

The p65 subunit of NF- κ B regulates I κ B by two distinct mechanisms

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Transcription factor NF- κ B (p50/p65) is generally localized to the cytoplasm by its inhibitor I κ B. Overproduced I κ B, free from NF- κ B, is rapidly degraded. Overexpression of p65 increases endogenous I κ B protein in both carcinoma and lymphoid cells by two mechanisms: protein stabilization and increased transcription of I κ B mRNA. In contrast, p65 Δ , a naturally occurring splice variant, fails to markedly augment I κ B protein levels. Both overexpressed p65 and coexpressed p50 are cytoplasmic, whereas p65 Δ is partly nuclear, indicating that the I κ B induced by p65 can maintain NF- κ B in the cytoplasm. Thus, p65 and I κ B are linked in an autoregulatory loop, ensuring that NF- κ B is held in the cytoplasm until cells are specifically induced to translocate it to the nucleus.

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NF- κ B is a widely expressed factor important for transcription of the immunoglobulin κ gene in mature B-lymphoid cells and for rapid response to external stimuli in other cells (Blank et al. 1992; Nolan and Baltimore 1992). NF- κ B is composed of two subunits, p50 and p65, that associate with a third protein, I κ B, in many cell types (Baeuerle and Baltimore 1989). p50 and p65 exhibit amino-terminal homology to the proto-oncogene *c-rel*, heterodimerize through interaction sites in their Rel homology domains, and bind to κ B sites in a variety of genes to stimulate transcription (Ghosh et al. 1990; Kieran et al. 1990; Zabel et al. 1991). In addition to its Rel homology region, p65 contains an acidic carboxyl terminus that functions in transcriptional activation (Nolan et al. 1991; Ruben et al. 1991; Fujita et al. 1992). A naturally occurring splice variant of p65, p65 Δ , which lacks 10 *rel*-homologous amino acids, has been suggested to transform Rat-1 fibroblasts and to exhibit differential expression during hemopoietic cell development (Narayanan et al. 1992). The 38-kD protein, I κ B- α , that copurifies with NF- κ B and inhibits its binding to DNA, can be dissociated from p50/p65 by deoxycholate (Baeuerle and Baltimore 1988; Zabel and Baeuerle 1991). A cDNA clone, MAD-3, originally isolated from adherent macrophages, encodes a protein that exhibits the properties of and contains peptide sequences from I κ B- α (Davis et al. 1991; Haskill et al. 1991; Link et al. 1992). Notable in its sequence are repeats of a motif present in *cdc10*, SWI4, SWI6 and the erythrocyte structural protein ankyrin (ankyrin repeat; Andrews and Herskowitz 1989; Lux et al. 1990).

The mechanisms controlling the levels of NF- κ B constituents are not yet established. Transient activation of

NF- κ B can be achieved in many cells by treatment with stimuli like active phorbol esters (PMA) or lipopolysaccharide (LPS; Sen and Baltimore 1986). PMA stimulates protein kinase C, which can phosphorylate I κ B in vitro, eliminating its ability to inhibit DNA binding by p50/p65 (Ghosh and Baltimore 1990). Thus, the control of NF- κ B may involve, at least partially, the controlled phosphorylation of I κ B.

We have found that p65 overexpression, directed either by transfection or by retroviral infection, causes a striking increase in endogenous I κ B- α protein. The induced I κ B- α is quite stable. It differs markedly in this sense from I κ B- α overproduced by plasmid-directed expression, which is short-lived. Combination of I κ B- α with p65 then appears to stabilize the inhibitor protein, explaining part of the increased I κ B- α produced by p65 expression. Overexpression of p65 also induces increased I κ B- α mRNA; run-on experiments show that the increase is the result of a highly specific stimulation of transcription. In contrast to p65, retrovirus-directed expression of the 10-amino-acid natural deletion mutant p65 Δ fails to increase intracellular levels of I κ B- α . p65 Δ binds with lower affinity to I κ B and does not activate transcription from κ B-dependent promoters (Ruben et al. 1992). The localization of overproduced p65 and p65 Δ reflects their ability to induce I κ B: p65 is cytoplasmic, whereas p65 Δ is partly nuclear. In addition, coexpressed p50 and p65 are maintained in the cytoplasm owing to the induced I κ B, whereas p50 alone is nuclear. These findings are explained most simply by the idea that I κ B- α induced by p65 maintains NF- κ B in the cytoplasm. Therefore, it appears that this system of proteins is autoregulated through the ability of p65 to determine the

level of I κ B- α protein. Similar conclusions were reached in two articles published while this paper was being reviewed (Brown et al. 1993; Sun et al. 1993).

Results

I κ B levels parallel those of p65, not p65 Δ

cDNAs coding for I κ B- α , p65, and p65 Δ were each inserted into a retroviral vector and then transfected by the calcium phosphate technique into 293 cells (human embryonic kidney cells transformed by the adenovirus E1A protein). Using Western blotting to detect the protein, untransfected cells exhibited nearly undetectable levels of p65, whereas lysates prepared 48 hr after transfection with each of three separately prepared constructs contained high levels of p65 (Fig. 1A, lanes 5–7). The untransfected cells also had virtually undetectable levels of I κ B- α , but the cells that made p65 contained a large quantity of I κ B- α (Fig. 1B, lanes 5–7). This inductive effect was specific to wild-type p65, because control cells transfected with the activated tyrosine kinase *v-abl*, the I κ B-like molecule *bcl-3*, or empty vector (pGD) alone showed no increase in I κ B- α (lanes 1–4,8). The levels of I κ B- α induced by p65 were surprisingly high. By Western blotting, the amount of p65-dependent I κ B- α was similar to that achieved through retroviral expression of the I κ B- α gene product alone (lane 9). In contrast, cells transfected with the p65 Δ construct produced p65 Δ but exhibited little, if any, induction of I κ B- α expression (lane 11) compared with cells transfected with the parent retroviral vector (lane 8); at most, a twofold increase might occur compared with the very marked increase induced by p65.

Time-course studies also suggested that the observed increases in I κ B- α expression depended on p65 expression. Lysates were prepared from separate dishes of cells that were transfected simultaneously with a p65 construct and harvested at the times indicated in Figure 2. Western blotting and detection with antibodies to p65 revealed low levels of the proteins until 24 hr after transfection. I κ B- α levels in the same lysates remained low at early time points and rose only when p65 was seen (Fig. 2B, lanes 7,8).

