



The Potential Therapeutic Effect of RNA Interference and Natural Products on COVID-19: A Review of the Coronaviruses Infection

Mohammad Reza Kalhori¹, Fatemeh Saadatpour², Ehsan Arefian², Masoud Soleimani³, Mohammad Hosien Farzaei^{4*}, Ina Yosifova Aneva⁵ and Javier Echeverría^{6*}

¹Medical Biology Research Center, Health Technology Institute, Kermanshah University of Medical Sciences, Kermanshah, Iran, ²Molecular Virology Lab, Department of Microbiology, School of Biology, College of Science, University of Tehran, Tehran, Iran, ³Department of Hematology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran, ⁴Medical Technology Research Center, Health Technology Institute, Kermanshah University of Medical Sciences, Kermanshah, Iran, ⁵Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, Sofia, Bulgaria, ⁶Departamento de Ciencias del Ambiente, Facultad de Química y Biología, Universidad de Santiago de Chile, Santiago, Chile

OPEN ACCESS

Edited by:

Per-Johan Jakobsson,
Karolinska Institutet (KI), Sweden

Reviewed by:

Priya Pusparajah,
Monash University Malaysia, Malaysia
Chun Yang,
The First Affiliated Hospital of Nanjing
Medical University, China

*Correspondence:

Mohammad Hosein Farzaei
mh.farzaei@gmail.com
Javier Echeverría
javier.echeverriam@usach.cl

Specialty section:

This article was submitted to
Ethnopharmacology,
a section of the journal
Frontiers in Pharmacology

Received: 13 October 2020

Accepted: 14 January 2021

Published: 23 February 2021

Citation:

Kalhori MR, Saadatpour F, Arefian E, Soleimani M, Farzaei MH, Aneva IY and Echeverría J (2021) The Potential Therapeutic Effect of RNA Interference and Natural Products on COVID-19: A Review of the Coronaviruses Infection. *Front. Pharmacol.* 12:616993. doi: 10.3389/fphar.2021.616993

The SARS-CoV-2 virus was reported for the first time in Wuhan, Hubei Province, China, and causes respiratory infection. This pandemic pneumonia killed about 1,437,835 people out of 61,308,161 cases up to November 27, 2020. The disease's main clinical complications include fever, recurrent coughing, shortness of breath, acute respiratory syndrome, and failure of vital organs that could lead to death. It has been shown that natural compounds with antioxidant, anticancer, and antiviral activities and RNA interference agents could play an essential role in preventing or treating coronavirus infection by inhibiting the expression of crucial virus genes. This study aims to introduce a summary of coronavirus's genetic and morphological structure and determine the role of miRNAs, siRNAs, chemical drugs, and natural compounds in stimulating the immune system or inhibiting the virus's structural and non-structural genes that are essential for replication and infection of SARS-CoV-2.

Keywords: coronavirus, miRNA, siRNA, natural products, phytochemicals, SARS-CoV-2

INTRODUCTION

Over the past 50 years, a wide range of human and animal diseases have been caused by coronaviruses (CoVs). Since the emergence of Severe Acute Respiratory Syndrome (SARS-CoV) Coronavirus in 2003, a considerable number of new Human coronaviruses (H-CoVs) have been identified (Ding et al., 2003). The appearance of Middle East Respiratory Syndrome (MERS-CoV) coronavirus in 2012 and the continuous occurrence of human cases further intensified attention to the importance of studying these viruses from different aspects (Centers for Disease Control and Prevention, 2012). Tolerance of new mutations and recombination has led to the evolution of this group of viruses, giving them the ability to transmit between species. In most cases, CoV infections are self-limiting, and after completing a reasonable period, the body recovers from the illness (Enjuanes et al., 2000). However, some strains of this family cause severe infections and have been the cause of widely spread epidemics during the last two decades (Mahase, 2020).

The SARS-CoV-2 virus was reported for the first time in Wuhan, Hubei Province, China, and causes respiratory infection. According to WHO statistics, on November 27, 2020, Coronavirus

disease 2019 (COVID-19) killed about 1,437,835 people out of 61,308,161 cases, and the most significant number of deaths and infected people were in the Americas (Chen et al., 2020b). In recent years, many treatment strategies such as prescribing antibiotics, the application of antiviral drugs (including Human Immunodeficiency Virus 1 (HIV-1) protease inhibitors, oseltamivir, and ribavirin), different corticosteroids, interferons, and natural human immunoglobulin have been used for treating patients suffering from H-CoVs. Recently, most treatment strategies for CoVs such as MERS-CoV, SARS-CoV, and Severe Acute Respiratory Syndrome coronavirus-2 (SARS-CoV-2) have been based on inhibiting viral agents involved in the replication, infection, or induction of host immune system agents to combat viral infection (Li and De Clercq, 2020; Prajapat et al., 2020). Numerous studies are being conducted today to prove the role of various biomolecules, including RNA interference (RNAi), fusion inhibitors, neutralizing antibodies, plant, or microbial metabolites, as antiviral compounds (Prajapat et al., 2020).

This review article tries to present some information about the biology, replication, infusion, and pathogenesis of coronavirus. Also, it has decided to explain the existing treatment strategies based on chemical drugs, natural compounds, small interfering RNA (siRNAs), and microRNA (miRNAs) that can be used to fight against coronavirus infection, especially SARS-CoV-2. For this purpose, keywords including natural product, flavonoid, polyphenols, phytochemicals, microRNA, siRNA, Coronavirus, COVID-19 up to August 2020 were searched and evaluated using Scopus, PubMed, WOS, and Google Scholar databases.

CORONAVIRUS

Coronavirus Classification and Taxonomy

Genome organization, genome homology, reproduction strategies in a host, and the virion's structural traits are criteria for CoVs classification by the International Committee for Taxonomy of Viruses (ICTV) (Casals et al., 1980). In terms of phylogenetics, CoVs belong to Coronaviridae, as the class of Nidovirales. Coronaviridae family includes two sub-families: Orthocoronavirinae and Torovirinae. The sub-family of Orthocoronavirinae consists of four genera: Alpha, Beta, Gamma, and Delta coronaviruses that are responsible for infection on a vast range of hosts, from mammals to birds (King et al., 2012). Human infection with CoVs was first reported in 1965 (Hamre and Procknow, 1966). H-CoV 229E and CoV-NL63 are human pathogens of the genus Alpha-CoV, causing common cold (Mcintosh et al., 1970; Monto, 1974). Severe Acute Respiratory Syndrome Coronavirus 1 (SARS-CoV-1), MERS-CoV, and SARS-CoV-2 are phylogenetically classified in the Beta-CoVs genus and have caused a high percentage of mortality in the human population during the last two decades (Zhou et al., 2020). The most common CoVs genes used for phylogenetic studies are nsp12 (RNA dependent RNA polymerase), nsp5 (chymotrypsin-like protease), nsp13 (helicase), nucleocapsid (N), and spike protein (S) (ICTV, 2009).

Genomic Structure

CoVs are positive single-strand RNA with 32–46% G + C content. They have the biggest genome size among all known RNA viruses (about 26.4–31.7 kb) (Lai and Cavanagh, 1997; Enjuanes et al., 2000). Depending on the strain, the genome in CoVs contains a different number of open reading frames (ORF). Nevertheless, ORF1a, ORF1b, envelope (E), protein S, protein N, and membrane protein (M) are present in all Human-CoV strains (Cavanagh, 1995; Enjuanes et al., 2000). The SARS-CoV-2 genome size varies from 29.8 to 29.9 kb and encodes 9,860 amino acids. The ORF1ab in SARS-CoV-2 is over 21 KB in length and covers two-thirds of the entire genome (Casella et al., 2020). According to recent reports, the ORF portion of the SARS-CoV-2 genome has a lower CG dinucleotide percentage than other coronaviruses, so ORF RNA translation is highly efficient (Wang Y. et al., 2020).

Coronavirus Replication Process

The replication cycle of CoVs occurs in several stages (Figure 1). In the initial stage, the virus binding proteins are attached to appropriate receptors on the host cell's surface. Then membrane fusion occurs, which finally leads to virus genome insertion in the host cell. In the second step, the RNA polymerase-dependent RNA gene is activated to translate the virus genome into structural proteins (Ziebuhr, 2005). During these steps, in addition to viral factors, some host cell factors can also enhance or inhibit this process.

Attachment; Entry and Cellular Factors Involvement

The first stage of virus entry starts with the attachment of S proteins to host ligands. Most HCoVs use endocytosis pathways for their entrance, while others can use direct membrane fusion (Gallagher and Buchmeier, 2001). The ectodomain S protein consists of a receptor-binding subunit (S1) and a membrane subunit (S2). After binding to the host cell surface receptors, the S1 subunit leads to a structural change in S2. The S2 proteolytic domain then fuses the virus membrane into the host membrane, leading to the viral genome's entry into the host cell. A partial mutation in the S protein that changes the amino acid sequence can significantly affect the virus's pathogenesis and tissue or cell susceptibility to infection (Li, 2016).

HCoV-NL63, SARS-CoV, and SARS-CoV-2, use Angiotensin-Converting Enzyme 2 (ACE2) to contaminate cells (Moore et al., 2004; Zhang et al., 2020a). Both types of alveolar cells (I and II types) and the endothelial cells express ACE2. It has been shown that MERS-CoV uses Dipeptidyl peptidase 4 (DPP4), a physiological ligand for adenosine deaminase, which is extensively expressed by the endothelial cells in various tissues (Raj et al., 2013). However, many host factors can limit the entry of the virus. According to cell culture mechanisms, a family of Interferon-Inducible Transmembrane proteins (IFITM) could limit S protein-dependent access in HCoV-229E and HCoV-NL63, which would also lead to intense reduction of the infection in SARS-CoV and MERS-CoV (Huang et al., 2011; Wrensch et al., 2014). It seems that IFITM inhibits the membrane fusion by

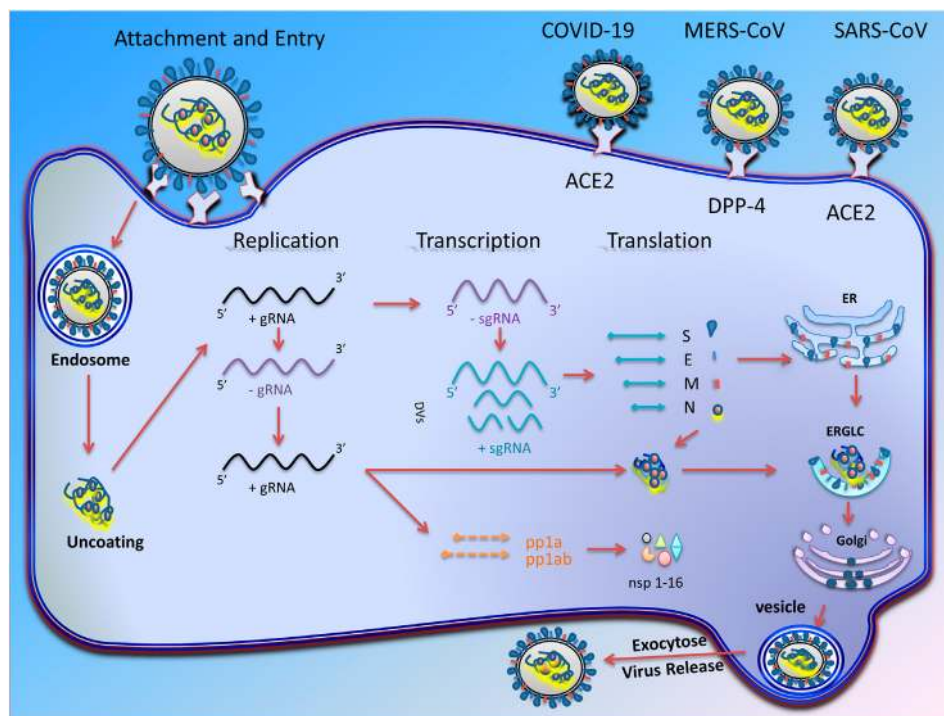


FIGURE 1 | Transcription and replication of coronavirus. Schematic picture showing the coronavirus infection starts with attachment of virus S proteins to receptors on the host cells. After successful fusion, ribonucleocapsid enters the cell cytoplasm and loses its coat to mRNA is released. RNA-dependent RNA synthesis in CoVs includes two different genome replication processes to achieve multiple copies of genomic RNAs (gRNAs) and transcription of sub-genomic RNAs (sg mRNAs) coding structural and accessory proteins. The gRNA eventually producing 16 non-structural proteins (nsp-16) that are involved in virus replication. After assembly and budding, the full virus particles are transferred to the cytoplasmic membrane and finally released through the exocytosis process.

preventing the virus fusion to the cell membrane, endosomal membranes, or through membrane fluidity. Moreover, it was shown that human coronavirus HCoV-OC43 utilizes IFITM2 and IFITM3 as receptors to facilitate cell entry (Zhao et al., 2014).

Transcription and Replication

In CoVs, RNA-dependent RNA polymerase amplifies the virus genome in two ways and eventually produces several copies of genomic RNAs (gRNAs) and subgenomic RNAs (sg mRNAs) (Ziebuhr, 2005). Generally, CoVs replication is initiated with the translation of ORF1a to produce polyprotein1a (pp1a) with 4,382 amino acids and polyprotein1ab (pp1ab) with 7,073 amino acids. Then, the ORF1b is translated through the ribosomal frameshifting mechanism (Lai, 1990; Ziebuhr, 2005). Eventually, each of these polyproteins is cleaved to produce 16 non-structural proteins (nsp-16), which are involved in the virus replication cycle (Mcintosh and Peiris, 2009). Some of the non-structural proteins (**Table 1**) that have critical roles in the virus's life cycle include helicase (nsp13), RNA-dependent RNA polymerase (RdRP) (nsp12 polymerase), 3-chymotrypsin-like protease (3CL^{Pro}) (nsp5 protease), and papain-like protease (PL^{Pro}) (nsp3 protease) (Lai, 1990; Sawicki et al., 2005).

Virus Assembly and Egress

The structural proteins (**Table 2**) and some membrane accessory proteins are translated inside ER-bound ribosomes, while N protein is translated via free ribosomes in the cytoplasm (Nal

et al., 2005). One of the distinct characteristics of CoVs is virion aggregation inside the ER and budding toward the Golgi apparatus (Cawood et al., 2007). Moreover, E, S, and N proteins interpolation to virions is modulated by M protein heterotypic interaction in the budding locus (Opstelten et al., 1995). After assembly and budding, the full virus particles are transferred to the cytoplasmic membrane and finally are released through the exocytosis process (**Figure 1**).

Coronaviruses Pathogenesis

Clinically, the pathogenesis of coronaviruses is divided into three stages. In the viremia phase, the virus enters the peripheral blood after the lungs are infected. Thus, the virus can reach its target tissues, such as the heart, kidneys, and gastrointestinal tract, through the bloodstream. The second phase is the pneumonia phase. Then the patient enters the recovery phase if the immune system can overcome the virus's attack at this stage. However, in patients with immunodeficiency, high blood pressure, diabetes, and the elderly, the immune system cannot effectively manage the infection, and a critical stage of the disease occurs (Peiris et al., 2003; Guan et al., 2020).

