

The Order of Sequence Alignment Can Bias the Selection of Tree Topology¹

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Sequential pairwise alignment of multiple sequences is a widely used procedure (Kruskal 1983). It is useful and generally successful when sequences within a set differ by relatively few substitutions. Although it is well known that differential substitution rates can artifactually bias the assessment of tree topology (Felsenstein 1978), it is not generally known that the order in which sequences are aligned can bias tree selection.

To test the effect of alignment order, the classical four-taxon test has been applied to the "tree of life" (Lake et al. 1984; Woese and Olsen 1986) by using alternative alignments and three reconstruction algorithms [maximum parsimony (Fitch 1971), transversion parsimony (Brown et al. 1982), and evolutionary parsimony (Lake 1987)]. There is enormous interest in this tree because it relates all known organisms and because its topology is expected to provide insight into the evolution of modern organisms. Because the tree spans large evolutionary distances, its topology has been difficult to establish.

By means of sequences from elongation factor Tu (EF-Tu), the most conserved protein sequence known to span the tree of life, it is shown that specific alignment orders systematically favor alternative trees. In particular, if taxa A and B are pairwise aligned and if C and D are pairwise aligned, the resulting alignment of the EF-Tu sequences more often gives the tree that has A and B as topological neighbors and C and D as topological neighbors, regardless of the tree reconstruction algorithm used. Because all three reconstruction algorithms produced the same tree for any particular alignment, unequal rate effects appear to be secondary for EF-Tu sequences. This indicates that order-dependent alignment biases are distinct from unequal rate effects and that, for some data, they could be as important as unequal rate effects.

Pairwise alignments of protein sequences were performed with the ALIGN program available in the Dayhoff package (Dayhoff et al. 1983). The penalty for a break was 6, and the mutation data matrix corresponded to 250 accepted point mutations with a bias of +2. These are reasonable values for the weights and correspond to those used in the examples in the description of the ALIGN program. [For an insightful discussion of alignment weights, see the paper by Fitch and Smith (1983); also see Waterman and Perlwitz (1984).] EF-Tu sequences were aligned as protein sequences to obtain more robust alignments and were back-translated into nucleic acid sequences (e.g., phe was translated as UUY, leu as YUN, arg as NGN, and ser as NNN) so that the maximum-, transversion-, and evolutionary-parsimony methods could be compared by equivalent data. Only positions consisting of a single nucleotide (i.e., U, C, A, or G but not R, Y, or N) in each of the four sequences were scored. These uniquely defined replacement sites are presumed to correspond to the most conserved nucleotide positions.

A multiple alignment of four sequences can be achieved by successively aligning

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three pairs of sequences. Let A, B, C, and D represent amino acid (or nucleotide) sequences, and let AB represent the alignment (*sensu* Kruskal 1983) of A with B, etc. [In this definition, an alignment consists of a matrix of two rows in which a match is indicated by a column with the same element above and below, a replacement (or substitution) is indicated by a column with different elements above and below, and a deletion in A (or an insertion in B) is represented by a column with a gap (—) above and with a nongap element below, etc.] If one sequence is common to all three pairs, I call these *star* alignments. If no sequence is common to more than two pairs, I call these *linear alignments*; there are 12 of them represented by a linear notation such as ABCD, where the four letters represent the four sequences and where the three adjacent pairs of letters—AB, BD, and DC—represent the three pairwise alignments used to generate the total alignment.

Pairwise-alignment algorithms do not distinguish between the order of the two sequences being aligned. Thus, the alignment of sequence A with B, AB, is equivalent to the alignment of B with A, BA. There are $4! = 24$ orderings of four letters (ABCD represents the alignment of A with B, B with C, and C with D) corresponding to four-taxon pairwise alignments, but because only the neighbors—and not their order—count, the alignment ABDC is equivalent to the alignment CDBA. This explains why it was stated earlier that there are only 12 ($=24/2$) independent linear, pairwise alignments of four sequences.

Multiple alignments were generated from sequential pairwise alignments as illustrated in table 1. In the upper example the AB alignment is aligned to the BC alignment by requiring the common (or guide) sequence B to have its two amino acid sequences aligned perfectly, leading to the introduction of an additional gap (*) in each of the AB and BC alignments. Once this has been done, the ABBC alignment on the left may be reduced to the ABC alignment on the right. [The ABC alignment is defined only within the length (or range) of B because B contains only gaps outside this range. Hence, B can not be used to relate the A and C sequences in regions where B does not exist. Additional rules could be devised to extend this range, but this seems

Table 1
Sequential Alignment of Two Alignments

Alignment of Two Alignments	Reduced Representation
AB aligned with BC: A: KN-ITGTS*QA B: KNMIT-AS*QA	A: KN-ITGTS-QA B: KNMIT-AS-QA
B: KNMIT*AS-QA C: K-MIT*AAKQM	C: K-MIT-AAKQM
BC aligned with CA: B: KNMITAS-QA C: K-MITAAKQM	B: KNMITAS-QA C: K-MITAAKQM
C: K*MITAAKQM A: K*NITGTSQA	A: K-NITGTSQA

