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The piriform, perirhinal, and entorhinal cortex in seizure generation

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Abbreviations:

4-AP – 4-aminopyridine

AOAA – amino-oxyacetic acid

AMPA – alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid

AT – area tempestas

CPP – 3-(2-carboxypiperazin-4-yl)propyl-1-phosphonic acid

EPN – endopiriform nucleus

EPSP – excitatory postsynaptic potentials

ERC – entorhinal cortex

GABA – gamma aminobutyric acid

NBQX – 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzo (f)-quinoxaline

NMDA – N-methyl-D-aspartate

PIRC – piriform cortex

PRC – perirhinal cortex

PSA-NCAM – polysialylated neural cell adhesion molecule

TLE – temporal lobe epilepsy

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ABSTRACT

Understanding neural network behavior is essential to shed light on epileptogenesis and seizure propagation. The interconnectivity and plasticity of the limbic and cortical regions of the mammalian brain provide the substrate for the hypersynchrony and hyperexcitability associated with epilepsy and seizure activity. Recurrent unprovoked seizures are the hallmark of epilepsy, and limbic epilepsy is the most common type of medically-intractable focal epilepsy in adolescents and adults that necessitates surgical evaluation. In this review, we describe the role and relationships among the piriform (PIRC), perirhinal (PRC), and entorhinal cortex (ERC) in seizure-generation and epilepsy. The inherent function, anatomy, and histological composition of these cortical regions are discussed. In addition, the neurotransmitters, intrinsic and extrinsic connections, and the interaction of these regions are described. Furthermore, we provide evidence based on clinical research and animal models that suggest that these cortical regions may act as key seizure-trigger zones and, even, epileptogenesis.

1. INTRODUCTION:

The rhinal and piriform cortices are highly epileptogenic regions that are broadly interconnected with other limbic and cortical areas. The functional architecture of these regions involves oscillatory network activity commonly associated with memory processing. As a result, the unique anatomical organization and connections of these regions can also lead to pathological hypersynchronization resulting in seizures. For example, the piriform cortex (PIRC) is part of the primary olfactory cortex, and odor can be a powerful trigger for memory [1]. The long-range projections of piriform cortex, including interconnections with amygdala, hippocampus, and rhinal cortex place this structure as a crucial hub in the ability of an odor to “trigger a feeling”, stir an autobiographical memory, or trigger seizure activity throughout the limbic system. The rhinal cortex is divided into the entorhinal (ERC) and perirhinal cortices (PRC), which serve as the major cortical communication relays for the hippocampus. Information flow from the rhinal cortex, through the hippocampus, and back to the rhinal cortex is a perfect example of a reverberatory, Hebbian cell assembly. Under normal conditions, this architecture plays a crucial role in the processing of declarative memory. Under pathological conditions, the rhinal-hippocampal loop tends towards excessive propagation and loop-gain amplification, which is a hallmark of temporal lobe epilepsy [2]. Herein, we review: 1) the normal anatomy and functional connectivity of these structures with an emphasis on the features that make them particularly prone to seizure initiation, 2) evidence for epilepsy-related pathology in these sites, and 3) evidence supporting the notion that these features make these sites critical “trigger zones” for seizure initiation in the temporal lobe.

2. PIRIFORM CORTEX (PIRC):

PIRC Functions

The PIRC is a part of olfactory cortex and its typical activities are olfactory processing and memory coding [1]. The posterior PIRC is activated in tasks that require categorization of odors [3]. Closely associated with the PIRC is the endopiriform nucleus (EPN), a large mass of multipolar cells located within the PIRC and has been implicated in seizure generation and propagation [4–12]. Although its functions remain largely unknown, the EPN may have a role in integrating gustatory and olfactory sensory information [13]. Furthermore, PIRC and EPN may also produce olfactory and gustatory semiology when affected by ictal discharges.

PIRC Anatomy and Histology

The PIRC is a large, cytoarchitectonically homogeneous area that extends over the ventrolateral surface of the forebrain, and constitutes a major component of primary olfactory cortex [1]. The PIRC consists of three layers: superficial plexiform layer (layer I), cell body layer (layer II), and layer III containing soma and fibers [14]. Adjacent to layer III is the well-defined EPN comprised of multipolar cells [15]. The EPN is also referred as layer IV of the PIRC since it spans the entire rostro-caudal dimension of the PIRC. The dorsal region of this nucleus consists of densely packed cells while the ventral region has more diffusely arranged cells [16]. The ventro-rostral aspect of the PIRC encompasses a functionally defined and histologically distinct chemoconvulsant trigger zone, located deep within the anterior PIRC, named area tempestas (AT) [4,5,17]. Compared to the rest of the PIRC, studies have demonstrated that AT possesses a decreased number of GAT-1 immunoreactivity with decreased density of GABAergic inputs at the axonal initial segments along with decreased concentration of cholecystinin positive interneurons [17]. EPN shares a number of properties with the PIRC such as excitatory discharges, hyperexcitability, and high speed of signal propagation. Together these features lead to a high potential for epileptogenicity [6,9,18,19]. Moreover, both the PIRC and EPN are exceedingly vulnerable to excitotoxic injury, and represent one of the earliest sites of neuronal

loss after status epileptics [20]. As discussed below, the extensive interconnections of the PIRC/EPN with other highly excitable brain regions may allow for the generation and spread of epileptiform activity within PIRC/EPN and throughout the brain [21].

PIRC Connections

The integrative connections between PIRC and several limbic nuclei likely facilitate propagation of ictal activity between these structures (Fig. 1). Afferents from the ERC constitute the most dense connections of PIRC [22]. In turn, the PIRC and EPN send efferent projections to the ERC [23,24]. There is also a direct connection from the PIRC to the amygdala [25,26] and a diffuse polysynaptic connection from the amygdala to the PIRC via the EPN [16], which may have significant implications for temporal lobe epilepsy (TLE). There are also pathways from the PIRC to the subiculum [16,23] and to areas 35 and 36 of PRC [24].

There are significant differences in the connectivity of the anterior and posterior subregions of the PIRC [27,28], suggesting functional specialization within the PIRC [29], which may be of particular importance to the functional topography related to seizure initiation. For example, local circuit GABAergic transmission appears to be stronger in the posterior subregion of the PIRC and become increasingly less prominent more anteriorly [21,27]. Interestingly, AT, crucial to seizure generation [4], is located in the anterior aspect of the PIRC, and has low density of GABAergic inputs to the axon initial segments [17]. Unlike the rest of the PIRC that receives olfactory input from mitral cells only, the AT also receives direct projections from tufted cells in the olfactory bulb [17] and provides most input from the PIRC to ventrolateral orbital cortex. These connections are both directly and via di- and trisynaptic connections incorporating pre-endopiriform nucleus and submedial and mediodorsal thalamus [17,29]. This interconnectivity may be important for seizure propagation and generalization.

Projections from the PIRC and EPN are very diffuse. Single neurons within posterior PIRC send axonal arbors to the insula, PRC, ERC, amygdala, and olfactory tubercle, with a

tendency to have no overlap with the arborization patches created by neighboring neurons [30]. Projections of the EPN parallel those of the PIRC, targeting ERC, PRC, insular, orbital, and other neocortical structures [8]. The EPN is the major route for information transfer along the rostrocaudal axis of PIRC/EPN and may facilitate seizure propagation longitudinally within this structure [8]. Moreover, EPN may play an especially important role in the spread of activity from EPN and PIRC to other cortical regions, because the EPN has a more diffuse topographic and laminar organization of axon terminals than does PIRC [8,23,31]. The extensive interconnectivity of EPN and PIRC with other epileptogenic sites within the temporal lobe (e.g., ERC, PRC) may provide a loop for positive feedback of activity and subsequent generalization of epileptic discharges.

3. PERIRHINAL CORTEX (PRC):

PRC functions

The PRC is involved in complex memory functions [32–34], such as object recognition, sensory representation, and spatial orientation [35]. Damage to PRC results in deficits in complex visual discrimination [36], object recognition memory [37,38], and attention to visual stimuli [39]. The PRC represents a component of the ventral visual stream, with interconnections to secondary visual regions being more robust than those with primary visual cortex [24]. This may support its involvement in higher visual information processing and memory [40] and may account for complex visual hallucinations in some temporal lobe seizures. Induction of theta activity in the PRC network results in the recall of vivid memories [40] and therefore, can represent a substrate for visual auras that accompany some TLE. PRC damage may also account for the feeling of *déjà vu* reported by a subset of TLE patients during the aura of their seizures [41]. Extensive damage to PRC in epilepsy may explain the cognitive decline, including primarily memory deficits [42,42] that occur in some patients with chronic TLE.

PRC Anatomy and Histology

The human PRC is located along the collateral sulcus in the ventromedial portion of the temporal lobe. Laterally, it borders the ERC with which it comprises the parahippocampal gyrus [43]. On the basis of cytoarchitectural, immunohistochemical, and physiological criteria, the PRC is further subdivided into two parallel cortical bands, areas 35 medially, and 36 laterally [43–45].

PRC Connections

Afferents: Both Areas 35 and 36 of PRC receive substantial input from ERC (Fig. 1). In addition to this input, PRC receives projections from ventral temporal association areas, PIRC, and other limbic-cortical areas (e.g., insula). The relative abundance and distribution of these inputs varies as a function of subregions of PRC (i.e., Area 35 vs. 36) [22].

Efferents: The PRC projects back to many of the same regions it receives input [22–24,31]. For example, the PIRC, insula, and subcortical limbic structures all have reciprocal connections with the PRC. Just as the pattern of afferents differs between Areas 35 and 36, the patterns of efferents also differ [24]. The PRC may represent a route by which seizure activity can propagate to distal cortex due to the heavy connections with the insula and sensory domains within frontal, temporal and parietal cortices. These projections may also help explain sensory symptoms associated with TLE including gustatory or autonomic symptoms related to insular cortex activation. There is a high degree of specialization within various subregions of the PRC (e.g., rostral PRC projects heavily to somatosensory cortex, while caudal PRC projects heavily to visual cortex) [24]. A relatively underexplored question is the degree to which topographic activation of the PRC may be associated with different patterns of propagation and/or symptoms associated with TLE.

