

Perspective

The Role of Lactylation in Mental Illness: Emphasis on Microglia

Adonis Sfera ^{1,*}, Carolina Klein ², Johnathan J. Anton ³, Zisis Kozlakidis ⁴ and Christina V. Andronescu ⁵

¹ Patton State Hospital, Riverside, CA 92369, USA

² Napa State Hospital, 2100 Napa Vallejo Hwy, Napa, CA 94558, USA

³ Department of Biomedical Sciences, California Baptist University, 8432 Magnolia Ave., Riverside, CA 92504, USA

⁴ International Agency for Research on Cancer, World Health Organization, 69000 Lyon, France

⁵ Department of Cell Biology, Stanford University, 450 Serra Mall, Stanford, CA 94305, USA

* Correspondence: adonis.sfera@dsh.ca.gov

Abstract: A paradigm shift is currently taking place in the etiopathogenesis of neuropsychiatric disorders as immunometabolism is replacing the earlier neurotransmitter model. According to the new concept, cellular bioenergetics drives information processing in the central nervous system; therefore, neuropathology is conceptualized as a direct consequence of impaired metabolism. Along the same lines, endoplasmic reticulum stress and gut barrier dysfunction are emerging as novel targets in schizophrenia and affective disorders, linking immune responses to cellular distress. Furthermore, microglia, the brain's innate immune cells, acquire energy through oxidative phosphorylation, while in the resting state, and glycolysis upon activation, contributing to lactate accumulation and reduced brain pH. The same metabolic signature characterizes neuropsychiatric disorders as the central nervous system derives adenosine triphosphate from aerobic glycolysis, upregulating lactate and generating an acidic environment. Although known for over three decades, the link between dysmetabolism and neuropathology was poorly defined until the discovery of brain-resident innate lymphoid cells, including natural killer cells, and lactylation of histone and nonhistone proteins. In this perspective article, we examine three anti-inflammatory microglial systems relevant for neuropsychiatry: lactate, oxytocin, and the aryl hydrocarbon receptor. We also discuss potential interventions for restoring microglial homeostasis.

Keywords: microglia; neuropathology; lactylation; oxytocin; aryl hydrocarbon receptor



Citation: Sfera, A.; Klein, C.; Anton, J.J.; Kozlakidis, Z.; Andronescu, C.V. The Role of Lactylation in Mental Illness: Emphasis on Microglia. *Neuroglia* **2023**, *4*, 119–140. <https://doi.org/10.3390/neuroglia4020009>

Academic Editor: Antonia Cianciulli

Received: 2 March 2023

Revised: 4 May 2023

Accepted: 8 May 2023

Published: 16 May 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Severe neuropsychiatric illnesses, including schizophrenia and affective disorders, have been associated with brain metabolic disturbances, including excessive aerobic glycolysis and upregulated lactate, emphasizing that in the central nervous system (CNS) bioenergetics and information processing are highly intertwined [1–4]. Aerobic glycolysis and upregulated lactate, known as the Warburg effect, are the preferred energy sources of cancer cells and likely drive the pathogenesis of neuropsychiatric disorders [5,6]. Although low levels of lactate are neuroprotective, in excess it was demonstrated to induce neuronal apoptosis and decreased glutamatergic and GABAergic neurotransmission, hallmarks of neuropathology [7,8]. Along this line, a novel schizophrenia (SCZ) study has found that oral flora-derived lactate crosses the blood–brain barrier (BBB), generating CNS acidity [9,10].

Microglia are the CNS-resident macrophages known for vigilantly scanning the brain parenchyma, searching for tissue damage, molecular debris, and invading pathogens [10,11]. When changes are detected, microglia become activated and bioenergetically dependent on lactate [10]. Inflamed microglia, a characteristic of several neuropsychiatric disorders,

release proinflammatory cytokines and adopt a neurotoxic phenotype [12]. Hence, targeting microglial lactate is an emerging immunometabolic strategy in neuropsychiatric disorders [13–15] (see the section on Potential Therapeutic Strategies).

Recent studies have revealed that lactate is an essential CNS metabolite which promotes synaptic plasticity and exerts anti-inflammatory, antipsychotic, and antidepressant properties [16–20]. These are mediated by suppression of nuclear factor kappa B (NF- κ B) via lactate signaling with its receptor hydroxycarboxylic acid receptor 1 (HCAR1) (also known as G-protein-coupled receptor 81 (GPR81)) [18]. In addition, lactate promotes axonal myelination as well as the rehabilitation of injured oligodendrocytes (OLs), highlighting a potential treatment modality for multiple sclerosis (MS) [21–23].

Lactylation is a post-translational modification (PTM) which utilizes lactate as a substrate; therefore, excessive availability of this metabolite promotes lactylation-mediated pathologies, including malignant transformation, Alzheimer’s disease (AD), and neuropsychiatric disorders [24–27]. Lysine lactylation (Kla), discovered in 2019 by Yingming Zhao, can be a physiological or pathological process, depending on the lactylation levels, which regulates gene transcription by metabolically reprogramming the neurons and glia [28,29] (Figure 1). In addition, the lactylation of nonhistone proteins, such as the high-mobility group box 1 (HMGB1), hypoxia-inducible factor 1 alpha (HIF-1 α), and the membrane-organizing extension spike protein (moesin), was associated with tumorigenesis and schizophrenia, further linking dysfunctional lactylation to various pathologies [30–32].

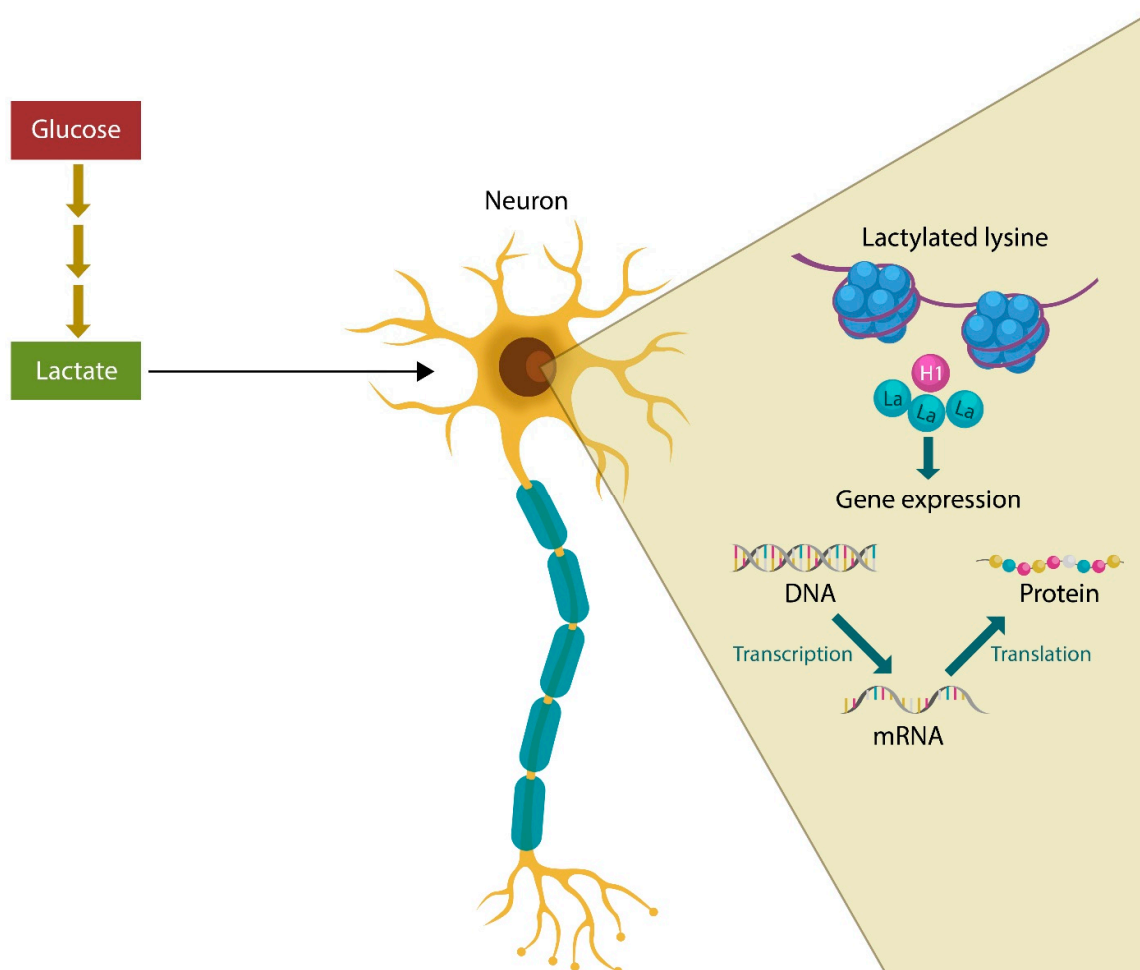


Figure 1. Excessive lactate derived from glucose metabolism can lead to excessive histone lysine lactylation (Kla), promoting pathologies, such as cancer, AD, schizophrenia, and related neuropsychiatric illnesses. Histone lactylation alters gene expression that in return affects translation and protein synthesis.

Another major discovery took place in 2022, when a subpopulation of oxytocin (OXT)-producing microglia was discovered, revealing a novel anti-inflammatory phenotype of these cells, likely distinct from the resting state [33,34]. Indeed, OXT signaling with OXT receptors (OXTRs), expressed on microglia, astrocytes, and oligodendrocytes, was reported to lower not only neuroinflammation but also depression, suicidal behavior, and psychosis, highlighting new potential pharmacological targets [35–44]. In addition, OXT promotes myelination and supports the nervous system white matter, likely explaining the reason this hormone was patented as an MS treatment (US patent 3274060A) [38,45,46].

The aryl hydrocarbon receptor (AhR), a cytosolic ligand-activated transcription factor, regulates the expression of numerous genes, including those involved in the response to environmental pollutants, polarization of microglia, myelination, and inflammation [47–49]. In the CNS, AhR exerts anti-inflammatory properties on microglia and astrocytes, promoting restorative neurogenesis and post-insult tissue regeneration [50]. However, in ischemic strokes, AhR may activate microglia, triggering inflammation, suggesting that this receptor is not only tissue-specific but also dependent on the pathology type [51]. In addition, AhR modulates meningeal innate lymphoid cells (ILCs), especially the protective CD56bright natural killer cells (NKC), promoting a tolerant, IL-10-generating phenotype [52,53]. Conversely, dysfunctional NKCs promote neuroinflammation by aberrantly eliminating the anti-inflammatory resting microglia, disrupting immune homeostasis by selectively sparing the activated, inflamed cells [54].

In this perspective article, we take a closer look at three anti-inflammatory microglial systems relevant for neuropsychiatry, lactate, OXT, and AhR, as well as potential interventions for restoring microglial homeostasis.

2. Lactate and Mental Illness

Major neuropsychiatric illnesses, including schizophrenia and affective disorders, have been associated with decreased brain pH, lactic acid accumulation and upregulated glycolysis [1,2,8]. From a bioenergetic perspective, the brain cells of mentally ill individuals resemble cancer cells, as adenosine triphosphate (ATP) is acquired through aerobic glycolysis, or the Warburg effect, decreasing the brain pH, while oxidative phosphorylation (OXPHOS) is downregulated.

Lactate plays a key role in the CNS where it functions as a metabolite as well as a neurotransmitter. Lactate is endowed with anti-inflammatory properties as it inhibits NK-kB, promotes myelination, and regulates microglial function [22,55]. Indeed, microglia possess the molecular machinery to shift metabolism according to phenotype; resting microglia rely on OXPHOS, while activated microglia utilize aerobic glycolysis [23].

Innate lymphoid cells (ILCs) signaling with microglia and the discovery of histone and nonhistone proteins lactylation have contributed to a better understanding of the role of aerobic glycolysis in neuropsychiatric disorders. Although lactate is neuroprotective and mediates synaptic plasticity, excessive lactylation can alter neuronal function, shifting them to an energy state akin to malignant cells [56–58] (see section Microglia, lactylation and mental illness). For example, aberrant neuronal cell cycle reentry and aneuploidy have been observed in patients with SCZ, suggesting that lactate triggers “reprimativization” of brain cells or the acquisition of stem cell properties [30,59–63]. This is further substantiated by the neuronal progenitor cells’ dependency on aerobic glycolysis, suggesting that local acidity promotes undifferentiated cellular phenotypes [64,65]. Indeed, in an acidic environment, somatic mammalian cells could be epigenetically reprogrammed into pluripotent stem cells, while L-lactate has been shown to induce neurogenesis in adult hippocampal neurons [66–68]. Moreover, exposed to lactate, cardiomyocytes were demonstrated to reenter the cell cycle in a manner reminiscent of neurons, emphasizing the link between low pH and postmitotic cells’ replication attempts [69].

Moesin is a member of the ezrin-radixin-moesin (ERM) family of proteins which are constituent parts of the cellular cytoskeleton, and thus implicated in mitosis [70]. In AD, moesin activates neuronal cell cycle reentry, indicating that lactylation of this protein

may promote neuropsychiatric pathology [71–73]. Indeed, moesin was implicated in schizophrenia and plays a major role in the aberrant NKCs activation [74–76] (Figure 2).

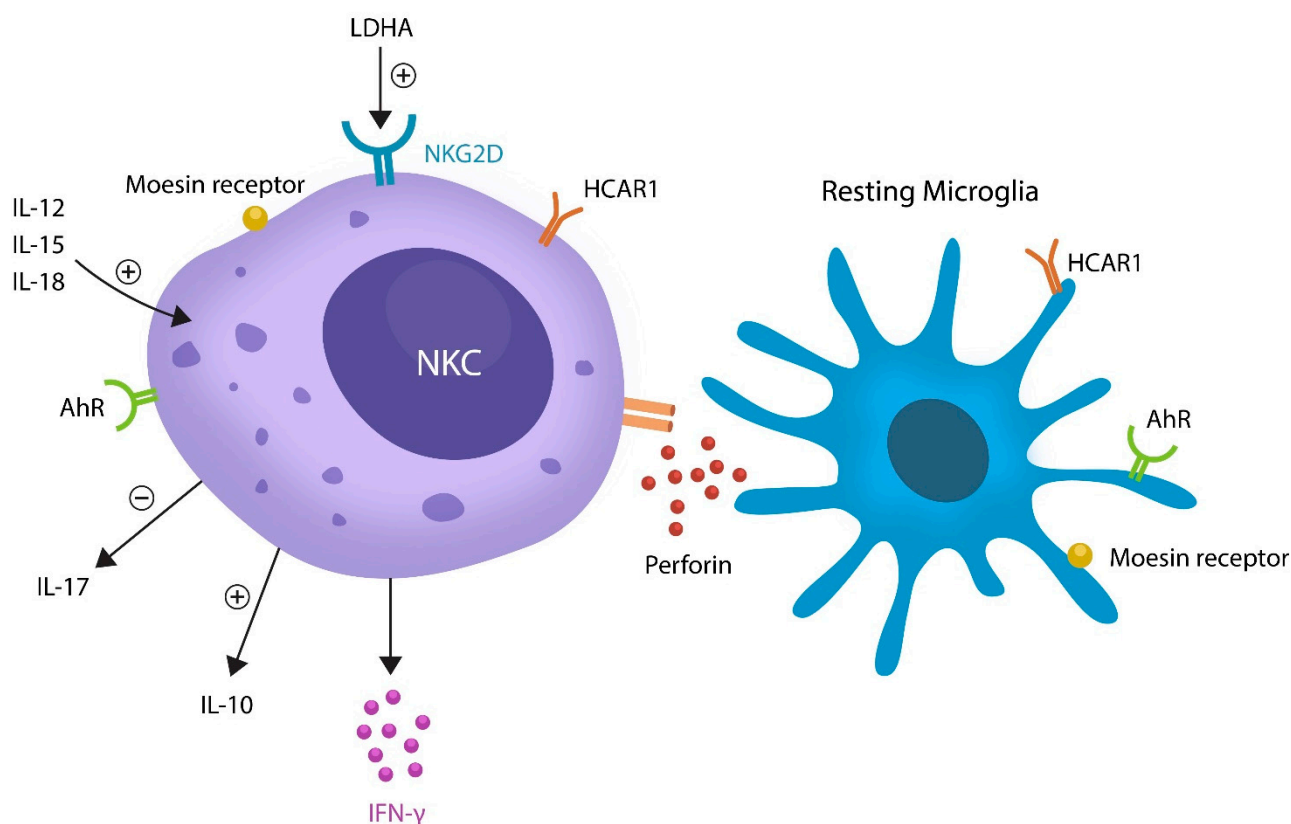


Figure 2. Aberrant elimination of resting microglia by lactate-activated NKCs. LDHA or LDH-5 can activate NKC via NKG2D receptors. IL-12, IL-15, and IL-18 activate NKCs, facilitating the release of IL-10, IL17, and IFN- γ . NKCs express HCAR1, AhR, and moesin receptors, proteins previously implicated in neuropathology. Activated NKCs release perforin, a toxic molecule known for eliminating resting microglia. Because activated microglia are not killed, this is a likely mechanism of tilting the balance toward inflammation.

