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The Neurodevelopmental Hypothesis of Schizophrenia, Revisited

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While multiple theories have been put forth regarding the origin of schizophrenia, by far the vast majority of evidence points to the neurodevelopmental model in which developmental insults as early as late first or early second trimester lead to the activation of pathologic neural circuits during adolescence or young adulthood leading to the emergence of positive or negative symptoms. In this report, we examine the evidence from brain pathology (enlargement of the cerebroventricular system, changes in gray and white matters, and abnormal laminar organization), genetics (changes in the normal expression of proteins that are involved in early migration of neurons and glia, cell proliferation, axonal outgrowth, synaptogenesis, and apoptosis), environmental factors (increased frequency of obstetric complications and increased rates of schizophrenic births due to prenatal viral or bacterial infections), and gene-environmental interactions (a disproportionate number of schizophrenia candidate genes are regulated by hypoxia, microdeletions and microduplications, the overrepresentation of pathogen-related genes among schizophrenia candidate genes) in support of the neurodevelopmental model. We relate the neurodevelopmental model to a number of findings about schizophrenia. Finally, we also examine alternate explanations of the origin of schizophrenia including the neurodegenerative model.

Key words: brain/genes/animal model/pathology/epidemiology/antiviral model/schizophrenia

The Neurodevelopmental Theory of Schizophrenia and Supportive Evidence

Schizophrenia is a neurodevelopmental disorder that affects youth in puberty and is manifested by a disruption in cognition and emotion along with negative (ie, avolition, alogia, apathy, poor or nonexistent social functioning) and positive (presence of hallucinations, delusions) symptoms. According to the neurodevelopmental hypothesis, the etiology of schizophrenia may involve pathologic processes, caused by both genetic and environmental factors, that begin before the brain approaches its adult anatomical state in adolescence.[1] These neurodevelopmental abnormalities, developing in utero as early as late first or early second trimester[2] for some and thereafter for others, have been suggested to lead to the activation of pathologic neural circuits during adolescence or young adulthood (sometimes owing to severe stress), which leads to the emergence of positive or negative symptoms or both.[2,3,4]

Earlier neuropathologic work indicated that some cases of schizophrenia result from embryologic maldevelopment.[5] E. Slater also referred to maldevelopmental similarities between temporal lobe epilepsy and schizophrenia and stressed their possible neuropathologic basis.[6] The emergence of evidence for cortical maldevelopment in schizophrenia and the development of several plausible animal models of schizophrenia,[7] which are based on various paradigms that produce behavioral abnormalities or altered sensitivity to dopaminergic drugs only in adolescent or adult animals, have strengthened the link between maldevelopment and schizophrenia. The concept of schizophrenia as a neurodevelopmental disorder is also consistent with other epidemiologic and clinical lines of evidence, discussed in the following sections.

A "2-hit" model proposed by Keshavan[8,9] works within the framework of the neurodevelopmental theory in which maldevelopment during 2 critical time points (early brain development and adolescence) combines to produce the symptoms associated with schizophrenia. According to this model, early developmental insults may lead to dysfunction of specific neural networks that would account for premorbid signs and symptoms observed in individuals that later develop schizophrenia.[8] At adolescence, excessive elimination of synapses and loss of plasticity may account for the emergence of symptoms.[8,9]

Congenital Abnormalities

Multiple markers of congenital anomalies indicative of neurodevelopmental insults have been found in schizophrenia.[10,11] Such anomalies include agenesis of corpus

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callosum, stenosis of sylvian aqueduct, cerebral hamartomas, and cavum septum pellucidum. Presence of low-set ears, epicanthal eye folds, and wide spaces between the first and second toes are suggestive of first trimester anomalies.[10,11] There is, however, support for abnormal dermatoglyphics in patients with schizophrenia indicating a second trimester event.[12,13] Multiple reports indicate the presence of premorbid neurologic soft signs in children who later develop schizophrenia.[14–16] Slight posturing of hands and transient choreoathetoid movements have been observed during the first 2 years of life in children who later developed schizophrenia.[15,17] Additionally, poor performance on tests of attention and neuromotor performance, mood and social impairment, and excessive anxiety have been reported to occur more frequently in high-risk children with a schizophrenic parent.[18,19] All these findings are consistent with schizophrenia as a syndrome of abnormal brain development.

Environmental Factors

There is a large body of epidemiologic research showing an increased frequency of obstetric and perinatal complications in schizophrenic patients.[20] The complications observed include periventicular hemorrhages, hypoxia, and ischemic injuries.[10,21] There is also a robust collection of reports indicating that environmental factors, especially viral infections, can increase the risk for development of schizophrenia.[22,23] Hare et al[24] and Machon et al[25] reported on excess of schizophrenic patients being born during late winter and spring as indicators of potential influenza infections being responsible for these cases. Indeed, the majority of nearly 50 studies performed in the intervening years indicate that 5%–15% excess schizophrenic births in the northern hemisphere occur during the months of January and March.[26–28] This excess winter birth has not been shown to be due to unusual patterns of conception in mothers or to a methodological artifact.[26,29] Machon et al[25] and Mednick et al[30] showed that the risk of schizophrenia was increased by 50% in Finnish individuals whose mothers had been exposed to the 1957 A2 influenza during the second trimester of pregnancy. Later, 9 out of 15 studies performed replicated Mednick's findings of a positive association between prenatal influenza exposure and schizophrenia.[2] These association studies showed that exposure during the 4th–7th months of gestation affords a window of opportunity for influenza virus to cause its teratogenic effects on the embryonic brain.[4] Additionally, 3 out of 5 cohort and case-control studies support a positive association between schizophrenia and maternal exposure to influenza prenatally.[31–33] Subsequent studies have now shown that other viruses such as rubella[34] may also increase the risk for development of schizophrenia in the affected progeny of exposed mothers.[26,34] A review by Brown[35] summarized that (1) there was a 10- to 20-fold risk of developing schizophrenia following prenatal exposure to rubella; (2) prenatal exposure to influenza in the first trimester increased 7-fold, and infection in early to midgestation increased risk 3-fold; and (3) presence of maternal antibodies against *Toxoplasma gondii* lead to 2.5-fold increased risk.[35] By far, the most exciting evidence linking viral exposure to development of schizophrenia was published by Karlsson et al,[23] who provided data suggestive of a possible role for retroviruses in the pathogenesis of schizophrenia.[22] Karlsson et al[23] identified nucleotide sequences homologous to retroviral polymerase genes in the cerebrospinal fluid of 28.6% of subjects with schizophrenia of recent origin and in 5% of subjects with chronic schizophrenia. In contrast, such retroviral sequences were not found in any individuals with noninflammatory neurological illnesses or in normal subjects.[22,23] More recently, Perron et al.[220] using an immunoassay to quantify serum levels of human endogenous retrovirus type W family GAG and envelope (ENV) proteins in subjects with schizophrenia and matched controls. Positive antigenemia for ENV was found in 23 of 49 (47%) and for GAG in 24 of 49 (49%) of patients with schizophrenia. In contrast, for control subjects only 1 of 30 (3%) for ENV and 2 of 49 (4%) for GAG were positive in blood donors ($p < .01$ for ENV; $p < .001$ for GAG), providing further evidence of an association between retroviruses and schizophrenia.[220] The upshot of these studies and previous epidemiological reports is that schizophrenia may represent the shared phenotype of a group of disorders whose etiopathogenesis involves the interaction between genetic influences and environmental risks, such as viruses operating on brain maturational processes.[22] Moreover, identification of potential environmental risk factors, such as influenza virus or retroviruses such as endogenous retroviral-9 family and the human endogenous retrovirus-W species observed by Karlsson et al,[23] will help in targeting early interventions at repressing the expression of these transcripts. An alternate approach would be to vaccinate against influenza thus influencing the course and outcome of schizophrenia in the susceptible individuals.[22]

There are at least 2 mechanisms that may be responsible for transmission of viral effects from the mother to the fetus. (1) *Via direct viral infection*: There are clinical, as well as direct experimental, reports[36–39] showing that human influenza A viral infection of a pregnant mother may cause transplacental passage of viral load to the fetus. In a series of reports, Aronsson and colleagues used human influenza virus (A/WSN/33, a neurotropic strain of influenza A virus) on day 14 of pregnancy, to infect pregnant C57BL/6 mice intranasally. Viral RNA and nucleoprotein were detected in fetal brains, and viral RNA persisted in the brains of exposed offspring for at least 90 days of postnatal life thus showing evidence for transplacental passage of influenza virus in mice and the persistence of viral components in the brains of progeny into young

adulthood.[38] Additionally, Aronsson et al[38] have demonstrated that 10–17 months after injection of the human influenza A virus into olfactory bulbs of TAP1 mutant mice, viral RNA encoding the nonstructural NS1 protein was detected in midbrain of the exposed mice. The product of *NS1* gene is known to play a regulatory role in the host cell metabolisms.[40] Several in vitro studies have also shown the ability of human influenza A virus to infect Schwann cells,[41] astrocytes, microglial cells and neurons,[36] and hippocampal GABAergic cells,[42,43] selectively causing persistent infection of target cells in the brain. (2) *Via induction of cytokine production*: Multiple clinical and experimental reports show the ability of human influenza infection to induce production of systemic cytokines by the maternal immune system, the placenta, or even the fetus itself.[44–48] New reports show presence of serologic evidence of maternal exposure to influenza as causing increased risk of schizophrenia in offspring.[4] Offspring of mothers with elevated immunoglobulin G and immunoglobulin M levels, as well as antibodies to herpes simplex virus type 2, during pregnancy have an increased risk for schizophrenia.[49] Cytokines such as interleukin (IL)-1β, IL-6, and tumor necrosis factor α (TNF-α) are elevated in the pregnant mothers after maternal infection[44,45,48] and after infection in animal models.[47,48] All these cytokines are known to regulate normal brain development and have been implicated in abnormal corticogenesis.[50–52] Additionally, expression of messenger RNAs (mRNAs) for cytokines in the central nervous system (CNS) is developmentally regulated both in man and in mouse,[53–57] emphasizing the significant role that cytokines play during neurodevelopment. IL-1β, IL-6, and TNF-α cross the placenta and are synthesized by mother,[58] by the placenta,[59] and by the fetus.[59] Maternal levels of TNF-α and IL-8 have been shown to be elevated in human pregnancies in which the offspring goes on to develop schizophrenia.[4,59] A more relevant series of studies in different animal models for schizophrenia show that maternal infection with human influenza mimic poly I:C, a synthetic double-stranded RNA that stimulates a cytokine response in mice, can cause abnormalities in prepulse inhibition (PPI)[60] or, after maternal exposure to E. coli cell wall endotoxin lipopolysaccharide, cause disruption of sensorimotor gating in the offspring.[61] Finally, maternal exposure to poly I:C also causes disrupted latent inhibition in rat.[62] All these models suggest that direct stimulation of cytokine production by infections or immunogenic agents cause disruptions in various brain structural or behavioral indices of relevance to schizophrenia. Other factors associated with increased schizophrenic births include famine during pregnancy,[63,64] Rh factor incompatibility,[65] and autoimmunity due to infectious agents.[66]

A number of animal models are currently in use to study schizophrenia and identify potential new therapies (reviewed by Carpenter and Koenig[7]). Our laboratory has studied the effects of prenatal human influenza viral infection on day 9 of pregnancy in BALB/c and C57BL/6 mice and their offspring. These studies showed the deleterious effects of influenza on growing brains of exposed offspring. Briefly, embryonic day 9 (E9) pregnant BALB/c mice were exposed to influenza A/NWS/33 (H1N1) or vehicle, following determination of viral dosage, causing sublethal lung and upper respiratory infection. Pregnant mice were allowed to deliver pups. The day of delivery was considered day 0. Prenatally infected murine brains from postnatal day 0 showed significant reductions in reelin-positive cell counts in layer I of neocortex and other cortical layers ($P < .0001$) when compared with controls.[67] Whereas layer I Cajal-Retzius cells produced significantly less reelin in infected animals, the same cells showed normal production of calretinin and neuronal nitric oxide synthase (nNOS) when compared with control brains.[67] This work has recently been confirmed by Meyer et al[68] who also observed a decrease in reelin-positive cells in medial prefrontal cortex (PFC) following poly I:C exposure on E9 and E17.

Additionally, prenatal viral infection on E9 resulted in various behavioral abnormalities.[60] These included abnormal exploratory behavior, reflecting difficulty handling stress, similar to what is observed in schizophrenia. The offspring of exposed mice showed significantly less time exploring their environment vs control mice.[60] Moreover, the offspring of exposed mice contacted each other less frequently than the control mice, suggesting altered social behavior.[60] Finally, the offspring of exposed mice displayed an abnormal acoustic startle response,[60] similar to PPI deficits in untreated schizophrenic subjects.[69] Administration of antipsychotic agents chlorpromazine (a typical agent) and clozapine (an atypical agent), agents which treat schizophrenic symptoms and correct PPI deficits in patients, caused significant increases in PPI in the exposed mice vs controls, correcting the PPI deficits.[60] The response by offspring of exposed mice to both antipsychotics shows that our animal model has predictive validity for positive symptoms of schizophrenia.[60]

Our laboratory has previously shown that infection of BALB/c mice at E9 has deleterious effects on brain morphology[67,70] (figure 1). Prenatally infected brains from P0 displayed decreases in neocortical and hippocampal thickness.[67] Moreover, brains at P0 displayed increased pyramidal cell density and significantly reduced pyramidal cell nuclear size.[70] By adulthood (P98), there continued to be an increase in pyramidal cell density and nonpyramidal cell density and a significant reduction in pyramidal cell nuclear size.[70] Taken together, these data suggest that prenatal viral infection at E9 (late first trimester) causes persistent deleterious changes in brain morphology. Morphometric analysis of brain also revealed numerous defects following infection of C57BL/6 mice at E18 (late second trimester). Analysis of brain and lateral ventricular volume areas in postnatal brains showed significant atrophy of the brain volume by

