

The Network of Tumor Microtubes: An Improperly Reactivated Neural Cell Network With Stemness Feature for Resistance and Recurrence in Gliomas

Xinyue Wang^{1,2}, Jianhao Liang² and Haitao Sun^{1,2,3*}

¹ Clinical Biobank Center, Microbiome Medicine Center, Department of Laboratory Medicine, Zhujiang Hospital, Southern Medical University, Guangzhou, China, ² Neurosurgery Center, The National Key Clinical Specialty, The Engineering Technology Research Center of Education Ministry of China on Diagnosis and Treatment of Cerebrovascular Disease, Guangdong Provincial Key Laboratory on Brain Function Repair and Regeneration, The Neurosurgery Institute of Guangdong Province, Zhujiang Hospital, Southern Medical University, Guangzhou, China, ³ Key Laboratory of Mental Health of the Ministry of Education, Guangdong–Hong Kong–Macao Greater Bay Area Center for Brain Science and Brain–Inspired Intelligence, Southern Medical University, Guangzhou, China

OPEN ACCESS

Edited by:

Yunqing Li, Kennedy Krieger Institute, United States

Reviewed by:

Frank Winkler, Heidelberg University, Germany Luciana N. S. Andrade, University of São Paulo, Brazil

*Correspondence:

Haitao Sun 2009sht@smu.edu.cn; msunhaitao1988@126.com

Specialty section:

This article was submitted to Neuro-Oncology and Neurosurgical Oncology, a section of the journal Frontiers in Oncology

Received: 17 April 2022 Accepted: 18 May 2022 Published: 29 June 2022

Citation:

Wang X, Liang J and Sun H (2022) The Network of Tumor Microtubes: An Improperly Reactivated Neural Cell Network With Stemness Feature for Resistance and Recurrence in Gliomas. Front. Oncol. 12:921975. doi: 10.3389/fonc.2022.921975 Gliomas are known as an incurable brain tumor for the poor prognosis and robust recurrence. In recent years, a cellular subpopulation with tumor microtubes (TMs) was identified in brain tumors, which may provide a new angle to explain the invasion, resistance, recurrence, and heterogeneity of gliomas. Recently, it was demonstrated that the cell subpopulation also expresses neural stem cell markers and shares a lot of features with both immature neurons and cancer stem cells and may be seen as an improperly reactivated neural cell network with a stemness feature at later time points of life. TMs may also provide a new angle to understand the resistance and recurrence mechanisms of glioma stem cells. In this review, we innovatively focus on the common features between TMs and sprouting axons in morphology, formation, and function. Additionally, we summarized the recent progress in the resistance and recurrence mechanisms of gliomas with TMs and explained the incurability and heterogeneity in gliomas with TMs. Moreover, we discussed the recently discovered overlap between cancer stem cells and TM-positive glioma cells, which may contribute to the understanding of resistant glioma cell subpopulation and the exploration of the new potential therapeutic target for gliomas.

Keywords: glioma, tumor microtubes, tunneling nanotubes, cancer stem cells, resistance, heterogeneity, invasion, brain tumor microenvironment

1

Abbreviations: TM, tumor microtube; NSC, neural stem cell; CNS, central nervous system; Gap43, growth-associated protein 43; Cx43, connexin 43; ICWs, intercellular calcium waves; SVZ, subventricular zone; NPCs, neural progenitor cells; BTSCs, brain tumor stem-like cells; TMZ, temozolomide.

1 INTRODUCTION

Gliomas represented approximately 25% of primary CNS (central nervous system) tumors and 80% of malignant tumors (1), characterized by poor prognosis with a median overall survival of approximately 15 months (2). Gliomas share a lot of morphological features with glial cells in the normal brain, such as astrocytes, oligodendrocytes, and ependymal cells, consequently classified morphologically (3). Of note, there is an increasing recognition of the important role of molecular characteristics in the classification of gliomas, such as the mutation of Isocitrate dehydrogenase (IDH) and 1p/19q codeletion, for the distinctive clinical manifestations of heterogeneous cell subpopulations (3). Gliomas are still known as an incurable brain tumor for the poor prognosis and robust recurrence (3). Increasing attention has been paid to the mechanism related to their widespread infiltration and robust resistance and self-repairment, particularly the heterogeneity of the resistant cellular subpopulation (4). For example, a cellular subpopulation of glioma cells with NSC (neural stem cell) marker expression, usually forming tumor microtubes (TMs), was identified (5-7), which may be related to the high resistance of gliomas to all existing standard therapies and recurrence (8-11). Furthermore, the TM-positive cell subpopulation may provide a new angle on the explanation of the mechanisms and the heterogeneity of glioma cells (12).

Tumor microtubes are ultralong membrane protrusions extended by glioma cells, which are demonstrated to facilitate glioma cell widespread infiltration and treatment resistance (12, 13). Two morphologically, molecularly, and functionally different TM subtypes have been identified: non-connecting ones, which are crucial for glioma invasion and proliferation, and interconnecting ones, which build Cx43-separated membrane tube connections between individual glioma cells (14). Ramírez-Weber and his colleagues first detected membrane tubes for intercellular connections in Drosophila development (15). Subsequently, accumulating cell types have been identified to form and utilize membrane tubes to exchange various cellular components, which are exemplified by organelles (16), pathogens including HIV (17-19) and prions (20), and genetic material (12, 21). The membrane tubes have gained various names: membrane nanotubes, tunneling nanotubes, and cytonemes (12). For the distinctive characteristics of the membrane tubes in gliomas, they were specially termed "tumor microtubes" (TMs) (12).

2 TMS AND TM-POSITIVE CELLS SHARE A LOT OF FEATURES WITH SPROUTING AXONS AND IMMATURE NEURONS

Recently, it was demonstrated that the dysregulated and disordered glioma progression and malignancy are, in essence, parallel to the directed and ordered CNS development and function based on the morphological characteristics, mechanism of cellular proliferation, migration, and communication (22, 23). In line with this, TMs and

the TM-positive glioma cell network share a lot of features with sprouting axons and immature neuroblasts (**Table 1**).

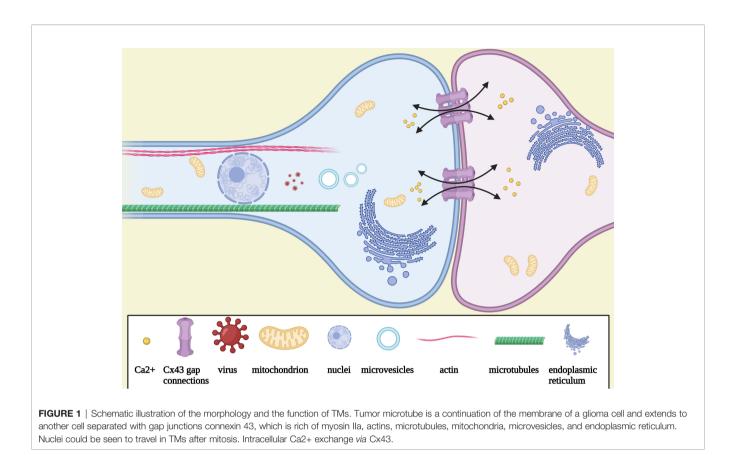
2.1 Morphology and Formation

Different TM subtypes are morphologically and molecularly heterogeneous. A non-connecting tumor microtube is an ultralong membrane protrusion extended by a glioma cell, while an interconnecting tumor microtube is a continuation of the membrane of a glioma cell and extends to another cell separated with gap junctions (Figure 1) (12, 14). TMs are 1.7 µm in width on average, and the maximum length reaches more than 500 μ m, while tunneling nanotubes are less than 1 μ m in width and 30 µm in length on average; the life span of a tumor microtube reaches more than 200 days, while that of tunneling nanotubes is up to 60 min (Table 1) (24). It can also be seen in vivo that the leading edges of invasive TMs are morphologically parallel with the axonal growth cones of normal sprouting axons (12, 25). Together, these findings suggest morphological parallels between invasive TMs and normal sprouting axons. Additionally, immunohistochemistry showed that the tumor microtubes were rich of myosin IIa, actins, and microtubules, which play an important role in the generation of contractile forces necessary for the movement of glioma cells (12). Likewise, myosin, actin, and microtubules are also known to be rich in the protrusions of neural precursor cells for migration (26). Furthermore, three key molecular players of TMs have been identified: Ttyh1 (14), Gap43 (growth-associated protein 43)

TABLE 1 | Characteristics of TMs in comparison with human sprouting neuron axons.