Histone K1a, a reversible PTM, is regulated by EP300 and class I histone deacetylases (HDAC1–3), enzymes which add and remove lactate to and from the histone proteins [77–79]. Both HDAC1–3 and EP300 have been implicated in SCZ and/or the adverse effects of antipsychotic drugs via the brain-derived neurotrophic factor (BDNF) [80–82]. As BDNF gene histone modifications contribute to fear extinction, this system’s malfunction likely triggers K1a-mediated neuropsychiatric symptoms, such as excitation, social defeat, anxiety, and stress [83,84]. Interestingly, the neuroprotective properties of BDNF consist of preventing neuronal cell cycle reentry, further linking attempted mitosis to psychopathology [85].

NKCs are members of ILCs which are activated by lactate and sense pathological changes, migrating to the affected sites, including the brain parenchyma [86]. Lactate-activated NKCs were found to eliminate resting microglia but not the activated phenotype, suggesting that the pathological loss of anti-inflammatory microglia triggers neuroinflammation and the related pathology [54] (Figure 2). Moreover, as activated microglia have been linked to neurotoxicity and demyelination, pathological NKCs may predispose to autoimmune inflammation, including multiple sclerosis (MS) [12,87,88]. This is significant as impaired NKCs were found in the first episode of SCZ in antipsychotic-naïve individuals, suggesting that manipulation of these cells could ameliorate psychotic symptoms [89].

Recent studies have revealed an alternative source of brain lactate, the microbial community residing in various body compartments, including the oral cavity and the GI tract [1,9]. Indeed, the high lactate producers *Bifidobacterium* and *Lactobacillus* spp. have been shown to maintain the integrity of intestinal epithelial cells (IECs) and the survival of other commensals [90,91]. However, excessive lactate accumulation in the GI tract can trigger pathology by lowering the production of short-chain fatty acids (SCFA), biomolecules in charge of numerous CNS and peripheral physiological functions [92]. Indeed, the lactate-induced anti-inflammatory and immunosuppressive milieu can predispose to cancer as well as microbial and viral infections [93–95]. For example, lactate signaling with microglial HCAR1 promotes tissue regeneration and healing; however, it may also induce pathology by suppressing the phagocytosis of malignant or pathogen-infected cells [94–96].

Mental Illness, Lactylation, and Microbes

Histone K14 is not limited to mammals as it was also demonstrated in prokaryotes, linking microbial lactate to host neuropathology [97]. For example, *Escherichia coli* (*E. coli*) proteins YiaC and CobB induce histone lactylation, likely explaining the association of this bacterium with schizophrenia [98]. This was exemplified in the 2011 outbreak of *E. coli* in Germany which was accompanied by psychosis, linking this pathogen to mental illness [99,100]. In another example, urinary tract infection (UTI), caused primarily by *E. coli*, is known for the potential to trigger new-onset psychosis or the exacerbation of psychotic symptoms in stable patients with schizophrenia, further illustrating the link between this pathogen and mental illness [101,102].

Lactylation of nonhistone proteins, HMGB1, HIF-1 α , and moesin, was associated with both schizophrenia and *E. coli* infection, linking this microbe once more to neuropathology [30,31,103–106]. Moreover, *Toxoplasma gondii*, an intracellular protozoan, previously associated with SCZ, is another example of pathogen-mediated lactylation, causing host neuropathology [107]. Indeed, a recent study found that *Toxoplasma gondii*, containing 523 lactylated proteins, eliminates host inhibitory synapses by aberrantly activating microglia [108,109]. Moreover, *Toxoplasma gondii* was demonstrated to induce endoplasmic reticulum (ER) stress as well as intestinal barrier disruption, pathologies associated with mental illness [110,111].

Taken together, lactate is a CNS protective metabolite that not only contributes to energy metabolism and signaling but also serves as a substrate for lactylation. Excessive lactate accumulation from local metabolism or the microbiome may trigger lactylation-associated pathology, such as cancer, AD, or neuropsychiatric illness.

3. Oxytocin and Microglia in Mental Illness

Oxytocin (OXT)-oxytocin receptor (OXTR) signaling comprises a microglial anti-inflammatory pathway. OXT is a neuropeptide synthesized in the hypothalamus and stored in the posterior pituitary prior to being released into the systemic circulation. OXT is both a hormone and a neurotransmitter, the former mediating parturition and lactation, while the latter affects social behavior, affiliation, intimacy, and mother–infant bonding [112]. A recent study found that the OXT concentration in the brain can reach levels 1000 times higher than in the systemic circulation, highlighting the importance of this biomolecule for CNS function [113].

Low blood OXT levels were reported in patients with SCZ marked by negative symptoms, while intranasal administration of OXT was shown to ameliorate this condition [114,115]. Indeed, like lactate, OXT modulates synaptic plasticity and upregulates adult hippocampal neurogenesis, enhancing long-term potentiation and memory, properties that likely mediate this hormone’s antipsychotic and antidepressant properties [116–118].

OXT decreases microglial inflammation by lowering ER stress via translation initiation factor 2 alpha/transcription factor ATF4 (eIF-2 α -ATF4), a common autophagic pathway disrupted in SCZ and major depressive disorder (MDD) [119–121]. For example, eIF-2 α -ATF4 lowers ER stress by upregulating Sigma-1 receptors (Sig-1Rs), proteins involved in

several neuropsychiatric disorders. Indeed, several antidepressant and antipsychotic drugs, including fluvoxamine, sertraline, and haloperidol, lower ER stress, suggesting that the therapeutic properties of these agents could be mediated by eIF-2 α -ATF4 [122,123]. Aside from lowering ER stress, Sig-1Rs protect the intestinal barrier and BBB, linking eIF-2 α -ATF4 to microbial translocation [124,125]. It stands to reason, therefore, that strengthening the gut barrier and lowering ER stress in intestinal epithelial cells (IECs) are emerging strategies for neuropsychiatric disorders (Table 1) (clinical trials identifier NCT03183609) [126–131]).

Table 1. The action mechanisms of Lactate, OXT, and AhR.

Lactate	OXT	AhR	References
Deactivates microglia by NF-kB inhibition	Deactivates microglia via eIF-2 α -ATF4 pathway	Deactivates microglia via NF-kB	[4,15,50]
Excess lactate promotes K κ a	No known effect on lactylation	Increases LDHA and lactate	[24,132]
Lowers ER stress	Lowers ER stress	Lowers ER stress	[16,33,133]
Augments gut barrier function	Augments gut barrier	Augments gut barrier	[90,124,134]
Upregulated in schizophrenia	Downregulated in schizophrenia	Implicated in schizophrenia	[1,135,136]

Several studies have connected OXT with the selective serotonin reuptake inhibitors (SSRIs), emphasizing that this hormone may contribute, at least in part, to the action mechanism of antidepressant drugs [137,138]. Moreover, sexual dysfunction, a common SSRI adverse effect, was associated with decreased OXT blood levels, linking this hormone further to the serotonergic system [139,140]. This is significant as OXT exerts antidepressant properties of its own and may be indicated in situations when other antidepressants are to be avoided, such as during pregnancy and the postpartum period as well as in patients with sexual dysfunction.

Aside from the antidepressant action, OXT exerts antipsychotic properties, documented as early as the 1970s and 1980s, followed by clinical trials of intranasal OXT in 2010, studies which have produced encouraging results for schizophrenia patients [35,135,141] (clinical trials identifier NCT01621737).

In 2022, it was discovered that a subpopulation of microglial cells is capable of generating OXT, drawing the attention of researchers and clinicians to the anti-inflammatory properties of these cells [33,34]. Indeed, OXT signaling with OXTR, expressed on microglia, astrocytes, and oligodendrocytes, lowers neuroinflammation, depression, suicidal behavior, and psychosis, indicating that OXT signaling is a significant neuropsychiatric target [35–44] (see section Microglia, lactylation and mental illness).

Taken together, OXT and lactate exert antipsychotic and antidepressant properties through different mechanisms. Impaired OXT signaling may trigger neuropathology by increasing ER stress in IECs, disrupting the GI tract barrier.

4. Aryl Hydrocarbon Receptor and Mental Illness

ILCs are mucosa-anchored lymphocytes which express transcription factors instead of T- or B-cell receptors; they are activated by specific cytokines and generate their own cytokine output. ILCs, comprised of NKC, ILCs types 1, 2, 3, and regulatory ILCs (ILCreg), are situated at the biological barriers, including the gut and meninges [142,143]. Human ILCs express AhR which is abundantly represented on CD56bright NKC, emphasizing further the protective role of these lymphocytes [53,144]. Glycolytic enzymes, lactate dehydrogenase A (LDHA) or lactate dehydrogenase-5 (LDH-5), activate NKG2D receptors, regulating CD56bright NKC responses, suggesting that excessive lactate likely disrupts the function of these lymphocytes [144,145].

Aside from AhR and lactate, NKCs, including CD56bright, are activated by IL-12, IL-15, and IL-18, releasing interferon γ (IFN- γ) and IL-10 (Figure 2) [1–4]. In addition, AhR lowers the proinflammatory IL-17 previously implicated in both SCZ and MS [146–148].

Moreover, dysfunctional IL-12, IL-15, IL-18, IFN- γ , and IL-10 have been associated with schizophrenia, highlighting further the key role NKC in this pathology [149,150]. AhR also suppresses microglial and astrocytic inflammation, switching these cells from inflammatory to the restorative phenotype [51,151,152].

Aberrant elimination of resting microglia by NKCs is likely the root cause of neuroinflammation, documented in many neuropsychiatric disorders; therefore, rescuing these cells should be explored as a therapeutic strategy [53,54]. For example, the expansion of circulating CD56bright NKCs has been associated with improved MS symptoms, suggesting that these cells may also play a beneficial role in SCZ and affective disorders [153,154]. Indeed, a low CD56 lymphocyte count was demonstrated in antipsychotic-naïve patients with schizophrenia, and it was upregulated by treatment, emphasizing the beneficial properties of these cells [134] (for the role of AhR in microglial cells, see the section Microglia, lactylation and mental illness). As CD56bright NKCs express AhR, the clinical symptoms may respond to the pharmacological manipulation of this receptor [155] (discussed in the section on Potential Therapeutic Strategies).

In the gut, AhR protects the intestinal barrier by binding microbiota-derived metabolites as well as the SCFAs, linking dysfunctional AhR signaling to gut barrier disruption and microbial translocation [156,157].

Novel studies have shown that AhR functions in a tissue-specific manner, dependent on the ligand affinity, likely accounting for the variable, often opposite, responses elicited in one organ vs. another [158,159]. For example, endogenous AhR ligands, such as tryptophan and microbial metabolites, may exacerbate breast cancer while exerting salutary effects in colorectal carcinoma [132,160–163]. Moreover, in the gut, AhR enhances the barrier function while inducing liver toxicity, illustrating the difficulties encountered in trying to develop AhR therapeutics. Having said that, selective modulators, such as flavonoids and isoflavones, may offer a solution [164–167] (Table 2) (see the section on potential interventions).

Table 2. Potential interventions for restoring the homeostasis of anti-inflammatory microglia.

Lactate	OXT	AhR	References
L-lactate	Intranasal OXT	Flavonoids	[114,168,169]
Lactylation inhibitors	ARBs	Nattokinase	[169–171]
Demethylzeylasteral (DML)	ISRIB	Isoflavones	[172–174]
Sirtuins	PAK inhibitors	Laquinimod	[170,175,176]
Sodium oxamate	MIF-1	CH 22391	[177–179]
Gallnut extract CN102836354A	OT-1 (TC OT 39)	Triptans (Sumatriptan)	[180–182]
Galloflavin (NSC 107022)	WAY 267464	CD56bright NKCs	[183–185]

AhR promotes myelination and lowers ER stress, suggesting that MS, MDD, schizophrenia, and bipolar disorder may be exacerbated by the dysfunction of this protein [133,186]. Interestingly, carbidopa and the unique antipsychotic drug clozapine are AhR agonists, linking this receptor to the dopaminergic system and probably explaining both the efficacy as well as the side effects of these agents [187,188]. Moreover, dopamine is an AhR ligand, further implicating this receptor in mental illness [136]. Indeed, AhR was implicated in schizophrenia by the earlier studies, emphasizing a novel psychopharmacological target [189]. In a rodent model, clozapine attenuates experimental autoimmune encephalomyelitis (EAE), suggesting that the unique anti-suicide properties of this drug (not shared with other antipsychotic agents) may be mediated by AhR [190]. Yet another study saw that agranulocytosis, an autoimmune-like adverse effect of clozapine, was associated with DRB1*15:01, a rare human leukocyte antigen (HLA), and a major MS risk factor, linking agranulocytosis to this genetic marker [191,192].

Taken together, AhR exerts antidepressant and antipsychotic properties by several mechanisms: 1. microglial deactivation, 2. lowering ER stress, 3. enhancing the gut barrier, 4. upregulating lactate, and 5. myelination. These protective effects may explain the unique clozapine properties (Table 1).

5. Microbial Translocation Outside the GI Tract

In our previous work, we have discussed microbial translocation from the GI tract into the host systemic circulation, suggesting that the pathogenesis of several idiopathic diseases could be explained by the immune responses to extraintestinal microbial proteins [193]. As higher gut and BBB permeability markers, including the soluble form of CD14 (sCD14), were documented in neuropsychiatric illness, this pathology may, at least in part, result from microbial migration through the gut barrier [194–196].

Various antigens of GI tract microbes, including *Bacteroides*, *Bifidobacterium*, *Acinetobacter*, and *Pseudomonas*, have been shown to exhibit molecular mimicry with the host myelin basic protein (MBP), suggesting that autoantibodies against myelin, documented in MS, schizophrenia, and bipolar disorder, may be conventional immunoglobulins directed at the displaced microbes or their components [197–199]. In addition, as myelin contains a serotonin binding site, the autoantibodies against serotonin, found in MS and MDD, may be classical antibodies against serotonin-producing microbes, such as *Clostridia* or *staphylococci* [200,201]. Furthermore, bacterial N-acetylmuramyl dipeptide was reported to mimic myelin, likely triggering antibodies upon translocation into host tissues [202]. These findings are significant as they emphasize that strengthening the gut barrier to lower microbial migration likely comprises a novel therapeutic strategy in neuropsychiatric disorders. In fact, inflammation in response to translocated microbial proteins was previously associated with suicidal behavior, suggesting that gut barrier restoration should be instituted as a therapy in mental illness [203–205]. Indeed, some antidepressant drugs currently utilized in clinical practice were demonstrated to optimize the gut barrier function, suggesting an alternative, noncanonical, mechanism of action [206]. Microbial translocation across the gut barrier has been documented in MDD, suicidal behavior, bipolar disorder, aggression in SCZ, and neurodegeneration, suggesting that the loss of anti-inflammatory microglia plays a crucial role in neuropathology [196,202–205,207,208].

6. Potential Therapeutic Strategies

Lactate, OXT, and AhR are therapeutic targets rarely considered in the treatment of neuropsychiatric disorders; however, they play a major role in the CNS and immune metabolism.

7. Lactate

Novel studies have shown that peripheral administration of L-lactate exerts antidepressant effects by increasing hippocampal lactic acid which upregulates neurogenesis and inhibits microglial activation [168,209] (Table 2).

1. Lactylation inhibitors are compounds belonging to several classes of therapeutics, such as LDHA inhibitors, demethylzylasteral (DML), and phytotherapeutics, including polyphenols. Here, we review only the agents that can cross the BBB, lower ER stress, and strengthen the intestinal barrier [210,211]. These are as follows.
2. Sirtuins

Studies in oncology have reported that sirtuins (SIRT), especially SIRT2, can remove lactate from the histone lysine, showing that de-lactylation may comprise a novel strategy for microglial depolarization, an approach likely beneficial in MDD and SCZ [170]. The effect of SIRT in these disorders has been documented by earlier research [212–214].

