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Leading article

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## The prophylaxis of bacterial infections in neutropenic patients

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Bacterial sepsis remains a major cause of morbidity and mortality in patients rendered neutropenic following cytoreductive therapy for haematological malignancy. The introduction of more aggressive anti-neoplastic chemotherapeutic regimes in individuals with solid organ malignancies has resulted in increasing numbers of these patients experiencing the profound neutropenia (i.e. granulocyte counts of  $<0.1 \times 10^9/L$ ) that puts them at risk of infection. It is now over 30 years since Bodey *et al.*, in a landmark paper,<sup>1</sup> first quantified the risk of infection associated with severe neutropenia. Throughout this period much effort and considerable economic resources have been expended on developing effective approaches to prevent infection by bacteria and other microorganisms in this setting. Many of these strategies have fallen out of favour, having foundered on grounds of expense, patient unacceptability and consequent problems with compliance, and a lack of convincing scientific evidence to support their efficacy. One practice that has proved more durable than most is the use of antibacterial prophylaxis (ABP). This approach is not without its detractors and, given the problems attributed to ABP that have emerged in recent years, a re-examination of its risks and benefits is warranted.

When ABP was first introduced, Gram-negative bacteria—particularly members of the Enterobacteriaceae and *Pseudomonas aeruginosa*—were the most important bacterial pathogens encountered in neutropenic patients; infections were associated with significant case-fatality rates. Many of these bacteraemias were deemed to be autochthonous in origin, with the gut being the most likely source. The concept of ‘colonization resistance’, which underpinned the rationale for ABP, was developed by van der Waaij and co-workers<sup>2</sup> (reviewed in reference 3) following observations in laboratory animals. Mice with their own normal gut flora were able to resist colonization when challenged by exogenous Gram-negative aerobic bacteria, unless these were administered in very high doses. Conversely, in gnotobiotic mice, colonization was achieved

by inocula of 10–100 organisms. Germ-free mice that had first been colonized by anaerobic flora were able to resist colonization nearly as well as normal controls. Extrapolating this concept to humans, it was argued that prevention of infection from gut-derived Gram-negative bacteria during neutropenia could be achieved by administering antimicrobials that would selectively target these bacteria and that would leave the ‘beneficial’ anaerobes intact. Initial regimens comprised cocktails of orally administered non-absorbable antibacterials, often in conjunction with a polyene antifungal. Frequently, these agents were used in combination with oral antiseptics. These protocols were problematic to administer and patient compliance was poor.

Experience with this approach to ABP was mixed and results derived from published studies are difficult to analyse because of differences in the combinations used, the frequency of administration of the agents involved, and in the degree and duration of neutropenia experienced by trial subjects. In addition, there were variations in the timing of commencement of ABP relative to the onset of neutropenia and also in adjunctive measures such as dietary manipulation or nursing of patients in laminar air flow environments.

Following the observations of Hughes *et al.*,<sup>4</sup> who reported that co-trimoxazole prophylaxis against *Pneumocystis carinii* in children with acute leukaemia was also associated with a decrease in septicaemia and bacterial infections at other sites, this combination was adopted by many centres in preference to oral non-absorbable ABP regimens. Several studies (reviewed in reference 5), in which co-trimoxazole was compared with placebo or with other prophylactic agents, yielded conflicting results, with some studies failing to demonstrate significant differences in the number of febrile episodes, use of therapeutic antibacterials and mortality. Owing to marked differences in trial design, patient mix and data analysis, results of these investigations are difficult to compare. Furthermore, the statistical validity of the data obtained was compro-

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mised because the number of patients enrolled in these studies was often very low.<sup>6</sup> Despite the successes in the reduction of bacterial infections that were claimed for co-trimoxazole, a number of drawbacks were associated with its use, including toxicity such as hypersensitivity and prolongation of the period of neutropenia,<sup>7,8</sup> breakthrough infections with resistant Gram-negative bacteria,<sup>9</sup> and an increased risk of Gram-positive sepsis,<sup>10</sup> fungal infections<sup>8</sup> and *Clostridium difficile* colitis.<sup>11</sup>