TREATMENT

Vaccination is the best way to control the COVID-19 pandemic, and many efforts are being made to produce it. Nevertheless, its

TABLE 1 | Biological activities of coronavirus non-structural proteins.

| Proteins | Biological functions in virus particle | Effect on the host cellular response | Refs. |
|----------|---|--|----------------------------|
| Nsp1 | A virulence factor. The genus-specific marker. Highly divergent among CoVs | Interact with the 40S subunit of ribosome to prevent host mRNA translation. | Woo et al. (2010) |
| Nsp2 | Not reported. | Interaction with two host proteins, PHB1 and PHB2 and disruption of intracellular host cell surviving signaling pathway. | Cornillez-Ty et al. (2009) |
| Nsp3 | Cysteine-type endopeptidase activity at the N-terminus of the replicase polyprotein. | Plays a role in host membrane rearrangement. Downregulate mRNA levels of pro-inflammatory cytokines including CCL5, and CXCL10. | Lei et al. (2018) |
| Nsp4 | Nucleic acid binding by interaction with N protein, RNA-directed 5'-3' RNA-directed 5'-3' RNA polymerase activity. Marker for the coronavirus-induced DMVs. Interacts with nsp3 and nsp6 to formation of the replication complexes. | Responsible for suppressing IFN induction. Antagonize IFN production Not reported. | Angelini et al. (2013) |
| Nsp5 | Cleaves the C-terminus of replicase polyprotein at 11 sites | Bind an ADRP, may cleave host ATP6V1G1 and modifying host vacuoles intracellular pH | Lin et al. (2005) |
| Nsp6 | Necessary for viral replication. Mediates the DMV formation by rearrangement of host membrane. | Plays a role in the initial induction of autophagosomes from host reticulum endoplasmic. Limits the expansion of these phagosomes that are no longer able to deliver viral components to lysosomes. | Angelini et al. (2014) |
| Nsp7 | A primase in the form of heterohexadecamer dsRNA-encircling as ring structure. | Not reported. | Te Velthuis et al. (2012) |
| Nsp8 | A processivity factor for the RdRP. | | |
| Nsp9 | May participate in viral replication by acting as a ssRNA-binding protein. | Not reported. | Miknis et al. (2009) |
| Nsp10 | Interact with nsp1, nsp7, nsp14, and nsp16. Implicated in the regulation of polyprotein processing. Plays a pivotal role in viral transcription by stimulating both nsp14 3'-5' exoribonuclease, and nsp16 2'-O-methyltransferase activities | Not reported. | Bouvet et al. (2012) |
| Nsp12 | Responsible for replication and transcription of the viral RNA genome. | Not reported. | Ahn et al. (2012) |
| Nsp13 | Magnesium dependent helicase activity. Displaying RNA and DNA duplex-unwinding activities with 5' to 3' polarity. | Not reported. | Tanner et al. (2003) |
| Nsp14 | An exoribonuclease acting on both ssRNA and dsRNA in a 3' to 5' direction. | Interacts with DDX1 via N-terminus. | Denison et al. (2011) |
| Nsp15 | Acts as a proofreading exoribonuclease for RNA replication. Mn ²⁺ -dependent, uridylyate-specific enzyme, which leaves 2'-3'-cyclic phosphates 5' to the cleaved bond. | Modulation of the innate immune response. Essential to evade dsRNA sensors. | Ricagno et al., 2006 |
| Nsp16 | Mediates mRNA cap 2'-O-ribose methylation to the 5'-cap structure of viral mRNAs. | Essential to evade MDA5 recognition. Negatively regulating innate immunity. IFN antagonism. | Bouvet et al. (2010) |

production is time-consuming because it must first be proven to be immunogenic and effective. Therefore, prevention is currently the best treatment. There are two essential strategies for treatment this disease. The first step is a general treatment that reduces the symptoms and clinical complications of the patients, and the second is drug treatment.

General Treatment

The available treatment options for these patients include measuring and monitoring of vital signs such as heart function, kidney, liver, respiratory rate, and oxygen therapy if necessary. Moreover, patients must be getting enough water, sufficient calories, balance for electrolytes, and use of fever medicines such as acetaminophen and ibuprofen (Wang J. et al., 2020; Zimmermann and Curtis, 2020). Although the use of corticosteroids such as methylprednisolone in the SARS-CoV epidemic has been shown to improve some clinical symptoms, it

is not common in COVID-19. Nevertheless, it could be used only temporarily in patients with severe illness such as dyspnea and Acute Respiratory Distress Syndrome (ARDS) (Huang et al., 2020; Russell et al., 2020; Zhang et al., 2020b). Herbal medicines have the property to be used as a dietary supplement for relieving and reduce respiratory symptoms and other symptoms of COVID-19. They can improve the general condition of patients with mild disease severity. *Althaea officinalis* L. (Malvaceae), *Commiphora myrrha* (T.Nees) Engl. (Burseraceae), *Glycyrrhiza glabra* L. (Fabaceae), *Hedera helix* L. (Araliaceae), and *Sambucus nigra* L. (Viburnaceae) are some of the herbal medicines can be used as adjunctive therapy for mild COVID-19 (Silveira et al., 2020).

Coronavirus Drug Treatment

SARS-CoV-2 is a new virus from the Coronaviruses family, with 79% genomic similarity to SARS-CoV and 51.8% to MERS-CoV.

TABLE 2 | The biological functions of structural proteins of SARS and SARS-CoV 2.

| Protein | Post-translational modification | Biological functions in virus particle | Effect on the host cellular response | Refs. |
|---------|---|--|---|---|
| S | Disulfide bridge, Palmitoylation, N-glycosylation | Virulence factor. Responsible for recognition of the cellular receptor. Fusion of virus membrane with host endosome membrane. | Physically interaction with eIF3F. Modulation the expression of the pro-inflammatory cytokines IL 6 and 8 at a later stage of infection. | Bosch et al. (2003) and Nal et al. (2005) |
| M | O-glycosylation, N-glycosylation | Plays a central role in virus morphogenesis. Maintain the structure and assembly via its interactions with other viral proteins such as 3a and 7a, forms a complex with HE and S proteins. Promotes membrane curvature, binds to the nucleocapsid and participates in RNA packaging into the virus | Pro-apoptotic mediated activation of caspases 8 and 9. Suppress IFN I production mediated by RIG-I and inhibiting the translocation of IRF3 into the nucleus, IFN antagonism | Nal et al. (2005) and Siu et al. (2009) |
| E | Palmitoylation, glycosylation | Plays a central role in virus morphogenesis and assembly. Responsible for the curvature of the viral envelope. A virulence factor trafficking within the infected cells and budding of the virion. | Induction of the cell stress response and mitochondrial-mediated apoptosis. Disruption of the lung epithelium, Potential B cell antagonism. | An et al. (1999) and Liu et al. (2007) |
| N | O-glycosylation, ADP-ribosylation, Sumoylation, Phosphorylation | An RNA chaperone, associates with the viral genome in a helical nucleocapsid. Plays a fundamental role during virion assembly through its interactions with the viral genome and membrane protein M, E and nsp3. Plays an important role in enhancing the efficiency of sgRNA transcription and viral replication. | Modulate transforming growth factor-beta signaling by binding host smad3. Interfere with the function of IRF3. Inhibition of IFN I response. Induction of apoptosis. | Zhao et al. (2006) and Wu et al. (2009) |

Therefore, since the risk of SARS-CoV-2 vertical infection is the same as these two types of viruses, their treatment can be similar. Consequently, drugs that were previously effective for the treatment of SARS-CoV and MERS-CoV may also have therapeutic potential for the treatment of COVID-19 (Ren et al., 2020). Although a definitive cure for this new virus has not yet been discovered, previous studies suggest that drugs such as western medicines and natural products may have potential efficiency against COVID-19 and could be used to reduce the severity of the disease.

Interferon

Interferon type I is a member of the innate immune system produced and secreted in response to viral infections, including alpha and beta-interferon. Interferons exert their antiviral activity in two ways 1) cytotoxic T lymphocytes and macrophages stimulate the immune system to kill the virus by increasing or stimulating natural killer cells, 2) or inhibit virus replication in the host cell (Samuel, 2001; Sadler and Williams, 2008; Fensterl and Sen, 2009). Previous studies have shown that interferon- α , particularly its recombinant form (INF- α 2b), can alleviate symptoms and shorten the disease course of respiratory tract viral infections in the early phase (caused by influenza, SARS-CoV, and MERS-CoV) through inhibiting virus replication and decreasing the virus load. Therefore, these molecules may be useful in the treatment of COVID-19 (Zheng et al., 2004; Danesh et al., 2011; Falzarano et al., 2013; Khalid et al., 2015). However, according to the latest update (3.2020) of the COVID-19 Treatment Guidelines Panel, there is no comment about its therapeutic effect on COVID-19 (COVID-19 Treatment Guidelines Panel).

Lopinavir/Ritonavir

Lopinavir/ritonavir is a protease inhibitor that has previously been used against HIV due to its antiviral activity. Nevertheless, later in the SARS-CoV (an epidemic that occurred in 2003), it was discovered that lopinavir/ritonavir by 3CL^{pro}-inhibitor action could be useful for coronavirus treatment (Chu et al., 2004; Su et al., 2019). Due to the structural similarity of 3CL^{pro} in SARS-CoV2 and SARS-CoV, previous studies have suggested that this drug may be useful for treating SARS-CoV2 (Uzunova et al., 2020). Nevertheless, later it was found that these drugs do not have the desired effect in treating this infectious disease, and the coronavirus disease 2019 treatment guidelines (last updated: November 3, 2020) announced that its use is not recommended, except in a clinical trial (COVID-19 Treatment Guidelines Panel, 2019).

Ribavirin

Ribavirin is a nucleoside analog that has antiviral activity. Co-administration of lopinavir/ritonavir has a significant therapeutic effect against SARS-CoV and may reduce ARDS risk (Chu et al., 2004). However, according to the latest update (November 3, 2020) of the COVID-19 Treatment Guidelines Panel, there is no comment about its therapeutic effect on COVID-19 (COVID-19 Treatment Guidelines Panel).

Chloroquine

Chloroquine is an antiparasitic drug used against malaria, but recently it has been shown to have antiviral activity by blocking virus infection and could suppress SARS-CoV-2 infection *in vitro* (Wang M. et al., 2020; Colson et al., 2020). Although many physicians and specialists initially considered this drug to treat

COVID-19, its use is no longer recommended today (COVID-19 Treatment Guidelines Panel).

Favipiravir

One of the promising drugs in the treatment of COVID-19 is favipiravir. This drug has an inhibitory effect on RNA polymerase, and its early clinical trial showed that not only is the antiviral activity of this drug more significant than lopinavir/ritonavir, but it has fewer side effects (Cai et al., 2020). However, according to the latest update (November 3, 2020) of the COVID-19 Treatment Guidelines Panel, there is no comment about its therapeutic effect on COVID-19 (COVID-19 Treatment Guidelines Panel).

Remdesivir

Remdesivir is one of the drugs that can be used against RNA viruses such as SARS-CoV/MERS-CoV. *In vivo* and *in vitro* studies have shown that this adenosine analog by inhibiting RdRP has a therapeutic effect against COVID-19 and could be a choice for treating this new pandemic infectious disease (Holshue et al., 2020; Wang M. et al., 2020). This drug is currently recommended to treat COVID-19 according to the latest update (November 3, 2020) of the COVID-19 Treatment Guidelines Panel (COVID-19 Treatment Guidelines Panel).

Arbidol

Arbidol is an antiviral compound for prophylaxis and treatment of influenza. An *in vitro* study has proved that arbidol has a direct antiviral activity on SARS-CoV replication (Wang X. et al., 2020). A clinical trial of this drug on SARS-CoV-2 has also shown that it reduces high flow nasal catheter (HFNC) oxygen therapy, enhances the process of viral clearance and improves focal absorption on radiologic images (Xu et al., 2020a). Arbidol could inhibit host cell adhesion and SARS-CoV-2 spike glycoprotein trimerization. Furthermore, the simultaneous use of arbidol by Lopinavir/ritonavir has a more significant therapeutic effect than the Lopinavir/ritonavir group (Deng et al., 2020). However, according to the latest update (November 3, 2020) of the COVID-19 Treatment Guidelines Panel, there is no comment about its therapeutic effect on COVID-19 (COVID-19 Treatment Guidelines Panel).

Antithrombotic Therapy

Anticoagulants, like heparins, play a vital role in preventing arterial thromboembolism in patients with heart arrhythmias. One of the leading causes of death in patients with COVID-19 is myocardial and stroke infarction for unknown reasons. Therefore, if hospitalized patients with COVID-19 due to hemophilia or similar diseases do not have anticoagulant contraindications, administration of a prophylactic dose plays a vital role in reducing the complications COVID-19 in patients with symptoms of blood coagulation (such as low levels of antithrombin, increased fibrinogen and di-dimer) (Godino et al., 2020).

Natural Products as Antiviral Agents

Natural products are biochemical mixtures produced by living organisms in nature. The effectiveness of natural compounds in

ancient medicine was introduced thousands of years ago (Ji et al., 2009). These compounds, with various pharmacological traits, have antioxidant, anticancer, anti-inflammatory, and antiviral activities. Today, many of the chemical drugs used in medicine are derived from natural compounds (Frabasile et al., 2017). The therapeutic strategy for natural products and chemical drugs is to target the protein molecules needed for each stage of the virus's life cycle (Dias et al., 2012). This section discusses the natural compounds' role in the inhibition of coronavirus infection (Table 3; Figure 2; Figure 3; and Figure 4). It is important to emphasize that within the compounds listed in Table 3, only epigallocatechin-3-gallate (NCT04446065) and resveratrol and zinc picolinate combination therapy (NCT04542993) has been investigated in clinical phase-2 trials against COVID-19 infection.

Targeting S Protein, ACE2, and TMPRSS2 to Inhibit Cell Attachment and Entry

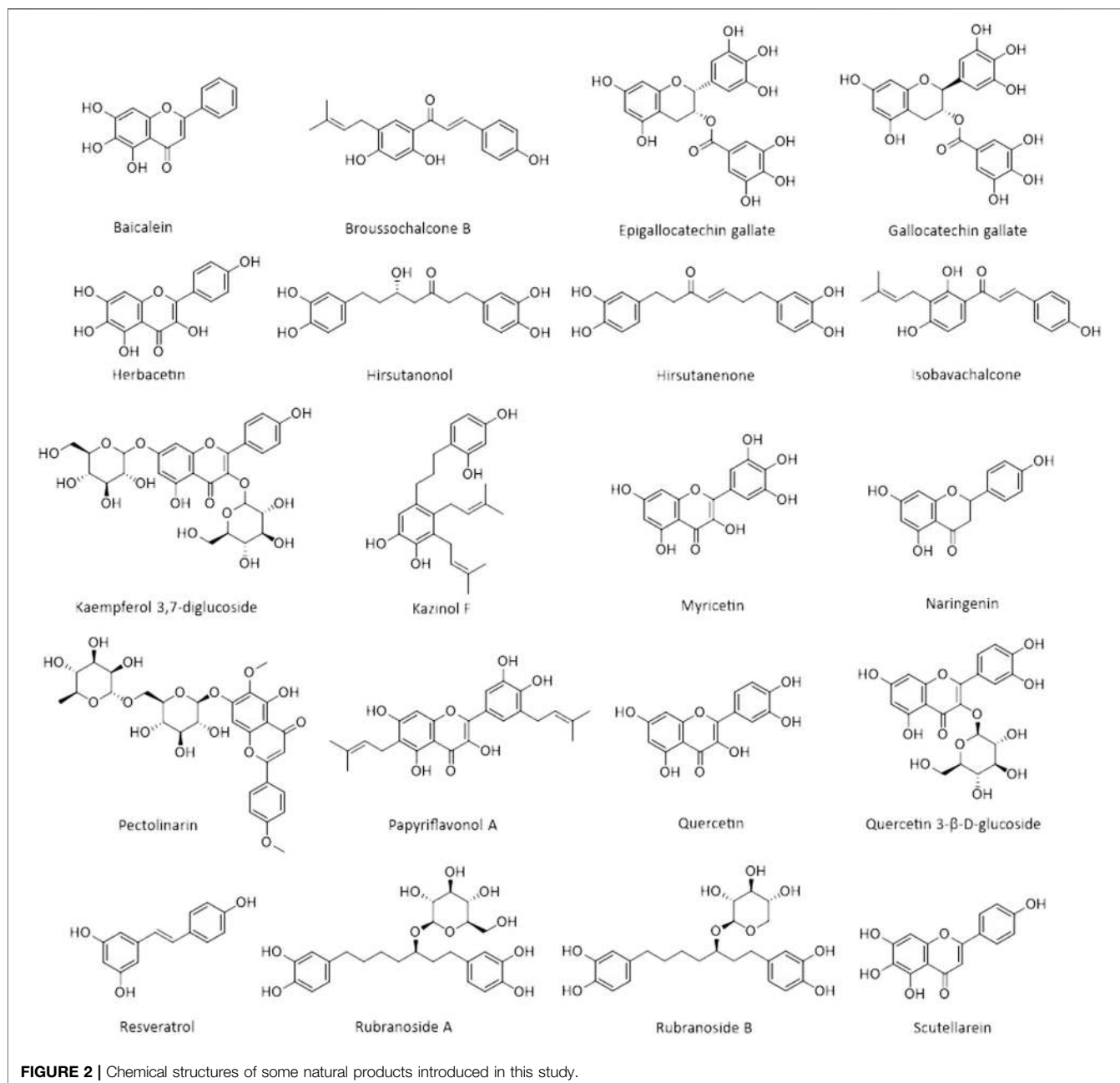
When a coronavirus enters the host cell, it must first, through its S protein, bind to the specific receptors at the surface of host cells. The particular receptor for SARS-CoV and SARS-CoV-2 is the ACE2 protein. To reduce the viral entry strength, the expression of the virus S protein can be inhibited or the expression of ACE2 in host cells can be reduced by natural products. Studies have shown that the amino acid sequence of SARS-CoV Spike protein is approximately 76.47%, similar to SARS-CoV-2 (Xu et al., 2020b). Since virus S protein is highly glycosylated, the use of plant-derived lectins, which tend to bind to glycosylated proteins, can prevent the virus from binding to receptors on the cell surface. Previous studies have shown that herbal compounds could be used as inhibitors for the human immune deficiency virus and SARS-CoV feline (Pyrce et al., 2007). Emodin belongs to the anthraquinones compounds (a group of natural compounds), which have anti-inflammatory, anticancer, and antioxidant activities (Izhaki, 2002). In a dose-dependent manner, emodin could significantly inhibit the ACE2 and S protein interaction in the biotinylated Enzyme-Linked Immunosorbent Assay (ELISA) method and Vero E6 cell line. Therefore, we can consider this compound as an antiviral therapeutic agent in the treatment of SARS-CoV (Ho et al., 2007). Some drugs and substances could reduce the expression of the ACE2 receptor. As seen, interleukin-4 and interferon- γ decreased this receptor in Vero E6 cells (De Lang et al., 2006). Molecular docking and structure-based drug design study reveal that berberine, thebaine, mangiferin, piperine, nimbin, and curcumin have an inhibitory effect against ACE2 receptor and spike glycoprotein of SARS-CoV-2. Among these, curcumin and nimbin have shown the highest interaction with ACE2 receptors and spike glycoprotein than other natural compounds and chemical drugs such as hydroxychloroquine, nafamostat, and captopril (Maurya et al., 2020).

