# THE NEUROBIOLOGY OF ANTIEPILEPTIC DRUGS

### Michael A. Rogawski\* and Wolfgang Löscher<sup>‡</sup>

Antiepileptic drugs (AEDs) provide satisfactory control of seizures for most patients with epilepsy. The drugs have the remarkable ability to protect against seizures while permitting normal functioning of the nervous system. AEDs act on diverse molecular targets to selectively modify the excitability of neurons so that seizure-related firing is blocked without disturbing non-epileptic activity. This occurs largely through effects on voltage-gated sodium and calcium channels, or by promoting inhibition mediated by GABA, ( $\gamma$ -aminobutyric acid, type A) receptors. The subtle biophysical modifications in channel behaviour that are induced by AEDs are often functionally opposite to defects in channel properties that are caused by mutations associated with epilepsy in humans.

ANTIEPILEPTIC DRUG
A drug that protects against the occurrence of epileptic seizures; an alternative to the term 'anticonvulsant drug'.

\*Epilepsy Research Section, National Institute of Neurological Disorders and Stroke. National Institutes of Health, Bethesda, Maryland 20892, USA <sup>‡</sup>Department of Pharmacology, Toxicology and Pharmacy, University of Veterinary Medicine, and Center for Systems Neuroscience, Bünteweg 17, D-30559 Hannover. Germany Correspondence to M.A.R. e-mail: michael.rogawski@nih.gov doi:10.1038/nrn1430

ANTIEPILEPTIC DRUGS (AEDs) are primarily intended to prevent epileptic seizures. They are also beneficial in diverse non-epileptic conditions and are commonly used in the treatment of migraine headache, neuropathic pain and bipolar affective disorder. AEDs can also be useful for treating essential tremor, myotonia, dystonia, anxiety disorders, schizophrenia, restless legs syndrome, social phobia, post-traumatic stress syndrome, and alcohol dependence and withdrawal<sup>1</sup>. Around two-dozen distinct molecular entities are marketed worldwide for epilepsy (BOX 1). To exhibit antiepileptic activity, a drug must act on one or more target molecules in the brain. These targets include ion channels, neurotransmitter transporters and neurotransmitter metabolic enzymes. The ultimate effects of these interactions are to modify the bursting properties of neurons and to reduce synchronization in localized neuronal ensembles. In addition, AEDs inhibit the spread of abnormal firing to distant sites, which is required for the expression of behavioural seizure activity. GENERALIZED ABSENCE SEIZURES, unlike other seizure types, are believed to result from thalamocortical synchronization. Interference with the rhythm-generating mechanisms that underlie the synchronized activity in this circuit is necessary to abort these seizures.

It is convenient to categorize AED actions according to those that involve (1) modulation of voltage-gated ion channels; (2) enhancement of synaptic inhibition; and (3) inhibition of synaptic excitation. Voltage-gated ion channels (including sodium, calcium and potassium channels) shape the subthreshold electrical behaviour of the neuron, allow it to fire action potentials, regulate its responsiveness to synaptic signals, contribute to the PAROXYSMAL DEPOLARIZATION SHIFT, and ultimately are integral to the generation of seizure discharges. In addition, voltage-gated ion channels are crucial elements in neurotransmitter release, which is required for synaptic transmission. Consequently, they are key targets for AEDs that inhibit epileptic bursting, synchronization and seizure spread. Synaptic inhibition and excitation are mediated by neurotransmitter-regulated channels; these channels permit synchronization of neural ensembles and allow propagation of the abnormal discharge to local and distant sites. AEDs that modify excitatory and inhibitory neurotransmission therefore can also suppress bursting and, when they inhibit synaptic excitation, can have prominent effects on seizure spread.

Which ion channels are relevant to the actions of AEDs? Studies in animal models have shown that protection from seizures can be achieved by blockade of sodium or calcium channels, and probably also through facilitation of potassium channels (as might be the case for the novel AED retigabine<sup>2</sup>). Antiepileptic effects are also produced by drugs that enhance inhibition mediated by GABA<sub>A</sub> ( $\gamma$ -aminobutyric acid, type A) receptors, or through effects on glycine systems, the regionally specific transmitter systems (including

#### Box 1 Currently approved antiepileptic drugs

The following antiepileptic drugs are registered in the United States and elsewhere: acetazolamide, carbamazepine, clonazepam, clorazepate, ethosuximide, ethotoin, felbamate, gabapentin, lamotrigine, levetiracetam, mephenytoin, methsuximide, oxcarbazepine, phenobarbital, phenytoin, primidone, tiagabine, topiramate, trimethadione, valproate, vigabatrin (available in Canada, Europe and elsewhere, but not the United States) and zonisamide.

The following additional agents are mainly used for the acute therapy of status epilepticus: diazepam, fosphenytoin, lorazepam, midazolam and propofol.

GENERALIZED ABSENCE SEIZURE A non-convulsive seizure that typically occurs in childhood. It is characterized by a sudden, brief impairment of consciousness, cessation of ongoing activity without loss of postural tone, and 3 Hz rhythmic cortical discharges of geneneralized onset. The usual duration is 5–10 seconds; several episodes can occur daily.

monoamines such as catecholamines, serotonin and histamine, and neuropeptides such as opioid peptides, galanin and neuropeptide Y) and the inhibitory neuromodulator adenosine<sup>3</sup>. In addition, blockade of glutamate receptors (including those of the NMDA (N-methyl-D-aspartate), AMPA ( $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid), kainate and group I METABOTROPIC mGluR1 and mGluR5 types) can also protect against seizures in animal models. In principle, it might be possible to prevent seizures by targeting any one or a combination of these systems. In fact, the development of AEDs by screening in animal models that are

non-biased with respect to mechanism has uncovered drugs that act by unique combinations of these mechanisms, and in some cases has identified drugs that act by entirely new mechanisms. As a result, marketed AEDs act by a remarkable diversity of actions, and no two marketed drugs work in exactly the same way.

#### Molecular targets of AED action

Voltage-gated sodium channels. Voltage-gated sodium channels are responsible for the rising phase of neuronal action potentials. When neurons are depolarized to action potential threshold, the sodium channel protein senses the depolarization and, within a few hundred microseconds, undergoes a conformational change that converts the channel from its closed (resting) non-conducting state to the open conducting state that permits sodium flux. Within a few milliseconds, the channel inactivates, terminating the flow of sodium ions. The channel must then be repolarized before it can be activated again by a subsequent depolarization. Brain sodium channels can rapidly cycle through the resting, open and inactivated states, allowing neurons to fire high-frequency trains of action potentials, as is required for normal brain function and for the expression of epileptic activity.

Table 1   Antiepileptic drugs and their molecular targets										
Drug	Sodium channels*	Calcium channels*	GABA system*	Glutamate receptors*	Partial seizure <sup>‡</sup>	GTC seizure <sup>‡</sup>	Absence seizure <sup>‡</sup>	Myoclonic seizure <sup>‡</sup>	Infantile spasms <sup>‡§</sup>	Lennox- Gastaut <sup>‡</sup>
Predominant sodium (and calcium) channel activity										
Phenytoin	I <sub>NaF</sub> , I <sub>NaP</sub>				+	+	-	-		
Carbamazepine	I <sub>NaF</sub>				+	+	-	-		
Oxcarbazepine	l <sub>NaF</sub>				+	+	-	-		
Lamotrigine	I <sub>NaF</sub>	HVA			+	+	+	(+/-)		+
Zonisamide	l <sub>NaF</sub>	T-type			+	+	(+)	(+)	(+)	(+)
Mixed, complex or poorly understood actions										
Valproate	I <sub>NaF</sub> ? I <sub>NaP</sub> ?	T-type?	↑ GABA turnover		+	+	+	+		(+)
Felbamate	I <sub>NaF</sub>	HVA	$GABA_AR$	NMDA	+	+	(+)			+
Topiramate	I <sub>NaF</sub> , I <sub>NaP</sub>	HVA	GABA <sub>A</sub> R	KA/AMPA	+	+	(+)	(+)	(+)	+
Ethosuximide	I <sub>NaP</sub> ?	T-type			-	-	+			
Gabapentin		ΗVΑ (α2δ)	↑ GABA turnover		+	+	-	-		
Levetiracetam <sup>  </sup>		HVA	Reverses DMCM		+	(+)	(+)	(+)		
Phenobarbital		HVA	GABA <sub>A</sub> R	AMPA	+	+	-			
GABA-mediated mechanisms										
Benzodiazepines			GABA <sub>A</sub> R		+	+	+	+		(+)
Vigabatrin			GABAT		+	+	_	-	+	(+)
Tiagabine			GABA- transporter		+	+	-			

\*Molecular targets. Not all molecular targets are shown; additional targets are discussed in the text.  $^{t}$ Clinical efficacy of drugs on symptoms. Clinical evidence: '+' indicates controlled trials or several open-label trials and general acceptance of utility; parentheses indicate less extensive base of evidence. '-' indicates evidence of lack of efficacy or worsening.  $^{s}$ A catastrophic epilepsy syndrome usually beginning in the first year of life in which there are typically 'Jackknife spasms' (myoclonic seizures involving the muscles of the neck, trunk and limbs, with nodding of the head and stiffening of the arms) and a disorganized cortical discharge termed hypsarrhythmia.  $^{\parallel}$ Levetiractam binds with high affinity to synaptic vesicle protein 2A (SV2A), a ubiquitous 90 kDa protein that is associated with synaptic vesicles and is believed to participate in the regulation of  $Ca^{2+}$ -dependent neurotransmitter release; SV2A-knockout mice exhibit seizures. AMPA,  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole-propionic acid; GABA,  $\gamma$ -aminobutyric acid; GABA, Receptor; GTC, generalized tonic-clonic; HVA, high voltage activated:  $I_{\text{NaP}}$ , fast sodium current;  $I_{\text{NaP}}$ , persistent sodium current; KA, kainate; NMDA, N-methyl- $\rho$ -aspartate. DMCM (methyl- $\rho$ -Archimethoxy-4-ethyl- $\rho$ -carboline-3-carboxylate) is a negative allosteric modulator of GABA, receptors. Adrenocorticotropic hormone and prednisolone are recognized treatments for infantile spasms. Lamotrigine is also effective in myoclonic astatic epilepsy, but causes worsening of other forms of myoclonic epilepsy. Gabapentin has been associated with focal myoclonus.

