Review

Open Access

The modular systems biology approach to investigate the control of apoptosis in Alzheimer's disease neurodegeneration Lilia Alberghina* and Anna Maria Colangelo

Address: Department of Biotechnology and Biosciences, Laboratory of Neuroscience "R. Levi-Montalcini", University of Milano-Bicocca, 20126 Milan, Italy

Email: Lilia Alberghina* - lilia.alberghina@unimib.it * Corresponding author

Published: 30 October 2006 BMC Neuroscience 2006, **7**(Suppl 1):S2 doi:10.1186/1471-2202-7-S1-S2

© 2006 Alberghina and Colangelo; licensee BioMed Central Ltd.

This is an open access article distributed under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by/2.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Apoptosis is a programmed cell death that plays a critical role during the development of the nervous system and in many chronic neurodegenerative diseases, including Alzheimer's disease (AD). This pathology, characterized by a progressive degeneration of cholinergic function resulting in a remarkable cognitive decline, is the most common form of dementia with high social and economic impact. Current therapies of AD are only symptomatic, therefore the need to elucidate the mechanisms underlying the onset and progression of the disease is surely needed in order to develop effective pharmacological therapies. Because of its pivotal role in neuronal cell death, apoptosis has been considered one of the most appealing therapeutic targets, however, due to the complexity of the molecular mechanisms involving the various triggering events and the many signaling cascades leading to cell death, a comprehensive understanding of this process is still lacking.

Modular systems biology is a very effective strategy in organizing information about complex biological processes and deriving modular and mathematical models that greatly simplify the identification of key steps of a given process.

This review aims at describing the main steps underlying the strategy of modular systems biology and briefly summarizes how this approach has been successfully applied for cell cycle studies. Moreover, after giving an overview of the many molecular mechanisms underlying apoptosis in AD, we present both a modular and a molecular model of neuronal apoptosis that suggest new insights on neuroprotection for this disease.

I. Introduction: the modular systems biology approach

Systems biology aims at describing biological processes as interactions of molecular components structured by regulatory wirings, whose dynamics in various conditions might be predicted by using mathematical models and computer simulations. The goal is to achieve the ability to integrate in a comprehensive model of pathways and networks the huge amount of data coming from postgenomic analysis in order to gain a better understanding of biological processes and a presently unattainable predictive ability.

For relatively simple processes, such as glycolysis, the structure of the pathway is quite well defined and allows us to readily construct the corresponding model. Neverth-

less, it has to be underlined that also in this relatively well known situation the systems biology approach has been able to identify a previously underestimated rate limiting step [1].

For other more complex processes, such as cell cycle, apoptosis, differentiation, transformation, we lack both a complete molecular understanding and the logic of their regulatory wirings. In order to develop a systems biology approach able to foster the understanding of complex bioprocesses, our laboratory has developed a modular systems biology approach and tested its validity in cell cycle studies.

The notion is by now largely accepted that cellular processes are carried out by modules, subsystems of interacting molecules (DNA, RNA, proteins and metabolites) that perform a given function in a way largely independent from the context [2]. The various modules are linked by governing interactions that follow general design principles well known in engineering, such as positive and negative feedbacks, switch, thresholds, amplification, error correction, etc. [3].

The modular systems biology approach, which is extensively discussed in ref. [4], starts with a global functional analysis of the process in order to identify both the known players and the main control functions active in the process. Then a modular blueprint of the process is constructed by gathering all the information previously collected. This low resolution blueprint is then used to derive a mathematical model that, tested by simulation, will show whether it is able to capture the essential features of the process. In the negative case, one has to go back to the drawing board and reconsider the blueprint structure; in the positive case, one can move on to identify the molecular components of each module following the so called 4M Strategy: Mining of literature and data banks; Manipulation by genetic means and by environmental changes of the module structure and function; Measurements of all putative regulatory components supposed to be active in the module (i.e. estimation of their concentration, localization, state of activation, time-course changes, etc.); Modeling and simulation: construction of a model at higher resolution than the previous one, in which the putative regulatory components of the module are linked by a specific flow of events. The simulation of this model will be compared with experimental results, in order to check its validity. It starts, therefore, an iteractive process of modeling, computer simulation, comparison with experimental data, new hypothesis-driven experiments, computer simulations, predictions that are going to be experimentally tested, which is the characteristic of the systems biology approach [5].

In this review we are going to apply the modular systems biology approach to give structure to the information available about the role of apoptosis in neurodegenerative diseases. Moreover, we construct the blueprint of neuronal apoptosis and present a tentative molecular model of the mechanisms underlying neuronal apoptosis. The roadmap describing the process is briefly reported in Figure 1.

In order to explain how the modular systems biology approach works in the definition of a molecular model of complex biological processes, we briefly summarize how

- Global functional analysis.
- Determination of the major functional modules involved in the process and of their regulatory connections.
- Determination of a basic modular blueprint of the process and validation of its dynamics.

Analysis of a module

- Environmental (i.e. changes in serum and growth factors availability) and genetic (i.e. gene dosage, deletion, point mutations) perturbations of the function of the module.
- Determination of content, localization, state of activity, etc. of putative components of the network.
- Post-genomic analysis.
- Development of a mathematical model of the module at a molecular level.
- Analysis of the model by simulations.
- Control and sensitivity analysis (robustness, connectivity, etc.).
- Predictions (to be experimentally tested).
- Iteractive replication of the previous steps until a satisfactory molecular model has been achieved.

Figure I

Scheme of the iterative roadmap of computational and experimental approaches applied in modular systems biology.

this approach has been successfully used in our laboratory for cell cycle studies.

2. The modular systems biology approach for cell cycle studies

Cell cycle requires the coordination of different processes: mass accumulation, DNA replication and faithful partitioning of genetic material. In the budding yeast Saccharomyces cerevisiae, the most relevant model organism for cell cycle studies, the coordination of mass accumulation with cell cycle progression relies on a cell sizer mechanism that allows DNA replication to start only when cells have reached a critical cell size, called Ps. Although this key regulatory control of cell cycle has been known for almost three decades [6-8], even recent cell cycle models [9,10] do not offer satisfactory molecular explanations for the setting of Ps. Besides, it is well known that external and metabolic signals are able to control the setting of the critical cell size [11], but we still ignore the detailed mechanisms that link signal transduction pathways to the cell cycle core molecular machine.

2.1. Defining the cell sizer regulatory network controlling entrance into S phase in yeast

In order to define the molecular mechanism underlying the cell sizer, the first step has been the drawing of a blueprint of the cell cycle [12] that states that the cell sizer is based on a threshold mechanism involving a growth-sensitive cyclin (Cln3) and an inhibitor of cyclin-dependent kinase (Cki), Far1, previously known to be involved in the arrest of the cell cycle in G1 in response to mating factors [13-15]. After the execution of the growth-sensitive threshold, a cascade of cyclin-dependent-kinases takes place, that controls the sequential progression of the cycle, terminated by an End function, that comprises events from the onset of mitosis to cell division and whose progression may be regulated by stress [12,16].

The second step has been the gathering of experimental evidence about the involvement of Far1 in cell cycle control: an increased level of Far1 increases cell size, whereas FAR1 Δ cells initiate bud emergence and DNA replication at a smaller size than wild type. Besides Cln3 Δ , FAR1 Δ and strains overexpressing Far1 do not show a drop in budding (a marker of the entrance into S phase) during an ethanol-glucose shift-up as wild type does. The activation of Cln3 · Cdk1, following the overcoming of the Cln3/ Far1 threshold, activates SBF- and MBF-dependent transcription. A second threshold given by Clb5,6 · Cdk1/Sic1 is required, together with the first one, to adjust the Ps value to carbon sources availability (i.e. low Ps in poor/ ethanol media, high Ps in rich/glucose media) [17].

A mathematical model accounting for the network identified so far has been implemented and shown to capture basic features of the G1 to S transition, such as the role of Cln3 and Far1 dosage on the setting of Ps [4].

The third step has focused more on the role of Sic1, another Cki, in the control of the entrance into S phase, focusing on the modulation by carbon source of the subcellular localization of Sic1 [18]. Sic1 and the cyclins Clb5 and Clb6 are the major regulators of the Cdk1 kinase activity required for the onset of DNA replication [19]. The complex Clb5, $6 \cdot$ Cdk1/Sic1, formed in G1 cells when Clb5,6 are synthesized due to MBF-mediated transcription, is activated by proteolysis of Sic1, primed by Cln1, $2 \cdot$ Cdk1 mediated phosphorylation on multiple sites and dependent on SCF complex [20].

The carbon source modulation of the setting of Ps mediated by Sic1 is dependent upon the fact that carbon source affects both the overall level and the subcellular localization of Sic1, that requires a newly characterized NLS to be imported in the nucleus. Besides, it has been shown that Sic1 facilitates nuclear accumulation of Clb5, thus playing a hitherto unrecognized positive role in promoting the G1 to S transition [18]. The kinetic parameters of the binding of Sic1 to cyclin/Cdk complex have been estimated, and evidence has been obtained *in vitro* showing that the strength of binding may be modulated by Sic1 protein phosphorylation [21].

It may be interesting to recall that Sic1 has been shown to be a functional homologue of the mammalian p27^{Kip1} with a conserved inhibitory domain [22]. Also p27^{Kip1}, together with p21^{Cip1}, has been shown to promote nuclear import of cognate cyclin D1/Cdk4, therefore the import of cyclins, that do not have obvious nuclear import signals, like cyclin D in mammals and Clb5 and Clb6 in yeast, appear to be imported into the nucleus piggy-backed on cargos (the complex cyclin/Cdk/Cki) that contains a protein (the Cki) with a NLS [23,24].

The fourth and last step, so far, has been to derive a mathematical model of the complex nucleus/cytoplasmic network, derived from the previous experimental findings integrated with the literature data, and to analyze various aspects of it by simulation [Barberis et al., 2006, submitted for publication]. Besides accounting for the behaviour of many known cell cycle mutants, the model shows that the critical cell size depends on the structure of the network and, in a more sensitive way, on few parameter values. The results of the simulations shed new light on the previously reported involvement of ribosome biogenesis on the setting of Ps [25-27].

3. Why modular systems biology approach for neurodegenerative diseases?

Taken together, the above mentioned results offer a strong support to the notion that the modular systems biology approach is very effective in fostering the identification of networks controlling complex bioprocesses and, therefore, it could be useful for a better understanding of neuronal apoptosis. Recently, mathematical models have been developed for CD95- and cytokine-induced apoptosis in mammalian cells [28,29], as well for Ca⁺⁺ signaling at dendritic spines and neurotransmitter receptor complexes underlying synaptic plasticity [30,31]. This review discusses the use of the modular systems biology approach to investigate apoptosis in neurodegenerative disorders and, in particular, Alzheimer's disease (AD). The basic idea is that structuring the apoptotic process in modules will allow us to better understand the regulatory wiring of the process.

As previously discussed, the first step requires the collection and organization of the information available about the process being considered. Therefore, the following paragraphs are going to gather current knowledge about functional analysis of apoptosis in AD neurodegeneration in order to derive the structure in modules of the blueprint, followed by an analysis of the key molecular players, in order to structure a tentative molecular model of neuronal apoptosis to be proposed for experimental validation.

4. Global functional analysis of apoptosis in AD

Neurodegenerative diseases are characterized by dysfunction and death of specific neuronal populations and, in many instances, by the presence of intracellular and extracellular aggregates of otherwise soluble proteins, a common feature in several disorders of the central nervous system (CNS), such as AD, Parkinson's (PD) and Huntington's diseases (HD) and Amyotrophic Lateral Sclerosis (ALS) [32-34].

Besides the genetic component of different molecular and cellular alterations, neuronal degeneration is contributed by the intervention of various factors and biochemical events that are relevant also to acute neuropathological conditions and including metabolic and oxidative stress, disruption of Ca⁺⁺ homeostasis and excitotoxicity, as well neurotrophic factor deprivation or traumatic injury [32,33,35-37]. For instance, energy depletion, excitotoxicity and oxidative stress are the major pathological mechanisms leading to neuronal death during ischemic stroke after oxygen and glucose deprivation due to transient or permanent reduction of the cerebral blood flow [38]. On the other hand, many biochemical mechanisms leading to cell death are shared by several neuropathological conditions. For example, excitotoxicity due to overactivation

of glutamate receptors represents a final common pathway for both acute neurological disorders, such as stroke, trauma and epileptic seizures, and chronic neurodegenerations like AD [35,39].

Regardless of which is the initial trigger and the neuronal population affected, neuronal death can occur through mechanisms involving necrosis or apoptosis, depending on the intensity and duration of the insult, both processes involving mitochondrial function [40,41]. Necrosis is a rapid form of death resulting from an acute toxic event that leads to complete failure of mitochondrial function and cellular homeostasis. Apoptosis is, instead, a slow and active process triggered by less severe insults and represents a mechanism for neurons to delay death, while activating various neuroprotective programs involving molecules and genes with anti-apoptotic function [42]. As discussed below, in this process, that is characterized by cell shrinkage, plasma membrane blebbing, nuclear chromatin condensation, DNA fragmentation and increase of a number of apoptotic markers, mitochondria play a pivotal role in cell decision, which is supposed to depend on a balance between apoptotic and anti-apoptotic signals. The initial trigger, in fact, induces a state of increased vulnerability to other toxic events and/or synergize with other apoptotic cascades, thus causing a persistent imbalance of neuronal homeostasis, mitochondrial dysfunction and neuronal death.

Increasing evidence indicates that apoptotic pathways occur in most neurological disorders. Besides the role of apoptosis in excitotoxicity and traumatic injury, there is evidence for apoptotic cell death in the penumbral region of ischemic brains [41,43,44]. Furthermore, apoptosis is currently regarded as the main form of death during neurodegenerative disorders like AD, PD, HD and ALS, as supported also by a number of studies in patients [45-47].

AD is a progressive neurodegeneration characterized by a disruption of synaptic function leading to a remarkable cognitive decline, including memory impairment and behavioural changes. The pathological hallmarks of AD include the presence of aggregates of amyloid- β peptide (A β) in neuritic plaques and intracellular neurofibrillary tangles (NFTs) of hyperphosphorylated tau (PHFtau), together with loss of neurons and synapses in the neocortex, hippocampus and other subcortical regions of the brain. These features are present both in sporadic AD and in early-onset familial form (FAD) which has been associated with genetic mutations of the β -amyloid precursor protein (APP) and the presenilin-1/2 (PS1/2) genes [34 and refs therein].

Due to the strong social impact of AD, a large number of studies have focused on the pathological mechanisms

underlying this debilitating disease that represents the most common form of dementia.

Neurochemical analyses indicate that neurotransmitter deficit, neuronal atrophy and synaptic dysfuction are early events in AD, as they occur also in subjects with Mild Cognitive Impairment (MCI), a high-risk pre-dementia state of the disease. Decrease of cholinergic markers and loss of basal forebrain cholinergic neurons has been observed in the brain of AD patients and senile dementia [48], and a strong correlation has been found between the degree of cognitive deficit and the decrease of synaptophysin immunoreactivity [49].

Brain imaging, electrophysiological and neuropsychological studies have shown that the degree of synaptic loss is correlated to levels of soluble oligomers of A β and occurs even before the appearance of A β plaques and NFT formation [50-54]. However, a reduction of neurotrophic support, in particular Nerve Growth Factor (NGF) [55-57] and Brain-Derived Neurotrophic Factor (BDNF) [58], is also believed to play a role in neuronal dysfunction, as discussed below.

Current therapies of AD are only symptomatic and include treatments with acetyl-cholinesterase inhibitors to improve cognitive function. However, effective therapies may require the development of pharmacological strategies that enable to modify and reverse the many biochemical alterations involved in the onset and progression of the disease.

Given its well established role in AD degeneration, apoptosis has lately gained much attention as a potential pharmacological target. However, although many of the apoptotic pathways have been extensively elucidated in terms of signaling [59-63] and differential gene expression [64,65], a comprehensive understanding of the apoptotic process in AD is still missing, due to the complex cross-talk between many different signaling cascades that are activated in a dynamic and coordinated fashion. Besides A β toxicity and deficit of neurotrophic support, neuronal death is triggered by a huge number of signals and biochemical mechanisms including microtubule transport impairment, decreased glucose metabolism, excitotoxicity, oxidative stress, mitochondrial dysfunction and inflammatory processes [37,59,61,66,67].

In order to apply the modular systems biology approach to apoptosis in AD, we will next review both the biochemical basis of the physio-pathological events underlying the two current AD hypothesis and the molecular mechanisms through which a number of diverse neurotoxic insults in AD may actually converge to convey apoptotic signals. Then, we will attempt for the first time to structure them in a molecular/modular model.