The citrus peel is rich in flavonoid and alkaloid compounds (such as naringin, naringenin, hesperetin, and hesperidin) that play an essential role in maintaining gastrointestinal health and improving the immune system. Naringin is an anti-inflammatory substance that decreases the inflammatory effect in LPS-induced RAW 264.7 macrophages and LPS-treated rats. Therefore, it can inhibit the increase of pro-inflammatory cytokines such as IL-6,

TABLE 3 | Different types of phytochemicals by their sources and function in the coronavirus infection.

| Compound name | Phytochemical class | Source | Function | Clinical trial stage | References |
|-----------------------------------|----------------------|---|---|-----------------------------------|--|
| Baicalein | Flavonoid | <i>Scutellaria baicalensis</i> Georgi, <i>Scutellaria lateriflora</i> L. (Lamiaceae) | Inhibit 3CL ^{pro} of SARS-CoV-1 Decreased the levels of IL-1 β and TNF- α in serum during the SARS-CoV-2 infection | — | Su et al. (2020) Song et al. (2020) |
| Broussonetone B | Polyphenol | <i>Broussonetia papyrifera</i> (L.) L'Hér. ex Vent. (Moraceae) | Inhibit PL ^{pro} of SARS-CoV-1 and MERS-CoV | — | Park et al. (2017) |
| Curcumin | Polyphenol | <i>Curcuma longa</i> L. (Zingiberaceae) | Inhibit the ACE2 receptor and spike glycoprotein of SARS-CoV-2 | — | Maurya et al. (2020). |
| Emodin | Anthraquinone | <i>Rheum palmatum</i> L. (Polygonaceae) | Inhibit the interaction of SARS-CoV-1 S protein and ACE2 | — | Ho et al. (2007) |
| | | | Inhibit 3 α protein channel of SARS-CoV-1 | — | Schwarz et al. (2011) |
| Epigallocatechin-3-gallate | Polyphenol | <i>Camellia sinensis</i> (L.) Kuntze (Theaceae) | Inhibit 3CL ^{pro} of SARS-CoV-1 and SARS-CoV-2 | Phase 2 (SARS-CoV-2) | Jang et al., (2020) and Nguyen et al. (2012) |
| Gallocatechin gallate | Polyphenol | <i>Camellia sinensis</i> (L.) Kuntze (Theaceae) | Inhibit 3CL ^{pro} of SARS-CoV-2 | — | Jang et al. (2020) |
| | | | Inhibit PL ^{pro} and CL ^{pro} of SARS-CoV-1 | — | Nguyen et al. (2012) and Park et al. (2016) |
| Herbacetin | Flavonoid | <i>Linum usitatissimum</i> L. (Linaceae) | Block the proteolytic activity of SARS-CoV-2 3CL ^{pro} | — | Jo et al. (2020a) |
| Hirsutanonol | Phenol | <i>Alnus glutinosa</i> (L.) Gaertn. (Betulaceae) | Inhibit PL ^{pro} and 3CL ^{pro} of SARS-CoV-1 | — | Park et al., 2012a |
| Hirsutenone | Phenol | <i>Alnus japonica</i> (Thunb.) Steud. (Betulaceae) | Inhibits PL ^{pro} activity of SARS-CoV-1 | — | Park et al. (2012a) and Park et al. (2012b) |
| Iodobananin | Oligo-oxa-adamantane | Natural product derivative | Inhibit NTPase/Helicase of SARS-CoV-1 | — | Tanner et al. (2005) |
| Isobavachalcone | Flavonoid | <i>Cullen corylifolium</i> (L.) Medik. (syn. <i>Psoralea corylifolia</i>) (Fabaceae) | Inhibit PL ^{pro} of SAR-CoV-1 | — | Kim et al. (2014) |
| Kaempferol 3,7-diglucoside | Glycosyloxyflavone | <i>Asplenium ruta-muraria</i> L., <i>Asplenium scolopendrium</i> L. (syn. <i>Asplenium altajense</i>) (Aspleniaceae) | Block virus ion channels in SARS-CoV-1 | — | Schwarz et al. (2014) |
| Kazinol F | Polyphenol | <i>Broussonetia papyrifera</i> (L.) L'Hér. ex Vent. (Moraceae) | Target SARS-CoV2-S spike protein Inhibit PL ^{pro} of SARS-CoV-1 and MERS-CoV | — | Pan et al. (2020) Park et al. (2017) |
| Myricetin | Polyphenol | <i>Myristica fragrans</i> Houtt. (Myristicaceae) | Inhibit helicase and nsP13 of SARS-CoV-1 | — | Yu et al. (2012) |
| Naringin | Flavonoid | <i>Citrus x aurantium</i> L., (Rutaceae) | Inhibit 3CL ^{pro} of SARS-CoV-1 | — | Nguyen et al. (2012) |
| Naringenin | Flavonoid | <i>Citrus</i> fruits (Rutaceae) | Two-Pore Channels (TPCs) inhibitor MERS-CoV | — | Pafumi et al. (2017) |
| | | | Block the enzymatic activity 3CL ^{pro} in SARS-CoV-1 | — | Jo et al. (2020b) |
| Pectolinarin | Flavonoid | <i>Cirsium chanroenicum</i> (Nakai) Nakai (Asteraceae) | Block the enzymatic activity of SARS-CoV-1 3CL ^{pro} | — | Jo et al. (2020b) |
| Papyriflavonol A | Flavonoid | <i>Broussonetia papyrifera</i> (L.) L'Hér. ex Vent. (Moraceae) | Inhibit PL ^{pro} of SARS-CoV-1 and MERS-CoV | — | Park et al. (2017) |
| Quercetin | Glycosyloxyflavone | <i>Allium cepa</i> L. (Amaryllidaceae) | Target SARS-CoV2-S spike protein | — | Pan et al., 2020 |
| Quercetin 3- β -D-glucoside | Flavonoid | <i>Passiflora subpeltata</i> Ortega (Passifloraceae) | Inhibit 3CL ^{pro} of MERS- CoV | — | Jo et al. (2019) |
| Resveratrol | Polyphenol | <i>Vitis vinifera</i> L. (Vitaceae) | Inhibit nucleocapsid protein translation in and reduced the MERS-CoV-mediated apoptosis | Phase 2 (SARS-CoV-2) ^a | Lin et al. (2017) |
| Rhoifolin | Flavonoid | <i>Toxicodendron succedaneum</i> (L.) Kuntze (syn. <i>Rhus succedanea</i>) (Anacardiaceae) | Inhibit 3CL ^{pro} SARS-CoV-1 | — | Nguyen et al. (2012) |
| Rubranoside A | Diarylheptanoids | <i>Alnus hirsuta</i> (Spach) Rupr. (syn. <i>Alnus sibirica</i>) (Betulaceae) | Inhibit 3CL ^{pro} and PL ^{pro} SARS-CoV-1 | — | Park et al. (2016) |
| Rubranoside B | Diarylheptanoids | <i>Alnus japonica</i> (Thunb.) Steud. (Betulaceae) | Inhibit 3CL ^{pro} and PL ^{pro} SARS-CoV-1 | — | Park et al. (2016) |
| Scutellarein | Flavonoid | <i>Scutellaria lateriflora</i> L. (Lamiaceae) | Inhibit helicase, nsP13 SARS-CoV-1 | — | Yu et al. (2012) |
| Vanillinbananin | Oligo-oxa-adamantane | Natural product derivative | Inhibit NTPase/Helicase of SARS-CoV-1 | — | Tanner et al. (2005) |

^aResveratrol and Zinc Picolinate combination therapy.



IL-1 β , iNOS, and COX-2 induced by LPS treatment. Molecular docking investigation showed that naringin, naringenin, and hesperetin could bind to ACE2, such as chloroquine. Furthermore, these compounds can bind to ACE2 at lower binding energy levels than chloroquine (docking energy of 5.7 kcal/mol). However, more *in vitro* and *in vivo* experimentation is needed to determine whether these compounds are more effective than chloroquine (da Silva Antonio et al., 2020). However, it should be noted that inhibition of ACE2 due to its important physiological activities (such as homeostasis of blood pressure, protection against pulmonary cell destruction, electrolyte retention, and water) may have detrimental effects on patients' life. In addition to

ACE2, Transmembrane Protein Serine 2 (TMPRSS2) is another essential protein for SARS-CoV-1 and SARS-CoV-2 infection. TMPRSS2 induces viral fusion with the host cell membrane through irreversible structural changes in viral S protein (Hoffmann et al., 2020). As a result, the inhibition of TMPRSS2 in the animal model's airways decreases the severity of lung damage after infection by SARS-CoV and MERS-CoV (Chikhale et al., 2020). Therefore, one of the strategies that could block the entry and restrict the pathogenesis of SARS-CoV-2 is the inhibition of TMPRSS2. The results of two independent studies using computational biology and molecular docking showed that some natural compounds such as neohesperidin, myricitrin, quercitrin, naringin, icariin, citicoline,

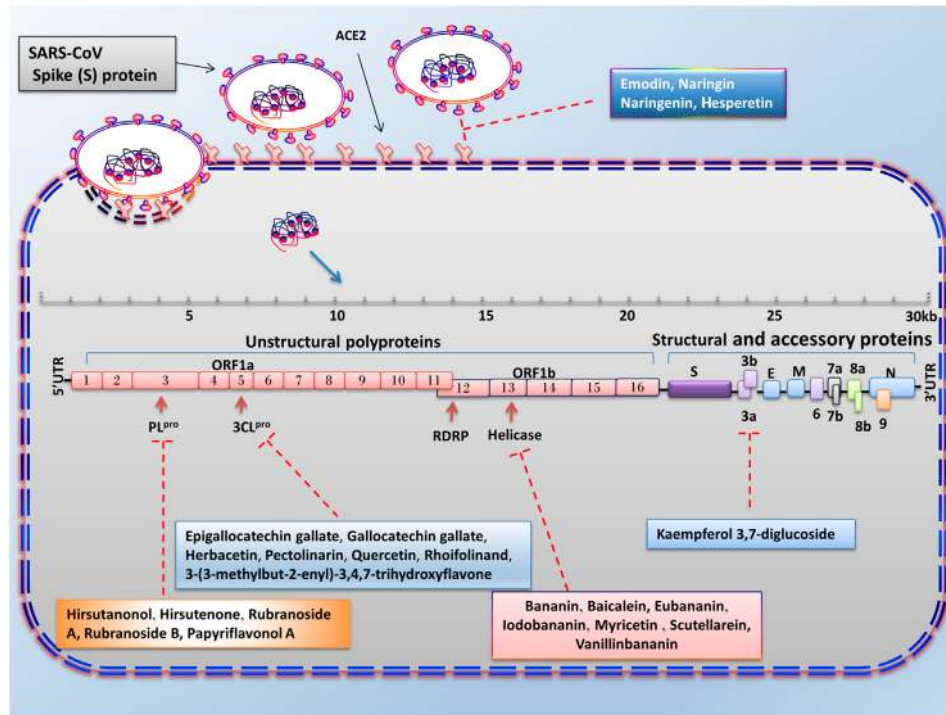


FIGURE 3 | Advantages of employing natural compounds against infection and replication of SARS-CoV. Some of the natural compounds by reducing the expression of 3CL^{pro}, PL^{pro}, helicase, and 3a genes can play a therapeutic role in inhibiting the SARS-CoV infection.

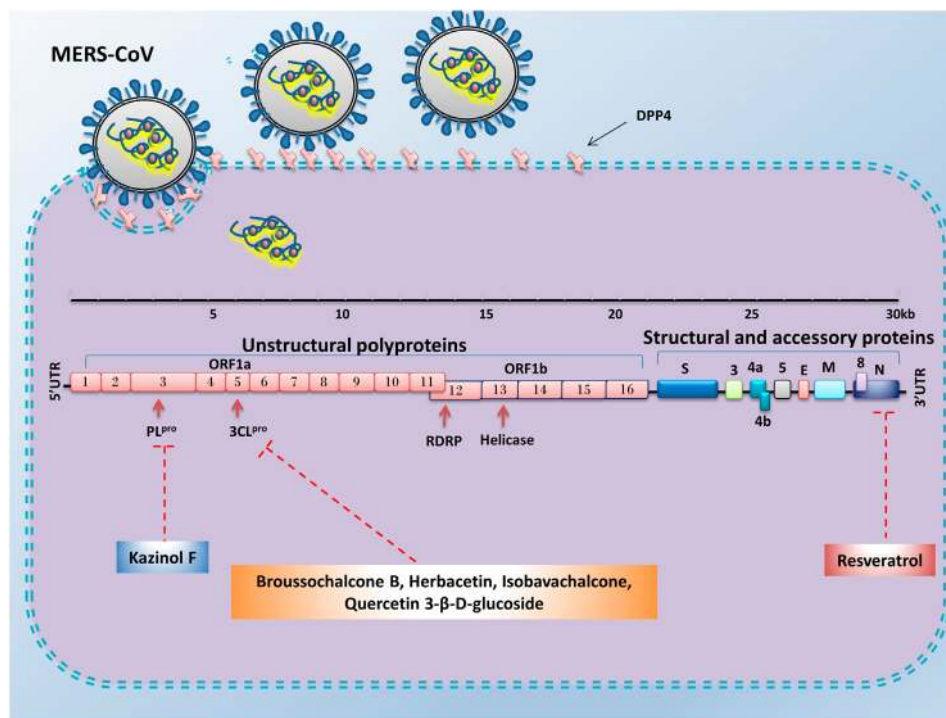


FIGURE 4 | Advantages of employing natural compounds against infection and replication of MERS-CoV. Some of the natural compounds by reducing the expression of nucleocapsid, 3CL^{pro}, and PL^{pro} genes could play a therapeutic role in inhibiting the MERS-CoV infection.

bianthraquinone, isogemichalcone B, and (-)-epigallocatechin-(3-O-methyl) gallate could be used as a TMPRSS inhibitor (Chikhale et al., 2020; Rahman et al., 2020).

Targeting Helicase and Inhibiting Virus Replication

SARS-CoV-2 Helicase protein is encoded by the nsp 13, a gene located downstream of the RdRP. The inhibition of this protein could reduce virus replication (Ivanov et al., 2004). Helicases could separate the double-stranded Nucleic Acid (NA) by using free energy that is obtained from the hydrolysis of Nucleoside Triphosphate (NTP). Therefore, this protein is a potential target for antiviral drug development. Previous studies revealed that various natural products could suppress helicase (unwinding or ATPase) activity. In 2005, Tanner et al. investigated the effect of adamantane-derived bananins on SARS-CoV helicase.

