

# The natural furanone (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone disrupts quorum sensing-regulated gene expression in *Vibrio harveyi* by decreasing the DNA-binding activity of the transcriptional regulator protein luxR

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## Summary

This study aimed at getting a deeper insight in the molecular mechanism by which the natural furanone (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone disrupts quorum sensing in *Vibrio harveyi*. Bioluminescence experiments with signal molecule receptor double mutants revealed that the furanone blocks all three channels of the *V. harveyi* quorum sensing system. In further experiments using mutants with mutations in the quorum sensing signal transduction pathway, the compound was found to block quorum sensing-regulated bioluminescence by interacting with a component located downstream of the Hfq protein. Furthermore, reverse transcriptase real-time polymerase chain reaction with specific primers showed that there was no effect of the furanone on *luxR<sub>Vh</sub>* mRNA levels in wild-type *V. harveyi* cells. In contrast, mobility shift assays showed that in the presence of the furanone, significantly lower

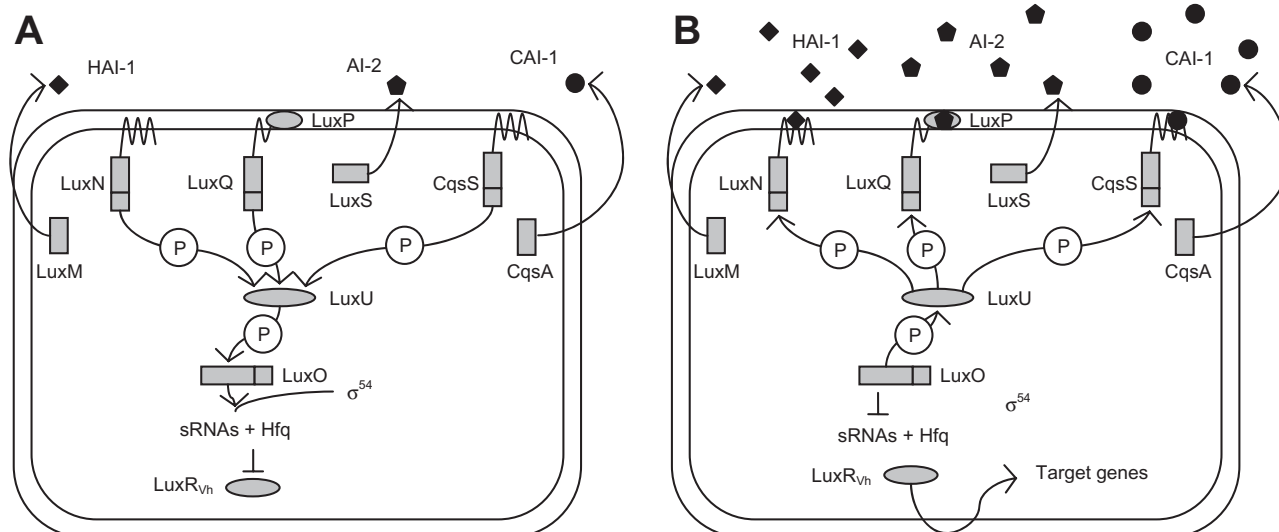
levels of the LuxR<sub>Vh</sub> response regulator protein were able to bind to its target promoter sequences in wild-type *V. harveyi*. Finally, tests with purified LuxR<sub>Vh</sub> protein also showed less shifts with furanone-treated LuxR<sub>Vh</sub>, whereas the LuxR<sub>Vh</sub> concentration was found not to be altered by the furanone (as determined by SDS-PAGE). Therefore, our data indicate that the furanone blocks quorum sensing in *V. harveyi* by rendering the quorum sensing master regulator protein LuxR<sub>Vh</sub> unable to bind to the promoter sequences of quorum sensing-regulated genes.

## Introduction

*Vibrio harveyi* is a Gram-negative, luminescent marine bacterium that can be found free-living in the water column as well as in association with marine animals (Thompson *et al.*, 2004). With the rapid development of the aquaculture industry, the species is becoming increasingly recognized as an important pathogen of marine vertebrates and invertebrates (Austin and Zhang, 2006). Because of the development and spread of antibiotic resistance in these bacteria, antibiotic treatments are becoming inefficient (Karunasagar *et al.*, 1994; Moriarty, 1998) and therefore, alternative control strategies are being developed. One of these strategies is the disruption of bacterial cell-to-cell communication, called quorum sensing (Defoirdt *et al.*, 2004).

Unlike many other Gram-negative bacteria, *V. harveyi* has been reported to use a multichannel quorum sensing system (Fig. 1). The first channel of this system is mediated by the Harveyi Autoinducer 1 (HAI-1), an acylated homoserine lactone (AHL) (Cao and Meighen, 1989). The second channel is mediated by the so-called Autoinducer 2 (AI-2), which is a furanosyl borate diester (Chen *et al.*, 2002). The chemical structure of the third autoinducer, called Cholerae Autoinducer 1 (CAI-1) is still unknown (Henke and Bassler, 2004a). All three autoinducers are detected at the cell surface and activate or inactivate target gene expression by a phosphorylation/dephosphorylation signal transduction cascade. Phenotypes that were found to be controlled by this quorum

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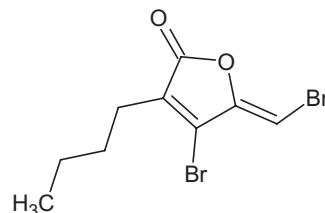
**Fig. 1.** Quorum sensing in *Vibrio harveyi*. The LuxM, LuxS and CqsA enzymes synthesize the autoinducers HAI-1, AI-2 and CAI-1 respectively. These autoinducers are detected at the cell surface by the LuxN, LuxP-LuxQ and CqsS receptor proteins respectively. A. At low signal molecule concentration, the receptors autophosphorylate and transfer phosphate to LuxO via LuxU. Phosphorylation activates LuxO, which together with  $\sigma^{54}$  activates the production of small regulatory RNAs (sRNAs). These sRNAs, together with the chaperone Hfq, destabilize the mRNA encoding the response regulator LuxR<sub>Vh</sub>. Therefore, in the absence of autoinducers, the LuxR<sub>Vh</sub> protein is not produced. B. In the presence of high concentrations of the autoinducers, the receptor proteins switch from kinases to phosphatases, which results in dephosphorylation of LuxO. Dephosphorylated LuxO is inactive and therefore, the sRNAs are not formed and the response regulator LuxR<sub>Vh</sub> is produced (adapted from Henke and Bassler, 2004a).

sensing system include bioluminescence (Bassler *et al.*, 1993) and the production of several virulence factors such as a type III secretion system (Henke and Bassler, 2004b), extracellular toxin (Manefield *et al.*, 2000) and a siderophore (Lilley and Bassler, 2000). Recently, we found that virulence of the bacterium towards the brine shrimp *Artemia franciscana* is also regulated by its quorum sensing system (Defoirdt *et al.*, 2005).

