Form1) males (Hobbs, 1972, 1989) is not possible as there are no male marbled crayfish (Scholtz *et al.*, 2003). However, marbled crayfish/*P. fallax* females can be discriminated from *P. alleni* by the annulus ventralis (Kawai *et al.*, 2009; Martin *et al.*, 2010). This important morphological character of cambarid females (Hobbs, 1972, 1989) is bell-shaped with an S-shaped sinus in marbled crayfish and *P. fallax*, but differently shaped in *P. alleni* (Fig. 1A). The specific morphology of the annulus ventralis is highly conserved among marbled crayfish from various sources (Kawai *et al.*, 2009; Martin *et al.*, 2010; Vogt *et al.*, 2015; Vojkovska *et al.*, 2014). Specific morphological characters that discriminate marbled crayfish from *P. fallax* are not known, which is likely related to the very recent speciation of marbled crayfish 20-30 years ago (Vogt *et al.*, 2015).

Their morphological similarities notwithstanding, life history traits have been found to be markedly different between marbled crayfish and *P. fallax* (Fig. 1B). Specifically, mean carapace length and clutch size were found to be significantly higher in adult marbled crayfish than in *P. fallax* (Vogt *et al.*, 2015). These findings suggest important phenotypic differences between marbled crayfish and *P. fallax* that are independent of the taxonomically relevant morphological characters.

Reproductive incompatibility. The marbled crayfish genome is triploid, which represents a major cytogenetic roadblock for meiotic chromosome segregation (Martin *et al.*, 2016; Vogt *et al.*, 2015). As such, most triploid organisms are apomictic parthenogens (Saura *et al.*, 1993) and thus reproductively isolated from sympatric sexually reproducing relatives. In crossbreeding experiments, marbled crayfish females and *P. fallax* males showed typical courtship behavior and mating (Vogt *et al.*, 2015). However, offspring of marbled crayfish females that had been mated with *P. fallax* males were exclusively female and exclusively showed the microsatellite markers of the marbled crayfish mother (Fig. 1C). These findings are in agreement with the notion that the *P. fallax* male did not contribute to the genome of the offspring and that the progeny is the product of apomictic parthenogenesis in the marbled crayfish mother. Taken together, the data suggest that marbled crayfish are reproductively isolated from *P. fallax* (Vogt *et al.*, 2015).

Genetic differences. Genetic analysis can be used to reveal independent species in the absence of distinctive morphological features (Bickford *et al.*, 2007). Furthermore, convergent evolution of morphological characters appears to be common in cambarids (Breinholt *et al.*, 2012), which further illustrates the value of genetic information for crayfish taxonomy (Owen *et al.*, 2015). Complete mitochondrial genome sequencing of marbled crayfish from diverse sources revealed their genetic homogeneity and strongly suggested a single origin of the known marbled crayfish population (Vogt *et al.*, 2015). In addition, mitochondrial genome sequence analysis identified considerable genetic differences between marbled crayfish and *P. fallax* (Fig. 1D). This was further supported by comparative whole-genome sequencing of marbled crayfish and their closest relatives, *P. fallax* and *P. alleni* (Gutekunst *et al.*, submitted). Altogether, these findings are consistent with the notion that marbled crayfish represent an independent species.

***Procambarus virginalis*, new species**

http://zoobank.org/NomenclaturalActs/CFEE97E8-558A-4A86-8496-85CF86A4716C

Marbled crayfish
Figures 2–3
Tables 1–2

Diagnosis. Body pigmented, eyes well developed. Rostrum with marginal spine and lacking median carina. Carapace with cervical spine. Areola 5 to 7 times as long as wide, constituting 30 to 35 percent carapace length (Tab. 1). Suborbital angle obtuse and weak. Postorbital ridge well developed with cephalic spine. Hepatic area punctate. Antennal scale approximately 2.5 times as long as wide, widest at midlength. Annulus ventralis bell-shaped, about 1.6 times as broad as long, bisected by narrow furrow leading caudally into median depression. Sinus originating on median line, continuing longitudinally on anterior half, then curving dextrally before curving caudally before continuing and terminating on median line. Sternum immediately cephalic to annulus with no tubercles or projections and not overhanging annulus. Unadorned bell-shaped postannular sclerite with central longitudinal furrow, width similar to annulus. First pleopods present. Mitochondrial DNA with guanine and cytosine at positions 8754 and 8783, respectively (GenBank accession number KT074364, see Tab. 2 for a list of discriminatory genetic variants).