The striking parallels between p65 and I κ B- α expression were not limited to transfected or adherent cells. To introduce the p65 retroviral expression construct into pre-B and B cells by retroviral transduction, we first cotransfected 293 cells with both the replication-defective p65 retrovirus construct and a clone encoding replication-competent Moloney murine leukemia virus (MuLV) to generate virus stocks that could infect murine 70Z/3 and WEHI 231 cells. Two days after transfection, lymphoid cells were added to the transfected (adherent) producer cells and cocultured for 24 hr. Medium containing the infected nonadherent lymphoid cells was then aspirated from the coculture. Lysates from the nonadherent cells were analyzed by Western blotting. Like the transfected 293 cells, both murine B-lymphoid cell types expressed elevated amounts of I κ B- α after introduction of the p65 gene (p65 levels in Fig. 3A; I κ B- α levels in Fig. 3B). The p65 and I κ B- α that we detected were unlikely to result from contamination of the nonadherent cell population by retrovirus-producing 293 cells for two reasons: (1) Free virus infection of 70Z/3 and WEHI 231 cells has been carried out with identical, but quantitatively less, impressive results (data not shown); (2) human nonadherent lymphoid (Namalwa) cells that are not infectable by ecotropic murine virus were cocultivated

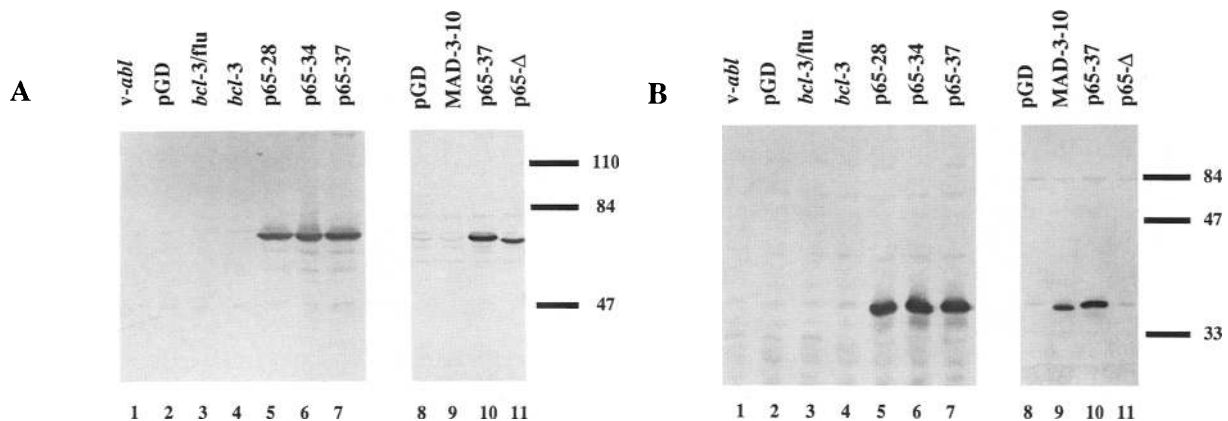


Figure 1. p65 and I κ B- α proteins in transfected cells. Human renal carcinoma (293) cells were transfected with plasmids directing expression of the genes indicated at the top of A and B. pGD (lanes 2, 8) is the parent retroviral vector without insert. *v-abl* refers to a construct containing the Abelson murine leukemia virus tyrosine kinase gene (lane 1). *bcl-3* and *bcl-3/flu* (lanes 3,4) refer to constructs containing a murine cDNA encoding the *bcl-3* proto-oncogene with or without a carboxy-terminal influenza virus HA epitope. p65-28, p65-34, and p65-37 (lanes 5–7) are independent clones of a p65 cDNA in the pGD vector. MAD-3-10 (lane 9) refers to a pGD plasmid containing the I κ B- α (MAD-3) cDNA. p65 Δ (lane 11) was derived from pGD-p65-34. Whole-cell protein lysates were boiled in SDS/mercaptoethanol, electrophoresed through denaturing gels, and transferred to nitrocellulose. One filter was incubated with anti-p65 serum (A); the second filter was incubated with antibody to I κ B- α (B). Molecular masses (in kilodaltons) are indicated by the numbers at right. Lanes 1–7 and 8–11 are from separate transfections.

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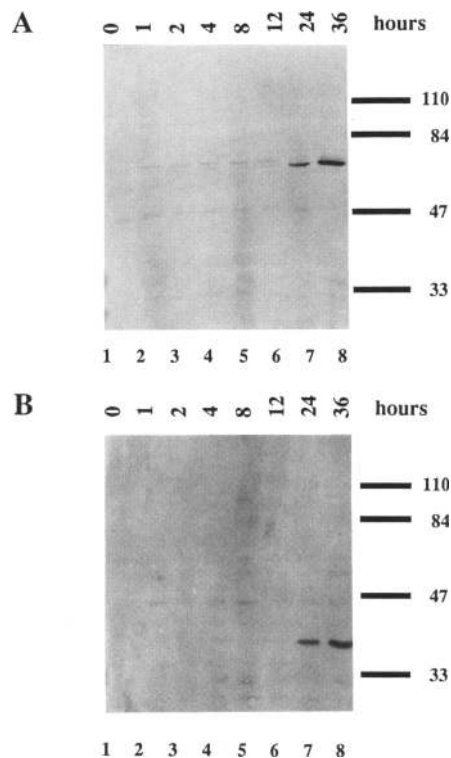


Figure 2. Time course of p65 and I κ B- α expression in transfected cells. 293 cells grown in separate 60-mm dishes were transfected simultaneously with pGD-p65-34 DNA as in Fig. 1. At times indicated (in hours) after addition of DNA as a calcium phosphate precipitate, cells from each dish were lysed and boiled in SDS/mercaptoethanol sample buffer. Duplicate polyacrylamide gels of the samples were transferred to nitrocellulose. One filter was incubated with p65 antiserum (A); the second filter was incubated with affinity-purified antibody to I κ B- α (B). Molecular masses (in kilodaltons) are indicated by the numbers at right. Bands demonstrating retrovirally expressed p65 and endogenous I κ B- α are visualized in the 24- and 36-hr time points (lanes 7,8).

with retrovirus-producing 293 cells by the same protocol and displayed no detectable level of p65 or I κ B- α (data not shown). We therefore conclude that I κ B- α levels in a variety of cell types may be determined by the levels of p65 expression in those cells.

The mechanism of increased I κ B expression is multifactorial

To determine the mechanism of increased I κ B- α expression in p65-transfected cells, we first carried out protein-labeling experiments. 293 cells were transfected with either a retroviral p65 construct or an I κ B- α (MAD-3) retroviral plasmid and labeled for 30 min with [35 S]methionine. Immunoprecipitations at the times indicated after labeling showed that the half-life of p65 was long, on the order of 12–24 hr (Fig. 4A; despite strongly denaturing lysis and wash conditions, a smaller protein that migrates at a size identical to overexpressed I κ B- α was

also observed in the immunoprecipitations of p65). Overexpressed I κ B- α was, in the absence of p65, a very short-lived protein with a $t_{1/2}$ of \sim 30 min (Fig. 4B). Immunoprecipitation of I κ B- α from p65-transfected cells (Fig. 4C,D) showed that protein stabilization was a significant factor in increasing the levels of the inhibitor. In the presence of p65, the half-life of I κ B- α was extended at least 25-fold to $>$ 12 hr (Fig. 4D). The upper limit of the half-life is not clearly established by the above experiment, which was carried out three times with no reproducible drop in the amount of protein detected 24 hr after labeling.