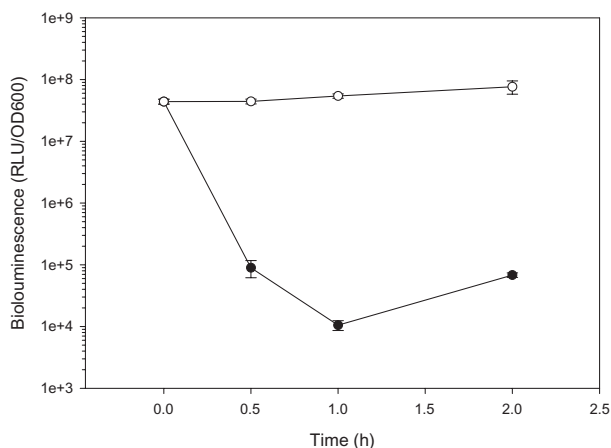
One of the mechanisms to disrupt bacterial quorum sensing is the use of halogenated furanones, such as the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone. These furanones (natural occurring compounds as well as synthetic derivatives) were found to disrupt the expression of different AHL-regulated phenotypes in several Gram-negative species, without affecting their growth (Hentzer and Givskov, 2003; Rasmussen and Givskov, 2006). Because of the structural similarity between AHL molecules and the furanones, it was originally hypothesized that these compounds disrupt AHL-mediated quorum sensing in the LuxI/LuxR<sub>Vf</sub>-type of quorum sensing system by competitively binding to the AHL receptor site of the *Vibrio fischeri* LuxR<sub>Vf</sub> protein (Givskov *et al.*, 1996). Later on, it was shown that the halogenated furanones promote rapid turnover of the LuxR<sub>Vf</sub>-type AHL receptor protein, reducing the amount of LuxR<sub>Vf</sub> available to interact with AHL and to act as transcriptional regulator (Manefield *et al.*, 2002). More recently, Koch and colleagues (2005) used site-directed mutagenesis to study interactions between LuxR<sub>Vf</sub> and

halogenated furanones. The authors could not conclude that the furanones bind to the AHL-binding cavity and suggested that furanones do not compete in a classic way with the signal molecules.

Meanwhile, several research groups provided evidence that halogenated furanones also disrupt quorum sensing-regulated gene expression in *V. harveyi* (Manefield *et al.*, 2000; Ren *et al.*, 2001; McDougald *et al.*, 2003; Defoirdt *et al.*, 2006). However, because the quorum sensing system of *V. harveyi* is quite different from the LuxI/LuxR<sub>Vf</sub>-type of quorum sensing system and does not contain a LuxR<sub>Vf</sub> homologue (Milton, 2006), the molecular mechanism of quorum sensing disruption in this species must be different from that in the LuxI/LuxR<sub>Vf</sub>-type of system. Consequently, in this study, we aimed at defining the molecular mechanism by which the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone (Fig. 2) disrupts quorum sensing-regulated gene expression in this different type of quorum sensing system.



**Fig. 2.** Structure of the natural furanone used in this study.



**Fig. 3.** Bioluminescence per unit cell density in wild-type *Vibrio harveyi* BB120 as a function of time, without (open symbols) and with (filled symbols) the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone (50 mg l<sup>-1</sup>). The error bars represent the standard deviation of three replicates. Note that RLU is the relative unit of luminescence reported by the Lumac Biocounter M2500 luminometer.

## Results

### *Impact of the furanone on quorum sensing-regulated bioluminescence of wild-type V. harveyi*

Bioluminescence is one of the phenotypes that is regulated by the *V. harveyi* quorum sensing system and therefore, in a first experiment, the impact of the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone on the bioluminescence of wild-type *V. harveyi* was determined. Strain BB120 was grown in LB<sub>20</sub> medium to high cell density in order to activate quorum sensing-regulated bioluminescence, after which the furanone was added to the medium at 50 mg l<sup>-1</sup>. The compound blocked bioluminescence of BB120, with over 3 log units difference between the furanone-treated and the untreated cultures already 0.5 h after the addition of the furanone (Fig. 3). Consistent with this, luciferase activity in protein lysates of furanone-treated BB120 cells was found to have decreased with approximately 1 log unit 0.5 h after the addition of 50 mg l<sup>-1</sup> furanone (Fig. 4). The bacterial alkaline phosphatase activities of the protein lysates were also measured in order to verify that the furanone had no effect on phenotypes that are not regulated by the quorum sensing system. As expected, there was no significant difference in bacterial alkaline phosphatase activities between furanone-treated and untreated cells (Fig. 4).

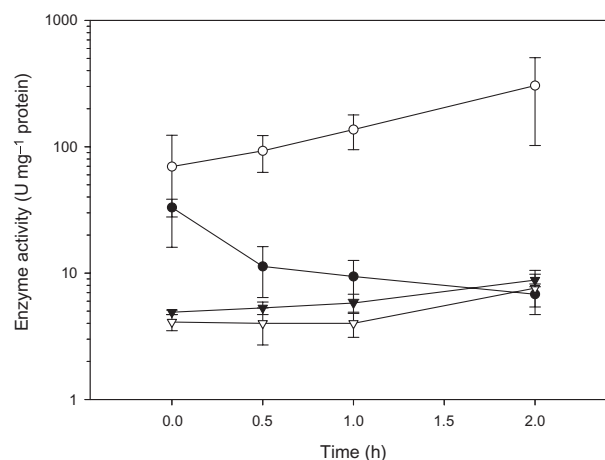
### *Impact of the furanone on bioluminescence of V. harveyi autoinducer receptor double mutants*

The *V. harveyi* quorum sensing system consists of three channels, with each channel being activated by a different type of signal molecule (Fig. 1). In order to determine

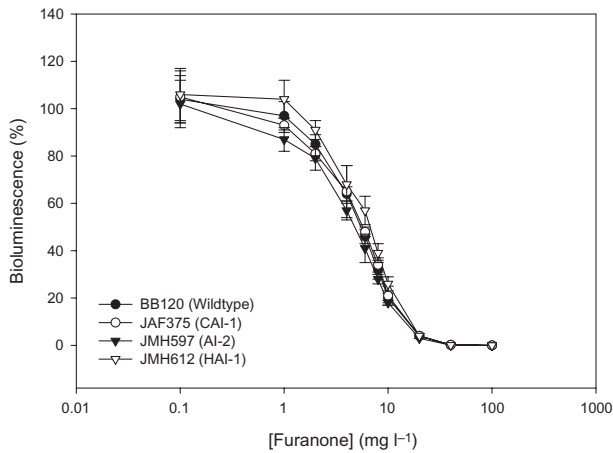
the impact of the furanone on the different channels, the signal molecule receptor double mutants JAF375 (sensor HAI-1<sup>-</sup>, sensor AI-2<sup>-</sup>, sensor CAI-1<sup>+</sup>), JMH597 (sensor HAI-1<sup>-</sup>, sensor AI-2<sup>+</sup>, sensor CAI-1<sup>-</sup>) and JMH612 (sensor HAI-1<sup>+</sup>, sensor AI-2<sup>-</sup>, sensor CAI-1<sup>-</sup>) were used. Because of the mutated receptors, bioluminescence in these mutants is only responsive to one of the three signal molecules (Henke and Bassler, 2004a). The mutants were grown to high cell densities, and after furanone addition, bioluminescence was found to be blocked in all three double mutants in a concentration-dependent way similar to the one obtained for the wild type (Fig. 5). This indicates that all three channels of the quorum sensing system were blocked.