4.1 The "Neurotrophic Factor Hypothesis" and AD neurodegeneration

The "*Neurotrophic Factor Hypothesis*" [68,69] is linked to the discovery of NGF by R. Levi-Montalcini and the first observations of cell death for neurons deprived of their target of innervation [70,71]. The neurotrophic activity of target-derived NGF was then followed by the identification of other molecules including BDNF, Neurotrophin-3 (NT-3) and Neurotrophin-4/5 (NT-4/5), with neurotrophic activity on different neuronal populations. All neurotrophins share similar sequences and structures along with variable domains that determine the specificity of their biological activity resulting from the interaction with their cognate tyrosine kinase (Trk) receptor [[72] and refs. therein].

Thus, the phenomenon of *apoptosis*, that is central to the Neurotrophic Factor Hypothesis, was first observed in the context of the discovery of NGF and its role on differentiation and survival of sympathetic and sensory neurons of the peripheral nervous system [70,71]. During development, neurotrophins regulate neuronal survival and differentiation, and determine the *pattern* of innervation and the expression of proteins that are crucial to a specific neuronal phenotype, like neurotransmitters and neuropeptides, neurotransmitter receptors and ion channels. According to the neurotrophic hypothesis, the correct pattern of innervation is dependent upon competition of developing neurons for a limited supply of growth factors secreted by target tissues, thus ensuring a balance between the number of neurons and the size of the innervated target [68,69].

Neurotrophins activity, however, is essential throughout adult life and aging, where they play a key role in the regulation of neuronal function and synaptic plasticity, as well in neuroprotection and repair [73]. In this context, neurotrophins function can be revelant to the development of neuronal dysfunction and degeneration.

The potential relevance of NGF to neuronal loss in AD is based on a large body of evidence, including the existence of a strong correlation between NGF expression in neocortex and hippocampus and expression of TrkA receptor and choline acetyltransferase (ChAT) activity in the basal forebrain, both during development [74-76] and in the adult [77,78]. Cholinergic neurons atrophy and memory deficit in aged rat and in primates could be prevented by intraventricular infusions of NGF [79-82], while it is induced by intraventricular administration of anti-NGF antibodies [55] or disruption of the NGF gene [56]. Moreover, there is evidence for reduced or defective retrograde transport of NGF in aged rats [83], while increased cortical and hippocampal NGF levels with reduction of NGF in the nucleus basalis have been found in AD patients [36].

Neverthless, it was shown that excitotoxic lesion of target neurons does not induce death of basal forebrain cholinergic neurons, but cell shrinkage and decreased ChAT activity, suggesting that in the adult these neurons are no longer dependent on NGF for acute survival but only for maintenance of their cholinergic phenotype that is crucial to the modulation of cortical and hippocampal synaptic plasticity and learning and memory processes [84].

Recently, the potential role of NGF in AD degeneration has been supported by the AD model of anti-NGF transgenic mice [57,85]. The conditional expression of antibodies in the adult induces cholinergic deficit and the appearance of structural changes (amyloid plaques, PHFtau and NFTs) resembling those found in human AD, thus establishing a strong correlation between NGF deprivation and cholinergic dysfunction. Moreover, the cholinergic deficits of these mice could be rescued by intranasal administration of NGF [86].

Besides NGF, it must be acknowledged also the role of BDNF in AD, given the well established role of this neurotrophin in synaptic plasticity of the hippocampus and the requirement for BDNF in long-term potentiation (LTP) during learning and memory processes [87]. The potential role of BDNF in AD is supported by several studies showing a reduction of BDNF mRNA and protein levels in the cortex and hippocampus of MCI and severe AD patients [58,88]. As this reduction also occurs at early stage of the disease, it clearly demonstrates a role for decreased BDNF in the cognitive impairment in AD, that even precedes the reduction of ChAT activity [89].

An intriguing issue about the relevance of neurotrophins in the onset and/or progression of AD is the recent finding that the neurotrophin precursor proNGF appears to be the major form of NGF in the brain and that proNGF levels are increased in the parietal cortex of AD patients, even at early stages of the disease and in subjects with MCI [90]. ProNGF levels were also found to be enhanced after CNS injury, together with upregulation of the p75 neurotrophin receptor and increased p75-mediated apoptosis [91]. Enhanced p75 levels are known to occur in various injuries of the nervous system, as shown by both in situ hybridization and immunohistochemistry analyses [92-94]. In AD patients, increased p75 expression [95] is accompanied by a 50% reduction in TrkA levels [96], thus providing a challenging interpretation for decreased NGF survival signaling and increased p75-induced apoptosis.

It is well known that levels of expression of NGF and its receptors TrkA and p75 determine a balance between survival and apoptosis responses, the latter induced by NGF in the absence of TrkA receptor or upon alteration of the ratio between the two receptors in cells that express both TrkA and p75 [97]. The discovery that NGF may be secreted also as proNGF and that this unprocessed form can induce neuronal death through preferential interaction with the p75 receptor suggest that NGF may play a role in neuronal survival also in the adult where life-death decision would be also dependent upon the ratio between secreted mature and proNGF [98,99].

Recently, also BDNF was found to be secreted as proBDNF by cultured neurons and shown to induce apoptosis of sympathetic neurons [100]. However, proNGF and proBDNF do not induce apoptosis in all p75-expressing neurons, as their pro-apoptotic activity appears to depend on co-expression of sortilin, a member of the newly discovered family of Vps10p-domain receptors [100,101]. Sortilin is expressed both during embryogenesis and in the adult, in neurons and non neuronal cells of different brain areas including cortex, hippocampus, medial and lateral septum and amygdala [102].

These findings add a further level of complexity to that already given by the large number of different biological activities triggered by NGF upon activation of TrkA and p75 receptors, as well as that resulting from the complex cross-talk between the molecular intermediates of the signaling cascades activated individually by each receptor. Moreover, new questions arise as to whether proneurotrophins may have their own functional role under physiological conditions, or whether they result from postranslational alterations of neurotrophin synthesis during brain damage. Interestingly, plasmin levels are reduced in the brain of AD patients [103], although plasmin is a proteinase that cleaves proNGF and proBDNF in vitro, but its activity in vivo is still uncertain. Moreover, a recent report showed a role of proBDNF-p75 in the enhancement of long-term depolarization (LTD) mediated by upregulation of the NR2B subunit of the Nmethyl-D-aspartate (NMDA) receptors [104].

The neurotrophic hypothesis of AD has led to some phase I clinical trials. One study was based on intraventricular administration of NGF in three AD patients [105], while the last trial involved a rhNGF gene therapy approach that resulted in some beneficial effect on cognitive function [106].

Due to the invasiveness of these strategies, together with the poor NGF permeability across the blood-brain barrier, many studies have also focused on pharmacological regulation of endogenous NGF expression [107]. Recently, a phase II clinical trial has been carried out by administration of a γ -aminobutyric acid (GABA) receptor antagonist in 110 MCI patients. [108]. The drug increased NGF and BDNF mRNA and protein levels in the rat cortex and hippocampus, and improved cognitive function in the patients. A second trial in AD patients is still in progress.

It should be emphasized that a NGF-based therapy of AD may not be sufficient to fully reverse the AD pathology. However, due to its role in neuroprotection and repair, it is expected that NGF may provide a useful tool to ameliorate the symptoms and slow down the progression of the disease, yielding anyhow a relevant accomplishment as compared to the present therapeutic interventions. A more effective AD therapy might be better accomplished by targeting also other specific mechanisms of the disease cascade, such as apoptosis.

4.2 AD and the Amyloid hypothesis

As mentioned, a clear role for increased production and accumulation of soluble $A\beta$ in the pathogenesis of AD has been supported by a large number of studies [34,50-54].

Generation of neurotoxic species of A β results from an abnormal metabolism of APP, a membrane protein localized at the presynaptic terminals [[109] and refs therein]. A non-amyloidogenic pathway involves cleavage of APP by α -secretase and production of a soluble secreted form of APP (sAPP α), which has several biological activities including cell survival, neurite outgrowth and protection against excitotoxicity [110]. Production of amyloidogenic A β occurs, instead, through sequential cleavage of APP at the N-terminus by β -secretase and in the transmembrane domain by γ -secretase, a multimeric complex whose proteolytic activity is regulated by binding of PS1/2 at its catalitic site. Cleavage sites of γ -secretase are located at positions 40 and 42, thus generating A β 40 or A β 42, respectively [109].

Although A β 42 is constitutively produced in the brain, mutations in the APP or PS1/2 genes (which alter the cleavage specificity of γ -secretase) lead to increased production and accumulation of A β 42, that is neurotoxic even before its aggregation in Aβ fibrils [111]. In fact, soluble Aβ (including Aβ oligomers) can compromise basal synaptic transmission and LTP in the hippocampus even in the absence of A β deposits [53,54]. Synaptic activity and learning are impaired also in APP transgenic mice [112]. Additional studies in cultured cortical neurons have shown that sublethal concentrations of AB decrease the activity of cyclic AMP-response element binding protein (CREB) and downstream gene expression, such as the activation of the BDNF promoter [113]. CREB is also a transcription factor for the NGF promoter [114]. Sublethal doses of Aß also interferes with BDNF-mediated pathways involved in protection of cortical neurons from apoptosis [115]. Given the role of BDNF in LTP during learning and memory [87], these data establish a strong link between A β , trophic support and learning-memory impairment in AD degeneration.

The correlation between $A\beta$ and neurotrophins function is supported by several studies showing that the amyloidogenic pathway seems to be favored by a failure of TrkAmediated NGF signaling and dysfunction of cholinergic neurons [109], where NGF regulates ChAT activity, choline uptake, acetylcholine synthesis and release [116]. In PC12 cells, APP transcription and secretion have been found to be regulated by NGF and its receptors TrkA and p75: TrkA activation increases APP processing and secretion, while p75 signaling increases APP transcription [117]. Recently, these data have been confirmed by *in vivo* studies showing that accumulation of A β in aging might depend on a switch of TrkA-p75 expression and that p75 enhances A β production in a NGF-dependent manner by a ceramide-mediated mechanism [118].

Likewise, the APP secretory pathway, which generates nonamyloidogenic sAPP α , is increased by activation of the phosphoinositides (IP3)/Protein kinase C (PKC) signaling pathways through stimulation of muscarinic acetylcholine receptor (mAChR)/G protein coupled receptors (GPCR) both *in vitro* and *in vivo* [119,120]. Cortical and hippocampal M1/M3 mAChR signaling is impaired in AD, although the number of receptors is unaltered [121], suggesting that an impaired mAChR signaling may favor the amyloidogenic pathway of APP. On the other hand, a reciprocal correlation also exists, since A β has been shown to decrease acetylcholine synthesis and impair muscarinic cholinergic signaling in cortical neurons and cells SN56 derived from basal cholinergic neurons [122,123].

Another mechanism that can contribute to synaptic dysfunction in AD is due to downregulation of the 2-amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) and NMDA receptors. In fact, A β -mediated cleavage of AMPA subtype of glutamate receptors has been shown to occur in a caspase-dependent manner both in AD brains and in cultured neurons [124], while recently A β has been found to promote the endocytosis of NMDA receptors through a mechanism involving Ca²⁺ influx and dephosphorylation of the NR2B subunit [125].

Besides its role in synaptic dysfunction and apoptosis, $A\beta$ also contributes to the formation of NFTs of tau, a protein that binds and stabilizes microtubules. Hyperphosphorylation of tau results in the detachments of PHFtau from microtubules and its aggregation as paired helical filaments of the NFTs, thus leading to microtubule destabilization, disruption of the axonal transport and dysfunction

of organelles like the Golgi apparatus. Furthermore, as described in paragraph 5.5, both A β and PHFtau production are increased by a mechanism involving Wnt signaling and glycogen synthase kinase-3 β (GSK-3 β) [126,127], whose activity is enhanced in AD brains [128], thus providing a link between the two principal structural and pathological components of AD and decreased trophic support [129].

Based on the evidence supporting the A β hypothesis, potential therapeutic strategies have been developed involving immuno-therapy or pharmacological modulation of A β production. Both these approaches have also led to some phase I and II clinical trials, whose results indicate a reduction of A β levels in the plasma, but not in the cerebrospinal fluid, with γ -secretase inhibitors, and some cases of plaques regression following vaccination [130,131].

5. Key players in neuronal apoptosis

Current knowledge of the AD pathology indicate that multiple factors, genetic and environmental, play different roles in determining the conditions that trigger the dysfunction of the cholinergic neurons and the dynamic and coordinated activation of a series of cascades that eventually lead to neuronal death. Before analyzing the distinct events triggering apoptosis in AD, it should be emphasized that most of them include oxidative stress and mitochondrial dysfunction. Thus, these organelles play a key role in the amplification of responses to apoptotic stimuli and, therefore, in cell death decision.

The main groups of molecules involved in the apoptotic cascade include molecules as different as Bcl-2 family members [60], caspases [61,67], adaptor proteins controlling the activation of initiator caspases, such as the Apoptotic Protease Activating Factor-1 (APAF-1), and members of the Tumor Necrosis Factor (TNF) receptor family containing death domains (DD) (p75 and CD95/Fas) [63,132,133], as well transcription factors, proteins involved in the regulation of cell cycle and growth arrest [134-136], heat-shock proteins [137], etc. An important issue is that all these factors are activated through independent signaling pathways whose molecular components are engaged in a complex cross-talk, thus influencing each other in a positive or negative manner.

The Bcl-2 family includes pro-apoptotic (Bax and Bad) and anti-apoptotic (Bcl-2 and Bcl- X_L) proteins, whose expression and activity is regulated through different signaling pathways including trophic factors deprivation, A β toxicity, cytokines and DD receptors activation (Fas and p75) [60,132,138-140]. The central element of the apoptotic cascades is the activation of caspases [61,67,141], a family of cystein proteases (initiator and effector cas-

pases) that can activate DNase and promote the cleavage of enzymes, cytoskeletal proteins (actin and spectrin, fodrin) and ion channels, such as the AMPA receptors [124,140]. Among other proteases that are activated in AD, calpains are activated by increased intracellular Ca²⁺ levels [142].

All these molecules are involved in two main apoptotic pathways. The intrinsic pathway is centered on mitochondrial dysfunction and is regulated by Bcl-2/Bax proteins [60,139,143]. Traslocation of Bax homodimers to mitochondria leads to release of cytochrome *C* to the cytosol and its binding to APAF-1 with formation of the apoptosome and recruitment of procaspase-9. Oligomerization of APAF-1 promotes the allosteric activation of caspase-9. This initiator caspase can in turn activate the executioner caspases -3 and -7, thus leading to activation of proteolysis and DNA fragmentation. This pathway is activated by most of the known apoptotic triggers in AD, also because is central to cell death caused by oxidative stress.

An extrinsic pathway is, instead, activated by DD receptors [63,132,133]. For instance, interaction of Fas ligand (Fas L) with Fas/CD95 through association with an intracellular adaptor protein Fas-Associated Death Domain (FADD) is followed by activation of the initiator caspase-8 and the effector caspase-3 [141]. The two pathways are not mutually exclusive as, for instance, the mitochondrial intrinsic cascade can also be activated by caspase 8- mediated cleavage of Bid into a truncated tBid, followed by its traslocation to mitochondria where it causes the release of cytochrome *C* [144,145], as well the Apoptosis-Inducing Factor (AIF) which mediates a caspase-independent pathway [146,147].

5.1 Does NGF deprivation or proNGF play a role in AD apoptosis?

While NGF-mediated apoptosis during the development is well established [68,69], the role of NGF on survival of cholinergic neurons in the adult is recently gaining stronger experimental support [57,85]. In fact, besides the role of reduced retrograde transport of NGF [36,83] and the new evidence obtained in adult anti-NGF transgenic mice directly linking NGF deprivation to cholinergic deficits [57,85], there might be also a potential role of proNGF in triggering cell death through the p75 receptor [90] whose levels are increased in AD patients along with decreased TrkA expression [95,96].

The aim of this review is the construction of a molecular model of apoptosis in AD. Given the existence of a crosstalk between the apoptotic cascades activated by different apoptotic triggers and that their molecular components are involved in reciprocal modulation by the NGF signaling, we will first review the main molecular mechanisms underlying the control of neuronal survival and death by NGF/proNGF through their interaction with the TrkA and p75 receptors. Co-expression of the two receptors determines the formation of high affinity binding sites that potentiate TrkA signaling under conditions of low NGF concentration, however both receptors can also activate independent signaling cascades leading to survival or apoptosis, depending on cell context [rev. in ref. [148]].