The PRC is a “gate-keeping” area that mediates bidirectional information transfer between

temporal neocortex and ERC [46]. It is worth noting that while most communication takes place through a slow, stepwise progression through cortical areas, a minor portion of axons bypass the PRC [45] directly activating the lateral ERC. Moreover, some ERC axons extend to area 35 and the temporal neocortex. These projections may allow for spread of pathological excitatory activity by circumventing the PRC gate. In conditions such as chronic epilepsy, the reciprocal interconnectivity of PRC with other hyperexcitable structures positions the PRC as a relay region within the limbic network for the generation and propagation of epileptiform activity.

4. ENTORHINAL CORTEX (ERC):

ERC functions

The ERC, located in the anterior parahippocampal gyrus, serves as the major interface between the hippocampus and sensory cortices [43]. Its close connection with cortical regions involved in the processing of visual and spatial information suggests that the ERC is an important substrate for visuo-spatial functioning [22,47]. Moreover, hippocampal memory function depends on an intact ERC [48] and appears to be well-conserved across species. For example, human neuroimaging studies have found correlations between parahippocampal activity and spatial working memory [49], and with learning and recall of topographic information [50]. In monkeys, single neurons within the ERC fire in response to visual input from specific locations in the environment, forming memories of spatial environments [51]. Moreover, “grid neurons” of the ERC have been well-described in rodents and primates [47,48,52,53]. As with PIRC and PRC, the extensive connections between ERC and other limbic brain regions suggest it is a major relay for seizure activity.

ERC Anatomy and Histology

In humans, the ERC, consisting of Brodmann areas 28 and 34, is situated bilaterally

between the sulcus semiannularis and sulcus rhinalis in the rostral parahippocampal gyrus [54,55]. It is an allocortical structure, consisting of six layers, with a striking paucity of cells in layer IV (lamina denticans), which separates the outer cell-rich layers (V and VI) from the inner cell-rich layers (II, and III) [55,56]. Layer III may be particularly important for seizure propagation as it consists of pyramidal projection neurons that target the hippocampus and other regions. For a detailed review of ERC anatomy and cytoarchitectonics, see Insausti (1993) [57].

Like the PRC, the ERC is divided in two major sub-fields, the rostral-lateral (LERC) and the caudal-medial ERC (MERC) [58,59]. These two divisions differ in their histology, morphology and physiology. The LERC has a number of morphological inhomogeneities and the neurons are organized in patches surrounded by thick radially-oriented bundles of fibers. Conversely, the MERC is more reminiscent of the neocortex, with a distinctly laminar appearance and columnar organization of neurons and fibers [55]. As will be discussed in the later sections, this laminar structure may have a special role in signal amplification and epileptogenesis.

ERC Connections

Afferents: A major input to both LERC and MERC originates within PIRC and accounts for about one-third of the input to these regions (Fig. 1). Other inputs originate in temporal and frontal cortices. Less abundant sources of input include insular, cingulate, parietal, and occipital cortex, with the relative abundance of these inputs varying between the MERC and LERC.

Efferents: In addition to its rostro-caudal organization, the ERC is also organized into bands that project to different septo-temporal levels of the dentate gyrus [24,58]. As other structures discussed in this review, the distribution of efferent projections differs between subregions of ERC. PIRC is the major recipient of input from both subdivisions of ERC of projections to inferotemporal, retrosplenial, posterior parietal, frontal, and insular cortices originating in medial and lateral domains. Similar to PIRC and PRC, many of the connections from ERC are

reciprocated [60,61]. This ability for bidirectional information transfer, while presumably important for normal information processing, is yet another example of a cortico-cortical loop that can amplify and propagate seizure discharges.

The hippocampus is a key area to seizure propagation and has a series of parallel ERC-hippocampus-ERC loops that underlie memory functions [62–64]. In the TLE, one of the most common types of medically intractable epilepsy [65,66], ictal synchronization within hippocampus is a pre-requisite to spread of activity to other cortical areas [67–69] and the hippocampal-ERC communication may serve as a critical substrate for propagation of epileptic activity of ERC origin to hippocampus.

Intrinsic circuitry

Despite histological data that strongly suggest the presence of substantial fiber tracts between the ERC and PRC, the functional strength of connections in the healthy brain is debatable [60]. Previously, the PRC was considered to be a relay nucleus for the bidirectional communication between the neocortex and the ERC. However, more recent data suggests the PRC is an important center for feed-forward inhibition of the ERC [45,46,60,70]. Extracellular recordings and optical imaging studies demonstrated that PRC–ERC and ERC–PRC interactions produce only limited neuronal activation with a relatively low probability for the transfer of neocortical inputs to ERC. This highly restricted and controlled information transfer is essential in a recurrent circuit to prevent loop-gain amplification. Therefore, in the healthy brain, the excitatory activity of the ERC is controlled by inhibitory interactions in the network that are activated by extrinsic projections. The PRC may act as a “gate” for spread of excitatory activity between the ERC and neocortex. While conditions of normal hyperexcitability can facilitate the propagation of olfactory-driven activity from the ERC to the PRC [71,72], malfunction of intrinsic

control mechanisms can result in hyperexcitability of the ERC and, consequently, epileptic seizures.

5. SUMMARY OF FUNCTIONAL ANATOMY:

Together, PIRC/EPN, PRC, and ERC form a unique and widely connected network that is ideally organized to engage the hippocampus (and the rest of the limbic system) in rhythmic activity. While this rhythmic activity is a part of normal brain function, malfunctions in the circuitry can easily lead to pathological synchronization and seizure spread. Seizures do not propagate randomly through the brain, but rather utilize circuits that normally support highly controlled recurrent activity. Through this network, activity can spread to distal cortical and subcortical regions along pathways that are also used in normal motor system [3,73–75].

In the sections above, we have described the anatomical features that position PIRC/EPN, PRC and ERC as sites through which seizure discharges can amplify and propagate. While these regions are capable of autonomous generation of ictal discharges, the importance of their interconnections is underscored by data demonstrating a significant reduction in ictal activity when the connections between these regions are disrupted [76]. These observations imply that the communication between the ictogenic structures is a key component for amplification and synchronization of ictal bursts and may have particular relevance to understanding the effects of temporal lobe resection for the treatment of epilepsy. Even “selective” amygdalohippocampectomy (aside from radiofrequency lesions) involve resection of the anterior aspects of the middle and inferior temporal gyrus, disrupting many connections intrinsic to the temporal lobe in addition to removing temporal cortical tissue, amygdala, and hippocampus.

Below we discuss conditions that may lead to failures in the intrinsic gating properties of these structures. These pathologies and subsequent aberrant gating are hypothesized to underlie the emergence of spontaneous seizures in chronic epilepsy.

6. MORPHOLOGICAL AND HISTOLOGICAL CHANGES DUE TO EPILEPSY:

The transition from normal network activity to hypersynchronous activity characteristic of a seizure may indicate a subtle change in the interconnections or functions within PIRC, PRC, ERC, or the broader limbic network. A common histopathological finding in TLE is neuronal loss and inflammation. It is possible to speculate that these changes may result in the network perturbations necessary for epilepsy or the comorbid memory and cognitive deficits. However, the degree to which these histopathological findings are a cause or an effect of recurrent seizures remains a matter of debate. In the section below, we will discuss evidence for neuropathology in PIRC, PRC and ERC in the context of epilepsy and epileptogenic insults.

Neuroimaging

In patients with TLE, volumetric analyses reveal volume loss of the anterior temporal lobe on the epileptic side [41,77–79], bilateral reduction in temporal neocortical gray matter, and reduction in temporal white matter [80]. Changes in volume of temporal cortex appear to be specific to TLE, and are not seen in extratemporal epilepsy [77,79]. Analysis of substructures within the TLE has also reveal volumetric changes, including in the hippocampus, PRC, and ERC [79]. In one study, 52% of patients with medically intractable TLE have reduction in ipsilateral ERC volume [77]. Moreover, the extent of atrophy in ERC correlates with ipsilateral hippocampal volume and the duration of epilepsy [77].

While it remains controversial if damage to piriform and rhinal structures leads to epilepsy, or rather if the recurrent seizures characteristic of epilepsy lead to damage, some patients have atrophic tissue in the temporal lobe at what appears to be the site of seizure origin. For example, in some patients with TLE, ERC appears to display epileptiform activity before the hippocampus, but only in patients with atrophy of ERC [81]. Interestingly, the degree of atrophy in ERC correlates with hypersynchronization between ERC and hippocampus. These data suggest that the atrophic ERC may be the epicenter of epileptogenic activity with dissemination to other limbic structures via an elaborate network of neuronal connections of the ERC.

Histopathology

Morphological changes, such as neuronal loss and reactive gliosis, are prominent both in hippocampus and extrahippocampal areas (e.g., ERC, PRC) in patients with TLE [20,82,83]. Animal models parallel these findings with ERC, PIRC, and PRC showing some of the most severe damage as soon as 72 hours after status epilepticus [20]. Even sublayers of these structures show differential vulnerability. For example, there is a profound and relatively selective loss of neurons within layer III of the ERC in patients with TLE [82]. In animal models, loss of neurons within layer III of ERC can occur as soon as 24 hours after injury, and progresses with time [84]. Morphological changes in the CA1, CA3, and the hilus of the dentate gyrus showed a continuous degeneration of neurons up to 7 days following experimental status epilepticus [85]. Synaptic reorganization and atrophy of layer III of the ERC produces a robust change in evoked and spontaneous activity in parts of the ERC and hippocampus that result in increased excitability and susceptibility to reentrant activation of the hippocampal–ERC loop via disinhibition of local networks in CA1 [86].

Interestingly, neuronal loss in layer III shows a marked sparing of parvalbumin-expressing interneurons within the medial ERC [87] with predominate loss of glutamatergic neurons[84] suggesting that a simple depletion of inhibition is insufficient to account for the hyperexcitable phenotype. However, others have described depletions of GABAergic neurons and a reduction in the number of GABAergic synapses after status epilepticus[88] suggesting that loss of inhibitory tone may contribute to a hyperexcitable state permissive to seizures. In addition, low-threshold-spiking (LTS) and fast-spiking (FS) interneurons within the medial entorhinal cortex creates a recurrent inhibitory network which is central to stability in grid formation [89].