Demethylzylasteral (DML) inhibits lactylation of histone H3, a protein associated with MDD, suggesting that it could exert antidepressant properties [172,215].

3. LDH inhibitors Lactate-HCAR1 signaling is regulated by LDHA or LDH-5, glycolytic enzymes upregulated by psychological stress, suggesting that inhibiting these en-

zymes could be therapeutic for neuropsychiatric illness [216]. LDH inhibitors, include the following:

- 3.1. Sodium oxamate, a structural analog of pyruvate, is known for lowering ER stress by promoting protective autophagy, properties beneficial for neuropsychiatric disorders [177,217]. To the best of our knowledge this compound has not been evaluated for psychiatric conditions.
- 3.2. Chinese Gallnut Extract from *Galla chinensis* is a traditional Chinese medicine and a potent LDHA inhibitor which was patented for the treatment of MDD and is currently in clinical trials (Chinese patent CN102836354A) (US patent 10098854B2) (clinical trials identifier NCT04080752) [180,218].
- 3.3. Galloflavin or NSC 107022 is a polyphenol found in berries which upregulates SIRT6, a key regulator of neuronal mitochondria [183,219]. Dysfunctional SIRT6 was associated with depression-like behavior in mice, suggesting potential benefits for humans with MDD [220]. Galloflavin has been patented as a modulator of protein kinases that can treat a variety of conditions, ranging from cancer to neuropsychiatric disorders (US patent 20030187007A1).

8. OXT

OXT signaling with OXTRs has been shown to inhibit activated microglia and increase the number of NKC, a phenomenon documented in schizophrenia [221]. Recently, an OXT-generating microglial population was identified, suggesting that anti-inflammatory microglia may regulate many physiological functions that could cause pathology when disrupted [222,223].

8.1. Angiotensin IV (ANG IV) as an OXT Agonist

Several studies demonstrated that microglia express a functional renin-angiotensin system (RAS), including angiotensin II type 1 receptors (AT-1Rs) [224]. Conversely, AT-1R blockers (ARBs), especially the BBB-crossing candesartan, have been found therapeutic in schizophrenia and MDD, linking dysfunctional RAS to inflamed microglia [171]. This is significant as angiotensin II (ANG II) is a negative OXT regulator; thus, ARBs may not only depolarize these cells but also enhance OXT release [33,34,224,225]. Indeed, angiotensin IV was reported to upregulate OXT in vitro and in vivo, probably by inhibiting oxytocinase, the hydrolyzing enzyme of this hormone [226].

OXT inhibits eIF-2 α -ATF4 signaling, a pathway implicated in ER stress, suggesting that the beneficial neuropsychiatric effects of OXT can likely be attributed to autophagy [44]. Indeed, the antiretroviral drug nelfinavir has been shown to promote ER stress by eIF2 α phosphorylation; in contrast, the integrated stress response inhibitor (ISRIB), a protein with antidepressant and antipsychotic properties, inhibits nelfinavir-induced ER stress via eIF2 α dephosphorylation [173]. The related drugs, p21-activated kinase (PAK) inhibitors, were found beneficial in SCZ, emphasizing new potential targets in this disorder [175]. IRIB was patented in Australia as a broad therapeutic agent that may be useful in pathologies involving learning, memory, immunity, intermediary metabolism, insulin production, and resistance to unfolded protein stress (Australian patent 2020229748A1).

8.2. OXT Fragments

OXT, comprised of nine amino acids, was found to retain biological properties even when cleaved in shorter segments. For example, melanocyte-inhibiting factor-1 (MIF-1), the N-terminal segment of oxytocin, possesses antidepressant and antipsychotic properties, suggesting that OXTR can be activated by related small ligands [178,227]. MIF-1 was patented for cancer, autoimmune diseases, fibrotic diseases, inflammation, and neurodegenerative diseases (international patent WO2009040045A2). Several OXT-like molecules have also been studied, including lipo-oxytocin-1 (LOT-1), a synthetic OXTR agonist (also known as TC OT 39), and WAY-267464, an OXTR agonist and vasopressin 1 receptor (V1AR) antagonist [181,184]. The efficacy of these ligands for neuropsychiatric disorders has not been

evaluated in clinical trials; however, several patents were obtained for use in neurological and psychiatric disorders (international patent WO2018107216A1) (EP2326341B1).

9. AhR

AhR binds diverse ligands derived from the environment, diet, microbes, and cellular metabolism, enabling cells to adapt to the changing exogenous or endogenous conditions. Drugs, chemicals, and plant-derived compounds, including isoflavones, attach with different affinities to AhR [228]. Nattokinase, a soybean fermented by *Bacillus subtilis var. natto*, contains AhR-binding flavonoids [169]. Indeed, several isoflavones, including, soybeans, genistein, daidzein, glycitein, formononetin, biochanin, and phytoestrogens, have been found to exert antidepressant and antipsychotic properties [174,229,230].

- Laquinimod, a quinolone-3-carboxamide developed for the treatment of MS, is a selective AhR ligand that demonstrated encouraging results in animal models of MDD and anxiety [176].
- CH223191 is an antagonistic selective AhR ligand with a favorable profile for neuropsychiatric disorders [179]. This compound was patented in the US for cancer treatment but has still to enter clinical trials for neuropsychiatric disorders (US patent 20160175278A1).
- Sumatriptan, a selective 5-hydroxy-triptamine (5-HT1) receptor agonist, activates human AhR, exerting anti-inflammatory properties, suggesting beneficial effects on activated microglia [182,231].
- Dimethyl fumarate, indicated for MS, is not an AhR ligand; however, it upregulates the protective AhR-expressing CD56bright NKCs which eliminate the detrimental autoreactive immune cells to avert autoimmunity [232,233]. For this reason, CD56bright NKCs expansion via AhR or transplantation may comprise a potential therapeutic strategy for neuropsychiatric illness. For example, CD56bright NK cells allogeneic or stem cell transplantation, a treatment used for acute myeloid leukemia, could be therapeutic in both MS and schizophrenia [185,234].

10. Microglia, Lactylation, and Mental Illness

Although microglia comprise less than 10% of the brain cells, it plays a major role in neuropsychiatric pathology [235]. Resting microglia are neuroprotective as evidenced by in vivo two-photon imaging of fluorescent-labeled neurons and microglia, demonstrating that these cells participate in the plasticity of neuronal circuits [236]. Resting microglia actively communicate with other brain cells, including astrocytes, neurons, and ILCs [237–239]. Recent studies have shown that microglia express monocarboxylate transporters (MCTs) and HCAR1, suggesting that lactate plays an important role in the reprogramming of these cells [240,241]. Numerous studies have found that young and healthy brains rely on glycolysis and lactate, while aging brains lose glycolytic ability [242]. For example, dysfunctional glycolysis was associated with AD pathology as well as schizophrenia [243,244]. Excessive lactate generation contributes to lactylation of histone and nonhistone proteins, engendering pathology [245].

Microglia are brain resident macrophages that express OX40 receptors, suggesting that these proteins contribute to anti-inflammatory microglia, a likely protection against mental illness [246,247].

Microglia express AhR, proteins involved in several pathologies, including glioblastoma [248]. AhR is a new receptor of interest in schizophrenia, not only because it binds dopamine but also because schizophrenia-associated exogenous toxins, such as plasticizers, are ligands at this receptor [249,250]. In addition, many protective metabolites synthesized by gut microbes, including indole and tryptophan, are AhR ligands, suggesting that the gut–brain axis likely operates through this receptor [251].

It has been reported that aberrantly activated microglia become neurotoxic and engage in the elimination of healthy neurons or synapses, a pathology involved in schizophrenia [252–254].

11. Limitations

Many compounds described above are phytotherapeutics, primarily natural products, such as sirtuins, or traditional foods, including nattokinase and the Chinese Gallnut Extract, that have been used by humans for hundreds of years. The ability of these molecules to act as AhR ligands suggests beneficial effects in neuropsychiatric illness, and clinical trials should be initiated to assess their efficacy. Likewise, the ability of sirtuins to remove histone lysine lactate has reawakened the interest in these compounds, previously studied for neurodegenerative disorders, as potential medical foods or dietary interventions.

Furthermore, several compounds mentioned above have been patented in different countries and some are currently in clinical trials, while others have not reached that stage yet. However, the profile of these agents, estimated by their action mechanism described in the patent application, suggests that they should be evaluated as antidepressant or antipsychotic agents. For example, dimethyl fumarate was patented in Europe for the treatment of MS; however, as this drug exerts neuroprotective as well as anti-inflammatory effects on astrocytes and microglia, it likely possesses antidepressant and antipsychotic properties which should be evaluated in clinical trials.

Additionally, we need to state that most of the pathophysiological mechanisms described in this article are newly elucidated. As the details become increasingly available, it would be much easier to prioritize different potential therapeutic targets. Moreover, understanding these mechanisms will eventually allow for an in-depth stratification of neuropsychiatric patients, and a move toward an improved precision medicine approach.

12. Conclusions

Increasing awareness of the shortcomings of the neurotransmitter paradigm contributed to the development of a new concept, metabolism-driven information processing. This model, illustrated by the previously known association of neuropsychiatric pathology with excessive aerobic glycolysis, was put in perspective by the discovery of ILCs and lactylation. The consequences of excessive brain lactate and increased local acidity include the following:

1. The aberrant elimination of anti-inflammatory resting microglia by pathologically activated NKC.
2. The reactivation of the neuronal cell cycle with resultant aneuploidy or apoptosis as these cells lack the molecular machinery to complete mitosis.
3. Several interventions may restore the homeostasis of anti-inflammatory microglia, including 1. lowering lactylation, 2. enhancing OXT signaling, 3. modulating AhR, and 4. expanding CD56NKs. These strategies lower the ER stress and strengthen the gut barrier, limiting microbial migration outside of the GI tract, thus decreasing the risk of microglial activation by translocated bacterial antigens.

More studies are needed to clarify the role of lactylation of histone and nonhistone proteins in mental illnesses and the interaction of lactylation with meningeal ILCs.

Where authors are identified as personnel of the International Agency for Research on Cancer/WHO, the authors alone are responsible for the views expressed in this article and they do not necessarily represent the decisions, policy, or views of the International Agency for Research on Cancer/WHO.

Author Contributions: Conceptualization, A.S., C.K. and Z.K.; methodology, C.V.A.; formal analysis, J.J.A. and C.V.A.; data curation, J.J.A.; writing—original draft preparation, A.S.; writing—review and editing, C.K. and Z.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: Where authors are identified as personnel of the International Agency for Research on Cancer/WHO, the authors alone are responsible for the views expressed in this article and they do not necessarily represent the decisions, policy or views of the International Agency for Research on Cancer/WHO.

Abbreviations

NF- κ B = nuclear factor kappa B; HCAR1 = hydroxycarboxylic acid receptor 1; GPR81 = G-protein-coupled receptor 81; K_{la} = lysine lactylation; OXT = oxytocin; OXTRs = receptors; AhR = aryl hydrocarbon receptor; NKCs = natural killer cells; ERM = ezrin-radixin-moesin; HDAC1 = class I histone deacetylases; ILC = innate lymphoid cells; HMGB1 = high-mobility group box 1; HIF-1 α = hypoxia-inducible factor 1 α ; BDNF = brain-derived neurotrophic factor; IEC = intestinal epithelial cells; SCFA = short-chain fatty acids; ISRIB = integrated stress response inhibitor; MIF-1 = melanocyte-inhibiting factor-1; OXPHOS = oxidative phosphorylation; V1AR = vasopressin 1 receptor; SCZ = schizophrenia.

References

1. Rowland, L.M.; Pradhan, S.; Korenic, S.; Wijtenburg, S.A.; Hong, L.E.; Edden, R.A.; Barker, P.B. Elevated brain lactate in schizophrenia: A 7 T magnetic resonance spectroscopy study. *Transl. Psychiatry* **2016**, *6*, e967. [[CrossRef](#)] [[PubMed](#)]
2. Dogan, A.E.; Yuksel, C.; Du, F.; Chouinard, V.-A.; Öngür, D. Brain lactate and pH in schizophrenia and bipolar disorder: A systematic review of findings from magnetic resonance studies. *Neuropsychopharmacology* **2018**, *43*, 1681–1690. [[CrossRef](#)] [[PubMed](#)]
3. Sullivan, C.R.; Mielnik, C.A.; Funk, A.; O'Donovan, S.M.; Bentea, E.; Pletnikov, M.; Ramsey, A.J.; Wen, Z.; Rowland, L.M.; McCullumsmith, R.E. Measurement of lactate levels in postmortem brain, iPSCs, and animal models of schizophrenia. *Sci. Rep.* **2019**, *9*, 5087. [[CrossRef](#)]
4. Pruett, B.S.; Meador-Woodruff, J.H. Evidence for altered energy metabolism, increased lactate, and decreased pH in schizophrenia brain: A focused review and meta-analysis of human postmortem and magnetic resonance spectroscopy studies. *Schizophr. Res.* **2020**, *223*, 29–42. [[CrossRef](#)]
5. Vallée, A.; Vallée, J.-N. Warburg effect hypothesis in autism Spectrum disorders. *Mol. Brain* **2018**, *11*, 1. [[CrossRef](#)]
6. Park, H.-J.; Choi, I.; Leem, K.-H. Decreased Brain pH and Pathophysiology in Schizophrenia. *Int. J. Mol. Sci.* **2021**, *22*, 8358. [[CrossRef](#)]
7. Li, J.; Chen, L.; Qin, Q.; Wang, D.; Zhao, J.; Gao, H.; Yuan, X.; Zhang, J.; Zou, Y.; Mao, Z.; et al. Upregulated hexokinase 2 expression induces the apoptosis of dopaminergic neurons by promoting lactate production in Parkinson's disease. *Neurobiol. Dis.* **2022**, *163*, 105605. [[CrossRef](#)]
8. Hollnagel, J.-O.; Cesetti, T.; Schneider, J.; Vazetdinova, A.; Valiullina-Rakhmatullina, F.; Lewen, A.; Rozov, A.; Kann, O. Lactate Attenuates Synaptic Transmission and Affects Brain Rhythms Featuring High Energy Expenditure. *iScience* **2020**, *23*, 101316. [[CrossRef](#)]
9. Krzyściak, W.; Karcz, P.; Bystrowska, B.; Sz wajca, M.; Bryll, A.; Śmierciak, N.; Ligezka, A.; Turek, A.; Kozicz, T.; Skalniak, A.E.; et al. The Association of the Oral Microbiota with the Effects of Acid Stress Induced by an Increase of Brain Lactate in Schizophrenia Patients. *Biomedicines* **2023**, *11*, 240. [[CrossRef](#)]
10. Ghosh, S.; Castillo, E.; Frias, E.S.; Swanson, R.A. Bioenergetic regulation of microglia. *Glia* **2017**, *66*, 1200–1212. [[CrossRef](#)]
11. Nimmerjahn, A.; Kirchhoff, F.; Helmchen, F. Resting Microglial Cells Are Highly Dynamic Surveillants of Brain Parenchyma in Vivo. *Science* **2005**, *308*, 1314–1318. [[CrossRef](#)] [[PubMed](#)]
12. Block, M.L.; Zecca, L.; Hong, J.-S. Microglia-mediated neurotoxicity: Uncovering the molecular mechanisms. *Nat. Rev. Neurosci.* **2007**, *8*, 57–69. [[CrossRef](#)] [[PubMed](#)]
13. Tauffenberger, A.; Fiumelli, H.; Almustafa, S.; Magistretti, P.J. Lactate and pyruvate promote oxidative stress resistance through hormetic ROS signaling. *Cell Death Dis.* **2019**, *10*, 653. [[CrossRef](#)] [[PubMed](#)]
14. Monsorno, K.; Buckinx, A.; Paolicelli, R.C. Microglial metabolic flexibility: Emerging roles for lactate. *Trends Endocrinol. Metab.* **2022**, *33*, 186–195. [[CrossRef](#)] [[PubMed](#)]
15. Tang, Y.; Le, W. Differential Roles of M1 and M2 Microglia in Neurodegenerative Diseases. *Mol. Neurobiol.* **2016**, *53*, 1181–1194. [[CrossRef](#)]
16. Karnib, N.; El-Ghandour, R.; El Hayek, L.; Nasrallah, P.; Khalifeh, M.; Barmo, N.; Jabre, V.; Ibrahim, P.; Bilen, M.; Stephan, J.S.; et al. Lactate is an antidepressant that mediates resilience to stress by modulating the hippocampal levels and activity of histone deacetylases. *Neuropsychopharmacology* **2019**, *44*, 1152–1162. [[CrossRef](#)] [[PubMed](#)]
17. Carrard, A.; Elsayed, M.; Margineanu, M.; Boury-Jamot, B.; Fragnière, L.; Meylan, E.M.; Petit, J.-M.; Fiumelli, H.; Magistretti, P.J.; Martin, J.-L. Peripheral administration of lactate produces antidepressant-like effects. *Mol. Psychiatry* **2016**, *23*, 392–399. [[CrossRef](#)] [[PubMed](#)]