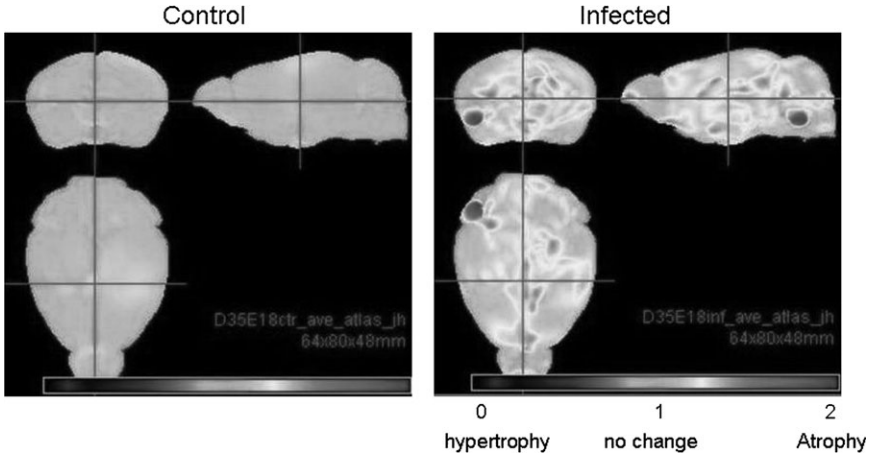

Fig. 1. Magnetic Resonance Imaging Reveals Significant ($P < .05$) Brain Atrophy in Multiple Brain Areas of the 35-d-Old Virally Infected Mouse Offspring (Right Panel) as Compared With Sham-Infected Mice (Left Panel). Originally published in Fatemi et al.[72]

approximately equal to 4% ($P < .05$) in P35 offspring of exposed mice.[71] There were significant reductions in volume for the cerebellum ($P < .001$) and hippocampus ($P < .00005$) at P35.[71] Fractional anisotropy of corpus callosum revealed white matter atrophy on P35 offspring ($P < .0082$) of exposed mice.[71]

Brain gene expression also changes in response to prenatal viral infection.[71–73] Gene expression data showed a significant ($P < .05$) at least 1.5-fold up- or downregulation of genes in frontal (43 upregulated and 29 downregulated at P0, 16 upregulated and 17 downregulated at P14, and 86 upregulated and 24 downregulated at P56), hippocampal (129 upregulated and 46 downregulated at P0, 9 upregulated and 12 downregulated at P14, and 45 upregulated and 17 downregulated at P56), and cerebellar (120 upregulated and 37 downregulated at P0, 11 upregulated and 5 downregulated at P14, and 74 upregulated and 22 downregulated at P56) areas of mouse offspring.[71] Several genes, which have been previously implicated in etiopathology of schizophrenia, were shown to be affected significantly ($P < .05$) in the same direction and the magnitude of change was validated by quantitative real-time polymerase chain reaction (qRT-PCR)[71] (table 1). There were also several genes that were known to be involved in influenza-mediated RNA processing and that were upregulated in all 3 brain areas and continued to be present at P0, eg, NS1 influenza–binding protein and aryl hydrocarbon receptor nuclear translocator genes.[71]

Prenatal viral infection may lead to the development of schizophrenia in multiple ways (figure 2). One way is via an epigenetic mechanism in which hypermethylation of promoters by molecules such as DNA methyltransferase 1 (*DNMT1*) results in altered expression of schizophrenia candidate genes. DNMT1 mRNA has been shown to be increased in brains of subjects with schizophrenia.[74] Activation of DNMT1, in turn, hypermethylates promoters for reelin and glutamic acid decarboxylase (GAD)67-kDa protein genes resulting in decreased levels of these molecules.[75] These changes contribute to abnormal brain development and altered γ-aminobutyric acid (GABA) signaling and subsequent genesis of schizophrenia. Maternal infection may also lead to activation of the maternal immune response leading to altered levels of cytokines including IL-1β, IL-6, and TNF-α that regulate normal brain development and are altered following maternal infection.[44,45,48] Changes may lead to abnormal cortical development[50–52] and, ultimately, schizophrenia. Prenatal viral expression may also lead to altered expression of genes that are involved in cell-cell communication and changes in cell structure due to chronic actin depolymerization (S.H. Fatemi and M. Peoples, unpublished observations, 2007). Aquaporin 4 is localized to astrocytes and ependymal cells in brain and is involved with water transport.[76,77] Aquaporin 4 protein expression is significantly decreased at postnatal day 35 in neocortex in BALB/c mice following infection at E9[78] possibly resulting in altered cell morphology. Similarly, chronic actin depolymerization may alter gene expression in schizophrenia (S.H. Fatemi and M. Peoples, unpublished observations, 2007). nNOS, which is associated with F-actin, displays altered expression following prenatal viral infection at E9 and may show altered expression following actin disruption. Actin depolymerization (with cytochalasin D) causes internalization of NR1 subunit of *N*-methyl-D-aspartate (NMDA) and therefore decreased NMDA currents leading to altered signaling, but it is unknown whether this occurs in schizophrenia. Our laboratory has observed significant increase in nNOS at P35 and a significant decrease at P56 that may lead to altered synaptogenesis and excitotoxicity in neonatal brains.[79]

Table 1. Microarray and qRT-PCR Results for Selected Affected Genes in E18 Infected Mice

Gene	Symbol	Area	PD	Microarray Fold Change	Microarray *P* Value	Gene Relative to Normalizer (qRT-PCR)	qRT-PCR *P* Value
Cdc42 guanine nucleotide exchange factor (GEF) 9 (Collybistin)	*Arhgef*	PFC	P0	2.79	**.023**	1.25	**.026**
Aryl hydrocarbon receptor nuclear translocator	*Arnt*	Cer	P0	*	*	1.51	**.024**
		Hipp	P0	*	*	2.08	**.021**
		PFC	P0	*	*	1.45	**.006**
Death-associated protein kinase 1	*Dapk1*	Cer	P0	2.18	**.0051**	1.22	**.013**
DEAD (Asp-Glu-Ala-Asp) box polypeptide 3, Y-linked	*Dby*	Cer	P56	4.00	**.044**	6.71	**.005**
Ephrin B2	*Efnb2*	Hipp	P0	2.33	**.035**	2.41	**.021**
V-erb-a erythroblastic leukemia viral oncogene homolog 4 (avian)	*Erbb4*	Hipp	P0	3.43	**.0055**	2.10	**.048**
Influenza virus NS1A–binding protein	*Ivns1abp*	Cer	P0	*	*	1.22	**.0004**
		Hipp	P0	*	*	1.91	.079
		PFC	P0	*	*	1.02	.86
Myelin transcription factor 1–like	*Myt1l*	Hipp	P0	2.36	**.039**	2.03	.051
Neurexophilin	*Nxph2*	Hipp	P0	4.15	**.0006**	3.50	**.010**
Sema domain, immunoglobulin domain, short basic domain, secreted, (semaphorin) 3A	*Sema3a*	Hipp	P0	3.52	**.018**	3.49	**.013**
SRY-box–containing gene 2	*Sox2*	Cer	P0	2.16	**.008**	1.30	**.010**
Transferrin receptor	*Trfr2*	Cer	P0	2.27	**.045**	1.57	**.026**
Ubiquitously transcribed tetratricopeptide repeat gene, Y chromosome	*Uty*	Cer	P56	3.68	**.033**	5.35	**.018**

Note: Data taken from Fatemi et al.[71] E, embryonic; PD, postnatal date; PFC, prefrontal cotex; Cer, cerebellum; Hipp, hippocampus; qRT-PCR, quantitative real-time polymerase chain reaction; *, not changed in microarray; bold values, $p < 0.05$.

Genetics

The mode of transmission in schizophrenia is unknown and most likely complex and non-Mendelian.[10,80] Chromosomal abnormalities show evidence for involvement of a balanced reciprocal translocation between chromosomes 1q42 and 11q14.3, with disruption of disrupted in schizophrenia 1 and 2 (*DISC1* and *DISC2*) genes on 1q42, being associated with schizophrenia.[80,81] Additionally, an association between a deletion on 22q11, schizophrenia, and velocardiofacial syndrome has been reported.[82] Mice with similar deletions exhibit sensorimotor gating abnormalities.[83]

Linkage and association studies[80,84,85] show 12 chromosomal regions containing 2181 known genes[84] and 9 specific genes[80] as being involved in etiology of schizophrenia.[80] Variations/polymorphisms in 9 genes including neuregulin 1 (*NRG1*), dystrobrevin-binding protein 1 (*DTNBP1*), G72 and G30, regulator of G-protein signaling 4 (*RGS4*), catechol-*O*-methytransferase (*COMT*), proline dehydrogenase (*PRODH*), *DISC1* and *DISC2*, serotonin 2A receptor, and dopamine receptor D3 (*DRD3*) have been associated with schizophrenia (table 2). However, of the various candidate genes, there is no single gene whose genetic association to schizophrenia has been replicated in every study.[86]

Another means of studying the genetic basis of schizophrenia uses the technique of DNA microarray.[87,88] These studies are based on discovering genes either repressed or stimulated significantly in well-characterized postmortem brain tissues from subjects with schizophrenia and matched healthy controls and peripheral lymphocytes obtained from schizophrenic and matched healthy controls and antipsychotic-treated brains of rodents (table 3). Genes involved in drug response or in etiopathogenesis of schizophrenia can be compared and studied to better understand the mechanisms responsible for this illness.[87]

Biological markers consistent with prenatal occurrence of neurodevelopmental insults in schizophrenia include changes in the normal expression of proteins that are involved in early migration of neurons and glia, cell proliferation, axonal outgrowth, synaptogenesis, and apoptosis. Some of these markers have been investigated in studies of various prenatal insults in potential animal models for schizophrenia thus helpful in deciphering the molecular mechanisms for genesis of schizophrenia.[7]

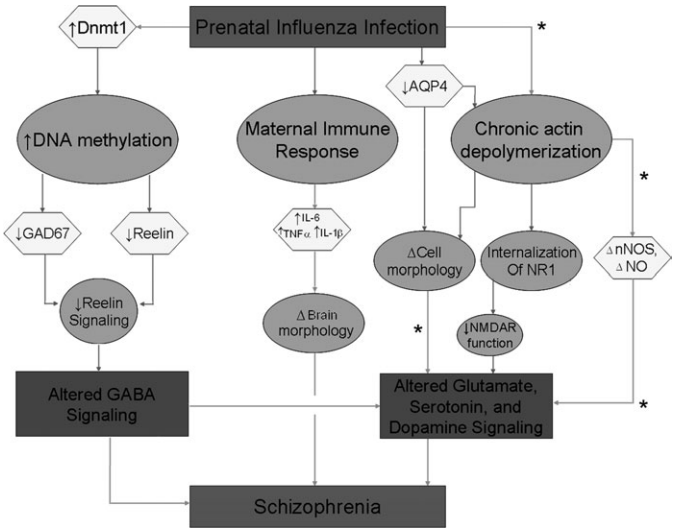

Fig. 2. A Hypothesis of How Prenatal Viral Infection Could Contribute to the Development of Schizophrenia. Prenatal viral infection may lead to (1) activation of DNA methyltransferase 1 (DNMT1) that in turn changes methylation of promoters for a variety of genes leading to altered levels of molecules such as glutamic acid decarboxylase 67-kDa protein (GAD67) and reelin (S.H. Fatemi, unpublished observations).[74,75] These changes may result in abnormal development and altered γ-aminobutyric acid (GABA) signaling and subsequent genesis of schizophrenia; (2) activation of the maternal immune response leading to altered levels of cytokines including interleukin (IL)-1β, IL-6, and tumor necrosis factor α (TNF-α)[44,45,48] that regulate normal brain development.[50–52] Changes may lead to abnormal cortical development and, ultimately, schizophrenia; and (3) altered expression of genes that are involved in cell-cell communication and changes in cell structure due to chronic actin depolymerization (S.H. Fatemi and M. Peoples, unpublished observations, 2007) may lead to dysregulation of multiple signaling systems that have been observed in schizophrenia. *, Pathways that require more substantial support.

Several recent reports implicate various gene families as being involved in pathology of schizophrenia using DNA microarray technology, ie, genes involved in signal transduction,[89–98] cell growth and migration,[91] myelination,[89,99] regulation of presynaptic membrane function,[92,93] and γ-aminobutyric acid–mediated (GABAergic) function.[89,94] By far, the most well-studied and replicated data deal with genes involved in oligodendrocyte- and myelin-related functions. Hakak et al[89] using mostly elderly schizophrenic and matched control dorsolateral prefrontal cortex (DLPFC) homogenates showed downregulation of 5 genes whose expression is enriched in myelin-forming oligodendrocytes, which have been implicated in the formation and maintenance of myelin sheaths. Later, Tkachev et al[99] using area 9 homogenates from Stanley Brain Collection showed significant downregulation in several myelin- and oligodendrocyte-related genes such as proteolipid protein 1,[96] myelin-associated glycoprotein, oligodendrocyte-specific protein CLDN11, myelin oligodendrocyte glycoprotein, myelin basic protein, neuroregulin receptor v-erb-a erythroblastic leukemia viral oncogene homolog 3 (*ERBB3*), transferrin, olig 1, olig 2, and SRY Box 10. [99] Mirnics et al[92] showed downregulation of genes involved in presynaptic function in the PFC such as methylmaleimide-sensitive factor, synapsin II, synaptojanin 1, and synaptotagmin 5. Vawter et al[93] showed downregulation of histidine triad nucleotide–binding protein and ubiquitin-conjugating enzyme E2N. Another important family of genes involved in schizophrenia are genes involved in glutamate and GABAergic function[220,221]. Hakak et al[89] showed an upregulation of several genes involved in GABA transmission, such as GAD65- and 67-kDa protein genes. However, several reports have shown decreases in these proteins in schizophrenia.[97,98,100] Hashimoto et al[94] showed a downregulation of parvalbumin gene, and Vawter et al[93] showed downregulation of glutamate receptor α-amino-3-hydroxyl-5-methyl-4-isoxazoleproprionate (AMPA). Another gene family of import in schizophrenia deals with signal transduction. Hakak et al[89] showed upregulation of several postsynaptic signal transduction pathways known to be regulated by dopamine, consistent with the dopamine hypothesis of schizophrenia[95,101] such as cAMP-dependent protein kinase subunit RII-β and nel-related protein 2. In a similar vein, Mirnics and Lewisl[90] also showed downregulation of RGS4 gene in PFC of schizophrenia. Recently in a study of temporal gyrus, Bowden et al,[102] found that a number of genes related to neurotransmission (*GRIN2B, GRIP2, SYT7*), neurodevelopment (*DAB1, SEMA5A*), and intracellular signaling (*PIK3R1, CACNG2*) were significantly altered.[102] Chung et al[91] showed upregulation of heat shock 70 gene in schizophrenic brain.[91] A number of schizophrenia candidate genes have been found to change in PFC over the course of the life span in brain samples from control subjects via microarray: (1) *RGS4* and glutamate receptor metabotropic 3 (*GRM3*) expression decreased across the age range, (2) *PRODH* and *DARPP32* expression increased with age, and (3) *NRG1*, *ERBB3*, and nerve growth factor receptor showed altered expression during the years of greatest risk for the development of schizophrenia.[103]

Interaction Between Genes and Environment

Genetic risk factors may also interact with obstetric complications to increase risk of schizophrenia,[104–106] and it has been suggested known susceptibility genes for schizophrenia were more likely than randomly selected genes to be regulated by hypoxia/ischemia.[107] Nicodemus et al[108] recently tested whether a set of 13 schizophrenia susceptibility genes thought to be regulated in part by hypoxia statistically interact with obstetric complications. Four genes: v-AKT murine thymoma viral oncogene homolog 1, brain-derived neurotrophic factor, *DTNBP1*, and *GRM3* showed significant interactions,[76] and all 4 have been shown to have neuroprotective roles.[107]

Table 2. Risk Genes for Schizophrenia

Gene	Abbreviation	Locus
Neuregulin	*NRG1*	8p12–p21
Dysbindin	*DTNBP1*	6p22
G72	*G72*	13q34
D-amino acid oxidase	*DAAO*	12q24
RGS4	*RGS4*	1q21–22
Catechol-*O*-methyltransferase	*COMT*	22q11
Proline dehydrogenase	*PRODH*	22q11
Reelin	*RELN*	7q22

Note: Data taken from Sullivan et al,[80] Le-Niculescu et al,[87] and Wedenoja et al.[218].