TrkA signaling

The role of TrkA signaling in neuronal survival is known to be mediated by receptor dimerization and autophosphorylation on Tyr-490 and Tyr-785. Recruitment to p-Tyr490 of shc (src homologous and collagen-like) and other adaptor proteins is critical to the activation of two independent pathways: ras-Extracellular signal-Regulated Kinases 1/2 (ERK1/2) and Phosphatidylinositol-3 Kinase (PI3K)/Akt (Protein kinase B) pathways.

Ras-ERK1/2 activation leads to ERK1/2 translocation to the nucleus where they phosporylate Elk-1 and regulate the activity of transcription factors and co-activators, such as *c-fos* and CREB Binding Protein (CBP), involved in the control of gene expression. ERK1/2 can also phosphorylate Rsk (Ribosomal S6 Kinase), a cell cycle-regulated kinase that phosphorylates the S6 protein of the 40S ribosomal subunit. Rsk can in turn phosphorylate the IkB α subunit of Nuclear Factor-Kappa B (NF-kB) and CREB, whose activity is known to regulate genes involved in neurite outgrowth, synaptic plasticity and cell cycle.

The PI3K/Akt pathway, instead, regulates neuronal survival by modulating the phosphorylation of members of the bcl-2 family. Akt-mediated phosphorylation of Bad favors its interaction with 14-3-3 protein and prevents it from binding to Bcl-X_L. As a consequence, Bcl-X_L is free to bind Bax, thus preventing the formation of Bax homodimers that are pro-apoptotic. In fact, Bax homodimers traslocate to mitochondria and determine the alteration of the membrane potential and release of cytochrome *C* [149]. Akt-mediated survival has been shown to involve also phosphorylation and inhibition of procaspase-9 [150].

Other important substrates of Akt include Forkhead (FKH) transcription factors, GSK-3 β and p53 [151-153]. Non-phosphorylated FKHs proteins are localized in the nucleus and activate gene transcription, while phosphorylation by Akt results in their association to 14-3-3 protein and retention in the cytoplasm [151]. FKHs proteins regulate genes controlling cell cycle, cell metabolism, oxidative stress and cell death, such as Fas L, which has been shown to be overexpressed in AD brains [63] and involved in A β -induced apoptosis in neuronal cultures [63].

FKHs proteins have also been found to be phosphorylated by a mechanism involving Jun N-terminal Kinase (JNK) and p66Shc during A β -induced apoptosis [154], a process that seems to occur also *in vivo*. It has been suggested that relocation of FKHs to the cytosol would result in downregulation of target genes, such as Mn-Superoxide dismutase (MnSOD), thus leading to increased accumulation of reactive oxigen species (ROS), oxidative stress and cell death. Also GSK-3 β , which has been shown to be involved in apoptosis following NGF deprivation and A β neurotoxicity [128,129], is inactivated by phosphorylation through the PI3K/Akt signaling [152], as well by Wntmediated activation of PKC [155].

A second survival element of the PI3K/Akt pathway is through inhibition of the JNK signaling, as well as the activation of NF-kB. Phosphorylation and degradation of IkB, the inhibitory subunit that sequesters the NF-kB complex in the cytoplasm, releases NF-kB which can traslocate to the nucleus and activate the transcription of genes that promote resistance to apoptosis, such as MnSOD, Bcl-2 and members of the Inhibitor of Apoptosis Proteins (IAPs) family.

Phosphorylation of Tyr785 of TrkA receptors determines, instead, activation of phospholipase C- γ (PLC- γ) and production of two intracellular second messenger: diacylglycerol (DAG), a potent activator of PKC, and Inositol 1,4,5-P₃(IP3) which stimulates the release of Ca²⁺ from the endoplasmic reticulum (ER) and the activity of Ca²⁺-dependent proteins (Ca²⁺-calmodulin protein kinase) and phosphatases.

Experimental evidence of cell death following NGF deprivation have been obtained in vitro, mainly in PC12 cells and sympathetic neurons, where apoptosis following growth factor deprivation occurs mainly through an intrinsic pathway activated upon a failure of the TrkA/ PI3K/Akt signaling, followed by translocation of Bax to mitochondria and release of cytochrome [138,139,156]. Decreased TrkA/PI3K/Akt signaling would also result in the activation of JNK and induction of p53 [153], as well FKH-mediated transcription [151]. Although NGF deprivation has been found to induce Fas and Fas L expression, activation of an extrinsic pathway may contribute, but does not seem to be required for apoptosis after NGF deprivation [139].

p75 signaling

The p75 receptor belongs to the TNF receptor family because of the presence of a DD in the cytoplasmic region. This property confers to this receptor the ability to control cell survival/apoptosis depending on cell context, mainly the presence/absence of TrkA receptors [rev. in ref. [148]].

Increasing evidence has been accumulated so far about the role of p75 in neuronal death both *in vitro* and *in vivo* [93,153,157]. For instance, p75 has been shown to mediate apoptosis of developing sympathetic neurons [153], cultured cortical and hippocampal neurons [157], as well in mature oligodendrocytes [158]. Moreover, overexpression of p75 contributes to neuronal death of cholinergic neurons [93]. However, the precise molecular mechanisms of the p75-mediated cell death are still controversial. In fact, regardless of the presence of a DD, the apoptotic cascade is quite different from that induced by other TNF receptors family members. Signaling through p75 does not appear to activate caspase-8, the end point of a typical extrinsic pathway induced by Fas and other related members of the TNF receptors.

Several adaptor proteins have been found to interact with the DD of p75, including: NRIF1/2 (Neurotrophin-Receptor Interacting Factor 1 and 2) and NADE (Neurotrophin-Associated cell Death Executor) both involved in apoptosis; SC-1 (Schwann cell-1) and NRAGE (Neurotrophin Receptor-interacting M<u>AGE</u> homologue) that are associated to growth arrest and cell cycle events; RIP2 (Receptor-Interacting Protein 2), which possesses a CARD (Caspase Recruitment Domain) and TRAF-4/6 (TNF Receptor Associated Factor-4 and -6), both linked to survival responses [159-161].

The p75-mediated apoptosis have been found to occur mainly through activation of the sphingomyelinase/ceramide and JNK pathways [162]. The precise mechanism underlying sphingomyelinase activation by p75 is still unclear, but in oligodendrocytes there is a strict correlation between p75-mediated generation of ceramide and rac/JNK activation [158,163]. It is intriguing that ceramide has been shown to directly inhibit ERK and PI3K/ Akt signaling [164,165] thus evoking the same apoptotic cascade induced by NGF deprivation. This inhibition is part of the well known reciprocal regulation between the two NGF receptors, as ceramide signaling is also inhibited by PI3K/Akt. In cortical neurons ceramide induces activation of the mitochondrial apoptotic cascade and is associated to dephosphorylation of Akt, BAD, FKH and GSK-3β [165].

The mitochondrial intrinsic pathway following p75 signaling is also mediated by the JNK pathway through stabilization of p53 [153] which targets several genes, including Bax. Thus, p53 is a common component of the pathways activated by the two NGF receptors, both TrkA following NGF withdrawal and p75 signaling. The mitochondrial cascade leading to activation of the initiator caspase -9, and then caspases -3 and -7, has been found to be mediated by a JNK-dependent pathway following interaction of p75 with NRAGE [159]. JNK/p53 also triggers the

induction of Fas L which promotes apoptosis through binding to Fas receptor [132]. Fas L and Fas receptor expression are elevated both in AD and in cultured neurons treated with A β [63].

Another target of p75 signaling is the activation of NF-kB [166]. Recently, *in vitro* studies have shown that NF-kB activation can be mediated by the interaction of p75 with RIP2 and TRAF-6 [160,161]. TRAF-6 activates the protein kinase NFkB-Interacting Kinase (NIK), which phosphorylates the Inhibitor of IkB Kinase (IKK), thus leading to phosphorylation and degradation of IkB and translocation of NF-kB to the nucleus.

Induction of NF-kB by p75-mediated signaling has been observed in many cell types, under different conditions [166,167] and mostly linked to a survival function. In general, NF-kB is regarded as an anti-apoptotic transcription factor and its activation is believed to be fundamental in neuroprotection and repair following brain injury and upregulation of p75 expression. Thus, it is reasonable to believe that activation of this transcription factor is one of the mechanisms through which cells are able to control between life and death decisions.

Finally, p75^{NTR} receptor has been found to directly interact with RhoA, a member of the Ras-GTP binding proteins and a regulator of actin assembly in many cell types, suggesting a role for this receptor in the neurite outgrowth and reorganization of the actin cytoscheleton, a function that is negatively modulated by neurotrophin binding [168].

5.2 Aeta toxicity and oxidative stress

Oxidative stress and mitochondrial dysfunction are common players in the apoptotic cascades leading to neuronal death in AD and are induced by all the known triggers, including growth factor deprivation and A β toxicity both *in vitro* and *in vivo* [37,169]. A β can induce apoptosis directly by increasing generation of ROS and mitochondrial dysfunction. ROS accumulation determines oxidation of proteins, peroxidation of membrane lipids and DNA damage. Oxidative stress causes then a dysfunction of Na⁺/K⁺- and Ca²⁺-ATPases, alteration of glucose and glutamate transport, excessive Ca²⁺ influx, ATP depletion, membrane depolarization and translocation of cytochrome *C* to the cytoplasm [rev. in ref [42]].

It should be briefly mentioned that further contribution to oxidative stress is also given by Fe²⁺, Cu²⁺ and Zn²⁺ ions [42], whose levels are increased in the senile plaques of AD patients. Several *in vitro* and *in vivo* studies have demonstrated that these ions are implicated in A β deposition [170].

Besides the evidence demonstrating the involvement of A β in the impairment of synaptic activity, A β toxicity has been found to be a central element of the apoptotic neuronal death in AD, as extensively demonstrated both *in vivo* [37,48,62,63] and *in vitro* by using either primary cultured neurons or PC12 cells [59,66,171]. A β -induced apoptosis of cortical neurons is characterized by internucleosomal fragmentation after only a 6 hr-exposure to A β and is greatly increased by expression of p75 [172-174], thus linking A β toxicity to the aberrant expression of p75 and TrkA in AD [95,96].

Aβ-induced apoptosis is mediated by activation of the Stress-activated protein kinases (SAPK) p38 Mitogen Activated Protein Kinase (p38MAPK) and JNK, followed by activation of both NF-kB and p53 [171,174,175]. Active forms of JNK and p38 kinase have been found in hippocampal and cortical neurons of AD brains, suggesting their potential implication in hyperphosphorylation of tau and NFT formation [176].

Recently, p75-mediated cell death in A β neurotoxicity has also been shown to occur through interaction of p75 with a newly identified P75-Like Apoptosis-Inducing Death Domain (PLAIDD) [177]. Moreover, a new protein has been identified, A β -Binding Alcohol Dehydrogenase (ABAD), that is highly expressed in the brain and has been clearly shown to link A β -toxicity to the mitochondrial oxidative stress [178].

In hippocampal neurons, sympathetic neurons and PC12 cells, $A\beta$ -toxicity has been found to be mediated by activation caspases-9 and -3, as well by caspase-2 [171,179].

In AD, caspases -9, -3, -6 and -8 colocalize with NFT, suggesting a direct role of caspases in cleavage of tau and formation of NFT [61,67,180]. Caspase-8 in the brain of AD patients has been also shown to be induced through a mechanism mediated by the TNF-Related Apoptosis-Inducing Ligand (TRAIL) in response to A β toxicity. Evidence from *in vitro* studies also indicates that TRAIL might provide a substantial contribution to the progression of the neurodegeneration during chronic inflammatory responses [181]. Other caspases that are found overexpressed in postmortem brains from AD patients include caspases-2, -5 and -7 [62].

However, besides the strong presence of the many activated caspases, also increased NF-kB activity has been observed in neurons and glia of brain sections from AD patients, as well in cultured primary neurons exposed to A β [182]. Thus, it is quite clear that also *in vivo* apoptosis involves activation of several apoptotic cascades, as well as signaling pathways that lead to trascription of neuro-

protective genes in an attempt to rescue neurons from cell death.

5.3 Excitotoxicity: role of Ca^{2+} and calpains in cell death and NFT formation

As previously mentioned, excitotoxicity plays a role in many acute and chronic degenerative diseases and exacerbates oxidative stress through a mechanism leading to disruption of Ca^{2+} homeostasis and increase of ROS [183].

Ca²⁺ is a critical regulator of synaptic plasticity because of its role in LTP and LTD, postranslational modification of proteins involved in neurite elongation and synaptic activity (actin-binding proteins and ion channels), activation of kinases, such as Protein Kinase A (PKA), Calcium-Calmodulin Kinase II (CaMKII), and ras-Mitogen Activated Kinase (MAPK), as well as the activity of transcription factors, like Elk-1, CREB and Activator Protein-1 (AP-1) [184]. Moreover, cytosolic free Ca²⁺ mobilized from the intracellular stores of the ER has been shown to be also a second messenger for NGF-TrkA signaling linked to the activation of PLC- γ [185]. However, persistent alterations of Ca2+ homeostasis following excitotoxicity and oxidative stress can contribute to neuronal apoptosis in AD [142,186]. One target for increased intracellular Ca²⁺ levels are calpains [142], cystein proteases involved in the regulation of cytoskeletal remodeling. Calpains were found to participate to apoptosis through cleavage of Bax [186], followed by release of cytochrome C and activation of caspase-3, as well as by direct activation of caspases-7 and -12.

Calpain activation has been detected in the brains of AD patients in proximity of NFT [187] and suggested to play a role in the regulation of tau phosphorylation and NFT formation by a mechanism involving the cleavage of p35 to p25 [188] and activation of cdk5 [188,189]. Cdk5, a member of the cyclin-dependent kinase family not linked to cell cycle, is highly expressed in neurons and, together with its regulatory protein p35, is required for neurite outgrowth. In AD, accumulation of p25 correlated with enhanced cdk5 kinase activity, while expression of p25/ cdk5 in cortical neurons was found to cause increased tau phosphorylation, disruption of cytoskeleton and apoptosis [189]. Inhibition of cdk5 or calpain activity reduced cell death in A\beta-treated cortical neurons, suggesting that the p25/cdk5 complex mediates also Aβ-neurotoxicity [188].

On the other hand, there is increasing evidence showing that NFT formation and apoptosis might also involve an impairment of the ubiquitin-proteasome (UB-proteasome) system, whose activity is required for degradation of short-lived and damaged proteins, such as misfolded or modified proteins in AD. *In vitro* studies have shown that

failure of the proteasome system is an early event in several apoptotic conditions including NGF deprivation of sympathetic neurons [190] and exposure of cerebellar granule cells to reduced K+ levels [191]. The latter has been found to be associated with caspase-3 activation and accumulation of ubiquitinated proteins in the cytosol [191].

There is evidence for impaiment of the UB-proteasome system also in AD. Among the others, a mutant form of ubiquitin (Ub⁺¹) has been found in the brain of AD patients, that would result in dominant inhibition of the UB-proteasome system [192] and might play a critical role in accumulation of misfolded tau proteins forming NFT.

5.4 Vascular factors and brain metabolism

A contribution to increased A β production and deposition was found in individuals carrying the ϵ 4 allele of the Apolipoprotein E (APOE) gene. APOE is a plasma lipoprotein that functions as the major cholesterol carrier to the brain, thus linking hypercholesterolaemia and alteration of brain cholesterol homeostasis to the risk of developing AD. This correlation has been suggested also by several epidemiological studies, as well by the evidence that statins, which reduce cholesterol levels, also decrease the risk of AD [rev. in refs [193,194]].

Atherosclerosis is also a risk factor for AD, as reduced cerebral blood flow can cause reduced glucose utilization in AD patients [195]. Other vascular factors in AD include diabetes and alteration of serum levels of insulin and Insulin Growth Factor-1 (IGF-1), two important regulators of brain energy balance. In fact, IGF-1 has been found to be implicated in modulation of synaptic plasticity [196,197], regulation of A β levels [198] and neuroprotection against A β toxicity [199] by a mechanism involving Akt and GSK-3 β [200,201].

Insulin/IGF-1 serum levels and signaling are impaired in AD patients suggesting a link also between AD pathology and defective signaling by these two important regulators of the brain energy balance. Thus, it is conceivable that a decline in metabolic and hormonal function in aging, together with accumulation of oxidative stress, can influence neuronal vulnerability to other concomitant insults and result in neuronal degeneration through activation of apoptotic cascades.

5.5 IGF-1, Wnt signaling and GSK-3 β

The relevance of IGF-1 in neuroprotection developes around the activity GSK-3 β , which is a central element of the IGF-1/Wnt signaling, and also a target of the PI3K/Akt pathway.