To investigate the importance of transcription or mRNA stabilization to the observed increase in I κ B- α protein levels, poly(A)-containing RNA from 293 cells transfected with retroviral p65, p65 Δ , or pGD vectors was isolated and hybridized with p65- and I κ B- α -specific probes. Specifically bound probe was quantitated using a PhosphorImager. Cells transfected with the p65 or p65 Δ constructs exhibited the expected 20- to 50-fold increases of p65 mRNA (Fig. 5A). Importantly, the p65-transfected cells contained approximately seven times as much I κ B- α mRNA as did cells transfected with p65 Δ or the pGD vector (Fig. 5B). Thus, p65 can both stabilize I κ B- α protein and increase the steady-state concentration of its mRNA.

To determine whether the increased I κ B- α mRNA was the result of increased transcription of the gene, we carried out nuclear run-on studies. We used retroviruses to introduce *rel-* or I κ B-related genes into NIH-3T3 cells and obtain run-on RNA free of contaminating plasmid sequences. Viruses transducing p50, an influenza hem-

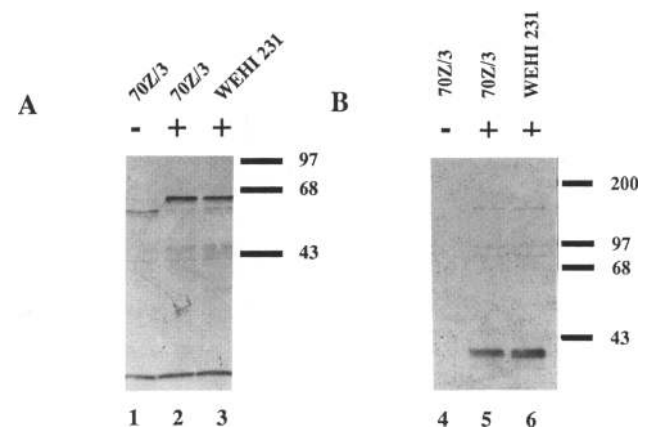


Figure 3. Retrovirus-induced p65 and I κ B- α proteins in lymphoid cells. 70Z/3 pre-B cells were infected with Moloney MuLV alone (-, lanes 1,4) or with pGD-p65 and MuLV helper virus (+, lanes 2,5); WEHI 231 cells were infected with pGD-p65 and MuLV helper virus (+, lanes 3,6). Twenty-four hours after cocultivation with adherent cells producing the indicated viruses, lymphoid cells were aspirated from the culture, pelleted by centrifugation, and lysed in RIPA buffer. Lysates were then subjected to gel electrophoresis and Western blotting as in Figs. 1 and 2. (A) Proteins detected by p65-specific antiserum; (B) proteins detected by I κ B- α -specific antibodies.