NOTE.—Two pairwise alignments of sequences are aligned by reference to a common sequence. At the top left the AB pair is aligned with respect to the BC pair through the common B sequence. At the bottom left the BC pair is aligned with the CA pair through the common C sequence. On the left, hyphens (—) represent gaps introduced when the initial pairs (AB and BC upper, or AB and CA lower) were aligned, and asterisks (*) represent gaps introduced when two alignments were aligned with each other. Asterisks have been changed to hyphens on the right. The final result is shown in a reduced form at the right. A triple alignment ABC is commonly not the same as a triple alignment BCA.

an unnecessary complication for this paper, since positions containing gaps will not be scored.]

One can easily show that the alignment ABC is equivalent to the alignment CBA, since B is used as the guide sequence for both alignments. Furthermore, alignments are associative; that is, the alignment (AB)(CD) is equivalent to (ABC)(D), where the brackets indicate the order in which alignments are combined. [The alignment (ABC)(D) is equivalent to the alignment (ABC)(CD) since the left and right C sequences are identical. Likewise, (AB)(CD) is equivalent to (ABC)(CD). Hence (ABC)(D) is equivalent to (AB)(CD).] Although sequential alignments are associative, they are not in general commutative. This is shown by the example in the bottom half of table 1, where the alignment BCA is calculated. It is clear that the alignment BCA is not equivalent to ABC. A collection of alignments is thus a semigroup under alignment, as pointed out by a reviewer.

The four-taxon tree is the traditional vehicle for testing reconstruction algorithms, and the best-known four-taxon test concerns the tree of life, which relates all known groups of organisms. Hence it will be used to illustrate the effects that alignment has on tree selection. In its unrooted form, the tree of life relates the Halobacteria (H), the Eubacteria (B), the Eukaryotes (K), and the Eocytes (E). The best-studied proteins that are found in all known organisms are the DNA-dependent RNA polymerase, the ATP synthetases, and the protein synthesis factor EF-Tu (EF-1 alpha in eukaryotes). The EF-Tu sequences are the least divergent of the three proteins and were used in the present study. This reduced complications introduced by unequal rate effects.

Amino acid sequences were taken from each of four representative taxa. *Escherichia coli* was selected as the traditional eubacterium (Yokota et al. 1980), but analogous results were obtained with EF-Tu sequences from *Spirulina platensis* (cyanobacterium) and *Thermotoga martima* (thermophilic eubacterium). *Halobacterium marismortui* was chosen as the H sequence (Baldacci et al. 1990). *Thermococcus celer* was selected as an E sequence (Auer et al. 1990). *Saccharomyces cerevisiae* was chosen as the K representative (Nagata et al. 1984) because of its central phylogenetic position and because its sequence appears to have undergone relatively fewer substitutions than have either other single-celled E or most metazoans. For sequences B, E, H, and K, the 12 linear alignments are EKBH, KEBH, EKHB, KEHB, EHBK, HEBK, EHKB, HEKB, HKBE, KHBE, HKEB, and KHEB. In reduced form, EHBK is an alignment of four rows.

Four taxa may be related in only three unrooted trees, as shown in figure 1. In the archaeobacterial tree in figure 1A, K are not topologically closest to either E or H. In the halobacterial tree, shown in figure 1B, K are topologically closest to H, and in the eocyte tree, shown in figure 1C, the K are topologically closest to E.

For the six alignments shown in table 2A, the order of alignment strongly influences the topology. In particular, (1) those alignments in which E is aligned with H and in which K is aligned with B support the archaeobacterial tree, (2) those alignments in which H is aligned with K and in which B is aligned with E support the halobacterial tree, and (3) those in which E is aligned with K and in which B is aligned with H support the eocyte tree. The order of alignment dominates the topology.

The observations are consistent with the following simple explanation: When one aligns the E sequence with the K sequences and then aligns the B and H sequences, the sequences are, in effect, being fit to the tree that has E and K on one side of the central branch and B and H on the other side. In this instance the eocyte topology will be emphasized. Similarly, when one fits E with H and fits B with K, the archaeobacterial tree is favored, and so forth.