554 | JULY 2004 | VOLUME 5 www.nature.com/reviews/neuro

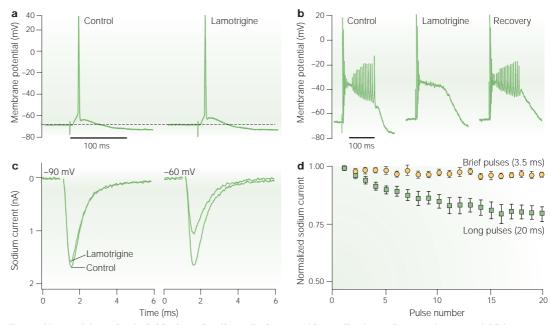


Figure 1 | Lamotrigine selectively blocks epileptiform discharges without affecting ordinary action potential firing. **a** | Intracellular recording showing lack of effect of lamotrigine (50  $\mu$ M) on evoked excitatory synaptic response with superimposed sodium-dependent action potentials. **b** | Recording under magnesium-free conditions (to enhance NMDA (*N*-methyl-D-aspartate) receptor responses) and in the presence of 20  $\mu$ M bicuculline (to block GABA<sub>A</sub> ( $\gamma$ -aminobutyric acid, type A) receptors) showing that lamotrigine selectively inhibits late epileptiform after-discharges without affecting initial action potential responses. Intracellular recordings are from rat CA1 hippocampal neurons evoked with Schaffer collateral/commissural fibre stimulation. **c** | Whole-cell voltage-clamp recording from CNallA-1 cells (CHO-K1 cells stably transfected with cDNA encoding rat brain type IIA sodium channels) illustrating voltage-dependent inhibition of sodium current by lamotrigine (50  $\mu$ M). Currents were activated by step-depolarization to 0 mV from a 30 s conditioning prepulse at the indicated membrane potentials. **d** | Accumulation of block of sodium current with repetitive depolarization (11 Hz) from –90 to 0 mV with pulses of two durations illustrating use-dependence. Modified, with permission, from REF.17 © (1995) Springer-Verlag.

PAROXYSMAL DEPOLARIZATION SHIFT
Abnormal prolonged depolarization with repetitive spiking characteristic of neurons in epileptic cortical zones that are reflected as interictal discharges in the electroencephalogram.

METABOTROPIC RECEPTOR A G-protein-coupled receptor that may indirectly influence the activity of ion channels but does not itself serve as a channel.

GENERALIZED TONIC-CLONIC SEIZURE

A convulsive seizure involving the entire body, usually characterized by muscle rigidity violent rhythmic muscle contractions and loss of consciousness.

PARTIAL SEIZURE
Seizure resulting from a
localized brain disturbance; also
referred to as 'focal seizure'.

SODIUM CHANNEL INACTIVATION
Entry of the sodium channel into a set of states that are distinct from the open and closed states that prevents the channel from reopening until there has been sufficient time for recovery, thereby preventing persistent sodium flux during long depolarizations.

Modulation of the gating of brain sodium channels is believed to account, at least in part, for the ability of several AEDs to protect against generalized tonic-clonic and PARTIAL SEIZURES. These AEDs include phenytoin, lamotrigine, carbamazepine, oxcarbazepine<sup>4,5</sup> and zonisamide<sup>6</sup>, and possibly felbamate<sup>7</sup>, topiramate<sup>8</sup> and valproate<sup>9-13</sup> (TABLE 1). These drugs block high-frequency repetitive spike firing, which is believed to occur during the spread of seizure activity, without affecting ordinary ongoing neural activity (FIG. 1a,b). This accounts for their ability to protect against seizures without causing a generalized impairment of brain function. At hyper-polarized membrane potentials, clinically relevant concentrations of sodium channel-blocking AEDs such as phenytoin, carabamazepine and lamotrigine block sodium channels only weakly<sup>13-17</sup>. However, when the membrane is depolarized, there is a marked increase in the degree of tonic inhibition (FIG. 1c). Moreover, the inhibitory potency is strongly 'use-dependent', so the block accumulates with prolonged or repetitive activation (FIG. 1d). These properties are explained by preferential binding of the drugs to inactivated conformations of the channel. These agents act mainly on action potential firing; the drugs do not directly alter excitatory or inhibitory synaptic responses. However, the effect on action potentials translates into reduced transmitter output at synapses, and glutamate release might be inhibited more strongly than that of other neurotransmitters, including GABA<sup>18,19</sup>

In recent years, the mechanism of SODIUM CHANNEL INACTIVATION has been elucidated and this has provided an opportunity to clarify the way in which phenytoin and other sodium channel-blocking AEDs promote channel inactivation. The normal predominant (fast) inactivation process results from occlusion of the intracellular mouth of the channel by a short loop of amino-acid residues between domains III and IV of the sodium channel α-subunit that serves as a 'hinged lid'20 (FIG. 2). An additional inactivation process, referred to as 'slow inactivation', begins to come into play with more prolonged depolarizations, such as might occur in association with epileptiform activity<sup>21</sup>. Whereas fast sodium channel inactivation is characterized by rapid (millisecond timescale) onset and recovery, leading to changes in the available pool of sodium channels over the time course of a single action potential, onset and recovery of slow inactivation require several seconds. Slow inactivation contributes to slow spike-frequency adaptation and to the termination of action potential bursts occurring as a result of prolonged or repetitive neuronal depolarization.

The molecular rearrangement that leads to slow inactivation is unknown, but it is distinct from that of fast inactivation. Single channel recordings show that phenytoin does not alter the conductance of the open state of sodium channels<sup>22</sup>. Phenytoin also does not seem to 'stabilize' the normal process of fast inactivation of the sodium channel; in fact, the drug can block

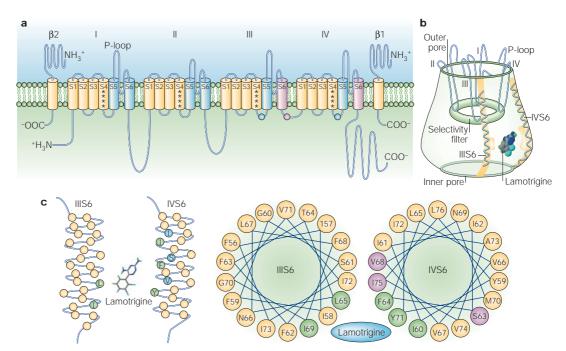


Figure 2 | **Antiepileptic drugs (AEDs) and sodium channels. a** | Primary structures of the subunits of brain type II voltage-gated sodium channels. The main  $\alpha$ -subunit, consisting of four homologous repeats (I–IV), is shown flanked by the two auxiliary  $\beta$ -subunits. Cylinders represent probable  $\alpha$ -helical transmembrane segments. Blue  $\alpha$ -helical segments (S5, S6) form the pore region. Asterisks, S4 voltage sensor; pink circle, inactivation particle in inactivation gate loop ('tethered pore blocker'); blue circles, sites implicated in forming the receptor for the inactivation gate; IIIS6 and IVS6 (pink segments) are regions of modulatory drug binding, including sodium channel-blocking AEDs. **b** | Schematic illustration of the sodium channel pore. The S5 and S6 transmembrane  $\alpha$ -helical segments from each homologous repeat (I–IV) form the four walls of the pore. The outer pore mouth and ion selectivity filter are formed by re-entrant P-loops. The key  $\alpha$ -helical S6 segments in repeat III and IV, which contain the AED binding sites, are highlighted in yellow. A lamotrigine molecule is illustrated in association with its binding sites. **c** | Amino-acid residues that constitute the lamotrigine binding site in transmembrane segments IIIS6 and IVS6. Left, extended view of the  $\alpha$ -helical segments. Substitution of the green residues by alanine results in greater than fivefold reduction in lamotrigine binding affinity for the inactivated state of the channel; substitution of blue residues reduces binding by two- to fivefold. Lamotrigine is shown within the channel pore. Right, axial view (helical wheel representation) of amino-acid side chains according to ideal  $\alpha$ -helix with 3.6 residues per turn. Modified, with permission, from REF. 35 © (2001) American Society for Biochemistry and Molecular Biology.

enzymatically treated channels that lack fast inactivation<sup>23</sup>. Rather, phenytoin induces a non-conducting state of the channel that is similar to channel inactivation. Recovery from drug block of the channel occurs much more slowly than does recovery from block by the intrinsic pore blocker<sup>23</sup>. This, in part, accounts for phenytoin's selective ability to block high-frequency firing: when recovery is slow, the block can accumulate during repetitive activation of the channel.

Interestingly, sodium channel mutations that are associated with accelerated recovery from inactivation and increased sodium channel activity (producing a 'gain-of-function') can lead to enhanced seizure susceptibility, as in the epilepsy syndrome GEFS+ type 2 (REF. 24). However, the syndrome is associated with a mutation (T875M) that paradoxically promotes slow inactivation<sup>25</sup>. Computational modelling, with the assumption that recovery from slow inactivation is also promoted, has shown that the mutation can be associated with an enhanced propensity to fire repetitive action potentials<sup>26</sup>. Alternatively, it is conceivable that the mutation does reduce excitability and that the epileptic phenotype results because the firing of inhibitory neurons is affected. It is notable that pharmacologically induced impairment

of sodium channel inactivation with AEDs such as phenytoin can also promote seizure activity in some circumstances<sup>27,28</sup>; whether this occurs as a result of effects on inhibitory neurons remains to be determined.

Another important feature of the block of sodium channels by phenytoin and lamotrigine (and one that distinguishes these drugs from local anaesthetics) is its slow onset<sup>29,30</sup>. However, once binding occurs it is tight, and unbinding (recovery from block) is slow. Slow binding has two important implications. First, it implies that the time course of sodium currents are not altered in the presence of the drug and therefore the kinetic properties of normal action potentials are not perturbed. Second, slow binding means that inhibition of action potentials does not occur with firing that has been induced by synaptic depolarizations of ordinary length. Rather, long depolarizations are required, possibly as long as a few seconds or more. In focal epilepsies, the cellular events that characterize ICTAL discharges are sustained depolarizations that evoke intermittent high-frequency bursts of action potentials. Such depolarizations provide the conditions required for drug binding and block. On the other hand, cortical interictal discharges are shorter and their frequency is generally not reduced by sodium

GEFS+

A pleotropic autosomal dominant epilepsy syndrome in which there are febrile seizures in childhood and afebrile generalized seizures that persist beyond 6 years of age, including absences, myoclonic seizures, atonic seizures and myoclonic-astatic seizures. The syndrome is distinct from common benign febrile seizures.

ICTAL Relating to an epileptic seizure.

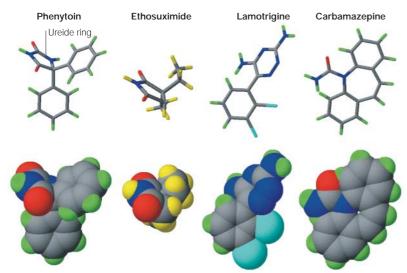


Figure 3 | Stick and space-filling views showing diphenyl moiety common to the sodiumchannel-blocking antiepileptic drugs phenytoin, lamotrigine and carbamazepine. Although ethosuximide has a heterocyclic five-member ring comparable to the ureide ring of phenytoin, it is not a diphenyl structure and does not block fast sodium currents.

channel-blocking AEDs (except lamotrigine)<sup>31</sup>. Overall, sodium channel-blocking AEDs are expected to have little effect on the physiological generation of action potentials in neurons at normal resting potential levels. The voltage-and use-dependent characteristics of the block would come into play only during pathological, sustained depolarizing events associated with high-frequency discharges, therefore allowing protection against seizures without interference with normal function.