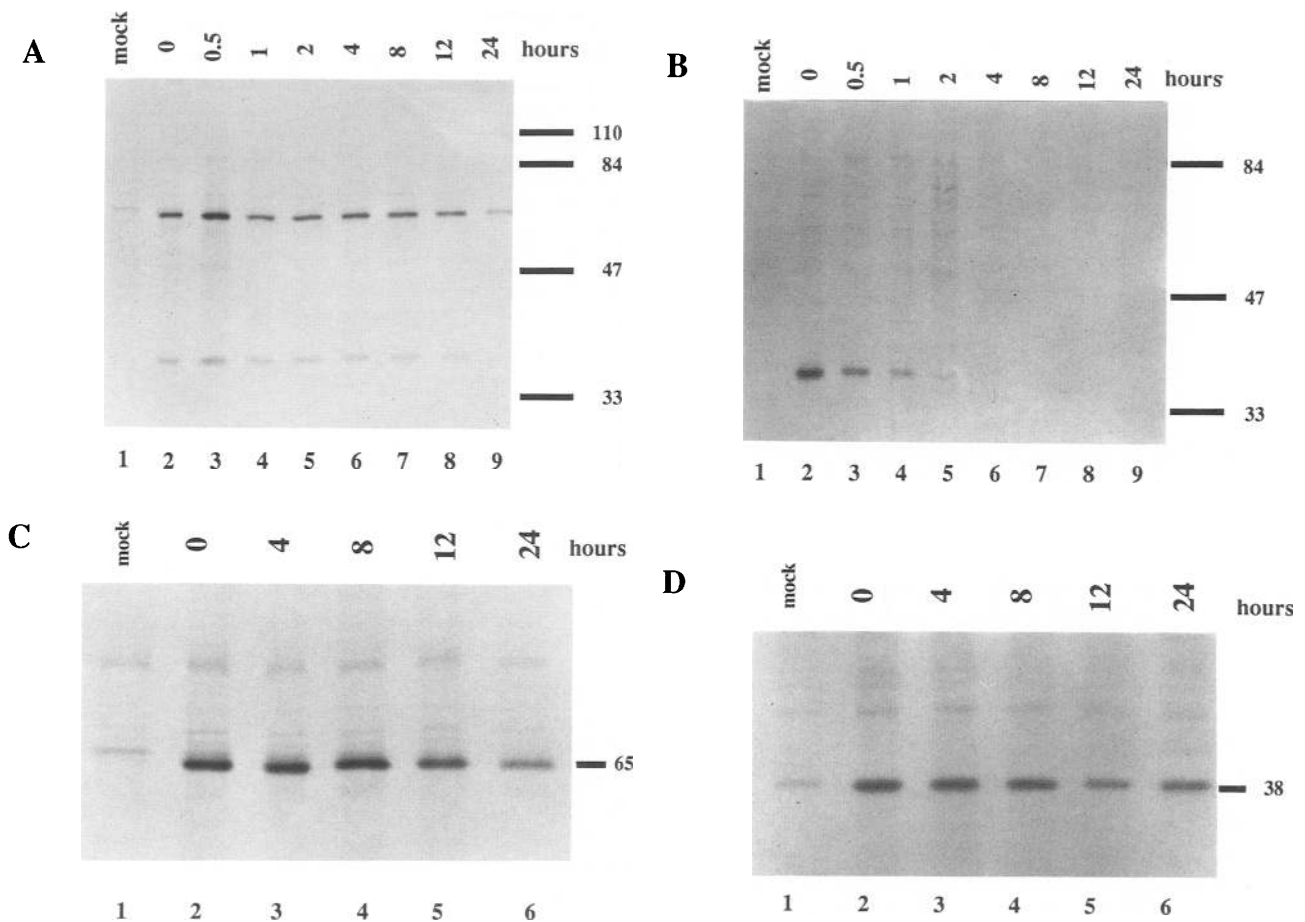


Figure 4. Pulse-chase labeling of p65 and I κ B- α proteins in transfected cells. (A) Separate plates of 293 cells were transfected with pGD-p65-34 DNA or pGD vector DNA (mock, lane 1). Thirty-six hours after addition of the DNA, all plates were starved in methionine-free medium for 30 min, and pulse labeled with [35 S]methionine for 30 min. All cells were then refed with normal medium. Just after feeding, lysates were prepared from mock-transfected cells and pGD-p65-transfected cells (time 0, lane 2). Lysates from separate plates were then prepared at the times indicated (in hours) and stored at -70°C . All samples were then immunoprecipitated using anti-p65 serum, resolved by electrophoresis on SDS-polyacrylamide gels, and autoradiographed. (B) Separate plates of 293 cells were transfected with retroviral DNAs encoding I κ B- α (pGD-MAD-3-10) or pGD vector alone (mock) and processed as described in A. These I κ B- α -transfected cell lysates were then immunoprecipitated using affinity-purified anti-I κ B- α antibody, analyzed on SDS-polyacrylamide gels, and autoradiographed. (C,D) Separate plates of 293 cells were transfected with pGD-p65-34 DNA or pGD vector DNA (mock, lane 1). Thirty-six hours after the addition of the DNA, all plates were labeled with [35 S]methionine, refed, and lysed at the indicated times (in hours) after labeling as before. Lysates from each plate were then divided. Half of the lysate from a given time point was immunoprecipitated using anti-p65 serum (C); the other half was precipitated with anti-I κ B- α antibody (D). The precipitates were analyzed on separate SDS-polyacrylamide gels and autoradiographed as shown.

agglutinin (HA) epitope-tagged variant of p50, p65, p65 Δ , bcl-3, and I κ B- α were tested. RNAs from the nuclei of infected cells were *in vitro* labeled and hybridized to immobilized plasmid DNAs coding for various *rel*- and I κ B-related genes, as well as others (Fig. 6). In each case, the increased transcription of the retrovirally transduced gene was detected. The only endogenous gene that responded to any retroviral infection was the I κ B- α gene; its transcription rate increased 5- to 10-fold after infection by the p65-encoding retrovirus (lane 3) but not after infection with the other viruses. Although strong binding between p50 and both bcl-3 and I κ B- γ have been demonstrated (Franzoso et al. 1992; Wulczyn et al. 1992), no transcriptional response to p50 overproduction was evi-

dent among any of the genes tested. Thus, p65 specifically increases the rate of I κ B- α transcription, accounting for the increased steady-state amount of I κ B- α mRNA.

Localization of p65

A central tenet of previous thinking about I κ B/NF- κ B regulation has been the idea that functional I κ B tethers NF- κ B in the cytoplasm of cells where it is dormant. Therefore, if the I κ B- α protein induced by p65 is functional, overexpressed p65 should be confined to the cytoplasm. To determine the subcellular localization of p65, we used helper-containing retroviral stocks to transmit the p65 gene to NIH-3T3 cells for immunofluores-

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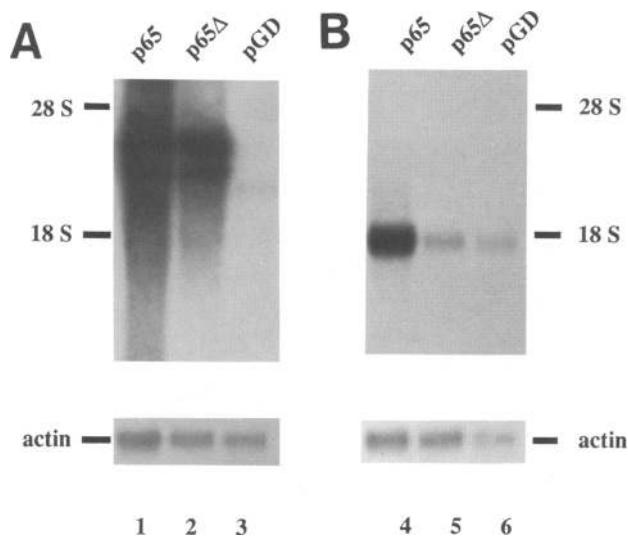


Figure 5. p65 and I κ B- α mRNAs in transfected cells. 293 cells were transfected with DNA constructs encoding p65 or p65 Δ expressed from a retroviral promoter, or with the empty retroviral vector (pGD), as described in Materials and methods. Poly(A)-containing RNA (2.5 μ g/lane) from each population of cells was fractionated by electrophoresis through a formaldehyde-agarose gel, transferred to nitrocellulose, and hybridized with 32 P-labeled probe. The transfected expression construct is indicated at the *top* of each lane. Bars at the *right* indicate the positions of 18S and 28S RNAs, determined by ethidium bromide staining of a separate lane from the gel. (A) Filter 1 was hybridized to a p65-specific probe. Intensely hybridizing bands represent the RNA transcribed from the p65 (lane 1) and p65 Δ (lane 2) expression constructs. The weak band in lane 3 represents endogenous p65 mRNA. Hybridization of the same filter to an actin probe is shown below. (B) Filter 2 was hybridized to an I κ B- α -specific probe. The band visualized in lanes 4–6 represents endogenous I κ B- α mRNA. Hybridization of the same filter to an actin probe is shown below.

cence studies. Under the staining conditions used, endogenous p65 was essentially undetectable (Fig. 7A,B). As predicted, retrovirally transduced wild-type p65 was almost completely limited to the cytoplasm (Fig. 7C). In contrast, the transcriptionally inactive p65 Δ was found in both the nucleus and the cytoplasm of infected cells (Fig. 7D).

If the p65 regulatory effect is the central mode of NF- κ B regulation, then the induced I κ B should maintain the whole NF- κ B complex in the cytoplasm. To test this possibility, we simultaneously infected NIH-3T3 cells with retroviral stocks that could transmit NF- κ B p50 (tagged at its amino terminus with a 9-amino-acid influenza virus HA epitope and detectable by a mouse monoclonal antibody; Field et al. 1988) or p65 (detectable by specific rabbit antiserum). Immunofluorescence staining 2 days after infection (Fig. 8) revealed populations of cells singly infected by either p50- or p65-transducing viruses, cells simultaneously infected by viruses transducing both genes, and uninfected cells. Figure 8A is the phase photomicrograph of a field in which one cell overexpressing

p65 is stained with a fluorescein-conjugated secondary antibody (Fig. 8B); neighboring cells lack expression. Examination using a rhodaminated anti-mouse antibody (Fig. 8C) shows that the p65-expressing cell is also expressing p50 and that both molecules are wholly cytoplasmic. As a control, another cell in the field expresses p50 but not p65. Its fluorescence is entirely nuclear. Thus, the I κ B- α induced by p65 maintains the p50/p65 NF- κ B complex in the cytoplasm, even at very high levels of expression. In contrast, p50 alone induces only a small amount of I κ B (data not shown), and virtually all of the protein is found in the nucleus.