18. Yang, K.; Fan, M.; Wang, X.; Xu, J.; Wang, Y.; Tu, F.; Gill, P.S.; Ha, T.; Liu, L.; Williams, D.L.; et al. Lactate promotes macrophage HMGB1 lactylation, acetylation, and exosomal release in polymicrobial sepsis. *Cell Death Differ.* **2021**, *29*, 133–146. [[CrossRef](#)]
19. Gao, Y.; Zhou, H.; Liu, G.; Wu, J.; Yuan, Y.; Shang, A. Tumor Microenvironment: Lactic Acid Promotes Tumor Development. *J. Immunol. Res.* **2022**, *2022*, 3119375. [[CrossRef](#)]
20. Ishihara, S.; Hata, K.; Hirose, K.; Okui, T.; Toyosawa, S.; Uzawa, N.; Nishimura, R.; Yoneda, T. The lactate sensor GPR81 regulates glycolysis and tumor growth of breast cancer. *Sci. Rep.* **2022**, *12*, 6261. [[CrossRef](#)]
21. Ichihara, Y.; Doi, T.; Ryu, Y.; Nagao, M.; Sawada, Y.; Ogata, T. Oligodendrocyte Progenitor Cells Directly Utilize Lactate for Promoting Cell Cycling and Differentiation. *J. Cell. Physiol.* **2016**, *232*, 986–995. [[CrossRef](#)] [[PubMed](#)]
22. Cerexhe, L.; Easton, C.; Macdonald, E.; Renfrew, L.; Sculthorpe, N. Blood lactate concentrations during rest and exercise in people with Multiple Sclerosis: A systematic review and meta-analysis. *Mult. Scler. Relat. Disord.* **2021**, *57*, 103454. [[CrossRef](#)] [[PubMed](#)]
23. Keytsman, C.; Hansen, D.; Wens, I.; Eijnde, B.O. Exercise-induced lactate responses in Multiple Sclerosis: A retrospective analysis. *Neurorehabilitation* **2019**, *45*, 99–106. [[CrossRef](#)] [[PubMed](#)]
24. Pan, R.-Y.; He, L.; Zhang, J.; Liu, X.; Liao, Y.; Gao, J.; Liao, Y.; Yan, Y.; Li, Q.; Zhou, X.; et al. Positive feedback regulation of microglial glucose metabolism by histone H4 lysine 12 lactylation in Alzheimer's disease. *Cell Metab.* **2022**, *34*, 634–648.e6. [[CrossRef](#)]
25. Xie, Y.; Hu, H.; Liu, M.; Zhou, T.; Cheng, X.; Huang, W.; Cao, L. The role and mechanism of histone lactylation in health and diseases. *Front. Genet.* **2022**, *13*, 949252. [[CrossRef](#)]
26. Liberti, M.V.; Locasale, J.W. The Warburg Effect: How Does it Benefit Cancer Cells? *Trends Biochem. Sci.* **2016**, *41*, 211–218, Erratum in *Trends Biochem. Sci.* **2016**, *41*, 287. [[CrossRef](#)]
27. Zhao, Y.; Xu, H. Microglial lactate metabolism as a potential therapeutic target for Alzheimer's disease. *Mol. Neurodegener.* **2022**, *17*, 36. [[CrossRef](#)]
28. Sabari, B.R.; Zhang, D.; Allis, C.D.; Zhao, Y. Metabolic regulation of gene expression through histone acylations. *Nat. Rev. Mol. Cell Biol.* **2017**, *18*, 90–101. [[CrossRef](#)]
29. Zhang, D.; Tang, Z.; Huang, H.; Zhou, G.; Cui, C.; Weng, Y.; Liu, W.; Kim, S.; Lee, S.; Perez-Neut, M.; et al. Metabolic regulation of gene expression by histone lactylation. *Nature* **2019**, *574*, 575–580. [[CrossRef](#)]
30. Okazaki, S.; Boku, S.; Otsuka, I.; Mouri, K.; Aoyama, S.; Shiroywa, K.; Sora, I.; Fujita, A.; Shirai, Y.; Shirakawa, O.; et al. The cell cycle-related genes as biomarkers for schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2016**, *70*, 85–91. [[CrossRef](#)]
31. Luo, Y.; Yang, Z.; Yu, Y.; Zhang, P. HIF1 α lactylation enhances KIAA1199 transcription to promote angiogenesis and vasculogenic mimicry in prostate cancer. *Int. J. Biol. Macromol.* **2022**, *222*, 2225–2243. [[CrossRef](#)] [[PubMed](#)]
32. Andersson, U.; Yang, H. HMGB1 is a critical molecule in the pathogenesis of Gram-negative sepsis. *J. Intensive Med.* **2022**, *2*, 156–166. [[CrossRef](#)] [[PubMed](#)]
33. Klein, B.Y.; Tamir, H.; Ludwig, R.J.; Glickstein, S.B.; Welch, M.G. Colostrum oxytocin modulates cellular stress response, inflammation, and autophagy markers in newborn rat gut villi. *Biochem. Biophys. Res. Commun.* **2017**, *487*, 47–53. [[CrossRef](#)] [[PubMed](#)]
34. Maejima, Y.; Yokota, S.; Ono, T.; Yu, Z.; Yamachi, M.; Hidema, S.; Nollet, K.E.; Nishimori, K.; Tomita, H.; Yaginuma, H.; et al. Identification of oxytocin expression in human and murine microglia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2022**, *119*, 110600. [[CrossRef](#)]
35. Pedersen, C.A.; Gibson, C.M.; Rau, S.W.; Salimi, K.; Smedley, K.L.; Casey, R.L.; Leserman, J.; Jarskog, L.F.; Penn, D.L. Intranasal oxytocin reduces psychotic symptoms and improves Theory of Mind and social perception in schizophrenia. *Schizophr. Res.* **2011**, *132*, 50–53. [[CrossRef](#)]
36. Macdonald, K.; Feifel, D. Oxytocin in schizophrenia: A review of evidence for its therapeutic effects. *Acta Neuropsychiatr.* **2012**, *24*, 130–146. [[CrossRef](#)]
37. Caldwell, H.K.; Stephens, S.L.; Young, W.S. Oxytocin as a natural antipsychotic: A study using oxytocin knockout mice. *Mol. Psychiatry* **2009**, *14*, 190–196. [[CrossRef](#)]
38. Jahangard, L.; Shayganfar, M.; Ghiasi, F.; Salehi, I.; Haghighi, M.; Ahmadpanah, M.; Bahmani, D.S.; Brand, S. Serum oxytocin concentrations in current and recent suicide survivors are lower than in healthy controls. *J. Psychiatr. Res.* **2020**, *128*, 75–82. [[CrossRef](#)]
39. Panaro, M.A.; Benameur, T.; Porro, C. Hypothalamic Neuropeptide Brain Protection: Focus on Oxytocin. *J. Clin. Med.* **2020**, *9*, 1534. [[CrossRef](#)]
40. Parris, M.S.; Grunebaum, M.F.; Galfalvy, H.C.; Andronikashvili, A.; Burke, A.K.; Yin, H.; Min, E.; Huang, Y.-Y.; Mann, J.J. Attempted suicide and oxytocin-related gene polymorphisms. *J. Affect. Disord.* **2018**, *238*, 62–68. [[CrossRef](#)]
41. Baudon, A.; Creusot, E.C.; Althammer, F.; Schaaf, C.P.; Charlet, A. Emerging role of astrocytes in oxytocin-mediated control of neural circuits and brain functions. *Prog. Neurobiol.* **2022**, *217*, 102328. [[CrossRef](#)] [[PubMed](#)]
42. Jokinen, J.; Chatzittofis, A.; Hellström, C.; Nordström, P.; Uvnäs-Moberg, K.; Åsberg, M. Low CSF oxytocin reflects high intent in suicide attempters. *Psychoneuroendocrinology* **2012**, *37*, 482–490. [[CrossRef](#)] [[PubMed](#)]
43. Yuan, L.; Liu, S.; Bai, X.; Gao, Y.; Liu, G.; Wang, X.; Liu, D.; Li, T.; Hao, A.; Wang, Z. Oxytocin inhibits lipopolysaccharide-induced inflammation in microglial cells and attenuates microglial activation in lipopolysaccharide-treated mice. *J. Neuroinflammation* **2016**, *13*, 77. [[CrossRef](#)] [[PubMed](#)]

44. Inoue, Y.U.; Miwa, H.; Hori, K.; Kaneko, R.; Morimoto, Y.; Koike, E.; Asami, J.; Kamijo, S.; Yamada, M.; Hoshino, M.; et al. Targeting Neurons with Functional Oxytocin Receptors: A Novel Set of Simple Knock-In Mouse Lines for Oxytocin Receptor Visualization and Manipulation. *Eneuro* **2022**, *9*, ENEURO.0423-21.2022. [[CrossRef](#)] [[PubMed](#)]
45. Mairesse, J.; Zinni, M.; Pansiot, J.; Hassan-Abdi, R.; Demene, C.; Colella, M.; Charriaut-Marlangue, C.; Novais, A.R.B.; Tanter, M.; Maccari, S.; et al. Oxytocin receptor agonist reduces perinatal brain damage by targeting microglia. *Glia* **2018**, *67*, 345–359. [[CrossRef](#)] [[PubMed](#)]
46. De Cagna, F.; Fusar-Poli, L.; Damiani, S.; Rocchetti, M.; Giovanna, G.; Mori, A.; Politi, P.; Brondino, N. The Role of Intranasal Oxytocin in Anxiety and Depressive Disorders: A Systematic Review of Randomized Controlled Trials. *Clin. Psychopharmacol. Neurosci.* **2019**, *17*, 1–11. [[CrossRef](#)]
47. Sorg, O. AhR signalling and dioxin toxicity. *Toxicol. Lett.* **2014**, *230*, 225–233. [[CrossRef](#)]
48. Climaco-Arvizu, S.; Domínguez-Acosta, O.; Cabañas-Cortés, M.A.; Rodríguez-Sosa, M.; Gonzalez, F.J.; Vega, L.; Elizondo, G. Aryl hydrocarbon receptor influences nitric oxide and arginine production and alters M1/M2 macrophage polarization. *Life Sci.* **2016**, *155*, 76–84. [[CrossRef](#)]
49. Sahebnaasagh, A.; Hashemi, J.; Khoshi, A.; Saghafi, F.; Avan, R.; Faramarzi, F.; Azimi, S.; Habtemariam, S.; Sureda, A.; Khayatkashani, M.; et al. Aromatic hydrocarbon receptors in mitochondrial biogenesis and function. *Mitochondrion* **2021**, *61*, 85–101. [[CrossRef](#)]
50. Di Giaimo, R.; Durovic, T.; Barquin, P.; Kociaj, A.; Lepko, T.; Aschenbroich, S.; Breunig, C.T.; Irmmler, M.; Cernilogar, F.M.; Schotta, G.; et al. The Aryl Hydrocarbon Receptor Pathway Defines the Time Frame for Restorative Neurogenesis. *Cell Rep.* **2018**, *25*, 3241–3251.e5. [[CrossRef](#)]
51. Tanaka, M.; Fujikawa, M.; Oguro, A.; Itoh, K.; Vogel, C.F.A.; Ishihara, Y. Involvement of the Microglial Aryl Hydrocarbon Receptor in Neuroinflammation and Vasogenic Edema after Ischemic Stroke. *Cells* **2021**, *10*, 718. [[CrossRef](#)] [[PubMed](#)]
52. Fan, X.; Wang, S.; Hu, S.; Yang, B.; Zhang, H. Host-microbiota interactions: The aryl hydrocarbon receptor in the acute and chronic phases of cerebral ischemia. *Front. Immunol.* **2022**, *13*, 967300. [[CrossRef](#)] [[PubMed](#)]
53. Moreno-Nieves, U.Y.; Mundy, D.C.; Shin, J.H.; Tam, K.; Sunwoo, J.B. The aryl hydrocarbon receptor modulates the function of human CD56^{bright} NK cells. *Eur. J. Immunol.* **2018**, *48*, 771–776. [[CrossRef](#)] [[PubMed](#)]
54. Lünemann, A.; Lünemann, J.D.; Roberts, S.; Messmer, B.; da Silva, R.B.; Raine, C.S.; Münz, C. Human NK Cells Kill Resting but Not Activated Microglia via NKG2D- and Nkp46-Mediated Recognition. *J. Immunol.* **2008**, *181*, 6170–6177. [[CrossRef](#)]
55. Peter, K.; Rehli, M.; Singer, K.; Renner-Sattler, K.; Kreutz, M. Lactic acid delays the inflammatory response of human monocytes. *Biochem. Biophys. Res. Commun.* **2015**, *457*, 412–418. [[CrossRef](#)]
56. Hagihara, H.; Shoji, H.; Otabi, H.; Toyoda, A.; Katoh, K.; Namihira, M.; Miyakawa, T. Protein lactylation induced by neural excitation. *Cell Rep.* **2021**, *37*, 109820. [[CrossRef](#)]
57. Jiang, J.; Huang, D.; Jiang, Y.; Hou, J.; Tian, M.; Li, J.; Sun, L.; Zhang, Y.; Zhang, T.; Li, Z.; et al. Lactate Modulates Cellular Metabolism Through Histone Lactylation-Mediated Gene Expression in Non-Small Cell Lung Cancer. *Front. Oncol.* **2021**, *11*, 647559. [[CrossRef](#)]
58. Mertens, J.; Herdy, J.R.; Traxler, L.; Schafer, S.T.; Schlachetzki, J.C.M.; Böhnke, L.; Reid, D.A.; Lee, H.; Zangwill, D.; Fernandes, D.P.; et al. Age-dependent instability of mature neuronal fate in induced neurons from Alzheimer's patients. *Cell Stem Cell* **2021**, *28*, 1533–1548.e6. [[CrossRef](#)]
59. Yurov, Y.B.; Vorsanova, S.G.; Demidova, I.; Kolotii, A.D.; Soloviev, I.V.; Iourov, I.Y. Mosaic Brain Aneuploidy in Mental Illnesses: An Association of Low-level post-zygotic Aneuploidy with Schizophrenia and Comorbid Psychiatric Disorders. *Curr. Genom.* **2018**, *19*, 163–172. [[CrossRef](#)]
60. Shegay, P.V.; Zabolotneva, A.A.; Shatova, O.P.; Shestopalov, A.V.; Kaprin, A.D. Evolutionary View on Lactate-Dependent Mechanisms of Maintaining Cancer Cell Stemness and Reprimitivization. *Cancers* **2022**, *14*, 4552. [[CrossRef](#)]
61. Fan, Y.; Abrahamsen, G.; McGrath, J.J.; Mackay-Sim, A. Altered Cell Cycle Dynamics in Schizophrenia. *Biol. Psychiatry* **2012**, *71*, 129–135. [[CrossRef](#)] [[PubMed](#)]
62. Katsel, P.; Davis, K.L.; Li, C.; Tan, W.; Greenstein, E.; Hoffman, L.B.K.; Haroutunian, V. Abnormal Indices of Cell Cycle Activity in Schizophrenia and their Potential Association with Oligodendrocytes. *Neuropsychopharmacology* **2008**, *33*, 2993–3009. [[CrossRef](#)] [[PubMed](#)]
63. Lineweaver, C.H.; Bussey, K.J.; Blackburn, A.C.; Davies, P.C.W. Cancer progression as a sequence of atavistic reversions. *Bioessays* **2021**, *43*, e2000305. [[CrossRef](#)]
64. Alvarez, Z.; Hyroššová, P.; Perales, J.C.; Alcántara, S.; Ova, P.H. Neuronal Progenitor Maintenance Requires Lactate Metabolism and PEPCK-M-Directed Cataplerosis. *Cereb. Cortex* **2014**, *26*, 1046–1058. [[CrossRef](#)] [[PubMed](#)]
65. Odenwelder, D.C.; Lu, X.; Harcum, S.W. Induced pluripotent stem cells can utilize lactate as a metabolic substrate to support proliferation. *Biotechnol. Prog.* **2020**, *37*, e3090. [[CrossRef](#)]
66. Goldman, S.; Pulsinelli, W.A.; Clarke, W.Y.; Kraig, R.P.; Plum, F. The Effects of Extracellular Acidosis on Neurons and Glia in vitro. *J. Cereb. Blood Flow Metab.* **1989**, *9*, 471–477. [[CrossRef](#)]
67. Tan, Z.; Chu, D.Z.V.; Chan, Y.J.A.; Lu, Y.E.; Rancati, G. Mammalian Cells Undergo Endoreduplication in Response to Lactic Acidosis. *Sci. Rep.* **2018**, *8*, 2890. [[CrossRef](#)]
68. Lev-Vachnish, Y.; Cadury, S.; Rotter-Maskowitz, A.; Feldman, N.; Roichman, A.; Illouz, T.; Varvak, A.; Nicola, R.; Madar, R.; Okun, E. L-Lactate Promotes Adult Hippocampal Neurogenesis. *Front. Neurosci.* **2019**, *13*, 403. [[CrossRef](#)]