Feature	TMs	Axons
Width	Mean, 1.7 µm	0.08–0.4 μm
Length	Maximum,>500 µm	Maximum, 20 μm Minimum,<1 mm Maximum.>1 m
Lifetime	Days, up to 200	years
Content		,
Actin	+	+
Mitochondria	+	+
Protein	+	+
Endoplasmic reticulum	+	+
Microvesicles	+	+
Microtubules	+	+
Myosin X	-	+
Myosin IIa	+	-
β III tubulin	-	+
Voltage-gated Ca ²⁺ channel	+	+
Ttyh1	+	+
Gap43	+	+
Cx43	+	+
Functions		
Nucleus transmission	+	?
cell migration	+	?
Mitochondria transmission	+	+
Protein transmission	+	+
Propagation of ICWs	+	+
Pathogen spread	+	+
Microvesicle transmission	+	+

+: positive; -: negative; ?: uncertain.



(12), and Connexin 43 (12). The gap junction protein connexin 43 is highly expressed at TMs integrated into the network (12). Cx43 immunoreactivity was most frequently only expressed at one end of a TM connecting two cells, suggesting that the one end of a TM is continuous with the cell membrane; the other end is a membrane boundary separated with a gap junction (12). Since the presence of Cx43 is usually accompanied with poor prognosis while the absence of Cx43 is accompanied with a reduced tumor size and improved survival, it was once viewed as an important molecular driver of tumor microtubes. However, some controversial views suggest that Cx43 did not always promote the growth and function of tumor microtubes (27). Recently, it was demonstrated that Cx43 may play an important role in the communication and maintain the integrity of the network, which is exemplified by empowering gliomas to acquire resistance to oxidative stress (28). Gap43 was demonstrated to be a key molecular player in driving TM outgrowth, which is highly expressed in the growth of cone-like tips of the TMs, driving the growth of TMs and TM-dependent astrocytoma cell migrations (12, 24). The knockdown of GAP-43 in mouse brain blocked both TM-non-connected glioma cell invasion and proliferation and intercellular TM connection (12). According to the recent research in mouse models, Ttyh1 also plays a key role in the growth of invasive non-connecting TMs, while the knockdown of Ttyh1 contributed to a dramatically declined proportion of invasive TM-non-connected glioma cells (14). Of note, TMconnected glioma cells appear to be uncompromised by

interference with Ttyh1; it was shown that TM-connected glioma cells were not affected in the absence of Ttyh1 (14). Together, these findings suggest that the two TM subtypes are molecularly and functionally heterogeneous. Similarly, Ttyh1 and Gap43 are also highly concentrated in axonal growth cones during neurite outgrowth, driving developmental and regenerative axon growth (29-31) and neuronal progenitor cell migrations (32). The overexpression of Ttyh1 and Gap43 appears to induce neurite outgrowth not only in neuronal (31, 33) but also in non-neuronal cells (34), while the downregulation of expression of them inhibit the neurite outgrowth (35). Furthermore, a recent study in mouse brain showed that intercellular adhesion and signaling provided by p120-catenindependent adherens junctions is crucial for both TM-nonconnected glioma cell invasion and the TM-connected network (36), which may be highly reminiscent of epithelial tumors regulated by p120 signaling for anchorage-independent growth, anoikis, resistance, and metastasis (37-39).

2.2 Function

2.2.1 Communication Network

Interconnecting TMs are shown to be involved in the molecular, organelle, and vesicle transport and intercellular calcium wave (ICW) propagation (12). Cells can be electrically coupled over long distances *via* membrane tubes associated with gab junctions (12). Thus, interconnecting TMs can mediate the bidirectional spread of ICWs between connecting cells through gap junctions

(12). ICWs play an important role in the communication of glioma cells with each other and in the coordination in the multicellular network (40, 41). Spontaneous widespread ICWs are rare to be seen in mature brains under physiological conditions; however, it can be observed in the communication of neural stem and progenitor cells to coordinate the proliferation and differentiation (42, 43).

2.2.2 Cell Migration

Non-connecting tumor microtubes can also promote the invasion and proliferation of the glioma cells. Immunohistochemistry showed that the tumor microtubes were rich of actins, myosin IIa, and microtubules, and three-dimension scanning electron microscopy (3D SEM) showed that mitochondria and microvesicles traveled quickly and frequently in the tubes, implying active movement, ATP production, and vesicle trafficking (12, 44). It has been shown that the nucleus is able to travel in cellular membrane tubes after mitosis, which may be concerned with the invasion, treatment resistance, and cellular selfrepairment and regeneration of glioma cells (12). Notably, it has been demonstrated that the synaptic and electrical integration described above also drives a calcium-dependent activation of glioma cell invasion (45) and promotes glioma proliferation (46).

According to the "seed and soil" hypothesis, it is not tumor cells that determine where to metastasize (47, 48). In contrast, the premetastatic niche is a prerequisite for the subsequent metastases of tumor cells (47, 48), supporting the fact that metastasis occurs only in selected organs but not in other organs, although tumor cells reach the vascular system of all organs (49). Residing in a specific niche is necessary for the survival and metastases of specific cells (50). The neurogenic niche in the subventricular zone (SVZ) of human lateral ventricles (51) is composed of neural stem cells (NSCs) and progenitor cells (NPCs), ependymal cells, astrocytes, microglia, macrophages, neurons, and extracellular matrix and associated vessels (52). The neurogenic niche is known to play an important role in the sustainment of the stem cell properties, including cell proliferation and self-renewal (53, 54). Similarly, in the perivascular niches in brain tumors, a cellular subpopulation of brain tumor stem-like cells (BTSCs) with NSC marker expression, including nestin and CD133, has also been identified (22). The perivascular niches were demonstrated to be related to the proliferation, self-renewal, invasion, and stemness of the tumor cells (55-57). Consequently, it is critical for tumor cells to be localized in the perivascular niches. It was seen that glioma stem-like cells extended TMs to move to the perivascular niche (12). A similar migration can be observed in the self-repairment of regeneration-damaged tumor cells (12). Likewise, it has been demonstrated that gliomas travel the same extracellular routes with migrant neural stem cells and neural progenitor cells during normal CNS development and damage repair (58-60). Above all, glioma cell migration induced by the growth of TMs is necessary for the invasion, malignancy progression, and recurrence of gliomas.

Normal neurodevelopment depends on the regulation of intracellular mechanisms, interactions with the microenvironment, and signaling pathways (22). Studies on TMs support the hypothesis

that glioma progression may be seen as neurodevelopment with the loss of regulation and improperly reactivated, exploiting developmental pathways and molecules for TM formation and cell invasion (14, 22). The growth of TMs and the formation of a TMconnected network just like neurodevelopment are improperly reactivated at later time points of life.

2.2.3 "Dendritic" Function in Electrical and Synaptic Integration of Glioma Into Neural Circuits

The newly discovered glutamatergic synaptic input observed in mouse models and resected patient tumor material with glutamate receptors of the AMPA subtype are usually located on tumor microtubes (45, 46), and activity-dependent, nonsynaptic potassium currents (46) can activate intercellular calcium currents in the glioma cell network to drive a calciumdependent activation of glioma cell invasion and promote glioma proliferation, which indicate that TMs may have a "dendritic" function for cancer cells. These findings are highly reminiscent of stem-cell populations regulated by a glutamatergic synaptic input, such as neuronal (61) and oligodendrocyte precursor cells (62) during normal neurodevelopment and function.

3 THE ROLE OF TMS IN NEW EXPLANATION FOR THE MECHANISMS OF RESISTANCE AND RECURRENCE OF GLIOMAS

It has been shown that TMs can mediate depolarization signals when subjected to stimulation such as radiotherapy and chemotherapy (12, 13). Apparently, even a slight fluctuation in intracellular calcium levels can cause great damage to intracellular homeostasis and impair cells and contribute to apoptotic cell death in glioma cells (63). It has been demonstrated that the synchronicity of the calcium peak of TM-connected cells is better than that of non-connected cells. implying that interconnecting tumor microtubes contribute to redistribute intracellular calcium to keep it at a nonlethal level and maintain intracellular homeostasis via membrane tubes connecting two cells and their forming networks to withstand adverse events (12). Intracellular calcium levels in cells without radiotherapy and TM-connected cells with radiotherapy were very homogeneous, while unconnected cells developed a high variability of calcium levels with radiotherapy (12). It has been shown that after radiotherapy, the vast majority of TMconnected glioma cells were protected from cell death, while most of the TM-unconnected and TM-negative cells died (12, 14). Moreover, glioblastoma cells may hijack neighboring nonmalignant astrocytes to transfer cGAMP via gap junctions as a result that activate the cGAS-STING pathway and release cytokines including IFN α and TNF to promote tumor metastasis (64) as it was demonstrated that melanoma cells can connect to active astrocytes via gap junctions to resist chemotherapy (65).