Discussion

NF- κ B p50/p65 heterodimers in most cells are held in the cytoplasm by an inhibitor molecule, I κ B. In many cell types, specific signals such as LPS, PMA, or TNF- α can release p50/p65 from I κ B, causing transport of transcriptionally active heterodimers to the nucleus. This scheme requires that inhibitor levels in unstimulated cells always equal or exceed transcription factor levels. Our studies suggest that the importance of controlling transcription from κ B-dependent genes is such that higher organisms have evolved at least two ways to assure parity of NF- κ B and I κ B levels. First, p65-dependent stabilization of I κ B- α protein can increase inhibitor half-life at least 25-fold. Because p65 is stable, but free I κ B- α is unstable, cells must make a large molar excess of I κ B- α that is available to be stabilized by any increased synthesis of p65. Second, I κ B- α mRNA levels are at least sevenfold inducible by p65 as a consequence of increased transcription of the I κ B- α gene. Therefore, even larger increases of p65 can trigger increased synthesis of I κ B- α .

Stabilization of I κ B- α

We found that the half-life of retrovirally expressed I κ B- α , free from p65, is short (\sim 30 min). Similar to findings by Sun et al. (1993) and Brown et al. (1993), published while this paper was being reviewed, we have observed the rapid disappearance of endogenous I κ B after PMA or LPS treatment of pre-B cells, presumably reflecting the difference in half-life between the unstable free and longer-lived bound forms (H.-C. Liou, M.L. Scott, and D. Baltimore, unpubl.). Two structural features of I κ B- α predict a short half-life for the free protein. The two amino acids just following the initiating methionine (F and Q) are by the amino-end rule "destabilizing" residues (Bachmair and Varshavsky 1989); interestingly, two lysine residues, 18 amino-acids carboxy-terminal to the Q, are at a distance such that one or both could serve as acceptors for ubiquitin and target the protein for degradation (Herzhko 1991). Also, the presence of PEST sequences associated with rapid protein turnover has been noted previously (Rogers et al. 1986; Haskill et al. 1991). Thus, binding of I κ B- α to p65 must prevent the inhibitor molecule from interacting with one or more degradative pathways

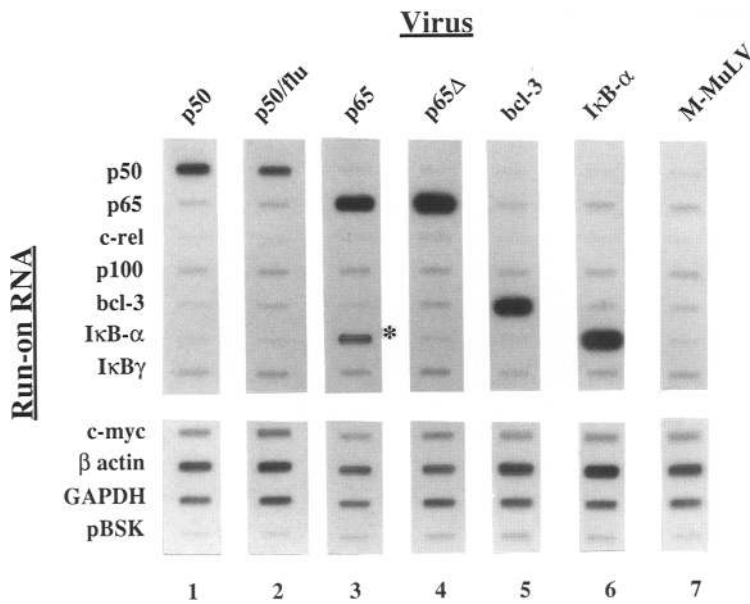


Figure 6. Nuclear run-on RNAs from Rel- and I κ B-expressing cells. Retrovirus stocks transducing the genes indicated at the top (Virus) were used to infect NIH-3T3 cells. Thirty-six hours after infection, 32 P-labeled nuclear run-on RNA was prepared from each population of infected cells and hybridized to slot-blotted plasmids containing the cDNAs indicated at the left (see Materials and methods). The asterisk (*) indicates the increased I κ B- α transcription in p65-expressing cells.

in the cell. In unstimulated cells, I κ B- α is probably made in excess, with the unbound fraction being continually degraded, such that fluctuations in the level of p65 will not lead to the unprogrammed nuclear appearance of NF- κ B (p50/p65) or (p65) $_2$. This suggests a mechanism by which cells that have cytoplasmic NF- κ B (p50/65) can also have nuclear (p50) $_2$ (KBF-1; Kieran et al. 1990). If p50 were made in excess over p65, its transport to the nucleus would not be inhibited by I κ B- α . Consistent with this idea, we have observed that overexpression of p50 induces a small amount of I κ B- α but has no effect on *bcl-3* or I κ B- γ , the two regulators that interact most strongly with p50 itself (Franzoso et al. 1992; Inoue et al. 1992; Liou et al. 1992; Wulczyn et al. 1992).

I κ B- α protein levels are also regulated by p65-dependent changes in I κ B mRNA. We have shown here that retrovirus-directed expression of murine p65 can cause at least a sevenfold increase in I κ B- α mRNA in transfected 293 cells. Nuclear run-on studies of RNA from NIH-3T3 cells demonstrated that most of the effect arises from increased transcription (Fig. 6). Although we cannot rule out a small contribution from post-transcriptional events, the simplest explanation for our findings involves the presence of κ B sequences in the I κ B- α promoter. These sequences could bind (p65) $_2$, p50/p65, or p65 complexed with another *rel*-related protein. Two recent findings are consistent with this hypothesis. First, we have demonstrated that (p65) $_2$ can function as a transcriptional activator in vitro (Fujita et al. 1992). Second, R. de Martin and F. Bach (pers. comm.) have sequenced the promoter region of a porcine genomic locus that encodes a structural and functional homolog of I κ B- α ; this promoter contains two κ B sites that are functional in EMSA assays. Increased transcription of I κ B- α is probably a physiologic response that can control increases in p65 or NF- κ B beyond those that can be accommodated by protein stabilization.