AD is characterized by increased levels of GSK-3 β which, in its active (dephosphorylated) state, is a key regulator of

NFT formation [126]. GSK-3 β is phosphorylated (and inactivated) through a PI3K/Akt signaling pathway induced by IGF-1, NGF and any growth factor whose activity is mediated by receptors with intrinsic tyrosine kinase activity [201], including also Fibroblast Growth Factor-2 (FGF-2). Down-regulation of PI3K/Akt results in active GSK-3 β and renders neurons more vulnerable to A β /JNK/p53 cascade. In fact, GSK-3 β is activated, along with caspases, during NGF deprivation-induced cell death of PC12 cells [129], A β - toxicity and oxidative stress [152] by a mechanism mediated by reduced PI3K/Akt signaling and JNK/p53 activation, thus providing a link between NGF deprivation and the two pathological components of AD.

Among other evidence indicating a role in neuroprotection, IGF-1, NGF and FGF-2 have been shown to protect hippocampal neurons against $A\beta$ toxicity [199,201-203].

GSK-3 β is also inactivated by the mitogenic IGF-1/Wnt pathway. GSK-3 β inactivation results in: *i*) decreased tau phosphorylation and *ii*) accumulation of β -catenin followed by translocation to the nucleus and regulation of cyclin D1 expression and c-myc [204]. The Wnt signaling can be inhibited by p53 leading to active GSK-3 β which determines phosphorylation of β -catenin for degradation by the ubiquitin-proteosome system, increased A β production and increased tau phosphorylation. Thus, GSK-3 β provides also a link between decreased IGF-1-mediated trophic support and A β toxicity/tau phosphorylation/cell cycle events [152].

5.6 Apoptosis and cell cycle events

An intriguing issue about the apoptotic processes underlying neurodegenerative diseases is raised by the question that neuronal death is accompanied by cell cycle events, suggesting that neurons attempt to divide before dying. Moreover, there is increasing evidence that neuronal apoptosis can arise from a failure of degenerating neurons to re-entry the cell cycle. Mature neurons exit the cell cycle and remain quiescent in the G0 phase, however, active cell cycle proteins are present in neurons of AD brain, including cyclins, cell cycle kinases, as well as their activators and inhibitors [134-136,205,206]. Cell cycle proteins and complexes found in AD brains include cyclinD, cdk4 and Ki67, as well the presence of cyclinE/cdk2 which controls G1/S phase transition, indicating an alteration of cell cycle events in AD neurons.

An increase in cyclin D1 expression has been demonstrated also in A β -induced apoptosis of differentiated neurons [206]. In addition, several *in vitro* studies have shown that apoptosis induced by NGF deprivation occurs together with the expression of cell cycle proteins cdk4 and cdk6 and their cognate cyclins, and that cdk inhibitors p16^{Ink4}, p21^{Cip1} and p27^{Kip1}, as well the DN-cdk4/6, promoted survival of differentiated PC12 cells and sympathetic neurons [207,208]. Besides supporting survival, p16^{Ink4} and p21^{Cip1} have been shown to reconstitute NGF-dependent neurite outgrowth in PC12 cells [209].

Similarly, elevated expression of the cdk inhibitor p16^{Ink4} and other members of the INK4-family (p15^{Ink4b}, p18^{Ink4c} and p19^{Ink4d}), which bind complexes of cdk4/6-cyclinD, were found also in AD, mainly in neurons with NFT and neuritic plaques [135]. Several other studies have demonstrated increased expression of cell cycle proteins, including Proliferating Cell Nuclear Antigen (PCNA), cyclins B, C and D in the brain of AD patients, both in neurons and astrocytes. Moreover, cell cycle events appear to occur at early stages of the disease (in MCI) and precede neuronal cell death [136,210,211].

Many factors may induce neurons to attempt a cell cycle re-entry. One is the overexpression of growth factors with mitogenic/differentiation activity, such as NGF, IGF-1/2 and Epidermal Growth factor (EGF), as well Tumor Growth Factor- α 1 (TGF α 1) and FGF-2 secreted by reactive astrocytes and microglia in response to brain damage. Most of these growth factors signal through the ras-MAPK pathway [212] that is involved in the regulation of G0/G1 transition. At an early stage of AD, elevated expression of p21ras co-localizes with p16^{Ink4} in pyramidal neurons [213]. ERK activation is also linked to c-myc phosphorylation in AD.

Cell cycle re-entry may occur also through mechanisms mediated by GSK-3 β following phosphorylation (and inactivation) by either a PI3K/Akt signaling or the mitogenic Wnt pathway that is a central element of IGF-1 signaling. For instance, the inactivation of GSK-3 β results in the upregulation of cyclin D1 expression in carcinoma cells [204].

Furthermore, cell cycle re-entry might also be triggered by oxidative stress through activation of the stress-activated protein kinases JNK-SAPK and p38-SAPK2, which control stress responses in AD [176], suggesting a possible link between the occurrence of cell cycle reentry and increased oxidative stress and neuronal vulnerability in aging. Accordingly, neurons in AD brains are characterized by extensive DNA damage and increased p53 expression. However, in PC12 cells and cortical neurons, exposure to A β induces upregulation of cdk4/6, phosphorylation of the retinoblastoma protein pRb/107, release of Rb from the transcription factor complex E2F-DP and activation of E2F-mediated trascription of genes required for S phase transition, but these events occur in a p53-independent manner [214]. Cell cycle components have been proved to be expressed also in FAD mutants of APP, where cell cycle entry can be mediated by the APP-binding protein 1 (APP-BP1) [215] or p21-Activated Kinase3 (PAK3) [216]. Recently, a reduced PAK pathway was associated to cognitive deficits in AD [217].

5.7 Inflammatory processes

Finally, another relevant factor in the pathogenesis of AD is given by neuroinflammatory processes, a common response to brain damage mediated by the proinflammatory cytokines Interleukin(IL)-1 α (IL-1 α), IL-1 β , TNF α and TGF- β 1, chemokines and the prostaglandin E2 (PGE2) released by reactive astrocytes [218].

Many of these inflammatory mediators have been shown to mediate protection by inducing the synthesis of growth factor. For instance, NGF while under normal conditions is localized in neurons, during inflammation is synthesized also by microglia and reactive astrocytes, where it is upregulated by cytokines (IL-1 α , IL-1 β and IL-6) and other inflammatory mediators, such as Interferon- γ (IFN- γ) and Lipopolysaccharide (LPS) [reviewed in ref. [107]]. This is a common response to CNS injury and it is linked to the role of NGF in neuroprotection and repair. NGFmediated activation of both TrkA and p75, the latter being increased following brain damage, may promote neuronal survival through activation of NF-kB [182], which can also be directly induced by most of the cytokines.

Microglial activation have also a phagocytosis function for removal of apoptotic cells, thus limiting the spread of toxins to the adjacent tissues. Neverthless, it is believed that the inflammatory processes contribute to exacerbation of the AD pathology and rapid progression of the disease. The activation of microglia causes chronic inflammation by inducing an autotoxic loop due to secretion of proinflammatory cytokines (TNF α and IL-1 β), acute-phase proteins and overproduction of free radicals (nitric oxide and superoxide) [218,219]. An amplification of the inflammatory processes is also due to activation of the complement system that has been shown to be also directly induced by A β [220].

The role of inflammatory processes in AD progression may explain the protection by non- steroid anti-inflammatory drugs observed in epidemiological studies [221], due to both cyclooxygenase-1 and -2 (COX1-2) inhibition of PGE2 synthesis and modulation of γ -secretase [222]. A different mechanism of protection may instead be mediated by the naturally occuring anti-inflammatory cytokines, such as IL-10 which has been shown to prevent glutamate toxicity through inhibition of caspase-3 [223].

6. Toward a blueprint of neuronal apoptosis

It is clear from the experimental evidence summarized in the two previous paragraphs that neuronal apoptosis in AD is quite a complex process, thus posing many problems to the current efforts of acquiring a comprehensive understanding of the phenomenon and ordering of the sequential events.

Recently, attempts to obtain this information have involved different approaches, including microarray analysis of expression profiles [64,65]. One of these studies revealed that $A\beta$ -induced apoptosis of cultured cortical neurons involves the differential expression of 956 genes, and 70 of them were common to those whose expression was altered during apoptosis of cerebellar granule cells by serum and potassium deprivation as well [64]. These genes included neurotransmitter receptors, neuropeptides, ion channels, growth factors, transcription factors, enzymes, structural proteins, and so on. These data strongly suggest that neuronal apoptosis displays several regulatory mechanisms that are common to other apoptotic processes triggered by various insults and in a cell type-independent manner.

Neverthless, gene expression analysis can only provide a partial picture of the whole process. The notion is well accepted that apoptosis is an active process requiring *de novo* protein synthesis [224]. However, many events are based on translational or posttranslational modifications, such as allosteric modifications, phosphorylations, compartimentalization, as well as cellular intermediates that belong to different classes of molecules, such as Ca²⁺, ceramide, etc.

As mentioned in the first paragraph of this review, a modular systems biology approach has been shown to be a useful strategy for organizing information and providing molecular models of complex bioprocesses. A similar instrumental benefit might be obtained for apoptosis in chronic neurodegenerative diseases.

Given the complexity of the phenomenon in AD, a large body of data needs to be organized and structured first in a low-level model (a blueprint) and further in a more detailed molecular model. A recent mathematical model of CD95-induced apoptosis [28] is used in this review as backbone to start to organize the data gathered on neuronal apoptosis. The proposed basic blueprint for neuronal apoptosis shown in Figure 2 considers a trigger submodule (the DISC-system in the Bentele *et al.* paper), an actuator (the caspase submodule), a feed-forward module (mitochondrial dysfunction), a brake (IAP/Survivin/Cki) and the many degradative responses of apoptosis (degradation of proteins, poly(ADP-ribose)polymerase (PARP), DNA and so on). As shown in Figure 2, the structure of the basic blueprint indicates quite clearly that the activation of caspases is due to one specific event, although it depends on various triggers. The mitochondria submodule, that has a feed-forward effect on caspases, may be further stimulated by an autocatalytic process, while a negative feed-back on caspases depends on the IAP/Survivin/Cki module. The fully fledged caspases induce apoptotic death by degrading the various cellular components.

The structure of the blueprint makes it possible to analyze the condition in which the caspases activation is not fully expressed and investigate the role that both the IAP/Survivin/Cki module and the anti-apoptotic molecules acting on mitochondria may have in protecting neurons from death.

Let us now go through the various submodules of the blueprint and analyze how the findings summarized in paragraphs 4 and 5 fit with the picture.

6.1. The triggers

It is quite clear from Figure 3 that the events able to trigger neuronal programmed cell death are numerous and quite diverse, and possibly in cross-talk.

Because of its pivotal role for maintenance of the cholinergic phenotype, one of the key triggers might be the alteration of NGF availability and/or transport, and failure of the TrkA-mediated signaling (via ras-ERK1/2 and PI3K pathways), as described in paragraphs 4.1 and 5.1. On the other hand, increased pro-NGF levels in AD [90] might also play an important role in triggering neuronal apoptosis through p75 receptor [91], whose upregulation is common to a variety of neuronal injuries. Indeed, although the precise p75 signaling is not completely understood, p75 can induce apoptosis through a neurotrophindependent activation of the sphingomyelinase/ceramide and JNK/p53 pathways followed by induction of Bax and Fas L, thus involving the activation of the mitochondrial intrinsic cascade and, perhaps, also an extrinsic apoptotic pathway, given that Fas and FasL have been found in senile plaques in AD and suggested to participate Aβinduced neuronal death [63].

A β is a another key trigger of neuronal apoptosis in AD and its neurotoxicity is enhanced by expression of the p75 receptor. Besides the involvement of the ceramide/JNK pathway, A β toxicity leads to accumulation of ROS, thus further increasing a condition of oxidative stress that is peculiar to the brain in aging and represents the third main element strongly implicated in neuronal apoptosis in AD, through activation of the mitochondrial intrinsic cascade. Moreover, ROS-mediated activation of caspase-3, can further stimulate sphingomyelinase and ceramide



Figure 2

The basic blueprint of neuronal apoptosis showing the different modules and how they are interconnected by positive and negative feed-backs.

production, thus generating a feed-forward loop. An extrinsic pathway is also activated by $A\beta$ through the induction of TRAIL and activation of caspase-8.

As mentioned, an important role in neuroprotection is played by other growth factors, such as FGF-2 and IGF-1, meaning that availability of these growth factors can influence the balance of pro- and anti-apoptotic mediators (Bcl-2/Bax, caspases, NF-kB, etc.) and push toward the rescue of cells rather than their death. The relevance of trophic support, mainly NGF and IGF-1, and other growth factor signaling through trk receptors and activation of PI3K/Akt and PKC, is also linked to the key role played by GSK-3 β in the regulation of both A β accumulation and NFT formation, as well to cell cycle events.

Another important element is the alteration of Ca^{2+} homeostasis that can be caused by both oxidative stress and excitotoxicity, a common pathway in many neuropathological conditions [35]. Besides the role in cell death, Ca^{2+} gives a further contribution to the formation of NFT by activating calpains and p25-cdk5 complexes which increase tau phosphorylation [188,189].

Moreover, among the other mechanisms triggering cell death and described in paragraph 4, we must acknowledge also the genetic mutations (APP, PS1/2, APOE) that cause an alteration of APP processing and accumulation of neurotoxic A β , although the role of the genetic factors is limited to the familial forms of AD.

6.2. The caspases

This module is characterized by one initiator caspase (caspase-8 or caspase-9) activated by the trigger events and several executioner caspases (caspases-3, -6, -7). Caspases are synthesized as inactive pro-caspases that are cleaved at the internal proteolytic site to yield the active caspase. The initiator caspases, once activated, will activate the downstream effector caspases, which in turn will cleave their substrates, such as PARP.

In the extrinsic pathway this module is initiated by the association of the DD receptor to its ligand and recruitment of an adaptor molecule and procaspase-8 to form the DISC (Death Inducing Signaling Complex). Procaspase-8 is then cleaved to active caspase-8 which in turn activates the effector caspase-3 [141]. The extrinsic pathway mediated by DD receptors in AD is believed to be prevalently linked to Fas-FasL, TRAIL and cytokines activity during inflammation.

The intrinsic or "mitochondrial" pathway, instead, includes the activation of the initiator caspase-9 mediated by the heptameric apoptosome complex. Formation of apoptosome is initiated by the interaction of the cytoplasmic APAF-1 with cytochrome *C* derived by the dysfunction of mitochondria. Once recruited to the apoptosome, procaspase-9 acquires autocatalytic activity and then activates also the executioner caspases-3 and -7. Caspase-3 activates, by proteolytic cleavage, both the executioner caspase-6 and caspase-2 which, in turn, activates a feed-

Trigger events

- A β neuritic plaques and NFTs formation
- Decreased availability of NGF/alteration of NGF processing
- Alteration of trkA/p75 receptors expression
- Impairment of M1/M3 mAchR signaling
- Oxidative stress
- Excitotoxicity
- Alteration of Ca²⁺ homeostasis
- Production of pro-inflammatory cytokines (TNF- α and IL-1 β)
- Decreased trophic support (IGF-1/Wnt signaling/GSK3β)
- Impairment of the proteasome system
- Vascular factors (hypercholesterolemia and decreased glucose metabolism)
- Genetic mutations (APP, PS1/2, APOE ε4 allele)

Figure 3

A list of the many genetic and environmental events that contribute to the activation of apoptotic cascades in AD neurodegeneration.

forward loop by stimulating mitochondria to release more cytochrome *C* and activate more caspase-9.

At this point we moved to collect the available information in a molecular model (Figure 4) structured in modules according to Figure 2, although we do not claim it to be exhaustive and definitive, but a useful tool for a systems biology approach. When considering caspases, Figure 4 shows that the overall dynamics of this submodule sees distinct triggers that independently activate pathways of proteolysis that converge on the three executioner caspases, whose activity is strongly controlled by a feed-forward loop acting on mitochondria. This process is in part sustained by autocatalytic events, such as the activation of caspase-2 by caspase-3 involved in the feed-forward loop, as well the activation of caspase-6 by caspase-3 that allows an amplification of the final proteolytic events controlling protein degradation and DNA fragmentation.

In addition, we can observe that in AD the majority of the cell death signals converge on the mitochondrial intrinsic

pathway and that the two main caspase cascades are not independent on each other. A cross-talk between the two pathways is mediated by caspase-8, cleavage of Bid and traslocation of tBid to mitochondria where, directly or indirectly, it causes the release of cytochrome *C*. Also this cross-talk between the DD receptor signal and mitochondria leads to an amplification of caspases activation.