69. Ordoño, J.; Pérez-Amodio, S.; Ball, K.; Aguirre, A.; Engel, E. The generation of a lactate-rich environment stimulates cell cycle progression and modulates gene expression on neonatal and hiPSC-derived cardiomyocytes. *Biomater. Adv.* **2022**, *139*, 213035. [[CrossRef](#)]
70. Rosenblatt, J. Mitosis: Moesin and the Importance of Being Round. *Curr. Biol.* **2008**, *18*, R292–R293. [[CrossRef](#)]
71. Congdon, E.E.; Sigurdsson, E.M. Tau-targeting therapies for Alzheimer disease. *Nat. Rev. Neurol.* **2018**, *14*, 399–415. [[CrossRef](#)] [[PubMed](#)]
72. Beckmann, A.; Ramirez, P.; Gamez, M.; Gonzalez, E.; De Mange, J.; Bieniek, K.F.; Ray, W.J.; Frost, B. Moesin is an effector of tau-induced actin overstabilization, cell cycle activation, and neurotoxicity in Alzheimer’s disease. *iScience* **2023**, *26*, 106152. [[CrossRef](#)] [[PubMed](#)]
73. Gu, J.; Zhou, J.; Chen, Q.; Xu, X.; Gao, J.; Li, X.; Shao, Q.; Zhou, B.; Zhou, H.; Wei, S.; et al. Tumor metabolite lactate promotes tumorigenesis by modulating MOESIN lactylation and enhancing TGF- β signaling in regulatory T cells. *Cell Rep.* **2022**, *40*, 111122. [[CrossRef](#)] [[PubMed](#)]
74. Satooka, H.; Matsui, M.; Ichioka, S.; Nakamura, Y.; Hirata, T. The ERM protein moesin regulates natural killer cell homeostasis in vivo. *Cell. Immunol.* **2022**, *371*, 104456. [[CrossRef](#)] [[PubMed](#)]
75. Ben-Shmuel, A.; Sabag, B.; Biber, G.; Barda-Saad, M. The Role of the Cytoskeleton in Regulating the Natural Killer Cell Immune Response in Health and Disease: From Signaling Dynamics to Function. *Front. Cell Dev. Biol.* **2021**, *9*, 609532. [[CrossRef](#)] [[PubMed](#)]
76. Bowden, N.A.; Weidenhofer, J.; Scott, R.J.; Schall, U.; Todd, J.; Michie, P.T.; Tooney, P.A. Preliminary investigation of gene expression profiles in peripheral blood lymphocytes in schizophrenia. *Schizophr. Res.* **2006**, *82*, 175–183. [[CrossRef](#)]
77. Moreno-Yruela, C.; Zhang, D.; Wei, W.; Bæk, M.; Liu, W.; Gao, J.; Danková, D.; Nielsen, A.L.; Bolding, J.E.; Yang, L.; et al. Class I histone deacetylases (HDAC1–3) are histone lysine delactylases. *Sci. Adv.* **2022**, *8*, eabi6696. [[CrossRef](#)]
78. Yu, J.; de Belle, I.; Liang, H.; Adamson, E.D. Coactivating Factors p300 and CBP Are Transcriptionally Crossregulated by Egr1 in Prostate Cells, Leading to Divergent Responses. *Mol. Cell* **2004**, *15*, 83–94. [[CrossRef](#)]
79. Gilbert, T.; Zürcher, N.R.; Wu, C.J.; Bhanot, A.; Hightower, B.G.; Kim, M.; Albrecht, D.S.; Wey, H.-Y.; Schroeder, F.A.; Rodriguez-Thompson, A.; et al. PET neuroimaging reveals histone deacetylase dysregulation in schizophrenia. *J. Clin. Investig.* **2018**, *129*, 364–372. [[CrossRef](#)]
80. Martínez-Pinteño, A.; Gassó, P.; Prohens, L.; Segura, A.G.; Parellada, M.; Saiz-Ruiz, J.; Cuesta, M.J.; Bernardo, M.; Lafuente, A.; Mas, S.; et al. Identification of EP300 as a Key Gene Involved in Antipsychotic-Induced Metabolic Dysregulation Based on Integrative Bioinformatics Analysis of Multi-Tissue Gene Expression Data. *Front. Pharmacol.* **2021**, *12*, 729474. [[CrossRef](#)]
81. Tan, M.; Shen, L.; Hou, Y. Epigenetic modification of BDNF mediates neuropathic pain via miR-30a-3p/EP300 axis in CCI rats. *Biosci. Rep.* **2020**, *40*, BSR20194442. [[CrossRef](#)] [[PubMed](#)]
82. Gören, J.L. Brain-derived neurotrophic factor and schizophrenia. *Ment. Health Clin.* **2016**, *6*, 285–288. [[CrossRef](#)] [[PubMed](#)]
83. Bredy, T.W.; Wu, H.; Crego, C.; Zellhoefer, J.; Sun, Y.E.; Barad, M. Histone modifications around individual BDNF gene promoters in prefrontal cortex are associated with extinction of conditioned fear. *Learn. Mem.* **2007**, *14*, 268–276. [[CrossRef](#)] [[PubMed](#)]
84. Fan, H.; Yang, F.; Xiao, Z.; Luo, H.; Chen, H.; Chen, Z.; Liu, Q.; Xiao, Y. Lactylation: Novel epigenetic regulatory and therapeutic opportunities. *Am J Physiol Endocrinol Metab.* **2023**, *324*, E330–E338. [[CrossRef](#)] [[PubMed](#)]
85. Boutahar, N.; Reynaud, E.; Lassabliere, F.; Borg, J. Brain-derived neurotrophic factor inhibits cell cycle reentry but not endoplasmic reticulum stress in cultured neurons following oxidative or excitotoxic stress. *J. Neurosci. Res.* **2010**, *88*, 2263–2271. [[CrossRef](#)] [[PubMed](#)]
86. Sedgwick, A.J.; Ghazanfari, N.; Constantinescu, P.; Mantamadiotis, T.; Barrow, A.D. The Role of NK Cells and Innate Lymphoid Cells in Brain Cancer. *Front. Immunol.* **2020**, *11*, 1549. [[CrossRef](#)]
87. Marzan, D.E.; Brügger-Verdon, V.; West, B.L.; Liddelow, S.; Samanta, J.; Salzer, J.L. Activated microglia drive demyelination via CSF1R signaling. *Glia* **2021**, *69*, 1583–1604. [[CrossRef](#)]
88. Gaffney, D.O.; Jennings, E.Q.; Anderson, C.C.; Marentette, J.O.; Shi, T.; Oxvig, A.-M.S.; Streeter, M.D.; Johannsen, M.; Spiegel, D.A.; Chapman, E.; et al. Non-enzymatic Lysine Lactoylation of Glycolytic Enzymes. *Cell Chem. Biol.* **2020**, *27*, 206–213.e6. [[CrossRef](#)]
89. Tarantino, N.; Leboyer, M.; Bouleau, A.; Hamdani, N.; Richard, J.R.; Boukouaci, W.; Ching-Lien, W.; Godin, O.; Bengoufa, D.; Le Corvoisier, P.; et al. Natural killer cells in first-episode psychosis: An innate immune signature? *Mol. Psychiatry* **2021**, *26*, 5297–5306. [[CrossRef](#)]
90. Lee, Y.-S.; Kim, T.-Y.; Kim, Y.; Lee, S.-H.; Kim, S.; Kang, S.W.; Yang, J.-Y.; Baek, I.-J.; Sung, Y.H.; Park, Y.-Y.; et al. Microbiota-Derived Lactate Accelerates Intestinal Stem-Cell-Mediated Epithelial Development. *Cell Host Microbe* **2018**, *24*, 833–846.e6. [[CrossRef](#)]
91. Wang, S.P.; Rubio, L.A.; Duncan, S.; Donachie, G.E.; Holtrop, G.; Lo, G.; Farquharson, F.M.; Wagner, J.; Parkhill, J.; Louis, P.; et al. Pivotal Roles for pH, Lactate, and Lactate-Utilizing Bacteria in the Stability of a Human Colonic Microbial Ecosystem. *Msystems* **2020**, *5*, e00645-20. [[CrossRef](#)] [[PubMed](#)]
92. Tang, C.-F.; Wang, C.-Y.; Wang, J.-H.; Wang, Q.-N.; Li, S.-J.; Wang, H.-O.; Zhou, F.; Li, J.-M. Short-Chain Fatty Acids Ameliorate Depressive-like Behaviors of High Fructose-Fed Mice by Rescuing Hippocampal Neurogenesis Decline and Blood–Brain Barrier Damage. *Nutrients* **2022**, *14*, 1882. [[CrossRef](#)] [[PubMed](#)]
93. Caslin, H.L.; Ababayehu, D.; Pinette, J.A.; Ryan, J.J. Lactate Is a Metabolic Mediator That Shapes Immune Cell Fate and Function. *Front. Physiol.* **2021**, *12*, 688485. [[CrossRef](#)] [[PubMed](#)]

94. Kennedy, L.; Glesaaen, E.R.; Palibrk, V.; Pannone, M.; Wang, W.; Al-Jabri, A.; Suganthan, R.; Meyer, N.; Austbø, M.L.; Lin, X.; et al. Lactate receptor HCAR1 regulates neurogenesis and microglia activation after neonatal hypoxia-ischemia. *Elife* **2022**, *11*, e76451. [[CrossRef](#)] [[PubMed](#)]
95. Errea, A.; Cayet, D.; Marchetti, P.; Tang, C.; Kluza, J.; Offermanns, S.; Sirard, J.-C.; Rumbo, M. Lactate Inhibits the Pro-Inflammatory Response and Metabolic Reprogramming in Murine Macrophages in a GPR81-Independent Manner. *PLoS ONE* **2016**, *11*, e0163694. [[CrossRef](#)]
96. Nicola, R.; Madar, R.; Okun, E. HCAR1-Mediated L-Lactate Signaling Suppresses Microglial Phagocytosis. *NeuroMol. Med.* **2022**, *24*, 399–404. [[CrossRef](#)]
97. Dong, H.; Zhang, J.; Zhang, H.; Han, Y.; Lu, C.; Chen, C.; Tan, X.; Wang, S.; Bai, X.; Zhai, G.; et al. YiaC and CobB regulate lysine lactylation in *Escherichia coli*. *Nat. Commun.* **2022**, *13*, 6628. [[CrossRef](#)]
98. Wiwanitkit, V. Psychosis and *E. coli* Infection: A Forgotten Issue. *Indian J. Psychol. Med.* **2012**, *34*, 407–408. [[CrossRef](#)]
99. Kleimann, A.; Toto, S.; Eberlein, C.K.; Kielstein, J.T.; Bleich, S.; Frieling, H.; Sieberer, M. Psychiatric Symptoms in Patients with Shiga Toxin-Producing *E. coli* O104:H4 Induced Haemolytic-Uraemic Syndrome. *PLoS ONE* **2014**, *9*, e101839. [[CrossRef](#)]
100. Fouladkhah, A.; Geornaras, I.; Yang, H.; Sofos, J.N. Lactic acid resistance of Shiga toxin-producing *Escherichia coli* and multidrug-resistant and susceptible *Salmonella* Typhimurium and *Salmonella* Newport in meat homogenate. *Food Microbiol.* **2013**, *36*, 260–266. [[CrossRef](#)]
101. Graham, K.L.; Carson, C.M.; Ezeoke, A.; Buckley, P.F.; Miller, B.J. Urinary Tract Infections in Acute Psychosis. *J. Clin. Psychiatry* **2014**, *75*, 379–385. [[CrossRef](#)] [[PubMed](#)]
102. Thomas, J.; McEvoy, J.P.; Miller, B.J. Urinary tract infection, inflammation, and cognition in phase 1 of the Clinical Antipsychotic Trials of Intervention Effectiveness Study. *Ann. Clin. Psychiatry* **2019**, *31*, 242–248. [[PubMed](#)]
103. Tadié, J.-M.; Bae, H.-B.; Banerjee, S.; Zmijewski, J.W.; Abraham, E. Differential activation of RAGE by HMGB1 modulates neutrophil-associated NADPH oxidase activity and bacterial killing. *Am. J. Physiol. -Cell Physiol.* **2012**, *302*, C249–C256. [[CrossRef](#)]
104. Lin, A.E.; Beasley, F.C.; Olson, J.; Keller, N.; Shalwitz, R.A.; Hannan, T.; Hultgren, S.J.; Nizet, V. Role of Hypoxia Inducible Factor-1 α (HIF-1 α) in Innate Defense against Uropathogenic *Escherichia coli* Infection. *PLoS Pathog.* **2015**, *11*, e1004818. [[CrossRef](#)]
105. Manosalva, C.; Quiroga, J.; Hidalgo, A.I.; Alarcón, P.; Anseoleaga, N.; Hidalgo, M.A.; Burgos, R.A. Role of Lactate in Inflammatory Processes: Friend or Foe. *Front Immunol.* **2022**, *12*, 808799. [[CrossRef](#)]
106. Al-Dujaili, A.H.; Mousa, R.F.; Al-Hakeim, H.K.; Maes, M. High Mobility Group Protein 1 and Dickkopf-Related Protein 1 in Schizophrenia and Treatment-Resistant Schizophrenia: Associations With Interleukin-6, Symptom Domains, and Neurocognitive Impairments. *Schizophr. Bull.* **2020**, *47*, 530–541. [[CrossRef](#)]
107. Torrey, E.F.; Yolken, R.H. *Toxoplasma gondii* and Schizophrenia. *Emerg. Infect. Dis.* **2003**, *9*, 1375–1380. [[CrossRef](#)]
108. Zhao, W.; Yu, H.; Liu, X.; Wang, T.; Yao, Y.; Zhou, Q.; Zheng, X.; Tan, F. Systematic identification of the lysine lactylation in the protozoan parasite *Toxoplasma gondii*. *Parasites Vectors* **2022**, *15*, 180. [[CrossRef](#)]
109. Carrillo, G.L.; Ballard, V.A.; Glausen, T.; Boone, Z.; Teamer, J.; Hinkson, C.L.; Wohlfert, E.A.; Blader, I.J.; Fox, M.A. *Toxoplasma* infection induces microglia-neuron contact and the loss of perisomatic inhibitory synapses. *Glia* **2020**, *68*, 1968–1986. [[CrossRef](#)]
110. Obed, C.; Wu, M.; Chen, Y.; An, R.; Cai, H.; Luo, Q.; Yu, L.; Wang, J.; Liu, F.; Shen, J.; et al. *Toxoplasma gondii* dense granule protein 3 promotes endoplasmic reticulum stress-induced apoptosis by activating the PERK pathway. *Parasites Vectors* **2022**, *15*, 276. [[CrossRef](#)]
111. Briceño, M.P.; Nascimento, L.A.C.; Nogueira, N.P.; Barenco, P.V.C.; Ferro, E.A.V.; Rezende-Oliveira, K.; Goulart, L.R.; Alves, P.T.; Barbosa, B.D.F.; Lima, W.R.; et al. *Toxoplasma gondii* Infection Promotes Epithelial Barrier Dysfunction of Caco-2 Cells. *J. Histochem. Cytochem.* **2016**, *64*, 459–469. [[CrossRef](#)] [[PubMed](#)]
112. Walter, M.H.; Abele, H.; Plappert, C.F. The Role of Oxytocin and the Effect of Stress During Childbirth: Neurobiological Basics and Implications for Mother and Child. *Front. Endocrinol.* **2021**, *12*, 742236. [[CrossRef](#)] [[PubMed](#)]
113. Ludwig, M.; Leng, G. Dendritic peptide release and peptide-dependent behaviours. *Nat. Rev. Neurosci.* **2006**, *7*, 126–136. [[CrossRef](#)] [[PubMed](#)]
114. Sabe, M.; Zhao, N.; Crippa, A.; Strauss, G.P.; Kaiser, S. Intranasal Oxytocin for Negative Symptoms of Schizophrenia: Systematic Review, Meta-Analysis, and Dose-Response Meta-Analysis of Randomized Controlled Trials. *Int. J. Neuropsychopharmacol.* **2021**, *24*, 601–614. [[CrossRef](#)] [[PubMed](#)]
115. Kéri, S.; Kiss, I.; Kelemen, O. Sharing secrets: Oxytocin and trust in schizophrenia. *Soc. Neurosci.* **2009**, *4*, 287–293. [[CrossRef](#)]
116. Matsuzaki, M.; Matsushita, H.; Tomizawa, K.; Matsui, H. Oxytocin: A therapeutic target for mental disorders. *J. Physiol. Sci.* **2012**, *62*, 441–444. [[CrossRef](#)]
117. Lin, Y.-T.; Chen, C.-C.; Huang, C.-C.; Nishimori, K.; Hsu, K.-S. Oxytocin stimulates hippocampal neurogenesis via oxytocin receptor expressed in CA3 pyramidal neurons. *Nat. Commun.* **2017**, *8*, 537. [[CrossRef](#)]
118. Marlin, B.J.; Froemke, R.C. Oxytocin modulation of neural circuits for social behavior. *Dev. Neurobiol.* **2016**, *77*, 169–189. [[CrossRef](#)]
119. Costa-Mattioli, M.; Gobert, D.; Stern, E.; Gamache, K.; Colina, R.; Cuello, C.; Sossin, W.; Kaufman, R.; Pelletier, J.; Rosenblum, K.; et al. eIF2 α Phosphorylation Bidirectionally Regulates the Switch from Short- to Long-Term Synaptic Plasticity and Memory. *Cell* **2007**, *129*, 195–206. [[CrossRef](#)]
120. Trinh, M.A.; Kaphzan, H.; Wek, R.C.; Pierre, P.; Cavener, D.R.; Klann, E. Brain-Specific Disruption of the eIF2 α Kinase PERK Decreases ATF4 Expression and Impairs Behavioral Flexibility. *Cell Rep.* **2012**, *1*, 676–688. [[CrossRef](#)]