TABLE 1. Morphometric parameters of *P. virginalis*.

	Holotype	Paratype 1	Paratype 2	Paratype 3	Paratype 4
Carapace					
length	53	45	40	41	33
width	24	21	18	19	15
height	26	22	19	20	16
Areola					
length	18	15	13	14	11
width	3	3	2	2	2
Rostrum					
length	16	13	11	12	8
width	9	8	6	6	5
Right chela					
palm length	16	12	9	10	7
palm width	11	9	7	9	6
dactyl length	18	14	12	13	10
Abdomen					
length	38	37	33	34	26
width	23	19	17	18	14

All measurements are indicated in mm.

Holotypic female. See Fig. 2. Cephalothorax ovate, slightly depressed. Abdomen narrower than cephalothorax (23 and 24 mm, respectively). Carapace slightly higher than wide at caudodorsal margin of cervical groove (Fig. 3A, B). Areola 6 times as long as wide, its length comprising 34% of the carapace length. Surface of carapace punctate. Rostrum slightly deflected ventrally with converging slender margins. Very short acumen, nearly reaching distal end of ultimate segment of antennular peduncle. Base of acumen marked by small marginal spines. Dorsal surface concave with punctations and no median carina. Postorbital ridge well developed, bearing spine at cephalic extremity. Suborbital angle weak and obtuse. Cervical spine twice the size of neighboring ones on caudal flank of cervical groove.

Abdomen slightly shorter than carapace (51 and 53 mm, respectively). Pleura of abdominal segments subtruncate. Telson and uropod with terminal setae. Telson with characteristic lateral spine and shallow lateral depressions. Mesial ramus with median ridge terminating in spine.

Antenna with small spiniform tubercle on ischium; flagellum reaching slightly beyond caudal margin of telson. Antennal scale (Fig. 3B) 2.5 times as long as wide, widest at midlength; lamellar area 2 times as wide than thickened lateral part. Third maxilliped extending cephalically to level of penultimate podomere of antennule, ventral surface of basal podomere densely covered with setae.

Right chela (Fig. 3C) elongate and depressed. Palm punctate and longer than wide (16 and 11 mm, respectively); mesial ridge with row of 8 small and subacute tubercles. Surface of fingers setiferous dentations. Opposable margin of fixed finger with 9 tubercles. Most distal tubercle at distal 1/4 of fixed finger larger and corneous. Surface of dactyl punctate, mesial margin with 4 small and subacute tubercles in proximal 1/4. Opposable margin of dactyl with row of 8 tubercles on proximal 2/3. Tip of dactyl elongate and pointed. Carpus of cheliped longer than broad with distinct oblique furrow dorsally, tuberculate mesially, dorsomesially and ventromesially. Ventral surface with 2 spines on distal margin. Merus with few weak spines dorsally, two larger ones distally. Mesial surface punctate, ventral surface with mesial row of 15 spines and lateral row of 13 spines. Ischium with dorsolateral tubercle and ventromesial row of 4 tubercles.

Annulus ventralis (Fig. 3D) as described in "Diagnosis".