Their results showed that iodobananin, vanillinbananin, bananin, and eubananin had an inhibitory effect on the helicase protein's ATPase activity with IC_{50} rates in the range 0.5–2.8 μM . Furthermore, using Fluorescence Resonance Energy Transfer (FRET) it was found that these four compounds could also inhibit unwinding helicase activity. The authors also observed that these compounds were ineffective against *E. coli* helicase but only affected SARS helicase, so they did not have a general helicase inhibitor activity. Finally, the results of cytopathic effects in fetal rhesus kidney-4 cells (FRhK-4) and RT-PCR demonstrated that these compounds could reduce virus replication without being toxic to the cell (Tanner et al., 2005). In 2012, the results of a study by Yu et al. showed that of the sixty-four natural compounds (flavonoids), only myricetin and scutellarein could reduce SARS-CoV helicase activity. The events showed that these two compounds could reduce Helicase ATPase activity by up to 90% at a dose of 10 μM without any toxicity effects on the healthy breast cell (MCF10A). Moreover, further analysis has shown that they do not change the helicase unwinding activity (Yu et al., 2012). Baicalein is another natural product that has an inhibitory effect on SARS-CoV (nsp13) helical activity. Studies have shown that this compound has no inhibitory effect on the dsDNA-unwinding activity of nsP13; however, it can decrease helicase ATPase activity up to 60% with an IC_{50} value 0.47 μM (Keum et al., 2013).

Targeting 3CL^{PRO}, PL^{PRO} Protease and Inhibiting Virus Protein Processing

Another important antiviral strategy is the use of specific inhibitors against viral proteases such as 3CL^{PRO} and PL^{PRO}, which play an essential role in the processing and maturation of proteins and virus replication. The SARS-CoV-2 genome (positive single-stranded RNA), after translation, is capable of producing two polypeptides, pp1a and pp1ab. Finally, these two polypeptides, cleaved by 3CL^{PRO} or PL^{PRO} activity, produce sixteen non-structural proteins (Thiel et al., 2003; Muramatsu et al., 2016). Flavonoids are a group of phenolic compounds found in plants with anti-inflammatory, antiviral, antioxidant, and anticancer activities. There have been many reports of reduced

coronavirus infection with these compounds (Tapas et al., 2008). *In vitro* studies in *Pichia pastoris* GS115 showed that epigallocatechin-3-gallate, quercetin, and gallic acid could reduce the expression of SARS-CoV 3CL^{PRO}. Molecular docking experiments and kinetic enzyme studies also revealed that these three compounds with IC_{50} rates in the range 47–73 μM could reduce protease activity up to 80%. In the meantime, gallic acid's effect was more significant than the other compounds (Nguyen et al., 2012). Rhoifolinand, herbacetin, and pectolarin are other flavonoid compounds that have an adverse influence on 3CL^{PRO}. Studies with FRET protease assays and absorption spectroscopic studies have shown that these compounds can bind to 3CL^{PRO} and significantly inhibit its protease activity (drug concentration less than 40 μM in IC_{50}) (Jo et al., 2020b). In another study, Jo et al. examined the inhibitory effects of several flavonoid compounds against MERS-CoV 3CL^{PRO}. Their results reveal the isobavachalcone, quercetin 3- β -D-glucoside, and herbacetin had noticeable inhibitory actions with IC_{50} values of 35.85, 37.03, 40.59 μM , respectively (Jo et al., 2019). The 3CL^{PRO} of SARS-CoV-2 is highly conserved among all CoVs and has approximately 96% similarity with SARS-CoV-1. Due to its essential role in viral replication, it is a potential therapeutic target for COVID-19 (Xu et al., 2020c). Since natural products derived from microbial sources have a unique chemical diversity compared to plant products, more than 50% of FDA-approved natural compound-based medicines are derived from microbial compounds. The virtual screening and molecular binding by Sayed et al. showed that citriquinochroman, holyrine B, proximicin C, and several other microbial compounds could inhibit 3CL^{PRO} of SARS-CoV-2 (Sayed et al., 2020). In addition to microbial compounds, molecular docking and MD simulation studies have shown that some marine natural products including hydroxypentafuhalol, pentaphloretol B, and luteolin-7-rutinoside (ΔG about -14.6 to -10.7 kcal/mol) can also inhibit 3CL^{PRO} of SARS-CoV-2 (Gentile et al., 2020).

The Papain-like protease (PL^{PRO}), is another protease that controls the proliferation of SARS-CoV-2 and is known as a potential target for treating this virus. Previous studies have shown that *Alnus japonica* (Thunb.) Steud. (Betulaceae) has anticancer, anti-inflammatory, and anti-influenza properties. The study of Kim et al. showed that the natural phenolic compounds (diarylheptanoids) prepared from this plant could change the proteolytic activity of PL^{PRO}. The fluorometric assay results showed that among the nine extracted substances, hirsutenone, hirsutanonol, rubranoside B, and rubranoside A had a dose-dependent inhibitory effect against PL^{PRO}. Among these compounds, hirsutenone had a remarkable inhibitory effect on SARS-CoV PL^{PRO} (IC_{50} = 4.1 μM) and 3CL^{PRO} (IC_{50} = 36.2 μM) enzyme activity. Further study showed that this substance, containing an α , β -unsaturated carbonyl group with a catechol moiety in the backbone, and the presence of this structure played an essential role in its inhibitory effects (Park et al., 2012a). In addition, polyphenols derived from the root of *Broussonetia papyrifera* (L.) L'Hér. ex Vent. (Moraceae) have been shown to have good inhibitory potential against SARS-CoV and MERS-CoV proteases. Park et al. investigated the inhibitory effect of ten

natural compounds derived from this plant on SARS-CoV and MERS-CoV proteases. The effect of these compounds on SARS-CoV proteases showed that the papyriflavonol A (Broussonol E) was the most effective inhibitor of PL^{Pro} (IC₅₀ value 3.7 μM) and 3-(3-methylbut-2-enyl)-3,4,7-trihydroxyflavone was the most useful inhibitor of 3CL^{Pro} (IC₅₀ value of 30.2 μM). Additionally, broussonol B (Bavachalcone), with a concentration of 27.9 μM (IC₅₀) and kazinol F with a concentration of 39.5 μM (IC₅₀), were able to reduce the activity of MERS-CoV 3CL^{Pro} and MERS-CoV PL^{Pro}, respectively (Park et al., 2017).

Targeting Nucleocapsid (N) Protein to Inhibit Virus Infection and Replication

Resveratrol is a natural compound with anti-inflammatory, antioxidant, and anticancer properties (Yeung et al., 2019; Filardo et al., 2020). The previous study demonstrated that this compound has antiviral activity and can inhibit viral infections caused by Herpes Simplex Virus (HSV), Respiratory Syncytial Virus (RSV), and Epstein-Barr Virus (EBV) (Faith et al., 2006; Zang et al., 2011; De Leo et al., 2012). Also, resveratrol by repressing the expression of MERS-CoV N protein in the Vero E6 cell line, could reduce RNA expression, viral yield, and replication of MERS-CoV. Additionally, it significantly decreases the virus's infection and enhances infected cells' survival by repressing Caspase 3 cleavage (Lin et al., 2017). Other natural compounds that can reduce coronavirus infection risk include the alkaloids fangchinoline, cepharanthine, and tetrandrine. The results of a study by Kim et al. showed that these compounds could inhibit the expression of pro-inflammatory cytokines (IFN-α1, IL-6, IFN-β1, IL-8, and IL-1) caused by human coronavirus OC43 infection in the MRC-5 cell line. These three natural compounds can also decrease the OC43 replication by inhibiting N protein expression and reducing the cytotoxic effect of this virus in the MRC-5 cell line, and increasing the proliferation and survival of MRC-5 human lung cells (Kim et al., 2019).

Targeting 3A Protein and Inhibiting Virus Release

The production and release of the virus require some ion channels in the host cell membrane. Therefore, inhibition of these ion channels played a significant role in inhibiting viral infections, so one antiviral strategy is to use compounds that can restrain these channels (Liang and Li, 2010). One of these ion channels is the cation-selective channel (3a protein) generated by the ORF3a of the SARS-CoV genome. Kaempferol glycoside is a natural flavonol found in a variety of plants. Previous studies have shown that this compound has antiviral properties (IC₅₀ value of 2.3 μM) and can inhibit the expression of 3a protein SARS CoV (an ion channel) in *Xenopus* oocyte as a model system (Schwarz et al., 2014).

RNA INTERFERENCE

RNA interference (RNAi) are RNA molecules found in many eukaryotes that inhibit gene expression by targeted 3'UTR of

mRNA molecules. Today, siRNA and miRNA are the most common type of RNAi used for gene silencing. RNAi origin can be endogenous (originating in the cell) and exogenous (coming from a virus or laboratory tools). Exogenous RNAi can be transmitted to cells using electroporation, viral vectors, liposomes, and calcium phosphate (Sohrab et al., 2018). When inserting into the cell, synthetic siRNAs are cleaved by Dicer in the cytoplasm. After placement in the RNA-Induced Silencing Complex (RISC) and Ago2, RNAi molecules (Figure 5) become single-stranded RNA and can destroy the target mRNA or inhibit its translation (Ding et al., 2018).

The Therapeutic Effect of siRNA on Coronaviruses

Synthetic siRNAs have about 21–23 bp length and perform their role by inhibiting gene expression at the post-transcription level. Unlike miRNAs, each siRNA is designed against a specific gene, so it can only impede that gene expression (Taxman et al., 2006). The siRNAs are first inserted into the cell as long double-stranded RNAs and then are cleaved by RNase III (Dicer) in the cytoplasm to become small dsRNAs of approximately twenty-one base pairs. This dsRNA later enters the RISC and converts to single-stranded RNA (ssRNA). If RISC and siRNAs complex could find the specific target site on mRNA, it could cleave the mRNA, and cellular exonucleases could invade to destroy the target mRNA (Wu and Chan, 2006). Studies have shown that this type of RNAi has the potential to act as antiviral agent to reduce replication and infection of many viruses such as HIV, Flock House Virus (FHV), Hepatitis C Virus (HCV), and Hepatitis B Virus (HBV) (Hamasaki et al., 2003; Liu et al., 2017; Shahid et al., 2017; Taning et al., 2018). Previous studies have shown that siRNA could target different parts of the virus genome to reduce the replication and infection of SARS-CoV (Figure 6A). For example, targeting the nsp1 gene (from nucleotides 250–786) by siRNA is one of the best ways to control SARS-CoV. Because targeting this area of the genome inhibits virus propagation, pathogenesis, and replication in Vero E6 cells (Ni et al., 2005). Another strategy that can help to suppress infection and replication of SARS-CoV is inhibiting the virus S protein or its specific receptor ACE2.

The SARS-CoV Spike protein is an essential viral surface glycoprotein for identifying target cells and interacting with ACE2 host cell receptors (Gallagher and Buchmeier, 2001). A study by Zhang et al. reveals that inhibiting expression of S protein using specific siRNAs could reduce the viral titers, infection, and replication of SARS-CoV in Vero E6 and 293T cells (Zhang et al., 2004). RNAi technology is also a useful instrument to suppress ACE2 expression at the host cells' surface to counteract SARS-CoV infection. In addition to the lungs, ACE2 is expressed on the surface of the bronchial, renal, duodenum, colon, gastrointestinal, and cardiovascular cells (Donoghue et al., 2000). Inhibition of the ACE2 protein in Vero E6 cell lines using siRNA could reduce replication, copies number, and infection of severe acute respiratory syndrome-associated coronavirus (Lu et al., 2008). Li et al. investigated the effect of siSC5 (nsp12 region) and siSC2 (spike protein) against SARS-CoV on *in vitro* and *in vivo* models. Their results showed

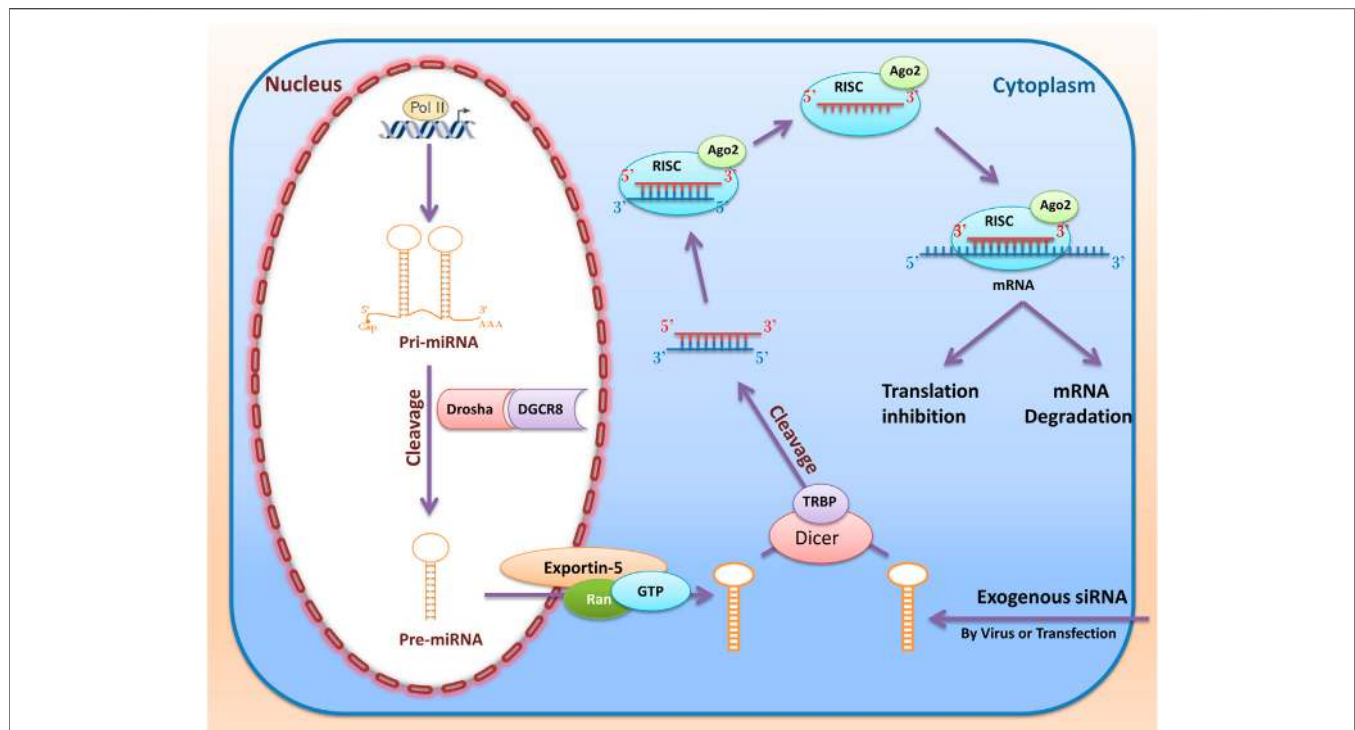


FIGURE 5 | RNA interference (miRNAs and siRNAs) biogenesis and function. The miRNAs first transcribed from the nucleus genome as pri-miRNA. Then pri-miRNA cleavage with Drosha and DGCR8 and converted to pre-miRNA. Then, RanGTP and exportin 5 cause the pre miRNA to be transported from the nucleus to the cytoplasm and cleavage by Dicer and TRBP. Ultimately, after entering the RISC complex, mature miRNA can be attached to the target mRNA and perform their function by destroying mRNA or inhibiting the translation. When inserting into the cell, synthetic siRNAs are cleaved by Dicer in the cytoplasm. This dsRNA enters RISC, and if cold finds the target mRNA, the mRNA is cleaved by the RISC and Ago2.

that these two siRNAs by inhibited virus replication in FRHK-4 cells could reduce the virus infection's effects and symptoms without having any toxicity effect on *Rhesus macaque* (dosages of 10–40 mg/kg) (Li et al., 2005a).