Following their introduction in the early-mid-1980s, the fluoroquinolones were embraced with some enthusiasm as agents for ABP. This was because of their increased activity against Gram-negative bacilli, particularly *P. aeruginosa*, compared with co-trimoxazole. In addition, they were not myelosuppressive and did not appear to have the problems of hypersensitivity associated with co-trimoxazole. Furthermore, the lack of anti-anaerobic activity of these compounds would preserve the anaerobic component of the gut microflora that is of crucial importance to the concept of colonization resistance. Whilst there is no shortage of clinical trials (summarized in reference 12) of quinolones versus placebo or other ABP regimes, there have been few trials in which different quinolones have been compared.<sup>13-15</sup> Similarly, there is a lack of data on the use of quinolone ABP in patients with solid organ malignancies.<sup>16,17</sup> Once again, methodological differences and very small numbers of patients<sup>11</sup> militate against meaningful comparisons between these studies. In addition, there are few published reports on the use of quinolone prophylaxis in the community setting.<sup>18</sup> Nevertheless, proponents of quinolone ABP point to data which demonstrate that many, but not all,<sup>11,17</sup> of these regimens are associated with fewer Gram-negative bacteraemias than controls.<sup>12</sup> Some investigations, however, do not show a reduction in numbers of unexplained febrile episodes in patients receiving quinolone ABP.<sup>14,19</sup> Whether this is because systemic absorption of quinolones from the gut is merely converting blood cultures that, otherwise, would have been positive into 'no growth' specimens or whether this is due to other reasons, such as absorption of endotoxins, remains unresolved.<sup>14</sup> With regard to other parameters such as the use of broad-spectrum antibacterials for suspected or documented infection, duration of hospital stay and mortality, the advantages of ABP with quinolones (and co-trimoxazole) are less clear-cut. Donnelly and co-workers,<sup>20</sup> for example, report that despite receiving prophylaxis, nearly 80% of patients in their trial required therapy with broad-spectrum antimicrobials. Moreover, not all studies claim to demonstrate superiority of quinolones against comparator regimes. In their study of 230 patients, Donnelly *et al.*<sup>20</sup> noted that patients receiving co-trimoxazole and colistin had fewer febrile days, a reduced frequency of infectious episodes and a delayed onset of fever compared with those receiving quinolones. Bacteraemia due to resistant Gram-negative bacilli, however, only occurred in the former group.<sup>20</sup>

Whatever the perceived advantages of quinolones, the problem of superinfection with Gram-positive bacteria that are not susceptible to these compounds cannot be ignored. This was highlighted in the very first study that described the use of quinolones in neutropenia<sup>21</sup> and the problem remains a major disadvantage of ABP, even though some investigators have not observed this phenomenon.<sup>22</sup> Cruciani *et al.*,<sup>23</sup> in their meta-analysis of 13 trials in which quinolones were compared with co-trimoxazole, oral non-absorbable agents or placebo, concluded that quinolones were not effective in preventing Gram-positive bacteraemia.