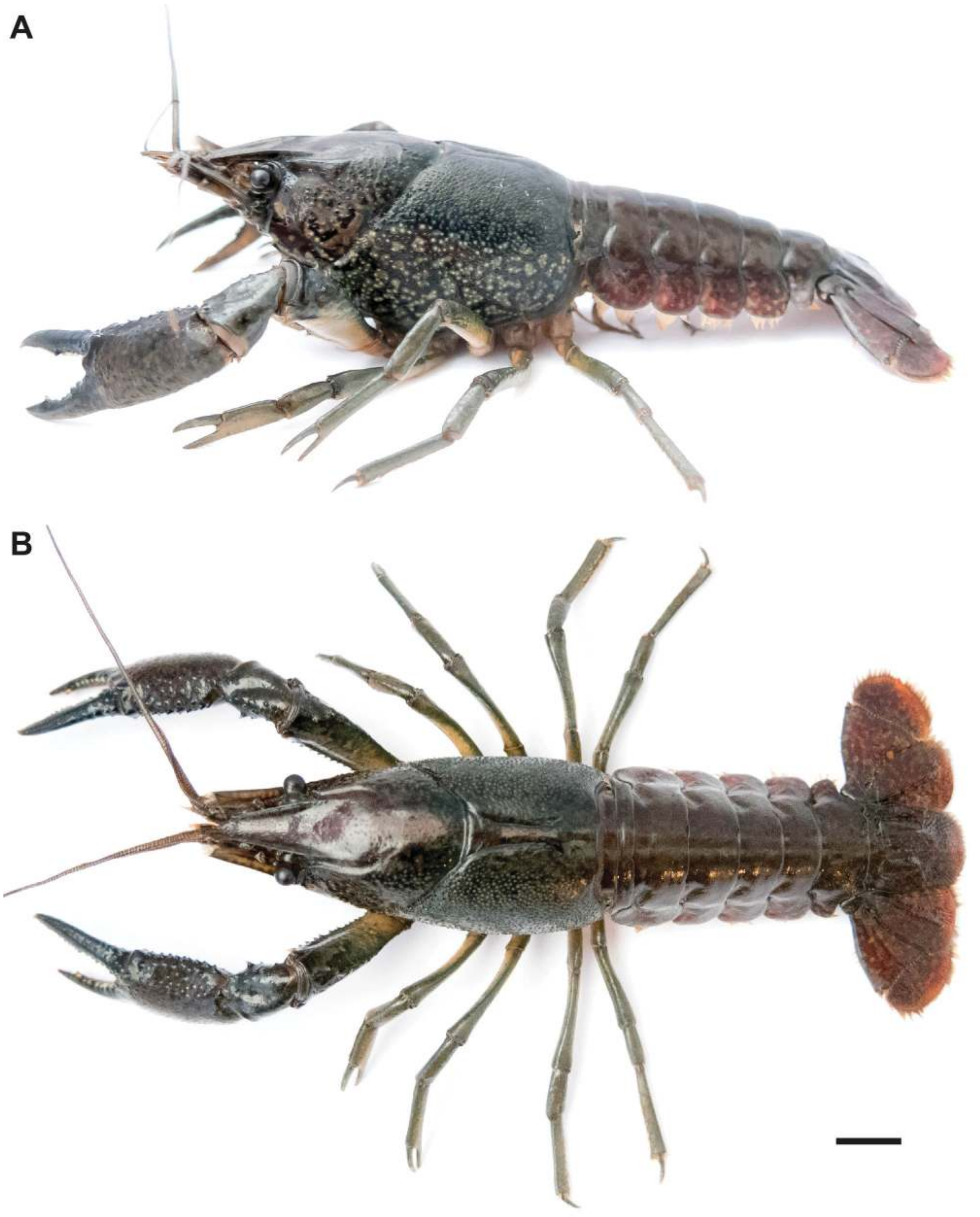

FIGURE 2. *Procambarus virginalis* new species holotype (A) lateral and (B) dorsal views. Bar: 1 cm.