6.3. The speeder: the mitochondrial submodule

The speeder submodule involves processes that integrate the initial apoptotic signaling which otherwise could proceed to either cell death or recovery from stress events. The main actors are mitochondria which play a crucial role in caspase-dependent and independent apoptotic signaling through the interaction with both pro-apoptotic (Bax, Bak, Bad, Bid, Bik and Bim) and anti-apoptotic proteins (Bcl-2, Bcl-X_L and Bcl-w) [149]. Following trigger events, their activation and association in different homo- and hetero-dimers function as a "sensor" of the level of cellular stress and damage. Proapoptotic proteins induce cell death by regulating the formation of transition pore



Figure 4

A molecular model of neuronal apoptosis organized in submodules. The central part of the model (green panel) represents the mitochondria submodule and all the molecules that regulate its function as "sensor" of the cellular stress responses (yellow panel). Dark blue and green arrows indicate survival and apoptotic pathways, respectively; orange arrows indicate inhibitions. Red arrows indicate the positive feed-back, while in purple and light blue are indicated the cross-talk between extrinsic/intrinsic apoptotic cascades and the survival regulatory control by IAPs, respectively. More information is the text.

(MTP) in the outer mitochondrial membrane. Mitochondrial membrane permeability (MMP) determines the release of cytochrome *C*, Smac/DIABLO, AIF and endonuclease G (Endo G). Traslocation of cytochrome *C* to the cytoplasm is crucial to the activation of the initiator caspase-9 and the executioner caspases-3 and -7, followed by cleavage of PARP and activation of Caspase-activated DNase (CAD) and DNA fragmentation. AIF is, instead, the final effector of a caspase-independent pathway that causes extensive DNA fragmentation, while Smac/DIA-BLO participate in the final steps of both the extrinsic and intrinsic pathway by inhibiting the IAP proteins. ROSmediated activation of the intrinsic apoptotic pathway involves the release of several mitochondrial proteins, such as cytochrome *C*, AIF and Smac/DIABLO. MMP is determined by Bax and Bak homodimers which are formed upon allosteric conformational changes and dimerization in response to trigger signals. Bax and Bak activation is mediated by Bad, Bik, Bid, and Bim and is due to either the preferential interaction with Bcl-2 and Bcl-X_L, or their release from these anti-apoptotic proteins, thus shifting the balance of the different heterodimers toward the formation of the proapoptotic Bax and Bak homodimers. In sympathetic neurons, following growth factor deprivation and failure of PI3K/Akt signaling, Bad is activated by dephosphorylation, dissociation from the molecular chaperone 14-3-3 and association with Bcl-X_L. In viable cells, Bcl-X_L binds Bax and Bak, thus keeping them in an inactive form. Bid and Bim can be also activated by a direct cleavage by caspase-8, as well as by the lysosomal protease cathepsin, in response to ceramide, ROS and JNK.

The model presented in Figure 4 indicates that mitochondria play a significant role in the amplification of the apoptotic processes initiated by growth factor deprivation, DD receptors activation (p75 and Fas), increased levels of intracellular Ca²⁺ linked to excitotoxicity and ROS accumulation in response to the many brain insults.

6.4. The brake: the IAP/Survivin/Cki submodule

In a scenario that mainly sees the propagation of various apoptotic stimuli that may compromise mitochondrial function and converge into the activation of a series of caspases and other effectors of cell death, a brake submodule is given by the IAP proteins (Figure 4). The IAP family of proteins include XIAP (X-linked IAP), c-IAP-1/2 and Survivin which are potent regulators of caspase activity. They function by inhibiting both the activation of pro-caspases and the enzymatic activity of active caspases, such as caspases-3, -7 and -9.

The pro-survival activity of IAPs, however, can in turn be inhibited by Smac/DIABLO that, once released from mitochondria, can bind IAP and promote activation of caspase-3. This modulation represents one of the many examples of the fine regulatory control of apoptosis. An interesting connection between cell cycle and apoptosis is given by the antiapoptotic activity of the Cki p 27^{Kip1} and $p21^{Cip1}$. When these proteins are phosphorylated by Akt, they are translocated to the cytoplasm where they may have antiapoptotic activity [225-227]. More specifically, $p27^{Kip1}$ has been shown to be cleaved by caspases in the p-23 and p-15 N-terminal peptides that have a strong antiapoptotic activity [226].

Although, as shown above, there is enough information to sketch the lines of structure of the various submodules of the apoptotic process in general, investigations are needed to identify the protein networks actually present in neuronal cells in various neurodegenerative disorders in order to make faithful mathematical models able to yield accurate predictions.

The application of the roadmap of modular systems biology to investigate neuronal apoptosis is expected to improve pathways and network identification with high efficiency.

7. Neuroprotection and control of apoptosis

Current therapies for Alzheimer's disease as well as for other neurodegenerative disorders are solely symptomatic [228]. The efforts of the pharmaceutical industry are now focusing on several steps of the hypothesized pathogenic pathways: for instance, prevention of production of neurodegenerative factors (plaques and NFT), targeting mediators of deleterious signaling cascades and trophic support. How does this approach fit with the blueprint of neuronal apoptosis discussed so far?

The concept of neuroprotection implies the reversibility or at least the significant slow down of the apoptotic process. It is quite clear that apoptosis cannot be reversed when a fully fledged activation of executioner caspases has been achieved. It is therefore reasonable to suppose that the passage from cells under apoptotic stress to dying cells requires the full activity of the positive feed-back loop based on mitochondria. What is the role of caspase-2 in this process? Which are the manipulations more likely to damp down the apoptotic stress? What are the markers of mild to severe apoptotic stress? How do the cells of a population/tissue respond to a specific apoptotic trigger? How will the overall activity of the cell, such as metabolic remodeling due to general environmental stress response in chromatin remodeling and ensuing changes in gene expression [229] affect neuronal apoptosis?

The molecular map drawn in Figure 4 attempts to compile the essential molecular pathways that chatacterize neuronal apoptosis. As said before, it does not pretend to be exhaustive, but only opens the way to a systems biology approach of neuronal apoptosis. First of all it presents a trigger module that includes the many genetic and environmental factors listed in Figure 3. Each triggering insult can activate pathways that independently lead to specific modifications/alterations of neuronal homeostasis and eventually initiate a cell death program.

The process is complicated by the existence of an intense cross-talk between the various molecular intermediates that directly or indirectly influence each other, as well as many other parameters controlling cellular homeostasis. The result is the sequential intervention of a large number of metabolic and signaling cascades that may also act synergistically.

Another important element is the redundancy of signalings leading to a specific effect. For instance, NFT formation is induced through activation (dephosphorylation) of GSK-3 β due to downregulation of PI3K/Akt, as well as by activation of the p25-cdk5 complex following Ca²⁺⁻ dependent activation of calpains and cleavage of p35, and both pathways can become synergistically activated upon trophic factors deprivation and ROS accumulation, suggesting that cells use more than one route to respond to stimuli.

A further level of amplification is present in the caspase submodule. As shown in Figure 4, caspases are activated both by the many stimuli that trigger the mitochondrial



Figure 5

A drawing exemplifying the active "stand-by" status of cells exposed to the various stressful events involved in AD apoptosis. Neuronal death occurs when the cell(s) reaches a point of "No-return", where an overwhelming mitochondria pressure is no longer compensated by the many protective pathways that the cell(s) has adopted.

intrinsic cascade (Bax, citochrome C/Apaf-1, caspase-9 and caspases-3/-7), and by the extrinsic pathways (DD receptors, caspase-8, caspase-3). Besides these cascades (green arrows), the caspase submodule is based on three mechanisms: *i*) autocatalytic events leading to the activation of many proteolytic enzymes that, once activated, greatly accelerate the degradation processes; *ii*) a feed-forward loop based on mitochondria and involving caspases and Bcl-2 family members (red arrows), and *iii*) a crosstalk between the extrinsic and intrinsic cascades (purple arrows).

This evidence indicates that mitochondria, besides playing an essential role in providing energy for all the cellular activities, also represent the central element of the apoptotic machinery, where signals from the different cellular compartments converge and are integrated. Thus, they play a pivotal role in the amplification of the apoptotic processes initiated by DD receptors activation (p75 and Fas), down-regulation of growth factor signaling, mitochondrial influx of Ca²⁺ released by ER and ROS accumulation in response to many brain insults. This function allows mitochondria to be the main "*sensor*" of the stress thresholds reached by neurons after an insult and continuously adjust their response. This role of mitochondria as "*sensor*" of the cellular conditions might be particularly relevant in AD and other chronic neurodegenerations, given that neurons, more than other cells, rather than dying, attempt to activate many protective programs that can delay the death decision, thus generating a situation of "*stand-by*". This situation is exemplified in Figure 5 showing cells that accumulate under stress pressure following exposure to toxic stimuli and/or stressful events. The idea is that after the activation of a trigger event, a cell may activate in part the caspase pathway without reaching a fully activated state.

In this context, the activity of IAPs and Survivin, which can inhibit caspases, appears relevant, as well as their inhibition by Smac/DIABLO. As shown in Figure 4, IAPs can be induced by NF-kB and Akt, which can also inhibit the intrinsic apoptotic pathway through phosphorylation of both caspase-9 and its inactive form [150]. This modulation represents, indeed, another example of the fine regu-

latory control of apoptosis, and is consistent with the hypothesis that the final outcome (life or death) depends not only on which players are active in a given moment, as this decision also revolves around to what extent each pro-apoptotic and anti-apoptotic element is active, in terms of levels and persistency. Thus, it seems that the amplifications and brake mechanisms are meant to add a further level of complexity to the final steps controlling the life/death decisions that eventually will depend on the amount of mitochondrial pressure. As illustrated in Figure 5, cells will die when a "Point of No-Return" is reached. Beyond this checkpoint the anti-apoptotic mechanisms are no longer effective against a persistent condition of energy depletion following mitochondrial dysfunction. Thus, apoptosis would take place upon engulfment of an over-loaded homeostatic system.

Another element that has recently gained a lot of attention is the building up of evidences for cell cycle proteins and DNA tetraploidy in neurons in brain regions involved in AD degeneration, suggesting that neurons may enter aberrant cell cycle before undergoing apoptosis.

An appealing issue, as well as an interesting field for future directions, is the antiapoptotic activity of the Cki p27^{Kip1} and p21^{Cip1}. If, on the one hand, there is evidence indicating that cell cycle events are upstream of caspases activity [214], suggesting that they may function like a "*last-chance*" mechanism engaged by stressed cells to overcome and/or recover from endangering conditions, on the other hand, the role of Cki is interesting as potential inhibitors of apoptosis upon their translocation to cytoplasm followed by caspases-3/8-mediated cleavage [226] (Figure 4). The release of an active Cyclin/cdk complex would then explain the abortive onset of cell cycle.

However, the presence of cell cycle proteins has been detected also in the brain of rodents and humans under normal conditions, suggesting that processes linked to cell cycle events might have a physiological, rather than pathological role [230], and require further investigation. The presence of so many regulatory links yields complex and often counter-intuitive dynamics in response to changing conditions, therefore underlying the need for accurate modeling and extensive simulations to better understand the system and allow the development of a rational drug discovery process for neurodegenerative disorders.

In this context, there is a remarkable need to identify the check-points that represent the points of no-return for over-stressed cells. Given the existence of so many actors, it is clear the importance of deriving mathematical models that can allow us to identify these critical check-points, thereby allowing to focus on selected relevant pathways and speed up the discovery process of rate limiting steps useful for neuroprotection.

Conclusion

In conclusion, this review has started from an overview of the current knowledge of the molecular events underlying apoptosis in neurodegenerative diseases, with particular attention to AD. Following the established strategy of modular systems biology, the data collected have been first used to identify the blueprint of the relevant modules of neuronal apoptosis and their wirings. Then, a molecular model of neuronal apoptosis has been constructed and discussed as a framework in which to investigate how various anti-apoptotic pathways may have a role in neuroprotection. This approach may offer new insights to the development of new therapeutic strategies against AD.

Acknowledgements

This work was in part supported by Associazione Levi-Montalcini.

This article has been published as part of *BMC Neuroscience* Volume 7, Supplement 1, 2006: Problems and tools in the systems biology of the neuronal cell. The full contents of the supplement are available online at <u>http://</u>www.biomedcentral.com/bmcneurosci/7?issue=S1.

References

- Rossel S, van der Weijden CC, Kruckeberg A, Bakker BM, Westerhoff HV: Loss of fermentative capacity in baker's yeast can partly be explained by reduced glucose uptake capacity. *Mol Biol Rep* 2002, 29:255-257.
- Hartwell LH, Hopfield JJ, Leibler S, Murray AW: From molecular to modular cell biology. Nature 1999, 402:C47-C52.
- 3. Nurse P: Systems biology: understanding cells. Nature 2003, 424:883.
- Alberghina L, Rossi RL, Porro D, Vanoni M: A modular systems biology analysis of cell cycle entrance into S-phase. Systems Biology Definitions and Perpectives 2005, 13:325-347.
- 5. Kitano H: Systems biology: a brief overview. Science 2002, 295:1662-1664.
- 6. Nurse P, Thuriaux P: Controls over the timing of DNA replication during the cell cycle of fission yeast. *Exp Cell Res* 1977, 107:365-375.
- Alberghina L, Mariani L: Control of cell growth and division. In Biomathematics and Cell Kinetics Edited by: Valleron AJ. Macdonald PDM. Elsevier-North-Holland; 1978:89-102.
- Tyson CB, Lord PG, Wheals AE: Dependency of size of Saccharomyces cerevisiae cells on growth rate. J Bacteriol 1979, 138:92-98.
- 9. Rupes I: Checking cell size in yeast. Trends Genet 2002, 18:479-485.
- Chen KC, Calzone L, Csikasz-Nagy A, Cross FR, Novak B, Tyson JJ: Integrative analysis of cell cycle control in budding yeast. *Mol Biol Cell* 2004, 15:3841-3862.
- Alberghina L, Porro D: Quantitative flow cytometry: analysis of protein distributions in budding yeast. A mini-review. Yeast 1993, 9:815-823.
- 12. Alberghina L, Porro D, Cazzador L: Toward a blueprint of cell cycle. Oncogene 2001, 20:1128-1134.
- 13. Chang F, Herskowitz I: Identification of a gene necessary for cell cycle arrest by a negative growth factor of yeast: FARI is an inhibitor of a GI cyclin, CLN2. *Cell* 1990, 63:999-1011.
- 14. Tyers M, Tokiwa G, Futcher B: Comparison of the Saccharomyces cerevisiae GI cyclins: Cln3 may be an upstream activator of Cln1, Cln2 and other cyclins. *EMBO J* 1993, 12:1955-1968.