121. Timberlake, M., II; Dwivedi, Y. Linking unfolded protein response to inflammation and depression: Potential pathologic and therapeutic implications. *Mol. Psychiatry* **2019**, *24*, 987–994. [[CrossRef](#)] [[PubMed](#)]
122. Mitsuda, T.; Omi, T.; Tanimukai, H.; Sakagami, Y.; Tagami, S.; Okochi, M.; Kudo, T.; Takeda, M. Sigma-1Rs are upregulated via PERK/eIF2 α /ATF4 pathway and execute protective function in ER stress. *Biochem. Biophys. Res. Commun.* **2011**, *415*, 519–525. [[CrossRef](#)] [[PubMed](#)]
123. Cobos, E.J.; Entrena, J.M.; Nieto, F.R.; Cendán, C.M.; Del Pozo, E. Pharmacology and Therapeutic Potential of Sigma1 Receptor Ligands. *Curr. Neuropharmacol.* **2008**, *6*, 344–366. [[CrossRef](#)]
124. Almási, N.; Török, S.; Dvorácskó, S.; Tömböly, C.; Csonka, Á.; Baráth, Z.; Murlasits, Z.; Valkusz, Z.; Pósa, A.; Varga, C.; et al. Lessons on the Sigma-1 Receptor in TNBS-Induced Rat Colitis: Modulation of the UCHL-1, IL-6 Pathway. *Int. J. Mol. Sci.* **2020**, *21*, 4046. [[CrossRef](#)]
125. Liu, D.; Yang, L.; Liu, P.; Ji, X.; Qi, X.; Wang, Z.; Chi, T.; Zou, L. Sigma-1 receptor activation alleviates blood–brain barrier disruption post cerebral ischemia stroke by stimulating the GDNF–GFR α 1–RET pathway. *Exp. Neurol.* **2021**, *347*, 113867. [[CrossRef](#)] [[PubMed](#)]
126. Kowalczyk, M.; Kowalczyk, E.; Kwiatkowski, P.; Łopusiewicz, Ł.; Talarowska, M.; Sienkiewicz, M. Cellular Response to Unfolded Proteins in Depression. *Life* **2021**, *11*, 1376. [[CrossRef](#)]
127. Kamarudin, M.N.A.; Parhar, I. Emerging therapeutic potential of anti-psychotic drugs in the management of human glioma: A comprehensive review. *Oncotarget* **2019**, *10*, 3952–3977. [[CrossRef](#)]
128. Liśkiewicz, P.; Kaczmarczyk, M.; Misiak, B.; Wroński, M.; Bąba-Kubiś, A.; Skonieczna-Żydecka, K.; Marlicz, W.; Bienkowski, P.; Misera, A.; Pełka-Wysiecka, J.; et al. Analysis of gut microbiota and intestinal integrity markers of inpatients with major depressive disorder. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2020**, *106*, 110076. [[CrossRef](#)]
129. Mao, J.; Hu, Y.; Ruan, L.; Ji, Y.; Lou, Z. Role of endoplasmic reticulum stress in depression (Review). *Mol. Med. Rep.* **2019**, *20*, 4774–4780. [[CrossRef](#)]
130. Patel, S.; Sharma, D.; Kalia, K.; Tiwari, V. Crosstalk between endoplasmic reticulum stress and oxidative stress in schizophrenia: The dawn of new therapeutic approaches. *Neurosci. Biobehav. Rev.* **2017**, *83*, 589–603. [[CrossRef](#)]
131. Hayashi, T. Conversion of psychological stress into cellular stress response: Roles of the sigma-1 receptor in the process. *Psychiatry Clin. Neurosci.* **2015**, *69*, 179–191. [[CrossRef](#)] [[PubMed](#)]
132. Safe, S.; Jin, U.-H.; Park, H.; Chapkin, R.S.; Jayaraman, A. Aryl Hydrocarbon Receptor (AHR) Ligands as Selective AHR Modulators (SAHRMs). *Int. J. Mol. Sci.* **2020**, *21*, 6654. [[CrossRef](#)] [[PubMed](#)]
133. Shackelford, G.; Sampathkumar, N.K.; Hichor, M.; Weill, L.; Meffre, D.; Juricek, L.; Laurendeau, I.; Chevallier, A.; Ortonne, N.; Larousserie, F.; et al. Involvement of Aryl hydrocarbon receptor in myelination and in human nerve sheath tumorigenesis. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E1319–E1328. [[CrossRef](#)] [[PubMed](#)]
134. Saraste, M.; Irjala, H.; Airas, L. Expansion of CD56Bright natural killer cells in the peripheral blood of multiple sclerosis patients treated with interferon-beta. *Neurol. Sci.* **2007**, *28*, 121–126. [[CrossRef](#)] [[PubMed](#)]
135. Feifel, D.; Macdonald, K.; Nguyen, A.; Cobb, P.; Warlan, H.; Galangue, B.; Minassian, A.; Becker, O.; Cooper, J.; Perry, W.; et al. Adjunctive Intranasal Oxytocin Reduces Symptoms in Schizophrenia Patients. *Biol. Psychiatry* **2010**, *68*, 678–680. [[CrossRef](#)] [[PubMed](#)]
136. Schubert, K.O.; Föcking, M.; Cotter, D.R. Proteomic pathway analysis of the hippocampus in schizophrenia and bipolar affective disorder implicates 14-3-3 signaling, aryl hydrocarbon receptor signaling, and glucose metabolism: Potential roles in GABAergic interneuron pathology. *Schizophr. Res.* **2015**, *167*, 64–72. [[CrossRef](#)]
137. Uvnäs-Moberg, K.; Björkstrand, E.; Hillegaard, V.; Ahlenius, S. Oxytocin as a possible mediator of SSRI-induced antidepressant effects. *Psychopharmacology* **1999**, *142*, 95–101. [[CrossRef](#)]
138. Galbally, M.; Watson, S.J.; Keelan, J.A.; Spigset, O.; Lewis, A. The relationship between oxytocin blood concentrations and antidepressants over pregnancy and the postpartum. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2020**, *109*, 110218. [[CrossRef](#)]
139. Humble, M.B.; Bejerot, S. Orgasm, Serotonin Reuptake Inhibition, and Plasma Oxytocin in Obsessive-Compulsive Disorder. Gleaning From a Distant Randomized Clinical Trial. *Sex Med.* **2016**, *4*, e145–e155. [[CrossRef](#)]
140. Abbasiazari, M.; Heidari-Kord, M.; Mazaheri-Meybodi, A.; Eshraghi, A.; Bayati, N. Plasma Oxytocin Level and Sexual Dysfunction in Depressed Women Treated by Either Fluoxetine or Citalopram: A Pilot Clinical Trial. *Iran. J. Pharm. Res.* **2018**, *17*, 408–414.
141. Bujanow, W. Is Oxytocin an Anti-Schizophrenic Hormone? *Can. Psychiatr. Assoc. J.* **1974**, *19*, 323. [[CrossRef](#)] [[PubMed](#)]
142. Zheng, M.; Zhu, J. Innate Lymphoid Cells and Intestinal Inflammatory Disorders. *Int. J. Mol. Sci.* **2022**, *23*, 1856. [[CrossRef](#)] [[PubMed](#)]
143. Robinette, M.L.; Fuchs, A.; Cortez, V.S.; Lee, J.S.; Wang, Y.; Durum, S.K.; Gilfillan, S.; Colonna, M.; Immunological Genome Consortium. Transcriptional programs define molecular characteristics of innate lymphoid cell classes and subsets. *Nat. Immunol.* **2015**, *16*, 306–317. [[CrossRef](#)]
144. Crane, C.A.; Austgen, K.; Habarthur, K.; Hofmann, C.; Moyes, K.W.; Avanesyan, L.; Fong, L.; Campbell, M.J.; Cooper, S.; Oakes, S.A.; et al. Immune evasion mediated by tumor-derived lactate dehydrogenase induction of NKG2D ligands on myeloid cells in glioblastoma patients. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 12823–12828. [[CrossRef](#)]
145. Wensveen, F.M.; Jelenčić, V.; Polić, B. NKG2D: A Master Regulator of Immune Cell Responsiveness. *Front. Immunol.* **2018**, *9*, 441. [[CrossRef](#)] [[PubMed](#)]

146. Borovcanin, M.M.; Minic Janicijevic, S.; Jovanovic, I.P.; Gajovic, N.M.; Jurisevic, M.M.; Arsenijevic, N.N. Type 17 Immune Response Facilitates Progression of Inflammation and Correlates with Cognition in Stable Schizophrenia. *Diagnostics* **2020**, *10*, 926. [[CrossRef](#)] [[PubMed](#)]
147. Matusевич, D.; Kivisäkk, P.; He, B.; Kostulas, N.; Özenci, V.; Fredrikson, S.; Link, H. Interleukin-17 mRNA expression in blood and CSF mononuclear cells is augmented in multiple sclerosis. *Mult. Scler. J.* **1999**, *5*, 101–104. [[CrossRef](#)] [[PubMed](#)]
148. Dawidowski, B.; Górnjak, A.; Podwalski, P.; Lebiecka, Z.; Misiak, B.; Samochowicz, J. The Role of Cytokines in the Pathogenesis of Schizophrenia. *J. Clin. Med.* **2021**, *10*, 3849. [[CrossRef](#)]
149. Al-Asmary, S.M.; Kadasah, S.; Arfin, M.; Tariq, M.; Al-Asmari, A. Genetic variants of interleukin-10 gene promoter are associated with schizophrenia in Saudi patients: A case-control study. *North Am. J. Med. Sci.* **2014**, *6*, 558–565. [[CrossRef](#)]
150. Warre-Cornish, K.; Perfect, L.; Nagy, R.; Duarte, R.R.R.; Reid, M.J.; Raval, P.; Mueller, A.; Evans, A.L.; Couch, A.; Ghevaert, C.; et al. Interferon- γ signaling in human iPSC-derived neurons recapitulates neurodevelopmental disorder phenotypes. *Sci. Adv.* **2020**, *6*, eaay9506. [[CrossRef](#)]
151. Ramirez, J.-M.; Brembilla, N.C.; Sorg, O.; Chicheportiche, R.; Matthes, T.; Dayer, J.-M.; Saurat, J.-H.; Roosnek, E.; Chizzolini, C. Activation of the aryl hydrocarbon receptor reveals distinct requirements for IL-22 and IL-17 production by human T helper cells. *Eur. J. Immunol.* **2010**, *40*, 2450–2459. [[CrossRef](#)] [[PubMed](#)]
152. Chen, W.C.; Chang, L.H.; Huang, S.S.; Huang, Y.J.; Chih, C.L.; Kuo, H.C.; Lee, Y.H.; Lee, I.H. Aryl hydrocarbon receptor modulates stroke-induced astrogliosis and neurogenesis in the adult mouse brain. *J. Neuroinflamm.* **2019**, *16*, 187. [[CrossRef](#)]
153. van Venrooij, J.A.E.M.; Fluitman, S.B.A.H.A.; Lijmer, J.G.; Kavelaars, A.; Heijnen, C.J.; Westenberg, H.G.M.; Kahn, R.S.; Wied, C.C.G.-D. Impaired Neuroendocrine and Immune Response to Acute Stress in Medication-Naive Patients With a First Episode of Psychosis. *Schizophr. Bull.* **2012**, *38*, 272–279. [[CrossRef](#)] [[PubMed](#)]
154. Gross, C.C.; Ahmetpahic, D.; Ruck, T.; Schulte-Mecklenbeck, A.; Schwarte, K.; Jörgens, S.; Scheu, S.; Windhagen, S.; Graefe, B.; Melzer, N.; et al. Alemtuzumab treatment alters circulating innate immune cells in multiple sclerosis. *Neurol. Neuroimmunol. Neuroinflamm.* **2016**, *3*, e289. [[CrossRef](#)]
155. Chu, C.-S.; Li, D.-J.; Chu, C.-L.; Wu, C.-C.; Lu, T. Decreased IL-1ra and NCAM-1/CD56 Serum Levels in Unmedicated Patients with Schizophrenia Before and After Antipsychotic Treatment. *Psychiatry Investig.* **2018**, *15*, 727–732. [[CrossRef](#)] [[PubMed](#)]
156. Postal, B.G.; Ghezzal, S.; Aguanno, D.; André, S.; Garbin, K.; Genser, L.; Brot-Laroche, E.; Poitou, C.; Soula, H.; Leturque, A.; et al. AhR activation defends gut barrier integrity against damage occurring in obesity. *Mol. Metab.* **2020**, *39*, 101007. [[CrossRef](#)]
157. Ghiboub, M.; Verburgt, C.M.; Sovran, B.; Benninga, M.A.; de Jonge, W.J.; Van Limbergen, J.E. Nutritional Therapy to Modulate Tryptophan Metabolism and Aryl Hydrocarbon-Receptor Signaling Activation in Human Diseases. *Nutrients* **2020**, *12*, 2846. [[CrossRef](#)]
158. Marinelli, L.; Martin-Gallausiaux, C.; Bourhis, J.-M.; Béguet-Crespel, F.; Blottière, H.M.; Lapaque, N. Identification of the novel role of butyrate as AhR ligand in human intestinal epithelial cells. *Sci. Rep.* **2019**, *9*, 643. [[CrossRef](#)]
159. Tagliabue, S.G.; Faber, S.C.; Motta, S.; Denison, M.S.; Bonati, L. Modeling the binding of diverse ligands within the Ah receptor ligand binding domain. *Sci. Rep.* **2019**, *9*, 10693. [[CrossRef](#)]
160. Wu, K.K. Cytoguardin: A Tryptophan Metabolite against Cancer Growth and Metastasis. *Int. J. Mol. Sci.* **2021**, *22*, 4490. [[CrossRef](#)]
161. Peyraud, F.; Guegan, J.-P.; Bodet, D.; Cousin, S.; Bessede, A.; Italiano, A. Targeting Tryptophan Catabolism in Cancer Immunotherapy Era: Challenges and Perspectives. *Front. Immunol.* **2022**, *13*, 807271. [[CrossRef](#)] [[PubMed](#)]
162. Juhász, C.; Nahleh, Z.; Zitron, I.; Chugani, D.C.; Janabi, M.Z.; Bandyopadhyay, S.; Ali-Fehmi, R.; Mangner, T.J.; Chakraborty, P.K.; Mittal, S.; et al. Tryptophan metabolism in breast cancers: Molecular imaging and immunohistochemistry studies. *Nucl. Med. Biol.* **2012**, *39*, 926–932. [[CrossRef](#)] [[PubMed](#)]
163. Papadimitriou, N.; Gunter, M.J.; Murphy, N.; Gicquiau, A.; Achaintre, D.; Brezina, S.; Gumpenberger, T.; Baierl, A.; Ose, J.; Geijssen, A.J.M.R.; et al. Circulating tryptophan metabolites and risk of colon cancer: Results from case-control and prospective cohort studies. *Int. J. Cancer* **2021**, *149*, 1659–1669. [[CrossRef](#)] [[PubMed](#)]
164. Pessione, E. Lactic acid bacteria contribution to gut microbiota complexity: Lights and shadows. *Front Cell Infect Microbiol.* **2012**, *2*, 86. [[CrossRef](#)]
165. Takeda, T.; Komiya, Y.; Koga, T.; Ishida, T.; Ishii, Y.; Kikuta, Y.; Nakaya, M.; Kurose, H.; Yokomizo, T.; Shimizu, T.; et al. Dioxin-induced increase in leukotriene B4 biosynthesis through the aryl hydrocarbon receptor and its relevance to hepatotoxicity owing to neutrophil infiltration. *J. Biol. Chem.* **2017**, *292*, 10586–10599. [[CrossRef](#)]
166. Yu, M.; Wang, Q.; Ma, Y.; Li, L.; Yu, K.; Zhang, Z.; Chen, G.; Li, X.; Xiao, W.; Xu, P.; et al. Aryl Hydrocarbon Receptor Activation Modulates Intestinal Epithelial Barrier Function by Maintaining Tight Junction Integrity. *Int. J. Biol. Sci.* **2018**, *14*, 69–77. [[CrossRef](#)]
167. Goya-Jorge, E.; Gonza, I.; Bondue, P.; Douny, C.; Taminiou, B.; Daube, G.; Scippo, M.-L.; Delcenserie, V. Human Adult Microbiota in a Static Colon Model: AhR Transcriptional Activity at the Crossroads of Host–Microbe Interaction. *Foods* **2022**, *11*, 1946. [[CrossRef](#)]
168. Chen, X.; Zhang, Y.; Wang, H.; Liu, L.; Li, W.; Xie, P. The regulatory effects of lactic acid on neuropsychiatric disorders. *Discov. Ment. Health* **2022**, *2*, 8. [[CrossRef](#)]
169. Guo, N.; Jiang, Y.W.; Song, X.R.; Li, Y.Y.; Liu, Z.M.; Fu, Y.J. Effect of *Bacillus natto* solid-state fermentation on the functional constituents and properties of Ginkgo seeds. *J. Food Biochem.* **2019**, *43*, e12820. [[CrossRef](#)]
170. Zhang, Y.; Anoopkumar-Dukie, S.; Davey, A.K. SIRT1 and SIRT2 Modulators: Potential Anti-Inflammatory Treatment for Depression? *Biomolecules* **2021**, *11*, 353. [[CrossRef](#)]