In addition to the cell loss associated with excitotoxic seizure injury (i.e. status epilepticus), the activation and proliferation of microglia may contribute to secondary injury. For example, within the rodent hippocampus, prolonged seizure activity can trigger the proliferation and differentiation of neural progenitor cells [90,91], resulting in the aberrant and ectopic neurogenesis in the hilus of the hippocampus. However, in extrahippocampal sites (i.e. PIRC, ERC) cellular proliferation and differentiation manifests as increased numbers of astrocytes and oligodendrocytes [83]. The production of chemokines by prolonged seizures may be of particular importance for triggering aberrant neurogenesis in the hippocampus, reactive gliosis, and astrocytosis in the temporal cortex [83].

As soon as five hours after generalized seizures, a marked elevation in the expression of IL-1 β can be detected in the endothelial cells of the blood vessels in the PIRC [11]. IL-1 β is a modulator of inflammatory response and apoptosis [92,93]. The effect of IL-1 β and TNF- α on hyperexcitability and seizure development have been shown in several studies [94]. However, some evidence suggest that TNF- α may decrease glutamate-mediated excitotoxicity via NF-kappaB-dependent up regulation of K2.2 channels [95]. In the PRC, astrocytes and microglia also express IL-1 β and display vascular endothelial expression of TNF- α , another vital

component of inflammatory response [92]. Within six weeks after generalized seizures, histopathological changes such as reactive astrocytosis and irregular microvascular proliferation appear in the allocortex including the PIRC, PRC, and ERC. Perivascular infiltration of C3 positive leukocytes has also been observed [96]. These changes are suggestive of inflammation and remodeling and provide a link between the epilepsy-induced inflammatory vascular processes, gliosis, and subsequent allocortical atrophy [11].

Reactive astrocytes and microglia are other markers of inflammatory process and appear in the areas of neuronal deterioration as early as 24 hours after the induction of kainate-induced animal model of status epilepticus. Inflammatory cells continue to rise, becoming prominent after 8 days with their number peaking at 30 days and subsequently declining almost to the baseline by 3 month after status; the initial rise in these cells mirrors the time course of spontaneous seizures in this model [97].

Markers of neuronal remodeling and synaptic plasticity, such as polysialylated neural cell adhesion molecule (PSA-NCAM), are increased in ERC and hippocampus from human patients with TLE [98] and in rodent models of TLE [99,100]. Changes in neuromodulatory systems and co-transmitters also occur in TLE with striking increases in NPY and somatostatin immunoreactivity and dysregulation of NPY receptor expression in PIRC and ERC [96,101–103]. In TLE and its models, it is difficult to determine if changes in plasticity represents a compensatory or decompensatory response to initial injury. For a more detailed review of the role of inflammation in TLE, see Vezzani *et al.* (2011) [94].

In summary, epileptic seizures are associated with significant alterations in the histology of the PRC, PIRC and ERC. Cell loss, inflammation, and altered gene expression may shift the balance between inhibition and excitation to create a pro-epileptic environment. Because these structures gate information transfer within the temporal lobe and extratemporal targets, atrophy in these regions may facilitate seizure propagation throughout the forebrain.

7. EPILEPTOGENECITY:

Piriform Cortex - Area Tempestas

The deep anterior piriform cortex contains a highly circumscribed and functionally-defined chemoconvulsant trigger zone called area tempestas (AT). From this site, unilateral microinjection of picomole amounts of GABA receptor antagonists or glutamate receptor agonists can trigger limbic motor seizures in rats and nonhuman primates [4,5]. More recently, neuroimaging approaches have found focal fMRI activation and reduced GABA_A receptor binding in the anterior piriform cortex of human patients with focal epilepsy [10]. Not only is activation of AT sufficient to trigger complex partial seizures in animals, it also appears to be necessary for the response to systemically applied chemoconvulsants (i.e. bicuculline) [4,5] and nerve agents (i.e. soman) [104,105]. Enhancement of GABA-mediated inhibition within this structure markedly attenuates seizure activity evoked by systemic chemoconvulsant administration [4,5]. In adult animals, AT is generally accepted as a zone of high epileptogenicity, while in immature rats, there was no difference between the rate of kindling from AT and PIRC [14]. These findings suggest that developmental changes in excitability within these subregions may modulate their epileptogenicity.

Some studies suggest a crucial role for the deep layers of the PIRC and/or the EPN in the induction of epileptiform bursting in slice preparations [18,106,107]. While *in vitro* studies have demonstrated that PIRC/EPN slices are highly excitable across the entire anteroposterior axis, *in vivo* studies demonstrate that the anterior PIRC/EPN has the highest susceptibility to seizure initiation [4]. The differences between these findings may be explained by the nature of the slice model utilized in which the entire slice was disinhibited by reduction in extracellular chloride [7].

Seizure propagation from AT requires both the posterior PIRC and PRC; pharmacological inhibition of these structures significantly attenuates the severity of seizures evoked from AT [108]. Likewise, blockade of excitatory neurotransmission within PRC suppressed amygdala kindled seizures [109]. Together, convergent evidence from experiments in acute slices, pharmacological manipulation of the rat brain *in vivo*, and human neuroimaging provide evidence that PIRC, and in particular, AT, is a crucial site for seizure initiation, and PRC is a key relay for seizure propagation within the limbic network. In the sections below we will outline additional evidence from pharmacological and kindling studies supporting these conclusions.

Kindling

Animal epilepsy model studies using kindling suggest that PIRC and PRC have particularly low thresholds for epileptogenesis; kindling from these structures occurs at a faster rate (i.e., with fewer stimulations necessary to trigger Stage 5 seizures) as compared to other forebrain structures [110,111]. Interestingly, the faster rate of kindling from PIRC/PRC was seen in both kindling-prone and kindling-resistant rat strains [111]. Within PIRC, cells in Layer III have the lowest afterdischarge threshold, but electrode placement within the EPN results in the fastest rate of kindling [21]. PIRC lesions have only mild effects on kindling: Lesions to the anterior PIRC have no effects, lesions to the posterior PIRC increase afterdischarge thresholds but do not alter kindling rate, and lesions to central PIRC increase the number of Stage 3 seizures before animals fully kindled [112,113]. Low frequency stimulation of the central PIRC has a similar effect of hindering generalization of focal seizures [114] and decreasing the duration and stage of seizures in fully kindled animals. Thus, the PIRC is readily capable of kindling and plays an important role in development and generalization of seizures of different origins, which may have tremendous therapeutic potential. The PIRC may be the site of interictal discharge origination within the limbic circuitry of kindled animals – regardless of the

site of kindling [115]. Within PRC, kindling not only occurs with the fewest stimulations of any site within the forebrain, but the latency from stimulation to behavioral seizures is very brief. This brief latency, along with extensive projections from PRC to frontal cortex led McIntyre and Kelly to propose that PRC is a crucial relay that clonic motor convulsions are triggered after limbic seizures [71]. Supporting this hypothesis, damage to perirhinal cortex prevents motor seizures evoked by hippocampal kindling [71]. While both the PIRC and PRC are highly susceptible to generating epileptiform discharges, the PRC is generally involved in generating seizures in naïve animals, though the role of epilepsy trigger zone shifts to PIRC as a result of the process of epileptogenesis [110].

Epileptogenesis relies heavily on synaptic plasticity. For example, tetanic stimulation of the white matter of the PRC and Schaffer collaterals of the hippocampus suggesting reciprocal long-term potentiation between the hyperexcitable structures pertinent in epilepsy [116]. Short and long-term plasticity also implicate changes at the receptor level and refer to the alteration in both number and composition of certain receptor groups leading to decreased inhibition of some brain structures and increasing their propensity to produce excitatory bursts. An increase in the density of α -1 adrenergic receptors [117], modifications in the composition of NMDA and AMPA glutamate receptors [116], and changes in morphology and number of gluzinergetic (zinc-containing glutaminergic) terminals in PRC and PIRC [118] occur as part of the changes of synaptic plasticity elicited by induction of long term potentiation. These changes are believed to play a role in epilepsy by facilitation of further seizures. Garcia-Cairasco and colleagues used acoustic kindling to observe long-term morphological changes [119]. In the groups that underwent long-term kindling and 60 days of recovery, staining revealed mossy fiber sprouting in the ventral hippocampus and a significant change in the optical density of PRC in susceptible animals as compared to resistant ones suggesting that permanent structural changes are present soon after the initial seizure. Changes on the synaptic and cellular level collectively

create an imbalance between excitation and inhibition in PRC and PIRC which are predisposed to excitation and have the infrastructure for synchronization of neuronal activity that amplifies and propagates excitatory signals throughout the neocortex resulting in TLE with its other comorbidities.

Hippocampal kindling also induces long-term changes in the dynamics within the amygdala-piriform-perirhinal slice. After hippocampal kindling, spontaneous discharges in the slice originated within PIRC, whereas in slices from control animals, discharges originated within PRC. Unlike amygdala kindling, which produced strictly unilateral changes in excitability, kindling of dorsal hippocampus generated bilateral alterations. Even partial hippocampal kindling was linked to presence of spontaneous spikes in the PRC under hyperexcitable conditions [120]. These findings show a change in the region of seizure onset are consistent with *in vivo* observations that early in epilepsy seizures began with focal onset, while later seizures had more diffuse onset (including hippocampus, amygdala, piriform and entorhinal cortices) [121]. This provides evidence for the hypothesis that epileptogenesis is an active process, initiated in hyperexcitable trigger zones that progressively involves more structures with wider network distribution over time and emphasis should be placed on early therapeutic intervention. As discussed in the section on histopathology, this process is accompanied by structural and functional changes in the seizure network, including loss of functional complexity in the interneurons of epilepsy trigger zones [122].

