

## Review Article

# The role of menaquinones (vitamin K<sub>2</sub>) in human health

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### Abstract

Recent reports have attributed the potential health benefits of vitamin K beyond its function to activate hepatic coagulation factors. Moreover, several studies have suggested that menaquinones, also known as vitamin K<sub>2</sub>, may be more effective in activating extra-hepatic vitamin K-dependent proteins than phyloquinone, also known as vitamin K<sub>1</sub>. Nevertheless, present dietary reference values (DRV) for vitamin K are exclusively based on phyloquinone, and its function in coagulation. The present review describes the current knowledge on menaquinones based on the following criteria for setting DRV: optimal dietary intake; nutrient amount required to prevent deficiency, maintain optimal body stores and/or prevent chronic disease; factors influencing requirements such as absorption, metabolism, age and sex. Dietary intake of menaquinones accounts for up to 25% of total vitamin K intake and contributes to the biological functions of vitamin K. However, menaquinones are different from phyloquinone with respect to their chemical structure and pharmacokinetics, which affects bioavailability, metabolism and perhaps impact on health outcomes. There are significant gaps in the current knowledge on menaquinones based on the criteria for setting DRV. Therefore, we conclude that further investigations are needed to establish how differences among the vitamin K forms may influence tissue specificities and their role in human health. However, there is merit for considering both menaquinones and phyloquinone when developing future recommendations for vitamin K intake.

**Key words:** Vitamin K: Dietary recommendations: Menaquinones: Bone health: CVD: Bioavailability

There is increasing interest in the potential health benefits of vitamin K beyond its role in coagulation. Several studies have reported functions for vitamin K beyond its classic role, including the improvement of bone health<sup>(1)</sup>, and the reduction of vascular calcification and cardiovascular risk<sup>(2,3)</sup>. Moreover, several studies<sup>(2–4)</sup> have suggested that menaquinones, also known as vitamin K<sub>2</sub>, could be more effective in these functions than phyloquinone, also known as vitamin K<sub>1</sub>. Nevertheless, menaquinones are generally not taken into consideration when developing dietary recommendations for vitamin K. Present recommendations for dietary vitamin K

are defined for phyloquinone intake only, and are based on median intakes of phyloquinone in certain regions, such as North America<sup>(5–7)</sup>. In some cases, the effects of phyloquinone on coagulation have also been accounted for. For healthy adults, adequate intakes of vitamin K range from 55 to 90 µg/d for adult women and 65–120 µg/d for adult men.

The International Life Sciences Institute (ILSI) Europe has selected experts on vitamin K from academia and industry to review the need for specific dietary reference values (DRV) for menaquinones. To achieve this objective, the expert group conducted a thorough review of existing literature on

**Abbreviations:** BMD, bone mineral density; DRV, dietary reference values; Gla, γ-carboxyglutamate; ILSI, International Life Sciences Institute; MGP, matrix γ-carboxyglutamate protein; MK-*n*, menaquinones; OC, osteocalcin; PIVKA-II, undercarboxylated prothrombin; PT, prothrombin time; uOC, undercarboxylated osteocalcin.

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dietary menaquinones and their role in human health to evaluate: (a) whether unique recommendations for menaquinone intake are justified at this time and (b) what additional information is needed to have strong scientific underpinnings for establishing DRV for menaquinones. In defining nutrient requirements, the selection of criteria to establish nutrient adequacy is an important step. For most nutrients, a hierarchy of criteria for nutrient adequacy can be established, ranging from the prevention of clinical deficiency to the optimisation of body stores or status. The goal is to have a low probability of nutrient inadequacy while minimising the potential risk of excess<sup>(8)</sup>. In light of this definition, we have reviewed the literature with a focus on the following criteria for setting DRV: chemical structure and function of menaquinones; dietary intake of menaquinones; absorption and metabolism of menaquinones; amount of menaquinones required to prevent deficiency, maintain optimal body stores and/or prevent chronic disease; factors influencing menaquinone requirements such as age, sex and safety. The evidence for individual menaquinones for each of these items is described and, if known, differences with phyloquinone are described. Based on this evidence, a conclusion on setting a DRV for menaquinones is drawn and recommendations for future research are made.

### Chemical structure and function of menaquinones

Vitamin K is a generic term for a number of structurally related compounds that are characterised by their common functional methylated naphthoquinone ring system, and an aliphatic side chain composed of a number of isoprenoid residues. All differences between the various forms of vitamin K originate from the differences in the length and the saturation degree of the side chain<sup>(9)</sup>. Phyloquinone is a single compound with a side chain of four isoprenoid residues, three of which are saturated (Fig. 1). Menaquinones, commonly found in nature, have side chains of varying length between four and thirteen isoprene residues, most of which are unsaturated<sup>(10)</sup>. However, some bacteria produce isoprenologues in which one or more of the prenyl units are saturated<sup>(9)</sup>. Menaquinones are generally denoted as MK-*n*, where *n* stands for the number of isoprene residues.