When the remaining six alignments are examined (table 2B), a similar but decreased effect is found. Although these alignments primarily support the eocyte tree, the greatest support for a given topology (or a tie for greatest support) occurs when

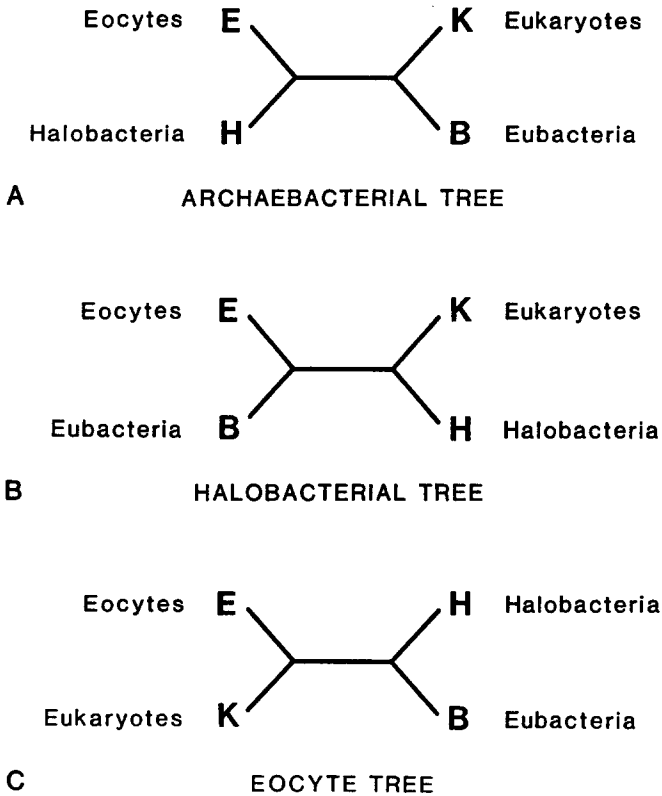


FIG. 1.—Three possible four taxon topologies for “tree of life.” The archaeobacterial tree, shown in A, relates the eocytes with halobacteria and relates the eukaryotes with eubacteria. The halobacterial tree, shown in B, relates the eocytes with eubacteria and relates the eukaryotes with halobacteria. The eocyte tree, shown in C, relates the eocytes with eukaryotes and relates the halobacteria with eubacteria.

its favored alignment order is used. Thus, if the EKHB or KEHB alignments is used, support for the eocyte tree is increased beyond the level that was found by using the EHKB, HEKB, HKEB, and KHEB alignments.

The differences between table 2A and table 2B suggest that the central alignment pair influences the strength of the effect. In all the six alignments in table 2A the B sequence is part of the central aligned pair. Because B is the most divergent of the four sequences [The length of the peripheral branch leading to B is consistently the longest for all alignments and topologies when measured by operator metrics (Lake 1988).], it appears that the effect is stronger if the two central sequences are highly diverged—and is weaker if they are less diverged. This effect was also noted for polymerase and ATP synthetases (J. A. Lake, unpublished results). Thus, when the divergent B sequence is part of the central pair, alignment effects dominate and determine the tree topology.

Whether analyzed by maximum parsimony, by transversion parsimony, or by evolutionary parsimony (see tables 2A and B), tree selection is principally determined by the alignment. Because the three methods have different sensitivities to unequal rates, these effects are probably not biasing the results. Hence, alignment effects are distinct from unequal rate effects. This implies that all the algorithms studied (including evolutionary parsimony, the least affected by unequal rates) are sensitive to sequential alignment effects.

For highly divergent sequences, the order of the alignments can dominate phy-

Table 2
Order of Alignment as Biasing Topology

ALIGNMENT	MAXIMUM PARSIMONY			TRANSVERSION PARSIMONY			EVOLUTIONARY PARSIMONY		
	Archaeobacterial Tree	Halobacterial Tree	Eocyte Tree	Archaeobacterial Tree	Halobacterial Tree	Eocyte Tree	Archaeobacterial Tree	Halobacterial Tree	Eocyte Tree
A: Central alignment including most divergent sequence (B):									
EHBK	<u>27</u>	6	21	<u>16</u>	2	9	<u>11</u>	0	5
HEBK	<u>31</u>	5	16	<u>14</u>	1	8	<u>8</u>	0	5
HKBE	15	<u>22</u>	20	5	<u>14</u>	9	5	<u>11</u> **	4
KHBE	13	<u>28</u>	21	5	<u>18</u>	10	5	<u>12</u> **	5
EKBH	12	7	<u>37</u>	5	4	<u>21</u>	2	2	<u>11</u>
KEBH	8	8	<u>37</u>	3	2	<u>21</u>	2	-1	<u>15</u> ***
B: Central alignment not including most divergent sequence (B):									
EHKB	18	6	<u>18</u>	8	3	<u>9</u>	<u>6</u>	3	3
HEKB	17	5	<u>19</u>	7	2	<u>9</u>	3	2	3
HKEB	13	8	<u>20</u>	5	3	<u>10</u>	1	2	7
KHEB	16	10	<u>19</u>	7	4	<u>10</u>	3	3	<u>5</u>
EKHB	14	8	<u>27</u>	5	4	<u>11</u>	3	3	<u>8</u> *
KEHB	15	8	<u>26</u>	6	3	<u>11</u>	2	2	<u>7</u>
C: Order-invariant alignment	7	2	<u>13</u>	3	0	<u>6</u>	3	1	<u>4</u>
D. Star alignments:									
StarB	13	13	<u>22</u>	6	7	<u>12</u>	5	4	5
StarE	13	8	<u>20</u>	5	2	<u>10</u>	0	0	6
StarH	15	11	<u>25</u>	6	5	<u>11</u>	4	5	<u>7</u>
StarK	16	12	<u>21</u>	6	3	<u>9</u>	4	3	4