8. PIRC and PRC PHARMACOLOGICAL STUDIES:

NMDA receptor

The role of N-methyl-D-aspartate (NMDA) receptors in the generation and propagation of seizures differs between anterior PIRC (i.e. AT), posterior PIRC, and the PRC. Within the AT,

blockade of NMDA receptors prevents bicuculline-evoked seizures [108,123] and attenuates amygdala-kindled seizures [109]. Similarly, within the PRC, NMDA receptor activation is necessary for seizure propagation. In contrast, blockade of NMDA receptors in posterior PIRC was without effect on AT-evoked seizure propagation [5,108,124]. The pattern differs slightly in the non-human primate: blockade of NMDA receptors within AT using 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzo (f)-quinoxaline (NBQX) or 3-(2-carboxypiperazin-4-yl)propyl-1-phosphonic acid (CPP) does not significantly protect against AT-evoked complex partial seizures [125]. Seizures evoked by soman, a nerve agent inhibitor of acetylcholinesterase, show some resemblance with AT-evoked seizures in terms of the profile of NMDA receptor involvement across regions. For example, NMDA receptor blockade in the posterior PIRC offered no protection against soman-induced seizures, while NMDA receptor blockade in the PRC more than doubled the latency to seizure onset after soman exposure [126]. This pattern differs with respect to NMDA receptor antagonism in AT, which is without effect against soman administration [105]. In the *ex vivo* guinea pig brain, seizure activity evoked with the potassium channel blocker 4-aminopyridine (4-AP) is actually worsened by blockade of NMDA receptors. Finally, in slice models, blockade of NMDA-receptor mediated neurotransmission attenuates ictal and interictal-like activity evoked by 4-AP [127], by high extracellular potassium concentrations [128], and spontaneous and evoked discharges from pilocarpine [129]. Thus, NMDA mediated neurotransmission plays a pivotal role in the propagation of seizure activity within the PIRC/PRC network.

AMPA

Alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) receptor mediated neurotransmission is critical for seizure initiation and propagation within anterior and posterior PIRC along with PRC. For example, systemic administration of the AMPA receptor antagonist, NBQX, produced a dose-dependent decrease in the severity of seizures evoked from AT [124].

Moreover, microinjection of NBQX into either AT, posterior PIRC or PRC antagonized AT-evoked seizures with doses as low as 500 pmol. Unlike the role of NMDA receptors, the role of AMPA receptors in this circuit is conserved between rats and monkeys: AMPA receptor blockade within AT suppressed AT-evoked seizures in the monkey [125]. Blockade of AMPA receptors within PRC is also effective against soman-evoked seizures [126]. In the guinea pig model, blockade of AMPA receptors significantly attenuated seizures triggered by 4-AP within piriform cortex, entorhinal cortex, and hippocampus [130].

AMPA receptor desensitization within AT appears to play a pivotal role in controlling the duration of seizure activity. For example, co-application of cyclothiazide (which blocks AMPA receptor desensitization) and bicuculline triggers a self-sustaining status epilepticus [123]. Moreover, overexpression of slow desensitizing (GluR1 flip), but not fast desensitizing (GluR1 flop) AMPA receptors within AT significantly increased the severity of bicuculline evoked seizures. In all cases, AT-evoked status epilepticus was insensitive to NMDA receptor blockade but was prevented by AMPA blockade [123]. Within the monkey, given the relative propensity of AT-evoked seizures to result in status epilepticus, and the insensitivity of AT-evoked seizures to NMDA receptor activation, it could be speculated that AMPA receptors within PIRC may be slow to desensitize.

At the level of synaptic physiology, AMPA antagonism with DNQX can eliminate epileptiform events in the slice [19]. This effect is associated with decreased buildup in multiunit activity preceding epileptiform discharges. Interestingly, this effect occurs at a concentration below that required to completely block monosynaptic EPSPs suggesting that the disruption in epileptiform activity by AMPA blockade results from disruption of a multisynaptic process. EPSPs mediated by AMPA receptors may be the basis for the epileptiform events in the PIRC. In contrast to NMDA receptors, which contribute but are not indispensable for seizure generation, AMPA receptors appear to be a key component to triggering seizures.

Together, the experimental data discussed above indicate that AMPA-mediated (as compared to NMDA-mediated) neurotransmission plays a relatively larger role in seizure generation within the PIRC/PRC network.

GABA

GABA-mediated (in particular GABA_A) neurotransmission plays a critical role in controlling hyperexcitability of neuronal circuits. Disinhibition of hyperexcitable regions by loss of GABAergic inhibition has been long proposed as a mechanism of seizure propagation [131]. As previously discussed, epileptogenic insults can change the balance between excitation and inhibition within PIRC/PRC in several ways, including by loss of inhibitory neurons. Multiple studies have demonstrated the pivotal role for GABA_A-mediated neurotransmission in PIRC and PRC for seizure control. For example, picomole amounts of GABA antagonist administered within AT or in posterior PIRC can trigger limbic motor seizure in rat [4]. In the *ex vivo* guinea pig brain, microinjection of bicuculline in the anterior PIRC results in interictal spiking and spontaneous epileptiform potentials [132]. Interestingly, in a PIRC slice model of epileptiform activity evoked by 4-AP blockade of GABA_A mediated transmission abolishes ictal-like discharges, but leaves periodic interictal activity intact [133].

Activation of GABA_A receptors by pretreatment of PIRC or PRC with a muscimol protected animals from development of AT-evoked seizures [108]. The effect of muscimol was unique to PIRC and PRC with no seizure prevention observed when it was placed into areas such as amygdala or hippocampus [108]. This suggests that proper functioning of GABA_A receptors is critical for prevention of ictal activity in PIRC and PRC, two areas that are hyperexcitable under normal conditions and serving as epilepsy trigger zones [134]. These data demonstrate that ERC is capable of producing focal epileptiform discharges, but also has the necessary functional connections to facilitate propagation of these discharges to other brain

areas. Febrile seizures upregulate the expression of GABA_A receptors in neonatal granule cells. The increase in GABA_A receptors causes a reversal in the direction of granule cell migration. Furthermore, preventing GABA-mediated action by using bumetanide, which inhibits that Na⁺-K⁺-2-Cl⁻ co-transporter, susceptibility to limbic seizures is reduced [135,136].

ERC function is significantly altered by epileptogenic insults. For example, unilateral damage to ERC caused by amino-oxyacetic acid (AOAA) produced long-lasting (1 mo to 1.5 yr) changes in extracellular responses to white matter stimulation in the superficial layers of the medial ERC. Extracellular responses in slices from AOAA treated animals were prolonged and repetitive and intracellular recordings from residual principal cells revealed prolonged, repetitive excitatory postsynaptic potentials (EPSPs). This pattern differed significantly from the brief EPSPs and single discharges seen in slices from controls. In neurons located in the deep layers of ERC, the initial synaptic response to white matter stimulation did not differ between AOAA and control treated animals. However, in a subset of deep neurons in AOAA treated animals, repetitive action potentials were seen after a brief delay [137]. All together, these changes are consistent with those needed to facilitate the generation of sustained synchronous activity.

9. ERC PHARMACOLOGICAL STUDIES:

As in PIRC/PRC, glutamatergic and GABAergic neurotransmission within ERC are essential for seizure propagation. However, relatively fewer studies have directly manipulated ERC neurotransmitter signaling *in vivo*. In the sections below, we will outline the *in vivo* and *in vitro* pharmacological studies that have established ERC as a key site for seizure origin and propagation.

Glutamatergic transmission

The role of glutamatergic neurotransmission within ERC is very similar to its role in the PIRC. Not only glutamate antagonists have a differential effect on epileptiform activity depending on the type of receptor blocked, but the effect of the antagonists also differ based on the type of discharges. Glutamatergic transmission has a function in both seizure generation in the deep layers and seizure propagation in the superficial layers of the ERC. Moreover, the process of kindling induces long-lasting changes in glutamatergic neurotransmission within ERC [138–141], along with other limbic regions.

AMPA

In the slice, the role of AMPA-receptor mediated neurotransmission is consistent across ages and methods of inducing epileptiform activity. AMPA antagonists suppress seizure-like events caused by increases in extracellular potassium, bath application of 4-AP, decrease in extracellular magnesium concentration [142,143] and pilocarpine application [144]. Moreover, AMPA antagonists also attenuate a subset of interictal discharges [145]. *Ex vivo*, in the isolated guinea pig brain, AMPA receptor blockade completely suppresses seizure activity evoked either electrically from lateral ERC [72] or by 4-AP perfusion [130].

While the effects of *in vivo* manipulation of glutamatergic neurotransmission within ERC on seizure propagation remain understudied, the compelling slice and *ex vivo* data above suggest an instrumental role for AMPA-receptor mediated glutamatergic signaling in seizure generation within ERC.

NMDA

In contrast to AMPA-receptor mediated transmission, NMDA receptor mediated signaling does not appear to play as consistently vital a role in seizure propagation within ERC. NMDA

receptor antagonists may have no effect in slice models [142,144,146,147], and in the *ex vivo* guinea pig model [72]. However, other authors have found that NMDA antagonists decrease the probability and speed of propagation, but do not abolish the propagation completely [148].

Some studies have suggested that NMDA-receptor-mediated transmission plays a particularly important role in the genesis of late epileptiform events, as compared to early events [149]. In slices taken from rats that had received status epilepticus, blockade of NMDA receptors partially normalized neuronal activity [150,151]. As with AMPA-mediated transmission, there is a dearth of data regarding effects of focal entorhinal pharmacological manipulations on seizure propagation. In conclusion, glutamatergic transmission in particular mediated by AMPA receptors is vital for seizure genesis within ERC and propagation both within ERC and to other structures.

GABA

A failure of the inhibitory control mechanisms that usually dominate interactions between PRC and ERC may underlie seizure initiation and propagation. In the *ex vivo* guinea pig brain, disinhibition by application of bicuculline with concentrations as low as 20 μ M resulted in spontaneous seizures that spread to the amygdala more rapidly than electrically evoked seizures [72]. Moreover, when compared to electrically-evoked seizures, the distribution area of bicuculline-induced seizures was greater, involving the posterior PIRC and larger areas over the medial amygdala. Both the speed of propagation and the extent of seizures were proportional to the concentration of bicuculline. In this preparation, interictal activity is first detected in ERC, before propagating to other regions and progressing to fast ictal discharges [59]. Interestingly, during the onset of seizure activity in this model, principal (excitatory) neurons are silent – ictal activity is generated by hypersynchronous and sustained firing of interneurons [152].

In addition to effects on generation of ictal discharges, blockade of GABA-mediated transmission in the ERC stimulates generation of interictal-like discharges, which consisted of an initial paroxysmal depolarizing shift followed by afterdischarges. The initial component of the interictal discharges, namely, the paroxysmal depolarizing shifts can be blocked in a progressive fashion, while the second component, the afterdischarges, are abolished strictly in all-or-none manner [153]. Interictal-like discharges represent an intrinsic electrophysiological property of the ERC and support the hypothesis that this region is highly epileptogenic. Similarly, spontaneous sharp waves in the temporal cortex were abolished by administration of GABA antagonists bicuculline [154] or μ -receptor agonists [155], while removal of antagonists restored paroxysmal depolarizing activity [154], effectively suggesting that the synchronization currents were mediated solely by GABA transmission.