Studies using mixtures of phenytoin, carbamazepine and lamotrigine have revealed that these drugs bind to a common recognition site on sodium channels32. Although these three compounds are structurally dissimilar, they do contain a common motif of two phenyl groups separated by one or two C-C or C-N single bonds (1.5–3 Å) (FIG. 3). These two phenyl groups are probably crucial binding elements. Mutational analysis has provided evidence for the location of the AED binding domain. As illustrated in FIG. 2, the S5 and S6 segments are believed to form the lining of the sodium channel pore. Specific phenylalanine (F1764) and tyrosine (Y1771) residues in the S6 segment of domain IV are crucial for use-dependent block by phenytoin and lamotrigine<sup>33,34</sup>. Indeed, adjacent residues spread across 160° of the circumference of the IVS6 segment that flanks F1764 (FIG. 2c) are all relevant to the interaction. When the sodium channel is inactivated, the structure of the channel pore is altered, making the interaction with these residues more favourable. Channel gating is believed to be associated with rotation of the segment containing F1764, Y1771 and the surrounding amino acids so that these residues are brought into the pore, thereby facilitating drug binding. In addition, mutational analysis has revealed that the pore-lining residues leucine 1465 and isoleucine 1469 in IIIS6 also form a portion of the high-affinity binding site for sodium channel-blocking AEDs35. The aromatic rings in

the antiepileptic molecules might interact with the aromatic side chains of the crucial residues in the IVS6 segment or with the nonpolar side chains of the IIIS6 residues. It is noteworthy that this region of the sodium channel is also implicated in inactivation gating<sup>36</sup>.

In addition to effects on the fast sodium current that is responsible for action potentials, AEDs might also act by blocking the Persistent Sodium Current. The persistent sodium current is ascribed to the current that flows as a result of the overlap between the voltage ranges for sodium channel activation and inactivation ('window' current)<sup>37</sup> and probably arises from alternate gating of the channels that are responsible for fast sodium currents38, although certain sodium channel isoforms, most notably Na. 1.6, exhibit greater non-inactivating current and probably contribute more significantly to the persistent current<sup>37,39</sup>. The current carried by the persistent openings is a minute fraction (0.7-4%) of the fast current. However, this current might have a key role in regulating excitability near firing threshold because it is largely unopposed by other voltage-activated currents in this range of membrane potentials. Moreover, there is evidence that the persistent sodium current contributes to the initiation and maintenance of epileptiform activity<sup>40</sup>. Several authors have reported that phenytoin<sup>40–43</sup>, valproate<sup>11</sup> and topiramate<sup>8</sup> inhibit the persistent sodium current at concentrations lower than those that block fast sodium current. The selective reduction of late, persistent sodium channel openings might contribute to the ability of these drugs to protect against seizures with minimal interference in normal function.

Changes in sodium channels that augment the persistent sodium current might contribute to epileptogenesis. In animal models of epileptogenesis, the voltage-dependent properties of sodium channels are altered so that there is greater persistent sodium current <sup>44–46</sup>. This has been attributed to reduced expression of the  $\beta 1$  and  $\beta 2$  auxiliary sodium channel subunits, which is predicted to lead to a depolarizing shift in sodium channel inactivation and greater overlap between the activation and inactivation curves so that the window current is amplified <sup>47</sup>. Moreover, sodium channel mutations that are associated with epilepsy in mice and humans (GEFS+) have been found to enhance the persistent sodium current <sup>48</sup>.

Voltage-gated calcium channels. Voltage-gated calcium channels, like sodium channels, are multisubunit protein complexes that permit ion flux when they are gated open by membrane depolarization<sup>49</sup>. Calcium channels are broadly grouped into high voltageactivated (HVA) and low voltage-activated families. HVA channels — which are further subgrouped as L-, R-, P/Q- and N-types — require strong membrane depolarization for gating and are largely responsible for the regulation of calcium entry and neurotransmitter release from presynaptic nerve terminals. HVA calcium channels represent potential AED targets, as blockade of these channels inhibits neurotransmitter release<sup>50</sup>. These channels consist of an  $\alpha 1$  protein (encoded by one of seven genes) that forms the channel pore and voltage sensor, along with several auxiliary subunits.

INTERICTAL DISCHARGES
Distinctive waves or complexes
that can be recorded between
seizures in the
electroencephalogram of
individuals with epilepsy.
Generally brief in duration, but
can have various morphologies
described as 'sharp wave,' 'spike'
or 'spike-and-slow-wave'.

PERSISTENT SODIUM CURRENT A small component of the sodium current that does not inactivate.

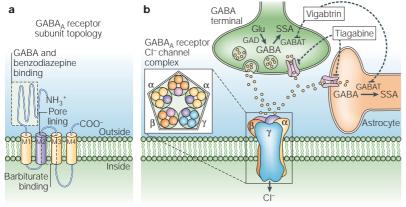


Figure 4 | GABA receptors and antiepileptic drugs. a | Membrane topology of GABA, ( $\gamma$ -aminobutyric acid, type A) receptor subunits. Residues identified as essential for agonist (GABA) and benzodiazepine binding are in the extracellular amino terminus. Barbiturates bind to the membrane-spanning M2 and M3 segments. b | Inhibitory GABA synapse illustrating synthesis of GABA in a presynaptic terminal from glutamate through the GABA-shunt of the tricarboxylic acid (Krebs) cycle by glutamate decarboxylase (GAD). In neurons and astrocytes, GABA is degraded by GABA transaminase (GABAT) to succinic semialdehyde (SSA); vigabatrin is an irreversible inhibitor of GABAT. Cylinders represent uptake transporters, which are inhibited by tiagabine. The pentameric structure of GABA $_{\rm A}$  receptors is represented to the left. Small circles indicate membrane-spanning  $\alpha$ -helical segments corresponding to those shown in  $\alpha$ . M2 segments form the CI- channel pore.

Of particular interest from the perspective of AED mechanisms are the  $\alpha 2\delta$ -1 and  $\alpha 2\delta$ -2 auxiliary subunits, which bind the AED gabapentin and its analogues with high affinity<sup>51,52</sup>. Gabapentin, the lipophilic 3-cycylohexyl analogue of GABA, was originally synthesized in an attempt to develop a brain-penetrant GABA agonist. However, gabapentin lacks activity at GABA, receptors. Rather, the α2δ-binding affinities of gabapentin and analogues such as pregabalin correlate in a stereoselective fashion with their antiepileptic potency, strongly implicating these subunits as a relevant target<sup>53,54</sup>. The α2δ-subunits are highly glycosylated products of a single gene that is post-translationally cleaved into  $\alpha$ 2- and  $\delta$ -peptides, which are then covalently linked by a disulphide bridge.  $\alpha 2\delta$ -1 and  $\alpha 2\delta$ -2 are believed to form complexes with many calcium channel types (represented by different α1 isoforms), allosterically enhancing current amplitude and also promoting channel trafficking to the membrane. In particular, the auxiliary subunits markedly augment the current through P/Q-type calcium channels (formed from the  $\alpha 1A/Ca_{u}2.1$  principal calcium channel subunit)55,56. Mutagenesis experiments have shown that the gabapentin binding site is probably confined to the  $\alpha 2$  protein and the external (rather than the transmembrane or internal) portion of the associated  $\delta$  component<sup>57,58</sup>.

The functional consequences of gabapentin binding to  $\alpha2\delta\text{-}1$  and  $\alpha2\delta\text{-}2$  are still not fully defined. Initial studies of the effect of gabapentin on neuronal calcium currents yielded inconsistent effects. However, several recent studies with dorsal root ganglion neurons have confirmed that the drug inhibits HVA calcium current in a concentration-dependent fashion at clinically relevant concentrations ( $\sim\!10\text{--}100\,\mu\text{M})$ , although the effect is relatively small $^{59\text{--}61}$ . Nevertheless, the effect on

calcium current probably translates into a reduction in excitatory transmission, as shown by inhibition of potassium-evoked glutamate release from brain slices<sup>62</sup> and suppression of excitatory postsynaptic currents, at least in spinal dorsal horn neurons<sup>63</sup>. Interestingly, increased expression of  $\alpha 2\delta$ -1 in pain-sensitive dorsal root ganglion neurons and in the dorsal spinal cord might contribute to the hyperexcitability of pain pathways in neuropathic pain syndromes, and the ability of gabapentin to inhibit excitatory synaptic responses in the spinal cord is potentiated under such pathological circumstances<sup>1</sup>. It will be of interest to determine whether changes in subunit expression could similarly occur in epilepsy and affect gabapentin responsiveness. Although the  $\alpha 2\delta$  subunit is at present the most plausible target for gabapentin, the drug has been found to increase GABA synthesis and turnover<sup>64</sup>, and it seems to elevate GABA levels in the human brain, possibly through competitive inhibition of the system-L branched-chain-amino acid transporter or through reversal of the GABA-uptake transporter<sup>65,66</sup>.

In addition to gabapentin, several other AEDs can, in part, act through inhibition of HVA calcium channels. For example, phenobarbital can block these channels, but its effects on  $\mathsf{GABA}_A$  receptors are probably more important in its antiepileptic activity  $^{67}$ . Lamotrigine can also inhibit HVA (N- and P/Q-type) calcium channels  $^{68,69}$ ; it does not affect low voltage-activated T-type calcium channels  $^{70}$ . Similarly, levetiracetam at clinically relevant concentrations has been reported to produce a small inhibition of calcium current  $^{71}$ , with a predominant affect on N-type channels  $^{72}$ .

In contrast to HVA calcium channels, whose main function is regulation of neurotransmitter release, low voltage-activated (T-type) calcium channels are believed to regulate neuronal firing by participating in bursting and intrinsic oscillations<sup>73</sup>. In the thalamus, these channels are crucial for the abnormal oscillatory behaviour that underlies generalized absence seizures<sup>74</sup>. In thalamic relay neurons and thalamic reticular neurons — GABA interneurons that modulate and synchronize thalamic output — T-type calcium channels generate 'lowthreshold calcium spikes', which trigger a burst of action potentials mediated by sodium channels<sup>75</sup>. The T-type calcium channel family (Ca<sub>.</sub>3) consists of the  $\alpha$ 1G,  $\alpha$ 1H and α1I subunits, which are ~30% homologous to HVA subunits in their putative membrane-spanning regions. Thalamic relay neurons express high levels of α1G subunits, whereas thalamic reticular neurons express high levels of  $\alpha 1I$  and moderate levels of  $\alpha 1H$  subunits<sup>76</sup>. Thalamic neurons from transgenic mice that lack  $\alpha 1G$ subunits fail to fire in burst mode and the mice are resistant to pharmacologically induced absence seizures, strongly supporting a role for thalamic neuron bursting mediated by T-type calcium channels in absence epilepsy<sup>77</sup>. Conversely, T-type calcium currents are substantially larger and Ca, 3 mRNA expression is greater in thalamic reticular neurons in a rat strain that exhibits absence seizures (the genetic absence epilepsy rat from Strasbourg or GAERS) than in seizure-free control rats<sup>78,79</sup>. Recently, 12 missense mutations were found in

558 JULY 2004 VOLUME 5 www.nature.com/reviews/neuro

POLYGENIC

A characteristic that is controlled by different genes, each of which has only a small role in the phenotype.