Table 3. Candidate Genes: Postmortem Studies and Animal Models

Gene	Abbreviation	Postmortem	Animal model
Adenosine A2A receptor	*ADORA2A*	+	+
Apolipoprotein D	*APOD*	+	+
CDC42 guanine nucleotide exchange factor 9	*ARHGEF9*	+	+
Complexin 2	*CPLX2*	+	+
Distal-less homeobox 1	*DLX1*	+	—
Dopamine receptor D1	*DRD1*	+	—
Dopamine receptor D2	*DRD2*	+	+
$GABA_A$ receptor, subunit A1	*GABRA1*	+	—
$GABA_A$ receptor, subunit A5	*GABRA5*	+	+
$GABA_B$ receptor 1	*GABBR1*	+	—
Glutamic acid decarboxylase 2	*GAD2*	+	—
Glial fibrillary acidic protein	*GFAP*	+	+
Glutamate receptor, ionotropic, AMPA1	*GRIA1*	+	—
Glutamate receptor, ionotropic, AMPA2	*GRIA2*	+	—
Myelin and lymphocyte protein	*MAL*	+	—
Myelin basic protein	*MBP*	+	+
Neuronal PAS domain protein 1	*NPAS1*	+	+
Proteolipid protein	*PLP1*	+	—
Reelin	*RELN*	+	+
Regulator of G-protein signaling 4	*RGS4*	+	—
Short stature homeobox 2	*SHOX2*	+	—
Synapsin II	*SYN2*	+	—

Note: Data taken from Fatemi,[5] Fatemi et al,[67] Fatemi et al,[71] and Le-Niculescu et al.[87] AMPA, α-amino-3-hydroxyl-5-methyl-4-isoxazoleproprionate; PAS, PER, ARNT, SIM.

In a study of schizophrenia candidate genes, Schmidt-Kastner et al[107] found that at least 50% were regulated by hypoxia and/or were expressed in the vasculature.[107] These genes included *CHRNA7, COMT, GAD1, NRG1, RELN*, and *RGS4*.[107] The authors proposed that the interaction of genes and "internal" environmental factors, in this case hypoxia, result in developmental perturbations leading to a predisposition to schizophrenia.[107] However, additional external factors would have to come into play postnatally for the full development of schizophrenia.[107]

Another approach to studying the genetic contribution is to examine rare structural variants including microduplications and microdeletions. These have previously been shown to underlie illnesses including neurological and neurodevelopmental syndromes.[109] Two recent reports by Walsh et al[110] and the International Schizophrenia Consortium[111] have used this approach in subjects with schizophrenia. Walsh et al[110] found that novel deletions and duplications of genes were present in 5% of controls compared with 15% of subjects with schizophrenia ($P < .0008$) and 25% of subjects with early-onset schizophrenia ($P < .0001$). The majority of genes identified were disproportionately associated with pathways important for brain development, including synaptic long-term transmission, NRG signaling, axonal guidance, and integrin signaling.[110] A large-scale genome-wide survey of copy number variants (CNVs) performed by the International Schizophrenia Consortium[111] revealed that subjects with schizophrenia were 1.15 times more likely to have a higher rate of CNVs than controls.[111] Associations with schizophrenia were found for large deletions of regions on chromosomes 1, 15, and 22 impacting a number of genes.[111]

Interestingly, 19 of the genes impacted in both articles have also been significantly upregulated or downregulated following prenatal viral infection at embryonic days 9, 16, and 18 with our animal model (table 4) providing further convergence between our model and human genetic data. Two genes, v-erb-a erythroblastic leukemia viral oncogene homolog 4 (*Erbb4*) and solute carrier family 1 (glial high-affinity glutamate transporter), member 3 (*Slc1a3*), have been previously associated with schizophrenia.[112,113] Erbb4 gene codes for a transmembrane tyrosine kinase receptor for NRG1.

Table 4. Novel Structural Variants in Genomic DNA That Delete or Duplicate Genes in Subjects With Schizophrenia and Controls Similar to Genes Significantly Altered Following Prenatal Viral Infection

	Chromosomal Abnormality in Subjects with Schizophrenia				Microarray of Virally Infected Mice			
Name	Gene	Chr	Dup/Del	Disease	Area	Inf Date	PD	Regulation
Ankyrin repeat domain 35[110]	*Ankrd35*	1	Del		Cer Cer Hipp	E9 E16 E16	P56 P56 P56	Up Down Up
B-cell CLL/lymphoma 9[110]	*Bcl-9*	1	Del		Hipp	E16	P0	Up
Lix1-like[111]	*Lix1l*	1	Del		Hipp	E16	P0	Up
v-erb-a erythroblastic leukemia viral oncogene homolog 4 (avian) [111]	*Erbb4*	2	Del	Scz	Cer Hipp PFC Cer Hipp	E16 E16 E16 E18 E18	P14 P0 P56 P56 P0	Up Up Up Up Up
S-phase kinase-associated protein 2 (p45)[111]	*Skp2*	5	Del		Cer	E16	P56	Up
Solute carrier family 1 (glial high-affinity glutamate transporter), member 3 (aka EAAT1)[111]	*Slc1a3*	5	Del	Scz	Cer Hipp PFC	E16 E16 E16	P56 P0 P14	Up Up Up
Cation-chloride cotransporter–interacting protein-1 (Solute carrier family 12 [potassium/chloride transporters], member 9)[111]	*Slc12a9*	7	Dup		PFC	E18	P14	Down
Membrane-associated guanylate kinase, inverted 2[111]	*Magi2*	7	Dup		Hipp	E16	P0	Up
Myeloid/lymphoid or mixed-lineage leukemia 3[111]	*Mll3*	7	Dup		Cer	E16	P56	Up
Putative homeodomain transcription factor 2[111]	*Phtf2*	7	Dup		Hipp	E18	P14	Up
PTK2 protein tyrosine kinase 2[111]	*Ptk2*	8	Dup		Hipp	E16	P0	Up
SWI/SNF–related, matrix-associated, actin-dependent regulator of chromatin, subfamily a, member 2[111]	*Smarca2*	9	Dup		Cer Hipp Cer	E16 E16 E18	P56 P0 P56	Up Up Up
Discs, large homolog 2 (Drosophila)[111]	*Dlg2*	11	Del		Hipp	E16	P0	Up
Kruppel-like factor 13[111]	*Klf13*	15	Del		Cer	E18	P0	Up
Myotubularin-related protein 10[110]	*Mtmr10*	15	Del		Cer	E16	P56	Down
Apoptosis-inducing factor, mitochondrion-associated 3[110]	*Aifm3*	22	Del		PFC	E16	P14	Up
Goosecoid-like[110]	*Gsc1*	22	Del		Cer	E9	P56	Up
*Hpa*II tiny fragments locus 9c[110]	*Ht9c*	22	Del		Cer	E16	P56	Up
Solute carrier family 25 (mitochondrial carrier, citrate transporter), member 1[110]	*Slc25a1*	22	Del		Cer	E16	P14	Up

Note: Data taken from Walsh et al,[110] Stone et al,[111] and S.H. Fatemi, unpublished observations, 2008; Chr, chromosome; Dup, duplication; Del, deletion; Inf, infected; PD, postnatal date; Cer, cerebellum; CLL, chronic lymphocytic leukemia; Hipp, hippocampus; PFC, prefrontal cortex; Scz, schizophrenia; E, embryonic.

This gene is involved in neuron and glial proliferation, differentiation, and migration processes. Binding of Erbb4 and NRG1 leads to NMDA receptor current propagation, a process that is apparently defective in schizophrenia.[112] A recent report shows that polymorphisms in *NRG1* are associated with gray and white matter alterations in childhood-onset schizophrenia,[114] a striking similarity seen in our viral model of schizophrenia, where brain atrophy also occurs in puberty in the exposed mice.[71] The significant increase in *Erbb4* mRNA we have observed may be due to decreases in levels of *NRG1* in the exposed mice.[71] Erbb4 also

interacts with 2 other genes common to both lists: discs, large homolog 2[115] and membrane-associated guanylate kinase, inverted 2 at neuronal synapses.[116]

Slc1a3 codes for a glutamate transporter found on glial cells that functions to regulate neurotransmitter concentrations at excitatory glutamatergic synapses.[117,118] Slc1a3 has been shown to be elevated in thalamus of subjects with schizophrenia.[113] We have also observed elevated levels of Slc1a3 mRNA following prenatal viral infection at E16 in cerebellum at P56, in hippocampus at P0, and in PFC at P14 (S.H.F., unpublished observations, 2008). The 506-kb deletion that disrupts Slc1a3 also disrupts S-phase kinase-associated protein 2 (Skp2), which suppresses apoptosis mediated by DNA damage,[118] and leads to the formation of a chimeric transcript.[110] Interestingly, Skp2 mRNA is similarly elevated in cerebellum at P56 following prenatal viral infection at E16 (S.H. Fatemi, unpublished observations, 2008).

Further analysis of some of the virally regulated brain genes in the exposed progeny that were also similarly disrupted in subjects with schizophrenia by microdeletions or microduplications included (1) *Hpa*II tiny fragments locus 9c, which is involved in nucleic acid metabolism, has recently been shown to be associated with a deficit in sustained attention within schizophrenia in a Taiwanese cohort[119]; (2) protein tyrosine kinase 2, also known as focal adhesion kinase, which is involved in axonal outgrowth[120]; and (3) SWI/SNF–related, matrix-associated, actin-dependent regulator of chromatin, subfamily a, member 2, which is involved in cell differentiation and may be involved in the conversion of oligodendrocyte precursor cells to neural stem cells.[121]

A recent study by Carter[122] has demonstrated the importance of the interaction of genes related to the life cycles of pathogens and schizophrenia. Carter examined 245 schizophrenia candidate genes and found that 21% interact with influenza virus, 22% interact with herpes simplex virus 1, 18% interact with cytomegalovirus, 12.6% interact with rubella, and 16% interact with *Toxoplasma gondii*.[122] These percentages suggest a general overrepresentation of pathogen-related genes in the set of schizophrenia candidate genes. These genes code for ligand-activated receptors (fibroblast growth factor receptor 1 [FGFR1]), adhesion molecules (neuronal cell adhesion molecule 1 [NCAM1]), molecules involved with intracellular traffic (DISC1), among others.[122] Carter[122] suggests that the variability observed in gene association studies may be partly explained by presence/absence of the pathogen that would affect the strength of association.

Brain Pathology

A consistent observation in schizophrenia is the enlargement of the cerebroventricular system. The abnormalities are present at onset of disease, progress slowly,[1] and are unrelated to the duration of illness or treatment regimen.[10] Additionally, cerebroventicular enlargement distinguishes affected from unaffected discordant monozygotic (MZ) twins. A large number of computed tomography and magnetic resonance imaging (MRI) studies indicate lateral and third ventricular enlargement and widening of cortical fissures and sulci.[123] Furthermore, gross brain abnormalities have been identified in DLPFC, hippocampus, cingulate cortex, and superior temporal gyrus.[10,124] Some reports also indicate presence of brain structural abnormalities in individuals at high risk for development of schizophrenia and in unaffected first-degree relatives of subjects with schizophrenia.[125] More recently, studies of white matter tracts show evidence of disorganization and lack of alignment in white fiber bundles in frontal and temporoparietal brain regions in schizophrenia.[126]

Numerous reports have documented the presence of various neuropathologic findings in postmortem brains of patients with schizophrenia.[127] These findings consist of cortical atrophy, ventricular enlargement, reduced volume of amygdala and parahippocampal gyrus, and cell loss and volume reduction in thalamus.[127,128] Several cytoarchitectural studies give credence to the idea of early abnormal laminar organization and orientation of neurons in subjects with schizophrenia including (1) decreased entorhinal cellularity in superficial layers I and II, incomplete clustering of neurons in layer II, and the presence of clusters in deeper layers where they are normally not found[129]; (2) findings similar to those in the entorhinal cortices in PFC and cingulate cortex[127,130,131]; and (3) reduced nicotinamide alanine dinucleotide phosphate (NADPH)-diaphorase (NOS)–positive cells (remnants of the embryonic subplate zone) in cortical layers I and II and increased density in deep layers (subcortical white layer or the putative vestigial subplate zone) in DLPFC and hippocampal and lateral temporal cortices.[132] Specific regions of the frontal cortex are associated with schizophrenia, most notably the DLPFC (for a review see Bunney and Bunney[130]) as well as the orbitofrontal cortex, medial PFC, and ventromedial PFC.[133–135] Changes in the frontal cortex include abnormal translocation of NADPH-diaphorase–positive cells[132] and reduced gray matter volume.[136] Hippocampal abnormalities include disturbed cytoarchitecture, abnormal translocation of NADPH-diaphorase–positive cells, and an overall reduction in volume.[132,137] A greater prevalence of hippocampal shape anomaly, characterized by a rounded shape, medial location, and a deep collateral sulcus, has been found in familial schizophrenia patients.[138] There is also evidence of irregular arrangement of neurons in the entorhinal cortex and disoriented pyramidal cells in CA1–CA3 subfields in subjects with schizophrenia when compared with controls.[139,140] Moreover, there is evidence of biochemical changes, including glutamatergic and GABAergic dysfunction in

the hippocampus of subjects with schizophrenia.[141,220,221] In cerebellum, reduced cell size in Purkinje cells have been observed.[127] Structural MRI studies have shown cerebellar atrophy associated with schizophrenia.[142–144] More recently, however, a study has shown an increase in cerebellar volume in subjects with schizophrenia.[145] Additionally, functional MRI investigations using cognitive tests have demonstrated decreased activation in cerebellum of schizophrenic patients.[146–148]

Several recent reports using MRI and diffusion tensor imaging have shown reduced white and gray matter diffusion anisotropy in patients with schizophrenia.[149–151] In brain white matter, water diffusion is highly anisotropic, with greater diffusion in the direction parallel to axonal tracts. Thus, reduced anisotropy of water diffusion has been proposed to reflect compromised white matter integrity.[150] Reductions in white matter anisotropy reflect disrupted white matter connections, which is consistent with the disconnection model of schizophrenia.[152] Reduced white matter diffusion anisotropy has been observed in prefrontal, parietooccipital, splenium of corpus callosum, arcuate and uncinate fasiculus corpus callosum, parahippocampal gyri, and deep frontal perigenual regions of schizophrenic patients.[150,153–157] It is conceivable that downregulation of genes affecting production of myelin-related proteins, as well as other components of axons, may lay the foundation for white matter abnormalities that develop later in life in subjects who become schizophrenic.[98,99] Recently, the dysregulation of white matter metabolites have been observed in elderly patients with schizophrenia.[158] Compared with healthy subjects, patients with schizophrenia displayed lower *N*-acetyl compounds, lower myoinosotol, and higher glutamate and glutamine in white matter regions.[158] The authors suggest lower *N*-acetyl compounds may indicate reduced neuronal content, lower myoinosotol may suggest decreased glial content or dysfunction, while the elevated glutamate and glutamine could be due to excess neuronal release of glutamate or glial dysfunction in glutamate reuptake.[149] A more recent study by the same group found that elderly patients with schizophrenia with elevated levels of glutamate and glutamine in white matter had lower negative positive and negative syndrome scale (PANSS) scores but greater deficits in executive function.[159] Table 5 summarizes the findings of selected research articles on brain abnormalities observed in subjects with schizophrenia.