Of particular concern has been bacteraemia with viridans group streptococci (VGS). These infections are characterized by significant morbidity including respiratory distress syndrome and endocarditis.<sup>24</sup> Case fatality rates may be as high as 30%.<sup>25</sup> It should be noted that, although several studies<sup>20,26,27</sup> have identified ABP as a risk factor for VGS sepsis, other predisposing factors have also been reported. These include high-dose cytosine chemotherapy,<sup>24,28</sup> use of antacids,<sup>29</sup> the presence of mucositis,<sup>24</sup> oral herpes simplex virus infection<sup>30</sup> and, indeed, co-trimoxazole prophylaxis.<sup>31</sup> The occurrence of VGS sepsis in quinolone ABP is, perhaps, not altogether surprising given the relative insusceptibility of the bacteria to earlier antimicrobials of this class. For example, McWhinney *et al.*<sup>32</sup> reported that of 47 blood culture isolates of VGS from neutropenic patients, only 4.3% were susceptible to ciprofloxacin and ofloxacin at breakpoints of 1 and 2 mg/L, respectively. It has been suggested that ofloxacin use is less likely to be associated with VGS sepsis than other quinolones.<sup>33</sup> Although most attention has focused on VGS infection in quinolone ABP, infections with other Gram-positive bacteria, such as *Stomatococcus mucilaginosus*<sup>34,35</sup> and coagulase-negative staphylococci,<sup>36,37</sup> have also been documented.

Strategies to counter the problem of VGS have centred around the addition of other agents with increased anti-Gram-positive activity to ABP regimens. The rationale for this practice is supported by animal work, in which survival rates in irradiated mice were higher in those animals receiving a combination of penicillin and ofloxacin compared with a group receiving only ofloxacin.<sup>38</sup> Given that some neutropenic populations may have a high prevalence of penicillin-resistant VGS<sup>39</sup> and that blood culture isolates of these bacteria from neutropenic patients are frequently penicillin resistant,<sup>32</sup> it is not altogether surprising that studies have reported an increase in infections with strains of VGS intermediately susceptible, or resistant, to penicillin.<sup>40,42</sup> Subsequent approaches have included the addition of macrolides,<sup>33,42</sup> rifampicin<sup>14</sup> and vancomycin<sup>41</sup> to quinolone ABP, with varying degrees of success. However, the potential for the selection of resistant bacteria, especially vancomycin-resistant enterococci must be acknowledged.<sup>43</sup> Although much attention has been focused on Gram-positive infections in patients receiving

quinolone ABP, there is increasing awareness of superinfection with resistant Gram-negative pathogens including *Escherichia coli*,<sup>44,45</sup> *P. aeruginosa*<sup>46,47</sup> and *Stenotrophomonas maltophilia*<sup>47</sup> as well as more unusual bacterial species such as *Leptotrichia buccalis*.<sup>48</sup>

With the introduction of newer agents of the quinolone class which have increased activity against Gram-positive bacteria in general,<sup>49</sup> and VGS in particular,<sup>50</sup> whilst retaining good activity against the Enterobacteriaceae and *P. aeruginosa*, is there further hope for quinolone ABP? Clearly, it is too early to predict the role of these newer compounds in this setting but it should be noted that many, although not all, of these newer drugs have significant in-vitro activity against anaerobes and would thus undermine the concept of colonization resistance. Well designed trials that avoid some of the pitfalls encountered by early investigations, particularly small numbers of subjects and the inclusion of patients with marked variations in the degree and duration of neutropenia, are awaited.

Although ABP will continue to be championed by many, perhaps it is a practice that has outstayed its welcome. As has been noted,<sup>5</sup> the response rate with timely administered empirical therapy is, currently, gratifyingly high, thus calling into question whether the putative benefits of ABP outweigh the risks of antimicrobial resistance and adverse effects.<sup>51</sup> Although this may be regarded by some as an overly pessimistic view, there are alternative strategies for preventing bacterial infections in the neutropenic patient that do not rely on administration of prophylactic antimicrobials. These include use of recombinant colony stimulating factors,<sup>52</sup> bone marrow protective agents such as amifostine,<sup>53</sup> which also has general cytoprotective properties and thus may reduce mucositis<sup>54</sup> (itself a risk factor for bacteraemia in neutropenia) and negative bone marrow regulators, including MIP1 $\alpha$ .<sup>55</sup> Technical developments have also led to a reawakening of interest in the use of prophylactic granulocyte transfusions which may also play a role in preventing infection.<sup>56</sup> Whilst the benefit of these new approaches remains to be assessed in full, the role of simple, if technologically unexciting measures such as good hand hygiene, care of vascular access devices and appropriate dietary advice, should also not be forgotten.