However, when the impairment is beyond their capacity of resistance, what would they do? It has been shown that the death of the TM-connected tumor cell network resulted in a rapid extension of the TMs of neighboring glioma cells into the lesion region, new TMs were extended toward the dead cells, and within a few days, new nuclei were transmitted through the tumor microtubes to the cells to facilitate the process of selfrepairment (13). The density of tumor cells in that region increased significantly and gradually even exceed those of unlesioned brain regions over time to improve the resistance of damaged tumor cells and maintain the integrity of the tumor cell network (13) (Figure 2). In contrast, non-TM-connected glioma cells are infrequently expected to observe the self-repairing mechanism. It has been demonstrated that the number of TMs of astrogliomas is usually more than those of oligodendrocytes, while the length of the TMs of astrogliomas is also frequently longer than those of oligodendrocytes (12). Simultaneously, the number and the length of TMs appear to have a positive correlation of the grade and poor prognosis (12). According to the studies of Venkataramani et al. (45) and Venkatesh et al. (46), the electrical and synaptic integration of the TM-connected glioma cell network into neural circuits may dramatically promote glioma cell proliferation.

4 TMS PROVIDES A NEW ANGLE TO UNDERSTAND THE INCURABILITY AND HETEROGENEITY IN GLIOMAS

Malignant gliomas are still regarded as a type of incurable tumor. Current clinical standard treatments for gliomas include radiotherapy, chemotherapy particularly with temozolomide (TMZ) (66, 67), and surgical resection (68). However, as a result of the widespread dissemination and robust recurrence of gliomas, the existing therapies are all limited.

After surgical resection, it has been shown that the vast majority of glioma cells tend to recur at or around the resection margin and even excessively proliferate (69), which means that, in fact, the resection of gliomas and the woundhealing response of a normal brain appear to promote the recurrence of gliomas. The phenomenon has also been observed in other tumors and has been attributed to the effect of wound-healing factors (70, 71) and growth factors (72). TMs may promote a new angle of explanation to the characteristic. S. Weil et al. proved that new TMs will be extended toward the margin of the resected region to promote the process of selfrepairment after resection (13). Thus, more and more glioma cells recur, neighboring the margin and eventually exceeding the tumor cell density before the resected and that of the unresected region (13). Similarly, the similar self-repairment mechanism also applies to the reaction of gliomas after radiotherapy (12).

Additionally, although radiochemotherapy with temozolomide (TMZ) is still a mainstream therapy for gliomas, the poor diagnosis is attributed to the innate and acquired drug resistance of gliomas (68). O6-methylguanine-DNA methyltransferase (MGMT) protein, which is effective to repair DNA damage and consequently avoid cell apoptosis, is a major known mechanism for TMZ resistance (73). Clinical studies have shown that MGMT promoter hypermethylation in approximately half of gliomas appears to predict a better treatment response to TMZ (74). However, the nonresponders indicate that the novel mechanisms of the drug resistance should be explored further. It has also been demonstrated that gliomas with TMs connected are more resident to TMZ than those without TMs and also appear to excessively

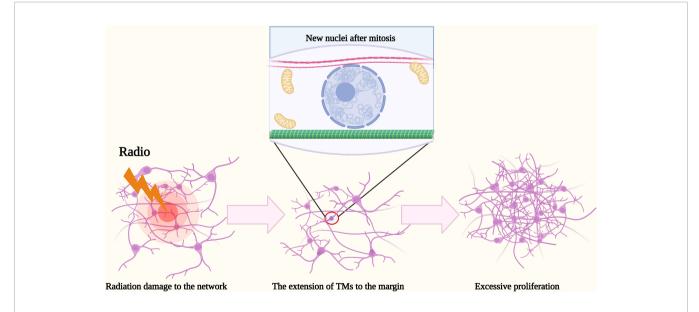


FIGURE 2 | Schematic illustration of the excessive proliferation of glioma cells after radiation. After radiation damage to the network, the death of TM-connected tumor cell network resulted in a rapid extension of TMs of neighboring glioma cells into the margin. Within a few days, new nuclei were transmitted there through the TMs to facilitate the recurrence of glioma cells. The density of tumor cells in that region increased significantly and gradually even exceed those of unlesioned brain regions over time.

proliferate (13), which indicates an important role played in the TMZ resistance by the TM network. It may be attributed to the gap junction connections *via* Cx43 in glioma cells, which has been demonstrated to have a marked negative correlation of the TMZ resistance that may, in part, be attributed to mitochondrial apoptosis (75–77). Consequently, TM may be a potential therapeutic target for the TMZ resistance of gliomas. Recent research reveals that meclofenamate as a connexin43 blocker can impair the integrity of the TM network and increase the TMZ sensitivity of TMZ-resistant glioma cells (78).

Based on the discussion above, the number and length of TMs are highly influenced by the tumor type and grade, with a marked positive correlation of treatment resistance and poor prognosis (12). The explanation may be applied to the 1p/19q co-deleted gliomas.

Gliomas with 1p/19q co-deletion frequently predict a favorable diagnosis and a high responsiveness to various therapies, such as radiotherapy And chemotherapy, which is a characteristic for oligodendrogliomas but absent in astrocytomas (79). However, the potential molecular mechanisms remain indistinct. The function of the tumor microtubes and their forming networks may provide an explanation of this phenomenon. It has been demonstrated that TMs in 1p/19q non-co-deleted gliomas are frequently more and longer than those in 1p/19q co-deleted ones and the RNA-Seq gene expression data revealed that the gap junction protein Cx43 and core pathways driving neurite formation and the growth of neurite-like TMs are highly expressed in 1p/19q non-co-deleted gliomas (12). Since the genes of neurotrophic factors such as NGF and NT-4 (also called NTF4) that play a crucial role in the expression of GAP-43 and TTYH1 are demonstrated to be located on both chromosomal parts 1p and 19q, 1p/19q codeleted gliomas may lack TMs due to downregulation of GAP-43 and their corresponding receptors (TrkA, TrkB), making them more susceptible to various therapies. As a consequence, the prognosis of oligodendroglioma is more favorable than that of astrocytoma (2, 66, 80-82).

5 TM-CONNECTED GLIOMA CELLS SHARE A LOT OF FEATURES WITH GLIOMA STEM-LIKE CELLS

There is a view pointing out that the tumor development is a more abnormal organ development than tumor cell clones, implying that the principles of normal stem cell biology may also be applied to tumor development and cancer stem cells (83). The first purification of cancer stem-like cells is a leukemiainitiating cell isolated by Dick and his colleagues (84). Subsequently, a subpopulation of BTSCs was identified and also becomes a major area of interest within the field of neurooncology (5, 6). The BTSCs share a lot of features with normal neural stem cells (NSCs), such as they both have the ability of self-renewal and differentiation (22). CD133 (57), OCT4, NANOG and SOX2 (85) are all involved in well-established stemness markers in gliomas. However, it has been demonstrated that cells expressing different CSCs markers in gliomas are not distinguished by distinct functional properties or transcriptomic profiles; the difference is more likely to be a result of intrinsic tumor plasticity induced by the microenvironment (86). In recent years, there is growing evidence that BTSCs may play an important role in the treatment resistance and robust recurrence in gliomas (8, 9, 22). The studies of TMs may provide a new angle to understand and explain the associated mechanism in BTSCs.