### *Impact of the furanone on bioluminescence of constitutively luminescent V. harveyi mutants with mutations in the quorum sensing signal transduction cascade*

Because the three signal molecules have quite different chemical structures (Henke and Bassler, 2004a), we suspected that the furanone did not block quorum sensing-regulated bioluminescence by competing with the autoinducers for receptor sites but rather by interfering with the quorum sensing signal transduction. In order to test this hypothesis, the effects of the furanone on bioluminescence of mutants that were fixed in the high cell-density configuration at different stages in the quorum sensing signal transduction cascade were investigated. The mutants JAF553 and JAF483 contain a point mutation in the *luxU* and *luxO* genes, respectively, that render the LuxU and LuxO proteins incapable of phosphorelay (Freeman and



**Fig. 4.** Luciferase (circles) and bacterial alkaline phosphatase (triangles) activities in protein lysates of wild-type *Vibrio harveyi* BB120 as a function of time, without (open symbols) and with (filled symbols) the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone (50 mg l<sup>-1</sup>). The error bars represent the standard deviation of four independent experiments.



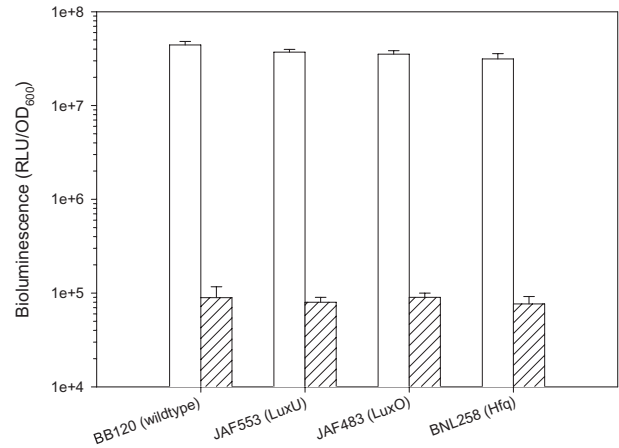
**Fig. 5.** Bioluminescence of the *Vibrio harveyi* wild type (BB120) and the double mutants JAF375 (sensor HAI-1<sup>-</sup>, sensor AI-2<sup>-</sup>, sensor CAI-1<sup>-</sup>), JMH597 (sensor HAI-1<sup>-</sup>, sensor AI-2<sup>+</sup>, sensor CAI-1<sup>-</sup>) and JMH612 (sensor HAI-1<sup>+</sup>, sensor AI-2<sup>-</sup>, sensor CAI-1<sup>-</sup>) as a function of the concentration of the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone. Luminescence measurements were performed 0.5 h after the addition of the furanone. For each strain, the bioluminescence without the addition of furanone was set at 100% and the other samples were normalized accordingly. The error bars represent the standard deviation of three replicates.

Bassler, 1999a,b). Strain BNL258 has a Tn5 insertion in the *hfq* gene, resulting in a non-functional Hfq protein (Lenz *et al.*, 2004). Hence, because of the nature of these mutations, the three mutants are constitutively luminescent and therefore, blocking luminescence in one of them would indicate that the furanone acts downstream of the mutated component. The furanone, at 50 mg l<sup>-1</sup>, blocked luminescence in all three mutants (Fig. 6). In wild-type *V. harveyi*, the Hfq protein acts together with small regulatory RNAs to destabilize the mRNA of the master regulator LuxR<sub>Vh</sub> (see Fig. 1). In the mutant BNL258, a transposon insertion has rendered the Hfq protein non-functional and therefore, in this mutant, the *luxR<sub>Vh</sub>* mRNA cannot be destabilized by the quorum sensing signal transduction system, resulting in a constitutively expressed bioluminescence (Lenz *et al.*, 2004). The fact that the furanone blocked bioluminescence in this mutant indicates that it acts downstream of Hfq, i.e. at the level of the *luxR<sub>Vh</sub>* mRNA and/or the LuxR<sub>Vh</sub> protein.

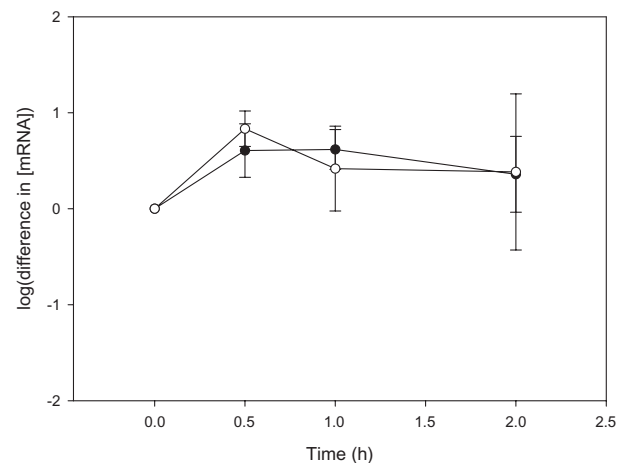
#### Impact of the furanone on *luxR<sub>Vh</sub>* mRNA levels

In order to verify whether the furanone indeed affects the quorum sensing master regulator, an experiment was set up in which the impact of the addition of the compound on *luxR<sub>Vh</sub>* mRNA was studied. Wild-type *V. harveyi* BB120 was grown to high cell density, after which the natural furanone was added at 50 mg l<sup>-1</sup>. The addition of the furanone resulted in a rapid decrease in luminescence (Fig. 3). *luxR<sub>Vh</sub>* mRNA levels were quantified relatively by