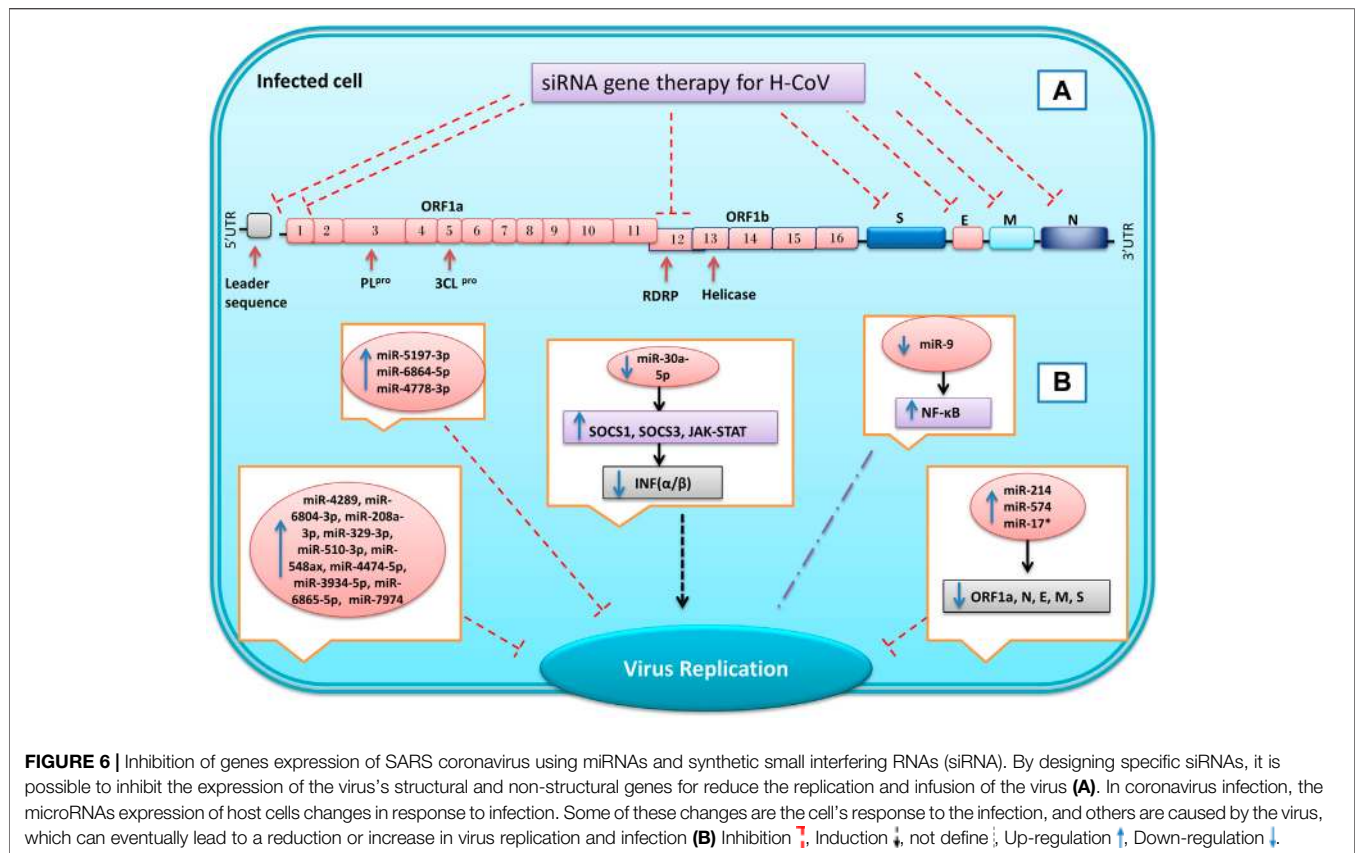
The spike protein structure determines the type of host cells which could be infected by a virus. However, variations of S protein are seen among the various strains of coronavirus. Therefore, one of the essential strategies for controlling viral infections targets the conserved genome areas between various coronaviruses (Kuo et al., 2000). Hence, Wang et al. tried to use two specific siRNAs against the conserved sequence of the SARS-CoV. The study results showed that by targeting two regions (14,450–14,468 and 15,877–15,895), encoding RNA polymerase could inhibit expression of RNA polymerase, N protein, 3CL^{Pro} and also reduce replication and cytopathic effect of the virus (Wang et al., 2004). Among the two COVID-19 proteases, the nsp3 sequence encoding papain-like protease is less conserved, while the nsp5 sequence encoding 3CL^{Pro} is highly conserved and can be selected as a potential target of siRNA for COVID-19 treatment (Liu et al., 2020).

The RdRP gene encodes a key enzyme for the replication of the virus. In SARS-CoV-2, this gene is located in ORF1b by 645 bp long and has been reported to be highly conserved, so it has the potential to target several siRNAs. Also, investigations of the RdRP sequence have shown that this gene has no genetic similarity to human genes and other coronaviruses (Wu et al.,

2020). Lu et al.'s investigation shows using specific siRNA can reduce SARS-CoV RdRP expression by more than 90% in HeLa and 239T cell lines, leading to inhibition of plaque formation in Vero E6 cells. Therefore, its suppression could be considered a suitable therapeutic target in COVID-19 patients (Lu et al., 2004). The siRNAs could target other essential coronavirus genes. For example, by designing three different siRNAs, Shi et al. were able to reduce the expression of N, E, and M genes of SARS-CoV in Vero E6 cells (Yi et al., 2005). Furthermore, the replication of SARS-CoV could be inhibited by targeting the leader sequence using specific siRNA in Vero E6 cells (Li et al., 2005b).

Although previous studies have shown that RNAi has antiviral potential, it appears that viruses can use these molecules to their advantage. MERS-CoV could infect both human and bat cell lines. Nevertheless, its pathogenic power and replication are varied between the bat (*Eptesicus fuscus*) cells and human (A549, MRC5, and Huh7) cell lines. In human cells, MERS-CoV shut-down interferon antiviral responses in the innate immunity system, unlike in bat cells by inhibition of IRF3 (a critical activator for INF β expression) (Banerjee et al., 2019).

Complemented palindromic small RNAs (cpsRNAs) are a group of small RNAs produced by mammalian and invertebrate viruses. There are sequences in the SARS-CoV (ORF3b) genome that have the origin of a cpsRNA called SARS-CoV-cpsR-19. The apoptosis assay events indicate that SARS-CoV-cpsR-19 could induce apoptosis in HeLa cells by



increasing the caspase 3 and BAX/BCL2 ratio and may play an essential role in SARS-CoV pathogenesis (Liu et al., 2018). It has been shown that some viruses (such as the Ebola virus and influenza A) could preserve themselves from RNAi-based immune systems and facilitate their replication by using Viral Suppressors of RNA silencing (VSR). Studies of Cui et al. have shown a short hairpin RNA (a novel VSR) in the SARS-CoV nucleocapsid protein sequence that defeats RNAi-triggered suppression (Cui et al., 2015). Moreover, N protein overexpression in Neuro-2a cells efficiently inhibits Dicer-mediated dsRNA cleavage and could increase replication and titration of MHV-A59 (a close relative to SARS-CoV in Coronaviridae family) viruses. The SARS-CoV-2 (N) nucleocapsid protein can also act as an escape agent from the immune system and contribute to its pathogenicity. N proteins have high homology (94%) of the amino acid sequences among coronaviruses. A recent study has shown that the N protein of SARS-CoV-2 has VSR activity that can antagonize RNAi in both effectors (recognition and cleavage of viral dsRNA by Dicer) and initiation (siRNA biogenesis) steps (Mu et al., 2020).

MicroRNAs

The miRNAs are a group of small non-coding RNAs with twenty-two nucleotides of length. These molecules are first transcribed from the nucleus genome (exons, introns, and intergenic regions) as pri-miRNA with a length of two hundred nucleotides to several thousand nucleotides (Booton and Lindsay, 2014). It is

noteworthy that each pri-miRNA can be a precursor to several mature miRNAs. Then, first processing of pri-miRNA starts with RNase III (Drosha) and its cofactor (DGCR) to form pre-miRNA, a hairpin with a length of about sixty nucleotides. In the next step, RanGTP and exportin 5 cause the pre-miRNA to be transported from the nucleus to the cytoplasm. Dicer and TRBP performed the second processing in the cytoplasm to create a double-stranded RNA molecule with a length of about twenty-two nucleotides (Bartel, 2004). Ultimately, after entering the RISC complex, one of the strings (passenger strand) is destroyed, and mature miRNA (guide strand) can be attached to the target mRNA. MiRNAs perform their function by destroying mRNA or inhibiting the translation (Vazquez, 2006). Previous studies have shown that a miRNA alone can regulate the expression of several different genes by its function. Meanwhile, various miRNAs can simultaneously control the expression of one mRNA. However, about 60% of human genes can be regulated by these molecules (Kalhori et al., 2020). For the first time, Lecellier et al. (2005) reported that cellular miR-32 could reduce virus replication of the primate foamy virus (PFV-1) by targeting the viral RNA genome (Lecellier et al., 2005).

MicroRNA Regulates the Innate Immune System, Virus Replication, and Pathogenesis of Coronavirus

The innate immune system is the body's first defense against viruses and bacteria. This system's principal cells include macrophages, dendritic cells, natural killer cells, monocytes, and granulocytes

(Leon-Icaza et al., 2019). Viruses, to increase replication, suppress the host's innate immune system via reducing INF α/β production. For example, Japanese Encephalitis Virus (JEV), Dengue Virus (DENV), and Enterovirus 71 (EV71) are able to inhibit the overexpression of INF α/β in response to viral infection by enhancing the expression of miR-146a in infected cells (Wu et al., 2013; Ho et al., 2014; Sharma et al., 2015). Viruses can also decrease the innate immune system by inhibiting the expression or function of some miRNAs. For instance, in oligodendrogloma cells, the Borna Disease Virus (BDV) can inhibit the expression of miR-155 by its specific phosphoprotein, thus inhibiting INF α/β overexpression in response to viral infection and reducing the innate immune system (Zhai et al., 2013). Therefore, one of the main antiviral components of the intrinsic immune system is type I interferons (INF α/β).

Coronaviruses can prevent the induction of the immune system in response to viral infection with different strategies. *In vivo* and *in vitro* studies of Ma et al. on the Transmissible Gastroenteritis Virus (TGEV), a member of the *alpha-coronavirus* family, showed that this virus could downregulate miR-30a-5p expression. Furthermore, they found that virus replication was facilitated by reducing IFN-I signaling cascades via removing the inhibitory effect of miR-30a-5p on INF negative regulators (such as SOCS1, SOCS3, and JAK-STAT) (Ma et al., 2018). Therefore, the overexpression of miRNAs in infected cells could increase the innate immune system and may be considered as a therapeutic approach for treatment.

Although it has previously been reported that miRNAs play a significant role in regulating the eukaryotic gene, subsequent studies showed that these nano molecules can also alter the virus's replication to increase or decrease its infection (**Figure 6B**). For example, miR-122 plays an essential role in the pathogenesis and replication of the Hepatitis C virus. The miR-122, to increase the virus's stability and replication bind to the 5' non-translated regions (NTRs) of the virus and repress RNA degradation via exonucleases. Therefore, the knockout of this miRNA in Huh-7 cells could reduce HCV replication (Jopling et al., 2005). In contrast, miR-32 could negatively alter the replication of the PFV-1 by targeting viral genes in the human HEK-293T cell line (Lecellier et al., 2005).

Some viruses have sequences (hairpin) in their genomes similar to miRNAs and can regulate the gene expression of the host cell or virus (Grundhoff and Sullivan, 2011; Kincaid and Sullivan, 2012). A computational approach study by Hassan et al. showed there are several hairpins in the genome of the MERS that could act as a precursor for thirteen miRNAs, which were significantly similar to human miRNAs. Their study showed ten miRNAs (miR-4289, miR-6804-3p, miR-208a-3p, miR-329-3p, miR-510-3p, miR-548ax, miR-4474-5p, miR-3934-5p, miR-6865-5p, and miR-7974) of these, miRNAs do not have any known specific biological function in humans or animals at all. Nevertheless, miR-18a, miR-628, and miR-342-3p had a biological role in humans related to Basal Cell Carcinoma (BCC) of the skin, malignant glioblastoma, and late-stage prion disease, respectively (Hasan et al., 2014). Numerous studies and reports suggest that some miRNAs have antiviral activity and can be used against influenza, HIV, HBV, and

poliovirus (PV) (Sanghvi and Steel, 2012; Zhang et al., 2013; Shim et al., 2016; Hamada-Tsutsumi et al., 2019). On the other hand, viruses could alter the gene and miRNA expression profile in the host cell. For example, miR-146a and miR-130b upregulated by the human T Cell Leukemia Virus (HTLV-1) in PBMC cells (Bouzar and Willems, 2008).

Nucleocapsid (N) protein is a structural protein that has the same function in all coronaviruses. The human coronavirus CoV-OC43 could inhibit miR-9 function by its N protein and increase NF- κ B expression in 253T cells. However, it is unclear whether upregulation NF- κ B is a suitable response for virus replication or a secondary inhibitor for virus replication (Lai et al., 2014). Infection of bronchoalveolar stem cells (BASICs) by SARS-CoV reveals that this virus could upregulate the expression of miR-574-5p, miR-214, and miRNAs-17* 2–4 fold. Moreover, overexpression of these miRNAs could repress SARS-CoV replication by targeting the four viral structure proteins (E, S, M, N), and orf1a (Mallick et al., 2009). Unfortunately, to date, not many *in vivo* and *in vitro* studies have been performed on RNAi's role in inhibiting the COVID-19, and most studies have been performed base on bioinformatics and *in silico* studies. Qingfei Paidu decoction (QFPD) contains twenty-one traditional Chinese medicines that have been used to treat COVID-19 since February 7, 2020. Chen et al.'s molecular docking study revealed that QFPD can bind to structural and non-structural proteins of COVID-19. They also found that miR-183 and miR-130A/B/301 predict targets of QFPD, and QFPD by these microRNAs may exert anti-SARS-CoV-2 activity (Chen et al., 2020a). In a bioinformatics approach study performed by Khan et al., it was found that several miRNAs can have antiviral properties in infections caused by SARS-CoV-1 and SARS-CoV-2. For example, evidence has shown miR-323a-5p, miR-622, miR-198, and miR-654-5p for SARS-CoV-1 and miR-323a-5p, miR-20b-5p, miR-17-5p for SARS-CoV-2 have antiviral roles by targeting the ORF1ab and the S region (Khan et al., 2020).

Therefore, it is likely miRNA with low side effects can be used as a therapeutic agent for the COVID-19 treatment. Differences in miRNA expression profiles in individuals are probably one reason why COVID-19 causes death in some people and causes only brief symptoms in others. As a result, microRNAs have recently emerged as a critical factor in increasing or inhibiting the potential of viral infection. We hope that clinical and preclinical research can use them in gene therapy as antiviral agents soon.

CONCLUSION

Today, the whole world is suffering from a pandemic disease called COVID-19, which has caused deaths in many developed and developing countries. Despite all the advances in human medicine, we have not yet been able to find a suitable treatment for this viral disease. The use of molecular or pharmacological methods to control infection or virus replication requires identifying essential genes involved in infection and replication of the virus. Two strategies are suggested to treat this disease. The first step is to reduce the virus's infection by preventing the virus from attaching to its specific receptor. The next step is to reduce the

virus's replication by inhibiting the virus's structural and non-structural genes. Medicinal plants and natural products are a good option for preventing and treating viral infections, especially COVID-19, due to their lower cost, lower side effects, and natural origin compared with chemical drugs. These compounds could increase efficiency and strengthen the host immune system against many infections and diseases due to their inherent properties. In the treatment of COVID-19, these compounds can reduce virus infection or replication by repressing the virus's coupling to the host cell's receptors or by inhibiting the expression of structural and non-structural genes. Moreover, RNAi (siRNA and miRNA) could inhibit viral infections, especially COVID-19, by inhibiting essential virus genes or inducing a host immune system. Therefore, the simultaneous use of natural compounds and RNAi can play a critical role in the treatment of SARS-CoV-2 and restraining this pandemic pneumonia.