Localization

The fluorescence data on p65 subcellular localization that we have presented here differ from the results of Beg et al. (1992) and Zabel et al. (1993), who found that overexpressed p65 localized to the nucleus. At least three experimental details distinguish our work from theirs. First, we used wild-type p65 and I κ B- α detected with rabbit sera specific to them, not proteins amino-terminally modified with an influenza virus HA epitope detected with an epitope-specific monoclonal antibody. Although the epitope-tagged proteins are largely similar in in vitro function to their normal counterparts, it is sometimes difficult to be certain that their in vivo behavior is identical. To rule out this possibility, we have also carried out coinfection experiments similar to those shown in Figure 8 using viruses that encode wild-type p50 and p65. Detection with p50-specific antiserum revealed a population of cells expressing cytoplasmic p50, which is not seen in cells transduced by p50 virus alone (data not shown). Second, the other studies were carried out using vectors containing a cytomegalovirus (CMV) promoter that, in our hands, yields expression levels at least 10-fold higher than the retrovirus-based constructs that we have employed. This partly results from the presence of κ B sites in the CMV promoter itself. Expressed proteins may therefore up-regulate their own synthesis. A third potential explanation involves the cell types studied. Some studies (Beg et al. 1992) were carried out in COS cells that can amplify the transfected constructs, whereas our investigation employed renal carcinoma, pre-B, and B-lymphoid cells without the ability to amplify. It is our belief that the last two differences are probably the source of the discrepancy. We suspect that in our experiments, the amount of p65 does not exceed the capacity of a cell to respond by synthesis of an equimolar amount of I κ B- α . In the other studies there is prob-

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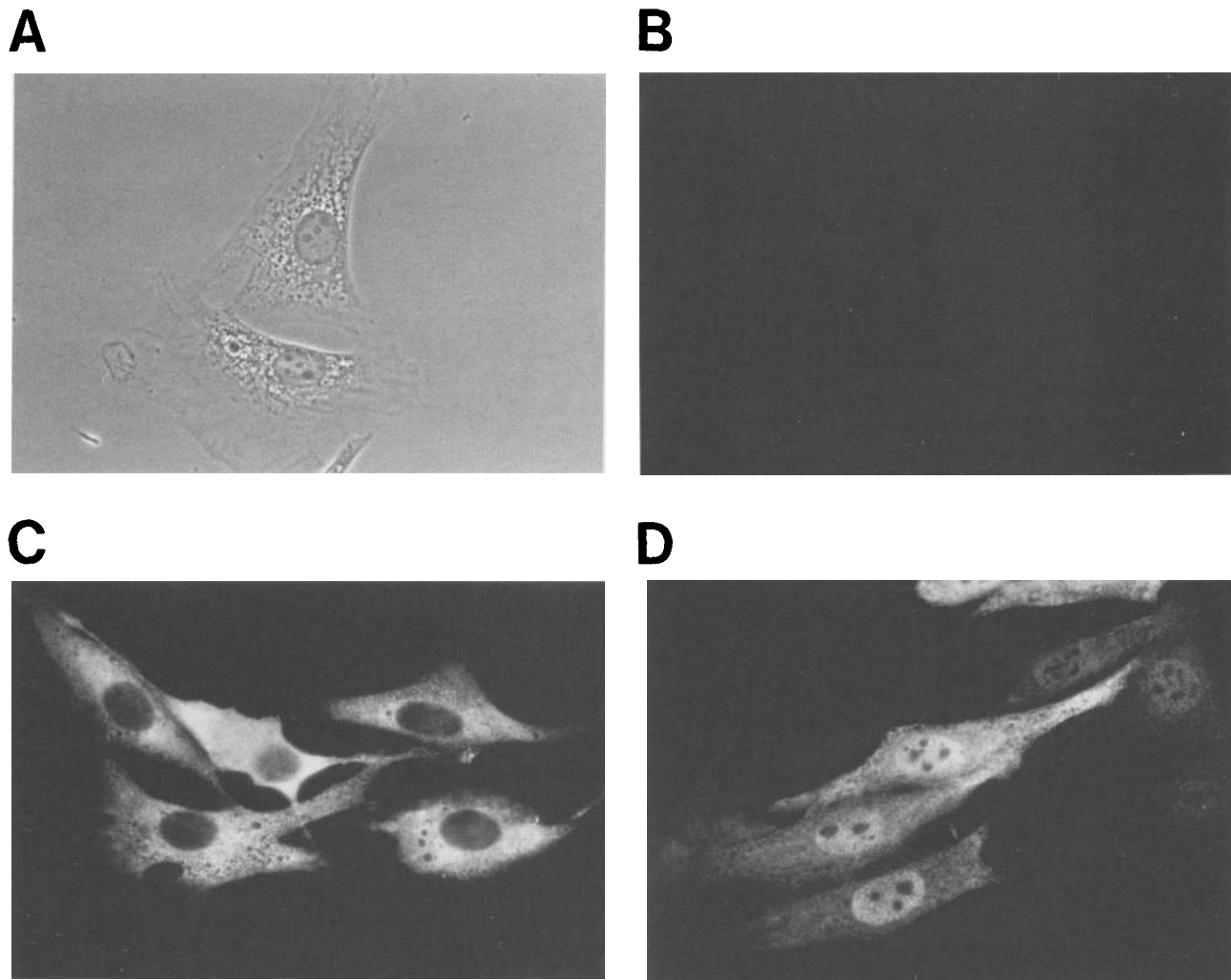


Figure 7. Immunolocalization of p65 and p65 Δ in NIH-3T3 cells. (A) A phase photomicrograph of cells infected with Moloney MuLV (helper) virus alone (60 \times objective). (B) A fluorescence micrograph of the cells seen in A. This sample was stained with p65-specific primary antibody (1 : 300 dilution) and secondary rhodaminated donkey antirabbit serum (1 : 200 dilution), and was then photographed under the conditions used to identify the cells shown in C and D. (C) NIH-3T3 cells infected with the p65-transducing retroviral construct pGD-p65-34 were stained with p65-specific serum and photographed (60 \times objective) under conditions identical to those employed in B. Greater than 50% of the cells in this sample expressed detectable p65 (all cytoplasmic). (D) NIH-3T3 cells infected with the p65-transducing retroviral construct pGD-p65- Δ 4 were stained with p65-specific serum and photographed (60 \times objective) under conditions identical to those employed in B. Greater than 30% of the cells in this sample expressed detectable p65 Δ , all with a mixture of cytoplasmic and nuclear staining.

ably such an excess of p65 that some or most of it is free from I κ B- α and is therefore transported to the nucleus. It seems unlikely that cells can make such an excess of p65 from their endogenous gene that it would overwhelm the I κ B response system. In this regard it is probably significant that transfection of a plasmid containing the genomic p65 promoter and natural exons causes increases in p65 and I κ B- α similar to those with retroviral constructs (W. Sha, M. Scott, and D. Baltimore, unpubl.).