NOTE.—Data are scores for the respective trees. In all three parsimony methods used to analyze aligned sequences, each of the three trees is associated with particular patterns of nucleotide occurrence. The scores for maximum parsimony and for transversion parsimony are the number of sequence positions at which the nucleotide pattern supports a particular topology. The scores for evolutionary parsimony are the number of sequence positions that support minus the number that oppose a topology. The tree with the greatest support is deemed “most parsimonious.” Only nucleotide positions without gaps were used in the tree construction analyses. The tree topology supported by the most counts is underlined (ties are not indicated). The six, of 12 possible, alignments shown in A correspond to those in which the central alignment includes the most divergent sequence, (B). In B the central alignment pair does not include the most divergent sequence, (B). Two alternative alignment strategies that are less influenced by tree topologies are shown in C and D. In C the order-independent alignment includes only the portion of the alignment that is common to three diverse linearly sequential alignments. Star alignments are analyzed in D. The StarB alignment uses the B sequence for reference; the StarE alignment uses the E sequence; etc.

* $P < .05$, by χ^2 test (as in Lake 1987).

** $P < .03$, by χ^2 test (as in Lake 1987).

*** $P < .01$, by χ^2 test (as in Lake 1987).

logenetic reconstructions. Furthermore, the alignment artifact is likely to have wide-ranging consequences, since almost all alignments are constructed by first aligning the pairs of sequences that are most similar. Whether calculated by computer or by eye, these types of alignments predispose the algorithms toward the tree that has the least divergent taxa as neighbors. Even true multiple-taxon alignment algorithms (Sankoff and Cedergren 1983) suffer from this distortion, which can enter through the choice of distances to be minimized (J. A. Lake, unpublished results). Until we can understand more completely the subtle relationships between sequence alignment and topology determination, some suggestions for obtaining multiple alignments seem useful.

A direct solution to the alignment problem is to search for alignments that are *independent* of the sequential alignments. This procedure can be computationally intensive but is potentially useful. If one can find subsets of an alignment that are common to all of the possible sequential alignments, then it can be argued that the subset is reasonably free of topological alignment biases. The subset that is common to the EKBH, HKBE, and EHBK alignments is shown in the Appendix. I call this an *order-independent alignment*, and the analysis of it is shown in table 2C. Although this alignment is easy to calculate for four taxa, for large data sets this calculation can become computationally intensive. For example, in one study of the tree of life (Lake 1988), ~1,200 individual four-taxon trees were analyzed. Since each four-taxon tree requires 12 independent alignments, one would need to calculate some 14,000 four-taxon alignments in order to use this alignment method. Nevertheless, this is still a feasible computation.

Another type of pairwise, sequential alignment—the star alignment—requires less computation (Lake 1988). In this alignment, one selects a reference sequence and aligns all other sequences to it. If K were selected as a reference, then one would calculate BK, EK, and HK and combine them. Four star alignments are possible for four taxa, and their analyses are shown in table 2D. For them, all three methods are consistent with the same topology found for the order-independent alignment. This suggests that topological distortions are less for the star and order-independent alignments than for the linear sequential alignments, for these data. The results in table 2D are not significantly different when the divergent B sequence is used as a reference, but in general it would seem unwise to use a divergent sequence as a reference. An obvious benefit is that star alignments require substantially less calculation, since all taxa can be referenced to a single sequence. For the tree of life (Lake 1988), only 31 separate pairwise alignments were required for their combination into the complete 32-taxon star alignment.

Notably, it appears that the four sequences analyzed here tend to support the eocyte tree. Whether this is an effect observed for these four sequences or a statement about the tree of life would take us beyond the scope of the present paper and require the analysis of additional data. Nevertheless, additional EF-Tu genes from eocytes are being sequenced. If order-independent alignments of these sequences—the most conserved protein sequences yet found—should also support the eocyte tree, this would argue strongly for it.

Most important, the present work shows that alignment order introduces topological distortions that are distinct from—and, in the present example, more significant than—unequal rate effects. As additional, longer sequences—and even complete genomes—become available, our attempts to reconstruct the past will become even more ambitious. Almost certainly, consideration of the artifacts introduced by alignment order will play a major role in these studies.