Explanatory Capacity of the Neurodevelopmental Model of Schizophrenia

Epidemiology of Schizophrenia

Schizophrenia affects 1% of the adult population in the world.[160] The point prevalence of schizophrenia is about 5/1000 population,[161] and the incidence is about 0.2/1000 per year.[161] This incidence rate was reported to be comparable in most societies[162]; however, recent studies suggest greater variability.[161] Schizophrenia has an earlier onset in males with mean ages of onset of 20 and 25 years in males and females, respectively.[10,161] Reports have indicated, however, that there are no sex differences in the lifetime risk of developing schizophrenia.[163] However, a meta-analysis by Aleman et al[164] of studies of the incidence of schizophrenia found that overall there was evidence for a sex difference in the risk of developing schizophrenia. Interestingly, in countries with a medium development index, the sex difference was not apparent.[164] The authors suggest that factors related to industrialization may play a role.[164] While age of first psychotic episode is generally during adolescence, 23.5% of patients with schizophrenia experience their first episode after age 40 years.[165–167] The prevalence of early adult onset, following extensive remodeling of the brain circuitry during adolescence, rather than onset evenly distributed by age, lends credence to the neurodevelopmental model.

Heritability of Schizophrenia

Emerging evidence points to schizophrenia as a familial disorder with a complex mode of inheritance and variable expression.[10,80,168] While single-gene disorders like Huntington disease have homogenous etiologies, complex-trait disorders like schizophrenia have heterogeneous etiologies emanating from interactions between multiple genes and various environmental insults.[80] Twin studies of schizophrenia suggest concordance rates of 45% for MZ twins and 14% for dizygotic twins.[10,169] Consistent with this, a recent meta-analytic study showed a heritability of 81% for schizophrenia.[169] Despite this high genetic predisposition, an 11% point estimate was suggested for the effects of environmental factors on liability to schizophrenia.[80,169] The interaction of genes and the environment (as discussed in "The Neurodevelopmental Theory of Schizophrenia and Supportive Evidence"), particularly in utero is likely to be very important. Additionally, adoption studies show a lifetime prevalence of 9.4% in the adopted-away offspring of schizophrenic parents vs 1.2% in control adoptees.[170] The adoption studies also clearly show that postnatal environmental factors do not play a major role in etiology of schizophrenia.[80] However, this issue remains controversial and needs to be interpreted carefully in view of the ample support for effects of environment on schizophrenia development.

Drug Abuse and the Development of Schizophrenia

Drug abuse has also been linked to the development of schizophrenia. It has been demonstrated that administration of D-amphetamine (which acts on the dopaminergic tracts) to healthy volunteers leads to production of psychotic symptoms and worsens psychosis in schizophrenic

Table 5. Summary of Selected Brain Abnormalities Observed in Subjects With Schizophrenia

Study	Brain Region	Method	Pathological Change
Northoff et al[123]	Ventricles and cerebral cortex	CT	Lateral and third ventricular enlargement and widening of cortical fissures and sulci
Davis et al[126]	Frontal and temporoparietal regions	MRI	Disorganization and lack of alignment in white fiber bundles
Akbarian et al[132]	Frontal lobe, DLPFC, hippocampus, and lateral and temporal cortices	Histochemical staining	Abnormal translocation of NADPH-diaphorase–positive cells in DLPFC and hippocampal and lateral temporal cortices
Wolf et al[136]	Frontal cortex	VBM	Reduced gray matter volume
Glantz and Lewis,[173] Pierri et al[174]	DLPFC	Histochemical staining	Reduction in pyramidal cell spine density and somal volume
Weiss et al[137]	Hippocampus	MRI	Reduced hippocampal volume
Connor et al[138]	Hippocampus	MRI	Altered hippocampal shape
Arnold et al,[139] Luts et al[140]	Hippocampus	Histochemical staining	Disoriented pyramidal cells in CA1–CA3 subfields
Arnold[129]	Entorhinal cortex	Histochemical staining	Decreased cellularity and incomplete or abnormal clustering
Arnold and Trojanowski[127]	Cerebellum	Histochemical staining	Reduced Purkinje cell size
Uematsu et al,[142] DeLisi et al,[143] Nopoulos et al[144]	Cerebellum	MRI	Cerebellar atrophy
Goldman et al[145]	Cerebellum	MRI	Increased cerebellar volume
Ardekani et al[150]	Corpus callosum, left superior temporal gyrus, parahippocampal gyri, middle temporal gyri, inferior parietal gyri, medial occipital lobe, and the deep frontal perigenual region	MRI	Reduced fractional anisotropy
Kubicki et al[151]	Cingulate fasciculus	DTI	Reduced area and fractional anisotropy
Buchsbaum et al[153]	Prefrontal cortex	MRI	Reduced fractional anisotropy
Lim and Helpern[149]	Prefrontal cortex and right parietal-occipital region	DTI	Reduced fractional anisotropy
Foong et al[155]	Corpus callosum	DTI	Reduced fractional anisotropy
Agartz et al[156]	Splenium of the corpus callosum	DTI	Reduced fractional anisotropy
Burns et al[157]	Left uncinate fasciculus and left arcuate fasciculus	DTI	Reduced fractional anisotropy

Note: CT, computed tomagraphy; MRI, magnetic resonance imaging; DLPFC, dorsolateral prefrontal cortex; VBM, voxel-based morphometry; DTI, diffusion tensor imaging.

subjects.[10] Moreover, heavy cannabis use in adolescence may lead to the development of later schizophrenia and that this is mediated by dopamine.[171] However, hallucinogens like lysergic acid diethylamide (LSD) or psilocybin (acting on serotonin system) or dissociative anesthetics like ketamine or phencyclidine (acting on glutamate system) also cause psychotic symptoms[10,172] suggesting that alterations of the dopaminergic system alone are not solely responsible for the development of schizophrenia.

Pyramidal Cell Abnormalities and Schizophrenia

As mentioned in "The Neurodevelopmental Theory of Schizophrenia and Supportive Evidence," there are numerous neuroanatomical deficits in the brains of schizophrenic subjects. Glantz and Lewis[173] observed that pyramidal cells located in layer III of the DLPFC of subjects with schizophrenia exhibited a 23% reduction in spine density when compared with normal controls suggesting a decrease in excitatory inputs to these cells.[173]

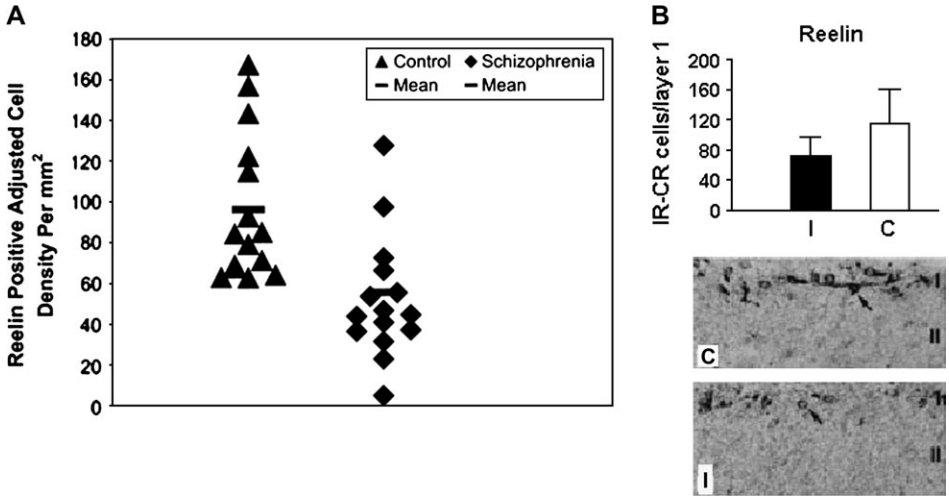

Fig. 3. Reelin is Reduced in Hippocampus of Individuals With Schizophrenia and in Cerebral Cortex Following Prenatal Viral Infection. A. The values expressed on the y-axis are reelin-positive adjusted cell densities per square millimeter localized to hippocampal CA4 areas in control and schizophrenic subjects. The number of brains used is 15 (control) and 15 (schizophrenic). Each point is the mean for 2–4 sections analyzed per brain. A crossbar localized over each scatter plot represents mean Reelin-positive adjusted cell density value per group. Mean values for schizophrenic subjects are significantly reduced when compared with control values (analysis of variance, $P < .05$). B. The top panel shows a graph depicting the hemispheric Reelin-positive Cajal-Retzius (CR) cell counts in layer I of the cortex of prenatally infected (I) and sham-infected control (C) animals. The number of Reelin-positive CR cells was significantly reduced in infected brains compared with control brains ($P < .0001$). The lower panel shows light micrographs of layer I–II in coronal sections of prenatally infected and sham-infected cortex. Originally published in Fatemi et al.[67,179]

These same cells also exhibit a 9.2% reduction in somal volume.[174] Taken together, the authors conclude that these findings indicate disruption of the thalamocortical and corticocortical circuits.[174] As with other alterations including reduced fractional anisotropy of the white matter or altered hippocampal volume or shape, these changes in pyramidal cells suggest neurodevelopmental dysfunction.[222]

The Role of the Reelin and GABAergic Signaling Systems in Schizophrenia

Several studies now implicate the pathological involvement of *RELN* gene or its protein product in schizophrenia. Reelin helps in normal lamination of the brain during embryogenesis and affects synaptic plasticity in adulthood.[5,175,176] Impagnatiello et al[177] used northern and western blotting and immunocytochemistry to show reductions in reelin mRNA and protein in cerebellar, hippocampal, and frontal cortices of patients with schizophrenia and psychotic bipolar disorder. Reduction in reelin was associated with significant decreases in GAD67-kDa protein in the same postmortem brains.[178] A later immunocytochemical report[179] showed significant reductions in reelin immunoreactivity in schizophrenic and bipolar patients. However, these authors detected similar decreases in hippocampal reelin protein levels in nonpsychotic bipolar and depressed subjects, suggesting that reelin deficiency may not be limited to subjects with psychosis alone.[179,223] Fatemi et al[98] subsequently demonstrated significant reductions in Reelin, as well as GAD65-kDa and GAD 67-kDa proteins, in cerebella of subjects with schizophrenia, bipolar disorder, and major depression[98,180] as well as in mice following prenatal viral infection (figure 3). Further confirmatory data relating to Reelin abnormalities in brains of schizophrenic patients were demonstrated by Eastwood et al,[181] who showed a trend for reduction in Reelin mRNA in cerebella of schizophrenic subjects; these reductions in Reelin mRNA correlated negatively with semaphorin 3A. The authors suggested that these findings were consistent with an early neurodevelopmental origin for schizophrenia and that the reciprocal changes in Reelin and semaphorin 3A may be indicative of a mechanism that affects the balance between inhibitory and trophic factors regulating synaptogenesis.[181]

Effects of Various Antipsychotics on Brain Genes Involved in Neurodevelopment of Schizophrenia

Pharmacotherapy is the primary mode of treatment for the psychotic symptoms of schizophrenia. All drugs currently used to treat schizophrenia mediate their actions through the dopamine D2 receptor.[182] With the exception of aripiprazole, which acts as a partial agonist, both typical and atypical antipsychotics are antagonists of the D2 receptor.[183–185] Dopamine hyperactivity may contribute to psychotic symptoms and that dopamine antagonists like chlorpromazine treat the psychotic symptoms.[10]

Clozapine is a dibenzodiazepine and the prototype for most of atypical antipsychotics (agents that may treat positive, negative, or cognitive symptoms of schizophrenia have decreased liability for extrapyramidal symptoms (EPS) and tardive dyskinesia [TD], may be effective for a proportion of treatment-nonresponsive patients and exhibit greater 5HT2 over D2 receptor antagonism and do not cause hyperprolactinema).[186,187] Clozapine has been shown to be effective in treatment-resistant schizophrenia.[188] Thus, clozapine remains the only antipsychotic agent to date that is Food and Drug Administration approved for treatment-resistant schizophrenia.[189] Additionally, other studies have shown superiority of clozapine vs typical agents in treatment of total psychopathology, EPS, and TD and categorical response to treatment.[124] Clozapine reduces positive, negative, and cognitive symptoms of schizophrenia without causation of EPS, TD, or hyperprolactinemia.[10] Additionally, clozapine has been shown to reduce depression and suicidality.[10,124]

The time course over which antipsychotic agents take effect is variable. In an analysis of studies measuring antipsychotic response during the first 4 weeks of treatment, Agid et al[190] found that there was a reduction in total scores of the Brief Psychiatric Rating Scale and the Positive and Negative Syndrome Scale (PANSS) of 13.8% during week 1, 8.1% during week 2, 4.2% during week 3, and 4.7% during week 4. The authors hold that these results reject the ''delayed onset'' model of antipsychotic action; rather, antipsychotic response begins within the first week and accumulates over time.[190] However, Emsley et al,[191] using a benchmark of a 20% improvement in total score on the PANSS for clinical response, found that 22.5% of subjects with first-episode schizophrenia did not achieve clinical response until 4 weeks of treatment or later. Taken together, these studies demonstrate the variability of time to antipsychotic response.