The p65 Δ splice variant both fails to stabilize I κ B- α and fails to induce elevated transcription of I κ B- α mRNA, properties that likely cause its localization to the nucleus. It has recently been reported that p65 Δ interacts only weakly with I κ B- α , possibly because the splice variant has little ability to dimerize with itself or

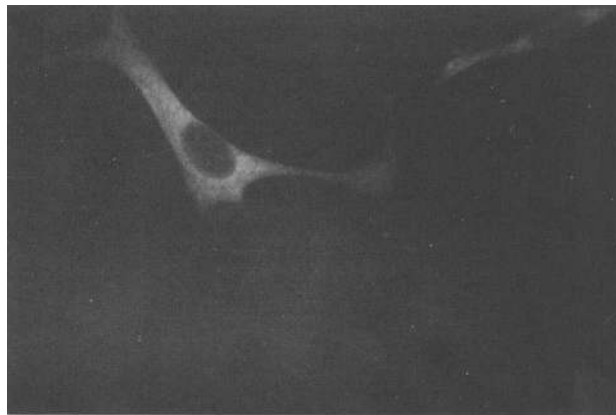
other Rel variants (Beg et al. 1992). Thus, it may be the affinity of I κ B- α for its full-length p65 target that stabilizes the inhibitor protein (Ganchi et al. 1992). Such high-affinity binding could mask a site of interaction with a proteolytic system or could be the signal for a modification of I κ B- α that makes it insensitive to a proteolytic system. How these properties relate to the transforming potential of p65 Δ (Narayanan et al. 1992) remains to be determined.

Materials and methods

Viruses and cells

p65 virus inserts were prepared by PCR using as template the murine p65-5 cDNA clone (Nolan et al. 1991). One microgram

A



C

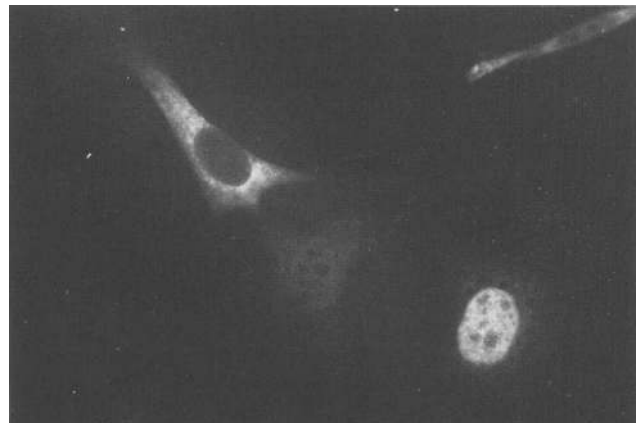


Figure 8. Immunolocalization of p50/flu and p65 in NIH-3T3 cells. (A) A phase photomicrograph of cells coinfecting with virus stocks transmitting p50 and p65 (60 \times objective). Cells were simultaneously stained with rabbit anti-p65 serum and a mouse monoclonal antibody specific for the influenza virus HA epitope. These antibodies were detected with fluorescein-conjugated donkey anti-rabbit serum and rhodaminated donkey anti-mouse serum as shown in B and C. (B) A photomicrograph of the same field of cells, using barrier filters to show green fluorescence (p65). Only the cell at the upper left exhibits cytoplasmic p65 protein. (C) A photomicrograph of the same cells seen in A and B, this time using barrier filters specific for red fluorescence (p50/flu). The cell at the lower right, containing only endogenous p65, shows a predominantly nuclear pattern of staining for p50. The cell at the upper left contains both retrovirally expressed p50 and p65; here, the p50 fluorescence is cytoplasmic.

of template DNA was amplified for 10 cycles under conditions including 10% DMSO and 4 mM MgCl₂ described by the manufacturer of *Taq* polymerase (Stratagene) to yield a 1.6-kb product with the following structure: *SalI*-*EcoRI*-Kozak-p65-*EcoRI*-*Clal*. After gel isolation, this product was inserted into the pGD retroviral vector (Daley et al. 1990) at its *XhoI* and *Clal* sites, removing the neomycin-resistance cassette from the virus. Three independent clones (pGD-p65-28, pGD-p65-34, and pGD-p65-37) were identified; all yielded the same increase in I κ B- α levels described in the text. A p65 Δ mutation, deleting amino acids EDIEVYFTGP from the murine cDNA, was introduced using a 30-base oligonucleotide to prime second-strand synthesis of uracil-substituted pBSK-p65-5 phagemid DNA (Ausubel et al. 1989). A clone containing the p65 Δ mutation was identified by DNA sequencing. The *SphI*-*BsiWI* fragment carrying this deletion was inserted into the pGD-p65-34 retroviral construct. Clones pGD-p65 Δ -4 and pGD-p65 Δ -11 were selected on the basis of *SphI*, *BsiWI*, and *SacI*-*EcoRI* digestion (the last yielding a 228-bp fragment 30 bp shorter than wild type) for further characterization. The human I κ B- α (MAD-3) cDNA was amplified under the conditions described above, using 51-mer and 48-mer primers to generate a *BamHI*-*EcoRI*-I κ B- α -*EcoRI*-*SalI* product. This gel-isolated product was ligated to a modified pGD vector generated by *XhoI* digestion of the parent plasmid, followed by addition of a *BclII* linker to the blunted site. The *BclII*

cloning vector (pGDB) so generated was digested with *BclII* and *SalI* to allow insertion of the corresponding MAD-3 cDNA. Retroviral constructs encoding murine *bcl-3* and influenza HA epitope-tagged or wild-type murine p50 were also prepared and are described elsewhere (Nolan et al. 1993).

Virus stocks and virus-producing cells were prepared by the method of W. Pear, G. Nolan, M. Scott, and D. Baltimore (in prep.) using pGD-p65, p65 Δ , or I κ B- α constructs and a replication-competent helper virus clone pZAP (Goff et al. 1982). In cocultivation experiments, 70Z/3 or WEHI 231 cells (10⁶/ml in RPMI) were added to the adherent producer cell culture. Twenty-four hours thereafter, the medium containing nonadherent cells was aspirated and lysates for Western blotting were prepared as described below.

Protein immunoblotting

Lysates of transfected 293 cells or infected lymphoid cells were prepared at either 48 hr or at the times indicated after the addition of the p65 or I κ B- α DNAs as calcium phosphate precipitates. Cells were washed once with cold calcium/magnesium-free PBS and scraped from the plates in RIPA buffer [25 mM Tris-Cl at pH 7.4, 0.5% Triton X-100, 0.5% deoxycholate, 0.1% SDS, 0.5 mM EDTA, 150 mM NaCl, 1 mM PMSF, 10 μ g/ml aprotinin (Sigma)], 1.0 ml/100-mm plate of cells. After clarifi-

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cation in an Eppendorf centrifuge (14000g for 10 min), 5–10 μ l of each boiled lysate was applied to 10% polyacrylamide–SDS gels, electrophoresed, and transferred to supported nitrocellulose.