APPENDIX

Some alignments of the EF-Tu sequences used in the present paper are listed in fig. A1. The sequences are referenced by the following code: B (eubacterium), E (eocyte), H (halobacterium), and K (eukaryote). This is followed by four letters describing the alignment that

K	STAR	MKKEK	-----	SHINVVVIGH	VDSGKST	---T---	TGHLI	-----	FKC	GGIDKRTIEK	FE	---KEAAE	LKQSGFKYAW	V	---LDLKA
E	EKBB	MAKKE	-----	PHINIVFVIGH	VDBGKST	---T---	IGRL	-----	YOT	ANIPENI1KK	FE	---EMGE	KGK-SFKFAW	V	---MDRLE
B	EKBB	MSKEKFERTK	-----	PHVNVGTIGH	VDRGKIT	---L---	TAAIT	-----	TVLAKTY	CGAAR	---A	FD	---Q	-----	---IDNAPE
H	EKBB	MSDEQ	-----	-HQNLAIIGH	VDBGKST	---L---	---VG	-----	RLLYETG	SVFEB	---V	IE	---QKKEE	AEEKGKGFFE	FAYVMDLAE
E	EKBE	MAKKE	-----	PHINIVFVIGH	VDBGKST	---L---	---VDG	-----	ENI1KKFEEM	GEKKG	---S	FKFAW	-----	-----	---MDRLE
B	EKBE	MSKEKFERTK	-----	PHVNVGTIGH	VDRGKIT	---L---	---TAAIT	-----	TVLAKTY	CGAAR	---A	FD	---Q	-----	---IDNAPE
H	EKBE	MSDEQ	-----	-HQNLAIIGH	VDBGKST	---L---	---VGRLL	-----	---YET	GSVPFVIEQG	HK	---EEAAE	KGKGGFFEPAY	V	---MDRLAE
E	EHBK	MAKKEP	-----	-BINIVFVIGH	VDBGKST	---T---	---IG	-----	RLLFDIA	NIPEN	---I	IK	---KF-EE	MGEKGS-FK	FAYVMDRLE
B	EHBK	MSKEKFERTK	-----	PHVNVGTIGH	VDRGKIT	---L---	---TAAIT	-----	TVLAKTY	CGAAR	---A	FD	---Q	-----	---IDNAPE
H	EHBK	MSDEQ	-----	-HQNLAIIGH	VDBGKST	---L---	---VG	-----	RLLYETG	SVFEB	---V	IE	---QKKEE	AEEKGKGFFE	FAYVMDLAE
K	COMMON	MKKEK	-----	-BINVVVIGH	VDSGKST	-----	-----	-----	-----	-----	-----	-----	-----	-----	---LDLKA
K	STAR	ERERGITIDI	ALWKFET-P-	KYQVTVIDAP	GERDFIKNMI	TGTSQADCAI	LIIAGGVGEF	EAGISKDQGT	REHALLAFTL	GVRQLIVAVN					
E	EKBB	ERERGITIDV	AHTKFET-PH	RY-ITIIDAP	GERDFVKNMI	TGASQADAAV	LVVA-VTD	---GMP	---QT	KEHAFIARTL	GNNILVAVN				
B	EKBB	EKARGITIND	SHVEYDT-P	TRHYAVDPCP	GERDFVKNMI	TGAAQMDGAI	LVVAAADGGM	P	-----	---QT	REHILLGRQV	GVPIYIVFLN			
H	EKBB	ERERGITIDI	AHQEFST-D	TYDFIIVDCP	GERDFVKNMI	TGASQADNAV	LVVAAADGQV	P	-----	---QT	QEHVFLARTL	GIGELIVAVN			
E	EKBE	ERERGITIDV	AHTKFET-P	HRYITIIDAP	GERDFVKNMI	TGASQADAAV	LVVAVTDGVM	P	-----	---QT	KEHAFIARTL	GNNILVAVN			
B	EKBE	EKARGITIND	SHVEYDT-P	TRHYAVDPCP	GERDFVKNMI	TGAAQMDGAI	LVVAAADGGM	P	-----	---QT	REHILLGRQV	GVPIYIVFLN			
H	EKBE	ERERGITIDV	AHQEFST-D	TYDFIIVDCP	GERDFVKNMI	TGASQADNAV	LVVAA	---DDGV	---QPQT	QEHVFLARTL	GIGELIVAVN				
E	EHBK	ERERGITIDV	AHTKFET-PH	RY-ITIIDAP	GERDFVKNMI	TGASQADAAV	LVVAVTDGVM	P	-----	---QT	KEHAFIARTL	GNNILVAVN			
B	EHBK	EKARGITIND	SHVEYDT-P	TRHYAVDPCP	GERDFVKNMI	TGAAQMDGAI	LVVAAADGGM	P	-----	---QT	REHILLGRQV	GVPIYIVFLN			
H	EHBK	ERERGITIDI	AHQEFST-D	TYDFIIVDCP	GERDFVKNMI	TGASQADNAV	LVVAAADGQV	P	-----	---QT	REHAFIARTL	GIGELIVAVN			
K	COMMON	ERERGITIDI	ALWKFET-	---VTVIDAP	GERDFIKNMI	TGTSQADCAI	LIIA	-----	---QT	QEHVALLARTL	GVRQLIVAVN				