It has been hypothesized that antipsychotic agents affect various brain genes, leading to changes in synaptic structure and function that may underlie clinical response.[192] Olanzapine is a second-generation antipsychotic agent that, like clozapine, exhibits greater 5HT2A than D2 antagonism[193] but does not share clozapine's propensity for agranulocytosis. One of the important genes upregulated by chronic olanzapine treatment is *Reln*[88] (figure 4). Recent reports show that Reelin receptor, apolipoprotein E receptor 2 (ApoER2), interacts with and alters, the conformation of NMDA receptors, NR2A and NR2B.[175] Additionally, Reelin induces tyrosine phosphorylation of NR2A and NR2B receptors in hippocampal tissue,[175] thus modulating NMDA receptor activity and synaptic plasticity in the hippocampus. Supporting evidence for the potential role of olanzapine in enhancing neuroplasticity was recently shown by Lieberman et al,[194] who demonstrated a cessation of brain gray matter loss in brains of patients with schizophrenia that were treated with olanzapine for 12 weeks and not in those treated for the same time period with haloperidol. Additionally, Wang and Deutch[195] have also shown that olanzapine prevented decreases in spine density of basilar

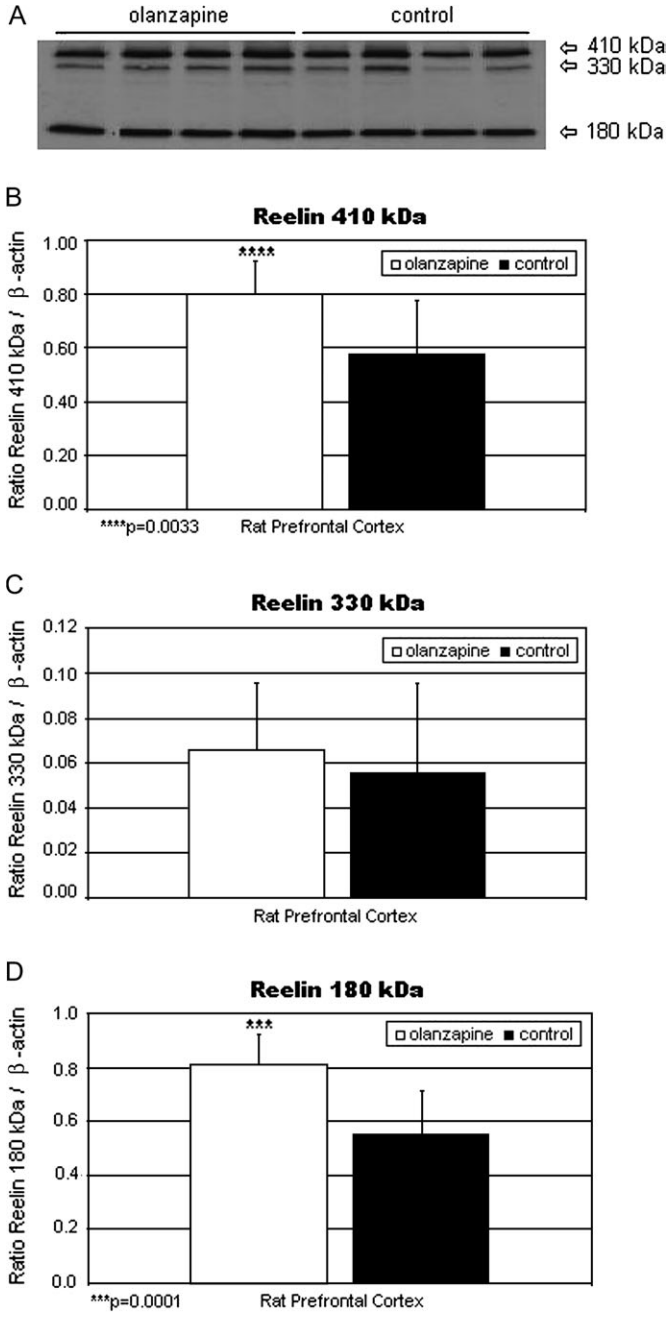

Fig. 4. (A) Reelin Bands of 410, 330, and 180 kDa From the Prefrontal Cortex Homogenates (70 μg protein per lane) of Representative Olanzapine-Treated and Control Rats Are Shown. Mean Reelin 410 (B), 330 (C), and 180 (D) kDa/β-actin ratios for olanzapine-treated (filled histogram bars) and control rats (unfilled histogram bars) are shown. Levels of Reelin 410 kDa/β-actin (B) and Reelin 180 kDa/β-actin (D) were significantly increased vs controls ($P = .0033$ and .0001, respectively). Reelin 330 kDa/β-actin (C) was nonsignificantly increased vs controls. Origianlly published in Fatemi et al.[88]

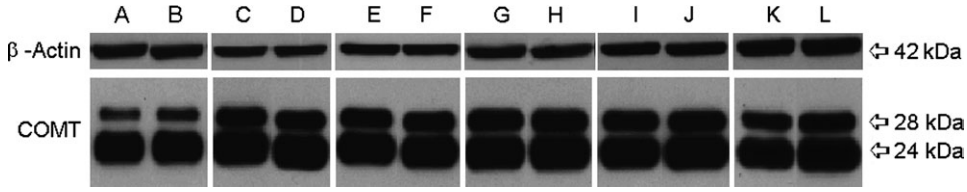

Fig. 5. Effects of Psychotropic Agents on Catechol-*O*-Methyltransferase (COMT) Expression in Rat Frontal Cortex. A, C, E, G, I, and K correspond to protein levels from frontal cortices of clozapine-, fluoxetine-, haloperidol-, lithium-, olanzapine-, and sodium valproate–treated rat brains, respectively. B, D, F, H, J, and L correspond to protein levels from frontal cortices of saline-treated rat brains, respectively. Originally published in Fatemi and Folsom[200].

dendrites on layers II, III, and IV of PFC pyramidal neurons in rats lesioned with 6-hydroxydopamine. Finally, olanzapine, but not haloperidol, increased expression of the polysialilated form of neural cell adhesion molecule in rat PFC, suggesting a possible role for this molecule in the efficacy of olanzapine.[196] NCAM appears early in development and is important during brain morphogenesis.

In recent years, COMT has drawn much interest as a modulator of PFC function, cognitive abilities, and the genetic disposition toward schizophrenia. COMT metabolizes catecholamines[197] and is known to modulate dopamine levels in the PFC.[198,199] Recently, our laboratory[200] conducted experiments testing a number of atypical antipsychotics, mood stabilizers, and antidepressants (clozapine, fluoxetine, haloperidol, lithium, olanzapine, and valproic acid [VPA]) to investigate which genes and proteins were affected by chronic treatment of the above agents. Rats were randomly assigned to 1 of the 6 drug groups or sterile saline and administered drug or diluent for 21 days. Microarray results showed a significant ($P < .05$), 2-fold decrease in COMT in PFC in all drug-treatment groups (except for olanzapine) when compared with controls. Protein levels for the 28-kDa membrane-bound isoform of COMT were significantly downregulated in VPA-treated PFC ($P = .0073$)[200] (figure 5). Protein levels for the 240kDa cytosolic isoform of COMT were significantly downregulated in PFC by clozapine ($P = .014$), lithium ($P = .0006$), olanzapine ($P = .046$), and VPA ($P = .0073$) and were significantly upregulated by fluoxetine ($P = .0063$) [200] (figure 5). In summary, as is evident (vide supra), various antipsychotics exert their clinical actions not only through classical neurotransmitters but also via numerous brain genes that may explain the variable course of clinical response. Some of these genes may also be involved in etiology of schizophrenia (e.g. Reelin).

Evidence in Support of Other Models of Schizophrenia

In addition to the neurodevelopmental model, there are alternative models that have been used to explain the etiology of schizophrenia. It is likely that due to the heterogeneous nature of schizophrenia that multiple factors interact to produce the disease state such as disruptions in the dopaminergic, serotonergic, and glutamatergic systems as well as neurodegenerative changes. With regard to epidemiology, a number of social factors have been shown to increase the risk of schizophrenia including urban birth and upbringing,[201] quality of maternal-child relationship,[202,203] and migration[204], a risk that increases when the immigrant group is a small minority indicating that isolation and lack of support may be important factors. An alternative explanation, however, may be that urban birth and migration may well be consistent with the neurodevelopmental hypothesis in that these represent, respectively, an environment in which one is exposed to more pathogens and an environment in which one may have not developed native antibodies or other resistances to pathogens. Abuse of drugs that affect the dopaminergic (amphetamine, cannabis), glutamatergic (PCP), or serotonergic (LSD) systems also may lead to psychotic symptoms and the development of schizophrenia. While many brain imaging and postmortem studies have yielded structural differences between subjects with schizophrenia and healthy controls, there are other reports showing no differences between schizophrenic patients and controls.[205] Moreover, there is debate as to whether the observed changes represent developmental or neurodegenerative changes or the result of antipsychotic medications.[206]

Critics of the neurodevelopmental model claim that it does not fully account for a number of features of schizophrenia, including the long gap between neurodevelopmental insult and the development of symptoms, the progressive clinical deterioration observed in some patients, and evidence of progressive changes in certain ventricular and cortical brain structures.[1,207–209] Longitudinal studies have demonstrated evidence of an increase in ventricular volume over a period of 2–4 years among first-episode patients.[143,210] Moreover, a decline in frontal lobe volume and posterior superior temporal gray matter volume over a period of 4 years has been reported in patients with chronic schizophrenia.[211] A mechanism to explain the progressive elements of schizophrenia is apoptosis, or programmed cell death (reviewed by Jarskog et al[212]), especially synaptic apoptosis in which apoptosis is localized to distal neurites without inducing immediate neuronal death.[213] In a series of studies of postmortem temporal cortex, Jarskog et al[214] found

reduced expression of Bcl-2, a molecule that protects against apoptosis, in schizophrenic brains. A further study showed that the ratio of proapoptotic molecule Bax to Bcl-2 was increased in the same region, suggesting that these neurons were receptive to apoptotic stimuli.[215] Interestingly, caspase 3, the caspase molecule most associated with apoptosis in the CNS[215] is not upregulated in temporal cortex of subjects with schizophrenia,[216] suggesting that chronic apoptosis is not taking place, in contrast to classic neurodegenerative disorders.[216] The vulnerability of neurons to proapoptotic insults such as oxidative stress and glutamate excitotoxicity could lead to selective dendritic and synaptic losses observed with schizophrenia.[212,222] However, the neurodegenerative model has been critiqued by Weinberger and McClure.[206] The authors point out that there is a lack of expression of genes involved with DNA fragmentation and response to injury from postmortem studies.[206] Moreover, longitudinal studies of cognitive function, which would serve as a measure of cortical neuronal system integrity, do not support a progression of loss of function that would be expected by the neurodegenerative hypothesis.[218]

A means to test for alternate theories to the neurodevelopmental model is through our animal model of prenatal viral infection. Longitudinal studies, in which animals are infected at specific gestational periods and then followed through late adulthood, with brains collected at specific postnatal time points, could help establish whether alternate models are valid. If important genes that have been linked to schizophrenia are not affected at early time points such as birth, childhood, adolescence, or early adulthood but are only turned on or off in mid-late adulthood, it would provide evidence against the neurodevelopmental model. Brain imaging experiments on animals from the same studies could help establish whether there is analagous progressive changes in ventricular or cortical structures observed in subjects with schizophrenia, providing evidence for the neurodegenerative model.

Conclusions

The vast majority of evidence supports a neurodevelopmental model of schizophrenia genesis. Evidence from genetic studies suggest a high degree of heritability of schizophrenia and point to a number of potential candidate genes that may be perturbed early in development leading ultimately to the development of psychotic symptoms. Genes involved with cell migration, cell proliferation, axonal outgrowth, myelination, synaptogenesis, and apoptosis are affected in subjects with schizophrenia, pointing to neurodevelopmental insults. Imaging studies have shown differences between the brains of subjects with schizophrenia and normal controls in a number of brain regions including the PFC, cerebellum, hippocampus, and amygdala. There is strong evidence from epidemiological studies and animal models that viral infection during pregnancy increases the risk for schizophrenia in the offspring. The presence of neurological soft signs in children who later develop schizophrenia also points to a neurodevelopmental etiology of schizophrenia.

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References

1. Rapoport JL, Addington AM, Frangou S, Psych MR. The neurodevelopmental model of schizophrenia: update 2005. *Mol Psychiatry*. 2005;10:434–449.
2. Fatemi SH. Prenatal viral infection, brain development and schizophrenia. In: Fatemi SH, ed. *Neuropsychiatric Disorders and Infection.* London, UK: Taylor and Francis; 2005.
3. Krapelin E. *Psychiatrie. 4th ed. Ein lehrbuch für studirende und ärzte [Psychiatry 4th Ed: A Textbook for Students and Physicians].* Leipzeig, Germany: Abel; 1893.
4. Brown AS, Begg MD, Gravenstein S, et al. Serologic evidence of prenatal influenza in the etiology of schizophrenia. *Arch Gen Psychiatry*. 2004;61:774–780.
5. Fatemi SH. Schizophrenia. In: Fatemi SH, Clayton PJ, eds. *Medical Basis of Psychiatry.* New York, NY: Humana Press; 2008:85–108.
6. Slater E, Beard AW, Glithero E. The schizophrenia-like psychoses of epilepsy. *Br J Psychiatry*. 1963;109:95–150.
7. Carpenter WT, Koenig JI. The evolution of drug development in schizophrenia: past issues and future opportunities. *Neuropsychopharmacology*. 2008;33:2061–2079.
8. Keshavan MS. Development, disease and degeneration in schizophrenia: a unitary pathophysiological model. *J Psychiatr Res*. 1999;33:513–521.
9. Keshavan MS, Hogarty GE. Brain maturational processes and delayed onset in schizophrenia. *Dev Psychopathol*. 1999;11:525–543.
10. Meltzer HY, Fatemi SH. Schizophrenia and other psychotic disorders. In: Ebert MH, Loosen PT, Nurcombe B, eds. *Current Diagnosis and Treatment in Psychiatry.* Norwalk, Conn: Appleton and Lange; 2000:260–277.
11. Lloyd T, Dazzan P, Dean K, et al. Minor physical anomalies in patients with first-episode psychosis: their frequency and diagnostic specificity. *Psychol Med*. 2008;38:71–77.
12. Bracha HS, Torrey EF, Gottesman II, Bigelow LB, Cunniff C. Second-trimester markers of fetal size in schizophrenia: a study of monozygotic twins. *Am J Psychiatry*. 1992;149:1355–1361.
13. Avila MT, Sherr J, Valentine LE, Blaxton TA, Thaker GK. Neurodevelopmental interactions conferring risk for schizophrenia: a study of dermatoglyphic markers in patients and relatives. *Schizophr Bull*. 2003;29:595–605.
14. Fish B, Marcus J, Hans S, Auerbach JG, Perdue S. Infants at risk for schizohrenia: sequelae of a genetic neurointegrative defect. *Arch Gen Psychiatry*. 1992;49:221–235.