171. Vasconcelos, G.S.; Dos Santos Júnior, M.A.; Monte, A.S.; da Silva, F.E.R.; Lima, C.N.C.; Moreira Lima Neto, A.B.; Medeiros, I.D.S.; Teixeira, A.L.; de Lucena, D.F.; Vasconcelos, S.M.M.; et al. Low-dose candesartan prevents schizophrenia-like behavioral alterations in a neurodevelopmental two-hit model of schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* **2021**, *111*, 110348. [[CrossRef](#)] [[PubMed](#)]
172. Pan, L.; Feng, F.; Wu, J.; Fan, S.; Han, J.; Wang, S.; Yang, L.; Liu, W.; Wang, C.; Xu, K. Demethylzeylasteral targets lactate by inhibiting histone lactylation to suppress the tumorigenicity of liver cancer stem cells. *Pharmacol. Res.* **2022**, *181*, 106270. [[CrossRef](#)] [[PubMed](#)]
173. Sidrauski, C.; McGeachy, A.M.; Ingolia, N.T.; Walter, P. The small molecule ISRIB reverses the effects of eIF2 α phosphorylation on translation and stress granule assembly. *Elife* **2015**, *4*, e05033. [[CrossRef](#)] [[PubMed](#)]
174. Hritcu, L.; Ionita, R.; Postu, P.A.; Gupta, G.K.; Turkez, H.; Lima, T.C.; Carvalho, C.U.S.; de Sousa, D.P. Antidepressant Flavonoids and Their Relationship with Oxidative Stress. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 5762172. [[CrossRef](#)] [[PubMed](#)]
175. Hayashi-Takagi, A.; Araki, Y.; Nakamura, M.; Vollrath, B.; Duron, S.G.; Yan, Z.; Kasai, H.; Haganir, R.L.; Campbell, D.A.; Sawa, A. PAKs inhibitors ameliorate schizophrenia-associated dendritic spine deterioration in vitro and in vivo during late adolescence. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 6461–6466. [[CrossRef](#)]
176. Gil-Ad, I.; Amit, B.H.; Hayardeni, L.; Tarasenko, I.; Taler, M.; Gueta, R.U.; Weizman, A. Effects of the Anti-Multiple Sclerosis Immunomodulator Laquinimod on Anxiety and Depression in Rodent Behavioral Models. *J. Mol. Neurosci.* **2014**, *55*, 552–560. [[CrossRef](#)]
177. Mao, N.; Fan, Y.; Liu, W.; Yang, H.; Yang, Y.; Li, Y.; Jin, F.; Li, T.; Yang, X.; Gao, X.; et al. Oxamate Attenuates Glycolysis and ER Stress in Silicotic Mice. *Int. J. Mol. Sci.* **2022**, *23*, 3013. [[CrossRef](#)]
178. Khan, R.S.; Yu, C.; Kastin, A.J.; He, Y.; Ehrensing, R.H.; Hsueh, H.; Stone, K.; Pan, W. Brain Activation by Peptide Pro-Leu-Gly-NH₂ (MIF-1). *Int. J. Pept.* **2010**, *2010*, 537639. [[CrossRef](#)]
179. Choi, E.-Y.; Lee, H.; Dingle, R.W.C.; Kim, K.B.; Swanson, H.I. Development of Novel CH223191-Based Antagonists of the Aryl Hydrocarbon Receptor. *Mol. Pharmacol.* **2011**, *81*, 3–11. [[CrossRef](#)]
180. Deiab, S.; Mazzi, E.; Eyunni, S.; McTier, O.; Mateeva, N.; Elshami, F.; Soliman, K.F.A. 1,2,3,4,6-Penta-O-galloylglucose within *Galla Chinensis* Inhibits Human LDH-A and Attenuates Cell Proliferation in MDA-MB-231 Breast Cancer Cells. *Evid. -Based Complement. Altern. Med.* **2015**, *2015*, 276946. [[CrossRef](#)]
181. Cid-Jofré, V.; Moreno, M.; Reyes-Parada, M.; Renard, G.M. Role of Oxytocin and Vasopressin in Neuropsychiatric Disorders: Therapeutic Potential of Agonists and Antagonists. *Int. J. Mol. Sci.* **2021**, *22*, 12077. [[CrossRef](#)] [[PubMed](#)]
182. Ala, M.; Ghasemi, M.; Mohammad Jafari, R.; Dehpour, A.R. Beyond its anti-migraine properties, sumatriptan is an anti-inflammatory agent: A systematic review. *Drug Dev. Res.* **2021**, *82*, 896–906. [[CrossRef](#)] [[PubMed](#)]
183. Manerba, M.; Vettraino, M.; Fiume, L.; Di Stefano, G.; Sartini, A.; Giacomini, E.; Buonfiglio, R.; Roberti, M.; Recanatini, M. Galloflavin (CAS 568-80-9): A Novel Inhibitor of Lactate Dehydrogenase. *Chemmedchem* **2011**, *7*, 311–317. [[CrossRef](#)] [[PubMed](#)]
184. Uba, A.I.; Radicella, C.; Readmond, C.; Scorese, N.; Liao, S.; Liu, H.; Wu, C. Binding of agonist WAY-267,464 and antagonist WAY-methylated to oxytocin receptor probed by all-atom molecular dynamics simulations. *Life Sci.* **2020**, *252*, 117643. [[CrossRef](#)] [[PubMed](#)]
185. Vukicevic, M.; Chalandon, Y.; Helg, C.; Matthes, T.; Dantin, C.; Huard, B.; Chizzolini, C.; Passweg, J.; Roosnek, E. CD56bright NK cells after hematopoietic stem cell transplantation are activated mature NK cells that expand in patients with low numbers of T cells. *Eur. J. Immunol.* **2010**, *40*, 3246–3254. [[CrossRef](#)]
186. Wang, Y.; Sun, J.; Zhu, K.; Wang, D.; Zhao, X.; Zhang, H.; Wu, S.; Wang, Y.; Wang, J. Microglial aryl hydrocarbon receptor enhances phagocytic function via SYK and promotes remyelination in the cuprizone mouse model of demyelination. *J. Neuroinflamm.* **2023**, *20*, 83. [[CrossRef](#)]
187. Fehsel, K.; Schwanke, K.; Kappel, B.; Fahimi, E.; Meisenzahl-Lechner, E.; Esser, C.; Hemmrich, K.; Haarmann-Stemann, T.; Kojda, G.; Lange-Asschenfeldt, C. Activation of the aryl hydrocarbon receptor by clozapine induces preadipocyte differentiation and contributes to endothelial dysfunction. *J. Psychopharmacol.* **2022**, *36*, 191–201. [[CrossRef](#)]
188. Korac, K.; Rajasekaran, D.; Sniegowski, T.; Schniers, B.; Ibrahim, A.; Bhutia, Y.D. Carbidopa, an activator of aryl hydrocarbon receptor, suppresses IDO1 expression in pancreatic cancer and decreases tumor growth. *Biochem. J.* **2022**, *479*, 1807–1824. [[CrossRef](#)]
189. Juricek, L.; Coumoul, X. The Aryl Hydrocarbon Receptor and the Nervous System. *Int. J. Mol. Sci.* **2018**, *19*, 2504. [[CrossRef](#)]
190. Robichon, K.; Sondhaus, S.; Jordan, T.W.; Keyzers, R.A.; Connor, B.; La Flamme, A.C. Localisation of clozapine during experimental autoimmune encephalomyelitis and its impact on dopamine and its receptors. *Sci. Rep.* **2021**, *11*, 2966. [[CrossRef](#)]
191. Mosca, L.; Mantero, V.; Penco, S.; La Mantia, L.; De Benedetti, S.; Marazzi, M.R.; Spreafico, C.; Erminio, C.; Grassi, L.; Lando, G.; et al. HLA-DRB1*15 association with multiple sclerosis is confirmed in a multigenerational Italian family. *Funct. Neurol.* **2017**, *32*, 83–88. [[CrossRef](#)] [[PubMed](#)]
192. Goldstein, J.I.; Jarskog, L.F.; Hilliard, C.; Alfirevic, A.; Duncan, L.; Fourches, D.; Huang, H.; Lek, M.; Neale, B.M.; Ripke, S.; et al. Clozapine-induced agranulocytosis is associated with rare HLA-DQB1 and HLA-B alleles. *Nat. Commun.* **2014**, *5*, 4757. [[CrossRef](#)] [[PubMed](#)]
193. Sfera, A.; Hazan, S.; Klein, C.; del Campo, C.M.Z.-M.; Sasannia, S.; Anton, J.J.; Rahman, L.; Andronescu, C.V.; Sfera, D.O.; Kozlakidis, Z.; et al. Microbial Translocation Disorders: Assigning an Etiology to Idiopathic Illnesses. *Appl. Microbiol.* **2023**, *3*, 212–240. [[CrossRef](#)]