REFERENCES

- Ahn, D.-G., Choi, J.-K., Taylor, D. R., and Oh, J.-W. (2012). Biochemical characterization of a recombinant SARS coronavirus nsp12 RNA-dependent RNA polymerase capable of copying viral RNA templates. *Arch. Virol.* 57, 2095–2104. doi:10.1007/s00705-012-6751404-x10.1007/s00705-012-1404-x
- An, S., Chen, C.-J., Yu, X., Leibowitz, J. L., and Makino, S. (1999). Induction of apoptosis in murine coronavirus-infected cultured cells and demonstration of E protein as an apoptosis inducer. *J. Virol.* 73, 7853–7859. doi:10.1128/JVI.73.9.7853-7859.1999
- Angelini, M. M., Akhlaghpour, M., Neuman, B. W., and Buchmeier, M. J. (2013). Severe acute respiratory syndrome coronavirus nonstructural proteins 3, 4, and 6 induce double-membrane vesicles. *MBio.* 4, e00524–e00513. doi:10.1128/mBio.00524-13
- Angelini, M. M., Neuman, B. W., and Buchmeier, M. J. (2014). Untangling membrane rearrangement in the nidovirales. *DNA Cell Biol.* 33, 122–127. doi:10.1089/dna.2013.2304
- Banerjee, A., Falzarano, D., Rapin, N., Lew, J., and Misra, V. (2019). Interferon regulatory factor 3-mediated signaling limits Middle-East respiratory syndrome (MERS) coronavirus propagation in cells from an insectivorous bat. *Viruses* 11, 152. doi:10.3390/v11020152
- Bartel, D. P. (2004). MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 116, 281–297. doi:10.1016/s0092-8674(04)00045-5
- Booton, R., and Lindsay, M. A. (2014). Emerging role of MicroRNAs and long noncoding RNAs in respiratory disease. *Chest* 146, 193–204. doi:10.1378/chest.13-2736
- Bosch, B. J., Van Der Zee, R., De Haan, C. A., and Rottier, P. J. (2003). The coronavirus spike protein is a class I virus fusion core protein: structural and functional characterization of the fusion core complex. *J. Virol.* 77, 8801–8811. doi:10.1128/jvi.77.16.8801-8811.2003
- Bouvet, M., Debarnot, C., Imbert, I., Selisko, B., Snijder, E. J., Canard, B., et al. (2010). *In vitro* reconstitution of SARS-coronavirus mRNA cap methylation. *PLoS Pathog.* 6 (4), e1000863. doi:10.1371/journal.ppat.1000863
- Bouvet, M., Imbert, I., Subissi, L., Gluais, L., Canard, B., and Decroly, E. (2012). RNA 3'-end mismatch excision by the severe acute respiratory syndrome coronavirus nonstructural protein nsp10/nsp14 exoribonuclease complex. *Proc. Natl. Acad. Sci USA* 109, 9372–9377. doi:10.1073/pnas.1201130109
- Bouzar, A. B., and Willems, L. (2008). How HTLV-1 may subvert miRNAs for persistence and transformation. *Retrovirology* 5, 101. doi:10.1186/1742-4690-5-101
- Cai, Q., Yang, M., Liu, D., Chen, J., Shu, D., Xia, J., et al. (2020). Experimental treatment with Favipiravir for COVID-19: an open-label control study. *Engineering* 6 (10), 1192–1198. doi:10.1016/j.eng.2020.03.007

AUTHOR CONTRIBUTIONS

The study was conceptualized by MS and EA. The methodology was given by EA and MF. Writing and original draft preparation was done by MK and FS. Writing, review, and editing were done by MF, IA, and JE. Funding acquisition was provided by MK and JE. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by the Kermanshah University of Medical Sciences (grant numbers 990512). JE gratefully acknowledges funding from CONICYT (PAI/ACADEMIA No 79160109).

- Casals, J., Tignor, G. H., Lack, E. E., Jenson, B., Smith, H. G., Healy, G. B., et al. (1980). International committee for Taxonomy of viruses. *Intervirology* 14, 228. doi:10.1159/000149188
- Casella, M., Rajnik, M., Cuomo, A., Dulebohn, S. C., and Di Napoli, R. (2020). “Features, evaluation and treatment coronavirus (COVID-19),” in *Statpearls* (Treasure Island, FL: StatPearls Publishing).
- Cavanagh, D. (1995). “The coronavirus surface glycoprotein,” in *The coronaviridae*. Boston, MA: Springer, 73–113. doi:10.1007/978-1-4899-1531-3_5
- Cawood, R., Harrison, S. M., Dove, B. K., Reed, M. L., and Hiscox, J. A. (2007). Cell cycle dependent nucleolar localization of the coronavirus nucleocapsid protein. *Cell Cycle* 6, 863–867. doi:10.4161/cc.6.7.4032
- Centers for Disease Control and Prevention (2012). Severe respiratory illness associated with a novel coronavirus--Saudi Arabia and Qatar, 2012. *MMWR Morb. Mortal. Wkly. Rep.* 61 (40), 820.
- Chen, J., Wang, Y. K., Gao, Y., Hu, L. S., Yang, J. W., Wang, J. R., et al. (2020a). Protection against COVID-19 injury by qingfei paidu decoction via anti-viral, anti-inflammatory activity and metabolic programming. *Biomed. Pharmacother.* 129, 110281. doi:10.1016/j.biopha.2020.110281
- Chen, Z. M., Fu, J. F., Shu, Q., Chen, Y. H., Hua, C. Z., Li, F. B., et al. (2020b). Diagnosis and treatment recommendations for pediatric respiratory infection caused by the 2019 novel coronavirus. *World J. Pediatr.* 16, 240–246. doi:10.1007/s12519-020-00345-5
- Chikhale, R. V., Gupta, V. K., Eldesoky, G. E., Wabaidur, S. M., Patil, S. A., and Islam, M. A. (2020). Identification of potential anti-TMPRSS2 natural products through homology modelling, virtual screening and molecular dynamics simulation studies. *J. Biomol. Struct. Dyn.* 1–16, 1–16. doi:10.1080/07391102.2020.1798813
- Chu, C., Cheng, V., Hung, I., Wong, M., Chan, K., Chan, K., et al. (2004). Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. *Thorax* 59, 252–256. doi:10.1136/thorax.2003.012658
- Colson, P., Rolain, J.-M., and Raoult, D. (2020). Chloroquine for the 2019 novel coronavirus SARS-CoV-2. *Int. J. Antimicrob. Agents* 55 (3), 105923. doi:10.1016/j.ijantimicag.2020.105923
- Cornillez-Ty, C. T., Liao, L., Yates, J. R., Kuhn, P., and Buchmeier, M. J. (2009). Severe acute respiratory syndrome coronavirus nonstructural protein 2 interacts with a host protein complex involved in mitochondrial biogenesis and intracellular signaling. *J. Virol.* 83, 10314–10318. doi:10.1128/JVI.00842-09
- COVID-19 Treatment Guidelines Panel (2019). *Coronavirus disease 2019 (COVID-19) treatment guidelines*. Bethesda, MD: National Institutes of Health. Available at: <https://www.covid19treatmentguidelines.nih.gov/>
- Cui, L., Wang, H., Ji, Y., Yang, J., Xu, S., Huang, X., et al. (2015). The nucleocapsid protein of coronaviruses acts as a viral suppressor of RNA silencing in mammalian cells. *J. virol.* 89, 9029–9043. doi:10.1128/jvi.01331-15
- Danesh, A., Cameron, C. M., León, A. J., Ran, L., Xu, L., Fang, Y., et al. (2011). Early gene expression events in ferrets in response to SARS coronavirus infection

- versus direct interferon- α 2b stimulation. *Virology* 409, 102–112. doi:10.1016/j.virol.2010.10.002
- da Silva Antonio, A., Wiedemann, L. S. M., and Veiga-Junior, V. F. (2020). Natural products' role against COVID-19. *RSC Adv.* 10, 23379–23393. doi:10.1039/D0RA03774E
- De Lang, A., Osterhaus, A. D., and Haagmans, B. L. (2006). Interferon-gamma and interleukin-4 downregulate expression of the SARS coronavirus receptor ACE2 in Vero E6 cells. *Virology* 353, 474–481. doi:10.1016/j.virol.2006.06.011
- De Leo, A., Arena, G., Lacanna, E., Oliviero, G., Colavita, F., and Mattia, E. (2012). Resveratrol inhibits Epstein Barr Virus lytic cycle in Burkitt's lymphoma cells by affecting multiple molecular targets. *Antiviral Res.* 96, 196–202. doi:10.1016/j.antiviral.2012.09.003
- Deng, L., Li, C., Zeng, Q., Liu, X., Li, X., Zhang, H., et al. (2020). Arbidol combined with LPV/r versus LPV/r alone against corona virus disease 2019: a retrospective cohort study. *J. Infect.* 81 (1), e1–e5. doi:10.1016/j.jinf.2020.03.002
- Denison, M. R., Graham, R. L., Donaldson, E. F., Eckerle, L. D., and Baric, R. S. (2011). Coronaviruses: an RNA proofreading machine regulates replication fidelity and diversity. *RNA Biol.* 8, 270–279. doi:10.4161/rna.8.2.15013
- Dias, D. A., Urban, S., and Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites* 2, 303–336. doi:10.3390/metabo2020303
- Ding, S. W., Han, Q., Wang, J., and Li, W. X. (2018). Antiviral RNA interference in mammals. *Curr Opin Immunol* 54, 109–114. doi:10.1016/j.coi.2018.06.010
- Ding, Y., Wang, H., Shen, H., Li, Z., Geng, J., Han, H., et al. (2003). The clinical pathology of severe acute respiratory syndrome (SARS): a report from China. *J. Pathol.* 200, 282–289. doi:10.1002/path.1440
- Donoghue, M., Hsieh, F., Baronas, E., Godbout, K., Gosselin, M., Stagliano, N., et al. (2000). A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9. *Circ Res* 87, e1–e9. doi:10.1161/01.res.87.5.e1
- Enjuanes, L., Brian, D., Cavanagh, D., Holmes, K., Lai, M., Laude, H., et al. (2000). *Coronaviridae. Virus Taxonomy, Classification and Nomenclature of viruses*. Editors M. A. Murphy, C. M. Fauquet, D. H. L. Bishop, S. A. Ghabrial, A. W. Jarvis, G. P. Martelli, et al. (New York, NY: Academic Press), 835–849.
- Faith, S. A., Sweet, T. J., Bailey, E., Booth, T., and Docherty, J. J. (2006). Resveratrol suppresses nuclear factor-kappaB in herpes simplex virus infected cells. *Antiviral Res.* 72, 242–251. doi:10.1016/j.antiviral.2006.06.011
- Falzarano, D., De Wit, E., Martellaro, C., Callison, J., Munster, V. J., and Feldmann, H. (2013). Inhibition of novel β coronavirus replication by a combination of interferon- α 2b and ribavirin. *Sci. Rep.* 3, 1686. doi:10.1038/srep01686
- Fenster, V., and Sen, G. C. (2009). Interferons and viral infections. *BioFactors* 35, 14–20. doi:10.1002/biof.6
- Filardo, S., Di Pietro, M., Mastromarino, P., and Sessa, R. (2020). Therapeutic potential of resveratrol against emerging respiratory viral infections. *Pharmacol Ther.* 214, 107613. doi:10.1016/j.pharmthera.2020.107613
- Frabasile, S., Koishi, A. C., Kuczera, D., Silveira, G. F., Verri, W. A., Jr, Dos Santos, C. N. D., et al. (2017). The citrus flavanone naringenin impairs dengue virus replication in human cells. *Sci. Rep.* 7, 41864. doi:10.1038/srep41864
- Gallagher, T. M., and Buchmeier, M. J. (2001). Coronavirus spike proteins in viral entry and pathogenesis. *Virology* 279, 371–374. doi:10.1006/viro.2000.0757
- Gentile, D., Patamia, V., Scala, A., Sciortino, M. T., Piperno, A., and Rescifina, A. (2020). Putative inhibitors of SARS-CoV-2 main protease from a library of marine natural products: a virtual screening and molecular modeling study. *Mar. Drugs* 18 (4), 225. doi:10.3390/md18040225
- Godino, C., Scotti, A., Mauerer, N., Mancini, N., Fominskiy, E., Margonato, A., et al. (2020). Antithrombotic therapy in patients with COVID-19? -rationale and evidence. *Int. J. Cardiol.* 35 (9), 2698–2706. doi:10.1016/j.ijcard.2020.09.064
- Grundhoff, A., and Sullivan, C. S. (2011). Virus-encoded microRNAs. *Virology* 411, 325–343. doi:10.1016/j.virol.2011.01.002
- Guan, W.-J., Ni, Z.-Y., Hu, Y., Liang, W.-H., Ou, C.-Q., He, J.-X., et al. (2020). Clinical characteristics of coronavirus disease 2019 in China. *N. Engl. J. Med.* 382 (18), 1708–1720. doi:10.1056/NEJMoa2002032
- Hamada-Tsutsumi, S., Naito, Y., Sato, S., Takaoka, A., Kawashima, K., Isogawa, M., et al. (2019). The antiviral effects of human microRNA miR-302c-3p against hepatitis B virus infection. *Aliment Pharmacol. Ther.* 49, 1060–1070. doi:10.1111/apt.15197
- Hamasaki, K., Nakao, K., Matsumoto, K., Ichikawa, T., Ishikawa, H., and Eguchi, K. (2003). Short interfering RNA-directed inhibition of hepatitis B virus replication. *FEBS Lett* 543, 51–54. doi:10.1016/S0014-5793(03)00400-9
- Hamre, D., and Procknow, J. J. (1966). A new virus isolated from the human respiratory tract. *Proc. Soc. Exp. Biol. Med.* 121, 190–193. doi:10.3181/00379727-121-30734
- Hasan, M. M., Akter, R., Ullah, M., Abedin, M., Ullah, G., and Hossain, M. (2014). A computational approach for predicting role of human microRNAs in MERS-CoV genome. *Adv. bioinformatics* 2014, 967946. doi:10.1155/2014/967946
- Ho, B. C., Yu, I. S., Lu, L. F., Rudensky, A., Chen, H. Y., Tsai, C. W., et al. (2014). Inhibition of miR-146a prevents enterovirus-induced death by restoring the production of type I interferon. *Nat. Commun.* 5, 3344. doi:10.1038/ncomms4344
- Ho, T.-Y., Wu, S.-L., Chen, J.-C., Li, C.-C., and Hsiang, C.-Y. (2007). Emodin blocks the SARS coronavirus spike protein and angiotensin-converting enzyme 2 interaction. *Antiviral Res.* 74, 92–101. doi:10.1016/j.antiviral.2006.04.014
- Hoffmann, M., Kleine-Weber, H., Schroeder, S., Krüger, N., Herrler, T., Erichsen, S., et al. (2020). SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. *Cell* 181, 271–280. doi:10.1016/j.cell.2020.02.052
- Holshue, M. L., DeBolt, C., Lindquist, S., Lofy, K. H., Wiesman, J., Bruce, H., et al. (2020). First case of 2019 novel coronavirus in the United States. *N. Engl. J. Med.* 382, 929–936. doi:10.1056/NEJMoa2001191
- Huang, C., Wang, Y., Li, X., Ren, L., Zhao, J., Hu, Y., et al. (2020). Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 395, 497–506. doi:10.1016/S0140-6736(20)30183-5
- Huang, I.-C., Bailey, C. C., Weyer, J. L., Radoshitzky, S. R., Becker, M. M., Chiang, J. J., et al. (2011). Distinct patterns of IFITM-mediated restriction of filoviruses, SARS coronavirus, and influenza A virus. *PLoS Pathog* 7 (1), e1001258. doi:10.1371/journal.ppat.1001258
- ICTV (2009). Virus taxonomy: 2009 release. Available at: https://talk.ictvonline.org/ictv-reports/ictv_9th_report/negative-sense-rna-viruses-2011/w/negra_viruses/209/orthomyxoviridae (Accessed November 30, 2020).
- Ivanov, K. A., Thiel, V., Dobbe, J. C., Van Der Meer, Y., Snijder, E. J., and Ziebuhr, J. (2004). Multiple enzymatic activities associated with severe acute respiratory syndrome coronavirus helicase. *J. Virol.* 78, 5619–5632. doi:10.1128/JVI.78.11.5619-5632.2004
- Izhaki, I. (2002). Emodin—a secondary metabolite with multiple ecological functions in higher plants. *New Phytol.* 155, 205–217. doi:10.1046/j.1469-8137.2002.00459.x
- Jang, M., Park, Y.-I., Cha, Y.-E., Park, R., Namkoong, S., Lee, J. I., et al. (2020). Tea polyphenols EGCG and theaflavin inhibit the activity of SARS-CoV-2 3CL-protease in vitro. *Evid. Based Complement. Alternat. Med.* 2020, 5630838. doi:10.1155/2020/5630838
- Ji, H. F., Li, X. J., and Zhang, H. Y. (2009). Natural products and drug discovery. Can thousands of years of ancient medical knowledge lead us to new and powerful drug combinations in the fight against cancer and dementia? *EMBO Rep.* 10, 194–200. doi:10.1038/embor.2009.12
- Jo, S., Kim, H., Kim, S., Shin, D. H., and Kim, M. S. (2019). Characteristics of flavonoids as potent MERS-CoV 3C-like protease inhibitors. *Chem. Biol. Drug Des.* 94, 2023–2030. doi:10.1111/cbdd.13604
- Jo, S., Kim, S., Kim, D. Y., Kim, M.-S., and Shin, D. H. (2020a). Flavonoids with inhibitory activity against SARS-CoV-2 3CLpro. *J. Enzyme Inhib. Med. Chem.* 35, 1539–1544. doi:10.1080/14756366.2019.169048010.1080/14756366.2020.1801672
- Jo, S., Kim, S., Shin, D. H., and Kim, M.-S. (2020b). Inhibition of SARS-CoV 3CL protease by flavonoids. *J. Enzyme Inhib. Med. Chem.* 35, 145–151. doi:10.1080/14756366.2019.1690480
- Jopling, C. L., Yi, M., Lancaster, A. M., Lemon, S. M., and Sarnow, P. (2005). Modulation of hepatitis C virus RNA abundance by a liver-specific microRNA. *Science* 309, 1577–1581. doi:10.1126/science.1113329
- Kalhori, M. R., Arefian, E., Atanaki, F. F., Kavousi, K., and Soleimani, M. (2020). miR-548x and miR-4698 controlled cell proliferation by affecting the PI3K/AKT signaling pathway in glioblastoma cell lines. *Sci. Rep.* 10, 1–12. doi:10.1038/s41598-020-57588-5
- Keum, Y.-S., Lee, J. M., Yu, M.-S., Chin, Y.-W., and Jeong, Y.-J. (2013). Inhibition of SARS coronavirus helicase by baicalein. *Bull Korean Chem. Soc.* 34, 3187–3188. doi:10.5012/bkcs.2013.34.11.3187