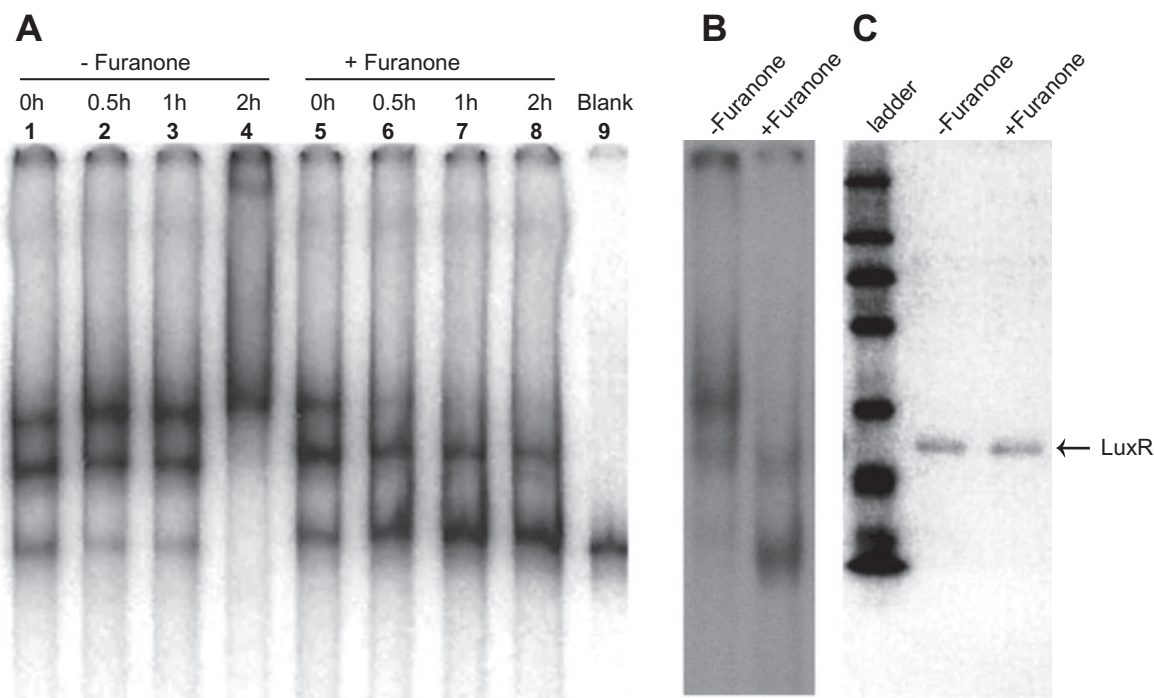
reverse transcriptase real-time polymerase chain reaction (PCR) with specific primers and the RNA polymerase A subunit (*rpoA*) mRNA was analysed as a control of a non-quorum sensing-regulated gene. For both genes, the mRNA levels followed a similar trend, with no significant difference between furanone-treated and untreated cells (Fig. 7). From these results, we conclude that the furanone has no effect on *luxR<sub>Vh</sub>* mRNA levels.



**Fig. 6.** Bioluminescence of wild-type *Vibrio harveyi* BB120 and the mutants JAF553 (*luxU* H58A), JAF483 (*luxO* D47A) and BNL258 (*hfq*:Tn5/*acZ*) without (white bars) and with (striped bars) the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone (50 mg l<sup>-1</sup>). Measurements were performed 0.5 h after the addition of the furanone. The error bars represent the standard deviation of three replicates. Note that RLU is the relative unit of luminescence reported by the Lumac Biocounter M2500 luminometer.



**Fig. 7.** Difference in *luxR<sub>Vh</sub>* (filled symbols) and *rpoA* (open symbols) mRNA levels between wild-type *Vibrio harveyi* BB120 cells treated with the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone (50 mg l<sup>-1</sup>) and untreated cells, as determined by reverse transcriptase real-time PCR with specific primers. The error bars represent the standard deviation of two independent experiments.



**Fig. 8.** LuxR<sub>Vh</sub> DNA binding as determined by mobility shifts.

A. Autoradiograph after 5% polyacrylamide gel electrophoresis of 5' <sup>32</sup>P-labelled *luxR<sub>Vh</sub>* promoter DNA containing the LuxR<sub>Vh</sub> binding sites, mixed with 0.5 µg of protein lysates from wild-type *Vibrio harveyi* BB120: lanes 1–4, addition of lysates taken at different time points from an untreated culture; lanes 5–8, addition of lysates taken at different time points from a culture treated with 50 mg l<sup>-1</sup> of the natural furanone (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone; last lane, blank (addition of lysate from the *luxR*-negative strain MR1130).

B. Autoradiograph after 5% polyacrylamide gel electrophoresis of 5' <sup>32</sup>P-labelled *luxR<sub>Vh</sub>* promoter DNA containing the LuxR<sub>Vh</sub> binding sites, mixed with purified LuxR<sub>Vh</sub>, which was incubated *in vitro* for 1 h with or without the furanone (50 mg l<sup>-1</sup>).

C. SDS-PAGE of the same samples as in panel B.

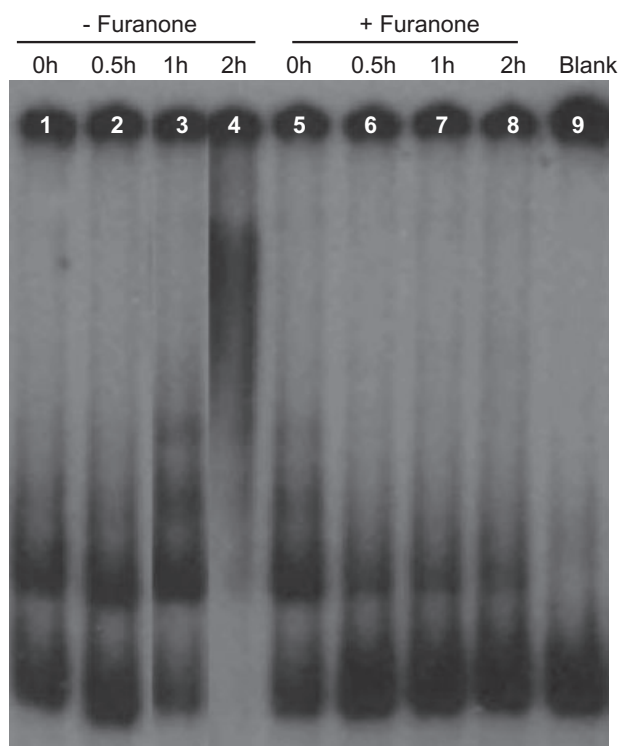
#### *Impact of the furanone on LuxR<sub>Vh</sub> protein levels and LuxR<sub>Vh</sub> binding to its target promoter sequences*

Mobility shifts using radiolabelled *luxR<sub>Vh</sub>* promoter DNA containing the LuxR<sub>Vh</sub> binding sites showed less shifts with cell lysates from furanone-treated cells when compared with untreated cells (Fig. 8A), indicating that there were significantly lower levels of the protein able to bind the promoter DNA. In order to quantify the difference in bound LuxR<sub>Vh</sub> levels between furanone-treated and untreated cells, a titration for mobility shifts of *luxR<sub>Vh</sub>* promoter DNA using the 2-h samples was performed. This titration revealed that 0.025 µg lysate of untreated cells gave the same shift as 0.5 µg lysate of furanone-treated cells (data not shown), which indicates that there was a 20-fold difference in LuxR<sub>Vh</sub> levels bound to the promoter DNA. The *luxCDABEGH* promoter DNA was also used to check for LuxR<sub>Vh</sub> shifts, with similar findings (Fig. 9). These observations could be explained by the furanone either decreasing translation or increasing turnover of LuxR<sub>Vh</sub>, or altering the protein in such a way that it is rendered unable to bind to its target promoter. Some additional tests were performed to clear this up. First, 50 mg l<sup>-1</sup> chloramphenicol was added to *V. harveyi*

BB120 cultures in order to stop translation of LuxR<sub>Vh</sub>. In these cultures, there was no effect on LuxR<sub>Vh</sub> mobility shifts in the absence of furanone throughout the 1 h of incubation, whereas in the presence of furanone, again less shifts occurred (data not shown). This suggests that LuxR<sub>Vh</sub> has quite a slow turnover and that the furanone acts on pre-existing LuxR<sub>Vh</sub>. Finally, the furanone was added to purified LuxR<sub>Vh</sub>, and after subsequent incubation of the mixture, again significantly lower levels of LuxR<sub>Vh</sub> were found to bind to the promoter DNA when compared with untreated LuxR<sub>Vh</sub> (Fig. 8B). Interestingly, SDS-PAGE showed that the concentration of LuxR<sub>Vh</sub> in the mixtures was not affected by the furanone (Fig. 8C). Together, these data lead to the conclusion that the furanone compound renders LuxR<sub>Vh</sub> unable to bind to its target promoter sequences, without degrading the protein.