The epileptogenicity is associated with changes on the molecular, cellular, and system levels that facilitate generation and propagation of seizures. The implication of these changes can be measured in part as a change in the latency to seizure generation under stimulating conditions. The latency to occurrence of the first seizure-like events after brain slices from SE-subjected animals were placed in Mg^{2+} -free artificial cerebrospinal fluid was significantly reduced at 4 and 8 weeks. This provides an evidence for escalation of brain excitability and neuronal hypersynchrony due to adaptations in brain plasticity, predisposing to recurrent epileptic seizures [156]. Exposure to a convulsant such as 4-AP produces lasting changes in the ERC suggesting plasticity that renders the area epileptogenic. Moreover, different network interactions within the hippocampus-ERC loop characterize control and epileptic slices in the pilocarpine model of epilepsy. For example, ERC-driven ictal discharges in pilocarpine-treated slices occur throughout the experiment and spread to the CA1/subicular area via the temporoammonic path [157]. These functional changes caused by seizure-induced cell damage lead to the development of epileptic seizures. The process is facilitated by a disinhibition of the ERC due to loss of feed-forward inhibition from the PRC and hippocampal outputs and possibly

sustained by the reverberant activity between ERC and CA1 networks that are excited via the temporoammonic path [157].

In conclusion, pharmacological means have demonstrated that ERC is an area that is highly susceptible to ictal discharges when exposed to low doses of excitatory compounds.

10. CONCLUSION:

In this review, we have described the functional neuroanatomy, pharmacology, and physiology of three limbic-cortical areas: ERC, PRC, and PIRC. These regions are highly interconnected with each other and with other limbic structures, and have features that make them particularly prone to the genesis and propagation of ictal discharges. While a great deal of attention has been paid to subcortical limbic structures in epilepsy (e.g., amygdala, hippocampus), the data reviewed herein clearly underscore the importance of these cortical regions for epileptogenesis.

Figure 1. Overview of the *functional connectivity of piriform, perirhinal, and entorhinal cortices.*

Many studies have investigated the connectivity of the piriform (PIRC), perirhinal (PRC), and entorhinal cortices (ERC) along with their relationship with other cortical and limbic structures. These regions are highly interconnected with each other and mediate bidirectional information from higher processing regions. Abnormal activity from these regions is hypothesized to cause hyperexcitability and excessive propagation associated with temporal lobe epilepsy along with perceptual disturbances such as hallucinations and auras.

DEDICATION:

We dedicate this article in memory of Dr. Karen Gale, Ph.D. who passed away on August 21, 2014. In addition to her extraordinary contributions to neuroscience, she was most notably the first to discover the region of the brain that triggers epileptic seizures called the Area Tempestas. A remarkable teacher, mentor, and advocate, Dr. Gale helped develop the Interdisciplinary Program in Neuroscience and Georgetown Women in Medicine, both at Georgetown University. Her passion and commitment to the scientific community at all levels will be remembered.

REFERENCES:

1. Neville KR, Haberly LB. Beta and gamma oscillations in the olfactory system of the urethane-anesthetized rat. *J Neurophysiol.* 2003;90: 3921–3930. doi:10.1152/jn.00475.2003
2. Cataldi M, Avoli M, de Villers-Sidani E. Resting state networks in temporal lobe epilepsy. *Epilepsia.* 2013;54: 2048–2059. doi:10.1111/epi.12400
3. Howard JD, Plailly J, Grueschow M, Haynes J-D, Gottfried JA. Odor quality coding and categorization in human posterior piriform cortex. *Nat Neurosci.* 2009;12: 932–938. doi:10.1038/nn.2324
4. Piredda S, Gale K. A crucial epileptogenic site in the deep prepiriform cortex. *Nature.* 1985;317: 623–625.
5. Piredda S, Gale K. Role of excitatory amino acid transmission in the genesis of seizures elicited from the deep prepiriform cortex. *Brain Res.* 1986;377: 205–210.
6. Demir R, Haberly LB, Jackson MB. Voltage imaging of epileptiform activity in slices from rat piriform cortex: onset and propagation. *J Neurophysiol.* 1998;80: 2727–2742.
7. Demir R, Haberly LB, Jackson MB. Sustained and accelerating activity at two discrete sites generate epileptiform discharges in slices of piriform cortex. *J Neurosci Off J Soc Neurosci.* 1999;19: 1294–1306.
8. Behan M, Haberly LB. Intrinsic and efferent connections of the endopiriform nucleus in rat. *J Comp Neurol.* 1999;408: 532–548.
9. Demir R, Haberly LB, Jackson MB. Epileptiform discharges with in-vivo-like features in slices of rat piriform cortex with longitudinal association fibers. *J Neurophysiol.* 2001;86: 2445–2460.
10. Laufs H, Richardson MP, Salek-Haddadi A, Vollmar C, Duncan JS, Gale K, et al. Converging PET and fMRI evidence for a common area involved in human focal epilepsies. *Neurology.* 2011;77: 904–910. doi:10.1212/WNL.0b013e31822c90f2
11. Mraovitch S. Isocortical hyperemia and allocortical inflammation and atrophy following generalized convulsive seizures of thalamic origin in the rat. *Cell Mol Neurobiol.* 2003;23: 773–791.
12. Vaughan DN, Jackson GD. The piriform cortex and human focal epilepsy. *Front Neurol.* 2014;5: 259. doi:10.3389/fneur.2014.00259

13. Sugai T, Yamamoto R, Yoshimura H, Kato N. Multimodal cross-talk of olfactory and gustatory information in the endopiriform nucleus in rats. *Chem Senses*. 2012;37: 681–688. doi:10.1093/chemse/bjs046
14. Sperber EF, Velísková J, Benenati B, Moshé SL. Enhanced epileptogenicity of area tempestas in the immature rat. *Dev Neurosci*. 1998;20: 540–545.
15. Tseng GF, Haberly LB. Deep neurons in piriform cortex. I. Morphology and synaptically evoked responses including a unique high-amplitude paired shock facilitation. *J Neurophysiol*. 1989;62: 369–385.
16. Krettek JE, Price JL. Projections from the amygdaloid complex and adjacent olfactory structures to the entorhinal cortex and to the subiculum in the rat and cat. *J Comp Neurol*. 1977;172: 723–752. doi:10.1002/cne.901720409
17. Ekstrand JJ, Domroese ME, Johnson DM, Feig SL, Knodel SM, Behan M, et al. A new subdivision of anterior piriform cortex and associated deep nucleus with novel features of interest for olfaction and epilepsy. *J Comp Neurol*. 2001;434: 289–307.
18. Hoffman WH, Haberly LB. Bursting-induced epileptiform EPSPs in slices of piriform cortex are generated by deep cells. *J Neurosci Off J Soc Neurosci*. 1991;11: 2021–2031.
19. Hoffman WH, Haberly LB. Kindling-induced epileptiform potentials in piriform cortex slices originate in the underlying endopiriform nucleus. *J Neurophysiol*. 1996;76: 1430–1438.
20. Scholl EA, Dudek FE, Ekstrand JJ. Neuronal degeneration is observed in multiple regions outside the hippocampus after lithium pilocarpine-induced status epilepticus in the immature rat. *Neuroscience*. 2013;252: 45–59. doi:10.1016/j.neuroscience.2013.07.045
21. Löscher W, Ebert U. The role of the piriform cortex in kindling. *Prog Neurobiol*. 1996;50: 427–481.
22. Burwell RD, Amaral DG. Cortical afferents of the perirhinal, postrhinal, and entorhinal cortices of the rat. *J Comp Neurol*. 1998;398: 179–205.
23. Luskin MB, Price JL. The laminar distribution of intracortical fibers originating in the olfactory cortex of the rat. *J Comp Neurol*. 1983;216: 292–302. doi:10.1002/cne.902160306
24. Agster KL, Burwell RD. Cortical efferents of the perirhinal, postrhinal, and entorhinal cortices of the rat. *Hippocampus*. 2009;19: 1159–1186. doi:10.1002/hipo.20578
25. Veening JG. Cortical afferents of the amygdaloid complex in the rat: an HRP study. *Neurosci Lett*. 1978;8: 191–195.

26. Wakefield C. The topographical organization and laminar origin of some cortico-amygdaloid connections. *Neurosci Lett*. 1980;20: 21–24.
27. Haberly LB. Parallel-distributed processing in olfactory cortex: new insights from morphological and physiological analysis of neuronal circuitry. *Chem Senses*. 2001;26: 551–576.
28. Schwabe K, Ebert U, Löscher W. The central piriform cortex: anatomical connections and anticonvulsant effect of GABA elevation in the kindling model. *Neuroscience*. 2004;126: 727–741. doi:10.1016/j.neuroscience.2004.04.022
29. Illig KR. Projections from orbitofrontal cortex to anterior piriform cortex in the rat suggest a role in olfactory information processing. *J Comp Neurol*. 2005;488: 224–231. doi:10.1002/cne.20595
30. Johnson DM, Illig KR, Behan M, Haberly LB. New features of connectivity in piriform cortex visualized by intracellular injection of pyramidal cells suggest that “primary” olfactory cortex functions like “association” cortex in other sensory systems. *J Neurosci Off J Soc Neurosci*. 2000;20: 6974–6982.
31. Luskin MB, Price JL. The topographic organization of associational fibers of the olfactory system in the rat, including centrifugal fibers to the olfactory bulb. *J Comp Neurol*. 1983;216: 264–291. doi:10.1002/cne.902160305
32. Rosen JB, Hitchcock JM, Miserendino MJ, Falls WA, Campeau S, Davis M. Lesions of the perirhinal cortex but not of the frontal, medial prefrontal, visual, or insular cortex block fear-potentiated startle using a visual conditioned stimulus. *J Neurosci Off J Soc Neurosci*. 1992;12: 4624–4633.
33. Zola-Morgan S, Squire LR, Ramus SJ. Severity of memory impairment in monkeys as a function of locus and extent of damage within the medial temporal lobe memory system. *Hippocampus*. 1994;4: 483–495. doi:10.1002/hipo.450040410
34. Wiig KA, Cooper LN, Bear MF. Temporally graded retrograde amnesia following separate and combined lesions of the perirhinal cortex and fornix in the rat. *Learn Mem Cold Spring Harb N*. 1996;3: 313–325.
35. Murray EA, Richmond BJ. Role of perirhinal cortex in object perception, memory, and associations. *Curr Opin Neurobiol*. 2001;11: 188–193.
36. Ryan L, Cardoza JA, Barense MD, Kawa KH, Wallentin-Flores J, Arnold WT, et al. Age-related impairment in a complex object discrimination task that engages perirhinal cortex. *Hippocampus*. 2012;22: 1978–1989. doi:10.1002/hipo.22069