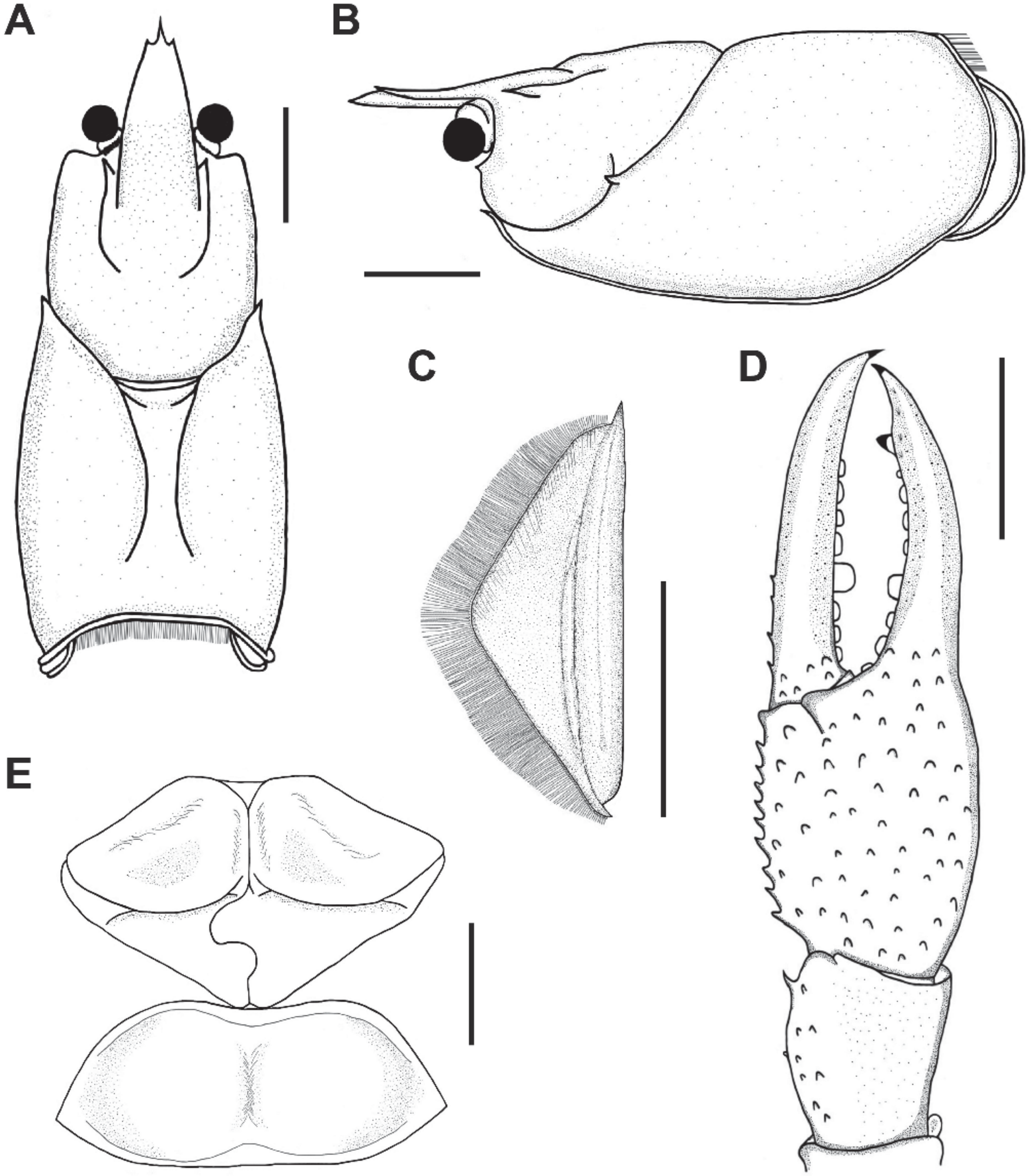

FIGURE 3. (A) Dorsal view of the carapace. (B) Lateral view of the carapace. (C) Dorsal view of the antennal scale. (D) Dorsal view of the right chela. (E) Ventral view of the annulus ventralis. Bars: 1 cm (A-D), 1 mm (E).

Color notes. The marmorated coloration of the carapace, which prompted the informal names "marbled crayfish" or "Marmorkrebs", is particularly prominent in aquarium stocks. It is often less pronounced in wild animals, which usually present in dark shades of brown or green with spotted patterns on the lateral cephalothorax and pleon (Fig. 2).

TABLE 2. Genetic markers for the discrimination of *P. virginalis*.