- 15. Jeoung DI, Oehlen LJ, Cross FR: Cln3-associated kinase activity in Saccharomyces cerevisiae is regulated by the mating factor pathway. Mol Cell Biol 1998, 18:433-441.
- 16. Alberghina L, Chiaradonna F, Vanoni M: Systems biology and the molecular circuits of cancer. ChemBioChem 2004, 5:1322-1333.
- 17. Alberghina L, Rossi RL, Querin L, Wanke V, Vanoni M: A cell sizer network involving Cln3 and Farl controls entrance into Sphase in the mitotic cycle of budding yeast. J Cell Biol 2004, . **167:**433-443.
- Rossi RL, Zinzalla VAM, Alberghina L: Subcellular localization of the cyclin dependent kinase inhibitor Sicl is modulated by the carbon source in budding yeast. Cell Cycle 2005, 4(12):
- 19. Murray AW: Recycling the cell cycle: cyclins revisited. Cell 2004, 116:221-234.
- 20 Nash P, Tang X, Orlicky S, Chen Q, Gertler FB, Mendenhall MD, Sicheri F, Pawson T, Tyers M: Multisite phosphorylation of a CDK inhibitor sets a threshold for the onset of DNA replication. Nature 2001, 414:541-521.
- 21. Barberis M, Pagano MA, DeGioia L, Marin O, Vanoni M, Pinna LA, Alberghina L: Čk2 regulates in vitro the activity of the yeast cyclin-dependent kinase inhibitor Sicl. Biochem Biophys Res Comm 2005, 336:1040-1048.
- 22. Barberis M, DeGioia L, Ruzzene M, Sarno S, Coccetti P, Fantucci P, Vanoni M, Alberghina L: The yeast cyclin-dependent kinase inhibitor Sicl and the mammalian p27Kipl are functional homologues with a structurally conserved inhibitory domain. Biochem J 2005, 387:639-647.
- 23. Peter M, Herskovitz I: Joining the complex: cyclin-dependent kinase inhibitory proteins and the cell cycle. Cell 1994, 79:181-184.
- 24. Cheng M, Olivier P, Diehl JA, Fero M, Roussel MF, Roberts JM, Sherr C]: The p21(Cip1) and p27(Kip1) CDK 'inhibitors' are essential activators of cyclin D-dependent kinases in murine fibroblast. EMBO / 1999, 18:1571-1583.
- 25. Jorgensen P, Rupes I, Sharom JR, Schneper L, Broach JR, Tyers M: A dynamic transcriptional network communicates growth potential to ribosome synthesis and critical cell size. Genes Dev 2004, 18:2491-2505.
- 26. Jorgensen P, Tyers M: How cells coordinate growth and division. Curr Biol 2004, 14:R1014-1027
- 27. Cipollina C, Alberghina L, Porro D, Vai M: SFP1 is involved in cell size modulation in respiro-fermentative growth conditions. Yeast 2005, 22:385-399.
- Bentele M, Lavrik I, Ulrich M, Stober S, Heermann DW, Kalthoff H, 28. Krammer PH, Eils R: Mathematical modeling reveals threshold mechanism in CD95-induced apoptosis. J Cell Biol 2004, 166:839-851
- 29. Janes KA, Albeck JG, Gaudet S, Sorger PK, Lauffenburger DA, Yaffe MB: A systems model of signaling identifies a molecular basis set for cytokine-induced apoptosis. Science 2005, **310:**1646-1653.
- 30. Naoki H, Sakumura Y, Ishii S: Local signaling with molecular diffusion as a decoder of Ca^{2+} signals in synaptic plasticity. Mol Systems Biol 2005. doi:10.1038/msb4100035
- Pocklington AJ, Cumiskey M, Armstrong JD, Grant SGN: The pro-31. teomes of neurotransmitter receptor complexes form modular networks with distributed functionality underlying plasticity and behaviour. Mol Systems Biol 2006. doi:10.1038/ msb4100041
- 32. Yankner BA: Mechanisms of neuronal degeneration in Alzheimer's disease. Neuron 1996, 16:921-932.
- 33. Wong PC, Rothstein JD, Price DL: The genetic and molecular mechanisms of motor neuron disease. Curr Opin Neurobiol 1998, 8:791-799.
- Selkoe DJ: Cell biology of protein misfolding: the examples of 34. Alzheimer's and Parkinson's diseases. Nat Cell Biol 2004, 6:1054-1061.
- 35. Lipton SA, Rosenberg PA: Excitatory amino acids as a final common pathway for neurologic disorders. N Engl | Med 1994, 330:6 3-622.
- 36. Scott SA, Mufson EJ, Weingartner JA, Skau KA, Crutcher KA: Nerve growth factor in Alzheimer's disease: increased levels throughout the brain coupled with declines in nucleus basalis. | Neurosci 1995, 15:6213-6221.

- 37. Yatin SM, Varadarajan S, Link CD, Butterfield DA: In vitro and in vivo oxidative stress associated with Alzheimer's amyloid β peptide (1–42). Neurobiol Aging 1999, 20:325-330. Dirnagl U, ladecola C, Moskowitz MA: Pathobiology of ischaemic
- 38. stroke: an integrated view. Trends Neurosci 1999, 22:391-397.
- Bradford HF: Glutamate, GABA and epilepsy. Prog Neurobiol 39. 1995. 47:477-511.
- 40. Ankarcrona M, Dypbukt JM, Bonfoco E, Zhivotovsky B, Orrenius S, Lipton SA, Nicotera P: Glutamate-induced neuronal death: a succession of necrosis or apoptosis depending on mitochondrial function. Neuron 1995, 15:961-973.
- Bonfoco E, Krainic D, Ankarcrona M, Nicotera P, Lipton SA: Apop-41. tosis and necrosis: Two distinct events induced, respectively, by mild and intense insults with N-methyl-D-aspartate or nitric oxide/superoxide in cortical cell cultures. Proc Natl Acad Sci USA 1995, 92:7162-7166.
- Mattson MP: Apoptosis in neurodegenerative disorders. Nat 42. Rev 2000, 1:120-129.
- 43. Yakovlev AG, Knoblach SM, Fan L, Fox GB, Goodnight R, Faden A: Activation of CPP32-like caspases contributes to neuronal apoptosis and neurological dysfunction after traumatic brain injury. | Neurosci 1997, 17:7415-7424.
- Velier JJ, Ellison JA, Kikly KK, Spera PA, Barone FC, Feuerstein GZ: Caspase-8 and caspase-3 are expressed by different populations of cortical neurons undergoing delayed cell death after focal stroke in the rat. J Neurosci 1999, 19:5932-5941.
- Cotman CW, Whittemore ER, Watt JA, Anderson AJ, Loo DT: Possible role of apoptosis in Alzheimer's disease. Ann N Y Acad Sci 1994, 747:36-49.
- Tatton NA, Maclean-Fraser A, Tatton WG, Perl DP, Olanow CW: A 46. fluorescent double-labeling method to detect and confirm apoptotic nuclei in Parkinson's disease. Ann Neurol 1998, 44:SI42-SI48.
- 47. Wu C-K, Thal L, Pizzo D, Hansen L, Masliah E, Geula C: Apoptotic signals within the basal forebrain cholinergic neurons in Alzheimer's disease. Exp Neurol 2005, 195:484-496
- Whitehouse PJ, Price DL, Struble RG, Clark AW, Coyle JT, Delon MR: Alzheimer's disease and senile dementia: loss of neurons in the basal forebrain. Science 1982, 215:1237-1239.
- 49. Masliah E, Mallory M, Alford M, DeTeresa R, Hansen LA, McKeel DW Jr, Morris JC: Altered expression of synaptic proteins occurs early during progression of Alzheimer's disease. Neurology 2001, 56:127-129.
- Selkoe DJ: Alzheimer's disease is a synaptic failure. Science 50. 2002, 298:789-791.
- Nestor PJ, Scheltens P, Hodges JR: Advances in the early detection of Alzheimer's disease. Nat Med 2004, 10(Suppl):S34-41.
- Lue LF, Kuo YM, Roher AE, Brachova L, Shen Y, Sue L, Beach T, Kurth 52. JH, Rydel RE, Rogers J: Soluble amyloid β peptide concentration as a predictor of synaptic change in Alzheimer's disease. Am | Pathol 1999, 155:853-862.
- 53. Freir DB, Holscher C, Herron CE: Blockade of long-term potentiation by β -amyloid peptides in the CAI region of the rat hippocampus in vivo. J Neurophysiol 2001, 85:708-713.
- Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, Wolfe MS, Rowan MJ, Selkoe DJ: Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo. Nature 2002, 416:535-539.
- Van der Zee CE, Lourenssen S, Stanisz J, Diamond J: NGF deprivation of adult rat brain results in cholinergic hypofunction and selective impairment in spatial learning. Eur J Neurosci 1995, 7:160-168
- Chen KS, Nishimura MC, Armanini MP, Crowley C, Spencer SD, Phillips HS: Disruption of a single allele of the nerve growth factor gene results in atrophy of basal cholinergc neurons and memory deficit. J Neurosci 1977, 17:7288-7296.
- 57. Ruberti F, Capsoni S, Comparini A, Di Daniel E, Franzot J, Gonfaloni S, Rossi G, Berardi N, Cattaneo A: Phenotypic knockout of Nerve Growth Factor in adult transgenic mice reveals severe deficit in basal forebrain cholinergic neurons, cell death in the spleen, and skeletal muscle dystrophy. J Neurosci 2000, 20:2589-2601.
- 58. Connor B, Young D, Yan Q, Faull RL, Synek B, Dragunow M: Brainderived neurotrophic factor is reduced in Alzheimer's disease. Brain Res Mol Brain Res 1997, 49:71-81.

- Loo DT, Copani A, Pike CJ, Whittemore ER, Walencewicz AJ, Cotman CW: Apoptosis is induced by β-amyloid in cultured central nervous system neurons. Proc Natl Acad Sci USA 1993, 90:7951-7955.
- Tortosa A, Lopez E, Ferrer I: Bcl-2 and Bax protein expression in Alzheimer's disease. Acta Neuropathol (Berl) 1998, 95:407-412.
- 61. Masliah E, Mallory M, Alford M, Tanaka S, Hansen LA: Caspase dependent DNA fragmentation might be associated with excitotoxicity in Alzheimer disease. J Neuropathol Exp Neurol 1998, 57:1041-1052.
- Pompl PN, Yemul S, Xiang Z, Ho L, Haroutunian V, Purohit D, Mohs R, Pasinetti GM: Caspase gene expression in the brain as a function of clinical progression of Alzheimer disease. Arch Neurol 2003, 60:369-376.
- 63. Su JH, Anderson AJ, Cribbs DH, Tu C, Tong L, Kesslack P, Cotman CW: Fas and Fas ligand are associated with neuritic degeneration in the AD brain and participate in beta-amyloid-induced neuronal death. *Neurobiol Dis* 2003, 12:182-193.
- Cavallaro S, D'Agata V, Alessi E, Coffa S, Alkon DL, Manickam P, Ciotti MT, Possenti R, Bonini P, Marlier L, Calissano P: Gene expression profiles of apoptotic neurons. *Genomics* 2004, 84:485-496.
- 65. Reddy PH, McWeeney S, Park BS, Manczak M, Gutala RV, Partovi D, Jung Y, Yau V, Searles R, Mori M, Quinn J: Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: upregulation of mitochondrial metabolism and apoptotic genes is a early cellular change in Alzheimer's disease. Hum Mol Genet 2004, 13:1225-1240.
- Pike CJ, Walencewicz AJ, Glabe CG, Cotman CW: Aggregationrelated toxicity of synthetic beta-amyloid protein in hippocampal cultures. Eur J Pharmacol 1991, 207:367-368.
- Rohn TT, Head E, Su JH, Anderson AJ, Bahr BA, Cotman CW, Cribbs DH: Correlation between caspase activation and neurofibrillary tangle formation in Alzheimer's disease. Am J Pathol 2001, 158:189-198.
- Yuen EC, Howe CL, Li Y, Holtzman DM, Mobley WC: Nerve growth factor and the neurotrophic hypothesis. Brain Dev 1996, 18:362-368.
- 69. Oppenheim RW: Cell death during development of the nervous system. Annu Rev Neurosci 1991, 14:453-501.
- 70. Levi-Montalcini R: Effects of mouse tumor transplantation on the nervous system. Ann N Y Acad Sci 1952, 55:330-44.
- Levi-Montalcini R: The nerve growth factor 35 years later. Science 1987, 237:1154-1162.
- Huang EJ, Reichardt LF: Neurotrophins: roles in neuronal development and function. Annu Rev Neurosci 2001, 24:677-736.
- Sofroniew MV, Howe CL, Mobley WC: Nerve Growth Factor signaling, neuroprotection, and neural repair. Annu Rev Neurosci 2001, 24:1217-1281.
- Mobley WC, Rutkowski JL, Tennekoon GI, Gemski J, Buchanan K, Johnston MV: Nerve growth factor increases choline acetyltransferase activity in developing basal forebrain neurons. Brain Res 1986, 387:53-62.
- Large TH, Bodary SC, Clegg DO, Weskamp G, Otten U, Reichardt LF: Nerve growth factor gene expression in the developing rat brain. Science 1986, 234:352-355.
- Li Y, Holtzman DM, Kromer LF, Kaplan DR, Chua-Couzens J, Clary DO, Knusel B, Mobley WC: Regulation of trkA and ChAT expression in developing rat basal forebrain: Evidence that both exogenous and endogenous NGF regulate differentiation of cholinergic neurons. J Neurosci 1995, 15:2888-2905.
 Goedert M, Fine A, Hunt SP, Ullrich A: Nerve growth factor
- 77. Goedert M, Fine A, Hunt SP, Ullrich A: Nerve growth factor mRNA in peripheral and central rat tissue and in the human central nervous system: lesion effect in the rat brain and levels in Alzheimer's disease. Brain Res 1986, 387:85-92.
- Whittemore SR, Ebendal T, Lärkfors L, Olson L, Seiger A, Stromberg I, Persson H: Developmental and regional expression of beta nerve growth factor messenger RNA and protein in the rat central nervous system. Proc Natl Acad Sci USA 1986, 83:817-821.
- 79. Fisher W, Wictorin K, Bjorklund A, Williams LR, Varon S, Gage FH: Amelioration of cholinergic neuron atrophy and spatial memory impairment in aged rats by nerve growth factor. *Nature* 1987, **329:**65-68.
- 80. Kromer LF: Nerve growth factor treatment after brain injury prevents neuronal death. Science 1987, 235:214-216.
- Koliatsos VE, Clatterbuck RE, Nauta HJW, Knusel B, Burton LE, Hefti FF, Mobley WC, Price DL: Human Nerve Growth Factor pre-

vents degeneration of basal forebrain cholinergic neurons in primates. Ann Neurol 1991, **30**:831-840.

- Tuszynski MH, Roberts J, Senut MC, U HS, Gage FH: Gene therapy in the adult primate brain: intraparenchymal grafts of cells genetically modified to produce Nerve Growth Factor prevent cholinergic neuronal degeneration. *Gene Ther* 1996, 3:305-314.
- Cooper JD, Lindholm D, Sofroniew MV: Reduced transport of I25I-NGF by cholinergic neurons and downregulated trkA expression in the medial septum of aged rats. *Neuroscience* 1994, 62:625-629.
- Sofroniew MV, Galletly NP, Isacson O, Svendsen CN: Survival of adult basal forebrain cholinergic neurons after loss of target neurons. Science 1990, 247:338-342.
- Capsoni S, Giannotta S, Cattaneo A: β-amyloid plaques in a model for sporadic Alzheimer's disease based on transgenic anti-Nerve Growth Factor antibodies. Mol Cell Neurosci 2002, 21:15-28.
- De Rosa R, Garcia AA, Braschi C, Capsoni S, Maffei L, Berardi N, Cattaneo A: Intranasal administration of Nerve Growth Factor (NGF) rescues recognition memory deficits in ADII anti-NGF transgenic mice. Proc Natl Acad Sci USA 2005, 102:3811-3816.
- Levine ES: Neurotrophins and hippocampal plasticity. In Neurobiology of theNeurotrophins Edited by: Mocchetti I. F.P. Graham Publishing Co; 2001:165-186.
- Holsinger RM, Schnarr J, Henry P, Castelo VT, Fahnestock M: Quantitation of BDNF mRNA in human parietal cortex by competitive reverse transcription-polymerase chain reaction: decreased levels in Alzheimer's disease. Brain Res Mol Brain Res 2000, 76:347-354.
- Peng S, Wuu J, Mufson EJ, Fahnestock M: Precursor form of brainderived neurotrophic factor and mature brain-derived neurotrophic factor are decreased in the pre-clinical stages of Alzheimer's disease. J Neurochem 2005, 93:1412-1421.
- Fahnestock M, Michalski B, Xu B, Coughlin MD: The precursor pronerve growth factor is the predominant form of nerve growth gactor in brain and is increased in Alzheimer's disease. Mol Cell Neurosci 2001, 18:210-220.
- Harrington AW, Leiner B, Blechschmitt C, Arevalo JC, Lee R, Mörl K, Meyer M, Hempstead BL, Yoon SO, Giehl KM: Secreted proNGF is a pathophysiological death-inducing ligand after adult CNS injury. Proc Natl Acad Sci USA 2004, 101:6226-6230.
- Reynolds ME, Brunello N, Mocchetti I, Wrathall JR: Localization of nerve growth factor receptor mRNA in contused rat spinal cord by in situ hybridization. Brain Res 1991, 559:149-153.
- Oh JD, Chartisathian K, Chase TN, Butcher LL: Overexpression of neurotrophin receptor p75 contributes to the excitotoxininduced cholinergic neuronal death in rat basal forebrain. Brain Res 2000, 853:174-185.
- Beattie MS, Harrington AW, Lee R, Kim JY, Boyce SL, Longo FM, Bresnahan JC, Hempstead BL, Yoon SO: ProNGF induces p75-mediated death of oligodendrocytes following spinal cord injury. *Neuron* 2002, 36:375-386.
- Mufson EJ, Kordower JH: Cortical neurons express nerve growth factor receptors in advanced age and Alzheimer's disease. Proc Natl Acad Sci USA 1992, 89:569-573.
- Counts SE, Nadeem M, Wuu J, Ginsberg SD, Saragovi HU, Mufson EJ: Reduction of cortical trkA but not p75NTR protein in earlystage Alzheimer's disease. Ann Neurol 2004, 56:520-531.
- Chao MV, Casaccia-Bonnefil P, Carter B, Chittka A, Kong H, Yoon SO: Neurotrophin receptors: mediators of life and death. Brain Res Rev 1998, 26:295-301.
- Lee R, Kermani P, Teng KK, Hempstead BL: Regulation of cell survival by secreted proneurotrophins. Science 2001, 294:1945-1948.
- 99. Chao MV, Bothwell M: Neurotrophins: to cleave or not to cleave. Neuron 2002, 33:9-12.
- 100. Teng HK, Teng KK, Lee R, Wright S, Tevar S, Almeida RD, Kermani P, Torkin R, Chen Z-Y, Lee FS, Kraemer RT, Nykjaer A, Hempstead BL: ProBDNF induces neuronal apoptosis via activation of a receptor complex of p75NTR and sortilin. J Neurosci 2005, 25:5455-5463.
- Nykjaer A, Lee R, Teng KK, Jansen P, Madsen P, Mielsen MS, Jacobsen C, Kliemannel M, Schwarz E, Willnow TE, Hempstead BL, Petersen

CM: Sortilin is essential for proNGF-induced neuronal cell death. *Nature* 2004, **427:**843-848.