UREIDE RING Heterocyclic ring formed from urea ( $CO(NH_2)_2$ ) as in phenytoin and barbiturates. In ethosuximide, the hydantoin ureide ring of phenytoin is substituted for a succinimide ring with a single nitrogen atom.

IONOTROPIC RECEPTOR Possessing an intrinsic channel (pore) that mediates transmembrane ion flux.

MYOCLONIC SEIZURE Sudden, brief, involuntary spasm (contraction-relaxation) of the tongue or muscles of the face, trunk, arms, legs or the entire body.

DRAVET'S SYNDROME
A rare intractable epilepsy
syndrome in which prolonged
generalized tonic, clonic or
tonic-clonic seizures occur
between 2 and 9 months of age
followed by myoclonic, tonicclonic, absence and partial
seizures in the second year of
life. The syndrome is associated
with delayed psychomotor and
speech development, and ataxia

JUVENILE MYOCLONIC EPILEPSY A common generalized epilepsy syndrome presenting between the ages of 8 and 26 years with early morning myoclonus, mainly affecting the upper extremities, and often associated with generalized tonic-clonic seizures and less frequently with absence seizures.

STATUS EPILEPTICUS Continuous seizure activity without recovery of consciousness or return to neurological function. the  $\alpha 1H$  gene CACNA1H in 14 of 118 patients with childhood absense epilepsy but not in 230 unrelated controls80. Khosravani et al.81 induced five of these mutations in the rat α1H (Ca, 3.2) T-type channel homologue and found that three were associated with significant gain-of-function effects on T-type channel activity that would predispose to hyperexcitability, including shifts of the activation voltage closer to the resting potential and of the inactivation voltage towards more depolarized potentials (so that there is increased channel availability for opening at the resting potential). Although these alterations in channel gating are compatible with the clinical phenotype, childhood absence epilepsy is likely to be POLYGENIC in origin so that variations in CACNA1H might only partially account for the disorder<sup>82</sup>.

Ethosuximide, an AED that is highly effective in the treatment of absence seizures but is ineffective in other seizure types, inhibits T-type (but not HVA) calcium channels in thalamic neurons<sup>83</sup>. This blocking action has been confirmed for all three cloned human T-type calcium channel types that are expressed in heterologous cells84. The potency is low so that at clinically relevant concentrations only a small fraction of channels are blocked. Nevertheless, the magnitude of the effect seems to be sufficient to reduce the amplitude of low-threshold spikes to an extent that prevents bursting and absence seizure activity. Ethosuximide does not block fast voltage-gated sodium currents at clinically relevant concentrations<sup>85,86</sup> and as a result it has no activity against partial or generalized tonic-clonic seizures (TABLE 1). Indeed, although it shares the UREIDE RING-like structure with phenytoin, it does not have the crucial phenyl groups that seem to be necessary for blockade of fast sodium currents (FIG. 3). Although it is now generally accepted that ethosuximide affects T-type calcium channels in thalamic neurons, blockade of other ionic currents, including the persistent sodium current, might also be relevant86 (but see REF. 43) and, at least in animal models, it seems that ethosuximide might also interact with neocortical regions that trigger absence seizure activity87. It has been reported that zonisamide, which might have activity against absence seizures (TABLE 1), also blocks T-type calcium channels88. Lamotrigine is a highly effective antiabsence drug that, as noted above, does not affect T-type calcium channels; the mechanism by which it protects in this seizure type is a mystery.

*GABA systems.* Potentiation of inhibitory neurotransmission mediated by GABA is a key mechanism of AED action. Neurons that use GABA as their neurotransmitter represent only a small fraction of neurons in regions that are key to epileptic activity, such as the neocortex, hippocampus and amygdala, and in these regions excitatory synapses can be several-fold more common than inhibitory ones<sup>89</sup>. However, these inhibitory connections are vital in restraining the natural tendency of recurrently connected excitatory neurons to undergo the transition through positive feedback into synchronized epileptiform discharges<sup>90</sup>.

GABA acts through fast chloride-permeable IONOTROPIC GABA, receptors and also through slower metabotropic G-protein-coupled GABA<sub>R</sub> receptors. Reduction in the efficacy of synaptic inhibition mediated by GABA, receptors — for example, with drugs that block  $\text{GABA}_{\scriptscriptstyle{A}}$  receptors, such as bicuculline and pentylenetetrazol — can lead to seizures. Moreover, mutations in the GABA, receptor  $\gamma$ 2 subunit (GABRG2) have been associated with various epilepsy syndromes including GEFS+ type 3, childhood absence epilepsy and severe MYOCLONIC epilepsy of infancy (DRAVET'S SYNDROME) 91-94 (TABLE 2). These mutations result in reduced GABA, receptor-mediated inhibitory function by various mechanisms including reduced surface expression of functional receptors and altered kinetics<sup>95</sup>. Similarly, a mutation in the  $\alpha$ 1 subunit (GABRA1) that markedly reduces the function of heteromeric GABA receptors containing the subunit has been associated with autosomal dominant Juvenile MYOCLONIC EPILEPSY in one family 96. Conversely, pharmacological enhancement of GABA, receptor-mediated inhibition is an effective antiepileptic approach. Indeed, bromide, the first effective epilepsy treatment, augments GABA, receptor-mediated inhibition by enhancing the sensitivity of GABA, receptors to GABA and increasing GABA, receptor currents owing to the threefold increase in permeability of Br- over Cl-(REFS 97-99). Although it has long been supplanted in clinical epilepsy therapy by less toxic agents, bromide salts continue to be widely used for treating epileptic dogs and cats in veterinary medicine. Many modern AEDs influence GABA, receptor inhibition (TABLE 1), either by interacting with GABA, receptors or by modifying the activity of enzymes and transporters so as to alter the dynamics of GABA. Drugs that act through these mechanisms typically have a broad spectrum of antiepileptic activity in human seizure disorders, although, with the exception of benzodiazepine receptor agonists, they are generally ineffective in absence seizures. Valproate and gabapentin increase GABA synthesis and turnover, and valproate and to a lesser extent gabapentin have a range of activities that overlaps those of drugs that are known to interact with GABA systems.

The antiepileptic activity of benzodiazepine-like agents occurs through positive allosteric modulation of  $GABA_{A}$  receptors containing the  $\gamma 2$  subunit, which is necessary for benzodiazepine modulation. GABA, receptors containing the α1 subunit are an important target for protection against seizures by benzodiazepine 100,101. The effects of benzodiazepines on tonic GABA receptor currents, which originate from GABA acting on extrasynaptic receptors, might be particularly important in their action<sup>102</sup>. In clinical practice, benzodiazepines have an important role in the acute treatment of status epilepticus. However, their use in chronic therapy is limited by the sedation and muscle relaxation that occurs at doses that are comparable to those that protect against seizures and, more importantly (because the side effects diminish over time), by the development of tolerance and dependence.

Table 2 | Ion channel genes implicated in human epilepsy syndromes Gene Functional role of expressed protein Epilepsy syndrome Voltage-gated ion channels SCN1A\* Neuronal (type I) voltage-gated sodium channel, GEFS+ type 2; severe myoclonic epilepsy of α-subunit (Na.1.1) SCN2A\* Neuronal (type II) voltage-gated sodium channel, Febrile seizures associated with afebrile seizures; α-subunit (Na. 1.2) benign familial neonatal-infantile seizures SCN1B\* Voltage-gated sodium channel, β-subunit GEFS+ type 1 T-type voltage-gated calcium channel CACNA1H\* Childhood absence epilepsy α1H-subunit (Ca, 3.2) KCNQ2\* M-type potassium channel subunit Benign neonatal epilepsy, type 1; myokymia with neonatal epilepsy KCNQ3\* M-type potassium channel subunit Benign neonatal epilepsy, type 2 KCNA1 K, 1.1 (Shaker-like) voltage-gated potassium Episodic ataxia/myokymia, type 1, sometimes with channel subunit partial seizures CLCN2 Voltage-gated chloride channel CIC2 Childhood absence epilepsy, type 3; juvenile absence epilepsy; epilepsy with grand mal seizures on awakening Neurotransmitter-gated ion channels GABRA1\* GABA, receptor, α1-subunit Juvenile myoclonic epilepsy GABRG2\* GABA, receptor, y2-subunit GEFS+ type 3; childhood absence epilepsy, type 2 CHRNA4 Neuronal nicotinic acetylcholine receptor, Nocturnal frontal lobe epilepsy, type 1 α4-subunit CHRNB2 Neuronal nicotinic acetylcholine receptor, Nocturnal frontal lobe epilepsy, type 3 **B**-subunit

Hypersynchronous activity in thalamocortical circuits is believed to underlie the 3-Hz spike-and-wave activity that is characteristic of generalized absence seizures. The anti-absence activity of benzodiazepines, such as clonazepam, probably results from their ability to 'desynchronize' these oscillations  $^{103}$ . This occurs through the enhancement of mutual inhibition in the thalamic reticular nucleus by effects on benzodiazepine-sensitive  $\alpha 3$ -containing GABA receptors. Thalamic reticular neurons exert an inhibitory influence on thalamocortical relay neurons that is necessary for de-inactivation of the T-type calcium currents that underlie bursting. Benzodiazepines reduce the inhibitory output of the reticular neurons and therefore prevent absence seizure activity.

Barbiturates such as phenobarbital also act as positive allosteric modulators of GABA<sub>A</sub> receptors, but with a different mode of action from benzodiazepines. At clinically relevant concentrations, phenobarbital does not increase the frequency of GABA-induced channel openings, but rather shifts the relative proportion of openings to favour the longest-lived open state associated with prolonged bursting (brief openings in rapid succession), thereby increasing the overall probability that the channel is open<sup>104</sup>. In addition, barbiturates act on other ion channel systems, including calcium and sodium channels, and this probably contributes to their therapeutic activity and might also be a factor in

side effects $^{67}$ . There is evidence that felbamate $^{105}$  and topiramate $^{106,107}$  might also act, in part, through positive modulation of GABA $_{\Lambda}$  receptors (TABLE 1).