K	STAR	KMDSVKW-DE	SRFQIVKET	SFNIKVKGVN	PKTVFF--VF	ISGWNQDNMI	E--AITNA--	-----	---P--	WYKGWEKETK	AGVVKGKTLG				
E	EKBB	KMDMVNY-DE	KKFKVAEAVQ	KQLLMLGY-	-KNPFI--IP	ISAWEGDNVV	K--KSDKM-	-----	---P--	WYNG	-----	---P	---	---	---PTL
B	EKBB	KCDMVDY-DE	ELLELVEMEV	RELLSQDYFP	GDDTPI--VR	-----	GSALK	A--LEGDA-	-----	---	---	---	---	---	---
H	EKBB	KMDLVYDGE	EYKQVVE-EV	KDLLTQVRFD	SENAKF--IP	-----	VSAFE	GDNTAAEES-	-----	EHGT	WY	---	---	---	---
E	EKBE	KMDMVNY-DE	KKFKVAEAVQ	KQLLMLGY-	-KNPFI--I-	-----	PIS	A--WEGDNVV	KKSDKMP-	WYN	---	---	---	---	---
B	EKBE	KCDMVDY-DE	ELLELVEMEV	RELLSQDYFP	GDDTPI--VR	-----	GSALK	A--LEGDA-	-----	---	---	---	---	---	---
H	EKBE	KMDLVYDGE	SEYKQVVE-EV	KDLLTQVRFD	SENAKF--IP	-----	VSAFE	GDNTAAEES-	-----	G--	---	---	---	---	---
E	EHBK	KMDMVNYDE	KFKAAVE-QV	KXLLLMGVYK	N-----FPIIP	-----	ISAVE	GDNVVKS-	-----	DKMP	WY	---	---	---	---
B	EHBK	KCDMVDY-DE	ELLELVEMEV	RELLSQDYFP	GDDTPI--VR	-----	GSALK	A--LEGDA-	-----	---	---	---	---	---	---
H	EHBK	KMDLVYDGE	EYKQVVE-EV	KDLLTQVRFD	SENAKF--IP	-----	VSAFE	GDNTAAEES-	-----	EHGT	WY	---	---	---	---
K	COMMON	KMDSVKW-	SRFQIVKET	SNFIKVKGVN	PKTVFF--VF	ISGWNQDNMI	E--AITNA--	-----	---P--	WYNG	-----	---P	---	---	---
K	STAR	EAIIDAIEQPS	RPTDKPLRLP	LQDVYKIGGI	GTVPVGRVET	GVIKPGMVVT	F--APAG--	---	---VTE--V	KS	---	---	---	---	---
E	EKBB	EALDQMPEPP	KPTDKPLRIP	IQDVYSISGR	GTVPVGRVET	GVLRVGDVVI	F--EPASTIF	HKPIQGE--V	KS	---	---	---	---	---	---
B	EKBB	DSY--IPEPE	RAIDKPLFLP	IEDVFSISGR	GTVTGTRVER	GILNKVGEVE	I--V--G--	---	---IKET--Q	KSTCTG--V	EMFRKLLDEG				
H	EKBB	E--LPAPE	PPTDAPLRLP	IQDVYISIGI	GTVPVGRVET	GILNTGDNVN	FQPS--D--	---	---VSGE--V	KT	---	---	---	---	---
E	EKBE	DO--MPEPP	KPTDKPLRIP	IQDVYSISGR	GTVPVGRVET	GVLRVGDVVI	I--F--E--	---	---PASTIFH	KPIQGEVSKI	EMHHEEVPKA				
B	EKBE	DSY--IPEPE	RAIDKPLFLP	IEDVFSISGR	GTVTGTRVER	GILNKVGEVE	I--V--G--	---	---IKET--Q	KST--CTGV	EMFRKLLDEG				
H	EKBE	EALNELPAE	PPTDAPLRLP	IQDVYISIGI	GTVPVGRVET	GILNTGDNVN	F--QPSD--	---	---VSGE--V	KT	---	---	---	---	---
E	EHBK	Q--MPEPP	KPTDKPLRIP	IQDVYSISGR	GTVPVGRVET	GVLRVGDVVI	F--APAG--	---	---VTE--V	KS	---	---	---	---	---
B	EHBK	DSY--IPEPE	RAIDKPLFLP	IEDVFSISGR	GTVTGTRVER	GILNKVGEVE	I--V--G--	---	---IKET--Q	KSTCTG--V	EMFRKLLDEG				
H	EHBK	E--LPAPE	PPTDAPLRLP	IQDVYISIGI	GTVPVGRVET	GILNTGDNVN	FQPS--D--	---	---VSGE--V	KT	---	---	---	---	---
K	COMMON	---IEQPS	RPTDKPLRLP	LQDVYKIGGI	GTVPVGRVET	GVIKPGMVV	---	---	---	---	---	---	---	---	---
K	STAR	VPGDNVGFNV	KNVSVKEIRR	GNVCGDAK--	--NDP--F	KGCA-----S	FNATVIVL--	NHPQOISA--	--GYSFVLDC	HTAHI	---	---	---	---	---
E	EKBB	LPGDNIGFNV	RGVCKNDIKR	GDVAGHTN--	--NPTVVRP	KD-----T	FKAQIIVL--	NHPTAITV--	--GYTPVLHA	HTLQV	---	---	---	---	---
B	EKBB	RAGENVGNFNV	RGIKREEIER	QOV--LAK--	--PGT--I	KPHT-----K	FSEVYVILSK	DEGRHPTFF	K-GYRFPQYF	RTTDV	---	---	---	---	---