- 102. Sarret P, Krzywkowski P, Segal L, Nielsen MS, Petersen CM, Maxella J, Stroh T, Beaudet A: Distribution of NTS3 receptor/sortilin mRNA and protein in the rat central nervous system. J Comp Neurol 2003, 461:483-505.
- 103. Ledesma MD, Da Silva JS, Crassaerts K, Delacourte A, De Strooper B, Dotti CG: Brain plasmin enhances APP alpha-cleavage and Abeta degradation and is reduced in Alzheimer's disease brains. EMBO Rep 2000, 1:530-535.
- 104. Woo NH, Teng HK, Siao C-J, Chiaruttini C, Pang PT, Milner TA, Hempstead BL, Lu B: Activation of p75^{NTR} by proBDNF facilitates hippocampal long-term depression. Nat Neurosci 2005, 8:1069-1077.
- 105. Eriksdotter Jonhagen ME, Nordberg A, Amberla K, Backman L, Ebendal T, Meyerson B, Olson L, Seiger, Shigeta M, Theodorsson E, Viitanen M, Winblad B, Wahlund LO: Intracerebroventricular infusion of nerve growth factor in three patients with Alzheimer's disease. Dementia Geriat Cogn Disord 1998, 9:246-257.
- 106. Tuszynski MH, Tahal L, Pay M, Salmon DP, SangU H, Bakay R, Patel P, Blesch A, Vahlsing HL, Ho G, Tong G, Potkin SG, Fallon J, Hansen L, Mufson EJ, Kordower JH, Gall C, Conner J: A phase I clinical trial of nerve growth factor gene therapy for Alzheimer disease. Nature Med 2005, 11:551-555.
- 107. Mocchetti I, Colangelo AM: Transcriptional regulation of NGF in the central nervous system. In Neurobiology of the Neurotrophins Edited by: Mocchetti I. F.P. Graham Publishing Co; 2001:457-497.
- 108. Froestl W, Gallaghere M, Jenkins H, Madrid A, Melcher T, Techman S, Mondatori CG, Pearlman R: SGS742: the first GABAB receptor antagonist in clinical trials. Biochem Pharmachol 2004, 68:1479-1487.
- 109. Roβner S, Ueberham U, Schliebs R, Perez-Polo JR, Bigl V: The regulation of amyloid precursor protein metabolism by cholinergic mechanisms and neurotrophin receptor signaling. Prog Neurobiol 1998, 56:541-569.
- 110. Perez RG, Zheng H, Van der Ploeg LH, Koo EH: The beta-amyloid precursor protein of Alzheimer's disease enhances neuron viability and modulates neuronal polarity. J Neurosci 1997, 17:9407-9414.
- 111. Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird TD, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad L, Viitanen M, Peskind E, Poorkaj P, Schellenberg G, Tanzi R, Wasco W, Lannfelt L, Selkoe D, Younkin S: Secreted amyloid β-protein similar to that in the senile plaques of Alzheimer's disease is increased *in vivo* by the presenilin I and 2 and APP mutations linked to familial Alzheimer's disease. Nature Med 1996, 2:864-870.
- 112. Chapman PF, White GL, Jones MW, Cooper-Blacketer D, Marshall VJ, Irizarry M, Younkin L, Good MA, Bliss TV, Hyman BT, Younkin SG, Hsiao KK: Impaired synaptic plasticity and learning in aged amyloid precursor protein transgenic mice. Nat Neurosci 1999, 2:271-276.
- 113. Tong L, Thornton PL, Balazs R, Cotman CW: β-amyloid-(1-42) impairs activity-dependent cAMP-response element-binding protein signaling in neurons at concentrations in which cell survival is not compromised. J Biol Chem 2001, 276:17301-17306.
- 114. McCauslin CS, Heath V, Colangelo AM, Mallei A, Mocchetti I, Johnson PF: C/EBP δ and CREB mediate inducible expression of Nerve Growth Factor in the central nervous system. J Biol Chem 2006, 281:17681-17688.
- 115. Tong L, Balazs R, Thornton PL, Cotman CW: β-amyloid peptide at sublethal concentrations downregulates Brain-Derived Neurotrophic Factor functions in cultured cortical neurons. J Neurosci 2004, 24:6799-6809.
- 116. Rylett RJ, Goddard S, Schmidt BM, Williams LR: Acetylcholine synthesis and release following continuous intracerebral administration of NGF in adult and aged Fisher-344 rats. J Neurosci 1993, 13:3956-3963.
- 117. Roβner S, Ueberham U, Schliebs R, Perez-Polo JR, Bigl V: p75 and trkA receptor signaling independently regulate APP mRNA expression, isoform composition and protein secretion in PC-12 cells. J Neurochem 1998, 71:757-766.
 118. Costantini C, Weindruch R, Della Valle G, Puglielli L: A TrkA-
- 118. Costantini C, Weindruch R, Della Valle G, Puglielli L: A TrkAtop75^{NTR} switch activates amyloid β-peptide generation during aging. *Biochem J* 2005, 391:59-67.

- 119. Nitsch RM, Growdon JH: Role of neurotransmission in the regulation of amyloid beta-protein precursor processing. Biochem Pharmacol 1994, 47:1275-1284.
- 120. Roβner S, Ueberham U, Yu J, Kirazov L, Schliebs R, Perez-Polo JR, Bigl V: In vivo regulation of amyloid precursor protein secretion in rat neocortex by cholinergic activity. Eur J Neurosci 1997, 9:2125-2134.
- 121. Jope RS, Song L, Powers RE: Cholinergic activation of phosphoinositide signaling is impaired in Alzheimer's disease brain. Neurobiol Aging 1997, 18:111-120.
- 122. Kelly JF, Furukawa K, Barger SW, Rengen MR, Mark RJ, Blanc EM, Roth GS, Mattson MP: **Amyloid**β peptide disrupts carbacholinduced muscarinic cholinergic signal transduction in cortical neurons. Proc Natl Acad Sci USA 1996, **93**:6753-6758.
- 123. Pedersen WA, Kloczewiak MA, Blusztajn JK: Amyloid β-protein reduces acetylcholine synthesis in a cell line derived from cholinergic neurons of the basal forebrain. Proc Natl Acad Sci USA 1996, 93:8068-8071.
- 124. Chan SL, Griffin WS, Mattson MP: Evidence for caspase-mediated cleavage of AMPA receptor subunits in neuronal apoptosis and Alzheimer's disease. J Neurosci Res 1999, 57:315-323.
- 125. Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, Nirn AC, Salter MW, Lombroso PJ, Gouras GK, Greengard P: Regulation of NMDA receptor trafficking by amyloid-β. Nature Neurosci 2005, 8:1051-1058.
- 126. Wagner U, Brownless J, Irving NG, Lucas FR, Salinas PC, Miller CC: Overexpression of the mouse dishevelled-I protein inhibits GSK-3beta-mediated phosphorylation of tau in transfected mammalian cells. FEBS Lett 1997, 411:369-372.
- 127. Barbato C, Canu N, Zambrano N, Serafino A, Minopoli G, Ciotti MT, Amadoro G, Russo T, Calissano P: Interaction of Tau with Fe65 links tau to APP. Neurobiol Dis 2005, 18:399-408.
- 128. Pei JJ, Braak E, Braak H, Grundke-Iqbal I, Iqbal K, Winblad B, Cowburn RF: Distribution of active glycogen synthase kinase 3beta (GSK-3beta) in brains staged for Alzheimer disease neurofibrillary changes. J Neuropathol Exp Neurol 1998, 58:1010-1019.
- 129. Bhat RV, Leonov S, Luthman J, Scott CW, Lee CM: Interactions between GSK3beta and caspase signalling pathways during NGF deprivation induced cell death. J Alzheimers Dis 2002, 4:291-301.
- 130. Nicoll JA, Wilkinson D, Holmes C, Steart P, Markham H, Weller RO: Neuropathology of human Alzheimer disease after immunization with amyloid-β peptide: a case report. Nat Med 2003, 9:448-452.
- 131. Hock C, Konietzko U, Streffer JR, Tracy J, Signorell A, Muller-Tillmanns B, Lemke U, Henke K, Moritz E, Garcia E, Wollmer MA, Umbricht D, de Querbain DJ, Hofmann M, Maddalena A, Papasotiropoulos A, Nitsch RM: Antibodies against β-amyloid slow cognitive decline in Alzheimer's disease. Neuron 2003, 38:547-554.
- 132. Raoul C, Pettmann B, Henderson CE: Active killing of neurons during development and following stress: a role for p75NTR and Fas? Curr Opin Neurobiol 2000, 10:111-117.
- 133. Ethell DW, Buhler LA: Fas ligand-mediated apoptosis in degenerative disorders of the brain. J Clin Immunol 2003, 23:439-446.
- 134. Nagy Z, Esiri MM, Cato AM, Smith AD: Cell cycle markers in the hippocampus in Alzheimer's disease. Acta Neuropathol 1997, 94:6-15.
- 135. Arendt T, Holzer M, Gartner U: Neuronal expression of cycline dependent kinase inhibitors of the INK4 family in Alzheimer's disease. J Neural Transm 1998, 105:949-960.
- 136. Yang Y, Mufson EJ, Herrup K: Neuronal cell death is preceded by cell cycle events at all stages of Alzheimer's disease. J Neurosci 2003, 23:2557.
- 137. Beere HM: 'The stress of dying': the role of heat shock proteins in the regulation of apoptosis. J Cell Sci 2004, 117:2641-2651.
- 138. Deshmukh M, Johnson EM Jr: Evidence of a novel event during neuronal death: Development of competence-to-die in response to cytoplasmic cytochrome c. Neuron 1998, 21:695-705.
- 139. Putcha GV, Harris CA, Moulder KL, Easton RM, Thompson CB, Johnson EM Jr: Intrinsic and extrinsic pathway signaling during neuronal apoptosis: lesson from the analysis of mutant mice. J Cell Biol 2002, 157:441-453.

- 140. Chong ZZ, Li F, Maiese K: Stress in the brain: novel cellular mechanisms of injury linked to Alzheimer's disease. Brain Res Brain Res Rev 2005, 49:1-21.
- Chang HY, Yang X: Proteases for cell suicide: functions and regulation of caspases. Microbiol Mol Biol Rev 2000, 64:821-846.
- 142. Chan SL, Mattson MP: Caspase and calpain substrates: role in synaptic plasticity and cell death. J Neurosci Res 1999, 58:167-190.
- 143. Putcha GV, Deshmukh M, Johnson EM Jr: Bax translocation is a critical event in neuronal apoptosis: regulation by neuroprotectants, Bcl-2, and caspases. *J Neurosci* 1999, 19:7476-7485.
 144. Li HL, Zhu H, Xu CJ, Yuan JY: Cleavage of BID by caspase 8
- 144. Li HL, Zhu H, Xu CJ, Yuan JY: Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. Cell 1998, 94:491-501.
- 145. Luo X, Budihardjo I, Zou H, Slaughter C, Wang XD: BID, a BCL-2 interacting protein, mediates cytochrome c release from mitochondria in response to activation of cell surface death receptors. Cell 1998, 94:481-490.
- 146. Plesnila N, Zinkel S, Le DA, Amin-Hanjani S, Wu Y, Qiu J, Chiarugi A, Thomas AA, Kohane DS, Korsmeyer SJ, Moskowitz MA: Bid mediates neuronal cell death after oxygen/glucose deprivation and focal cerebral ischemia. Proc Natl Acad Sci USA 2001, 98:15318-15323.
- 147. Culmsee C, Zhu C, Landshamer S, Becattini B, Wagner E, Pellechia M, Blomgren K, Plesnila N: Apoptosis-Inducing Factor triggered by Poly(ADP-Ribose) Polymerase and Bid mediates neuronal death after oxygen-glucose deprivation and focal cerebral ischemia. J Neurosci 2005, 25:10262-10272.
- 148. Chao MV: Neurotrophins and their receptors: a convergence point for many signalling pathways. Nat Rev Neurosci 2003, 4:299-309.
- 149. Ryungsa K: Unknotting the roles of Bcl-2 and Bcl-xL in cell death. Biochem Biophys Res Commun 2005, 333:336-343.
- Cardone MH, Roy N, Stennicke HR, Salvesen GS, Franke TF, Stanbridge E, Frisch S, Reed JC: Regulation of cell death protease caspase-9 by phosphorylation. Science 1998, 282:1318-1321.
- 151. Brunet A, Bonni A, Zigmond MJ, Lin MZ, Juo P, Hu LS, Anderson MJ, Arden KC, Blenis J, Greenberg ME: Akt promotes cell survival by phosphorylating and inhibiting a Forkhead transcription factor. Cell 1999, 96:857-868.
- 152. Caricasole A, Copani A, Caruso A, Caraci F, Iacovelli L, Sorino AM, Terstappen GC, Nicoletti F: The Wnt pathway, cell-cycle activation and b-amyloid: novel therapeutic strategies in Alzheimer's disease? Thends Pharmacol Sci 2003, 24:233-238.
- 153. Aloyz RS, Bamji SX, Pozniak CD, Toma JG, Atwal J, Kaplan DR, Miller FD: P53 is essential for developmental neuron death as regulated by the trkA and p75 neurotrophin receptors. J Cell Biol 1998, 143:1691-1703.
- 154. Smith WW, Norton DD, Gorospe M, Jiang H, Nemoto S, Holbrook NJ, Finkel T, Kusiak JW: Phosphorylation of p66Shc and forkhead proteins mediates Aβ toxicity. J Cell Biol 2005, 169:331-339.
- 155. Garrido JL, Godoy JA, Alvarez A, Bronfman M, Inestrosa NC: Protein kinase C inhibits amyloid beta peptide neurotoxicity by acting on members of the Wnt pathway. FASEB J 2002, 16:1982-1984.
- 156. Mesner PW, Winters TR, Green SH: Nerve Growth Factor withdrawal-induced cell death in neuronal PC12 cells resembles that in sympathetic neurons. J Cell Biol 1992, 119:1669-1680.
- Friedman WJ: Neurotrophins induce death of hippocampal neurons via the p75 receptor. J Neurosci 2000, 20:6340-6346.
- 158. Casaccia-Bonnefil P, Carter BD, Dobrowsky RT, Chao MV: Death of oligodendrocytes mediated by the interaction of nerve growth factor with its receptor p75. Nature 1996, 383:716-719.
- 159. Salehi AH, Xanthoudakis S, Barker PA: NRAGE, a p75 Neurotrophin Receptor-interacting protein, induces caspase activation and cell death through a JNK-dependent mitochondrial pathway. J Biol Chem 2002, 277:48043-48050.
- 160. Khursigara G, Bertin J, Yano H, Moffett H, DiStefano PS, Chao MV: A prosurvival function for the p75 receptor death domain mediated via the caspase recruitment domain Receptor-Interacting Protein 2. J Neurosci 2001, 21:5854-5863.
- 161. Khursigara G, Orlinick JR, Chao MV: Association of the p75 neurotrophin receptor with TRAF6. J Biol Chem 1999, 274:2597-2600.