According to the research of Xie and his colleagues in mouse models and resected patient material, TM-connected glioma cells share features with BTSCs (87). Compared with the TM-negative subpopulation, the TM-connected subpopulation performs significantly more genetic enrichment associated with both the embryonic stem cell status and the cell cycle (87). Nestin is known to be one of the best-established markers of cell stemness (57) and treatment resistance in gliomas (88). Compared with the TMnegative subpopulation, the TM-connected subpopulation performs a significantly higher expression of nestin and other stemness markers such as Musashi and Sox2 (87). Additionally, mice receiving the reimplantation of TM-connected glioma cells is shown to suffer obvious tumorigenesis with poor survival, while tumorigenesis fails to be detected in mice receiving the reimplantation of TM-negative glioma cells (87). The reimplanted TM-connected subpopulation is also demonstrated to reconstitute the heterogeneity of the TM content and network integration (87). Together, these findings suggest that TMconnected glioma cells share two typical features with BTSCs: proliferative potential and give rise to heterogeneous combinations of cells with different phenotypes (83).

Furthermore, after radiotherapy, it is shown that only nestinpositive tumor cells collectively extended more TMs and survive while nestin-negative tumor cells tend to fail to respond in such way and get impaired. Eventually, nestin-positive TM networkintegrated glioma cells account for the vast majority of the tumor cells after radiotherapy. However, notably, both nestin-positive subpopulation with or without a TM response survived better, implying that the resistance mechanism induced by nestin is, in part, independent of that of TMs (87). According to the discussion above, the new findings may suggest a relationship between TM proficiency and cellular stemness.

It has been shown that reducing tumor bulk can induce the excessive proliferation of particularly quiescent BTSCs (89, 90). According to the discussion above, a similar mechanism has been detected in the self-repairment process induced by TMs (12, 13). Consequently, we speculate that responsive TMs might also be related to the tumor self-repairment induced by BTSCs for further research.

As a result that most standard therapies for glioma target proliferating cells, one of the crucial mechanisms of CSC treatment resistance is to switch into a quiescent state, particularly when the CSCs suffer from various standard therapies and a recent research showed that quiescent human glioma stem cells drove tumor initiation, infiltration, and recurrence following chemotherapy (91), which may be related to the dormancy of tumor cells in the perivascular niches (92). For example, it has been demonstrated that a cellular subpopulation of slow-cycling CSCs in GBM plays an important role in TMZ treatment resistance, and the resistance will be suppressed with the ablation of the slow-cycling CSCs (88). It is demonstrated that glioma cells may extend TMs to migrate to the specific perivascular niches with nuclei transport and contractile forces provided (12). It is the specific perivascular niche that determines the proliferating or quiescent state of tumor cells (47, 48). Consequently, we speculate that the dormancy process might be related to the rapid migration to the quiescent pool with TMs. Moreover, we speculate that other resistance mechanisms of TMs, such as ICW propagation, might also be applied to BTSC resistance for further research.

Although Xie and his colleagues have only demonstrated that TM-connected glioma cell networks are enriched for certain stem-like behaviors (87), it is highly likely that also invasive TM-positive but non-TM-connected glioma cells have stem-like properties for their many common features shared with neural progenitor and stem cells, which have been discussed above. Therefore, further research should be considered to unravel which stem/progenitor cell-like subpopulations are enriched in TM-positive cancer cells (unconnected or connected subpopulation) on the RNA expression level and on cellular functional levels.

6 CLINICAL IMPLICATIONS

Current clinical standard treatments for gliomas include radiotherapy, chemotherapy particularly with temozolomide (TMZ), and surgical resection. However, as a result of the robust resistance and self-repairment of gliomas, the existing therapies are all limited and there is an urgent need for a new treatment. TMs may provide a novel potential therapeutic target for gliomas.

6.1 Inhibition of Gap Junction

Since the gap junction is critical to the formation and communication of the network of tumor microtubes (12), we can inhibit the function of the gap junction to impair the integrity of the network. The available inhibitors may include 1) carbenoxolone (an effective medicine for gastric ulcer treatment but also shows a strong inhibitory effect on astrocytoma ICWs) (93), 2) inhibitors of ICWpropagating molecules (IP3, ATP receptors), 3) other calcium antagonists, such as mibefradil, and 4) inhibitors of various types of connexins. However, since various types of connexins, particularly connexin43, have crucial and complex functions to sustain intracellular homeostasis, the selection of the inhibitors must discreetly and considerably concentrate on the target specificity. The recent studies of Schneider and his colleagues in the human neocortical slice model showed a clear road for the introduction of TM network-targeting therapies into clinical concepts, proposing MFA as the first TM-targeted FDAapproved drug (78). They demonstrated that in comparison with TMZ treatment alone, TMZ and connexin43 blocker meclofenamate (MFA) co-treatment dramatically reduced

interconnecting TMs and tumor bulk, achieving better therapeutic effect and prognosis (78). However, further research should be considered to evaluate the concentration levels of MFA within the human brain. In the environment, MFA monotherapy is being tested in patients with recurrent/progressive brain metastases from primary tumors in the United States (NCT02429570). In addition, based on the findings of Schneider et al. (78), a national phase I/II study of MFA/TMZ combination treatment in recurrent MGMT-methylated glioblastoma ("MecMeth" EudraCT2021-000708-39) is underway in Germany to measure the concentrations of MFA within gliomas, examine the safety and practicality of a combined MFA/TMZ strategy, and may acquire first insights into the effectiveness of MFA as the first clinically viable TM-targeted medication.

6.2 Blockade of Electrical and Synaptic Integration of TM-Connected Glioma Cells Into Neural Circuits

Glutamatergic synapses with the glutamate receptors of the AMPA subtype and activity-dependent potassium currents are crucial for glioma cell resistance, invasion, and proliferation (45, 46). The genetic and pharmacological blockade of AMPAR signaling may be applied to block the communication between neurons and glioma cells in order to inhibit the glioma progression and malignancy. According to clinical studies, epileptic seizures are common in individuals with gliomas (94), the recurrence or progression of which is linked to malignant glioma recurrence (95). The clinical manifestation was once thought to be due to the induction of glutamate released by brain tumors (96). However, a recent study suggested that excessive neural activity due to epilepsy might hasten glioma progression and malignancy (97), which is supported by the study of Venkataramani et al. (45). Together, these findings suggest that the clinically approved AMPAR-inhibiting antiepileptic medication perampanel (98) may be a new potential drug for gliomas. Lange et al. demonstrated antitumorigenic effects mediated by perampanel in vitro (99). In addition, the study of Salmaggi et al. has demonstrated that perampanel showed a pro-apoptotic effect on human glioma cell lines when used alone and also showed synergistic cooperativity when combined with TMZ (100). Against this backdrop, further preclinical studies and clinical trials should be considered.

6.3 Inhibition of the Growth of TMs

There are two known factors to be demonstrated to direct the growth of TMs: GAP-43 and Ttyh1 (12). Neurotrophic factors such as NGF and NT-4 (also called NTF4) promote the expression of GAP-43 (12). Silencing genes and disrupting signaling pathways related to the expression of GAP-43 and Ttyh1 may be potential choices.

6.4 Regulation of Associated Signaling Pathway in Tumor Microenvironment

Recent research showed that the downregulation of NOTCH1 is effective to promote the growth of TMs (101). However, since it was also demonstrated to play an important role in the perivascular niche of resistance in gliomas (101), additional

potential signaling pathways or specific blockers remain to be identified. Gritsenko et al. recently demonstrated that intercellular adhesion and signaling provided by p120-catenin– dependent adherens junctions is indispensable for TMconnected glioma cell progression and malignancy, implying that p120-catenin–dependent adherens junctions or their downstream effectors may be a potential target (36).

6.5 Hijack the Network for Drug Transport

It is a potential way of therapy to hijack the network to distribute injected toxic molecules, which are gap junction permeable (24). In addition, it has been demonstrated that lipopolysaccharideanchored macrophages can hijack tumor microtube networks for selective drug transport, serving as versatile bioactive carriers of drugs such as Dox and repressing tumor genesis (102).

7 CONCLUSION AND PERSPECTIVES

The newly discovered TMs and their forming network may provide a new angle to understand the resistance and recurrence of incurable gliomas. TMs share a lot of features in development and function with the axons of immature neural cells and may be seen as "electric synapse" connecting glioma cells and mediating intercellular communication. Both TMconnected cells and immature neural cells can be integrated into a multicellular network and enrich cell stemness (22, 87).