15. Walker EF. Developmentally moderated expressions of the neuropathology underlying schizophrenia. *Schizophr Bull*. 1994;20:453–480.
16. Barkus E, Stirling J, Hopkins R, Lewis S. The presence of neurological soft signs along the psychosis proneness continuum. *Schizophr Bull*. 2006;32:573–577.
17. Compton MT, Bollini AM, McKenzie ML, et al. Neurological soft signs and minor physical anomalies in patients with schizophrenia and related disorders, their first-degree biological relatives, and non-psychiatric controls. *Schizophr Res*. 2007;94:64–73.
18. Niemi LT, Suvisaari JM, Tuulio-Henriksson A, Lonnqvist JK. Childhood developmental abnormalities in schizophrenia: evidence from high-risk studies. *Schizophr Res*. 2003;60: 239–258.
19. Ott SL, Spinelli S, Rock D, Roberts S, Amminger GP, Erlenmeyer-Kimling L. The New York High-Risk Project: social and general intelligence in children at risk for schizophrenia. *Schizophr Res*. 1998;31:1–11.
20. Keshavan MS, Gilbert AR, Diwadkar VA. Neurodevelopmental theories. In: Lieberman JA, Stroup TS, Perkins DO, eds. *The American Psychiatric Publishing Textbook of Schizophrenia.* Washington, DC: American Psychiatric Publishing Inc.; 2006:69–84.
21. Gilmore JH, Murray RM. Prenatal and perinatal factors. In: Lieberman JA, Stroup TS, Perkins DO, eds. *The American Psychiatric Publishing Textbook of Schizophrenia.* Washington, DC: American Psychiatric Publishing Inc.; 2006:55–68.
22. Lewis DA. Retroviruses and the pathogenesis of schizophrenia. *Proc Natl Acad Sci USA*. 2001;94:4293–4294.
23. Karlsson H, Bachmann S, Schroder J, McArthur J, Torrey EF, Yolken RH. Retroviral RNA identified in the cerebrospinal fluids and brains of individuals with schizophrenia. *Proc Natl Acad Sci USA*. 2001;98:4634–4639.
24. Hare EH, Price JS, Slater E. Schizophrenia and season of birth. *Br J Psychiatry*. 1972;120:125–126.
25. Machon RA, Mednick SA, Schulsinger F. The interaction of seasonality, place of birth, genetic risk and subsequent schizophrenia in a high risk sample. *Br J Psychiatry*. 1983; 143:383–388.
26. Susser ES, Brown AS, Gorman JM. *Prenatal Exposures in Schizophrenia.* Washington, DC: American Psychiatric Press; 1999.
27. Boyd JH, Pulver AE, Stewart W. Season of birth: schizophrenia and bipolar disorder. *Schizophr Bull*. 1986;12:173–186.
28. Pallast EG, Jongbloet PH, Straatman HM. Excess seasonality of births among patients with schizophrenia and seasonal ovopathy. *Schizophr Bull*. 1994;20:269–276.
29. Pulver AE, Liang KY, Wolyniec PS. Season of birth among siblings of schizophrenic patients. *Br J Psychiatry*. 1992;160: 71–75.
30. Mednick SA, Machon RA, Huttunen MO. Adult schizophrenia following prenatal exposure to an influenza epidemic. *Arch Gen Psychiatry*. 1988;45:189–192.
31. Stober G, Franzek E, Beckmann J. The role of maternal infectious diseases during pregnancy in the aetiology of schizophrenia in offspring. *Eur Psychiatry*. 1992;7:147–152.
32. Wright P, Rakei N, Rifkin L, Murray RM. Maternal influenza, obstetric complications, and schizophrenia. *Am J Psychol*. 1995;152:1714–1720.
33. Mednick SA, Huttunen MO, Macon RA. Prenatal influenza infections and adult schizophrenia. *Schizophr Bull*. 1994;20:263–267.
34. Brown AS, Cohen P, Greenwald S, Susser E. Nonaffective psychosis after prenatal exposure to rubella. *Am J Psychiatry*. 2000;157:438–443.
35. Brown AS. Prenatal infection as a risk factor for schizophrenia. *Schizophr Bull*. 2006;32:200–202.
36. Nakai Y, Itoh M, Mizuguchi M, et al. Apoptosis and microglial activation in influenza encephalopathy. *Acta Neuropathol*. 2003;105:233–239.
37. Aronsson F, Robertson B, Ljunggren HG, Kristensson K. Invasion and persistence of the neuroadapted influenza virus A/WSN/33 in the mouse olfactory system. *Viral Immunol*. 2003;16:415–423.
38. Aronsson F, Lannebo C, Paucar M, Brask J, Kristensson K, Karlsson H. Persistence of viral RNA in the brain of offspring to mice infected with influenza A/WSN/33 during pregnancy. *J Neurovirol*. 2002;8:353–357.
39. Chen BY, Chang HH, Chiou HL, Lin DP. Influenza B virus-induced brain malformations during early chick embryogenesis and localization of tRNA in specific areas. *J Biomed Sci*. 2004;11:266–274.
40. Aronsson F, Karlsson H, Ljunggren HG, Kristensson K. Persistence of the influenza A/WSN/33 virus RNA at midbrain levels of immunodefective mice. *J Neurovirol*. 2001;7:117–124.
41. Levine J, Buchman CA, Fregien N. Influenza A virus infection of human Schwann cells in vitro. *Acta Otolaryngol*. 2003;123:41–45.
42. Brask J, Owe-Larsson B, Hill RH, Kristensson K. Changes in calcium currents and GABAergic spontaneous activity in cultured rat hippocampal neurons after a neurotropic influenza A virus infection. *Brain Res Bull*. 2001;55: 421–429.
43. Pearce BD, Valadi NM, Po CL, Miller AH. Viral infection of developing GABAergic neurons in a model of hippocampal disinhibition. *Neuroreport*. 2000;11:2433–2438.
44. Hillier SL, Witkin SS, Krohn MA, Watts DH, Kiviat NB, Eschenbach DA. The relationship of amniotic fluid cytokines and preterm delivery, amniotic fluid infection, histologic chorioamnionitis, and chorioamnion infection. *Am J Obstet Gynecol*. 1993;81:941–948.
45. Fortunado SJ, Menon RP, Swan KF, Menon R. Inflammatory cytokines (interleukins 1.6.8 and tumor necrosis factor-α) release from cultured fetal membranes in response to endotoxic lipopolysaccharide mirrors amniotic fluid. *Am J Obstet Gynecol*. 1996;174:1855–1862.
46. Fidel PL, Jr, Romero R, Wolf N, et al. Systemic and local cytokine profiles in endotoxin-induced preterm parturition in mice. *Am J Obstet Gynecol*. 1994;170:1467–1475.
47. Urkabo A, Jarskog LF, Lieberman JA, Gilmore JH. Prenatal exposure to maternal infection alters cytokine expression in the placenta, amniotic fluid, and fetal brain. *Schizophr Res*. 2001;47:27–36.
48. Yoon BH, Romero R, Moon J, et al. Differences in the fetal interleukin-6 response to microbial invasion of the amniotic cavity between term and preterm gestation. *J Matern Fetal Neonatal Med*. 2003;13:32–38.
49. Buka SL, Tsuang MT, Torrey EF, Klebanoff MA, Wagner RL, Yolken RH. Maternal cytokine levels during pregnancy and adult psychosis. *Brain Behav Immun*. 2001;15: 411–420.
50. Merrill JE. Tumor necrosis factor alpha, interleukin 1 and related cytokines in brain development: normal and pathological. *Dev Neurosci*. 1992;14:1–10.

51. Mehler MF, Kessler JA. Growth factor regulation of neuronal development. *Dev Neurosci*. 1994;16:180–195.
52. Mehler MF, Kessler JA. Hematolymphopoietic and inflammatory cytokines in neural development. *Trends Neurosci*. 1997;20:357–365.
53. Burns TM, Clough JA, Klein RM, Wood GW, Berman NE. Developmental regulation of cytokine expression in the mouse brain. *Growth Factors*. 1993;9:253–258.
54. Gadient RA, Otten U. Expression of interleukin-6 (IL-6) and interleukin-6 receptor (IL-6R) mRNAs in rat brain during postnatal development. *Brain Res*. 1994;637:10–14.
55. Pousset F. Developmental expression of cytokine genes in the cortex and hippocampus of the rat central nervous system. *Dev Brain Res*. 1994;81:143–146.
56. Mousa A, Seiger A, Kjaeldgaard A, Bakhiet M. Human first trimester forebrain cells express genes for inflammatory and anti-inflammatory cytokines. *Cytokine*. 1999;11:55–60.
57. Dziegielewska KM, Moller JE, Potter AM, Ek J, Lane MA, Saunders NR. Acute-phase cytokines IL-1β and TNFα in brain development. *Cell Tissue Res*. 2000;299:235–245.
58. McDuffie RS, Dabies JK, Leslie KK, Sherman MP, Gibbs RS. A randomized control trail of interleukin-1 receptor antagonist in a rabbit model of ascending infection in pregnancy. *Infect Dis Obstet Gynecol*. 2001;9:233–237.
59. Menon R, Swan KF, Lyden TW, Rote NS, Fortunado SJ. Expression of inflammatory cytokines (interleukin-1β and interleukin-6) in amniochorionic membranes. *Am J Obstet Gynecol*. 1995;172:493–500.
60. Shi L, Fatemi SH, Sidwell RW, Patterson PH. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. *J Neurosci*. 2003;23:297–302.
61. Borrell J, Vela JM, Arevalo-Martin A, Molina-Holgado E, Guaza C. Prenatal immune challenge disrupts sensorimotor gating in adult rats: implications for the etiopathogenesis of schizophrenia. *Neuropsychopharmacology*. 2002;26:204–215.
62. Zuckerman L, Weiner I. Post-pubertal emergence of disrupted latent inhibition following prenatal immune activation. *Psychopharmacology*. 2003;169:308–313.
63. Susser E, Neugebauer R, Hoek HW, et al. Schizophrenia after prenatal famine. Further evidence. *Arch Gen Psychiatry*. 1996;53:25–31.
64. Brown AS, Susser E. Prenatal nutritional deficiency and risk of adult schizophrenia. *Schizophr Bull*. 2008;34:1054–1063.
65. Hollister JM, Laing P, Mednick SA. Rhesus incompatibility as a risk factor for schizophrenia in male adults. *Arch Gen Psychiatry*. 1996;53:19–24.
66. Wright P, Murray RM. Schizophrenia: prenatal influenza and autoimmunity. *Ann Med*. 1993;25:497–502.
67. Fatemi SH, Emamian ES, Kist D, et al. Defective corticogenesis and reduction in Reelin immunoreactivity in cortex and hippocampus of prenatally infected neonatal mice. *Mol Psychiatry*. 1999;4:145–154.
68. Meyer U, Nyffeler M, Yee BK, Knuesel I, Feldon J. Adult brain and behavioral pathological markers of prenatal immune challenge during early/middle and late fetal development in mice. *Brain Behav Immun*. 2008;22:469–486.
69. Geyer MA, Braff DL, Swerdlow NR. Startle-response measures of information processing in animals: relevance to schizophrenia. In: Haug M, Whalen RE, eds. *Animal Models of Human Emotion and Cognition.* Washington, DC: American Psychiatric Press; 1999:103–116.
70. Fatemi SH, Earle JA, Kanodia R, et al. Prenatal viral infection leads to pyramidal cell atrophy and macrocephaly in adulthood: implications for genesis of autism and schizophrenia. *Cell Mol Neurobiol*. 2002;22:25–33.
71. Fatemi SH, Reutiman TJ, Folsom TD, et al. Maternal infection leads to abnormal gene regulation and brain atrophy in mouse offspring: implications for genesis of neurodevelopmental disorders. *Schizophr Res*. 2008;99:56–70.
72. Fatemi SH, Pearce DA, Brooks AI, Sidwell R. Prenatal viral infection in mouse causes differential expression of genes in brains of mouse progeny: a potential animal model for schizophrenia and autism. *Synapse*. 2005;57:91–99.
73. Fatemi SH, Reutiman TJ, Folsom TD, Sidwell R. The role of cerebellar genes in pathology of autism and schizophrenia. *Cerebellum*. 2008; In press.
74. Veldic M, Caruncho HJ, Liu WS. DNA-methyltransferase 1 mRNA is selectively overexpressed in telencephalic GABAergic interneurons of schizophrenia brains. *Proc Natl Acad Sci USA*. 2004;101:348–353.
75. Costa E, Dong E, Grayson DR, Guidotti A, Ruzicka W, Veldic M. Reviewing the role of DNA (cytosine-5) methyltransferase overexpression in the cortical GABAergic dysfunction associated with psychosis vulnerability. *Epigenetics*. 2007;2:29–36.
76. Papadopoulos MC, Manley GT, Krishna S, Verkman AS. Aquaporin-4 facilitates reabsorption of excess fluid in vasogenic brain edema. *FASEB J*. 2004;18:1291–1293.
77. Verkman AS, Binder DK, Bloch O, Auguste K, Papadopoulos MC. Three distinct roles of aquaporin-4 in brain function revealed by knockout mice. *Biochim Biophys Acta*. 2006;1758:1085–1093.
78. Fatemi SH, Folsom TD, Reutiman TJ, Sidwell RW. Viral regulation of aquaporin 4, connexin 43, microcephalin and nucleolin. *Schizophr Res*. 2008;98:163–177.
79. Fatemi SH, Cuadra AE, El-Fakahany EE, Thuras P. Prenatal viral infection causes alterations in nNOS expression in developing mouse brains. *Neuroreport*. 2000;11:1493–1496.
80. Sullivan PF, Owen MJ, O'Donovan MC, Freedman MD. Genetics. In: Lieberman JA, Stroup TS, Perkins DO, eds. *The American Psychiatric Publishing Textbook of Schizophrenia.* Washington, DC: American Psychiatric Publishing Inc; 2006:39–54.
81. Owen MJ, Craddock N, O'Donovan MC. Schizophrenia: genes at last? *Trends Genet*. 2005;21:518–525.
82. Murphy KC. Schizophrenia and velo-cardio-facial syndrome. *Lancet*. 2002;359:426–430.
83. Paylor R, McIlwain KL, McAninch R, et al. Mice deleted for the DiGeorge/velocardiofacial syndrome region show abnormal sensorimotor gating and learning and memory impairments. *Hum Mol Genet*. 2001;10:2645–2650.
84. Lewis CM, Levinson DF, Wise LH, et al. Genome scan meta-analysis of schizophrenia and bipolar disorder, part II: schizophrenia. *Am J Hum Genet*. 2003;73:34–48.
85. Sullivan PF, Eaves LJ, Kendler KS, Neale MC. Genetic case-control association studies in neuropsychiatry. *Arch Gen Psychiatry*. 2001;58:1015–1024.
86. Levitt P, Ebert P, Mirnics K, Nimgaonkar VL, Lewis DA. Making the case for a candidate vulnerability gene in schizophrenia: convergent evidence for regulator of G-protein signaling 4 (RGS4). *Biol Psychiatry*. 2006;60:534–537.
87. Le-Niculescu H, Balaraman Y, Patel S, et al. Towards understanding the schizophrenia code: an expanded convergent