37. Meunier M, Bachevalier J, Mishkin M, Murray EA. Effects on visual recognition of combined and separate ablations of the entorhinal and perirhinal cortex in rhesus monkeys. *J Neurosci Off J Soc Neurosci*. 1993;13: 5418–5432.
38. Zola-Morgan S, Squire LR, Ramus SJ. Severity of memory impairment in monkeys as a function of locus and extent of damage within the medial temporal lobe memory system. *Hippocampus*. 1994;4: 483–495. doi:10.1002/hipo.450040410
39. Bucci DJ, Burwell RD. Deficits in attentional orienting following damage to the perirhinal or postrhinal cortices. *Behav Neurosci*. 2004;118: 1117–1122. doi:10.1037/0735-7044.118.5.1117
40. Barbeau E, Wendling F, Régis J, Duncan R, Poncet M, Chauvel P, et al. Recollection of vivid memories after perirhinal region stimulations: synchronization in the theta range of spatially distributed brain areas. *Neuropsychologia*. 2005;43: 1329–1337. doi:10.1016/j.neuropsychologia.2004.11.025
41. Martin CB, Mirsattari SM, Pruessner JC, Pietrantonio S, Burneo JG, Hayman-Abello B, et al. Déjà vu in unilateral temporal-lobe epilepsy is associated with selective familiarity impairments on experimental tasks of recognition memory. *Neuropsychologia*. 2012;50: 2981–2991. doi:10.1016/j.neuropsychologia.2012.07.030
42. Shatskikh TN, Raghavendra M, Zhao Q, Cui Z, Holmes GL. Electrical induction of spikes in the hippocampus impairs recognition capacity and spatial memory in rats. *Epilepsy Behav EB*. 2006;9: 549–556. doi:10.1016/j.yebeh.2006.08.014
43. Insausti R, Amaral DG, Cowan WM. The entorhinal cortex of the monkey: II. Cortical afferents. *J Comp Neurol*. 1987;264: 356–395. doi:10.1002/cne.902640306
44. Saleem KS, Price JL, Hashikawa T. Cytoarchitectonic and chemoarchitectonic subdivisions of the perirhinal and parahippocampal cortices in macaque monkeys. *J Comp Neurol*. 2007;500: 973–1006. doi:10.1002/cne.21141
45. De Curtis M, Paré D. The rhinal cortices: a wall of inhibition between the neocortex and the hippocampus. *Prog Neurobiol*. 2004;74: 101–110. doi:10.1016/j.pneurobio.2004.08.005
46. Pelletier JG, Apergis J, Paré D. Low-probability transmission of neocortical and entorhinal impulses through the perirhinal cortex. *J Neurophysiol*. 2004;91: 2079–2089. doi:10.1152/jn.01197.2003
47. Killian NJ, Jutras MJ, Buffalo EA. A map of visual space in the primate entorhinal cortex. *Nature*. 2012;491: 761–764. doi:10.1038/nature11587

48. Lu L, Leutgeb JK, Tsao A, Henriksen EJ, Leutgeb S, Barnes CA, et al. Impaired hippocampal rate coding after lesions of the lateral entorhinal cortex. *Nat Neurosci.* 2013;16: 1085–1093. doi:10.1038/nn.3462
49. Courtney SM, Ungerleider LG, Keil K, Haxby JV. Object and spatial visual working memory activate separate neural systems in human cortex. *Cereb Cortex N Y N 1991.* 1996;6: 39–49.
50. Aguirre GK, Detre JA, Alsop DC, D’Esposito M. The parahippocampus subserves topographical learning in man. *Cereb Cortex N Y N 1991.* 1996;6: 823–829.
51. Rolls ET, O’Mara SM. View-responsive neurons in the primate hippocampal complex. *Hippocampus.* 1995;5: 409–424. doi:10.1002/hipo.450050504
52. Navratilova Z, Giocomo LM, Fellous J-M, Hasselmo ME, McNaughton BL. Phase precession and variable spatial scaling in a periodic attractor map model of medial entorhinal grid cells with realistic after-spike dynamics. *Hippocampus.* 2012;22: 772–789. doi:10.1002/hipo.20939
53. Sargolini F, Fyhn M, Hafting T, McNaughton BL, Witter MP, Moser M-B, et al. Conjunctive representation of position, direction, and velocity in entorhinal cortex. *Science.* 2006;312: 758–762. doi:10.1126/science.1125572
54. Leichnetz GR, Astruc J. The squirrel monkey entorhinal cortex: architecture and medial frontal afferents. *Brain Res Bull.* 1976;1: 351–358.
55. Amaral DG, Insausti R, Cowan WM. The entorhinal cortex of the monkey: I. Cytoarchitectonic organization. *J Comp Neurol.* 1987;264: 326–355. doi:10.1002/cne.902640305
56. Insausti R, Tuñón T, Sobreviela T, Insausti AM, Gonzalo LM. The human entorhinal cortex: a cytoarchitectonic analysis. *J Comp Neurol.* 1995;355: 171–198. doi:10.1002/cne.903550203
57. Insausti R. Comparative anatomy of the entorhinal cortex and hippocampus in mammals. *Hippocampus.* 1993;3 Spec No: 19–26.
58. Dolorfo CL, Amaral DG. Entorhinal cortex of the rat: organization of intrinsic connections. *J Comp Neurol.* 1998;398: 49–82.
59. Uva L, Gruschke S, Biella G, De Curtis M, Witter MP. Cytoarchitectonic characterization of the parahippocampal region of the guinea pig. *J Comp Neurol.* 2004;474: 289–303. doi:10.1002/cne.20121

60. Biella G, Uva L, de Curtis M. Propagation of neuronal activity along the neocortical-perirhinal-entorhinal pathway in the guinea pig. *J Neurosci Off J Soc Neurosci*. 2002;22: 9972–9979.
61. Biella G, Uva L, Hofmann UG, de Curtis M. Associative interactions within the superficial layers of the entorhinal cortex of the guinea pig. *J Neurophysiol*. 2002;88: 1159–1165.
62. Kibler AB, Durand DM. Orthogonal wave propagation of epileptiform activity in the planar mouse hippocampus in vitro. *Epilepsia*. 2011;52: 1590–1600. doi:10.1111/j.1528-1167.2011.03125.x
63. Andersen P, Bliss TV, Skrede KK. Lamellar organization of hippocampal pathways. *Exp Brain Res*. 1971;13: 222–238.
64. Acsády L, Káli S. Models, structure, function: the transformation of cortical signals in the dentate gyrus. *Prog Brain Res*. 2007;163: 577–599. doi:10.1016/S0079-6123(07)63031-3
65. Engel J Jr, McDermott MP, Wiebe S, Langfitt JT, Stern JM, Dewar S, et al. Early surgical therapy for drug-resistant temporal lobe epilepsy: a randomized trial. *JAMA J Am Med Assoc*. 2012;307: 922–930. doi:10.1001/jama.2012.220
66. Engel J Jr, Wiebe S, French J, Sperling M, Williamson P, Spencer D, et al. Practice parameter: temporal lobe and localized neocortical resections for epilepsy: report of the Quality Standards Subcommittee of the American Academy of Neurology, in association with the American Epilepsy Society and the American Association of Neurological Surgeons. *Neurology*. 2003;60: 538–547.
67. Umeoka SC, Lüders HO, Turnbull JP, Koubeissi MZ, Maciunas RJ. Requirement of longitudinal synchrony of epileptiform discharges in the hippocampus for seizure generation: a pilot study. *J Neurosurg*. 2012;116: 513–524. doi:10.3171/2011.10.JNS11261
68. Bartolomei F, Wendling F, Bellanger JJ, Régis J, Chauvel P. Neural networks involving the medial temporal structures in temporal lobe epilepsy. *Clin Neurophysiol Off J Int Fed Clin Neurophysiol*. 2001;112: 1746–1760.
69. Pallud J, Devaux B, Depaulis A. [Changes in spontaneous epileptic activity after selective intrahippocampal transection in a model of chronic mesial temporal lobe epilepsy]. *Neurochirurgie*. 2008;54: 135–140. doi:10.1016/j.neuchi.2008.02.002
70. Finch DM, Tan AM, Isokawa-Akesson M. Feedforward inhibition of the rat entorhinal cortex and subicular complex. *J Neurosci Off J Soc Neurosci*. 1988;8: 2213–2226.
71. Kelly ME, McIntyre DC. Perirhinal cortex involvement in limbic kindled seizures. *Epilepsy Res*. 1996;26: 233–243.