Position (KT074364)	*P. virginalis* (n=9)	*P. fallax* (n=5)	*P. alleni* (n=1)	*P. clarkii* (n=1)
3290	C	T	T	T
5856	C	T	T	T
8754	G	A	A	A
8783	C	T	T	T
10857	C	T	T	T
13366	T	C	C	C

Single nucleotide variants were determined by mapping of high-coverage Illumina sequencing data to the *P. virginalis* mitochondrial genome sequence (GenBank accession number KT074364). Numbers of analyzed animals are indicated below the species name.

Type locality. Lentic lake at Reilingen, Germany (Reilinger See, N49.296893, E8.544591). The holotype was selected from >150 animals that were collected using handnets and traps in August 2017. The lake has a size of 12 ha and a maximum depth of 20 m. The presence of the animals has been known by local fishermen for several years.

Disposition of types. The holotype (SMF 50818) and the paratypes 1 and 2 (SMF 50819) are deposited at the Senckenberg Natural History Museum (Frankfurt, Germany). The paratypes 3 and 4 remain in the author's collection.

Etymology. The name *Procambarus virginalis* is derived from the preliminary designation *Procambarus fallax* forma *virginalis* (Martin *et al.*, 2010) and reflects the species' unique parthenogenetic mode of reproduction.

Remarks. The oldest known record of *P. virginalis* is from a German biologist and hobby aquarist. In a personal conversation with the author (February 2017), he recalled obtaining an uncharacterized batch of "Texas crayfish" from a pet trader specializing in American insects and other invertebrates, at a trade fair in Frankfurt (Germany) in 1995. Animal numbers of this original stock increased rapidly and animals were subsequently distributed to other German aquarists, eventually reaching commercial traders and pet stores. The absence of male animals and the parthenogenetic mode of reproduction were soon recognized among aquarists and subsequently confirmed in the first scientific description of marbled crayfish (Scholtz *et al.*, 2003).

Acknowledgements

I thank various (former) members of my group for their help, in particular Ranja Andriantsoa for drawings, Sina Tönges for photographs, Julian Gutekunst for DNA sequence analysis and Günter Vogt for discussions. I also thank Ralf Bräuninger, Dominik Mattern and the Angelsportverein 1960 Reilingen e.V. for their cooperation and Zen Faulkes (The University of Texas Rio Grande Valley) for constructive comments. I am particularly indebted to Daniel P. Johnson for his advice during the drafting stage of this manuscript.

References

Alwes, F. & Scholtz, G. (2006) Stages and other aspects of the embryology of the parthenogenetic Marmorkrebs (Decapoda, Reptantia, Astacida). *Development Genes and Evolution,* 216 (4), 169–184.
https://doi.org/10.1007/s00427-005-0041-8

Bickford, D., Lohman, D.J., Sodhi, N.S., Ng, P.K., Meier, R., Winker, K., Ingram, K.K. & Das, I. (2007) Cryptic species as a window on diversity and conservation. *Trends in Ecology and Evolution,* 22 (3), 148–155.
https://doi.org/10.1016/j.tree.2006.11.004

Breinholt, J.W., Porter, M.L. & Crandall, K.A. (2012) Testing phylogenetic hypotheses of the subgenera of the freshwater crayfish genus Cambarus (Decapoda: Cambaridae). *PLoS ONE,* 7, e46105.
https://doi.org/10.1371/journal.pone.0046105

Chucholl, C., Morawetz, K. & Groß, H. (2012) The clones are coming – strong increase in Marmorkrebs *Procambarus fallax*

(Hagen, 1870) f. *virginalis* records from Europe. *Aquatic Invasions,* 7 (4), 511–519.
https://doi.org/10.3391/ai.2012.7.4.008

Chucholl, C. & Pfeiffer, M. (2010) First evidence for an established Marmorkrebs (Decapoda, Astacida, Cambaridae) population in Southwestern Germany, in syntopic occurrence with *Orconectes limosus* (Rafinesque, 1817). *Aquatic Invasions,* 5 (4), 405–412.
https://doi.org/10.3391/ai.2010.5.4.10

Crandall, K.A. & De Grave, S. (2017) An updated classification of the freshwater crayfishes (Decapoda: Astacidea) of the world, with a complete species list. *Journal of Crustacean Biology,* 37 (5), 1–39.
https://doi.org/10.1093/jcbiol/rux070