The findings may support the hypothesis that gliomas are initiated by cancer stem cells (5, 7, 57, 103). In essence, glioma progression can be seen as neurodevelopment improperly reactivated at later time points of life (22). The subpopulation of cancer stem cells has been demonstrated to play a crucial role in the robust resistance and recurrence in gliomas (7, 104). Recent research showed that glioma cells with stemness feature tend to extend TMs and integrate into a network to withstand adverse events (87), which might provide a new angle to

REFERENCES

- Ostrom QT, Patil N, Cioffi G, Waite K, Kruchko C, Barnholtz-Sloan JS. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2013-2017. *Neuro* Oncol (2020) 22(12 Suppl 2):iv1–iv96. doi: 10.1093/neuonc/noaa200
- Stupp R, Hegi ME, Mason WP, van den Bent MJ, Taphoorn MJ, Janzer RC, et al. Effects of Radiotherapy With Concomitant and Adjuvant Temozolomide Versus Radiotherapy Alone on Survival in Glioblastoma in a Randomised Phase III Study: 5-Year Analysis of the EORTC-NCIC Trial. *Lancet Oncol* (2009) 10(5):459–66. doi: 10.1016/S1470-2045(09) 70025-7
- Weller M, Wick W, Aldape K, Brada M, Berger M, Pfister SM, et al. Glioma. Nat Rev Dis Primers (2015) 1:15017. doi: 10.1038/nrdp.2015.17
- Nicholson JG, Fine HA. Diffuse Glioma Heterogeneity and Its Therapeutic Implications. *Cancer Discov* (2021) 11(3):575–90. doi: 10.1158/2159-8290.CD-20-1474
- Galli R, Binda E, Orfanelli U, Cipelletti B, Gritti A, De Vitis S, et al. Isolation and Characterization of Tumorigenic, Stem-Like Neural Precursors From Human Glioblastoma. *Cancer Res* (2004) 64(19):7011–21. doi: 10.1158/ 0008-5472.CAN-04-1364

understand the resistance and recurrence mechanism of glioma stem-like cells. However, further research should be undertaken to investigate the stem cell behavior of TM-connected nestinpositive glioma cells and the existence of the subpopulation of TM connected with the expression of other stemness markers. In previous studies, BTSCs and TM-connected glioma cells are always discussed as two independent subpopulations. In this review, we discuss the overlap between them and appeal to do a further comprehensive study. TMs connect glioma cell morphology with the molecular phenotype, suggesting network integration as a new potential signature of cancer stem cells. A similar mechanism may be also applied to other tumors with a membrane tube connection such as breast cancer (105, 106), cervix cancer (107), leukemia (108-110), and lung cancer (111), which require further research. Overall, TM-connected cells provide a novel potential therapeutic target subpopulation for gliomas and might also be a target of resistant cancer stem cells after further research.

AUTHOR CONTRIBUTIONS

The work presented here was carried out in collaboration among all authors. HS conceived this work. XW designed the review and drafted the manuscript. HS revised the manuscript. All authors read, commented on, and approved this manuscript.

FUNDING

Guangdong Basic and Applied Basic Research Foundation (2020A1515010038); the Presidential Foundation of Zhujiang Hospital of Southern Medical University (No. yzjj2018rc03); University students innovation and entrepreneurship project "Three-dimensional visualization of glioma vessels based on tissue clearing technique" (No.202112121004)

- Ignatova TN, Kukekov VG, Laywell ED, Suslov ON, Vrionis FD, Steindler DA. Human Cortical Glial Tumors Contain Neural Stem-Like Cells Expressing Astroglial and Neuronal Markers *In Vitro. Glia* (2002) 39 (3):193–206. doi: 10.1002/glia.10094
- Lathia JD, Mack SC, Mulkearns-Hubert EE, Valentim CL, Rich JN. Cancer Stem Cells in Glioblastoma. *Genes Dev* (2015) 29(12):1203–17. doi: 10.1101/ gad.261982.115
- Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, et al. Glioma Stem Cells Promote Radioresistance by Preferential Activation of the DNA Damage Response. *Nature* (2006) 444(7120):756–60. doi: 10.1038/ nature05236
- Sharanek A, Burban A, Laaper M, Heckel E, Joyal JS, Soleimani VD, et al. OSMR Controls Glioma Stem Cell Respiration and Confers Resistance of Glioblastoma to Ionizing Radiation. *Nat Commun* (2020) 11(1):4116. doi: 10.1038/s41467-020-17885-z
- Huang W, Zhong Z, Luo C, Xiao Y, Li L, Zhang X, et al. The miR-26a/AP-2α/Nanog Signaling Axis Mediates Stem Cell Self-Renewal and Temozolomide Resistance in Glioma. *Theranostics* (2019) 9(19):5497–516. doi: 10.7150/thno.33800
- Shi Y, Guryanova OA, Zhou W, Liu C, Huang Z, Fang X, et al. Ibrutinib Inactivates BMX-STAT3 in Glioma Stem Cells to Impair Malignant Growth

and Radioresistance. *Sci Transl Med* (2018) 10(443):eaah6816. doi: 10.1126/ scitranslmed.aah6816

- Osswald M, Jung E, Sahm F, Solecki G, Venkataramani V, Blaes J, et al. Brain Tumour Cells Interconnect to a Functional and Resistant Network. *Nature* (2015) 528(7580):93–8. doi: 10.1038/nature16071
- Weil S, Osswald M, Solecki G, Grosch J, Jung E, Lemke D, et al. Tumor Microtubes Convey Resistance to Surgical Lesions and Chemotherapy in Gliomas. *Neuro Oncol* (2017) 19(10):1316–26. doi: 10.1093/neuonc/nox070
- Jung E, Osswald M, Blaes J, Wiestler B, Sahm F, Schmenger T, et al. Tweety-Homolog 1 Drives Brain Colonization of Gliomas. J Neurosci (2017) 37 (29):6837–50. doi: 10.1523/JNEUROSCI.3532-16.2017
- Ramírez-Weber FA, Kornberg TB. Cytonemes: Cellular Processes That Project to the Principal Signaling Center in Drosophila Imaginal Discs. *Cell* (1999) 97(5):599–607. doi: 10.1016/S0092-8674(00)80771-0
- Rustom A, Saffrich R, Markovic I, Walther P, Gerdes HH. Nanotubular Highways for Intercellular Organelle Transport. *Science* (2004) 303 (5660):1007–10. doi: 10.1126/science.1093133
- Sowinski S, Jolly C, Berninghausen O, Purbhoo MA, Chauveau A, Köhler K, et al. Membrane Nanotubes Physically Connect T Cells Over Long Distances Presenting a Novel Route for HIV-1 Transmission. *Nat Cell Biol* (2008) 10 (2):211–9. doi: 10.1038/ncb1682
- Eugenin EA, Gaskill PJ, Berman JW. Tunneling Nanotubes (TNT) are Induced by HIV-Infection of Macrophages: A Potential Mechanism for Intercellular HIV Trafficking. *Cell Immunol* (2009) 254(2):142–8. doi: 10.1016/j.cellimm.2008.08.005
- Bracq L, Xie M, Benichou S, Bouchet J. Mechanisms for Cell-To-Cell Transmission of HIV-1. Front Immunol (2018) 9:260. doi: 10.3389/fimmu.2018.00260
- Gousset K, Schiff E, Langevin C, Marijanovic Z, Caputo A, Browman DT, et al. Prions Hijack Tunnelling Nanotubes for Intercellular Spread. *Nat Cell Biol* (2009) 11(3):328–36. doi: 10.1038/ncb1841
- Sartori-Rupp A, Cordero Cervantes D, Pepe A, Gousset K, Delage E, Corroyer-Dulmont S, et al. Correlative Cryo-Electron Microscopy Reveals the Structure of TNTs in Neuronal Cells. *Nat Commun* (2019) 10(1):342. doi: 10.1038/s41467-018-08178-7
- Jung E, Alfonso J, Osswald M, Monyer H, Wick W, Winkler F. Emerging Intersections Between Neuroscience and Glioma Biology. *Nat Neurosci* (2019) 22(12):1951–60. doi: 10.1038/s41593-019-0540-y
- Laug D, Glasgow SM, Deneen B. A Glial Blueprint for Gliomagenesis. Nat Rev Neurosci (2018) 19(7):393–403. doi: 10.1038/s41583-018-0014-3
- Osswald M, Solecki G, Wick W, Winkler F. A Malignant Cellular Network in Gliomas: Potential Clinical Implications. *Neuro Oncol* (2016) 18(4):479–85. doi: 10.1093/neuonc/now014
- Lowery LA, Van Vactor D. The Trip of the Tip: Understanding the Growth Cone Machinery. Nat Rev Mol Cell Biol (2009) 10(5):332–43. doi: 10.1038/ nrm2679
- Tsai JW, Bremner KH, Vallee RB. Dual Subcellular Roles for LIS1 and Dynein in Radial Neuronal Migration in Live Brain Tissue. *Nat Neurosci* (2007) 10(8):970–9. doi: 10.1038/nn1934
- Sin WC, Crespin S, Mesnil M. Opposing Roles of Connexin43 in Glioma Progression. *Biochim Biophys Acta* (2012) 1818(8):2058–67. doi: 10.1016/ j.bbamem.2011.10.022
- 28. Le HT, Sin WC, Lozinsky S, Bechberger J, Vega JL, Guo XQ, et al. Gap Junction Intercellular Communication Mediated by Connexin43 in Astrocytes is Essential for Their Resistance to Oxidative Stress. J Biol Chem (2014) 289(3):1345–54. doi: 10.1074/jbc.M113.508390
- Skene JH, Jacobson RD, Snipes GJ, McGuire CB, Norden JJ, Freeman JA. A Protein Induced During Nerve Growth (GAP-43) is a Major Component of Growth-Cone Membranes. *Science* (1986) 233(4765):783–6. doi: 10.1126/ science.3738509
- Goslin K, Schreyer DJ, Skene JH, Banker G. Development of Neuronal Polarity: GAP-43 Distinguishes Axonal From Dendritic Growth Cones. *Nature* (1988) 336(6200):672–4. doi: 10.1038/336672a0
- Aigner L, Arber S, Kapfhammer JP, Laux T, Schneider C, Botteri F, et al. Overexpression of the Neural Growth-Associated Protein GAP-43 Induces Nerve Sprouting in the Adult Nervous System of Transgenic Mice. *Cell* (1995) 83(2):269–78. doi: 10.1016/0092-8674(95)90168-X
- Haag D, Zipper P, Westrich V, Karra D, Pfleger K, Toedt G, et al. Nos2 Inactivation Promotes the Development of Medulloblastoma in Ptch1(+/-)