Rabbit sera specific to murine p65 were obtained after immunization with full-length protein derived from a baculovirus expression construct (Fujita et al. 1992). The serum was used at a dilution of 1 : 1000 on immunoblots. Affinity-purified antibodies specific to I κ B- α were prepared by immunizing rabbits with a glutathione S-transferase/I κ B- α fusion protein (Liou et al. 1992) and passing the immune sera over an Affigel 15 matrix (Bio-Rad) to which I κ B- α fusion protein had been coupled as per the manufacturer's instructions. Antibody bound to the column was eluted as specified (Harlow and Lane 1988) and used at a concentration of 0.5–1.0 μ g/ml. An alkaline phosphatase-coupled donkey anti-rabbit preparation (Jackson ImmunoResearch) was used to detect bound primary antibody in transfected cells.

In vivo protein labeling

293 cells seeded in 60 mm plates at 2.5×10^5 cells per plate were transfected with 5 μ g of retroviral DNA per plate and incubated 48 hr before labeling. After a 30-min period of starvation in methionine-free Dulbecco's minimal essential medium (DMEM) supplemented with 5% dialyzed fetal calf serum, fresh labeling medium with [35 S]methionine (Amersham, 800 Ci/mmol, 200 μ Ci/ml) was added for 30 min. At the end of labeling, radioactive medium was aspirated and replaced with standard DMEM/10% FCS (time 0). Lysates from the subsequent time points were prepared as described above and frozen at -70°C before immunoprecipitation. Bicinchoninate assays (Pierce) revealed less than twofold differences among the protein concentrations of all lysates. Each lysate was precleared by the addition of 5 μ l of preimmune rabbit serum for 30 min on ice; this serum was then collected on a 20- μ l bed volume of protein A–Sepharose 4B (Pharmacia) for 30 min.

To precipitate p65 from the precleared lysates of cells transfected with pGD–p65, 5 μ l of specific serum was added. After incubation on ice for 60 min, another 20- μ l bed volume of protein A–Sepharose (precoated with a cold lysate from untransfected 293 cells) was added for 30 min. The protein A–Sepharose beads were washed twice in RIPA buffer, twice in high salt buffer (1% Triton X-100, 0.5 M NaCl, 20 mM Tris-Cl at pH 7.6), and twice in low salt buffer (1% Triton X-100, 10 mM NaCl, 20 mM Tris-Cl at pH 7.6). After the last wash, each pellet was resuspended in 30 μ l of protein sample buffer and boiled for 5 min. Five microliters of each sample was loaded onto SDS–polyacrylamide gels and visualized by autoradiography.

A similar protocol was used to precipitate I κ B- α protein from cells transfected with either the p65 or I κ B- α expression constructs. Instead of whole sera, affinity-purified antibody (5 μ g per lysate) was used. As an additional demonstration of specificity, 5 μ g of affinity-purified antibody to I κ B- α was preincubated with 50 μ g of fusion protein for 30 min on ice and then added to precleared lysates and processed as described above. This preincubation eliminated the 38-kD-labeled band precipitated from either p65 or (I κ B- α)-transfected cells (not shown).

Immunofluorescence

Virus stocks transducing the p50, p65, and p65 Δ genomes were prepared by transient transfection of 293 cells as described above and used to infect NIH-3T3 cells. 3T3 cells were seeded onto glass coverslips at 10^4 cells/well in 24-well plates. The cells were exposed, 24–36 hr later, to the indicated viruses in 4 μ g/ml of Polybrene for 3 hr. For mixed infections, equal vol-

umes of viral supernatants from 293 cells singly transfected with p50 or p65 constructs were added to the plate of 3T3 cells. Cells were then refed with DMEM/10% calf serum and incubated for 36–48 hr. (For some experiments, cells were infected at the indicated density in 35-mm plastic dishes, incubated for 24–48 hr, and seeded onto coverslips for staining 1 day thereafter.) After washing in PBS, coverslips were fixed in PBS/4% paraformaldehyde for 10 min, quenched with 50 mM glycine (pH 8.0) in PBS for 10 min, and permeabilized with 1% Triton X-100/PBS for 10 min. Cells remaining on the coverslips were then blocked in a solution of PBS, 5% normal donkey serum, and 1 mg/ml of BSA for 60 min. Primary rabbit antiserum specific to p65 was diluted 1 : 300 in blocking solution and added to the coverslips for 45 min. The coverslips were subsequently washed for 4–12 hr in three or four changes of PBS and reblocked for 30 min. Secondary antibody (rhodaminated donkey antirabbit Fab₂; Jackson ImmunoResearch) diluted 1 : 200, as described above, was added for 30–45 min and washed away in PBS (two or three changes) for 1–3 hr before inversion and mounting on glass slides. Cells were visualized and photographed using a Nikon fluorescence microscope with a 60 \times objective.

RNA isolations and hybridizations

293 cells (10^7 cells/150-mm plate) were transfected with p65, p65 Δ , or empty vector (pGD) retroviral DNAs and incubated at 37°C for 48 hr. After lysis in an NP-40-containing solution as per standard procedures (Ausubel et al. 1989), total cytoplasmic RNAs obtained from each population of transfected cells were applied to an oligo(dT)–cellulose column. Samples (2.5 μ g) of each poly(A)-containing RNA were electrophoresed through formaldehyde–agarose gels, transferred to nitrocellulose, and hybridized to ^{32}P -labeled probes derived from gel-isolated p65 and MAD-3 cDNAs. Probe bound to each filter was detected by exposure to XAR film and quantitated by using a PhosphorImager (Molecular Dynamics). The filters were stripped and rehybridized to an actin probe, and the process was repeated to demonstrate equal loading of RNA in each lane.

Nuclear run-on assays were performed as described (Greenberg and Ziff 1984) using nuclei isolated from three 150-mm plates of NIH-3T3 cells, 36 hr after infection, with the indicated virus stocks prepared as described above. Plasmids (7.5 μ g) encoding p50 (Ghosh et al. 1990), p65 Δ (above), *c-rel* (Bull et al. 1990), human p100 (Schmid et al. 1991), *bcl-3* (Nolan et al. 1993), human I κ B- α , murine I κ B- γ (Liou et al. 1992), *c-myc* (Saksel et al. 1991), glyceraldehyde phosphate dehydrogenase, and β -actin were linearized, phenol extracted, and slot blotted to supported nitrocellulose before hybridization with labeled RNA.

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