#### Discussion

Disease caused by antibiotic resistant luminescent vibrios is a serious problem in the aquaculture industry (Austin and Zhang, 2006). Recent investigations have pointed out



**Fig. 9.** LuxR<sub>Vh</sub> DNA binding as determined by mobility shifts. Autoradiograph after 5% polyacrylamide gel electrophoresis of 5' <sup>32</sup>P-labelled *luxCDABEGH* promoter DNA containing the LuxR<sub>Vh</sub> binding sites, mixed with 0.5 µg of protein lysates from wild-type *Vibrio harveyi* BB120: lanes 1–4, addition of lysates taken at different time points from an untreated culture; lanes 5–8, addition of lysates taken at different time points from a culture treated with 50 mg l<sup>-1</sup> of the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone; last lane, blank (addition of lysate from the *luxR*-negative strain MR1130).

that disruption of the quorum sensing system of these bacteria by using halogenated furanones could be a promising alternative biocontrol strategy (Manefield *et al.*, 2000; Defoirdt *et al.*, 2006). In view of the potential practical applications of this type of compounds, it is of significant interest to define the mode of action of the furanones in these bacteria. Hence, in this study, we aimed at elucidating the molecular mechanism of quorum sensing disruption by the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone in *V. harveyi*.

In a first series of experiments, the impact of the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone on quorum sensing-regulated bioluminescence of *V. harveyi* wild type and quorum sensing mutants was determined. Importantly, halogenated furanones were shown before not to block bioluminescence when expressed from an external promoter (Givskov *et al.*, 1996), indicating that the biochemical function of the Lux proteins is not affected. Consistent with the work of Manefield and colleagues (2000), the compound was found to

block bioluminescence in wild-type *V. harveyi* BB120 in a concentration-dependent way. Moreover, luciferase activity was significantly decreased in protein lysates of furanone-treated BB120 cells. In order to determine the effect of the furanone on the three different channels of the *V. harveyi* quorum sensing system, its impact on bioluminescence of the signal molecule receptor double mutants JAF375, JMH597 and JMH612 (Henke and Bassler, 2004a) was studied. The compound was shown to block bioluminescence in all three double mutants in a pattern similar to the one obtained for the wild type. Because of the mutated receptors, bioluminescence in these mutants is only responsive to one of the three signal molecules, which implies that all three channels of the quorum sensing system were blocked. These data confirm the reports by Ren and colleagues (2001) and McDougald and colleagues (2003), who determined the impact of the furanone on the AI-2- and/or HAI-1-mediated channels of the system by using the AI-2 and/or HAI-1 receptor mutants BB886 and BB170 (which thus were still responsive to both HAI-1 and CAI-1 and AI-2 and CAI-1 respectively). Moreover, our results indicate that the furanone also blocks the CAI-1-mediated channel of the *V. harveyi* quorum sensing system.

Previously, the hypothesis that prevailed in literature was that the furanones disrupt quorum sensing in *V. harveyi* by displacing the signal molecules from their receptors (Manefield *et al.*, 2000; Ren *et al.*, 2001). However, because the three signal molecules have quite different chemical structures (although the exact structure of CAI-1 is still unknown; Henke and Bassler, 2004a), we suspected that the furanone did not block quorum sensing-regulated bioluminescence by competing with the autoinducers for receptor sites but rather by interfering with the quorum sensing signal transduction. In order to test this hypothesis, the effects of the furanone on bioluminescence of mutants that were fixed in the high cell-density configuration at different stages in the quorum sensing signal transduction cascade were investigated (i.e. LuxU, LuxO and Hfq). We found that the furanone blocked bioluminescence in the *hfq* mutant BNL258. Hfq is a chaperone protein that acts together with small regulatory RNAs to destabilize the mRNA of the master regulator LuxR<sub>Vh</sub>. The Hfq protein is non-functional in strain BNL258, resulting in constitutively expressed bioluminescence (Lenz *et al.*, 2004). The fact that the furanone blocked bioluminescence in this mutant indicates that it acts downstream of Hfq, i.e. at the level of the *luxR<sub>Vh</sub>* mRNA and/or the LuxR<sub>Vh</sub> protein and not by displacing the signal molecules from their receptors.

The quorum sensing master regulator protein LuxR<sub>Vh</sub> has been shown before to be a transcriptional activator that is required for expression of the *lux* operon (Swartzman *et al.*, 1992; Swartzman and Meighen, 1993).

Consequently, in a final series of experiments, the effect of the furanone on this response regulator protein was investigated. Reverse transcriptase real-time PCR with primers specific for *V. harveyi luxR<sub>Vh</sub>* revealed that the furanone has no effect on the concentration of the *luxR<sub>Vh</sub>* mRNA. In contrast, mobility shift assays showed that the concentration of the LuxR<sub>Vh</sub> response regulator protein able to bind to its target promoter sequences significantly decreased after furanone addition, both in intact cells and in purified LuxR<sub>Vh</sub> extracts. Interestingly, the concentration of the LuxR<sub>Vh</sub> protein was shown not to be affected by the furanone (as determined by SDS-PAGE analysis). LuxR<sub>Vh</sub> is a member of the TetR family of transcriptional regulators containing a helix–turn–helix DNA binding domain (Ramos *et al.*, 2005). Members of the TetR family that have been studied in depth, have been shown to bind DNA as dimers, but this has not yet been proven for LuxR<sub>Vh</sub>. Because halogenated furanones are known to be very reactive compounds (Hentzer and Givskov, 2003), we hypothesize that the furanone reacts with the LuxR<sub>Vh</sub> protein, thereby altering it in such a way that it cannot bind the DNA anymore, either by changing the structure of the DNA binding domain or the regions involved in dimer formation (that is, if LuxR<sub>Vh</sub> also needs to dimerize in order to bind DNA). However, the elucidation of the exact biochemical reaction mechanism between halogenated furanones and LuxR<sub>Vh</sub> (with the aid of mass spectrometry) will be part of future work at our laboratories.