The concentration of GABA in the brain is controlled by two pyridoxal-5'-phosphate-dependent enzymes, glutamate decarboxylase (GAD) and GABA transaminase (GABAT). The AED vigabatrin (γ-vinyl GABA) is a GABA analogue that acts as an irreversible suicide INHIBITOR of GABAT<sup>108</sup>. The drug initially binds reversibly to the pyridoxal-5'-phosphate cofactor and then binds irreversibly to the enzyme. Administration of vigabatrin leads to large elevations in brain GABA levels<sup>109</sup>. The antiepileptic properties of vigabatrin have been attributed to enhanced GABA-mediated inhibition. Paradoxically, vigabatrin does not lead to larger GABA, receptor-mediated synaptic responses, and it has generally been found to inhibit spontaneous and evoked synaptic GABA currents<sup>110-112</sup>. Rather, the drug probably acts by enhancing tonic GABA-mediated inhibition, owing to an increase in ambient GABA that is attributed either to a reversal of GABA transporters or to a shift in the equilibrium for GABA that favours increased extracellular GABA as a result of the marked elevation in intracellular GABA<sup>113</sup> or of effects on GABA uptake that are unrelated to GABAT<sup>114</sup>. The lack of strong sedation produced by vigabatrin is difficult to reconcile with this mechanism, which would be expected to induce a generalized enhancement of inhibitory tone. In this

IRREVERSIBLE SUICIDE
INHIBITOR
An inhibitor that is inactive until acted upon by the enzyme: the inhibitor binds to the enzyme as a substrate and a chemically reactive intermediate is generated that inactivates the enzyme.

regard it is notable that the elevation in extracellular GABA that is produced by vigabatrin might alter the dynamics of inhibitory synaptic function so as to reduce the fading of inhibitory mechanisms during repetitive activation of interneurons<sup>110</sup>. Such fading, which is dependent on GABA<sub>B</sub> receptors, is believed to be an important factor that permits focal epileptiform activity to develop into a full-blown seizure. Interference with this process could account for the selective suppression of seizures by vigabatrin. In support of a role for GABA<sub>B</sub> receptors in limiting epileptic activity, mice lacking functional GABA<sub>B</sub> receptors exhibit spontaneous seizures<sup>115</sup>.

The synaptic action of GABA is terminated by rapid reuptake into presynaptic terminals and surrounding glia by high-affinity plasma membrane GABA transporters, including GAT1, the most abundant of the transporters<sup>116</sup>. The AED tiagabine is a potent and selective competitive inhibitor of GAT1 that binds with high affinity to the transporter and prevents GABA uptake without itself being transported<sup>117</sup>. By slowing the reuptake of synaptically released GABA, tiagabine prolongs inhibitory postsynaptic potentials<sup>118–120</sup>. This action can be enhanced with repetitive activation, as is expected to occur during the synchronous discharge of interneurons associated with epileptic activity, so that the behavioural depression that would accompany indiscriminate enhancement of GABA inhibition is minimized.

Glutamate receptors. Ionotropic glutamate receptors are glutamate-gated cation channels that mediate the bulk of fast excitatory neurotransmission in the CNS<sup>121</sup>. Blockade of the NMDA and AMPA subtypes of these receptors protects against seizures in in vitro and in vivo models<sup>122</sup>; there is emerging evidence that kainate receptors are a potential antiepileptic target 123. Despite the voluminous preclinical literature, clinical trials with selective NMDA antagonists in the chronic treatment of epilepsy have been disappointing, although there are case reports that ketamine, a channelblocking NMDA receptor antagonist, might be useful in the treatment of refractory status epilepticus<sup>124–126</sup>. Nevertheless, several marketed AEDs might, at least in part, act through effects on ionotropic glutamate receptors. For example, felbamate, among other actions, inhibits NMDA receptors at clinically relevant concentrations<sup>127,128</sup>. NMDA receptors in mammalian neurons are hetero-oligomers formed by co-assembly of an obligatory NR1 subunit and at least one type of NR2 subunit. Felbamate is modestly more potent as an antagonist of NMDA receptors containing NR2B subunits<sup>129,130</sup>. Unlike the NR2A subunit, which is distributed ubiquitously in the CNS, expression of the NR2B subunit in the adult is largely restricted to the forebrain. NR2B selectivity could in part contribute to felbamate's low neurobehavioural toxicity in relation to other NMDA receptor antagonists, as the drug might target NMDA receptor-mediated synaptic transmission in forebrain areas that are necessary for seizure generation and avoid perturbing non-forebrain structures that

could mediate side effects. In addition, NR2B subunits are abundantly expressed in the immature brain and this could also account for felbamate's clinical utility in childhood seizure disorders such as the LENNOX-GASTAUT SYNDROME (TABLE 1).

AMPA receptors are key mediators of seizure spread and could have a role in seizure-induced brain damage<sup>122</sup>. They are therefore potentially important AED targets. Selective AMPA receptor antagonists are under investigation, but no currently marketed AED acts predominantly through this target. Like AMPA and NMDA receptors, kainate receptors are responsible for a portion of glutamate-mediated excitation at some synapses, including those in limbic regions that are relevant to epilepsy<sup>131</sup>. As well as contributing to postsynaptic excitation, kainate receptors on presynaptic axon terminals modulate glutamate release from excitatory afferents and they also have the unique property of suppressing GABA release from inhibitory interneurons<sup>132</sup>. The combination of postsynaptic excitation and suppression of inhibition endow kainate receptors with a unique potential to induce epileptic activity<sup>123</sup>. Moreover, selective kainate receptor antagonists can protect against seizures in brain slice and animal models<sup>133</sup>. So, kainate receptors are potential AED targets. Among its complex actions, topiramate selectively inhibits kainate receptors and, to a lesser extent, AMPA receptors<sup>134,135</sup>.

#### Perspective

The modern AED era — spanning a period of 150 years from the first use of bromide to the present day — has seen the introduction into clinical practice of a diverse group of effective and safe drugs that have provided immeasurable benefits for those afflicted with seizure disorders of all kinds. With only a few exceptions, these drugs were discovered by screening in animal models, and the underlying mechanisms are only now coming into clear focus. Remarkably, the set of molecular targets (TABLE 1) overlaps the rapidly enlarging roster of 'epilepsy genes' that have been identified by molecular genetics approaches (TABLE 2). Moreover, the way AEDs act to modulate the activity of these brain targets is often opposite to the functional defect in a genetic epilepsy syndrome. So, AEDs often act on ion channels to subtly reduce excitability whereas the mutations often cause correspondingly subtle gains-of-function in the same ion channels. This illustrates the pivotal importance of ion channels in epilepsy and indicates that inevitable additions to the rapidly enlarging list of ion channel epilepsy genes might provide promising new targets for the rational development of AEDs.

In the end, AEDs are only a step on the way to the ultimate goal of epilepsy medicine, which is to provide treatments that prevent epilepsy or reverse it. ANTIEPILEPTOGENIC Strategies are under development using various animal models of chronic epilepsy<sup>136,137</sup>. By targeting plasticity mechanisms that underlie the enhanced seizure susceptibility that often follows brain insults such as head trauma, status epilepticus or neonatal hypoxia, antiepileptogenic drugs of the future would prevent, or reverse, progressive worsening of the epileptic process.

LENNOX-GASTAUT SYNDROME A devastating paediatric epilepsy syndrome usually beginning between ages 1 to 8 years. It is characterized by multiple seizure types including tonic, atonic, atypical absence and myoclonic seizures. There is often impaired intellectual functioning and behavioural disturbances.

ANTIEPILEPTOGENIC Protection against the development of epilepsy, a state characterized by recurrent seizures. It is likely that antiepileptogenic drugs will have mechanisms of action that are distinct from traditional AEDs, as the molecular mechanisms that underlie epileptogenesis and ictogenesis probably differ. As in the case of symptomatic epilepsy therapies that have found enormous utility for indications other than epilepsy<sup>1</sup>, it can be expected that such antiepileptogenic approaches will be applied to the prevention of various progressive neurological disorders, such as neuropathic pain, and perhaps important psychiatric disorders as well.

- Rogawski, M. A. & Löscher, W. The neurobiology of antiepileptic drugs for the treatment of nonepileptic conditions. *Nature Med.* (in the press).
   A review of the actions of AEDs in conditions other than epilepsy, focusing on disease-related ion channel plasticity and long-term effects on signalling
- pathways.
   Tatulian, L., Delmas, P., Abogadie, F. C. & Brown, D. A. Activation of expressed KCNQ potassium currents and native neuronal M-type potassium currents by the anticonvulsant drug retigabine. *J. Neurosci.* 21, 5535–5545
- Rogawski, M. A. in Neurotherapeutics: Emerging Strategies (eds Pullan, L. & Patel, J.) 193–273 (Humana, Totowa, New Jersey, 1996).
- Schmutz, M., Brugger, F., Gentsch, C., McLean, M. J. & Olpe, H. R. Oxcarbazepine: preclinical anticonvulsant profile and putative mechanisms of action. *Epilepsia* 35 (Suppl. 5), 547–50 (1994)
- Ambrósio, A. F. et al. Inhibition of glutamate release by BIA 2-093 and BIA 2-024, two novel derivatives of carbamazepine, due to blockade of sodium but not calcium channels. Biochem. Pharmacol. 61, 1271–1275 (2001).
- Schauf, C. L. Zonisamide enhances slow sodium inactivation in *Myxicola*. *Brain Res.* 413, 185–188 (1987).
- Taglialatela, M., Öngini, E., Brown, A. M., Di Renzo, G. & Annunziato, L. Felbamate inhibits cloned voltage-dependent Na<sup>+</sup> channels from human and rat brain. *Eur. J. Pharmacol.* 316, 373–377 (1996).
- Taverna, S., Sancini, G., Mantegazza, M., Franceschetti, S & Avanzini, G. Inhibition of transient and persistent Narcurrent fractions by the new anticonvulsant topiramate. *J. Pharmacol. Exp. Ther.* 288, 960–968 (1999).
- Van den Berg, R. J., Kok, P. & Voskuyl, R. A. Valproate and sodium currents in cultured hippocampal neurons. Exp. Brain. Res. 93, 279–287 (1993).
- Albus, H. & Williamson, R. Electrophysiologic analysis of the actions of valproate on pyramidal neurons in the rat hippocampal slice. *Epilepsia* 39, 124–139 (1998).
- Taverna, S., Mantegazza, M., Franceschetti, S. & Avanzini, G. Valproate selectively reduces the persistent fraction of Nar current in neocortical neurons. *Epilepsy Res.* 32, 304–308 (1998)
- Xie, X. et al. Electrophysiological and pharmacological properties of the human brain type IIA Na\* channel expressed in a stable mammalian cell line. Pflugers Arch. 441, 425–433 (2001).
  - In this paper, studies using human brain type IIA voltage-gated sodium channels stably expressed in CHO cells showed voltage- and use-dependent block by phenytoin and lamotrigine, but not valproate and gabapentin.
- Remy, S., Urban, B. W., Elger, C. E. & Beck, H. Anticonvulsant pharmacology of voltage-gated Na\* channels in hippocampal neurons of control and chronically epileptic rats. Eur. J. Neurosci. 17, 2648–2658 (2003).
- Willow, M., Gonoi, T. & Catterall, W. A. Voltage clamp analysis of the inhibitory actions of diphenylhydantoin and carbamazepine on voltage-sensitive sodium channels in neuroblastoma cells. *Mol. Pharmacol.* 27, 549–558 (1985)
- Matsuki, N., Quandt, F. N., Ten Eick, R. E. & Yeh, J. Z. Characterization of the block of sodium channels by phenytoin in mouse neuroblastoma cells. J. Pharmacol. Exp. Ther. 228, 523–530 (1984).
- Ragsdale, D. S., Scheuer, T. & Catterall, W. A. Frequency and voltage-dependent inhibition of type IIA Na<sup>+</sup> channels, expressed in mammallan cell line, by local anesthetic, antiarrhythmic, and anticonvulsant drugs. *Mol. Pharmacol.* 40, 756–765 (1991).
  - Characterization of the actions of phenytoin and carbamazepine on recombinant rat brain sodium channels.
- Xie, X., Lancaster, B., Peakman, T. & Garthwaite, J. Interaction of the antiepileptic drug lamotrigine with recombinant rat brain type IIA Na<sup>+</sup> channels and with native Na<sup>+</sup> channels in rat hippocampal neurones. *Pflügers Arch.* 430, 437–446 (1995).
- Leach, M. J., Marden, C. M. & Miller, A. A. Pharmacological studies on lamotrigine, a novel potential antiepileptic drug: II.