72. Federico P, MacVicar BA. Imaging the induction and spread of seizure activity in the isolated brain of the guinea pig: the roles of GABA and glutamate receptors. *J Neurophysiol.* 1996;76: 3471–3492.
73. Haberly LB, Bower JM. Olfactory cortex: model circuit for study of associative memory? *Trends Neurosci.* 1989;12: 258–264.
74. Wilson DA, Stevenson RJ. Olfactory perceptual learning: the critical role of memory in odor discrimination. *Neurosci Biobehav Rev.* 2003;27: 307–328.
75. McIntyre DC, Gilby KL. Parahippocampal networks, intractability, and the chronic epilepsy of kindling. *Adv Neurol.* 2006;97: 77–83.
76. De Guzman P, D’Antuono M, Avoli M. Initiation of electrographic seizures by neuronal networks in entorhinal and perirhinal cortices in vitro. *Neuroscience.* 2004;123: 875–886.
77. Jutila L, Ylinen A, Partanen K, Alafuzoff I, Mervaala E, Partanen J, et al. MR volumetry of the entorhinal, perirhinal, and temporopolar cortices in drug-refractory temporal lobe epilepsy. *AJNR Am J Neuroradiol.* 2001;22: 1490–1501.
78. Lencz T, McCarthy G, Bronen RA, Scott TM, Inserni JA, Sass KJ, et al. Quantitative magnetic resonance imaging in temporal lobe epilepsy: relationship to neuropathology and neuropsychological function. *Ann Neurol.* 1992;31: 629–637.
doi:10.1002/ana.410310610
79. Bonilha L, Kobayashi E, Rorden C, Cendes F, Li LM. Medial temporal lobe atrophy in patients with refractory temporal lobe epilepsy. *J Neurol Neurosurg Psychiatry.* 2003;74: 1627–1630.
80. Lee JW, Andermann F, Dubeau F, Bernasconi A, MacDonald D, Evans A, et al. Morphometric analysis of the temporal lobe in temporal lobe epilepsy. *Epilepsia.* 1998;39: 727–736.
81. Bartolomei F, Khalil M, Wendling F, Sontheimer A, Régis J, Ranjeva J-P, et al. Entorhinal cortex involvement in human mesial temporal lobe epilepsy: an electrophysiologic and volumetric study. *Epilepsia.* 2005;46: 677–687. doi:10.1111/j.1528-1167.2005.43804.x
82. Du F, Whetsell WO Jr, Abou-Khalil B, Blumenkopf B, Lothman EW, Schwarcz R. Preferential neuronal loss in layer III of the entorhinal cortex in patients with temporal lobe epilepsy. *Epilepsy Res.* 1993;16: 223–233.
83. Jung K-H, Chu K, Lee S-T, Kim J-H, Kang K-M, Song E-C, et al. Region-specific plasticity in the epileptic rat brain: a hippocampal and extrahippocampal analysis. *Epilepsia.* 2009;50: 537–549. doi:10.1111/j.1528-1167.2008.01718.x

84. Drexel M, Preidt AP, Sperk G. Sequel of spontaneous seizures after kainic acid-induced status epilepticus and associated neuropathological changes in the subiculum and entorhinal cortex. *Neuropharmacology*. 2012;63: 806–817. doi:10.1016/j.neuropharm.2012.06.009
85. Langer M, Brandt C, Zellinger C, Löscher W. Therapeutic window of opportunity for the neuroprotective effect of valproate versus the competitive AMPA receptor antagonist NS1209 following status epilepticus in rats. *Neuropharmacology*. 2011;61: 1033–1047. doi:10.1016/j.neuropharm.2011.06.015
86. Empson RM, Heinemann U. The perforant path projection to hippocampal area CA1 in the rat hippocampal-entorhinal cortex combined slice. *J Physiol*. 1995;484 (Pt 3): 707–720.
87. Du F, Eid T, Lothman EW, Köhler C, Schwarcz R. Preferential neuronal loss in layer III of the medial entorhinal cortex in rat models of temporal lobe epilepsy. *J Neurosci Off J Soc Neurosci*. 1995;15: 6301–6313.
88. Kumar SS, Buckmaster PS. Hyperexcitability, interneurons, and loss of GABAergic synapses in entorhinal cortex in a model of temporal lobe epilepsy. *J Neurosci Off J Soc Neurosci*. 2006;26: 4613–4623. doi:10.1523/JNEUROSCI.0064-06.2006
89. Couey JJ, Witoelar A, Zhang S-J, Zheng K, Ye J, Dunn B, et al. Recurrent inhibitory circuitry as a mechanism for grid formation. *Nat Neurosci*. 2013;16: 318–324. doi:10.1038/nn.3310
90. Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH. Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci Off J Soc Neurosci*. 1997;17: 3727–3738.
91. Parent JM, Valentin VV, Lowenstein DH. Prolonged seizures increase proliferating neuroblasts in the adult rat subventricular zone-olfactory bulb pathway. *J Neurosci Off J Soc Neurosci*. 2002;22: 3174–3188. doi:20026296
92. Darlington GJ, Wilson DR, Lachman LB. Monocyte-conditioned medium, interleukin-1, and tumor necrosis factor stimulate the acute phase response in human hepatoma cells in vitro. *J Cell Biol*. 1986;103: 787–793.
93. Hartung HP, Schäfer B, Heininger K, Toyka KV. Recombinant interleukin-1 beta stimulates eicosanoid production in rat primary culture astrocytes. *Brain Res*. 1989;489: 113–119.
94. Vezzani A, French J, Bartfai T, Baram TZ. The role of inflammation in epilepsy. *Nat Rev Neurol*. 2011;7: 31–40. doi:10.1038/nrneurol.2010.178

95. Dolga AM, Granic I, Blank T, Knaus H-G, Spiess J, Luiten PGM, et al. TNF-alpha-mediates neuroprotection against glutamate-induced excitotoxicity via NF-kappaB-dependent up-regulation of K2.2 channels. *J Neurochem*. 2008;107: 1158–1167. doi:10.1111/j.1471-4159.2008.05701.x
96. Jamali S, Bartolomei F, Robaglia-Schlupp A, Massacrier A, Peragut J-C, Régis J, et al. Large-scale expression study of human mesial temporal lobe epilepsy: evidence for dysregulation of the neurotransmission and complement systems in the entorhinal cortex. *Brain J Neurol*. 2006;129: 625–641. doi:10.1093/brain/awl001
97. Drexel M, Preidt AP, Sperk G. Sequel of spontaneous seizures after kainic acid-induced status epilepticus and associated neuropathological changes in the subiculum and entorhinal cortex. *Neuropharmacology*. 2012;63: 806–817. doi:10.1016/j.neuropharm.2012.06.009
98. Mikkonen M, Soininen H, Kälviänen R, Tapiola T, Ylinen A, Vapalahti M, et al. Remodeling of neuronal circuitries in human temporal lobe epilepsy: increased expression of highly polysialylated neural cell adhesion molecule in the hippocampus and the entorhinal cortex. *Ann Neurol*. 1998;44: 923–934. doi:10.1002/ana.410440611
99. Rossi AR, Angelo MF, Villarreal A, Lukin J, Ramos AJ. Gabapentin administration reduces reactive gliosis and neurodegeneration after pilocarpine-induced status epilepticus. *PLoS One*. 2013;8: e78516. doi:10.1371/journal.pone.0078516
100. Pekcec A, Fuest C, Mühlenhoff M, Gerardy-Schahn R, Potschka H. Targeting epileptogenesis-associated induction of neurogenesis by enzymatic depolysialylation of NCAM counteracts spatial learning dysfunction but fails to impact epilepsy development. *J Neurochem*. 2008;105: 389–400. doi:10.1111/j.1471-4159.2007.05172.x
101. Kato N, Higuchi T, Friesen HG, Wada JA. Changes of immunoreactive somatostatin and beta-endorphin content in rat brain after amygdaloid kindling. *Life Sci*. 1983;32: 2415–2422.
102. Vezzani A, Monhemius R, Tutka P, Milani R, Samanin R. Functional activation of somatostatin- and neuropeptide Y-containing neurons in the entorhinal cortex of chronically epileptic rats. *Neuroscience*. 1996;75: 551–557.
103. Kang TC, Park SK, Do SG, Suh JG, Jo SM, Oh YS, et al. The over-expression of somatostatin in the gerbil entorhinal cortex induced by seizure. *Brain Res*. 2000;882: 55–61.
104. Myhrer T, Nguyen NHT, Enger S, Aas P. Anticonvulsant effects of GABA(A) modulators microinfused into area tempestas or substantia nigra in rats exposed to soman. *Arch Toxicol*. 2006;80: 502–507. doi:10.1007/s00204-006-0070-8

105. Myhrer T, Enger S, Aas P. Anticonvulsant efficacy of drugs with cholinergic and/or glutamatergic antagonism microinfused into area tempestas of rats exposed to soman. *Neurochem Res.* 2008;33: 348–354. doi:10.1007/s11064-007-9429-3
106. Hoffman WH, Haberly LB. Role of synaptic excitation in the generation of bursting-induced epileptiform potentials in the endopiriform nucleus and piriform cortex. *J Neurophysiol.* 1993;70: 2550–2561.
107. Doherty J, Gale K, Eagles DA. Evoked epileptiform discharges in the rat anterior piriform cortex: generation and local propagation. *Brain Res.* 2000;861: 77–87.
108. Halonen T, Tortorella A, Zrebeet H, Gale K. Posterior piriform and perirhinal cortex relay seizures evoked from the area tempestas: role of excitatory and inhibitory amino acid receptors. *Brain Res.* 1994;652: 145–148.
109. Holmes KH, Bilkey DK, Laverty R. The infusion of an NMDA antagonist into perirhinal cortex suppresses amygdala-kindled seizures. *Brain Res.* 1992;587: 285–290.
110. McIntyre DC, Plant JR. Long-lasting changes in the origin of spontaneous discharges from amygdala-kindled rats: piriform vs. perirhinal cortex in vitro. *Brain Res.* 1993;624: 268–276.
111. McIntyre DC, Kelly ME, Dufresne C. FAST and SLOW amygdala kindling rat strains: comparison of amygdala, hippocampal, piriform and perirhinal cortex kindling. *Epilepsy Res.* 1999;35: 197–209.
112. Schwabe K, Ebert U, Löscher W. Bilateral lesions of the central but not anterior or posterior parts of the piriform cortex retard amygdala kindling in rats. *Neuroscience.* 2000;101: 513–521.
113. Wahnschaffe U, Ebert U, Löscher W. The effects of lesions of the posterior piriform cortex on amygdala kindling in the rat. *Brain Res.* 1993;615: 295–303.
114. Yang L-X, Jin C-L, Zhu-Ge Z-B, Wang S, Wei E-Q, Bruce IC, et al. Unilateral low-frequency stimulation of central piriform cortex delays seizure development induced by amygdaloid kindling in rats. *Neuroscience.* 2006;138: 1089–1096. doi:10.1016/j.neuroscience.2005.12.006
115. Kairiss EW, Racine RJ, Smith GK. The development of the interictal spike during kindling in the rat. *Brain Res.* 1984;322: 101–110.
116. Supcun B, Ghadiri MK, Zeraati M, Stummer W, Speckmann E-J, Gorji A. The effects of tetanic stimulation on plasticity of remote synapses in the hippocampus-perirhinal cortex-amygdala network. *Synap N Y N.* 2012;66: 965–974. doi:10.1002/syn.21591