- 162. Dobrowsky RT, Werner MH, Castellino AM, Chao MV, Hannun YA: Activation of the sphingomyelin cycle through the low affinity neurotrophin receptor. Science 1994, 265:1596-1598.
- 163. Harrington AW, Kim JY, Yoon SO: Activation of Rac GTPase by p75 is necessary for JNK-mediated apoptosis. J Neurosci 2001, 22:156-166.
- 164. Zhou H, Summers SA, Birnbaum MJ, Pittman RN: Inhibition of Akt kinase by cell-permeable ceramide and its implications for ceramide-induced apoptosis. J Biol Chem 1998, 273:16568-16575.
- 165. Stoica BA, Movsesyan VA, LealV PM, Fade AI: Ceramide-induced neuronal apoptosis is associated with dephosphorylation of Akt, BAD, FKHR, GSK-3β, and induction of the mitochondrial-dependent intrinsic caspase pathway. *Mol Cell Neurosci* 2003, 22:365-382.
- 166. Carter BD, Kaltschmidt C, Kaltschmidt B, Offenhauser N, Bohm Matthaei R, Baeuerle PA, Barde Y-A: Selective activation of NF-kB by nerve growth factor through the neurotrophin receptor p75. *Science* 1996, 272:542-545.
- 167. Hamanoue M, Middleton G, Wyatt S, Jaffrey E, Hay RT, Davies AM: p75-mediated NF-kB activation enhances the survival response of developing sensory neurons to nerve growth factor. Mol Cell Neurosci 1999, 14:28-40.
- 168. Yamashita T, Tucker KL, Barde T: Neurotrophin binding to the p75 receptor modulates rho activity and axonal growth. Neuron 1999, 24:585-593.
- 169. Pike CJ, Ramezan-Arab N, Cotman CW: Beta-amyloid neurotoxicity in vitro : evidence of oxidative stress but not protection by antioxidants. J Neurochem 1997, 69:1601-1611.
- 170. Cherny RA, Atwood CS, Xilinas ME, Gray DN, Jones WD, McLean CA, Barnham KJ, Volitakis I, Fraser FW, Kim Y, Huang X, Goldstein LE, Moir RD, Lim JT, Beyreuther K, Zheng H, Tanzi RE, Masters CL, Bush Al: Treatment with a copper-zinc chelator markedly and rapidly inhibits beta-amyloid accumulation in Alzhe-imer's disease transgenic mice. Neuron 2001, 30:665-676.
- 171. Troy CM, Rabacchi SA, Xu Z, Maroney AC, Connors TJ, Shelanski ML, Greene LA: beta-amyloid-induced neuronal apoptosis requires c-Jun N-terminal kinase activation. J Neurochem 2001, 77:157-164.
- 172. Rabizadeh S, Bitker CM, Butcher LL, Bredesen DE: Expression of the low-affinity nerve growth factor receptor enhances βamyloid peptide toxicity. Proc Natl Acad Sci USA 1994, 91:10703-10706.
- 173. Yaar M, Zhai S, Pilch PF, Doyle SM, Eisenhauer PB, Fine RE, Gilchrest BA: Binding of beta-amyloid to the p75 neurotrophin receptor induces apoptosis. A possible mechanism for Alzheimer's disease. / Clin Invest 1997, 100:2333-2340.
- 174. Costantini C, Rossi F, Formaggio E, Bernardoni R, Cecconi D, Della-Bianca V: Characterization of the signaling pathway downstream p75 neurotrophin receptor involved in β-amyloid peptide-dependent cell death. J Mol Neurosci 2005, 25:141-156.
- 175. Yao M, Nguyen TV, Pike CJ: β-amyloid-induced neuronal apoptosis involves c-Jun N-terminal kinase-dependent downregulation of Bcl-w. J Neurosci 2005, 25:1149-1158.
- 176. Pei JJ, Braak E, Braak H, Grundke-Iqbal I, Iqbal K, Winblad B, Cowburn RF: Localization of active forms of C-jun kinase (JNK) and p38 kinase in Alzheimer's disease brains at different stages of neurofribillary degeneration. J Alzheimers Dis 2001, 3:41-48.
- 177. Hashimoto Y, Kaneko Y, Tsukamoto E, Frankowski H, Kouyama K, Kita Y, Niikura T, Aiso S, Bredesen DE, Matsuoka M, Nishimoto I: Molecular characterization of neurohybrid cell death induced by Alzheimer's amyloid-β peptides via p75NTR/ PLAIDD. J Neurochem 2004, 90:549-558.
- 178. Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N, Caspersen C, Chen X, Pollak S, Chaney M, Trinchese F, Liu S, Gunn-Moore F, Lue LF, Walker DG, Kuppusamy P, Zewier ZL, Arancio O, Stern D, Yan SS, Wu H: ABAD directly links Aβ to mitochondrial toxicity in Alzheimer's disease. Science 2004, 304:448-452.
- 179. Troy CM, Rabacchi SA, Friedman WJ, Frappier TF, Brown K, Shelanski ML: Caspase-2 mediates neuronal cell death induced by βamyloid. J Neurosci 2000, 20:1386-1392.
- 180. Guo H, Albrecht S, Bourdeau M, Petzke T, Bergeron C, LeBlanc AC: Active caspase-6 and caspase-6-cleaved tau in neuropil threads, neuritic plaques, and neurofibrillary tangles of Alzheimer's disease. Am J Pathol 2004, 165:523-531.

- 181. Cantarella G, Uberti D, Carsana T, Lombardo G, Bernardini R, Memo M: Neutralization of TRAIL death pathway protects human neuronal cell line from β-amyloid toxicity. Cell Death Differ 2003, 10:134-141.
- 182. Kaltschmidt B, Uherek M, Volk B, Baeuerle PA, Kaltschmidt C: Transcription factor NF-kB is activated in primary neurons by amyloid β peptides and in neurons surrounding early plaques from patients with Alzheimer's disease. Proc Natl Acad Sci USA 1997, 94:2642-2647.
- 183. Choi DW: Calcium and excitotoxic neuronal injury. Ann N Y Acad Sci 1994, 747:162-171.
- 184. Vanhoutte P, Barnier JV, Guibert B, Pages C, Besson MJ, Hipskind RA, Caboche J: Glutamate induces phosphorylation of Elk-1 and CREB, along with c-fos activation, via an extracellular signalregulated kinase-dependent pathway in brain slices. Mol Cell Biol 1999, 19:136-146.
- 185. De Bernardi MA, Rabin SJ, Colangelo AM, Brooker G, Mocchetti I: TrkA mediates the nerve growth factor-induced intracellular calcium accumulation. J Biol Chem 1996, 271:6092-6098.
- 186. Wood DE, Thomas A, Devi LA, Berman Y, Beavis RC, Reed JC, Newcomb EW: Bax cleavage is mediated by calpain during druginduced apoptosis. Oncogene 1998, 17:1069-1078.
- 187. Adamec E, Mohan P, Vonsattel JP, Nixon RA: Calpain activation in neurodegenerative diseases: confocal immunofluorescence study with antibodies specifically recognizing the active form of calpain 2. Acta Neuropathol 2002, 104:92-104.
- 188. Lee M-S, Kwon YT, Li M, Peng J, Friedlander RM, Tsai L-H: Neurotoxicity induces cleavage of p35 to p25 by calpain. Nature 2000, 405:360-364.
- 189. Patrick GN, Zukerberg L, Nikolic M, de la Monte S, Dikkes P, Tsai L-H: Conversion of p35 to p25 deregulates cdk5 activity and promotes neurodegeneration. Nature 2000, 402:615-622.
- 190. Sadoul R, Fernandez PA, Quiquerez AL, Martinou I, Maki M, Schroter M, Becherer JD, Irmler M, Tschopp J, Martinou JC: Involvement of the proteasome in the programmed cell death of NGFdeprived sympathetic neurons. *EMBO J* 1996, 15:3845-3852.
- 191. Canu N, Barbato C, Ciotti MT, Serafino A, Dus L, Calissano P: Proteasome involvement and accumulation of ubiquitinated proteins in cerebellar granule neurons undergoing apoptosis. J Neurosci 2000, 20:589-599.
- 192. Lam YA, Pickart CM, Alban A, Landon M, Jamieson C, Ramage R, Mayer RJ: Inhibition of the ubiquitin-proteasome system in Alzheimer's diease. Proc Natl Acad Sci USA 2000, 97:9902-9906.
- 193. Jick H, Zornberg GL, Jick SS, Seshadri S, Drachman DA: **Statins and** the risk of dementia. *Lancet* 2000, **356**:1627-1631.
- 194. Puglielli L, Tanzi RE, Kovacs DM: Alzheimer's disease: the cholesterol connection. Nat Neurosci 2003, 6:345-351.
- 195. Mattson MP, Pedersen WA, Duan W, Culmsee C, Camandola S: Cellular and molecular mechanisms underlying perturbed energy metabolism and neuronal degeneration in Alzheimer's and Parkinson's diseases. Ann N Y Acad Sci 1999, 893:154-175.
- 196. Castro-Alamancos MA, Torres-Aleman I: Long-term depression of glutamate-induced gamma-aminobutyric acid release in cerebellum by insulin-like growth factor I. Proc Natl Acad Sci USA 1993, 90:7386-7390.
- 197. Blair LA, Marshall J: IGF-I modulates N and L calcium channels in a PI3-kinase-dependent manner. Neuron 1997, 19:412-429.
- Carro E, Trejo JL, Gomez-Isla T, LeRoith D, Torres-Aleman I: Serum insulin-like growth factor I regulates brain amyloid beta levels. Nat Med 2002, 8:1390-1397.
- 199. Dore S, Kar S, Quirion R: Insulin-like growth factor I protects and rescues hippocampal neurons against β-amyloid- and human amylin-induced toxicity. Proc Natl Acad Sci USA 1997, 94:4772-4777.
- 200. Shaw M, Cohen P, Alessi DR: Further evidence that the inhibition of glycogen synthase kinase-3β by IGF-I is mediated by PDK1/PKB-induced phosphorylation of Ser-9 and not by dephosphorylation of Tyr-216. FEBS Lett 1997, 416:307-311.
- 201. Zheng WH, Kar S, Dore S, Quirion R: Insulin-like growth factor-I (IGF-1): a neuroprotective trophic factor acting via the Akt kinase pathway. J Neural Transm Suppl 2000, 60:261-272.
- 202. Aloe L: Intracerebral pretreatment with nerve growth factor prevents irreversible brain lesions in neonatal rats injected with ibotenic acid. *Biotechnology* 1987, **5**:1085-1086.

- 203. Cheng B, Mattson MP: NGF and bFGF protect rat and human central neurons against hypoglycemic damage by stabilizing calcium homeostasis. *Neuron* 1991, **7:**1031-1041.
- Tetsu O, McCormick F: Beta-catenin regulates expression of cyclin D1 in colon carcinoma cells. Nature 1999, 398:422-426.
- Herrup K, Neve R, Ackerman SL, Copani A: Divide and die: cell cycle events as triggers of nerve cell death. J Neurosc 2004, 24:9232-9239.
- Copani A, Condorelli F, Caruso A, Vancheri C, Sala A, Giuffrida SAM, Canonico PL, Nicoletti F, Sortino MA: Mitotic signaling by betaamyloid causes neuronal death. Faseb J 1999, 13:2225-2234.
- 207. Park DS, Levine B, Ferrari G, Greene LA: Cyclin dependent kinase inhibitors and dominant negative cyclin dependent kinase 4 and 6 promote survival of NGF-deprived sympathetic neurons. / Neurosci 1997, 17:8975-8983.
- Park DS, Morris EJ, Padmanabhan J, Shelanski ML, Geller HM, Greene LA: Cyclin-dependent kinases participate in death of neurons evoked by DNA-damaging agents. J Cell Biol 1998, 143:457-467.
- 209. Zeng G, Meakin SO: Overexpression of the signaling adapter FRS2 reconstitutes the cell cycle deficit of a nerve growth factor non-responsive trkA receptor mutant. J Neurochem 2002, 81:820-831.
- 210. McShea A, Harris PL, Webster KR, Wahl AF, Smith MA: Abnormal expression of the cell cycle regulators P16 and CDK4 in Alzheimer's disease. Am J Pathol 1997, 150:1933-1939.
- 211. Ueberham U, Hessel A, Arendt T: Cyclin C expression is involved in the pathogenesis of Alzheimer's disease. Neurobiol Aging 2003, 24:427-435.
- 212. Frasca G, Chiechio S, Vancheri C, Nicoletti F, Copani A, Sortino A: βamyloid-activated cell cycle in SH-SY5Y neuroblastoma cells: correlation with the MAP kinase pathway. J Mol Neurosci 2004, 22:231-236.
- 213. Luth HJ, Holzer M, Gertz HJ, Arendt T: Aberrant expression of nNOS in pyramidal neurons in Alzheimer's disease is higly co-localized with p21ras and p16INK4a. Brain Res 2000, 852:45-55.
- 214. Giovanni A, Keramaris E, Morris EJ, Hou ST, O'Hare M, Dyson N, Robertson GS, Slack RS, Park DS: E2F1 mediates death of β-amyloid-treated cortical neurons in a manner independent of p53 and dependent on Bax and caspase 3. J Biol Chem 2000, 275:11553-11560.
- 215. Chen Y, McPhie DL, Hirschberg J, Neve RL: The amyloid precursor protein-binding protein APP-BPI drives the cell cycle through the S-M checkpoint and causes apoptosis in neurons. J Biol Chem 2000, 275:8929-8935.
- 216. McPhie DL, Coopersmith R, Hines-Peralta A, Chen Y, Ivins KJ, Manly SP, Kozlowski MR, Neve KA, Neve RL: DNA synthesis and neuronal apoptosis caused by familial Alzheimer disease mutants of the amyloid precursor protein are mediated by the p21 activated kinase PAK3. J Neurosci 2003, 23:6914-6927.
- 217. Zhao L, Ma Q-L, Calon F, Harris-White ME, Yang F, Lim GP, Morihara T, Ubeda OJ, Ambergaokar S, Hansen JE, Weisbart RH, Teter B, Frautschy SA, Cole GM: Role of p21-activated kinase pathway defects in the cognitive deficits of Alzheimer disease. Nat Neurosci 2006, 9:234-242.
- 218. McGeer PL, McGeer EG: The inflammatory response system of brain: implication for therapy of Alzheimer and other neurodegenerative diseases. *Brain Res Rev* 1995, 21:195-218.
- Bruce-Keller AJ: Microglia-neuronal interactions in synaptic damage and recovery. J Neurosci Res 1999, 58:191-201.
 Rogers J, Cooper NR, Webster S, Schultz J, McGeer PL, Styren SD,
- 220. Rogers J, Cooper NR, Webster S, Schultz J, McGeer PL, Styren SD, Civin WH, Brachova L, Bradt B, Ward P, Lieberburg I: Complement activation by β-amyloid in Alzheimer disease. Proc Natl Acad Sci USA 1992, 89:10016-10020.
- 221. Rogers J, Kirby LC, Hempelman SR, Berry DL, McGeer PL, Kaszniak AW, Zalinski J, Cofield M, Mansukhani L, Willson P, Kogan F: Clinical trial of indomethacin in Alzheimer's disease. Neurology 1993, 43:1609-1611.
- 222. Weggen S, Eriksen JL, Das P, Sagi SA, Wang R, Pietrzik CU, Findlay KA, Smith TE, Murphy MP, Bulter T, Kang DE, Marquez-Sterling N, Golde TE, Koo EH: A subset of NSAIDs lower amyloidogenic Abeta42 independently of cyclooxygenase activity. Nature 2001, 414:212-216.
- 223. Bachis A, Colangelo AM, Vicini S, Doe PP, De Bernardi MA, Brooker G, Mocchetti I: Interleukin-10 prevents glutamate-mediated

cerebellar granule cell death by blocking caspase-3-like activity. / Neurosci 2001, 21:3104-3112.

- Martin DP, Schmidt RE, DiStefano PS, Lowry OH, Carter JC, Johnson EM Jr: Inhibitors of protein synthesis and RNA synthesis prevent neuronal death caused by Nerve Growth Factor deprivation. J Cell Biol 1988, 106:829-844.
 Suzuki A, Tsutomi Y, Akahane K, Araki T, Miura M: Resistance to
- 225. Suzuki A, Tsutomi Y, Akahane K, Araki T, Miura M: Resistance to Fas-mediated apoptosis: activation of caspase 3 is regulated by cell cycle regulator p21^{WAF1} and IAP gene family ILP. Oncogene 1998, 7:931-939.
- 226. Eymin B, Sordet O, Droin N, Munsch B, Haugg M, Van de Craen M, Vandenabeele P, Solary E: Caspase-induced proteolysis of the cyclin-dependent kinase inhibitor p27^{Kip1} mediates its antiapoptotic activity. Oncogene 1999, 18:4839-4847.
- 227. Blagosklonny MV: Are p27 an p21 cytoplasmic oncoproteins? Cell Cycle 2002, 1:391-393.
- 228. Hardy J, Selkoe DJ: The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002, **297**:353-356.
- 229. Alberghina L, Vai M, Vanoni M: Probing control mechanisms of cell cycle and ageing in budding yeast. Curr Genomics 2004, 5:615-627.
- 230. Ueberham U, Arendt T: The expression of cell cycle proteins in neurons and its relevance for Alzheimer's disease. Curr Drug Targets CNS Neurol Disord 2005, 4:293-306.