Feria, T.P. & Faulkes, Z. (2011) Forecasting the distribution of Marmorkrebs, a parthenogenetic crayfish with high invasive potential, in Madagascar, Europe, and North America. *Aquatic Invasions,* 6 (1), 55–67.
https://doi.org/10.3391/ai.2011.6.1.07

Gutekunst, J., Andriantsoa, R., Falckenhayn, C., Hanna, K., Stein, W., Rasamy, J.R. & Lyko, F. (submitted) Clonal genome evolution and rapid invasive spread of the marbled crayfish.

Hobbs, H.H.J. (1972) *Crayfishes (Astacidae) of North and Middle America. Biota of Freshwater Ecosystems. Identification Manual* 9. Environmental Protection Agency, Washington, DC, 177 pp.

Hobbs, H.H.J. (1989) An illustrated checklist of the American crayfishes (Decapoda: Astacidae, Cambaridae, and Parastacidae). *Smithsonian Contributions to Zoology,* 480, 1–236.
https://doi.org/10.5479/si.00810282.480

ICZN (1999) *International Code of Zoological Nomenclature.* Fourth Edition. The International Trust for Zoological Nomenclature, London, UK, 306 pp.

Jimenez, S.A. & Faulkes, Z. (2011) Can the parthenogenetic marbled crayfish Marmorkrebs compete with other crayfish species in fights? *Journal of Ethology,* 29 (1), 115–120.
https://doi.org/10.1007/s10164-010-0232-2

Jones, J.P.G., Rasamy, J.R., Harvey, A., Toon, A., Oidtmann, B., Randrianarison, M.H., Raminosoa, N. & Ravoahangimalala, O.R. (2009) The perfect invader: a parthenogenic crayfish poses a new threat to Madagascar's freshwater biodiversity. *Biological Invasions,* 11 (6), 1475–1482.
https://doi.org/10.1007/s10530-008-9334-y

Kawai, T., Scholtz, G., Morioka, S., Ramanamandimby, F., Lukhaup, C. & Hanamura, Y. (2009) Parthenogenetic alien crayfish (Decapoda: Cambaridae) spreading in Madagascar. *Journal of Crustacean Biology,* 29 (4), 562–567.
https://doi.org/10.1651/08-3125.1

Keller, N.S., Pfeiffer, M., Roessink, I., Schulz, R. & Schrimpf, A. (2014) First evidence of crayfish plague agent in populations of the marbled crayfish (*Procambarus fallax* forma *virginalis*). *Knowledge and Management of Aquatic Ecosystems,* 414 (15), 1–8.
https://doi.org/10.1051/kmae/2014032

Liptak, B., Mrugala, A., Pekarik, L., Mutkovic, A., Grula, D., Petrusek, A. & Kouba, A. (2016) Expansion of the marbled crayfish in Slovakia: beginning of an invasion in the Danube catchment? *Journal of Limnology,* 75 (2), 305–312.
https://doi.org/10.4081/jlimnol.2016.1313

Lokkos, A., Muller, T., Kovacs, K., Varkonyi, L., Specziar, A. & Martin, P. (2016) The alien, parthenogenetic marbled crayfish (Decapoda: Cambaridae) is entering Kis-Balaton (Hungary), one of Europe's most important wetland biotopes. *Knowledge and Management of Aquatic Ecosystems,* 417 (16), 1–9.
https://doi.org/10.1051/kmae/2016003

Martin, P., Dorn, N.J., Kawai, T., van der Heiden, C. & Scholtz, G. (2010) The enigmatic Marmorkrebs (marbled crayfish) is the parthenogenetic form of *Procambarus fallax* (Hagen, 1870). *Contributions to Zoology,* 79, 107–118. Available from: http://www.ctoz.nl/cgi/t/text/get-pdf?c=ctz;idno=7903a03 (accessed 9 December 2017)