- Neurochemical studies on the mechanism of action *Epilepsia* **27**, 490–497 (1986).
- Prakriya, M. & Mennerick, S. Selective depression of lowrelease probability excitatory synapses by sodium channel blockers. *Neuron* 26, 671–682 (2000).
- Goldin, A. L. Mechanisms of sodium channel inactivation Curr. Opin. Neurobiol. 13, 284–290 (2003).
- Fleidervish, I. A., Friedman, A. & Gutnick, M. J. Slow inactivation of Na<sup>+</sup> current and slow cumulative spike adaptation in mouse and guinea-pig neocortical neurones in slices. J. Physiol. (Lond.) 493, 83–97 (1996).
- Quandt, F. N. Modification of slow inactivation of single sodium channels by phenytoin in neuroblastoma cells. *Mol. Pharmacol.* 34, 557–565 (1988).
  - Analysis of phenytoin block of single sodium channels in membrane patches.
- 23. Kuo, C. C. & Bean, B. P. Na<sup>+</sup> channels must deactivate to recover from inactivation. Neuron 12, 819–829 (1994). A demonstration that phenytoin stabilizes sodium channels of hippocampal neurons in a nonconducting state, distinct from fast inactivation, from which recovery is slow and requires the channel to deactivate.
- Spampanato, J., Escayg, A., Meisler, M. H. & Goldin, A. L. Functional effects of two voltage-gated sodium channel mutations that cause generalized epilepsy with febrile seizures plus type 2. J. Neurosci. 21, 7481–7490 (2001).
- Alekov, A. K., Rahman, M. M., Mitrovic, N., Lehmann-Horn, F. & Lerche, H. Enhanced inactivation and acceleration of activation of the sodium channel associated with epilepsy in man. Eur. J. Neurosci. 13, 2171–2176 (2001).
- Spampanato, J., Aradi, I., Soltesz, I. & Goldin, A. L. Increased neuronal firing in computer simulations of sodium channel mutations that cause generalized epilepsy with febrile seizures plus. J. Neurophysiol. 91, 2040–2050 (2004)
- Vallaria, J. M., Bell, D. B. & Reichert, A. Progressive encephalopathy due to chronic hydantoin intoxication. *Am. J. Dis. Child.* 128, 27–34 (1974).
- Löscher, W., Cramer, S. & Ebert, U. Limbic epileptogenesis alters the anticonvulsant efficacy of phenytoin in Sprague-Dawley rats. *Epilepsy Res.* 31, 175–186 (1998).
- Kuo, C. C. & Bean, B. P. Slow binding of phenytoin to inactivated sodium channels in rat hippocampal neurons. *Mol. Pharmacol.* 46, 716–725 (1994).
   Phenytoin is found to bind tightly to the fast inactivated state of sodium channels but binding occurs slowly, a key characteristic enabling phenytoin to disrupt epileptic discharges with minimal effects on normal firing activity.
- Kuo, C. C. & Lu, L. Characterization of lamotrigine inhibition of Na\* channels in rat hippocampal neurones. *Br. J. Pharmacol.* 121, 1231–1238 (1997).
- Bazil, C. W. & Pedley, T. A. in Antiepileptic Drugs, 5th Edition (eds Levy, R. H., Mattson, R. H., Meldrum, B. S. & Perucca, E.)
   22–35 (Lippincott Williams & Wilkins, Philadelphia, 2002).
- 32. Kuo, C. C. A common anticonvulsant binding site for phenyloin, carbamazepine, and lamotrigine in neuronal Na channels. *Mol. Pharmacol.* 54, 712–721 (1998). Studies with mixtures of the main sodium channelblocking anticonvulsants demonstrate that these drugs bind to a similar recognition site on the inactivated conformation of the channel.
- Ragsdale, D. S., McPhee, J. C., Scheuer, T. & Catterall, W. A. Common molecular determinants of local anesthetic, antiarrhythmic, and anticonvulsant block of voltage-gated Na\* channels. Proc. Natl Acad. Sci. USA 93, 9270–9275 (1996).
  - Mutational analysis demonstrates the importance of F1764 and Y1771 in sodium channel transmembrane segment IVS6 in the binding of lidocaine and phenytoin.
- Liu, G. et al. Differential interactions of lamotrigine and related drugs with transmembrane segment IVS6 of voltagegated sodium channels. Neuropharmacology 44, 413–422 (2003).
  - Site-directed mutagenesis reveals residues of the IVS6 transmembrane segment of the sodium channel that represent points of interaction for lamotrigine.

- Yarov-Yarovoy, V. et al. Molecular determinants of voltagedependent gating and binding of pore-blocking drugs in transmembrane segment IIIS6 of the Na<sup>+</sup> channel α-subunit. J. Biol. Chem. 276, 20–27 (2001).
   Site-directed mutagenesis reveals residues of the
  - Site-directed mutagenesis reveals residues of the IIIS6 transmembrane segment of the sodium channel that represent points of interaction for lamotrigine and the local anesthetic etidocaine.
- McPhee, J. C., Ragsdale, D. S., Scheuer, T. & Catterall, W. A. A critical role for transmembrane segment IVS6 of the sodium channel α-subunit in fast inactivation. J. Biol. Chem. 270, 12025–12034 (1995).
- Smith, M. R., Smith, R. D., Plummer, N. W., Meisler, M. H. & Goldin, A. L. Functional analysis of the mouse Scn8a sodium channel. *J. Neurosci.* 18, 6093–6102 (1998).
- Taddese, A. & Bean, B. P. Subthreshold sodium current from rapidly inactivating sodium channels drives spontaneous firing of tuberomammillary neurons. *Neuron* 33, 587–600 (2002).
- Maurice, N., Tkatch, T., Meisler, M., Sprunger, L. K. & Surmeier, D. J. D1/D5 dopamine receptor activation differentially modulates rapidly inactivating and persistent sodium currents in prefrontal cortex pyramidal neurons. J. Neurosci. 21, 2268–2277 (2001).
- Segal, M. M. & Douglas, A. F. Late sodium channel openings underlying epileptiform activity are preferentially diminished by the anticonvulsant phenytoin. J. Neurophysiol. 77, 3021–3034 (1977).
  - This paper shows that the tetrodotoxin-sensitive, persistent sodium current that results from rare late openings of voltage-gated sodium channels is more sensitive to phenytoin than is the transient current. Such late openings are hypothesized to be important in initiating and continuing epileptiform activity.
- Chao, T. I. & Alzheimer, C. Effects of phenytoin on the persistent Na<sup>+</sup> current of mammalian CNS neurones. Neuroreport 6, 1778–1780 (1995).
- Lampl, İ., Schwindt, P. & Crill, W. Reduction of cortical pyramidal neuron excitability by the action of phenytoin on persistent Na\* current. J. Pharmacol. Exp. Ther. 284, 228–237 (1998).
- Niespodziany, I., Klitgaard, H. & Georg Margineanu, D. Is the persistent sodium current a specific target of anti-absence drugs? Neuroreport 15, 1049–1052 (2004).
- Ketelaars, S. O., Gorter, J. A., van Vilet, E. A., Lopes da Silva, F. H. & Wadman, W. J. Sodium currents in isolated rat CA1 pyramidal and dentate granule neurones in the poststatus epilepticus model of epilepsy. *Neuroscience* 105, 109–120 (2001)
- Ellerkmann, R. K. et al. Molecular and functional changes in voltage-dependent Nar channels following pilocarpineinduced status epilepticus in rat dentate granule cells. Neuroscience 119, 323–333 (2003).
- Agrawal, N., Alonso, A. & Ragsdale, D. S. Increased persistent sodium currents in rat entorhinal cortex layer V neurons in a post-status epilepticus model of temporal lobe epilepsy. *Epilepsia* 44, 1601–1604 (2003).
- Gastaldi, M., Robaglia-Schlupp, A., Massacrier, A., Planells, R. & Cau, P. mRNA coding for voltage-gated sodium channel \( \beta 2 \) subunit in rat central nervous system: cellular distribution and changes following kainate-induced seizures. Neurosci. Lett. 249, 53–56 (1998).
- Lossin, C., Wang, D. W., Rhodes, T. H., Vanoye, C. G. & George, A. L. Jr. Molecular basis of an inherited epilepsy. Neuron 34, 877–884 (2002).
  - Recombinant human voltage-gated sodium channel SCN1A was coexpressed in cultured mammalian cells with human accessory subunits  $\beta 1$  and  $\beta 2$ . GEFS+ mutations T875M, W1204R and R1648H in SCN1A were found to alter channel inactivation, resulting in persistent inward sodium current. This gain-offunction abnormality is hypothesized to represent the biophysical mechanism that is responsible for the epileptic phenotype.
- Catterall, W. A. Structure and regulation of voltage-gated Ca<sup>2+</sup> channels. Annu. Rev. Cell Dev. Biol. 16, 521–555 (2000).
- Turner, T. J. Calcium channels coupled to glutamate release Prog. Brain. Res. 116, 3–14 (1998).