The fact that the furanone affects the master regulator rather than selectively blocking one of the channels of the *V. harveyi* quorum sensing system is quite important with respect to possible practical applications because there seems to be a difference in the relative importance of the three channels for a successful infection of different hosts. Indeed, disrupting only the AI-2-mediated channel of the *V. harveyi* quorum sensing system has been shown to significantly increase survival of the brine shrimp *A. franciscana*, whereas the HAI-1-mediated channel had no effect on infection of the shrimp (Defoirdt *et al.*, 2005). In contrast, both the HAI-1- and AI-2-mediated channel needed to be disrupted in order to decrease mortality of gnotobiotic rotifers (*Brachionus plicatilis*) caused by *V. harveyi* (Tinh *et al.*, 2007). Because the furanone blocks all three channels of the system at once by acting at the end of the quorum sensing signal transduction cascade, it will not be necessary to develop different furanone compounds to protect different hosts. Consistent with this, the natural furanone was shown to protect both brine shrimp and rotifers from luminescent vibrios (Defoirdt *et al.*, 2006; Tinh *et al.*, 2007). In addition to this, several human pathogens, including *Vibrio cholerae*, *Vibrio parahaemolyticus* and *Vibrio vulnificus*, have been found to contain a LuxR<sub>Vh</sub> homologue (Jobling and Holmes, 1997; McCarter, 1998; McDougald

*et al.*, 2000). Moreover, the LuxR<sub>Vh</sub> homologues have been shown before to regulate virulence factor production in these species as well (Henke and Bassler, 2004b; Milton, 2006) and consequently, the furanones might be useful to control infections caused by these human pathogens.

In addition to affecting both *V. fischeri* LuxR<sub>Vf</sub> (Manfield *et al.*, 2002) and *V. harveyi* LuxR<sub>Vh</sub> (this study), covalent binding of this natural furanone to the *Escherichia coli* LuxS protein has been reported (Ren *et al.*, 2004). Hence, it can be hypothesized that these furanones react with certain reactive groups which are present in different regulatory proteins. It therefore seems that the macro-alga *Delisea pulchra* has evolved a sophisticated quorum sensing disruption-based defence system, producing compounds that affect different types of signal regulators in a non-growth inhibitory way. In view of the broad range of bacteria that can potentially colonize marine organisms, the production of broad spectrum quorum sensing-disrupting compounds by the alga probably confers a strong evolutionary advantage.

In conclusion, the results presented in this article show that the natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone disrupts quorum sensing-regulated gene expression in *V. harveyi* by decreasing the DNA-binding activity of the quorum sensing master regulator protein LuxR<sub>Vh</sub>. These results are of significant practical interest because they suggest that the furanone could be used as a broad spectrum biocontrol agent to protect a variety of hosts from different pathogenic vibrios in which virulence factor production is regulated by a LuxR<sub>Vh</sub> homologue.

## Experimental procedures

### Furanone preparation

The natural furanone (5*Z*)-4-bromo-5-(bromomethylene)-3-butyl-2(5*H*)-furanone was synthesized as described previously (Ren *et al.*, 2001). The furanone was dissolved and diluted in pure ethanol and stored at –20°C.

### Bacterial strains and growth conditions

The strains that were used in this study are shown in Table 1. All strains were grown in Luria–Bertani medium containing 20 g l<sup>-1</sup> NaCl (LB<sub>20</sub>) at 28°C under constant agitation. Spectrophotometry at OD<sub>600</sub> was used to measure growth. Luminescence was measured with a Lumac Biocounter M2500 luminometer (Lumac b.v., Landgraaf, the Netherlands).

For all experiments, *V. harveyi* strains were grown to an OD<sub>600</sub> of approximately 0.75, after which the furanone was added to the cultures in the appropriate concentration and the cultures were further incubated at 28°C with shaking. Unless otherwise indicated, samples were taken in triplicate 0.5 h after furanone addition.

**Table 1.** Strains used in this study.

Strain	Phenotype	Reference
<i>Vibrio harveyi</i> BB120	Wild type from which strains BNL258, JAF375, JAF483, JAF553, JMH597 and JMH612 are derived	Bassler <i>et al.</i> (1997)
<i>Vibrio harveyi</i> BNL258	<i>hfq::Tn5lacZ</i>	Lenz <i>et al.</i> (2004)
<i>Vibrio harveyi</i> JAF375	<i>luxN::Cm<sup>r</sup> luxQ::Kan<sup>r</sup></i>	Freeman and Bassler (1999a)
<i>Vibrio harveyi</i> JAF483	<i>luxO D47A</i> linked to Kan <sup>r</sup>	Freeman and Bassler (1999a)
<i>Vibrio harveyi</i> JAF553	<i>luxU H58A</i> linked to Kan <sup>r</sup>	Freeman and Bassler (1999b)
<i>Vibrio harveyi</i> JMH597	<i>luxN::Tn5 cqsS::Cm<sup>r</sup></i>	Henke and Bassler (2004a)
<i>Vibrio harveyi</i> JMH612	<i>luxPQ::Tn5 cqsS::Cm<sup>r</sup></i>	Henke and Bassler (2004a)
<i>Vibrio harveyi</i> MR1130	Isogenic <i>LuxR</i> mutant	Martin <i>et al.</i> (1989)

### Protein assays

Wild-type *V. harveyi* BB120 was used in the luciferase and bacterial alkaline phosphatase assays. Immediately before and 0.5, 1 and 2 h after the addition of the furanone, 2 units [OD<sub>600</sub> × vol (ml)] were pelleted, sonicated in 0.5 ml of 0.25 M Tris, pH 8 and the cellular debris removed (as described by Miyamoto *et al.*, 1990). A modified Lowry assay was used to determine the protein content (Markwell *et al.*, 1981). *In vitro* luciferase and bacterial alkaline phosphatase activities were determined as described by Gunsalus-Miguel and colleagues (1972) and Garen and Levinthal (1960).

### Primer design

Specific primers for the amplification of *luxR<sub>Vh</sub>* and RNA polymerase A subunit (*rpoA*) mRNA were designed using the Primer Express 2.0 software (Applied Biosystems, Foster City, USA). The primers were designed based on the consensus of the sequences that have been deposited before in GenBank. The combinations of the two primer sequences were blasted against GenBank. The primer sequences are shown in Table 2.

### RNA extraction and reverse transcriptase real-time PCR

The effect of the furanone on *luxR<sub>Vh</sub>* mRNA concentrations was studied in wild-type *V. harveyi* BB120. Immediately before and 0.5, 1 and 2 h after furanone addition, luminescence and cell density (OD<sub>600</sub>) of the cultures were measured and 0.5 ml of samples for RNA extraction were taken, which were immediately frozen in cold ethanol (−80°C). RNA was isolated using the Qiagen RNeasy Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. RNA extracts were treated with RNase-free DNase I (Fermentas, St. Leon-Rot, Germany), after which the RNA quantity was checked spectrophotometrically. RNA concentrations obtained were all around 300 ng μl<sup>-1</sup>. RNA quality was confirmed by electrophoresis. cDNA was obtained by reverse

**Table 2.** Primers used in this study.

Target	Primer	Sequence
<i>rpoA</i>	<i>rpoA</i> <sub>forward</sub>	5'-CGTAGCTGAAGCAAGATGA-3'
	<i>rpoA</i> <sub>reverse</sub>	5'-AAGCTGGAACATAACCACGA-3'
<i>luxR<sub>Vh</sub></i>	<i>LuxR</i> <sub>forward</sub>	5'-TCAATTGCAAAGAGACCTCG-3'
	<i>LuxR</i> <sub>reverse</sub>	5'-AGCAAACACTTCAAGAGCGA-3'

transcription using the Qiagen One Step RT-PCR kit (Qiagen), according to the manufacturer's instructions.