## References

1. Bodey, G. P., Buckley, M., Sathe, Y. S. & Freireich, E. J. (1966). Quantitative relationship between circulating leukocytes and infection in patients with acute leukemia. *Annals of Internal Medicine* **64**, 328–40.
2. van der Waaij, D., Berghuis-de Vries, J. M. & Lekkerkerk-van der Wees, J. E. (1971). Colonization resistance of the digestive tract in conventional and antibiotic-treated mice. *Journal of Hygiene* **69**, 405–11.
3. Vollaard, E. J. & Clasener, H. A. (1994). Colonization resistance. *Antimicrobial Agents and Chemotherapy* **38**, 409–14.
4. Hughes, W. T., Kuhn, S., Chaudhary, S., Feldman, S., Verzosa, M., Aur, R. J. *et al.* (1977). Successful chemoprophylaxis for *Pneumocystis carinii* pneumonitis. *New England Journal of Medicine* **297**, 1419–26.
5. Walsh, T. J., Karp, J., Hathorn, J. W. & Pizzo, P. A. (1994). Prevention of bacterial infections in neutropenic patients. *Baillière's Clinical Infectious Diseases* **1**, 469–98.
6. Weiser, B., Lange, M., Fialk, M. A., Singer, C., Szatrowski, T. H. & Armstrong, D. (1981). Prophylactic trimethoprim-sulfamethoxazole during consolidation chemotherapy for acute leukemia: a controlled trial. *Annals of Internal Medicine* **95**, 436–8.
7. Imrie, K. R., Prince, H. M., Couture, F., Brandwein, J. M. & Keating, A. (1995). Effect of antimicrobial prophylaxis on hematopoietic recovery following autologous bone marrow transplantation: ciprofloxacin versus co-trimoxazole. *Bone Marrow Transplantation* **15**, 267–70.
8. Wade, J. C., de Jongh, C. A., Newman, K. A., Crowley, J., Wiernik, P. H. & Schimpff, S. C. (1983). Selective antimicrobial modulation as prophylaxis against infection during granulocytopenia: trimethoprim-sulfamethoxazole vs. nalidixic acid. *Journal of Infectious Diseases* **147**, 624–34.
9. Wilson, J. M. & Guiney, D. G. (1982). Failure of oral trimethoprim-sulfamethoxazole prophylaxis in acute leukemia. Isolation of resistant plasmids from strains of Enterobacteriaceae causing bacteremia. *New England Journal of Medicine* **306**, 16–20.
10. Bow, E. J. & Ronald, A. R. (1993). Antibacterial chemoprophylaxis in neutropenic patients—where do we go from here? *Clinical Infectious Diseases* **17**, 333–7.
11. Lew, M. A., Kehoe, K., Ritz, J., Antman, K. H., Nadler, L., Takvorian, T. *et al.* (1991). Prophylaxis of bacterial infections with ciprofloxacin in patients undergoing bone marrow transplantation. *Transplantation* **51**, 630–6.
12. Patrick, C. C. (1997). Use of fluoroquinolones as prophylactic agents in patients with neutropenia. *Pediatric Infectious Disease Journal* **16**, 135–9.
13. GIMEMA Infection Program. (1991). Prevention of bacterial infection in neutropenic patients with hematologic malignancies; a randomized multicenter trial comparing norfloxacin with ciprofloxacin. *Annals of Internal Medicine* **115**, 7–12.
14. Bow, E. J., Mandell, L. A., Louie, T. J., Feld, R., Palmer, M., Zee, B. *et al.* (1996). Quinolone-based antibacterial chemoprophylaxis in neutropenic patients: effect of augmented gram-positive activity on infectious morbidity. *Annals of Internal Medicine* **125**, 183–90.
15. D'Antonio, D., Iacone, A., Fioritoni, G., Betti, S., di Girolamo, A., Piccolomini, R. *et al.* (1991). Antibacterial prophylaxis in granulocytopenic patients: a randomized study of ofloxacin versus norfloxacin. *Current Therapeutic Research* **50**, 304–11.
16. Carlson, J. W., Fowler, J. M., Saltzman, A. K., Carter, J. R., Chen, M. D., Mitchell, S. K. *et al.* (1994). Chemoprophylaxis with oral ciprofloxacin in ovarian cancer patients receiving taxol. *Gynecologic Oncology* **55**, 415–20.
17. Carlson, J. W., Fowler, J. M., Mitchell, S. K., Carson, L. F., Mayer, A. R. & Copeland L. J. (1997). Chemoprophylaxis with ciprofloxacin in ovarian cancer patients receiving paclitaxel: a randomized trial. *Gynecologic Oncology* **65**, 325–9.
18. Meisnerberg, B., Gollard, R., Brehm, T., McMillan, R. & Miller, W. (1996). Prophylactic antibiotics eliminate bacteraemia and allow safe out-patient management following high-dose chemotherapy