H	EKBB	EPGDNIGFNV	RGVCKDDIRK	GDV--CG--	--PAD--D	PPSVAE--T	FQAQIVVM--	---QHSPTV	TEGYTFVFA	HTAQV	---	---	---	---	---
E	EKBE	EPGDNIGFNV	RGVCKNDIKR	GDV--AGTN	NPEPT--V	RPKD-----V	FKAQIIVL--	---HPTAIT	V-GYTPVLHA	HTLQVAVRFE					
B	EKBE	RAGENVGVLL	RGIKREEIER	QOV--LAK--	--PGT--I	KPHT-----K	FSEVYVILSK	DEGRHPTFF	K-GYRFPQYF	RTTDV	---	---	---	---	---
H	EKBE	EPGDNIGFNV	RGVCKDDIRK	GDVCGFA--	--DDP--F	SVAE-----T	FQAQIVVM--	---QHSPTV	TEGYTFVFA	HTAQV	---	---	---	---	---
E	EHBK	LPGDNIGFNV	RGVCKNDIKR	GDV--AG--	--HTN--N	PPTVVRKPTD	FKAQIIVL--	---NHPTAIT	TVGYTPVLHA	HTLQV	---	---	---	---	---
B	EHBK	RAGENVGVLL	RGIKREEIER	QOV--LAK--	--PGT--I	KPHT-----K	FSEVYVILSK	DEGRHPTFF	K-GYRFPQYF	RTTDV	---	---	---	---	---
H	EHBK	EPGDNIGFNV	RGVCKDDIRK	GDV--CG--	--PAD--D	PPSVAE--ET	FQAQIVVM--	---QHSPTV	TEGYTFVFA	HTAQV	---	---	---	---	---
K	COMMON	VPGDNVGFNV	KNVSVKEIRR	GNV-----	--PAD--D	PPSVAE--S	FNATVIVL--	---QHSPTV	--GYSFVLDC	HTAHI	---	---	---	---	---
K	STAR	-----A	CRFDELLEKN	DRRSKKL--	---EDHPKF	-----LKS	GDAALVKFVP	S-----KP	MCVEAFSEYP	P	---	---	---	---	---
E	EKBB	-----A	VRFEQLLAKL	DPRTGNIV--	---EENPOF	-----TKT	GDSATVVLRP	T-----KP	MVIEPVKEIP	Q	---	---	---	---	---
B	EKBB	-----T	GTIE	-----	---PEGVEM	-----VMP	GDN--IKMIV	TLI	---BP	IAMD	---	---	---	---	---
H	EKBB	-----A	CTVE	-----	---SID	KKIDPSSGEV	AEENPDFIQN	GDA--AVTV	RPQ	---KP	LSIEP	---	---	---	---
E	EKBE	QLLAKLDPRT	GNI	-----	---V--	---EENPOF	---IKT	GDS--AIIVL	RPTKPMVIEP	VKEIP	---	---	---	---	---
B	EKBE	-----T	GTIE	-----	---L--	---PEGVEM	-----VMP	GDN--IKMIV	T	---	---	---	---	---	---
H	EKBE	-----A	CTVESIDKKI	DPSSGEVA--	---EENPDF	---IQN	GDAAVTVVRP	Q	---	---	---	---	---	---	---
E	EHBK	-----A	VRFE	-----	---QLL	AKLDPRTGNI	VEENPOFIKT	GDS--AIIVL	RPT	---	---	---	---	---	---
B	EHBK	-----T	GTIE	-----	---L--	---PEGVEM	-----VMP	GDN--IKMIV	TLI	---	---	---	---	---	---
H	EHBK	-----A	CTVE	-----	---SID	KKIDPSSGEV	AEENPDFIQN	GDA--AVTV	RPQ	---KP	LSIEP	---	---	---	---
K	COMMON	-----	-----	-----	---	---	---	---	---	---	---	---	---	---	---
K	STAR	AVRDMRQTVA	VGVIKSVDKT	EKAAKVTKAA	QKAARK										458
E	EKBB	AIRDMGQTV	AGMVISIQKA	E-----	-----										428
B	EKBB	AIREGRTVG	AGVV-----	---AKVLS	-----										394
H	EKBB	AIRDMGQTV	AG-----	---KVLGVN	ER	---	---	---	---	---	---	---	---	---	421
E	EKBE	AIRDMGQTV	AGMV-----	---YSIQKAE	-----										428
B	EKBE	AIREGRTVG	AGVV-----	---AKVLS	-----										394
H	EKBE	AIRDMGQTV	AGKVLGVN--	ER-----	-----										421
E	EHBK	AIRDMGQTV	AG-----	---MVISIQ	KAE	---	---	---	---	---	---	---	---	---	428
B	EHBK	AIREGRTVG	AGVV-----	---AKVLS	-----										394
H	EHBK	AIRDMGQTV	AG-----	---KVLGVN	ER	---	---	---	---	---	---	---	---	---	421
K	COMMON	AVRDMRQTVA	VG-----	---	---										237