- Khalid, M., Al Rabiah, F., Khan, B., Al Mobeireek, A., Butt, T. S., and Al Mutairy, E. (2015). Ribavirin and interferon- α 2b as primary and preventive treatment for Middle East respiratory syndrome coronavirus: a preliminary report of two cases. *Antivir. Ther. (Lond)* 20, 87–91. doi:10.3851/IMP2792
- Khan, M. A., Sany, M. R. U., Islam, M. S., and Islam, A. (2020). Epigenetic regulator miRNA pattern differences among SARS-CoV, SARS-CoV-2, and SARS-CoV-2 World-Wide isolates delineated the mystery behind the epic pathogenicity and distinct clinical characteristics of pandemic COVID-19. *Front. Genet.* 11, 765. doi:10.3389/fgene.2020.00765
- Kim, D. E., Min, J. S., Jang, M. S., Lee, J. Y., Shin, Y. S., Park, C. M., et al. (2019). Natural bis-benzylisoquinoline alkaloids-tetrandrine, fangchinoline, and cepharanthine, inhibit human coronavirus OC43 infection of MRC-5 human lung cells. *Biomolecules* 9, 696. doi:10.3390/biom9110696
- Kim, D. W., Seo, K. H., Curtis-Long, M. J., Oh, K. Y., Oh, J.-W., et al. (2014). Phenolic phytochemical displaying SARS-CoV papain-like protease inhibition from the seeds of *Psoralea corylifolia*. *J. Enzyme Inhib. Med. Chem.* 29, 59–63. doi:10.1371/journal.ppat.100301810.3109/14756366.2012.753591
- Kincaid, R. P., and Sullivan, C. S. (2012). Virus-encoded microRNAs: an overview and a look to the future. *PLoS Pathog.* 8, e1003018. doi:10.1371/journal.ppat.1003018
- King, A. M., Adams, M. J., Carstens, E. B., and Lefkowitz, E. J. (2012). "Virus taxonomy," in *Ninth report of the international committee on Taxonomy of viruses* (London, United Kingdom: Elsevier Academic Press), 486–487.
- Kuo, L., Godeke, G.-J., Raamsman, M. J., Masters, P. S., and Rottier, P. J. (2000). Retargeting of coronavirus by substitution of the spike glycoprotein ectodomain: crossing the host cell species barrier. *J. Virol.* 74, 1393–1406. doi:10.1128/JVI.74.3.1393-1406.2000
- Lai, F. W., Stephenson, K. B., Mahony, J., and Lichty, B. D. (2014). Human coronavirus OC43 nucleocapsid protein binds microRNA 9 and potentiates NF- κ B activation. *J. Virol.* 88, 54–65. doi:10.1128/JVI.02678-13
- Lai, M. M., and Cavanagh, D. (1997). The molecular biology of coronaviruses. *Adv. Virus Res.* 48, 1–100. doi:10.1016/S0065-3527(08)60286-9
- Lai, M. M. (1990). Coronavirus: organization, replication and expression of genome. *Annu. Rev. Microbiol.* 44, 303. doi:10.1146/annurev.mi.44.100190.001511
- Lecellier, C. H., Dunoyer, P., Arar, K., Lehmann-Che, J., Eyquem, S., Himber, C., et al. (2005). A cellular microRNA mediates antiviral defense in human cells. *Science* 308, 557–560. doi:10.1126/science.1108784
- Lei, J., Kusov, Y., and Hilgenfeld, R. (2018). Nsp3 of coronaviruses: structures and functions of a large multi-domain protein. *Antiviral Res.* 149, 58–74. doi:10.1016/j.antiviral.2017.11.001
- Leon-Icaza, S. A., Zeng, M., and Rosas-Taraco, A. G. (2019). MicroRNAs in viral acute respiratory infections: immune regulation, biomarkers, therapy, and vaccines. *ExRNA* 1 (1), 1. doi:10.1186/s41544-018-0004-7
- Li, B.-J., Tang, Q., Cheng, D., Qin, C., Xie, F. Y., Wei, Q., et al. (2005a). Using siRNA in prophylactic and therapeutic regimens against SARS coronavirus in Rhesus macaque. *Nat. Med.* 11, 944–951. doi:10.1038/nm1280
- Li, F. (2016). Structure, function, and evolution of coronavirus spike proteins. *Annu. Rev. Virol.* 3, 237–261. doi:10.1146/annurev-virology-110615-042301
- Li, G., and De Clercq, E. (2020). Therapeutic options for the 2019 novel coronavirus (2019-nCoV). *Nat. Rev. Drug. Discov.* 19, 149–150. doi:10.1038/d41573-020-00016-0
- Li, T., Zhang, Y., Fu, L., Yu, C., Li, X., Li, Y., et al. (2005b). SiRNA targeting the leader sequence of SARS-CoV inhibits virus replication. *Gene Ther.* 12, 751–761. doi:10.1038/sj.gt.3302479
- Liang, X., and Li, Z.-Y. (2010). Ion channels as antiviral targets. *Virol. Sin.* 25, 267–280. doi:10.1007/s12250-010-3136-y
- Lin, C.-W., Tsai, F.-J., Wan, L., Lai, C.-C., Lin, K.-H., Hsieh, T.-H., et al. (2005). Binding interaction of SARS coronavirus 3CL(pro) protease with vacuolar-H⁺ ATPase G1 subunit. *FEBS Lett.* 579, 6089–6094. doi:10.1016/j.febslet.2005.09.075
- Lin, S.-C., Ho, C.-T., Chuo, W.-H., Li, S., Wang, T. T., and Lin, C.-C. (2017). Effective inhibition of MERS-CoV infection by resveratrol. *BMC Infect. Dis.* 17, 144. doi:10.1186/s12879-017-2253-8
- Liu, C., Chen, Z., Hu, Y., Ji, H., Yu, D., Shen, W., et al. (2018). Complemented palindromic small RNAs first discovered from SARS coronavirus. *Genes* 9, 442. doi:10.3390/genes9090442
- Liu, C., Liang, Z., and Kong, X. (2017). Efficacy analysis of combinatorial siRNAs against HIV derived from one double hairpin RNA precursor. *Front. Microbiol.* 8, 1651. doi:10.3389/fmicb.2017.01651
- Liu, C., Zhou, Q., Li, Y., Garner, L. V., Watkins, S. P., Carter, L. J., et al. (2020). Research and development on therapeutic agents and vaccines for COVID-19 and related human coronavirus diseases. *ACS Cent. Sci.* 6, 315–331. doi:10.1021/acscentsci.0c00272
- Liu, D., Yuan, Q., and Liao, Y. (2007). Coronavirus envelope protein: a small membrane protein with multiple functions. *Cell Mol. Life Sci.* 64, 2043–2048. doi:10.1007/s00018-007-7103-1
- Lu, A., Zhang, H., Zhang, X., Wang, H., Hu, Q., Shen, L., et al. (2004). Attenuation of SARS coronavirus by a short hairpin RNA expression plasmid targeting RNA-dependent RNA polymerase. *Virology* 324, 84–89. doi:10.1016/j.virol.2004.03.031
- Lu, C.-Y., Huang, H.-Y., Yang, T.-H., Chang, L.-Y., Lee, C.-Y., and Huang, L.-M. (2008). siRNA silencing of angiotensin-converting enzyme 2 reduced severe acute respiratory syndrome-associated coronavirus replications in Vero E6 cells. *Eur. J. Clin. Microbiol. Infect. Dis.* 27, 709–715. doi:10.1007/s10096-008-0495-5
- Ma, Y., Wang, C., Xue, M., Fu, F., Zhang, X., Li, L., et al. (2018). The coronavirus transmissible gastroenteritis virus evades the type I interferon response through IRE1 α -mediated manipulation of the microRNA miR-30a-5p/SOCS1/3 axis. *J. Virol.* 92, e00728–e00718. doi:10.1128/JVI.00728-18
- Mahase, E. (2020). Coronavirus covid-19 has killed more people than SARS and MERS combined, despite lower case fatality rate. *BMJ* 368, m641. doi:10.1136/bmj.m641
- Mallick, B., Ghosh, Z., and Chakrabarti, J. (2009). MicroRNome analysis unravels the molecular basis of SARS infection in bronchoalveolar stem cells. *PLoS One* 4 (11), e7837. doi:10.1371/journal.pone.0007837
- Maurya, V. K., Kumar, S., Prasad, A. K., Bhatt, M. L. B., and Saxena, S. K. (2020). Structure-based drug designing for potential antiviral activity of selected natural products from Ayurveda against SARS-CoV-2 spike glycoprotein and its cellular receptor. *Virusdisease* 31, 179–193. doi:10.1007/s13337-020-00598-8
- Mcintosh, K., Kapikian, A. Z., Turner, H. C., Hartley, J. W., Parrott, R. H., and Chanock, R. M. (1970). Seroepidemiologic studies of coronavirus infection in adults and children. *Am. J. Epidemiol.* 91, 585–592. doi:10.1093/oxfordjournals.aje.a121171
- Mcintosh, K., and Peiris, J. (2009). "Coronaviruses," in *Clinical virology*. 3rd Edn. (Washington, DC: American Society of Microbiology Press), 1155–1171. doi:10.1128/9781555815981.ch51
- Miknis, Z. J., Donaldson, E. F., Umland, T. C., Rimmer, R. A., Baric, R. S., and Schultz, L. W. (2009). Severe acute respiratory syndrome coronavirus nsp9 dimerization is essential for efficient viral growth. *J. Virol.* 83, 3007–3018. doi:10.1128/JVI.01505-08
- Monto, A. S. (1974). Medical reviews. Coronaviruses. *Yale J. Biol. Med.* 47 (4), 234–251.
- Moore, M. J., Dorffman, T., Li, W., Wong, S. K., Li, Y., Kuhn, J. H., et al. (2004). Retroviruses pseudotyped with the severe acute respiratory syndrome coronavirus spike protein efficiently infect cells expressing angiotensin-converting enzyme 2. *J. Virol.* 78, 10628–10635. doi:10.1128/JVI.78.19.10628-10635.2004
- Mu, J., Xu, J., Zhang, L., Shu, T., Wu, D., Huang, M., et al. (2020). SARS-CoV-2-encoded nucleocapsid protein acts as a viral suppressor of RNA interference in cells. *Sci. China Life Sci.* 63, 1–4. doi:10.1007/s11427-020-1692-1
- Muramatsu, T., Takemoto, C., Kim, Y.-T., Wang, H., Nishii, W., Terada, T., et al. (2016). SARS-CoV 3CL protease cleaves its C-terminal autoprocessing site by novel subsite cooperativity. *Proc. Natl. Acad. Sci. U.S.A.* 113, 12997–13002. doi:10.1073/pnas.1601327113
- Nal, B., Chan, C., Kien, F., Siu, L., Tse, J., Chu, K., et al. (2005). Differential maturation and subcellular localization of severe acute respiratory syndrome coronavirus surface proteins S, M and E. *J. Gen. Virol.* 86, 1423–1434. doi:10.1099/vir.0.80671-0
- Nguyen, T. T. H., Woo, H.-J., Kang, H.-K., Kim, Y.-M., Kim, D.-W., Ahn, S.-A., et al. (2012). Flavonoid-mediated inhibition of SARS coronavirus 3C-like protease expressed in *Pichia pastoris*. *Biotechnol. Lett.* 34, 831–838. doi:10.1007/s10529-011-0845-8
- Ni, B., Shi, X., Li, Y., Gao, W., Wang, X., and Wu, Y. (2005). Inhibition of replication and infection of severe acute respiratory syndrome-associated