and autologous stem-cell rescue. *Supportive Care in Cancer* **4**, 364–9.

19. de Marie, S., van den Broek, P. J., Willemze, R. & van Furth, R. (1993). Strategy for antibiotic therapy in febrile neutropenic patients on selective antimicrobial decontamination. *European Journal of Clinical Microbiology and Infectious Diseases* **12**, 897–906.

20. Donnelly, J. P., Maschmeyer, G. & Daenen, S. (1992). Selective oral antimicrobial prophylaxis for the prevention of infection in acute leukaemia—ciprofloxacin versus co-trimoxazole plus colistin. *European Journal of Cancer* **28A**, 873–8.

21. Rozenberg-Arska, M., Dekker, A. W. & Verhoef, J. A. (1985). Ciprofloxacin for selective decontamination of the alimentary tract in patients with acute leukemia during remission induction treatment: the effect of fecal flora. *Journal of Infectious Diseases* **152**, 104–7.

22. Liang, R. H., Yung, R. W., Chan, T.-K., Chau, P.-Y., Lam, W.-K., So, S.-Y. *et al.* (1990). Ofloxacin versus co-trimoxazole for prevention of infection in neutropenic patients following cytotoxic chemotherapy. *Antimicrobial Agents and Chemotherapy* **34**, 215–8.

23. Cruciani, M., Rampazzo, R., Malena, M., Lazzarini, L., Todeschini, G., Messori, A. *et al.* (1996). Prophylaxis with fluoroquinolones for bacterial infections in neutropenic patients: a meta-analysis. *Clinical Infectious Diseases* **23**, 795–805.

24. Bochud, P.-Y., Eggiman, P., Calandra, T., van Melle, G., Saghafi, L. & Francioli, P. (1994). Bacteremia due to viridans streptococcus in neutropenic patients with cancer: clinical spectrum and risk factors. *Clinical Infectious Diseases* **18**, 25–31.

25. Bochud, P.-Y., Calandra, T. & Francioli, P. (1994). Bacteremia due to viridans streptococci in neutropenic patients: a review. *American Journal of Medicine* **97**, 256–64.

26. McWhinney, P. H. M., Gillespie, S. H., Kibbler, C. C., Hoffbrand, A. V. & Prentice, H. G. (1991). *Streptococcus mitis* and ARDS in neutropenic patients. *Lancet* **337**, 429.

27. Classen, D. C., Burke, J. P., Ford, C. D., Evershed, S., Aloia, M. R., Wilfahrt, J. K. *et al.* (1990). *Streptococcus mitis* sepsis in bone marrow transplant patients receiving oral antimicrobial prophylaxis. *American Journal of Medicine* **89**, 441–6.