Real-time PCR was performed with the specific *luxR<sub>Vh</sub>* and *rpoA* primers. Amplicon lengths are 84 bp and 197 bp for *luxR<sub>Vh</sub>* and *rpoA* respectively. Amplification was performed in 25 μl reaction mixtures using the SYBR Green PCR Master Mix kit (Applied Biosystems, Nieuwerkerk a/d IJssel, the Netherlands), according to the manufacturer's instructions, in optical 96-well reaction plates with optical caps (Applied Biosystems). The thermal profile was as follows: 95°C for 2 min and 95°C for 10 min followed by 40 cycles of 95°C for 30 s, 54°C for 1 min, and 60°C for 1 min. Amplicon dissociation curves were determined by constant fluorescent measurement during a final heating step at 60°C to 95°C at 0.1°C s<sup>-1</sup> ramping speed. The melting temperatures for the amplicons were 77°C and 82°C for *luxR<sub>Vh</sub>* and *rpoA* respectively. The template DNA in the reaction mixtures was amplified and monitored with an ABI Prism SDS 7000 instrument (Applied Biosystems). The difference in mRNA levels between furanone-treated and untreated extracts was calculated as follows:

$$\log(\text{difference in concentration}) = \Delta C_t / \text{regression coefficient}$$

with  $\Delta C_t$  the difference in  $C_t$  value between furanone-treated and untreated extracts. The regression coefficients between  $\log(\text{concentration})$  and  $C_t$  value were determined by analysing a 10-fold dilution series of a *V. harveyi* BB120 DNA extract and were −3.380 ( $R^2 = 0.998$ ) and −3.597 ( $R^2 = 0.993$ ) for *luxR<sub>Vh</sub>* and *rpoA* respectively.

### Mobility shift assays

The effect of the furanone on *LuxR<sub>Vh</sub>* protein concentrations was studied in wild-type *V. harveyi* BB120. Immediately before and 0.5, 1 and 2 h after the addition of the furanone, 2 units [OD<sub>600</sub> × vol (ml)] were pelleted, sonicated in 0.5 ml of 0.25 M Tris, pH 8 and the cellular debris removed (as described by Miyamoto *et al.*, 1990). In the experiment that aimed at testing whether the furanone affected translation or acted on pre-existing translates, cultures were treated with 50 mg l<sup>-1</sup> chloramphenicol Sigma-Aldrich (Bornem, Belgium) before furanone addition. Mobility shift assays were conducted as described previously (Swartzman and Meighen, 1993) using radiolabelled *luxR<sub>Vh</sub>* promoter DNA (Chatterjee *et al.*, 1996) or *luxCDABEGH* promoter DNA (Miyamoto *et al.*, 1996) containing the *LuxR<sub>Vh</sub>* binding sites. The amount of retarded DNA was quantified as described in Miyamoto and colleagues (1996) using a Fuji bioimage.



*Experiments with purified LuxR<sub>Vh</sub>*

LuxR<sub>Vh</sub> purification was carried out as described previously (Swartzman and Meighen, 1993). The purified LuxR<sub>Vh</sub> was stored in 50 mM NaPO<sub>4</sub>, 300 mM NaCl, pH 8.0. The protein concentration (as determined by Bio-Rad protein assays) was 0.2 mg ml<sup>-1</sup>. To glass tubes containing 40 µl of the protein preparation, 0.2 µl of absolute ethanol or 0.2 µl of furanone (10 mg ml<sup>-1</sup> in ethanol) were directly added. The tubes were incubated in a 37°C water bath for 1 h and then stored at 4°C. For mobility shift analyses, the samples were diluted 10-fold in storage buffer and 1 µl (0.02 µg) was assayed. For SDS-PAGE, 1 µg of protein was applied and 10% SDS-PAGE was performed according to Maniatis and colleagues (1982).

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**References**

- Austin, B., and Zhang, X.-H. (2006) *Vibrio harveyi*: a significant pathogen of marine vertebrates and invertebrates. *Lett Appl Microbiol* **43**: 119–124.
- Bassler, B.L., Wright, M., Showalter, R.E., and Silverman, M.R. (1993) Intercellular signaling in *Vibrio harveyi*-sequence and function of genes regulating expression of luminescence. *Mol Microbiol* **9**: 773–786.
- Bassler, B.L., Greenberg, E.P., and Stevens, A.M. (1997) Cross-species induction of luminescence in the quorum-sensing bacterium *Vibrio harveyi*. *J Bacteriol* **179**: 4043–4045.
- Cao, J.G., and Meighen, E.A. (1989) Purification and structural identification of an autoinducer for the luminescence system of *Vibrio harveyi*. *J Biol Chem* **264**: 21670–21676.
- Chatterjee, J., Miyamoto, C.M., and Meighen, E.A. (1996) Autoregulation of *luxR*: the *Vibrio harveyi lux*-operon activator functions as a repressor. *Mol Microbiol* **20**: 415–425.
- Chen, X., Schauder, S., Potier, N., Van Dorsselaer, A., Pelczar, I., Bassler, B.L., and Hughson, F.M. (2002) Structural identification of a bacterial quorum-sensing signal containing boron. *Nature* **415**: 545–549.
- Defoirdt, T., Boon, N., Bossier, P., and Verstraete, W. (2004) Disruption of bacterial quorum sensing: an unexplored strategy to fight infections in aquaculture. *Aquaculture* **240**: 69–88.
- Defoirdt, T., Bossier, P., Sorgeloos, P., and Verstraete, W. (2005) The impact of mutations in the quorum sensing systems of *Aeromonas hydrophila*, *Vibrio anguillarum* and *Vibrio harveyi* on their virulence towards gnotobiotically cultured *Artemia franciscana*. *Environ Microbiol* **7**: 1239–1247.
- Defoirdt, T., Crab, R., Wood, T.K., Sorgeloos, P., Verstraete, W., and Bossier, P. (2006) Quorum sensing-disrupting brominated furanones protect the gnotobiotic brine shrimp *Artemia franciscana* from pathogenic *Vibrio harveyi*, *Vibrio campbellii*, and *Vibrio parahaemolyticus* isolates. *Appl Environ Microbiol* **72**: 6419–6423.
- Freeman, J.A., and Bassler, B.L. (1999a) A genetic analysis of the function of LuxO, a two-component response regulator involved in quorum sensing in *Vibrio harveyi*. *Mol Microbiol* **31**: 665–677.
- Freeman, J.A., and Bassler, B.L. (1999b) Sequence and function of LuxU: a two-component phosphorelay protein that regulates quorum sensing in *Vibrio harveyi*. *J Bacteriol* **181**: 899–906.
- Garen, A., and Levinthal, C. (1960) A fine-structure genetic and chemical study of the enzyme alkaline phosphatase of *E. coli*. I. Purification and characterization of alkaline phosphatases. *Biochim Biophys Acta* **38**: 470–483.
- Givskov, M., de Nys, R., Manefield, M., Gram, L., Maximilien, R., Eberl, L., et al. (1996) Eukaryotic interference with homoserine lactone-mediated prokaryotic signalling. *J Bacteriol* **178**: 6618–6622.
- Gunsalus-Miguel, A., Meighen, E.A., Nicoli, M.Z., Nealson, K.H., and Hastings, J.W. (1972) Purification and properties of bacterial luciferases. *J Biol Chem* **247**: 398–404.
- Henke, J.M., and Bassler, B.L. (2004a) Three parallel quorum sensing systems regulate gene expression in *Vibrio harveyi*. *J Bacteriol* **186**: 6902–6914.
- Henke, J.M., and Bassler, B.L. (2004b) Quorum sensing regulates type III secretion in *Vibrio harveyi* and *Vibrio parahaemolyticus*. *J Bacteriol* **186**: 3794–3805.
- Hentzer, M., and Givskov, M. (2003) Pharmacological inhibition of quorum sensing for the treatment of chronic bacterial infections. *J Clin Invest* **112**: 1300–1307.
- Jobling, M.G., and Holmes, R.K. (1997) Characterization of *hapR*, a positive regulator of the *Vibrio cholerae* HA/protease gene *hap*, and its identification as a functional homologue of the *Vibrio harveyi luxR* gene. *Mol Microbiol* **26**: 1023–1034.
- Karunasagar, I., Pai, R., Malahti, G.R., and Karunasagar, I. (1994) Mass mortality of *Penaes monodon* larvae due to antibiotic-resistant *Vibrio harveyi* infection. *Aquaculture* **128**: 203–209.
- Koch, B., Liljefors, T., Persson, T., Nielsen, J., Kjelleberg, S., and Givskov, M. (2005) The LuxR receptor: the sites of interaction with quorum-sensing signals and inhibitors. *Microbiology* **151**: 3589–3602.
- Lenz, D.H., Mok, K.C., Lilley, B.N., Kulkarni, R.V., Wingreen, N.S., and Bassler, B.L. (2004) The small RNA chaperone Hfq and multiple small RNAs control quorum sensing in *Vibrio harveyi* and *Vibrio cholerae*. *Cell* **118**: 69–82.
- Lilley, B.N., and Bassler, B.L. (2000) Regulation of quorum sensing in *Vibrio harveyi* by LuxO and  $\delta 54$ . *Mol Microbiol* **36**: 940–954.
- McCarter, L.L. (1998) OpaR, a homolog of *Vibrio harveyi* LuxR, controls opacity of *Vibrio parahaemolyticus*. *J Bacteriol* **180**: 3166–3173.
- McDougald, D., Rice, S.A., and Kjelleberg, S. (2000) The marine pathogen *Vibrio vulnificus* encodes a putative homologue of the *Vibrio harveyi* regulatory gene *luxR*: a genetic and phylogenetic comparison. *Gene* **248**: 213–221.