562 JULY 2004 VOLUME 5

- Gee, N. S. et al. The novel anticonvulsant drug, gabapentin (Neurontin) binds to the α<sub>2</sub>δ subunit of a calcium channel. J. Biol. Chem. 271. 5768–5776 (1996).
  - A protein from guinea-pig cerebral cortex that binds [\*H]gabapentin with high affinity is solubilized and purified; the partial N-terminal amino-acid sequence is identical to the calcium channel \( \mathcal{\pi} - 52. Marais, E., Klugbauer, N. & Hofmann, F. Calcium channel  $\alpha_2\delta$  subunits structure and gabapentin binding. *Mol. Pharmacol.* **59**, 1243–1248 (2001).
- Bryans, J. S. et al. Identification of novel ligands for the gabapentin binding site on the α2δ subunit of a calcium channel and their evaluation as anticonvulsant agents. J. Med. Chem. 4, 1838–1845 (1998).
- Dooley, D. J., Donovan, C. M., Meder, W. P. & Whetzel, S. Z. Preferential action of gabapentin and pregabalin at P/Q-type voltage-sensitive calcium channels: inhibition of K\*-evoked [3H]-norepinephrine release from rat neocortical slices. Synapse 45, 171–190 (2002).
- Dolphin, A. C. et al. The effect of α2δ and other accessory subunits on expression and properties of the calcium channel α1G. J. Physiol. (Lond.) 519, 35–45 (1999).
- Arikkath, J. & Campbell, K. P. Auxiliary subunits: essential components of the voltage-gated calcium channel complex. *Curr. Opin. Neurobiol.* 13, 298–307 (2003).
- Brown, J. P. & Gee, N. S. Cloning and deletion mutagenesis of the α2δ calcium channel subunit from porcine cerebral cortex. J. Biol. Chem. 273, 25458–25465 (1998).
- Wang, M., Offord, J., Oxender, D. L. & Su, T.-Z. Structural requirement of the calcium channel subunit α2δ for gabapentin binding. *Biochem. J.* 342, 313–320 (1999).
- Martin, D. J. et al. Gabapentin-mediated inhibition of voltage-activated Ca<sup>2+</sup> channel currents in cultured sensory neurones is dependent on culture conditions and channel subunit expression. Neuropharmacology 42, 353–366 (2002).
- Sutton, K. G., Martin, D. J., Pinnock, R. D., Lee, K. & Scott, R. H. Gabapentin inhibits high-threshold calcium channel currents in cultured rat dorsal root ganglion neurones. *Brit. J. Pharmacol.* 135, 257–265 (2002).
- Sarantopoulos, C., McCallum, B., Kwok, W. M. & Hogan, Q. Gabapentin decreases membrane calcium currents in injured as well as in control mammalian primary afferent neurons. Reg. Anesth. Pain. Med. 27, 47–57 (2002).
- Dooley, D. J., Mieske, C. A. & Borosky, S. A. Inhibition of K\*evoked glutamate release from rat neocortical and hippocampal slices by gabapentin. *Neurosci. Lett.* 280, 107–110 (2000).
- Shimoyama, M., Shimoyama, N. & Hori, Y. Gabapentin affects glutamatergic excitatory neurotransmission in the rat dorsal horn. *Pain* 85, 405–414 (2000).
- Löscher, W., Honack, D. & Taylor, C. P. Gabapentin increases aminooxyacetic acid-induced GABA accumulation in several regions of rat brain. *Neurosci. Lett.* 128, 150–154 (1991).
- Petroff, O. A., Hyder, F., Rothman, D. L. & Mattson, R. H. Effects of gabapentin on brain GABA, homocarnosine, and pyrrolldinone in epilepsy patients. *Epilepsia* 41, 675–680 (2001)
- Errante, L. D., Williamson, A., Spencer, D. D. & Petroff, O. A Gabapentin and vigabatrin increase GABA in the human neocortical slice. *Epilepsy Res.* 49, 203–210 (2002).
- Ffrench-Mullen, J. M. H., Barker, J. L. & Rogawski, M. A.
  Calcium current block by (-)-pentobarbital, phenobarbital
  and CHEB but not (+)-pentobarbital in acutely isolated CA1
  neurons: comparison with effects on GABA-activated CIcurrent. J. Neurosci. 13, 3211–3221 (1993).
- Stefani, A., Spadoni, F., Siniscalchi, A. & Bérnardi, G. Lamotrigine inhibits Ca<sup>2+</sup> currents in cortical neurons: functional implications. Eur. J. Pharmacol. 307, 113–116 (1996).
- Wang, S.-J., Huang, C.-C., Hsu, K.-S., Tsai, J.-J. & Gean, P.-W. Inhibition of N-type calcium currents by lamotrigine in rat amygdalar neurones. *Neuroreport* 7, 3037–3040 (1904)
- Zona, C. et al. Levetiracetam does not modulate neuronal voltage-gated Na<sup>+</sup> and T-type Ca<sup>2+</sup> currents. Seizure 10, 279–286 (2001).
- Niespodziany, I., Klitgaard, H. & Margineanu, D. G. Levetiracetam inhibits the high-voltage-activated Ca<sup>2+</sup> current in pyramidal neurones of rat hippocampal slices. Neurosci. Lett. 306, 5–8 (2001).
- Lukyanetz, E. A., Shkryl, V. M. & Kostyuk, P. G. Selective blockade of N-type calcium channels by levetiracetam. *Epilepsia* 43, 9–18 (2002).
- Perez-Reyes, E. Molecular physiology of low-voltageactivated T-type calcium channels. *Physiol. Rev.* 83, 117–161 (2003).
- Huguenard, J. R. Low-threshold calcium currents in central nervous system neurons. *Annu. Rev. Physiol.* 58, 329–358 (1994)

- Suzuki, S. & Rogawski, M. A. T-type calcium channels mediate the transition between tonic and phasic firing in thalamic neurons. *Proc. Natl Acad. Sci. USA* 86, 7228–7232 (1989).
- Talley, E. M. et al. Differential distribution of three members of a gene family encoding low voltage-activated (T-type) calcium channels. J. Neurosci. 19, 1895–1911 (1999).
- Kim, D. et al. Lack of the burst firing of thalamocortical relay neurons and resistance to absence seizures in mice lacking α1G T-type Ca<sup>2+</sup> channels. Neuron 31, 35–45 (2001).
- Tsakiridou, E., Bertollini, L., de Curtis, M., Avanzini, G. & Pape, H. C. Selective increase in T-type calcium conductance of reticular thalamic neurons in a rat model of absence epilepsy. J. Neurosci. 15, 3110–3117 (1995).
- Talley, E. M., Solórzano, G., Depaulis, A., Perez-Reyes, E. & Bayliss, D. A. Low-voltage-activated calcium channel subunit expression in a genetic model of absence epilepsy in the rat. *Brain Res. Mol. Brain Res.* 75, 159–165 (2000).
- Chen, Y. et al. Association between genetic variation of CACNA1H and childhood absence epilepsy. Ann. Neurol. 54, 239–243 (2003).
- Khosravani, H. et al. Gating effects of mutations in the Ca,3.2 T-type calcium channel associated with childhood absence epilepsy. J. Biol. Chem. 279, 9681–9684 (2004).
- Heron, S. E. et al. Genetic variation of CACNA1H in idiopathic generalized epilepsy. Ann. Neurol. 55, 595–596 (2004).
- Coulter, D. A., Huguenard, J. R. & Prince, D. A. Characterization of ethosuximide reduction of low-threshold calcium current in thalamic relay neurons. *Ann. Neurol.* 25, 582–593 (1989). The inhibitory effect of ethosuximide on T-type calcium channels is hypothesized to account for its anti-absence activity.
- Gomora, J. C., Daud, A. N., Weiergräber, M. & Perez-Reyes, E. Block of cloned human T-type calcium channels by succinimide antiepileptic drugs. *Mol. Pharmacol.* 60, 1121–1132 (2001).
- McLean, M. J. & Macdonald, R. L. Sodium valproate, but not ethosuximide, produces use- and voltage-dependent limitation of high frequency repetitive firing of action potentials of mouse central neurons in cell culture. J. Pharmacol. Exp. Ther. 237, 1001–1011 (1986).
- Leresche, N. et al. On the action of the anti-absence drug ethosuximide in the rat and cat thalamus. J. Neurosci. 18, 4842–4853 (1998).
- Manning, J. P., Richards, D. A., Leresche, N., Crunelli, V. & Bowery, N. G. Cortical-area specific block of genetically determined absence seizures by ethosuximide. *Neuroscience* 123, 5–9 (2004).
- Kito, M., Maehara, M. & Watanabe, K. Mechanisms of T-type calcium channel blockade by zonisamide. Seizure 5, 115–119 (1996).
- Galarreta, M. & Hestrin, S. Frequency-dependent synaptic depression and the balance of excitation and inhibition in the neocortex. *Nature Neurosci.* 1, 587–594 (1998).
- Miles, R. & Wong, R. K. Inhibitory control of local excitatory circuits in the guinea-pig hippocampus. *J. Physiol. (Lond.)* 388, 611–629 (1987).
- Wallace, R. H. et al. Mutant GABA, receptor γ2-subunit in childhood absence epilepsy and febrile seizures. Nature Genet. 28, 49–52 (2001).

## This paper and reference 92 were the first to identify mutations in GABA<sub>A</sub> receptor subunits as a genetic basis of epilepsy.

- Baulac, S. et al. First genetic evidence of GABA<sub>A</sub> receptor dysfunction in epilepsy: a mutation in the γ2-subunit gene. Nature Genet. 28, 46–48 (2001).
- Kananura, C. et al. A splice-site mutation in GABRG2 associated with childhood absence epilepsy and febrile convulsions. Arch. Neurol. 59, 1137–1141 (2002).
- Harkin, L. A. et al. Truncation of the GABA<sub>A</sub>-receptor γ2 subunit in a family with generalized epilepsy with febrile seizures plus. Am. J. Hum. Genet. 70, 530–536 (2002).
- Bianchi, M. T., Song, L., Zhang, H. & Macdonald, R. L. Two different mechanisms of disinhibition produced by GABA, receptor mutations linked to epilepsy in humans. *J. Neurosci.* 22, 5321–5327 (2002).
- Cossette, P. et al. Mutation of GABRA1 in an autosomal dominant form of juvenile myoclonic epilepsy. Nature Genet. 31, 184–189 (2002).
- Gallagher, J. P., Higashi, H. & Nishi, S. Characterization and ionic basis of GABA-induced depolarizations recorded in vitro from cat primary afferent neurones. J. Physiol. (Lond.) 275, 263–282 (1978).
- Akaike, N., Inomata, N. & Yakushiji, T. Differential effects of extra- and intracellular anions on GABA-activated currents in bullfrog sensory neurons. J. Neurophysiol. 62, 1388–1399 (1989).
- Bormann, J., Hamill, O. P. & Sakmann, B. Mechanism of anion permedion through channels gated by glycine and γ-aminobutyric acid in mouse cultured spinal neurones. *J. Physiol. (Lond.)* 385, 243–286 (1987).