Mice by Deregulation of Gap43-Dependent Granule Cell Precursor Migration. *PloS Genet* (2012) 8(3):e1002572. doi: 10.1371/ journal.pgen.1002572

- 33. Stefaniuk M, Swiech L, Dzwonek J, Lukasiuk K. Expression of Ttyh1, a Member of the Tweety Family in Neurons *In Vitro* and *In Vivo* and its Potential Role in Brain Pathology. *J Neurochem* (2010) 115(5):1183–94. doi: 10.1111/j.1471-4159.2010.07023.x
- Zuber MX, Goodman DW, Karns LR, Fishman MC. The Neuronal Growth-Associated Protein GAP-43 Induces Filopodia in non-Neuronal Cells. *Science* (1989) 244(4909):1193–5. doi: 10.1126/science.2658062
- Aigner L, Caroni P. Absence of Persistent Spreading, Branching, and Adhesion in GAP-43-Depleted Growth Cones. J Cell Biol (1995) 128 (4):647–60. doi: 10.1083/jcb.128.4.647
- Gritsenko PG, Atlasy N, Dieteren CEJ, Navis AC, Venhuizen JH, Veelken C, et al. P120-Catenin-Dependent Collective Brain Infiltration by Glioma Cell Networks. Nat Cell Biol (2020) 22(1):97–107. doi: 10.1038/s41556-019-0443-x
- Dohn MR, Brown MV, Reynolds AB. An Essential Role for P120-Catenin in Src- and Rac1-Mediated Anchorage-Independent Cell Growth. J Cell Biol (2009) 184(3):437–50. doi: 10.1083/jcb.200807096
- van de Ven RA, Tenhagen M, Meuleman W, van Riel JJ, Schackmann RC, Derksen PW. Nuclear P120-Catenin Regulates the Anoikis Resistance of Mouse Lobular Breast Cancer Cells Through Kaiso-Dependent Wnt11 Expression. *Dis Model Mech* (2015) 8(4):373–84. doi: 10.1242/dmm.018648
- Schackmann RC, van Amersfoort M, Haarhuis JH, Vlug EJ, Halim VA, Roodhart JM, et al. Cytosolic P120-Catenin Regulates Growth of Metastatic Lobular Carcinoma Through Rock1-Mediated Anoikis Resistance. J Clin Invest (2011) 121(8):3176–88. doi: 10.1172/JCI41695
- Leybaert L, Sanderson MJ. Intercellular Ca(2+) Waves: Mechanisms and Function. *Physiol Rev* (2012) 92(3):1359–92. doi: 10.1152/physrev.00029.2011
- Scemes E, Giaume C. Astrocyte Calcium Waves: What They are and What They do. *Glia* (2006) 54(7):716–25. doi: 10.1002/glia.20374
- Lacar B, Young SZ, Platel JC, Bordey A. Gap Junction-Mediated Calcium Waves Define Communication Networks Among Murine Postnatal Neural Progenitor Cells. *Eur J Neurosci* (2011) 34(12):1895–905. doi: 10.1111/ j.1460-9568.2011.07901.x
- 43. Malmersjö S, Rebellato P, Smedler E, Planert H, Kanatani S, Liste I, et al. Neural Progenitors Organize in Small-World Networks to Promote Cell Proliferation. *Proc Natl Acad Sci U S A* (2013) 110(16):E1524–32. doi: 10.1073/pnas.1220179110
- 44. Pinto G, Saenz-de-Santa-Maria I, Chastagner P, Perthame E, Delmas C, Toulas C, et al. Patient-Derived Glioblastoma Stem Cells Transfer Mitochondria Through Tunneling Nanotubes in Tumor Organoids. *Biochem J* (2021) 478(1):21–39. doi: 10.1042/BCJ20200710
- Venkataramani V, Tanev DI, Strahle C, Studier-Fischer A, Fankhauser L, Kessler T, et al. Glutamatergic Synaptic Input to Glioma Cells Drives Brain Tumour Progression. *Nature* (2019) 573(7775):532–8. doi: 10.1038/s41586-019-1564-x
- Venkatesh HS, Morishita W, Geraghty AC, Silverbush D, Gillespie SM, Arzt M, et al. Electrical and Synaptic Integration of Glioma Into Neural Circuits. *Nature* (2019) 573(7775):539–45. doi: 10.1038/s41586-019-1563-y
- Paget S. The Distribution of Secondary Growths in Cancer of the Breast. 1889. *Cancer Metastasis Rev* (1989) 8(2):98–101. doi: 10.1016/S0140-6736 (00)49915-0
- Peinado H, Zhang H, Matei IR, Costa-Silva B, Hoshino A, Rodrigues G, et al. Pre-Metastatic Niches: Organ-Specific Homes for Metastases. *Nat Rev Cancer* (2017) 17(5):302–17. doi: 10.1038/nrc.2017.6
- Fidler IJ, Kripke ML. Metastasis Results From Preexisting Variant Cells Within a Malignant Tumor. *Science* (1977) 197(4306):893–5. doi: 10.1126/ science.887927
- Komsany A, Pezzella F. The Perivascular Niche. In: *Tumor Vascularization* (2020). p. 113–27. Amsterdam, Elsevier
- Sanai N, Tramontin AD, Quiñones-Hinojosa A, Barbaro NM, Gupta N, Kunwar S, et al. Unique Astrocyte Ribbon in Adult Human Brain Contains Neural Stem Cells But Lacks Chain Migration. *Nature* (2004) 427 (6976):740–4. doi: 10.1038/nature02301
- Bjornsson CS, Apostolopoulou M, Tian Y, Temple S. It Takes a Village: Constructing the Neurogenic Niche. *Dev Cell* (2015) 32(4):435–46. doi: 10.1016/j.devcel.2015.01.010