- coronavirus with plasmid-mediated interference RNA. *Antivir. Ther. (Lond)* 10, 527–533.
- Opstelten, D. J., Raamsman, M., Wolfs, K., Horzinek, M. C., and Rottier, P. (1995). Envelope glycoprotein interactions in coronavirus assembly. *J. Cell Biol.* 131, 339–349. doi:10.1083/jcb.131.2.339
- Pafumi, I., Festa, M., Papacci, F., Lagostena, L., Giunta, C., Gutla, V., et al. (2017). Naringenin impairs two-pore channel 2 activity and inhibits VEGF-induced angiogenesis. *Sci. Rep.* 7, 1–11. doi:10.1038/s41598-017-04974-1
- Pan, B., Fang, S., Zhang, J., Pan, Y., Liu, H., Wang, Y., et al. (2020). Chinese herbal compounds against SARS-CoV-2: puerarin and quercetin impair the binding of viral S-protein to ACE2 receptor. *Comput. Struct. Biotechnol. J.* 18, 3518–3527. doi:10.1016/j.csbj.2020.11.010
- Park, J.-Y., Jeong, H. J., Kim, J. H., Kim, Y. M., Park, S.-J., Kim, D., et al. (2012a). Diarylheptanoids from *Alnus japonica* inhibit papain-like protease of severe acute respiratory syndrome coronavirus. *Biol. Pharm. Bull.* 35, 2036. doi:10.1248/bpb.12-00623
- Park, J.-Y., Kim, J. H., Kim, Y. M., Jeong, H. J., Kim, D. W., Park, K. H., et al. (2012b). Tanshinones as selective and slow-binding inhibitors for SARS-CoV cysteine proteases. *Bioorg. Med. Chem.* 20 (19), 5928–5935. doi:10.1016/j.bmc.2012.07.038
- Park, J.-Y., Ko, J.-A., Kim, D. W., Kim, Y. M., Kwon, H.-J., Jeong, H. J., et al. (2016). Chalcones isolated from *Angelica keiskei* inhibit cysteine proteases of SARS-CoV. *J. Enzyme Inhib. Med. Chem.* 31 (1), 23–30. doi:10.3109/14756366.2014.1003215
- Park, J.-Y., Yuk, H. J., Ryu, H. W., Lim, S. H., Kim, K. S., Park, K. H., et al. (2017). Evaluation of polyphenols from *Broussonetia papyrifera* as coronavirus protease inhibitors. *J. Enzyme Inhib. Med. Chem.* 32, 504–512. doi:10.1080/14756366.2016.1265519
- Peiris, J. S. M., Chu, C.-M., Cheng, V. C.-C., Chan, K., Hung, I., Poon, L. L., et al. (2003). Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet* 361, 1767–1772. doi:10.1016/S0140-6736(03)13412-5
- Prajapat, M., Sarma, P., Shekhar, N., Avti, P., Sinha, S., Kaur, H., et al. (2020). Drug targets for corona virus: a systematic review. *Indian J. Pharmacol.* 52 (1), 56–65. doi:10.4103/ijp.IJP_115_20
- Pyrck, K., Berkhout, B., and Van Der Hoek, L. (2007). Antiviral strategies against human coronaviruses. *Infect. Disord. Drug. Targets* 7, 59–66. doi:10.2174/187152607780090757
- Rahman, N., Basharat, Z., Yousuf, M., Castaldo, G., Rastrelli, L., and Khan, H. (2020). Virtual screening of natural products against type II Transmembrane serine protease (TMPRSS2), the priming agent of coronavirus 2 (SARS-CoV-2). *Molecules* 25 (10), 2271. doi:10.3390/molecules25102271
- Raj, V. S., Mou, H., Smits, S. L., Dekkers, D. H., Müller, M. A., Dijkman, R., et al. (2013). Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature* 495, 251–254. doi:10.1038/nature12005
- Ren, L.-L., Wang, Y.-M., Wu, Z.-Q., Xiang, Z.-C., Guo, L., Xu, T., et al. (2020). Identification of a novel coronavirus causing severe pneumonia in human: a descriptive study. *Chin. Med. J. (Engl)* 133 (9), 1015–1024. doi:10.1097/CM9.0000000000000722
- Ricagno, S., Egloff, M.-P., Ulferts, R., Coutard, B., Nurizzo, D., Campanacci, V., et al. (2006). Crystal structure and mechanistic determinants of SARS coronavirus nonstructural protein 15 define an endoribonuclease family. *Proc. Natl. Acad. Sci. U.S.A.* 103, 11892–11897. doi:10.1073/pnas.0601708103
- Russell, C. D., Millar, J. E., and Baillie, J. K. (2020). Clinical evidence does not support corticosteroid treatment for 2019-nCoV lung injury. *Lancet* 395, 473–475. doi:10.1016/S0140-6736(20)30317-2
- Sadler, A. J., and Williams, B. R. (2008). Interferon-inducible antiviral effectors. *Nat. Rev. Immunol.* 8, 559–568. doi:10.1038/nri2314
- Samuel, C. E. (2001). Antiviral actions of interferons. *Clin Microbiol Rev* 14, 778–809. doi:10.1128/CMR.14.4.778-809.2001
- Sanghvi, V. R., and Steel, L. F. (2012). RNA silencing as a cellular defense against HIV-1 infection: progress and issues. *FASEB J.* 26, 3937–3945. doi:10.1096/fj.12-210765
- Sawicki, S. G., Sawicki, D. L., Younker, D., Meyer, Y., Thiel, V., Stokes, H., et al. (2005). Functional and genetic analysis of coronavirus replicase-transcriptase proteins. *PLoS Pathog.* 1 (4), e39. doi:10.1371/journal.ppat.0010039
- Sayed, A. M., Alhadrami, H. A., El-Gendy, A. O., Shamikh, Y. I., Belbahri, L., Hassan, H. M., et al. (2020). Microbial natural products as potential inhibitors of SARS-CoV-2 main protease (M(pro)). *Microorganisms* 8 (7), 970. doi:10.3390/microorganisms8070970
- Schwarz, S., Sauter, D., Wang, K., Zhang, R., Sun, B., Karioti, A., et al. (2014). Kaempferol derivatives as antiviral drugs against the 3a channel protein of coronavirus. *Planta Med.* 80, 177–182. doi:10.1055/s-0033-1360277
- Schwarz, S., Wang, K., Yu, W., Sun, B., and Schwarz, W. (2011). Emodin inhibits current through SARS-associated coronavirus 3a protein. *Antiviral Res.* 90, 64–69. doi:10.1016/j.antiviral.2011.02.008
- Shahid, I., Almalki, W. H., Alrabia, M. W., Mukhtar, M. H., Almalki, S. S. R., Alkahtani, S. A., et al. (2017). *In vitro* inhibitory analysis of consensus siRNAs against NS3 gene of hepatitis C virus 1a genotype. *Asian Pac. J. Trop. Med.* 10, 701–709. doi:10.1016/j.apjtm.2017.07.011
- Sharma, N., Verma, R., Kumawat, K. L., Basu, A., and Singh, S. K. (2015). miR-146a suppresses cellular immune response during Japanese encephalitis virus JaOArS982 strain infection in human microglial cells. *J. Neuroinflammation* 12, 30. doi:10.1186/s12974-015-0249-0
- Shim, B.-S., Wu, W., Kyriakis, C. S., Bakre, A., Jorquera, P. A., Perwitasari, O., et al. (2016). MicroRNA-555 has potent antiviral properties against poliovirus. *J. Gen. Virol.* 97, 659–668. doi:10.1099/jgv.0.000372
- Silveira, D., Prieto-Garcia, J. M., Boylan, F., Estrada, O., Fonseca-Bazzo, Y. M., Jamal, C. M., et al. (2020). COVID-19: is there evidence for the use of herbal medicines as adjuvant symptomatic therapy? *Front. Pharmacol.* 11, 581840. doi:10.3389/fphar.2020.581840
- Siu, K.-L., Kok, K.-H., Ng, M.-H. J., Poon, V. K., Yuen, K.-Y., Zheng, B.-J., et al. (2009). Severe acute respiratory syndrome coronavirus M protein inhibits type I interferon production by impeding the formation of TRAF3.TANK.TBK1/IKKepsilon complex. *J. Biol. Chem.* 284, 16202–16209. doi:10.1074/jbc.M109.008227
- Sohrab, S. S., El-Kafrawy, S. A., Mirza, Z., Kamal, M. A., and Azhar, E. I. (2018). Design and delivery of therapeutic siRNAs: application to MERS-coronavirus. *Curr. Pharm. Des.* 24, 62–77. doi:10.2174/1381612823666171109112307
- Song, J., Zhang, L., Xu, Y., Yang, D., Yang, S., Zhang, W., et al. (2020). The comprehensive study on the therapeutic effects of baicalin for the treatment of COVID-19 *in vivo* and *in vitro*. *Biochem. Pharmacol.* 183, 114302. doi:10.1016/j.bcp.2020.114302
- Su, B., Wang, Y., Zhou, R., Jiang, T., Zhang, H., Li, Z., et al. (2019). Efficacy and tolerability of lopinavir/ritonavir- and efavirenz-based initial antiretroviral therapy in HIV-1-infected patients in a tertiary care hospital in Beijing, China. *Front. Pharmacol.* 10, 1472. doi:10.3389/fphar.2019.01472
- Su, H.-X., Yao, S., Zhao, W.-F., Li, M.-J., Liu, J., Shang, W.-J., et al. (2020). Anti-SARS-CoV-2 activities *in vitro* of Shuanghuanglian preparations and bioactive ingredients. *Acta. Pharmacol. Sin.* 41, 1167–1177. doi:10.1038/s41401-020-0483-6
- Taning, C. N., Christiaens, O., Li, X., Swevers, L., Casteels, H., Maes, M., et al. (2018). Engineered flock house virus for targeted gene suppression through RNAi in fruit flies (*Drosophila melanogaster*) *in vitro* and *in vivo*. *Front. Physiol.* 9, 805. doi:10.3389/fphys.2018.00805
- Tanner, J. A., Watt, R. M., Chai, Y.-B., Lu, L.-Y., Lin, M. C., Peiris, J. M., et al. (2003). The severe acute respiratory syndrome (SARS) coronavirus NTPase/helicase belongs to a distinct class of 5' to 3' viral helicases. *J. Biol. Chem.* 278, 39578–39582. doi:10.1074/jbc.C300328200
- Tanner, J. A., Zheng, B.-J., Zhou, J., Watt, R. M., Jiang, J.-Q., Wong, K.-L., et al. (2005). The adamantane-derived bananins are potent inhibitors of the helicase activities and replication of SARS coronavirus. *Chem. Biol.* 12, 303–311. doi:10.1016/j.chembiol.2005.01.006
- Tapas, A. R., Sakarkar, D., and Kakde, R. (2008). Flavonoids as nutraceuticals: a review. *Trop J. Pharm. Res.* 7, 1089–1099. doi:10.4314/tjpr.v7i3.14693
- Taxman, D. J., Livingstone, L. R., Zhang, J., Conti, B. J., Iocca, H. A., Williams, K. L., et al. (2006). Criteria for effective design, construction, and gene knockdown by shRNA vectors. *BMC Biotechnol.* 6, 7. doi:10.1186/1472-6750-6-7
- Te Velthuis, A. J., Van Den Worm, S. H., and Snijder, E. J. (2012). The SARS-coronavirus nsp7+ nsp8 complex is a unique multimeric RNA polymerase capable of both de novo initiation and primer extension. *Nucleic Acids Res* 40, 1737–1747. doi:10.1093/nar/gkr893
- Thiel, V., Ivanov, K. A., Putics, A., Hertzog, T., Schelle, B., Bayer, S., et al. (2003). Mechanisms and enzymes involved in SARS coronavirus genome expression. *J. Gen. Virol.* 84, 2305–2315. doi:10.1099/vir.0.19424-0

- Uzunova, K., Filipova, E., Pavlova, V., and Vekov, T. (2020). Insights into antiviral mechanisms of remdesivir, lopinavir/ritonavir and chloroquine/hydroxychloroquine affecting the new SARS-CoV-2. *Biomed. Pharmacother.* 131, 110668. doi:10.1016/j.biopha.2020.110668
- Vazquez, F. (2006). Arabidopsis endogenous small RNAs: highways and byways. *Trends Plant Sci.* 11, 460–468. doi:10.1016/j.tplants.2006.07.006
- Wang, J., Qi, H., Bao, L., Li, F., and Shi, Y. (2020). A contingency plan for the management of the 2019 novel coronavirus outbreak in neonatal intensive care units. *Lancet Child Adolesc. Health* 4 (4), 258–259. doi:10.1016/S2352-4642(20)30040-7
- Wang, M., Cao, R., Zhang, L., Yang, X., Liu, J., Xu, M., et al. (2020). Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) *in vitro*. *Cell Res.* 30, 269–271. doi:10.1038/s41422-020-0282-0
- Wang, X., Cao, R., Zhang, H., Liu, J., Xu, M., Hu, H., et al. (2020). The anti-influenza virus drug, arbidol is an efficient inhibitor of SARS-CoV-2 *in vitro*. *Cell Discov.* 6, 28. doi:10.1038/s41421-020-0169-8
- Wang, Y., Mao, J., Wang, G., Qiu, Z., Yao, Q., and Chen, K. (2020). Human SARS-CoV-2 has evolved to reduce CG dinucleotide in its open reading frames. *Res. Squ.* 10, 12331. doi:10.1038/s41598-020-69342-y
- Wang, Z., Ren, L., Zhao, X., Hung, T., Meng, A., Wang, J., et al. (2004). Inhibition of severe acute respiratory syndrome virus replication by small interfering RNAs in mammalian cells. *J. Virol.* 78, 7523–7527. doi:10.1128/JVI.78.14.7523-7527.2004
- Woo, P. C., Huang, Y., Lau, S. K., and Yuen, K.-Y. (2010). Coronavirus genomics and bioinformatics analysis. *viruses* 2, 1804–1820. doi:10.3390/v2081803
- Wrensch, F., Winkler, M., and Pöhlmann, S. (2014). IFITM proteins inhibit entry driven by the MERS-coronavirus spike protein: evidence for cholesterol-independent mechanisms. *Viruses* 6, 3683–3698. doi:10.3390/v6093683
- Wu, C.-H., Yeh, S.-H., Tsay, Y.-G., Shieh, Y.-H., Kao, C.-L., Chen, Y.-S., et al. (2009). Glycogen synthase kinase-3 regulates the phosphorylation of severe acute respiratory syndrome coronavirus nucleocapsid protein and viral replication. *J. Biol. Chem.* 284, 5229–5239. doi:10.1074/jbc.M805747200
- Wu, C.-J., and Chan, Y.-L. (2006). Antiviral applications of RNAi for coronavirus. *Expert Opin. Investig. Drugs* 15, 89–97. doi:10.1517/13543784.15.2.89
- Wu, F., Zhao, S., Yu, B., Chen, Y. M., Wang, W., Song, Z. G., et al. (2020). A new coronavirus associated with human respiratory disease in China. *Nature* 579, 265–269. doi:10.1038/s41586-020-2008-3
- Wu, S., He, L., Li, Y., Wang, T., Feng, L., Jiang, L., et al. (2013). MiR-146a facilitates replication of dengue virus by dampening interferon induction by targeting TRAF6. *J. Infect* 67, 329–341. doi:10.1016/j.jinf.2013.05.003
- Xu, K., Chen, Y., Yuan, J., Yi, P., Ding, C., Wu, W., et al. (2020a). Clinical efficacy of arbidol in patients with 2019 novel coronavirus-infected pneumonia: a retrospective cohort study. *SSRN Elect. J.* 2020, 3542148. doi:10.2139/ssrn.3542148
- Xu, X., Chen, P., Wang, J., Feng, J., Zhou, H., Li, X., et al. (2020b). Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission. *Sci. China Life Sci.* 63, 457–460. doi:10.1007/s11427-020-1637-5
- Xu, Z., Peng, C., Shi, Y., Zhu, Z., Mu, K., Wang, X., et al. (2020c). Nelfinavir was predicted to be a potential inhibitor of 2019-nCoV main protease by an integrative approach combining homology modelling, molecular docking and binding free energy calculation. *BioRxiv* 2020, 921627. doi:10.1101/2020.01.27.921627
- Yeung, A. W. K., Aggarwal, B. B., Balacheva, A., Barreca, D., Battino, M., Belwal, T., et al. (2019). Resveratrol, a popular dietary supplement for human and animal health: quantitative research literature analysis. *Animal Sci. Papers Reports* 36 (4), 345–358.
- Yi, S., De Hua, Y., Xiong, J., Jie, J., Huang, B., and Jin, Y. X. (2005). Inhibition of genes expression of SARS coronavirus by synthetic small interfering RNAs. *Cell Res.* 15, 193–200. doi:10.1038/sj.cr.7290286
- Yu, M.-S., Lee, J., Lee, J. M., Kim, Y., Chin, Y.-W., Jee, J.-G., et al. (2012). Identification of myricetin and scutellarein as novel chemical inhibitors of the SARS coronavirus helicase, nsP13. *Bioorg. Med. Chem. Lett.* 22, 4049–4054. doi:10.1016/j.bmcl.2012.04.081
- Zang, N., Xie, X., Deng, Y., Wu, S., Wang, L., Peng, C., et al. (2011). Resveratrol-mediated gamma interferon reduction prevents airway inflammation and airway hyperresponsiveness in respiratory syncytial virus-infected immunocompromised mice. *J. Virol.* 85, 13061–13068. doi:10.1128/JVI.05869-11
- Zhai, A., Qian, J., Kao, W., Li, A., Li, Y., He, J., et al. (2013). Borna disease virus encoded phosphoprotein inhibits host innate immunity by regulating miR-155. *Antiviral Res.* 98, 66–75. doi:10.1016/j.antiviral.2013.02.009
- Zhang, H., Li, Z., Li, Y., Liu, Y., Liu, J., Li, X., et al. (2013). A computational method for predicting regulation of human microRNAs on the influenza virus genome. *BMC Syst. Biol.* 7 (S2), S3. doi:10.1186/1752-0509-7-S2-S3
- Zhang, H., Penninger, J. M., Li, Y., Zhong, N., and Slutsky, A. S. (2020a). Angiotensin-converting enzyme 2 (ACE2) as a SARS-CoV-2 receptor: molecular mechanisms and potential therapeutic target. *Intensive Care Med.* 46 (4), 586–590. doi:10.1007/s00134-020-05985-9
- Zhang, J., Zhou, L., Yang, Y., Peng, W., Wang, W., and Chen, X. (2020b). Therapeutic and triage strategies for 2019 novel coronavirus disease in fever clinics. *Lancet Respir. Med.* 8 (3), e11–e12. doi:10.1016/S2213-2600(20)30071-0
- Zhang, Y., Li, T., Fu, L., Yu, C., Li, Y., Xu, X., et al. (2004). Silencing SARS-CoV Spike protein expression in cultured cells by RNA interference. *FEBS Lett.* 560, 141–146. doi:10.1016/S0014-5793(04)00087-0
- Zhao, G., Shi, S.-Q., Yang, Y., and Peng, J.-P. (2006). M and N proteins of SARS coronavirus induce apoptosis in HPF cells. *Cell Biol. Toxicol.* 22, 313–322. doi:10.1007/s10565-006-0077-1
- Zhao, X., Guo, F., Liu, F., Cuconati, A., Chang, J., Block, T. M., et al. (2014). Interferon induction of IFITM proteins promotes infection by human coronavirus OC43. *Proc. Natl. Acad. Sci. U.S.A.* 111, 6756–6761. doi:10.1073/pnas.1320856111
- Zheng, B., He, M.-L., Wong, K.-L., Lum, C. T., Poon, L. L., Peng, Y., et al. (2004). Potent inhibition of SARS-associated coronavirus (SCOV) infection and replication by type I interferons (IFN- α/β) but not by type II interferon (IFN- γ). *J. Interferon Cytokine Res.* 24, 388–390. doi:10.1089/1079990041535610
- Zhou, P., Yang, X.-L., Wang, X.-G., Hu, B., Zhang, L., Zhang, W., et al. (2020). Discovery of a novel coronavirus associated with the recent pneumonia outbreak in humans and its potential bat origin. *BioRxiv* 2020, 914952. doi:10.1101/2020.01.22.914952
- Ziebuhr, J. (2005). The coronavirus replicase. *Coronavirus Rep. Rev. Gen.* 287, 57–94. doi:10.1007/3-540-26765-4_3
- Zimmermann, P., and Curtis, N. (2020). Coronavirus infections in children including COVID-19: an overview of the epidemiology, clinical features, diagnosis, treatment and prevention options in children. *Pediatr. Infect. Dis. J.* 39 (5), 355–368. doi:10.1097/INF.0000000000002660

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Kalhori, Saadatpour, Arefian, Soleimani, Farzaei, Aneva and Echeverria. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.