FIG. A1.—Various sequence alignments of EF-Tu

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corresponds to the code used in the text. The K STAR sequence is the yeast sequence used as a "star" reference to combine the three different alignments. The K COMMON sequence is the yeast sequence at only those positions where the EKBH, HKBE, and EHBK alignments are identical.

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LITERATURE CITED

- AUER, J., B. SPICKER, and A. BOCK. 1990. Nucleotide sequence of the gene for elongation factor EF-1 alpha for the extreme thermophilic archaebacterium *Thermococcus celer*. *Nucleic Acids Res.* **18**:3989.
- BALDACCI, B., F. GUINET, J. TILLIT, G. ZACCAI, and A.-M. DE RECONDO. 1990. Functional implications related to the gene structure of the elongation factor EF-Tu form *Halobacterium marismortui*. *Nucleic Acids Res.* **18**:507-511.
- BROWN, W. M., E. M. PRAGER, A. WANG, and A. C. WILSON. 1982. Mitochondrial DNA sequences of primates: tempo and mode of evolution. *J. Mol. Evol.* **18**:225-239.
- DAYHOFF, M. O., W. C. BARKER, and L. T. HUNT. 1983. Establishing homologies in protein sequences. *Methods Enzymol.* **91**:524-545.
- FELSENSTEIN, J. 1978. Cases in which parsimony or compatibility methods will be positively misleading. *Syst. Zool.* **27**:401-410.
- FITCH, W. M. 1971. Toward defining the course of evolution: minimum change for a specific tree topology. *Syst. Zool.* **20**:406-416.
- FITCH, W. M., and T. F. SMITH. 1983. Optimal sequence alignments. *Proc. Natl. Acad. Sci. USA* **80**:1382-1386.
- KRUSKAL, J. B. 1983. An overview of sequence comparison. Pp. 1-45 in D. SANKOFF and J. B. KRUSKAL, eds. *Time warps, string edits, and macromolecules*. Addison-Wesley, Reading, Mass.
- LAKE, J. A. 1987. A rate-independent technique for analysis of nucleic acid sequences: evolutionary parsimony. *Mol. Biol. Evol.* **4**:167-191.
- . 1988. Origin of the eukaryotic nucleus determined by rate-invariant analysis of rRNA sequences. *Nature* **331**:184-186.
- LAKE, J. A., E. HENDERSON, M. W. CLARK, and M. OAKES. 1984. Eocytes: a new ribosome structure indicates a kingdom with a close relationship to eukaryotes. *Proc. Natl. Acad. Sci. USA* **81**:3786-3790.
- NAGATA, S., K. NAGASHIMA, Y. TSUNETSUGU-YOKATA, K. FUJIMURA, M. MIYAZAKI, and Y. KAZIRO. 1984. Polypeptide chain elongation factor 1 alpha from yeast: nucleotide sequence of one of the two genes for EF-1 alpha from *Saccharomyces cerevisiae*. *EMBO J.* **3**:1825-1830.
- SANKOFF, D., and R. J. CEDERGREN. 1983. Simultaneous comparison of three or more sequences related by a tree. Pp. 253-263 in D. SANKOFF and J. B. KRUSKAL, eds. *Time warps, string edits, and macromolecules*. Addison-Wesley, Reading, Mass.
- WATERMAN, M., and M. D. PERLWITZ. 1984. Line geometries for sequence comparison. *Bull. Math. Biol.* **46**:567-577.
- WOESE, C. R., and G. J. OLSEN. 1986. Archaeobacterial phylogeny: perspectives on the urkingdoms. *Syst. Appl. Microbiol.* **7**:161-177.
- YOKOTA, T., H. SUGISAKI, M. TAKANAMI, and Y. KAZIRO. 1980. The nucleotide sequence of the cloned *tufA* gene of *Escherichia coli*. *Gene* **12**:25-31.

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