functional genomics approach. *Am J Med Genet B Neuropsychiatr Genet*. 2007;144:129–158.
88. Fatemi SH, Reutiman TJ, Folsom TD, et al. Chronic olanzapine treatment causes differential expression of genes in frontal cortex of rats as revealed by DNA microarray technique. *Neuropsychopharmacology*. 2006;31:1888–1899.
89. Hakak Y, Walker JR, Li C, et al. Genome-wide expression analysis reveals dysregulation of myelination-related genes in chronic schizophrenia. *Proc Natl Acad Sci USA*. 2001;98:4746–4751.
90. Mirnics K, Lewis DA. Genes and subtypes of schizophrenia. *Trends Mol Med*. 2001;7:169–174.
91. Chung C, Tallerico T, Seeman P. Schizophrenia hippocampus has elevated expression of chondrex glycoprotein gene. *Synapse*. 2003;50:29–34.
92. Mirnics K, Middleton FA, Marquez A, Lewis DA, Levitt P. Molecular characterization of schizophrenia viewed by microarray analysis of gene expression in prefrontal cortex. *Neuron*. 2000;28:53–67.
93. Vawter MP, Crook JM, Hyde TM, et al. Microarray analysis of gene expression in the prefrontal cortex in schizophrenia: a preliminary study. *Schizophr Res*. 2002;58:11–20.
94. Hashimoto T, Volk DW, Eggan SM, et al. Gene expression deficits in a subclass of GABA neurons in the prefrontal cortex of subjects with schizophrenia. *J Neurosci*. 2003;23:6315–6326.
95. Marcotte ER, Srivastava LK, Quirion R. cDNA microarray and proteomic approaches in the study of brain diseases: focus on schizophrenia and Alzheimer's disease. *Pharmacol Ther*. 2003;100:63–74.
96. Pongrac J, Middleton FA, Lewis DA, Levitt P, Mirnics K. Gene expression profiling with DNA microarrays: advancing our understanding of psychiatric disorders. *Neurochem Res*. 2002;27:1049–1063.
97. Akbarian S, Kim JJ, Potkin SG, et al. Gene expression for glutamic acid decarboxylase is reduced without loss of neurons in prefrontal cortex of schizophrenics. *Arch Gen Psychiatry*. 1995;52:258–278.
98. Fatemi SH, Stary JM, Earle JA. GABAergic dysfunction in schizophrenia and mood disorders as reflected by decreased levels of glutamic acid decarboxylase 65 and 67 kDa and Reelin proteins in cerebellum. *Schizophr Res*. 2005;72:109–122.
99. Tkachev D, Mimmack ML, Ryan MM, et al. Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. *Lancet*. 2003;362:798–805.
100. Benes FM, Berretta S. GABAergic interneurons: implications for understanding schizophrenia and bipolar disorder. *Neuropsychopharmacology*. 2001;25:1–27.
101. Seeman P. Atypical antipsychotics: mechanism of action. *Can J Psychiatry*. 2002;47:27–38.
102. Bowden NA, Scott RJ, Tooney PA. Altered gene expression in the superior temporal gyrus in schizophrenia. *Biol Psychiatry*. 2003;53:1086–1098.
103. Colantuoni C, Hyde TM, Mitkus S, et al. Age-related changes in the expression of schizophrenia susceptibility genes in the human prefrontal cortex. *Brain Struct Funct*. 2008;213:255–271.
104. Preti A, Cardascia L, Zen T, et al. Risk for obstetric complications and schizophrenia. *Psychiatry Res*. 2000;96:127–139.
105. Cannon TD, van Erp TG, Rosso IM, Marchetti M, Favaretto G, Miotto P. Fetal hypoxia and structural brain abnormalities in schizophrenic patients, their siblings, and controls. *Arch Gen Psychiatry*. 2002;59:35–41.
106. Boog G. Obstetrical complications and subsequent schizophrenia in adolescent and young adult offsprings: is there a relationship? *Eur J Obstet Gynecol Reprod Biol*. 2004;114:130–136.
107. Schmidt-Kastner R, van Os J, Steinbusch HMW, Schmitz C. Gene regulation by hypoxia and the neurodevelopmental origin of schizophrenia. *Schizophr Res*. 2006;84:253–271.
108. Nicodemus KK, Marenco S, Batten AJ, et al. Serious obstetric complications interact with hypoxia-regulated/vascular-expression genes to influence schizophrenia risk. *Mol Psychiatry*. 2008;13:873–877.
109. Lee JA, Lupski JR. Genomic rearrangements and gene copy-number alterations as a cause of nervous system disorders. *Neuron*. 2006;52:103–121.
110. Walsh T, McClellan JM, McCarthy SE, et al. Rare structural variants disrupt multiple genes in neurodevelopmental pathways in schizophrenia. *Science*. 2008;320:539–543.
111. The International Schizophrenia Consortium.Stone JL, O'Donovan MC, Gurling H, et al. Rare chromosomal deletions and duplications increase risk of schizophrenia. *Nature*. 2008;455:237–241.
112. Hahn CG, Wang HY, Cho DS, et al. Altered neuregulin 1-erbB4 signaling contributes to NMDA receptor hypofunction in schizophrenia. *Nat Med*. 2006;12:824–828.
113. Smith RE, Haroutunian V, Davis KL, Meador-Woodruff JH. Expression of excitatory amino acid transporter transcripts in the thalamus of subjects with schizophrenia. *Am J Psychiatry*. 2001;158:1393–1399.
114. Addington AM, Gornick MC, Shaw P, et al. Neuregulin 1 (8p12) and childhood-onset schizophrenia: susceptibility haplotypes for diagnosis and brain developmental trajectories. *Mol Psychiatry*. 2006;12:195–205.
115. Garcia RA, Vasudevan K, Buonanno A. The neuregulin receptor ErbB-4 interacts with PDZ-containing proteins at neuronal synapses. *Proc Natl Acad Sci USA*. 2000;97:3596–3601.
116. Buxbaum JD, Georgieva L, Young JJ, et al. Molecular dissection of NRG1-ERBB4 signaling implicates PTPRZ1 as a potential schizophrenia susceptibility gene. *Mol Psychiatry*. 2008;13:162–172.
117. Kirschner MA, Arriza JL, Copeland NG, et al. The mouse and human excitatory amino acid transporter gene (EAAT1) maps to mouse chromosome 15 and a region of syntenic homology on human chromosome 5. *Genomics*. 1994;22:631–633.
118. Kitagawa M, Lee SH, McCormick F. Skp2 suppresses p53-dependent apoptosis by inhibiting p300. *Mol Cell*. 2008;29:217–231.
119. Liu YL, Fann CS, Liu CM, et al. HTF9C gene of 22q11.21 region associates with schizophrenia having deficit-sustained attention. *Psychiatr Genet*. 2007;17:333–338.
120. Liu G, Beggs H, Jurgensen C, et al. Netrin requires focal adhesion kinase and Src family kinases for axon outgrowth and attraction. *Nat Neurosci*. 2004;7:1222–1232.
121. Kondo T, Raff MC. Chromatin remodeling and histone modification in the conversion of oligodendrocyte precursors to neural stem cells. *Genes Dev*. 2004;18:2963–2972.
122. Carter CJ. Schizophrenia susceptibility genes directly implicated in the life cycles of pathogens: cytomegalovirus, influenza, herpes simplex, rubella, and Toxoplasma gondii. *Schizophr Bull*. 2008; In press.

123. Northoff G, Waters H, Mooren I, et al. Cortical sulcal enlargement in catatonic schizophrenia: a planimetric CT study. *Psychiatry Res*. 1999;91:45–54.
124. Meltzer HY, Bobo WV, Heckers SH, Fatemi SH. Schizophrenia. In: Ebert MH, Loosen PT, Nurcombe B, eds. *Lange Current Series.* New York, NY: McGraw Hill; 2008: pp. 261–288.
125. Wright IC, Rabe-Hesketh S, Woodruff PW, David AS, Murray RM, Bullmore ET. Meta-analysis of regional brain volumes in schizophrenia. *Am J Psychiatry*. 2000;157: 16–25.
126. Davis KL, Stewart DG, Friedman JI, et al. White matter changes in schizophrenia: evidence for myelin-related dysfunction. *Arch Gen Psychiatry*. 2003;60:443–456.
127. Arnold SE, Trojanowski JQ. Recent advanced in defining the neuropathology of schizophrenia. *Acta Neuropathol*. 1997;92:217–231.
128. Andreasen NC. A unitary model of schizophrenia. Bleuler's ''Fragmented phrene'' as schizencephaly. *Arch Gen Psychiatry*. 1999;56:781–793.
129. Arnold SE. Cellular and molecular neuropathology of the parahippocampal region in schizophrenia. *Ann N Y Acad Sci*. 2000;911:275–292.
130. Bunney WE, Bunney BG. Evidence for a compromised dorsolateral prefrontal cortical parallel circuit in schizophrenia. *Brain Res Brain Res Rev*. 2000;31:138–146.
131. Chana G, Landau S, Beasley C, Everall IP, Cotter D. Two-dimensional assessment of cytoarchitecture in the anterior cingulate cortex in major depressive disorder, bipolar disorder, and schizophrenia: evidence for decreased neuronal somal size and increased neuronal density. *Biol Psychiatry*. 2003;53:1086–1098.
132. Akbarian S, Bunney WE, Jr, Potkin SG, et al. Altered distribution of nicotinamide—adenine—dinucleotide—phosphate-diaphorase cells in frontal lobe of schizophrenics implies disturbances of cortical development. *Arch Gen Psychiatry*. 1993;50:169–177.
133. Ritter LM, Meador-Woodruff JH, Dalack GW. Neurocognitive measures of prefrontal cortical dysfunction in schizophrenia. *Schizophr Res*. 2004;68:65–73.
134. Schiller D, Zuckerman L, Weiner I. Abnormally persistent latent inhibition induced by lesions to the nucleus accumbens core, basolateral amygdala and orbitofrontal cortex is reversed by clozapine but not by haloperidol. *J Psychiatr Res*. 2006;40:167–177.
135. Surguladze SA, Chu EM, Evans A, et al. The effect of long-acting risperidone on working memory in schizophrenia: a functional magnetic resonance imaging study. *J Clin Psychopharmacol*. 2007;27:560–570.
136. Wolf RC, Höse A, Frasch K, Walter H, Vasic N. Volumetric abnormalities associated with cognitive deficits in patients with schizophrenia. *Eur Psychiatry*. 2008;23:541–548.
137. Weiss AP, Dewitt I, Goff D, Ditman T, Heckers S. Anterior and posterior hippocampal volumes in schizophrenia. *Schizophr Res*. 2005;73:103–112.
138. Connor SE, Ng V, McDonald C, et al. A study of hippocampal shape anomaly in schizophrenia and in families multiply affected by schizophrenia or bipolar disorder. *Neuroradiology*. 2004;46:523–534.
139. Arnold SE, Hyman BT, Van Hoesen GW, Damasio AR. Some cytoarchitectural abnormalities of the entorhinal cortex in schizophrenia. *Arch Gen Psychiatry*. 1999;48: 625–632.
140. Luts A, Jonsson SA, Guldberg-Kjaer N, Brun A. Uniform abnormalities in the hippocampus of five chronic schizophrenic men compared with age-matched controls. *Acta Psychiatr Scand*. 1998;98:60–64.
141. Harrison PJ, Owen MJ. Genes for schizophrenia? Recent findings and their pathophysiological implications. *Lancet*. 2003;361:417–419.
142. Uematsu M, Kaiya H. Midsagittal cortical pathomorphology of schizophrenia: a magnetic resonance imaging study. *Psychiatry Res*. 1989;30:11–20.
143. DeLisi LE, Sakuma M, Tew W, Kushner M, Hoff AL, Grimson R. Schizophrenia as a chronic active brain process: a study of progressive brain structural change subsequent to the onset of schizophrenia. *Psychiatry Res*. 1997;74: 129–140.
144. Nopoulos PC, Ceilley JW, Gailis EA, Andreasen NC. An MRI study of cerebellar vermis morphology in patients with schizophrenia: evidence in support of the cognitive dysmetria concept. *Biol Psychiatry*. 1999;46: 703–711.
145. Goldman AL, Pezawas L, Mattay VS, et al. Heritability of brain morphology related to schizophrenia: a large-scale automated magnetic resonance imaging segmentation study. *Biol Psychiatry*. 2008;63:475–483.
146. Meyer-Lindenberg A, Poline JB, Kohn PD, et al. Evidence for abnormal cortical functional connectivity during working memory in schizophrenia. *Am J Psychiatry*. 2001;158: 1809–1817.
147. Riehemann S, Volz HP, Stutzer P, Smesny S, Gaser C, Sauer H. Hypofrontality in neuroleptic-naive schizophrenic patients during the Wisconsin Card Sorting Test–a fMRI study. *Eur Arch Psychiatry Clin Neurosci*. 2001;251: 66–71.
148. Kumari V, Gray JA, Goney GD, et al. Procedural learning in schizophrenia: a functional magnetic resonance imaging investigation. *Schizophr Res*. 2002;57:97–107.
149. Lim KO, Helpern JA. Neuropsychiatric applications of DTI—a review. *NMR Biomed*. 2002;15:587–593.
150. Ardekani BA, Nierenberg J, Hoptman MJ, Javitt DC, Lim KO. MRI study of white matter diffusion anisotropy in schizophrenia. *Neuroreport*. 2003;14:2025–2029.
151. Kubicki M, Westin CF, Nestor PG, et al. Cingulate fasciculus integrity disruption in schizophrenia: a magnetic resonance diffusion tensor imaging study. *Biol Psychiatry*. 2003;54:1171–1180.
152. Bullmore ET, Frangou S, Murray RM. The dysplastic net hypothesis: an integration of developmental and dysconnectivity theories of schizophrenia. *Schizophr Res*. 1997;28: 143–156.
153. Buchsbaum MS, Tang CY, Peled S, et al. MRI white matter diffusion anisotropy and PET metabolic rate in schizophrenia. *Neuroreport*. 1998;9:425–430.
154. Lim KO, Hedehus M, Moseley M, de Crespigny A, Sullivan EV, Pfefferbaum A. Compromised white matter tract integrity in schizophrenia inferred from diffusion tensor imaging. *Arch Gen Psychiatry*. 1999;56:367–374.
155. Foong J, Maier M, Clark CA, Barker GJ, Miller DH, Ron MA. Neuropathological abnormalities of the corpus callosum in schizophrenia: a diffusion tensor imaging study. *J Neurol Neurosurg Psychiatry*. 2000;68:242–244.
156. Agartz I, Andersson JL, Skare S. Abnormal brain white matter in schizophrenia: a diffusion tensor imaging study. *Neuroreport*. 2001;12:2251–2254.