Martin, P., Kohlmann, K. & Scholtz, G. (2007) The parthenogenetic Marmorkrebs (marbled crayfish) produces genetically uniform offspring. *Naturwissenschaften,* 94 (10), 843–846.
https://doi.org/10.1007/s00114-007-0260-0

Martin, P., Thonagel, S. & Scholtz, G. (2016) The parthenogenetic Marmorkrebs (Malacostraca: Decapoda: Cambaridae) is a triploid organism. *Journal of Zoological Systematics and Evolutionary Research,* 54 (1), 13–21.
https://doi.org/10.1111/jzs.12114

Novitsky, R.A. & Son, M.O. (2016) The first records of Marmorkrebs [*Procambarus fallax* (Hagen, 1870) f. *virginalis*] (Crustacea, Decapoda, Cambaridae) in Ukraine. *Ecologica Montenegrina,* 5, 44–46. Available from: https://www.biotaxa.org/em/article/view/19706/19060 (accessed 9 December 2017)

Owen, C.L., Bracken-Grissom, H., Stern, D. & Crandall, K.A. (2015) A synthetic phylogeny of freshwater crayfish: insights for conservation. *Philosophical Transactions of the Royal Society London B: Biological Sciences,* 370 (1662), 20140009.
https://doi.org/10.1098/rstb.2014.0009

Patoka, J., Buřič, M., Kolář, V., Bláha, M., Petrtýl, M., Franta, P., Tropek, R., Kalous, L., Petrusek, A. & Kouba, A. (2016) Predictions of marbled crayfish establishment in conurbations fulfilled: Evidences from the Czech Republic. *Biologia,* 71 (12), 1380–1385.
https://doi.org/10.1515/biolog-2016-0164

Saura, A., Lokki, J. & Suomalainen, E. (1993) Origin of polyploidy in parthenogenetic weevils. *Journal of Theoretical Biology,* 163 (4), 449–456.
https://doi.org/10.1006/jtbi.1993.1130

Scholtz, G., Braband, A., Tolley, L., Reimann, A., Mittmann, B., Lukhaup, C., Steuerwald, F. & Vogt, G. (2003) Ecology: Parthenogenesis in an outsider crayfish. *Nature,* 421, 806.
https://doi.org/10.1038/421806a

Seitz, R., Vilpoux, K., Hopp, U., Harzsch, S. & Maier, G. (2005) Ontogeny of the Marmorkrebs (marbled crayfish): a parthenogenetic crayfish with unknown origin and phylogenetic position. *Journal of Experimental Zoology Part A: Comparative Experimental Biology,* 303A (5), 393–405.
https://doi.org/10.1002/jez.a.143

Vogt, G. (2008) Investigation of hatching and early post-embryonic life of freshwater crayfish by in vitro culture, behavioral analysis, and light and electron microscopy. *Journal of Morphology,* 269 (7), 790–811.
https://doi.org/10.1002/jmor.10622

Vogt, G., Falckenhayn, C., Schrimpf, A., Schmid, K., Hanna, K., Panteleit, J., Helm, m., Schulz, R. & Lyko, F. (2015) The marbled crayfish as a paradigm for saltational speciation by autopolyploidy and parthenogenesis in animals. *Biology Open,* 4, 1583–1594.
https://doi.org/10.1242/bio.014241

Vogt, G., Huber, M., Thiemann, M., van den Boogaart, G., Schmitz, O.J. & Schubart, C.D. (2008) Production of different phenotypes from the same genotype in the same environment by developmental variation. *Journal of Experimental Biology,* 211, 510–523.
https://doi.org/10.1242/jeb.008755

Vogt, G., Tolley, L. & Scholtz, G. (2004) Life stages and reproductive components of the Marmorkrebs (marbled crayfish), the first parthenogenetic decapod crustacean. *Journal of Morphology,* 261 (3), 286–311.
https://doi.org/10.1002/jmor.10250

Vojkovska, R., Horka, I., Tricarico, E. & Duris, Z. (2014) New record of the parthenogenetic marbled crayfish *Procambarus fallax* f. *virginalis* from Italy. *Crustaceana,* 87 (11–12), 1386–1392.
https://doi.org/10.1163/15685403-00003365