- McDougald, D., Srinivasan, S., Rice, S.A., and Kjelleberg, S. (2003) Signal-mediated cross-talk regulates stress adaptation in *Vibrio* species. *Microbiology* **149**: 1923–1933.
- Manefield, M., Harris, L., Rice, S.A., de Nys, R., and Kjelleberg, S. (2000) Inhibition of luminescence and virulence in the black tiger prawn (*Penaeus monodon*) pathogen *Vibrio harveyi* by intercellular signal antagonists. *Appl Environ Microbiol* **66**: 2079–2084.
- Manefield, M., Rasmussen, T.B., Hentzer, M., Andersen, J.B., Steinberg, P., Kjelleberg, S., and Givskov, S. (2002) Halogenated furanones inhibit quorum sensing through accelerated LuxR turnover. *Microbiology* **148**: 1119–1127.
- Maniatis, T., Fritsch, E.F., and Sambrook, J. (1982) *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, NY, USA: Cold Spring Harbor Laboratory.
- Markwell, M.A.K., Haas, S.M., Tolbert, N.E., and Bieber, L.L. (1981) Protein determination in membrane and lipoprotein samples: manual and automated procedures. *Methods Enzymol* **72**: 296–303.
- Martin, M.O., Showalter, R.E., and Silverman, M. (1989) Identification of a locus controlling expression of luminescence genes in *Vibrio harveyi*. *J Bacteriol* **171**: 2406–2414.
- Milton, D.L. (2006) Quorum sensing in vibrios: complexity for diversification. *Int J Med Microbiol* **296**: 61–71.
- Miyamoto, C.M., Meighen, E.A., and Graham, A.F. (1990) Transcriptional regulation of *lux* genes transferred into *Vibrio harveyi*. *J Bacteriol* **172**: 2046–2054.
- Miyamoto, C.M., Chatterjee, J., Swartzman, E., Szittner, R., and Meighen, E.A. (1996) The role of the *lux* autoinducer in regulating luminescence in *Vibrio harveyi*; control of *luxR* expression. *Mol Microbiol* **19**: 767–775.
- Moriarty, D.J.W. (1998) Disease control in shrimp aquaculture with probiotic bacteria. In *Microbial Biosystems: New Frontiers. Proceedings of the Eighth International Symposium on Microbial Ecology*. Bell. C.R., Brylinsky, M. and Johnson-Green, P. (eds). Halifax, NS, Canada: Atlantic Canada Society for Microbial Ecology.
- Ramos, J.L., Martínez-Bueno, M., Molina-Henares, A.J., Terán, W., Watanabe, K., Zhang, X., et al. (2005) The TetR family of transcriptional repressors. *Microbiol Mol Biol Rev* **69**: 326–356.
- Rasmussen, T.B., and Givskov, M. (2006) Quorum-sensing inhibitors as anti-pathogenic drugs. *Int J Med Microbiol* **296**: 149–161.
- Ren, D., Sims, J., and Wood, T.K. (2001) Inhibition of biofilm formation and swarming of *Escherichia coli* by (5Z)-4-bromo-5-(bromomethylene)-3-butyl-2(5H)-furanone. *Environ Microbiol* **3**: 731–736.
- Ren, D., Bedzyk, L., Ye, R.W., Thomas, S.M., and Wood, T.K. (2004) Differential gene expression shows natural brominated furanones interfere with the autoinducer-2 bacterial signaling system of *Escherichia coli*. *Biotechnol Bioeng* **88**: 630–642.
- Swartzman, E., and Meighen, E.A. (1993) Purification and characterization of a poly-(dA-dT) *lux*-specific DNA binding protein from *Vibrio harveyi* and identification as LuxR. *J Biol Chem* **268**: 16706–16716.
- Swartzman, E., Silverman, M., and Meighen, E.A. (1992) The *luxR* gene product of *Vibrio harveyi* is a transcriptional activator of the *lux* promoter. *J Bacteriol* **174**: 7490–7493.
- Thompson, F.L., Iida, T., and Swings, J. (2004) Biodiversity of vibrios. *Microbiol Mol Biol Rev* **68**: 403–431.
- Tinh, N.T.N., Linh, N.D., Wood, T.K., Dierckens, K., Sorgeloos, P., and Bossier, P. (2007) Interference with the quorum sensing systems in a *Vibrio harveyi* strain alters the growth rate of gnotobiotically cultured rotifer *Brachionus plicatilis*. *J Appl Microbiol* (in press).