- Riquelme PA, Drapeau E, Doetsch F. Brain Micro-Ecologies: Neural Stem Cell Niches in the Adult Mammalian Brain. *Philos Trans R Soc Lond B Biol Sci* (2008) 363(1489):123–37. doi: 10.1098/rstb.2006.2016
- Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. Cell (2011) 144(5):646–74. doi: 10.1016/j.cell.2011.02.013
- Calabrese C, Poppleton H, Kocak M, Hogg TL, Fuller C, Hamner B, et al. A Perivascular Niche for Brain Tumor Stem Cells. *Cancer Cell* (2007) 11 (1):69–82. doi: 10.1016/j.ccr.2006.11.020
- Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, et al. Identification of a Cancer Stem Cell in Human Brain Tumors. *Cancer Res* (2003) 63(18):5821–8.
- Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, et al. Identification of Human Brain Tumour Initiating Cells. *Nature* (2004) 432 (7015):396–401. doi: 10.1038/nature03128
- Taverna E, Huttner WB. Neural Progenitor Nuclei IN Motion. Neuron (2010) 67(6):906–14. doi: 10.1016/j.neuron.2010.08.027
- Cuddapah VA, Robel S, Watkins S, Sontheimer H. A Neurocentric Perspective on Glioma Invasion. *Nat Rev Neurosci* (2014) 15(7):455–65. doi: 10.1038/nrn3765
- Cayre M, Canoll P, Goldman JE. Cell Migration in the Normal and Pathological Postnatal Mammalian Brain. *Prog Neurobiol* (2009) 88(1):41– 63. doi: 10.1016/j.pneurobio.2009.02.001
- Ohtaka-Maruyama C, Okamoto M, Endo K, Oshima M, Kaneko N, Yura K, et al. Synaptic Transmission From Subplate Neurons Controls Radial Migration of Neocortical Neurons. *Science* (2018) 360(6386):313–7. doi: 10.1126/science.aar2866
- Gibson EM, Purger D, Mount CW, Goldstein AK, Lin GL, Wood LS, et al. Neuronal Activity Promotes Oligodendrogenesis and Adaptive Myelination in the Mammalian Brain. *Science* (2014) 344(6183):1252304. doi: 10.1126/ science.1252304
- 63. Tombal B, Denmeade SR, Gillis JM, Isaacs JT. A Supramicromolar Elevation of Intracellular Free Calcium ([Ca(2+)](i)) is Consistently Required to Induce the Execution Phase of Apoptosis. *Cell Death Differ* (2002) 9 (5):561–73. doi: 10.1038/sj.cdd.4400999
- 64. Chen Q, Boire A, Jin X, Valiente M, Er EE, Lopez-Soto A, et al. Carcinoma-Astrocyte Gap Junctions Promote Brain Metastasis by cGAMP Transfer. *Nature* (2016) 533(7604):493–8. doi: 10.1038/nature18268
- 65. Lin Q, Balasubramanian K, Fan D, Kim SJ, Guo L, Wang H, et al. Reactive Astrocytes Protect Melanoma Cells From Chemotherapy by Sequestering Intracellular Calcium Through Gap Junction Communication Channels. *Neoplasia* (2010) 12(9):748–54. doi: 10.1593/neo.10602
- 66. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. Radiotherapy Plus Concomitant and Adjuvant Temozolomide for Glioblastoma. N Engl J Med (2005) 352(10):987–96. doi: 10.1056/ NEJMoa043330
- Buckner JC, Shaw EG, Pugh SL, Chakravarti A, Gilbert MR, Barger GR, et al. Radiation Plus Procarbazine, CCNU, and Vincristine in Low-Grade Glioma. *N Engl J Med* (2016) 374(14):1344–55. doi: 10.1056/NEJMoa1500925
- Weller M, van den Bent M, Hopkins K, Tonn JC, Stupp R, Falini A, et al. EANO Guideline for the Diagnosis and Treatment of Anaplastic Gliomas and Glioblastoma. *Lancet Oncol* (2014) 15(9):e395–403. doi: 10.1016/S1470-2045(14)70011-7
- Petrecca K, Guiot MC, Panet-Raymond V, Souhami L. Failure Pattern Following Complete Resection Plus Radiotherapy and Temozolomide is at the Resection Margin in Patients With Glioblastoma. J Neurooncol (2013) 111(1):19–23. doi: 10.1007/s11060-012-0983-4
- Ekblad L, Lindgren G, Persson E, Kjellén E, Wennerberg J. Cell-Line-Specific Stimulation of Tumor Cell Aggressiveness by Wound Healing Factors - a Central Role for STAT3. *BMC Cancer* (2013) 13:33. doi: 10.1186/1471-2407-13-33
- Mannino M, Yarnold J. Effect of Breast-Duct Anatomy and Wound-Healing Responses on Local Tumour Recurrence After Primary Surgery for Early Breast Cancer. *Lancet Oncol* (2009) 10(4):425–9. doi: 10.1016/S1470-2045 (09)70040-3
- 72. Licitra L, Perrone F, Tamborini E, Bertola L, Ghirelli C, Negri T, et al. Role of EGFR Family Receptors in Proliferation of Squamous Carcinoma Cells Induced by Wound Healing Fluids of Head and Neck Cancer Patients. *Ann Oncol* (2011) 22(8):1886–93. doi: 10.1093/annonc/mdq756

- Chen X, Zhang M, Gan H, Wang H, Lee JH, Fang D, et al. A Novel Enhancer Regulates MGMT Expression and Promotes Temozolomide Resistance in Glioblastoma. *Nat Commun* (2018) 9(1):2949. doi: 10.1038/s41467-018-05373-4
- 74. Kitange GJ, Carlson BL, Schroeder MA, Grogan PT, Lamont JD, Decker PA, et al. Induction of MGMT Expression is Associated With Temozolomide Resistance in Glioblastoma Xenografts. *Neuro Oncol* (2009) 11(3):281–91. doi: 10.1215/15228517-2008-090
- Murphy SF, Varghese RT, Lamouille S, Guo S, Pridham KJ, Kanabur P, et al. Connexin 43 Inhibition Sensitizes Chemoresistant Glioblastoma Cells to Temozolomide. *Cancer Res* (2016) 76(1):139–49. doi: 10.1158/0008-5472.CAN-15-1286
- Munoz JL, Rodriguez-Cruz V, Greco SJ, Ramkissoon SH, Ligon KL, Rameshwar P. Temozolomide Resistance in Glioblastoma Cells Occurs Partly Through Epidermal Growth Factor Receptor-Mediated Induction of Connexin 43. Cell Death Dis (2014) 5(3):e1145. doi: 10.1038/cddis.2014.111
- Gielen PR, Aftab Q, Ma N, Chen VC, Hong X, Lozinsky S, et al. Connexin43 Confers Temozolomide Resistance in Human Glioma Cells by Modulating the Mitochondrial Apoptosis Pathway. *Neuropharmacology* (2013) 75:539– 48. doi: 10.1016/j.neuropharm.2013.05.002
- Schneider M, Vollmer L, Potthoff AL, Ravi VM, Evert BO, Rahman MA, et al. Meclofenamate Causes Loss of Cellular Tethering and Decoupling of Functional Networks in Glioblastoma. *Neuro Oncol* (2021) 23(11):1885–97. doi: 10.1093/neuonc/noab092
- Lapointe S, Perry A, Butowski NA. Primary Brain Tumours in Adults. Lancet (2018) 392(10145):432–46. doi: 10.1016/S0140-6736(18)30990-5
- Mirimanoff RO, Gorlia T, Mason W, Van den Bent MJ, Kortmann RD, Fisher B, et al. Radiotherapy and Temozolomide for Newly Diagnosed Glioblastoma: Recursive Partitioning Analysis of the EORTC 26981/ 22981-NCIC CE3 Phase III Randomized Trial. J Clin Oncol (2006) 24 (16):2563–9. doi: 10.1200/JCO.2005.04.5963
- Jaeckle KA, Ballman KV, van den Bent M, Giannini C, Galanis E, Brown PD, et al. CODEL: Phase III Study of RT, RT + TMZ, or TMZ for Newly Diagnosed 1p/19q Codeleted Oligodendroglioma. Analysis From the Initial Study Design. *Neuro Oncol* (2021) 23(3):457–67. doi: 10.1093/neuonc/noaa168
- Perry JR, Laperriere N, O'Callaghan CJ, Brandes AA, Menten J, Phillips C, et al. Short-Course Radiation Plus Temozolomide in Elderly Patients With Glioblastoma. N Engl J Med (2017) 376(11):1027–37. doi: 10.1056/ NEJMoa1611977
- Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem Cells, Cancer, and Cancer Stem Cells. *Nature* (2001) 414(6859):105–11. doi: 10.1038/35102167
- Bonnet D, Dick JE. Human Acute Myeloid Leukemia is Organized as a Hierarchy That Originates From a Primitive Hematopoietic Cell. Nat Med (1997) 3(7):730–7. doi: 10.1038/nm0797-730
- Prager BC, Bhargava S, Mahadev V, Hubert CG, Rich JN. Glioblastoma Stem Cells: Driving Resilience Through Chaos. *Trends Cancer* (2020) 6(3):223–35. doi: 10.1016/j.trecan.2020.01.009
- Dirkse A, Golebiewska A, Buder T, Nazarov PV, Muller A, Poovathingal S, et al. Stem Cell-Associated Heterogeneity in Glioblastoma Results From Intrinsic Tumor Plasticity Shaped by the Microenvironment. *Nat Commun* (2019) 10(1):1787. doi: 10.1038/s41467-019-09853-z
- Xie R, Kessler T, Grosch J, Hai L, Venkataramani V, Huang L, et al. Tumor Cell Network Integration in Glioma Represents a Stemness Feature. *Neuro* Oncol (2021) 23(5):757–69. doi: 10.1093/neuonc/noaa275
- Chen J, Li Y, Yu TS, McKay RM, Burns DK, Kernie SG, et al. A Restricted Cell Population Propagates Glioblastoma Growth After Chemotherapy. *Nature* (2012) 488(7412):522–6. doi: 10.1038/nature11287
- Shankar A, Kumar S, Iskander AS, Varma NR, Janic B, deCarvalho A, et al. Subcurative Radiation Significantly Increases Cell Proliferation, Invasion, and Migration of Primary Glioblastoma Multiforme *In Vivo. Chin J Cancer* (2014) 33(3):148–58. doi: 10.5732/cjc.013.10095
- Gao X, McDonald JT, Hlatky L, Enderling H. Acute and Fractionated Irradiation Differentially Modulate Glioma Stem Cell Division Kinetics. *Cancer Res* (2013) 73(5):1481–90. doi: 10.1158/0008-5472.CAN-12-3429
- Xie XP, Laks DR, Sun D, Ganbold M, Wang Z, Pedraza AM, et al. Quiescent Human Glioblastoma Cancer Stem Cells Drive Tumor Initiation, Expansion, and Recurrence Following Chemotherapy. *Dev Cell* (2022) 57(1):32–46.e8. doi: 10.1016/j.devcel.2021.12.007