- Rudolph, U. et al. Benzodiazepine actions mediated by specific γ-aminobutyric acid, receptor subtypes. Nature 401, 796–800 (1999).
  - In mice engineered with a knock-in mutation (H101R) in the GABA, receptor  $\alpha 1$  subunit that confers benzodiazepine insensitivity, the benzodiazepine diazepam failed to exhibit sedative and amnesic activity and had reduced anticonvulsant activity, whereas the anxiolytic and muscle relaxant effects were unaffected, revealing the specific behavioural functions of GABA, receptors containing the  $\alpha 1$  subunit.
- Crestani, F., Martin, J. R., Möhler, H. & Rudolph, U. Mechanism of action of the hypnotic zolpidem in vivo. Br. J. Pharmacol. 131, 1251–1256 (2000)
- Bai, D. et al. Distinct functional and pharmacological properties of tonic and quantal inhibitory postsynaptic currents mediated by y-aminobutyric acid, receptors in hippocampal neurons. Mol. Pharmacol. 59, 814–824 (2001).
- 103. Sohaí, V. S., Keist, R., Rudolph, U. & Huguenard, J. R. Dynamic GABA<sub>A</sub> receptor subtype-specific modulation of the synchrony and duration of thalamic oscillations. J. Neurosci. 23, 3649–3657 (2003).
- Macdonald, R. L. & Olsen, R. W. GABA<sub>A</sub> receptor channels. *Annu. Rev. Neurosci.* 17, 569–602 (1994).
- Rho, J. M., Donevan, S. D. & Rogawski, M. A. Barbituratelike actions of the propanediol dicarbamates felbamate and meprobamate. J. Pharmacol. Exp. Ther. 280, 1383–1391 (1997).
- White, H. S., Brown, S. D., Woodhead, J. H., Skeen, G. A. & Wolf, H. H. Topiramate enhances GABA-mediated chloride flux and GABA-evoked chloride currents in murine brain neurons and increases seizure threshold. *Epilepsy Res.* 28, 167–179 (1997).
- Gordey, M., DeLorey, T. M. & Olsen, R. W. Differential sensitivity of recombinant GABA<sub>A</sub> receptors expressed in *Xenopus* oocytes to modulation by topiramate. *Epilepsia* 41 (Suppl. 1), S25–29 (2000).
- De Biase, D., Barra, D., Bossa, F., Pucci, P. & John, R. A. Chemistry of the inactivation of 4-aminobutyrate aminotransferase by the antiepileptic drug vigabatrin. J. Biol. Chem. 266, 20056–20061 (1991).
- 109. Löscher, W. & Horstermann, D. Differential effects of vigabatrin, γ-acetylenic GABA, aminooxyacetic acid, and valproate on levels of various amino acids in rat brain regions and plasma. Naunyn Schmiedeberg's Arch. Pharmacol. 349, 270–278 (1994).
- Jackson, M. F., Esplin, B. & Capek, R. Reversal of the activity-dependent suppression of GABA-mediated inhibition in hippocampal slices from y-vinyl GABA (vigabatrin)pretreated rats. *Neuropharmacology* 39, 65–74 (2000).
- Overstreet, L. S. & Westbrook, G. L. Paradoxical reduction of synaptic inhibition by vigabatrin. *J. Neurophysiol.* 86, 596–603 (2001).
- Wu, Y., Wang, W. & Richerson, G. B. Vigabatrin induces tonic inhibition via GABA transporter reversal without increasing vesicular GABA release. *J. Neurophysiol.* 89, 2021–2034 (2003).
- Richerson, G. B. & Wu, Y. Dynamic equilibrium of neurotransmitter transporters: not just for reuptake anymore. J. Neurophysiol. 90, 1363–1374 (2003).
- Löscher, W. Effect of inhibitors of GABA transaminase on the synthesis, binding, uptake, and metabolism of GABA. J. Neurochem. 34, 1603–1608 (1980).
- 115. Schuler, V. et al. Epilepsy, hyperalgesia, impaired memory, and loss of pre- and postsynaptic GABA<sub>B</sub> responses in mice lacking GABA<sub>B(1)</sub>. Neuron 31, 47–58 (2001).
- Sarup, A., Larsson, O. M. & Schousboe, A. GABA transporters and GABA-transaminase as drug targets. *Curr. Drug Target CNS Neurol. Disord.* 2, 269–277 (2003).
- Suzdak, P. D. & Jansen, J. A. A review of the preclinical pharmacology of tiagabine: a potent and selective anticonvulsant GABA uptake inhibitor. *Epilepsia* 36, 612–626 (1995).
- Thompson, S. M. & Gahwiler, B. H. Effects of the GABA uptake inhibitor tiagabine on inhibitory synaptic potentials in rat hippocampal slice cultures. *J. Neurophysiol.* 67, 1698–1701 (1992).
  - Recordings from CA3 hippocampal pyramidal cells demonstrate that tiagabine greatly prolongs the duration of monosynaptic inhibitory synaptic potentials and can also inhibit epileptiform bursting.
- 119. Engel, D. et al. Laminar difference in GABA uptake and GAT-1 expression in rat CA1. J. Physiol. (Lond.) 512, 643–649 (1998).
- 120. Jackson, M. F., Esplin, B. & Capek, R. Activity-dependent enhancement of hyperpolarizing and depolarizing γ-aminobutyric acid (GABA) synaptic responses following inhibition of GABA uptake by tlagabine. *Epilepsy Res.* 37, 25–36 (1999).

- Dingledine, R., Borges, K., Bowie, D. & Traynelis, S. F. The glutamate receptor ion channels. *Pharmacol. Rev.* 51, 7–61 (1999)
- Löscher, W. & Rogawski, M. in *Ionotropic Glutamate Receptors as Therapeutic Targets* (eds Lodge, D., Danysz, W. & Parsons, C. G.) 91–132 (F. P. Graham, Johnson City, Tennessee, 2002).
- Rogawski, M. A. et al. GluR5 kainate receptors, seizures, and the amygdala. Ann. NY Acad. Sci. 985, 150–162 (2003).
- 124. Sheth, R. D. & Gidal, B. E. Refractory status epilepticus: response to ketamine. *Neurology* **51**, 1765–1766 (1998).
- Boyd, S. G. & Dan, B. Cortical sensitivity in refractory status epilepticus. *Clin. Neurophysiol.* 111 (Suppl. 1), S156–164 (2000).
- Mewasingh, L. D. et al. Oral ketamine in paediatric nonconvulsive status epilepticus. Seizure 12, 483–489 (2003).
- Subramaniam, S. et al. Felbamate block of the N-methyl-oaspartate receptor. J. Pharmacol. Exp. Ther. 273, 878–886 (1995)
- 128. Kuo, C. C., Lin, B. J., Chang, H. R. & Hsieh, C. P. Use-dependent inhibition of the *N*-methyl-o-aspartate currents by felbamate: a gating modifier with selective binding to the desensitized channels. *Mol. Pharmacol.* 65, 370–380 (2004).
- 129. Kleckner, N. W., Glazewski, J. C., Chen, C. C. & Moscrip, T. D. Subtype-selective antagonism of *N*-methyl-o-aspartate receptors by felbamate: insights into the mechanism of action. *J. Pharmacol. Exp. Ther.* 289, 886–894 (1999).
- Harty, T. P. & Rogawski, M. A. Felbamate block of recombinant N-methyl-p-aspartate receptors: selectivity for the NR2B subunit. *Epilepsy Res.* 39, 47–55 (2000).
- 131. Bleakman, D. Kainate receptor pharmacology and physiology. *Cell. Mol. Life. Sci.* **56**, 558–566 (1999)

- Lerma, J., Paternain, A. V., Rodriguez-Moreno, A. & López-Garcia, J. C. Molecular physiology of kainate receptors. *Physiol. Rev.* 81, 971–998 (2001).
- Smolders, I. et al. Antagonists of GLU<sub>rs</sub>-containing kainate receptors prevent pilocarpine-induced limbic seizures. *Nature Neurosci.* 5, 796–804 (2002).
- Gryder, D. S. & Rogawski, M. A. Selective antagonism of GluR5 kainate-receptor-mediated synaptic currents by topiramate in rat basolateral amygdala neurons. *J. Neurosci* 23, 7069–7074 (2003).
  - A demonstration that topiramate inhibits AMPA and kainate receptor mediated synaptic excitation, but is more potent and effective against the kainate receptor-mediated component.
- 135. Kaminski, R. M., Banerjee, M. & Rogawski, M. A. Topiramate selectively protects against selzures induced by ATPA, a GluR5 kainate receptor agonist. Neuropharmacology 46, 1097–1104 (2004).
- Löscher, W. Animal models of epilepsy for the development of antiepileptogenic and disease-modifying drugs. A comparison of the pharmacology of kindling and models with spontaneous recurrent seizures. Epilepsy Res. 50, 105–123 (2002).
- White, H. S. Animal models of epileptogenesis. *Neurology* (Suppl. 5), S7–14 (2002).
- Lynch, B. A. et al. The synaptic vesicle protein SV2A is the binding site for the antiepileptic drug levetiracetam. Proc. Natl Acad. Sci. USA (in the press).
- Scheffer, I. E. & Berkovic, S. F. The genetics of human epilepsy. *Trends Pharmacol. Sci.* 24, 428–433 (2003).
- Escayg, A. et al. Coding and noncoding variation of the human calcium-channel 84-subunit gene CACNB4 in patients with idiopathic generalized epilepsy and episodic ataxia. Am. J. Hum. Genet. 66, 1531–1539 (2000).

- Jouvenceau A. et al. Human epilepsy associated with dysfunction of the brain P/Q-type calcium channel. Lancet 358, 801–807 (2001).
- Holtmann, M., Opp, J., Tokarzewski, M. & Korn-Merker, E. Human epilepsy, episodic ataxia type 2, and migraine. *Lancet* 359, 170–171 (2002).
- 143. Chioza, B. et al. Haplotype and linkage disequilibrium analysis to characterise a region in the calcium channel gene CACNA1A associated with idiopathic generalised epilepsy. Eur. J. Hum. Genet. 10, 857–864 (2002).
- 144. Chioza, B. *et al.* Association between the  $\alpha_{1a}$  calcium channel gene *CACNA1A* and idiopathic generalized epilepsy. *Neurology* **56**, 1245–1246 (2001).

Competing interests statement
The authors declare that they have no competing financial interests.



#### DATABASES

The following terms in this article are linked online to: Entrez Gene:

http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=gene CACNA1H | GABAT | GABRA1 | GAD | GAT1 | mGluR1 | mGluR5 | NR1 | NR2B

#### **FURTHER INFORMATION**

Encyclopedia of Life Sciences: http://www.els.net/epilepsy|epilepsy: management
Rogawski's homepage:
http://intra.ninds.nih.gov/Lab.asp?Org\_ID=464
Access to this interactive links box is free online.

564 JULY 2004 VOLUME 5