28. Richard, P., Amador del Valle, G., Moreau, P., Milpied, N., Felice, M.-P., Daeschler, T. *et al.* (1995). Viridans streptococcal bacteraemia in patients with neutropenia. *Lancet* **345**, 1607–9.

29. Elting, L. S., Bodey, G. P. & Keefe, B. H. (1992). Septicemia and shock syndrome due to viridans streptococci: a case control study of predisposing factors. *Clinical Infectious Diseases* **14**, 1201–7.

30. Ringden, O., Heimdahl, A., Lönnqvist, B., Malmborg, S. & Wilczek, H. (1984). Decreased incidence of viridans streptococcal septicaemia in allogeneic bone marrow transplant recipients after the introduction of acyclovir. *Lancet* **1**, 744.

31. Cohen, J., Donnelly, J. P., Worsley, A. M., Catovsky, D., Goldman, J. H. & Galton, D. A. (1983). Septicaemia caused by viridans streptococci in neutropenic patients with leukaemia. *Lancet* **2**, 1452–4.

32. McWhinney, P. H., Patel, S., Whiley, R. A., Hardie, J. M., Gillespie, S. H. & Kibbler, C. C. (1993). Activities of potential therapeutic and prophylactic antibiotics against blood culture isolates of viridans group streptococci from neutropenic patients receiving ciprofloxacin. *Antimicrobial Agents and Chemotherapy* **37**, 2493–5.

33. Kern, W. V., Hay, B., Kern, P., Marre, R. & Arnold, R. (1994). A randomized trial of roxithromycin in patients with acute leukemia and

bone marrow transplant recipients receiving fluoroquinolone prophylaxis. *Antimicrobial Agents and Chemotherapy* **38**, 465–72.

34. McWhinney, P. H., Kibbler, C. C., Gillespie, S. H., Patel, S., Morrison, D., Hoffbrand, A. V. *et al.* (1992). *Stomatococcus mucilaginosus*: an emerging pathogen in neutropenic patients. *Clinical Infectious Diseases* **14**, 641–6.

35. Van Tiel, F. H., Slangen, B. F., Schouten, H. C. & Jacobs, J. A. (1995). Study of *Stomatococcus mucilaginosus* isolated in a hospital ward using phenotypic characterization. *European Journal of Clinical Microbiology and Infectious Diseases* **14**, 193–8.

36. Winston, D. J., Ho, W. G., Nakao, S. L., Gale, R. P. & Champlin, R. E. (1986). Norfloxacin versus vancomycin/polymyxin for prevention of infection in granulocytopenic patients. *American Journal of Medicine* **88**, 884–90.

37. Oppenheim, B. A., Hartley, J. W., Lee, W. & Burnie J. P. (1989). Outbreak of coagulase negative staphylococci highly resistant to ciprofloxacin in a leukemia unit. *British Medical Journal* **299**, 294–7.

38. Brook, I. & Ledney, G. D. (1991). Ofloxacin and penicillin G combination therapy in prevention of bacterial translocation and animal mortality after irradiation. *Antimicrobial Agents and Chemotherapy* **35**, 1685–7.

39. Guiot, H. F., Corel, L. J. & Vossen, J. M. (1994). Prevalence of penicillin-resistant viridans streptococci in healthy children and in patients with malignant haematological disorders. *European Journal of Clinical Microbiology and Infectious Diseases* **13**, 645–50.

40. Krcmery, V. & Trupl, J. (1995). Bacteraemia due to penicillin-resistant *Streptococcus viridans* in cancer patients, before and after prophylaxis with penicillin. *Lancet* **346**, 1362–3.

41. Broun, E. R., Wheat, J. L., Kneebone, P. H., Sundblad, K., Hromas, R. A. & Tricot, G. (1994). Randomized trial of the addition of Gram-positive prophylaxis to standard antimicrobial prophylaxis for patients undergoing autologous bone marrow transplantation. *Antimicrobial Agents and Chemotherapy* **38**, 576–9.