- Chen W, Dong J, Haiech J, Kilhoffer MC, Zeniou M. Cancer Stem Cell Quiescence and Plasticity as Major Challenges in Cancer Therapy. *Stem Cells Int* (2016) 2016:1740936. doi: 10.1155/2016/1740936
- Parys B, Côté A, Gallo V, De Koninck P, Sik A. Intercellular Calcium Signaling Between Astrocytes and Oligodendrocytes *via* Gap Junctions in Culture. *Neuroscience* (2010) 167(4):1032–43. doi: 10.1016/j.neuroscience.2010.03.004
- 94. Weller M, Stupp R, Wick W. Epilepsy Meets Cancer: When, Why, and What to do About it? *Lancet Oncol* (2012) 13(9):e375–82. doi: 10.1016/S1470-2045 (12)70266-8
- Körber V, Yang J, Barah P, Wu Y, Stichel D, Gu Z, et al. Evolutionary Trajectories of IDH(WT) Glioblastomas Reveal a Common Path of Early Tumorigenesis Instigated Years Ahead of Initial Diagnosis. *Cancer Cell* (2019) 35(4):692–704.e12. doi: 10.1016/j.ccell.2019.02.007
- Buckingham SC, Campbell SL, Haas BR, Montana V, Robel S, Ogunrinu T, et al. Glutamate Release by Primary Brain Tumors Induces Epileptic Activity. *Nat Med* (2011) 17(10):1269–74. doi: 10.1038/nm.2453
- Huberfeld G, Vecht CJ. Seizures and Gliomas–Towards a Single Therapeutic Approach. Nat Rev Neurol (2016) 12(4):204–16. doi: 10.1038/nrneurol.2016.26
- Potschka H, Trinka E. Perampanel: Does it Have Broad-Spectrum Potential? *Epilepsia* (2019) 60 Suppl 1:22–36. doi: 10.1111/epi.14456
- Lange F, Weßlau K, Porath K, Hörnschemeyer J, Bergner C, Krause BJ, et al. AMPA Receptor Antagonist Perampanel Affects Glioblastoma Cell Growth and Glutamate Release *In Vitro. PloS One* (2019) 14(2):e0211644. doi: 10.1371/journal.pone.0211644
- 100. Salmaggi A, Corno C, Maschio M, Donzelli S, D'Urso A, Perego P, et al. Synergistic Effect of Perampanel and Temozolomide in Human Glioma Cell Lines. J Pers Med (2021) 11(5):390. doi: 10.3390/jpm11050390
- 101. Jung E, Osswald M, Ratliff M, Dogan H, Xie R, Weil S, et al. Tumor Cell Plasticity, Heterogeneity, and Resistance in Crucial Microenvironmental Niches in Glioma. *Nat Commun* (2021) 12(1):1014. doi: 10.1038/s41467-021-21117-3
- 102. Guo L, Zhang Y, Wei R, Wang C, Feng M. Lipopolysaccharide-Anchored Macrophages Hijack Tumor Microtube Networks for Selective Drug Transport and Augmentation of Antitumor Effects in Orthotopic Lung Cancer. *Theranostics* (2019) 9(23):6936–48. doi: 10.7150/thno.37380
- 103. Lathia JD, Gallagher J, Myers JT, Li M, Vasanji A, McLendon RE, et al. Direct In Vivo Evidence for Tumor Propagation by Glioblastoma Cancer Stem Cells. PloS One (2011) 6(9):e24807. doi: 10.1371/journal.pone.0024807
- 104. Paul R, Dorsey JF, Fan Y. Cell Plasticity, Senescence, and Quiescence in Cancer Stem Cells: Biological and Therapeutic Implications. *Pharmacol Ther* (2021) 231:107985. doi: 10.1016/j.pharmthera.2021.107985
- 105. Tishchenko A, Azorín DD, Vidal-Brime L, Muñoz MJ, Arenas PJ, Pearce C, et al. Cx43 and Associated Cell Signaling Pathways Regulate Tunneling

Nanotubes in Breast Cancer Cells. Cancers (Basel) (2020) 12(10):2798. doi: 10.3390/cancers12102798

- 106. Connor Y, Tekleab S, Nandakumar S, Walls C, Tekleab Y, Husain A, et al. Physical Nanoscale Conduit-Mediated Communication Between Tumour Cells and the Endothelium Modulates Endothelial Phenotype. *Nat Commun* (2015) 6:8671. doi: 10.1038/ncomms9671
- 107. Rimkutė L, Jotautis V, Marandykina A, Sveikatienė R, Antanavičiūtė I, Skeberdis VA. The Role of Neural Connexins in HeLa Cell Mobility and Intercellular Communication Through Tunneling Tubes. *BMC Cell Biol* (2016) 17:3. doi: 10.1186/s12860-016-0080-1
- 108. Polak R, de Rooij B, Pieters R, den Boer ML. B-Cell Precursor Acute Lymphoblastic Leukemia Cells Use Tunneling Nanotubes to Orchestrate Their Microenvironment. *Blood* (2015) 126(21):2404–14. doi: 10.1182/ blood-2015-03-634238
- 109. Burt R, Dey A, Aref S, Aguiar M, Akarca A, Bailey K, et al. Activated Stromal Cells Transfer Mitochondria to Rescue Acute Lymphoblastic Leukemia Cells From Oxidative Stress. *Blood* (2019) 134(17):1415–29. doi: 10.1182/ blood.2019001398
- 110. Marlein CR, Zaitseva L, Piddock RE, Robinson SD, Edwards DR, Shafat MS, et al. NADPH Oxidase-2 Derived Superoxide Drives Mitochondrial Transfer From Bone Marrow Stromal Cells to Leukemic Blasts. *Blood* (2017) 130 (14):1649–60. doi: 10.1182/blood-2017-03-772939
- 111. Lou E, Fujisawa S, Morozov A, Barlas A, Romin Y, Dogan Y, et al. Tunneling Nanotubes Provide a Unique Conduit for Intercellular Transfer of Cellular Contents in Human Malignant Pleural Mesothelioma. *PloS One* (2012) 7(3): e33093. doi: 10.1371/journal.pone.0033093

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Wang, Liang and Sun. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author (s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.