117. Gundlach AL, Burazin TC, Jenkins TA, Berkovic SF. Spatiotemporal alterations of central alpha 1-adrenergic receptor binding sites following amygdaloid kindling seizures in the rat: autoradiographic studies using [3H]prazosin. *Brain Res.* 1995;672: 214–227.
118. Galvis-Alonso OY, Cortes De Oliveira JA, Garcia-Cairasco N. Limbic epileptogenicity, cell loss and axonal reorganization induced by audiogenic and amygdala kindling in wistar audiogenic rats (WAR strain). *Neuroscience.* 2004;125: 787–802.
doi:10.1016/j.neuroscience.2004.01.042
119. Garcia-Cairasco N, Wakamatsu H, Oliveira JA, Gomes EL, Del Bel EA, Mello LE. Neuroethological and morphological (Neo-Timm staining) correlates of limbic recruitment during the development of audiogenic kindling in seizure susceptible Wistar rats. *Epilepsy Res.* 1996;26: 177–192.
120. McIntyre DC, Plant JR, Kelly ME. Dorsal hippocampal kindling produces long-lasting changes in the origin of spontaneous discharges in the piriform versus perirhinal cortex in vitro. *Epilepsy Res.* 2000;39: 191–200.
121. Bertram EH. Functional anatomy of spontaneous seizures in a rat model of limbic epilepsy. *Epilepsia.* 1997;38: 95–105.
122. Gavrilovici C, Pollock E, Everest M, Poulter MO. The loss of interneuron functional diversity in the piriform cortex after induction of experimental epilepsy. *Neurobiol Dis.* 2012;48: 317–328. doi:10.1016/j.nbd.2012.07.002
123. Fornai F, Busceti CL, Kondratyev A, Gale K. AMPA receptor desensitization as a determinant of vulnerability to focally evoked status epilepticus. *Eur J Neurosci.* 2005;21: 455–463. doi:10.1111/j.1460-9568.2005.03873.x
124. Tortorella A, Halonen T, Sahibzada N, Gale K. A crucial role of the alpha-amino-3-hydroxy-5-methylisoxazole-4-propionic acid subtype of glutamate receptors in piriform and perirhinal cortex for the initiation and propagation of limbic motor seizures. *J Pharmacol Exp Ther.* 1997;280: 1401–1405.
125. Malkova L, Forcelli PA, Wellman LL, Dybdal D, Dubach MF, Gale K. Blockade of glutamatergic transmission in perirhinal cortex impairs object recognition memory in macaques. *J Neurosci Off J Soc Neurosci.* 2015;35: 5043–5050.
doi:10.1523/JNEUROSCI.4307-14.2015
126. Myhrer T, Enger S, Aas P. Roles of perirhinal and posterior piriform cortices in control and generation of seizures: a microinfusion study in rats exposed to soman. *Neurotoxicology.* 2010;31: 147–153. doi:10.1016/j.neuro.2009.10.002
127. Salah A, Perkins KL. Persistent ictal-like activity in rat entorhinal/perirhinal cortex following washout of 4-aminopyridine. *Epilepsy Res.* 2011;94: 163–176.
doi:10.1016/j.eplepsyres.2011.01.017

128. Traynelis SF, Dingledine R. Potassium-induced spontaneous electrographic seizures in the rat hippocampal slice. *J Neurophysiol.* 1988;59: 259–276.
129. Benini R, Longo D, Biagini G, Avoli M. Perirhinal cortex hyperexcitability in pilocarpine-treated epileptic rats. *Hippocampus.* 2011;21: 702–713. doi:10.1002/hipo.20785
130. Carriero G, Uva L, Gnatkovsky V, Avoli M, de Curtis M. Independent epileptiform discharge patterns in the olfactory and limbic areas of the in vitro isolated Guinea pig brain during 4-aminopyridine treatment. *J Neurophysiol.* 2010;103: 2728–2736. doi:10.1152/jn.00862.2009
131. Prince DA, Wilder BJ. Control mechanisms in cortical epileptogenic foci. “Surround” inhibition. *Arch Neurol.* 1967;16: 194–202.
132. De Curtis M, Biella G, Forti M, Panzica F. Multifocal spontaneous epileptic activity induced by restricted bicuculline ejection in the piriform cortex of the isolated guinea pig brain. *J Neurophysiol.* 1994;71: 2463–2476.
133. Panuccio G, Sanchez G, Lévesque M, Salami P, de Curtis M, Avoli M. On the ictogenic properties of the piriform cortex in vitro. *Epilepsia.* 2012;53: 459–468. doi:10.1111/j.1528-1167.2012.03408.x
134. Giacchino JL, Somjen GG, Frush DP, McNamara JO. Lateral entorhinal cortical kindling can be established without potentiation of the entorhinal-granule cell synapse. *Exp Neurol.* 1984;86: 483–492.
135. Koyama R, Tao K, Sasaki T, Ichikawa J, Miyamoto D, Muramatsu R, et al. GABAergic excitation after febrile seizures induces ectopic granule cells and adult epilepsy. *Nat Med.* 2012;18: 1271–1278. doi:10.1038/nm.2850
136. Fujiwara-Tsakamoto Y, Isomura Y, Nambu A, Takada M. Excitatory GABA input directly drives seizure-like rhythmic synchronization in mature hippocampal CA1 pyramidal cells. *Neuroscience.* 2003;119: 265–275.
137. Scharfman HE, Goodman JH, Du F, Schwarcz R. Chronic changes in synaptic responses of entorhinal and hippocampal neurons after amino-oxyacetic acid (AOAA)-induced entorhinal cortical neuron loss. *J Neurophysiol.* 1998;80: 3031–3046.
138. Cincotta M, Young NA, Beart PM. Unilateral up-regulation of glutamate receptors in limbic regions of amygdaloid-kindled rats. *Exp Brain Res.* 1991;85: 650–658.
139. Prince HK, Conn PJ, Blackstone CD, Haganir RL, Levey AI. Down-regulation of AMPA receptor subunit GluR2 in amygdaloid kindling. *J Neurochem.* 1995;64: 462–465.

140. Lee S, Miskovsky J, Williamson J, Howells R, Devinsky O, Lothman E, et al. Changes in glutamate receptor and proenkephalin gene expression after kindled seizures. *Brain Res Mol Brain Res*. 1994;24: 34–42.
141. Behr J, Heinemann U, Mody I. Kindling induces transient NMDA receptor-mediated facilitation of high-frequency input in the rat dentate gyrus. *J Neurophysiol*. 2001;85: 2195–2202.
142. Luhmann HJ, Dzhalala VI, Ben-Ari Y. Generation and propagation of 4-AP-induced epileptiform activity in neonatal intact limbic structures in vitro. *Eur J Neurosci*. 2000;12: 2757–2768.
143. Borck C, Jefferys JG. Seizure-like events in disinhibited ventral slices of adult rat hippocampus. *J Neurophysiol*. 1999;82: 2130–2142.
144. Nagao T, Alonso A, Avoli M. Epileptiform activity induced by pilocarpine in the rat hippocampal-entorhinal slice preparation. *Neuroscience*. 1996;72: 399–408.
145. Avoli M, Panuccio G, Herrington R, D’Antuono M, de Guzman P, Lévesque M. Two different interictal spike patterns anticipate ictal activity in vitro. *Neurobiol Dis*. 2013;52: 168–176. doi:10.1016/j.nbd.2012.12.004
146. Lücke A, Nagao T, Köhling R, Avoli M. Synchronous potentials and elevations in [K⁺]_o in the adult rat entorhinal cortex maintained in vitro. *Neurosci Lett*. 1995;185: 155–158.
147. Avoli M, Barbarosie M, Lücke A, Nagao T, Lopantsev V, Köhling R. Synchronous GABA-mediated potentials and epileptiform discharges in the rat limbic system in vitro. *J Neurosci Off J Soc Neurosci*. 1996;16: 3912–3924.
148. Avoli M, Mattia D, Siniscalchi A, Perreault P, Tomaiuolo F. Pharmacology and electrophysiology of a synchronous GABA-mediated potential in the human neocortex. *Neuroscience*. 1994;62: 655–666.
149. Zhang CL, Gloveli T, Heinemann U. Effects of NMDA- and AMPA-receptor antagonists on different forms of epileptiform activity in rat temporal cortex slices. *Epilepsia*. 1994;35 Suppl 5: S68–73.
150. Fountain NB, Bear J, Bertram EH 3rd, Lothman EW. Responses of deep entorhinal cortex are epileptiform in an electrogenic rat model of chronic temporal lobe epilepsy. *J Neurophysiol*. 1998;80: 230–240.
151. Bear J, Fountain NB, Lothman EW. Responses of the superficial entorhinal cortex in vitro in slices from naive and chronically epileptic rats. *J Neurophysiol*. 1996;76: 2928–2940.

152. Gnatkovsky V, Librizzi L, Trombin F, de Curtis M. Fast activity at seizure onset is mediated by inhibitory circuits in the entorhinal cortex in vitro. *Ann Neurol*. 2008;64: 674–686. doi:10.1002/ana.21519
153. Jones RS, Lambert JD. Synchronous discharges in the rat entorhinal cortex in vitro: site of initiation and the role of excitatory amino acid receptors. *Neuroscience*. 1990;34: 657–670.
154. Köhling R, Lücke A, Straub H, Speckmann EJ, Tuxhorn I, Wolf P, et al. Spontaneous sharp waves in human neocortical slices excised from epileptic patients. *Brain J Neurol*. 1998;121 (Pt 6): 1073–1087.
155. Louvel J, Papatheodoropoulos C, Siniscalchi A, Kurcewicz I, Pumain R, Devaux B, et al. GABA-mediated synchronization in the human neocortex: elevations in extracellular potassium and presynaptic mechanisms. *Neuroscience*. 2001;105: 803–813.
156. Holtkamp M, Buchheim K, Elsner M, Matzen J, Weissinger F, Meierkord H. Status epilepticus induces increasing neuronal excitability and hypersynchrony as revealed by optical imaging. *Neurobiol Dis*. 2011;43: 220–227. doi:10.1016/j.nbd.2011.03.014
157. D’Antuono M, Benini R, Biagini G, D’Arcangelo G, Barbarosie M, Tancredi V, et al. Limbic network interactions leading to hyperexcitability in a model of temporal lobe epilepsy. *J Neurophysiol*. 2002;87: 634–639.

Figure 1.TIFF