42. Wimperis, J. Z., Baglin, T. P., Marcus, R. E. & Warren, R. E. (1991). An assessment of the efficacy of antimicrobial prophylaxis in bone marrow autografts. *Bone Marrow Transplantation* **8**, 363–7.

43. Plessis, P., Lamy, T., Donnio, P. Y., Autuly, F., Grulois I., Le Prise, P. Y. *et al.* (1995). Epidemiologic analysis of glycopeptide-resistant *Enterococcus* strains in neutropenic patients receiving prolonged vancomycin administration. *European Journal of Clinical Microbiology and Infectious Diseases* **14**, 959–63.

44. Carratala, J., Fernandez-Sevilla, A., Tubau, F., Callis, M. & Gudiol, F. (1995). Emergence of quinolone-resistant *Escherichia coli* bacteremia in neutropenic patients with cancer who have received prophylactic norfloxacin. *Clinical Infectious Diseases* **20**, 557–60.

45. Kern, W. V., Markus, A. & Andriof, E. (1994). Bacteremia due to fluoroquinolone-resistant *Escherichia coli* in two immunocompromised patients. *European Journal of Clinical Microbiology and Infectious Diseases* **13**, 161–5.

46. Cruciani, M., Concia, E. & Navarra, A. (1989). Prophylactic co-trimoxazole versus norfloxacin in neutropenic children—prospective randomized study. *Infection* **17**, 65–9.

47. Spanik, S., Trupl, J., Ilavská, I., Helpianska, L., Drgona, L., Demitrovicova, A. *et al.* (1996). Bacteremia and fungemia occurring during antimicrobial prophylaxis with ofloxacin in cancer patients; risk factors, etiology and outcome. *Journal of Chemotherapy* **8**, 387–93.

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48. Schwartz, D. N., Schable, B., Tenover, F. C. & Miller, R. D. (1995). *Leptotrichia buccalis* bacteremia in patients treated in a single bone marrow transplant unit. *Clinical Infectious Disease* **20**, 762–7.
49. Piddock, L. J. (1994). New quinolones and Gram-positive bacteria. *Antimicrobial Agents and Chemotherapy* **32**, 163–9.
50. Kerr, K. G., Armitage, H. T. & McWhinney, P. H. (1999). Activity of quinolones against viridans group streptococci isolated from blood cultures of patients with haematological malignancy. *Supportive Care in Cancer* **7**, 28–30.
51. Rubinstein, E., Potgieter, P., Davey, P. & Norrby, S. R. (1994). The use of fluoroquinolones in neutropenic patients—analysis of adverse effects. *Journal of Antimicrobial Chemotherapy* **34**, 7–19.
52. Lau, A. S., Lehman, D., Geertsma, F. R. & Yeung, M. C. (1996). Biology and therapeutic use of myeloid hematopoietic growth factors and interferons. *Pediatric Infectious Disease Journal* **15**, 563–75.
53. McCauley, D. L. (1997). Amifostine. A novel cytoprotective agent. *Cancer Practice* **5**, 189–91.
54. Griggs, J. J. (1998). Reducing the toxicity of anticancer therapy; new strategies. *Leukemia Research* **22**, Suppl. 1, S27–33.
55. Marshall, E., Woolford, L. B. & Lord, B. I. (1997). Continuous infusion of macrophage inflammatory protein MIP-1 $\alpha$  enhances leucocyte recovery and haemopoietic progenitor cell mobilization after cyclophosphamide. *British Journal of Cancer* **75**, 1715–20.
56. Grigg, A., Vecchi, L., Bardy, P. & Szer, J. (1996). G-CSF stimulated donor granulocyte collections for prophylaxis and therapy of neutropenic sepsis. *Australian and New Zealand Journal of Medicine* **26**, 813–8.