194. Johansson, V.; Jakobsson, J.; Fortgang, R.G.; Zetterberg, H.; Blennow, K.; Cannon, T.D.; Hultman, C.M.; Wetterberg, L.; Landén, M. Cerebrospinal fluid microglia and neurodegenerative markers in twins concordant and discordant for psychotic disorders. *Eur. Arch. Psychiatry Clin. Neurosci.* **2016**, *267*, 391–402. [[CrossRef](#)] [[PubMed](#)]
195. Weber, N.S.; Gressitt, K.L.; Cowan, D.N.; Niebuhr, D.W.; Yolken, R.H.; Severance, E.G. Monocyte activation detected prior to a diagnosis of schizophrenia in the US Military New Onset Psychosis Project (MNOOP). *Schizophr. Res.* **2018**, *197*, 465–469. [[CrossRef](#)] [[PubMed](#)]
196. Severance, E.G.; Gressitt, K.L.; Stallings, C.R.; Origoni, A.E.; Khushalani, S.; Leweke, F.M.; Dickerson, F.B.; Yolken, R.H. Discordant patterns of bacterial translocation markers and implications for innate immune imbalances in schizophrenia. *Schizophr. Res.* **2013**, *148*, 130–137. [[CrossRef](#)]
197. Kamaeva, D.A.; Smirnova, L.P.; Vasilieva, S.N.; Kazantseva, D.V.; Vasilieva, A.R.; Ivanova, S.A. Catalytic Antibodies in Bipolar Disorder: Serum IgGs Hydrolyze Myelin Basic Protein. *Int. J. Mol. Sci.* **2022**, *23*, 7397. [[CrossRef](#)]
198. von Zedtwitz, K.; Matteit, I.; Michel, M.; Feige, B.; Runge, K.; Denzel, D.; Schlump, A.; Nickel, K.; Schiele, M.A.; Berger, B.; et al. Anti-MOG autoantibody-associated schizophreniform psychosis. *Acta Neuropsychiatr.* **2021**, *34*, 47–54. [[CrossRef](#)]
199. Ponomarenko, N.; Durova, O.M.; Vorobiev, I.I.; Belogurov, A.A.; Telegin, G.B.; Suchkov, S.V.; Misikov, V.K.; Morse, H.C., 3rd; Gabibov, A.G. Catalytic activity of autoantibodies toward myelin basic protein correlates with the scores on the multiple sclerosis expanded disability status scale. *Immunol. Lett.* **2006**, *103*, 45–50. [[CrossRef](#)]
200. Schott, K.; Schaefer, J.-E.; Richartz, E.; Batra, A.; Eusterschulte, B.; Klein, R.; Berg, P.A.; Bartels, M.; Mann, K.; Buchkremer, G. Autoantibodies to serotonin in serum of patients with psychiatric disorders. *Psychiatry Res.* **2003**, *121*, 51–57. [[CrossRef](#)]
201. Zong, S.; Hoffmann, C.; Mané-Damas, M.; Molenaar, P.; Losen, M.; Martinez-Martinez, P. Neuronal Surface Autoantibodies in Neuropsychiatric Disorders: Are There Implications for Depression? *Front. Immunol.* **2017**, *8*, 752. [[CrossRef](#)] [[PubMed](#)]
202. Westall, F.C. Molecular Mimicry Revisited: Gut Bacteria and Multiple Sclerosis. *J. Clin. Microbiol.* **2006**, *44*, 2099–2104. [[CrossRef](#)] [[PubMed](#)]
203. Brundin, L.; Erhardt, S.; Bryleva, E.Y.; Achtyes, E.D.; Postolache, T.T. The role of inflammation in suicidal behaviour. *Acta Psychiatr. Scand.* **2015**, *132*, 192–203. [[CrossRef](#)] [[PubMed](#)]
204. Ohlsson, L.; Gustafsson, A.; Lavant, E.; Suneson, K.; Brundin, L.; Westrin, Å.; Ljunggren, L.; Lindqvist, D. Leaky gut biomarkers in depression and suicidal behavior. *Acta Psychiatr. Scand.* **2018**, *139*, 185–193, Erratum in *Acta Psychiatr. Scand.* **2020**, *142*, 423. [[CrossRef](#)] [[PubMed](#)]
205. Ahrens, A.P.; Sanchez-Padilla, D.E.; Drew, J.C.; Oli, M.W.; Roesch, L.F.W.; Triplett, E.W. Saliva microbiome, dietary, and genetic markers are associated with suicidal ideation in university students. *Sci. Rep.* **2022**, *12*, 14306. [[CrossRef](#)]
206. Eyzaguirre-Velásquez, J.; González-Toro, M.P.; González-Arancibia, C.; Escobar-Luna, J.; Beltrán, C.J.; Bravo, J.A.; Julio-Pieper, M. Sertraline and Citalopram Actions on Gut Barrier Function. *Dig. Dis. Sci.* **2020**, *66*, 3792–3802. [[CrossRef](#)]
207. Wang, C.; Zhang, T.; He, L.; Fu, J.-Y.; Deng, H.-X.; Xue, X.-L.; Chen, B.-T. Bacterial Translocation Associates With Aggression in Schizophrenia Inpatients. *Front. Syst. Neurosci.* **2021**, *15*, 704069. [[CrossRef](#)]
208. Bulgart, H.R.; Neczypor, E.W.; Wold, L.E.; Mackos, A.R. Microbial involvement in Alzheimer disease development and progression. *Mol. Neurodegener.* **2020**, *15*, 42. [[CrossRef](#)]
209. Liang, L.; Liu, P.; Deng, Y.; Li, J.; Zhao, S. L-lactate inhibits lipopolysaccharide-induced inflammation of microglia in the hippocampus. *Int. J. Neurosci.* **2022**, *23*, 1–8. [[CrossRef](#)]
210. Guglielmetti, S.; Bernardi, S.; Del Bo', C.; Cherubini, A.; Porrini, M.; Gargari, G.; Hidalgo-Liberona, N.; Gonzalez-Dominguez, R.; Peron, G.; Zamora-Ros, R.; et al. Effect of a polyphenol-rich dietary pattern on intestinal permeability and gut and blood microbioms in older subjects: Study protocol of the MaPLE randomised controlled trial. *BMC Geriatr.* **2020**, *20*, 77. [[CrossRef](#)]
211. Figueira, I.; Garcia, G.; Pimpão, R.C.; Terrasso, A.P.; Costa, I.; Almeida, A.F.; Tavares, L.; Pais, T.F.; Pinto, P.; Ventura, M.R.; et al. Polyphenols journey through blood-brain barrier towards neuronal protection. *Sci. Rep.* **2017**, *7*, 11456. [[CrossRef](#)] [[PubMed](#)]
212. Wang, D.; Tang, W.; Zhao, J.; Fan, W.; Zhang, Y.; Zhang, C. A Comprehensive Analysis of the Effect of SIRT1 Variation on the Risk of Schizophrenia and Depressive Symptoms. *Front. Genet.* **2020**, *11*, 832. [[CrossRef](#)] [[PubMed](#)]
213. Iacono, L.L.; Visco-Comandini, F.; Valzania, A.; Viscomi, M.T.; Coviello, M.; Giampà, A.; Roscini, L.; Bisicchia, E.; Siracusano, A.; Troisi, A.; et al. Adversity in childhood and depression: Linked through SIRT1. *Transl. Psychiatry* **2015**, *5*, e629. [[CrossRef](#)] [[PubMed](#)]
214. Zu, H.; Li, C.; Dai, C.; Pan, Y.; Ding, C.; Sun, H.; Zhang, X.; Yao, X.; Zang, J.; Mo, X. SIRT2 functions as a histone deacetylase and inhibits the proliferation and migration of neuroblastoma cells. *Cell Discov.* **2022**, *8*, 54. [[CrossRef](#)] [[PubMed](#)]
215. Peña, C.J.; Nestler, E.J. Progress in Epigenetics of Depression. *Prog. Mol. Biol. Transl. Sci.* **2018**, *157*, 41–66. [[CrossRef](#)]
216. Cui, B.; Luo, Y.; Tian, P.; Peng, F.; Lu, J.; Yang, Y.; Su, Q.; Liu, B.; Yu, J.; Luo, X.; et al. Stress-induced epinephrine enhances lactate dehydrogenase A and promotes breast cancer stem-like cells. *J. Clin. Investig.* **2019**, *129*, 1030–1046. [[CrossRef](#)]
217. Zhao, Z.; Han, F.; Yang, S.; Wu, J.; Zhan, W. Oxamate-mediated inhibition of lactate dehydrogenase induces protective autophagy in gastric cancer cells: Involvement of the Akt–mTOR signaling pathway. *Cancer Lett.* **2015**, *358*, 17–26. [[CrossRef](#)]
218. Ren, Y.-Y.; Zhang, X.-R.; Li, T.-N.; Zeng, Y.-J.; Wang, J.; Huang, Q.-W. Galla Chinensis, a Traditional Chinese Medicine: Comprehensive review of botany, traditional uses, chemical composition, pharmacology and toxicology. *J. Ethnopharmacol.* **2021**, *278*, 114247. [[CrossRef](#)]

219. Rahnasto-Rilla, M.; Järvenpää, J.; Huovinen, M.; Schroderus, A.-M.; Ihantola, E.-L.; Küblbeck, J.; Khadeer, M.; Moaddel, R.; Lahtela-Kakkonen, M. Effects of galloflavin and ellagic acid on sirtuin 6 and its anti-tumorigenic activities. *Biomed. Pharmacother.* **2020**, *131*, 110701. [[CrossRef](#)]
220. Mao, Q.; Gong, X.; Zhou, C.; Tu, Z.; Zhao, L.; Wang, L.; Wang, X.; Sun, L.; Xia, J.; Lian, B.; et al. Up-regulation of SIRT6 in the hippocampus induced rats with depression-like behavior via the block Akt/GSK3 β signaling pathway. *Behav. Brain Res.* **2017**, *323*, 38–46. [[CrossRef](#)]
221. Sever, I.H.; Ozkul, B.; Erisik Tanriover, D.; Ozkul, O.; Elgormus, C.S.; Gur, S.G.; Sogut, I.; Uyanikgil, Y.; Cetin, E.O.; Erbas, O. Protective effect of oxytocin through its anti-inflammatory and antioxidant role in a model of sepsis-induced acute lung injury: Demonstrated by CT and histological findings. *Exp. Lung Res.* **2021**, *47*, 426–435. [[CrossRef](#)] [[PubMed](#)]
222. Erbaş, O.; Altuntaş, İ. *Oxytocin and Neuroprotective Effects*; IntechOpen: London, UK, 2021. [[CrossRef](#)]
223. Gonzalez, A.; Hammock, E.A.D. Oxytocin and microglia in the development of social behaviour. *Philos. Trans. R. Soc. B Biol. Sci.* **2022**, *377*, 20210059. [[CrossRef](#)] [[PubMed](#)]
224. Labandeira-García, J.L.; Garrido-Gil, P.; Rodriguez-Pallares, J.; Valenzuela, R.; Borrajo, A.; Rodríguez-Perez, A.I. Brain renin-angiotensin system and dopaminergic cell vulnerability. *Front. Neuroanat.* **2014**, *8*, 67. [[CrossRef](#)] [[PubMed](#)]
225. Srinivasa, S.; Aulinas, A.; O'malley, T.; Maehler, P.; Adler, G.K.; Grinspoon, S.K.; Lawson, E.A. Oxytocin response to controlled dietary sodium and angiotensin II among healthy individuals. *Am. J. Physiol. Endocrinol. Metab.* **2018**, *315*, E671–E675. [[CrossRef](#)] [[PubMed](#)]
226. Gard, P.R.; Daw, P.; Mashhour, Z.S.; Tran, P. Interactions of angiotensin IV and oxytocin on behaviour in mice. *J. Renin-Angiotensin-Aldosterone Syst.* **2007**, *8*, 133–138. [[CrossRef](#)]
227. Kokare, D.M.; Singru, P.S.; Dandekar, M.P.; Chopde, C.T.; Subhedar, N.K. Involvement of alpha-melanocyte stimulating hormone (α -MSH) in differential ethanol exposure and withdrawal related depression in rat: Neuroanatomical-behavioral correlates. *Brain Res.* **2008**, *1216*, 53–67. [[CrossRef](#)]
228. Bialesova, L.; Novotna, A.; Macejova, D.; Brtko, J.; Dvorak, Z. Agonistic effect of selected isoflavones on arylhydrocarbon receptor in a novel AZ-AhR transgenic gene reporter human cell line. *Gen. Physiol. Biophys.* **2015**, *34*, 331–334. [[CrossRef](#)]
229. Venkataramaiah, C.; Payani, S.; Priya, B.L.; Pradeepkiran, J.A. Therapeutic potentiality of a new flavonoid against ketamine induced glutamatergic dysregulation in schizophrenia: In vivo and in silico approach. *Biomed. Pharmacother.* **2021**, *138*, 111453. [[CrossRef](#)]
230. Křížová, L.; Dadáková, K.; Kašparovská, J.; Kašparovský, T. Isoflavones. *Molecules* **2019**, *24*, 1076. [[CrossRef](#)]
231. Long, T.; He, W.; Pan, Q.; Zhang, S.; Zhang, D.; Qin, G.; Chen, L.; Zhou, J. Microglia P2X4R-BDNF signalling contributes to central sensitization in a recurrent nitroglycerin-induced chronic migraine model. *J. Headache Pain* **2020**, *21*, 4. [[CrossRef](#)]
232. Smith, M.; Calabresi, P.; Bhargava, P. Dimethyl fumarate treatment alters NK cell function in multiple sclerosis. *Eur. J. Immunol.* **2017**, *48*, 380–383. [[CrossRef](#)] [[PubMed](#)]
233. Nielsen, N.; Ødum, N.; Ursø, B.; Lanier, L.L.; Spee, P. Cytotoxicity of CD56bright NK Cells towards Autologous Activated CD4+ T Cells Is Mediated through NKG2D, LFA-1 and TRAIL and Dampened via CD94/NKG2A. *PLoS ONE* **2012**, *7*, e31959. [[CrossRef](#)] [[PubMed](#)]
234. Ruggeri, L.; Capanni, M.; Urbani, E.; Perruccio, K.; Shlomchik, W.D.; Tosti, A.; Posati, S.; Rogaia, D.; Frassoni, F.; Aversa, F.; et al. Effectiveness of Donor Natural Killer Cell Alloreactivity in Mismatched Hematopoietic Transplants. *Science* **2002**, *295*, 2097–2100. [[CrossRef](#)] [[PubMed](#)]
235. Monji, A.; Kato, T.; Kanba, S. Cytokines and schizophrenia: Microglia hypothesis of schizophrenia. *Psychiatry Clin. Neurosci.* **2009**, *63*, 257–265. [[CrossRef](#)]
236. Wake, H.; Moorhouse, A.J.; Jinno, S.; Kohsaka, S.; Nabekura, J. Resting Microglia Directly Monitor the Functional State of Synapses In Vivo and Determine the Fate of Ischemic Terminals. *J. Neurosci.* **2009**, *29*, 3974–3980. [[CrossRef](#)]
237. Jha, M.K.; Jo, M.; Kim, J.-H.; Suk, K. Microglia-Astrocyte Crosstalk: An Intimate Molecular Conversation. *Neurosci.* **2018**, *25*, 227–240. [[CrossRef](#)]
238. Correa, F.G.; Hernangómez, M.; Guaza, C. Understanding Microglia-Neuron Cross Talk: Relevance of the Microglia-Neuron Cocultures. *Methods Mol. Biol.* **2013**, *1041*, 215–229. [[CrossRef](#)]
239. Mills, J.; Ladner, L.; Soliman, E.; Leonard, J.; Morton, P.D.; Theus, M.H. Cross-Talk and Subset Control of Microglia and Associated Myeloid Cells in Neurological Disorders. *Cells* **2022**, *11*, 3364. [[CrossRef](#)]
240. Yang, S.; Qin, C.; Hu, Z.W.; Zhou, L.Q.; Yu, H.H.; Chen, M.; Bosco, D.B.; Wang, W.; Wu, L.J.; Tian, D.S. Microglia reprogram metabolic profiles for phenotype and function changes in central nervous system. *Neurobiol. Dis.* **2021**, *152*, 105290. [[CrossRef](#)]
241. Nijland, P.G.; Michailidou, I.; Witte, M.E.; Mizee, M.R.; van der Pol, S.M.; van Het Hof, B.; Reijkerkerk, A.; Pellerin, L.; van der Valk, P.; de Vries, H.E.; et al. Cellular distribution of glucose and monocarboxylate transporters in human brain white matter and multiple sclerosis lesions. *Glia* **2014**, *62*, 1125–1141. [[CrossRef](#)]
242. Goyal, M.S.; Vlassenko, A.G.; Blazey, T.M.; Su, Y.; Couture, L.E.; Durbin, T.J.; Bateman, R.J.; Benzinger, T.L.-S.; Morris, J.C.; Raichle, M.E. Loss of Brain Aerobic Glycolysis in Normal Human Aging. *Cell Metab.* **2017**, *26*, 353–360.e3. [[CrossRef](#)] [[PubMed](#)]
243. Hipkiss, A.R. Aging, Alzheimer's Disease and Dysfunctional Glycolysis; Similar Effects of Too Much and Too Little. *Aging Dis.* **2019**, *10*, 1328–1331. [[CrossRef](#)] [[PubMed](#)]
244. Zuccoli, G.S.; Guest, P.C.; Martins-De-Souza, D. Effects on Glial Cell Glycolysis in Schizophrenia: An Advanced Aging Phenotype? *Adv. Exp. Med. Biol.* **2019**, *1178*, 25–38. [[CrossRef](#)] [[PubMed](#)]

245. Dai, X.; Lv, X.; Thompson, E.W.; Ostrikov, K.K. Histone lactylation: Epigenetic mark of glycolytic switch. *Trends Genet.* **2021**, *38*, 124–127. [[CrossRef](#)]
246. Szeto, A.; Sun-Suslow, N.; Mendez, A.J.; Hernandez, R.I.; Wagner, K.V.; McCabe, P.M. Regulation of the macrophage oxytocin receptor in response to inflammation. *Am. J. Physiol. Endocrinol. Metab.* **2017**, *312*, E183–E189. [[CrossRef](#)]
247. Kaneko, Y.; Pappas, C.; Tajiri, N.; Borlongan, C.V. Oxytocin modulates GABAAR subunits to confer neuroprotection in stroke in vitro. *Sci. Rep.* **2016**, *6*, 35659. [[CrossRef](#)]
248. Buonfiglioli, A.; Hambardzumyan, D. Macrophages and microglia: The cerberus of glioblastoma. *Acta Neuropathol. Commun.* **2021**, *9*, 54. [[CrossRef](#)]
249. Genuis, S.J. Toxic causes of mental illness are overlooked. *Neurotoxicology* **2008**, *29*, 1147–1149. [[CrossRef](#)]
250. Krüger, T.; Long, M.; Bonefeld-Jørgensen, E.C. Plastic components affect the activation of the aryl hydrocarbon and the androgen receptor. *Toxicology* **2008**, *246*, 112–123. [[CrossRef](#)]
251. Scott, S.A.; Fu, J.; Chang, P.V. Microbial tryptophan metabolites regulate gut barrier function via the aryl hydrocarbon receptor. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 19376–19387. [[CrossRef](#)]
252. Zhang, J.; Velmeshev, D.; Hashimoto, K.; Huang, Y.; Hofmann, J.W.; Shi, X.; Chen, J.; Leidal, A.M.; Dishart, J.G.; Cahill, M.K.; et al. Neurotoxic microglia promote TDP-43 proteinopathy in progranulin deficiency. *Nature* **2020**, *588*, 459–465. [[CrossRef](#)] [[PubMed](#)]
253. Piccioni, G.; Mango, D.; Saidi, A.; Corbo, M.; Nisticò, R. Targeting Microglia-Synapse Interactions in Alzheimer’s Disease. *Int. J. Mol. Sci.* **2021**, *22*, 2342. [[CrossRef](#)] [[PubMed](#)]
254. Gober, R.; Ardalan, M.; Shiadeh, S.M.J.; Duque, L.; Garamszegi, S.P.; Ascona, M.; Barreda, A.; Sun, X.; Mallard, C.; Vontell, R.T. Microglia activation in postmortem brains with schizophrenia demonstrates distinct morphological changes between brain regions. *Brain Pathol.* **2021**, *32*, e13003. [[CrossRef](#)] [[PubMed](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.