157. Burns J, Job D, Bastin ME, et al. Structural disconnectivity in schizophrenia: a diffusion tensor magnetic resonance imaging study. *Br J Psychiatry*. 2003;182:439–443.
158. Chang L, Friedman J, Ernst T, Zhong K, Tsopelas ND, Davis K. Brain metabolite abnormalities in the white matter of elderly schizophrenic subjects: implication for glial dysfunction. *Biol Psychiatry*. 2007;62:1396–1404.
159. Friedman JI, Davis KL, Chang L, Ernst T, Tsopelas ND, Zhong K. Relationships between white matter metabolite abnormalities, cognitive and social functioning in elderly schizophrenic subjects. *Schizophr Res*. 2008;100:356–358.
160. American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorders (DSM-IV). Washington, DC: American Psychiatric Press; 1994.
161. Easton WW, Chen C-Y. Epidemiology. In: Lieberman JA, Stroup TS, Perkins DO, eds. *The American Psychiatric Publishing Textbook of Schizophrenia.* Washington, DC: American Psychiatric Publishing Inc; 2006:17–38.
162. Sartorius N, Jablensky A, Korten A, et al. Early manifestations and first-contact incidence of schizophrenia in different cultures. A preliminary report on the initial evaluation phase of the WHO Collaborative Study on determinants of outcome of severe mental disorders. *Psychol Med*. 1986;16:909–928.
163. Hales RE, Yudofsky SC, Talbott JA. *The American Psychiatric Press Textbook of Psychiatry.* Washington, DC: American Psychiatric Press; 1999.
164. Aleman A, Kahn RS, Selten JP. Sex differences in the risk of schizophrenia: evidence from meta-analysis. *Arch Gen Psychiatry*. 2003;60:565–571.
165. Harris MJ, Jeste DV. Late-onset schizophrenia: an overview. *Schizophr Bull*. 1988;14:39–55.
166. Howard R, Rabins PV, Seeman MV, Jeste DV. Late-onset schizophrenia and very-late-onset schizophrenia-like psychosis: an international consensus. The International Late-Onset Schizophrenia Group. *Am J Psychiatry*. 2000;157:172–178.
167. Palmer BW, McClure FS, Jeste DV. Schizophrenia in late life: findings challenge traditional concepts. *Harv Rev Psychiatry*. 2001;9:51–58.
168. Carter CS. Re-conceptualizing schizophrenia as a disorder of cognitive and emotional processing: a shot in the arm for translational research. *Biol Psychiatry*. 2006;60:1169–1170.
169. Sullivan PF, Kendler KS, Neale MC. Schizophrenia as a complex trait: evidence from a meta-analysis of twin studies. *Arch Gen Psychiatry*. 2003;60:1187–1192.
170. Asherson P, Mane R, McGiffin P. Genetics and schizophrenia. In: Mirsch SR, Weinberger DR, eds. *Schizophrenia.* Boston, Mass: Blackwell Scientific; 1995:253–274.
171. Arseneault L, Cannon M, Witton J, Murray RM. Causal association between cannabis and psychosis: examination of the evidence. *Br J Psychiatry*. 2004;184:110–117.
172. Javitt DC, Laruelle M. Neurochemical theories. In: Lieberman JA, Stroup TS, Perkins DO, eds. *The American Psychiatric Publishing Textbook of Schizophrenia.* Washington, DC: American Psychiatric Publishing Inc; 2006:85–116.
173. Glantz LA, Lewis DA. Decreased dendritic spine density on prefrontal cortical pyramidal neurons in schizophrenia. *Arch Gen Psychiatry*. 2000;57:65–73.
174. Pierri JN, Volk CL, Auh S, Sampson A, Lewis DA. Decreased somal size of deep layer 3 pyramidal neurons in the prefrontal cortex of subjects with schizophrenia. *Arch Gen Psychiatry*. 2001;58:466–473.
175. Beffert U, Weeber EJ, Durudas A, et al. Modulation of synaptic plasticity and memory by reelin involves differential splicing of the lipoprotein receptor apoer2. *Neuron*. 2005;47:567–579.
176. Fatemi SH. Reelin glycoprotein: structure, biology and roles in health and disease. *Mol Psychiatry*. 2005;10:251–257.
177. Impagnatiello F, Guidotti AR, Pesold C, et al. A decrease of Reelin expression as a putative vulnerability factor in schizophrenia. *Proc Natl Acad Sci USA*. 1998;95:15718–15723.
178. Guidotti A, Auta J, Davis JM, et al. Decrease in reelin and glutamic acid decarboxylase67 (GAD67) expression in schizophrenia and bipolar disorder: a postmortem brain study. *Arch Gen Psychiatry*. 2000;57:1061–1069.
179. Fatemi SH, Earle JA, McMenomy T. Reduction in Reelin immunoreactivity in hippocampus of subjects with schizophrenia, bipolar disorder and major depression. *Mol Psychiatry*. 2000;5:571–654–663.
180. Fatemi SH, Reutiman TJ, Folsom TF. The role of reelin in etiology and treatment of psychiatric disorders. In: Fatemi SH, ed. *Reelin Glycoprotein, Structure and Roles in Health and Disease.* New York, NY: Springer; 2008:317–340.
181. Eastwood SL, Law AJ, Everall IP, Harrison PJ. The axonal chemorepellant semaphorin 3A is increased in the cerebellum in schizophrenia and may contribute to its synaptic pathology. *Mol Psychiatry*. 2003;8:148–155.
182. Kapur S, Mamo D. Half a century of antipsychotics and still a central role for dopamine D2 receptors. *Prog Neuropsychopharmacol Biol Psychiatry*. 2003;27:1081–1090.
183. Carlsson A, Lindqvist M. Effect of chlorpromazine or haloperidol on formation of 3 methoxytyramine and normetanephrine in mouse brain. *Acta Pharmacol Toxicol (Copenh)*. 1963;20:140–144.
184. Davies MA, Sheffler DJ, Roth BL. Aripiprazole: a novel atypical antipsychotic drug with a uniquely robust pharmacology. *CNS Drug Rev*. 2004;10:317–336.
185. Tamminga CA, Carlsson A. Partial dopamine agonists and dopaminergic stabilizers, in the treatment of psychosis. *Curr Drug Targets CNS Neurol Disord*. 2002;1:141–147.
186. Lewis DA, Lieberman JA. Catching up on schizophrenia: natural history and neurobiology. *Neuron*. 2000;28:325–334.
187. Sim K, Cullen T, Ongur D, Heckers S. Testing models of thalamic dysfunction in schizophrenia using neuroimaging. *J Neural Transm*. 2006;113:907–928.
188. Kane J, Honigfeld G, Singer J, Meltzer H. Clozapine for the treatment-resistant schizophrenic. A double-blind comparison with chlorpromazine. *Arch Gen Psychiatry*. 1988;45:789–796.
189. Sharif Z, Bradford D, Stroup S, Lieberman J. Pharmacological treatment of schizophrenia. In: Nathan PE, Gorman J, eds. *A Guide to Treatments That Work.* 3rd ed. New York, NY: Oxford University Press; 2007:203–242.
190. Agid O, Kapur S, Arenovich T, Zipursky RB. Delayed-onset hypothesis of antipsychotic action: a hypothesis tested and rejected. *Arch Gen Psychiatry*. 2003;60:1228–1235.
191. Emsley R, Rabinowitz J, Medori R. Time course for antipsychotic treatment response in first-episode schizophrenia. *Am J Psychiatry*. 2006;163:743–745.
192. Chen ML, Chen CH. Microarray analysis of differentially expressed genes in rat frontal cortex under chronic risperidone treatment. *Neuropsychopharmacology*. 2005;30:268–277.
193. Fatemi SH, Meltzer HY. (2000). Binding of olanzapine to serotonin receptors. In: Tran PV, Bymaster FP, Tye N, Herrera JM, Breier A, Tollefson GD, eds. *Olanzapine (Zyprexa): A Novel Antipsychotic.*Philadelphia, Pa: Lippincott, Williams and Wilkins; 2000:25–30.

194. Lieberman JA, Tollefson GD, Charles C, et al. Antipsychotic drug effects on brain morphology in first-episode psychosis. *Arch Gen Psychiatry*. 2005;62:361–370.
195. Wang HD, Deutch AY. Olanzapine reverses dopamine depletion-induced dendritic spine loss in prefrontal cortical pyramidal neurons. *34th Annual Meeting of Society for Neuroscience.* October, 2004, San Diego, CA
196. Frasca A, Fumagalli F, Ter Horst J, Racagni G, Murphy KJ, Riva MA. Olanzapine, but not haloperidol, enhances PSA-NCAM immunoreactivity in rat prefrontal cortex. *Int J Neuropsychopharmacol*. 2008;11:591–595.
197. Männistö PT, Kaakkola S. Catechol-O-methyltransferase (COMT): biochemistry, molecular biology, pharmacology, and clinical efficacy of the new selective COMT inhibitors. *Pharmacol Rev*. 1999;51:593–628.
198. Karoum F, Chrapusta SJ, Egan MF. 3-Methoxytyramine is the major metabolite of released dopamine in the rat frontal cortex: reassessment of the effects of antipsychotics on the dynamics of dopamine release and metabolism in the frontal cortex, nucleus accumbens, and striatum by a simple two pool model. *J Neurochem*. 1994;63:972–979.
199. Tunbridge EM, Bannerman DM, Sharp T, Harrison PJ. Catechol-o-methyltransferase inhibition improves set-shifting performance and elevates stimulated dopamine release in the rat prefrontal cortex. *J Neurosci*. 2004;24:5331–5335.
200. Fatemi SH, Folsom TD. Catechol-O-methyltransferase gene regulation in rat frontal cortex. *Mol Psychiatry*. 2007;12:322–323.
201. Boydell J, van Os J, McKenzie K, Murray RM. The association of inequality with the incidence of schizophrenia–an ecological study. *Soc Psychiatry Psychiatr Epidemiol*. 2004;39:597–599.
202. Jones P, Rodgers B, Murray R, Marmot M. Child development risk factors for adult schizophrenia in the British 1946 birth cohort. *Lancet*. 1994;344:1398–1402.
203. Cannon M, Jones PB, Murray RM. Obstetric complications and schizophrenia: historical and meta-analytic review. *Am J Psychiatry*. 2002;159:1080–1092.
204. Cantor-Graae E, Selten JP. Schizophrenia and migration: a meta-analysis and review. *Am J Psychiatry*. 2005;162:12–24.
205. Foong J, Symms MR, Barker GJ, Maier M, Miller DH, Ron MA. Investigating regional white matter in schizophrenia using diffusion tensor imaging. *Neuroreport*. 2002;13:333–336.
206. Weinberger DR, McClure RK. Neurotoxicity, neuroplasticity, and magnetic resonance imaging morphometry: what is happening in the schizophrenic brain? *Arch Gen Psychiatry*. 2002;59:553–558.
207. Bertolino A, Kumra S, Callicott JH, et al. Common pattern of cortical pathology in childhood-onset and adult-onset schizophrenia as identified by proton magnetic resonance spectroscopic imaging. *Am J Psychiatry*. 1998;155:1376–1383.
208. Lieberman JA. Is schizophrenia a neurodegenerative disorder? A clinical and neurobiological perspective. *Biol Psychiatry*. 1999;46:729–739.
209. Rapoport JL, Giedd JN, Blumenthal J, et al. Progressive cortical change during adolescence in childhood-onset schizophrenia. A longitudinal magnetic resonance imaging study. *Arch Gen Psychiatry*. 1999;56:649–654.
210. Nair TR, Christensen TD, Kingsbury SJ, Kumar NG, Terry WM, Garver DL. Progression of cerebroventricular enlargement and the subtyping of schizophrenia. *Psychiatry Res*. 1997;74:141–150.
211. Mathalon DH, Sullivan EV, Lim KO, Pfefferbaum A. Progressive brain volume changes and the clinical course of schizophrenia in men: a longitudinal magnetic resonance imaging study. *Arch Gen Psychiatry*. 2001;58:148–157.
212. Jarskog LF, Glantz LA, Gilmore JH. Apoptotic mechanisms in the pathophysiology of schizophrenia. *Prog Neuropsychopharmacol Biol Psychiatry*. 2005;29:846–858.
213. Mattson MP, Duan W. "Apoptotic" biochemical cascades in synaptic compartments: roles in adaptive plasticity and neurodegenerative disorders. *J Neurosci Res*. 1999;58:152–166.
214. Jarskog LF, Gilmore JH, Selinger ES, Lieberman JA. Cortical bcl-2 protein expression and apoptotic regulation in schizophrenia. *Biol Psychiatry*. 2000;48:641–650.
215. Jarskog LF, Selinger ES, Lieberman JA, Gilmore JH. Apoptotic proteins in the temporal cortex in schizophrenia: high Bax/Bcl-2 ratio without caspase-3 activation. *Am J Psychiatry*. 2004;161:109–115.
216. Yuan J, Yankner BA. Apoptosis in the nervous system. *Nature*. 2000;407:802–809.
217. Rund BR. A review of longitudinal studies of cognitive functions in schizophrenia patients. *Schizophr Bull*. 1998;2: 425–435.
218. Wedenoja J, Loukola A, Tuulio-Henriksson A, et al. Replication of linkage on chromosome 7q22 and association of the regional Reelin gene with working memory in schizophrenia families. *Mol Psychiatry*. 2008;13:673–684.
219. Perron H, Mekaoui L, Bernard C, Veas F, Stefas I, Leboyer M. Endogenous retrovirus type W GAG and envelope protein antigenemia in serum schizophrenic patients. *Biol Psychiatry*. 2008;64:1019–1023.
220. Akbarian S. Restoring GABAergic signaling and neuronal synchrony in schizophrenia. *Am J Psychiatry*. 2008;165: 1507–1509.
221. Bullock WM, Cardon K, Bustillo J, Roberts RC, Perrone-Bizzozero NI. Altered expression of genes involved in GABAergic transmission and neuromodulation of granule cell activity in the cerebellum of schizophrenia patients. *Am J Psychiatry*. 2008;165:1594–1603.
222. Sweet RA, Henteleff RA, Zhang W, Sampson AR, Lewis DA. Reduced dendritic spine density in auditory cortex of subjects with schizophrenia. *Neuropsychopharmacology*. 2009;34: 374–389.
223. Fatemi SH, ed. Reelin Glycoprotein: Structure, Biology, and Roles in Health and Disease. New York: Springer.