MK-4 is unique among the menaquinones in that it is not synthesised by bacteria. Instead, MK-4 is alkylated from

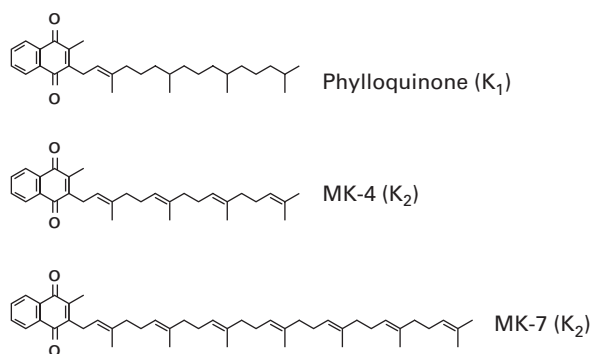


Fig. 1. Chemical structures of K vitamins. MK, menaquinone.

menadiione (vitamin K<sub>3</sub>), a synthetic form of vitamin K that is present in animal feeds, or is the product of tissue-specific conversion directly from dietary phyloquinone, with menadiione as the postulated intermediate<sup>(11,12)</sup>. There is also speculation that longer-chain menaquinones, such as MK-7, can be converted to MK-4 as well<sup>(13)</sup>. The most abundant menaquinones in the human diet are the short-chain MK-4, which originates from animal products, and the long-chain MK-7, MK-8, MK-9 and MK-10.

All forms of vitamin K have one well-known function. They all serve as a cofactor for the post-translational enzyme  $\gamma$ -glutamate carboxylase, which is established by the common naphthoquinone ring structure<sup>(14)</sup>. This enzyme converts certain protein-bound glutamate residues into  $\gamma$ -carboxyglutamate, generally known as Gla. Currently, seventeen members of the Gla protein family are known, including seven proteins involved in blood coagulation (all synthesised in the liver), osteocalcin (OC; bone), matrix Gla protein (MGP; mainly cartilage and vessel wall), growth arrest-specific protein 6, Gla-rich proteins, two proline-rich Gla proteins, two transmembrane Gla proteins, periostin and periostin-like factor<sup>(15)</sup>. With the exception of the clotting factors OC (bone formation) and MGP (inhibitor of soft tissue calcification), the physiological importance of these proteins is not yet fully understood<sup>(15)</sup>. At this time, conformation-specific assays are available for two extra-hepatic Gla proteins (OC and MGP).

### Dietary intake of menaquinones

Menaquinones generally are of microbial origin. Important dietary sources are cheese, curd and natto (a traditional Japanese food composed of fermented soya beans)<sup>(16)</sup>, while dietary phyloquinone is mainly found in green vegetables, notably spinach, broccoli, kale and Brussels sprouts<sup>(17,18)</sup>. Estimated intake of phyloquinone and menaquinones in The Netherlands and Germany has suggested that between 10 and 25% of total vitamin K intake are provided by menaquinones<sup>(16,19)</sup>. Information on the dietary intake of menaquinones is, however, limited. This is mainly due to the lack of complete food composition tables that list menaquinone concentrations in common foods. Currently, most food composition data for menaquinones are restricted to single foods such as cheese or yogurt<sup>(16,20)</sup>. However, a regularly updated and expanding food composition table of foods on the Dutch market is produced at VitaK's laboratories in The Netherlands.

Based on the content of menaquinones of certain foods, regional differences exist both for the form and the amount of menaquinones consumed. For example, in Japan, MK-7 intake has mostly been documented in the diet due to the consumption of natto, while long-chain menaquinones, MK-7 to MK-10, are predominantly consumed by high dairy intake populations, such as the Dutch population. In addition, cheese is the most important source of menaquinones in the European food supply, and therefore menaquinone concentrations in the cheeses of Dutch, German, Swiss, British and French origins were tested<sup>(16)</sup>. Since different lactic acid bacteria are used in European cheeses, a large variability in



menaquinone content among the cheeses was found<sup>(21)</sup>. However, it remains to be seen whether these data are applicable to non-European countries.

The few studies that have provided estimates for menaquinone intakes have been mainly performed among Japanese or European populations for which menaquinone-rich foods are present in the diet. A study by Kamao *et al.*<sup>(20)</sup> measured the menaquinone content of foods and the estimated intake of MK-4 and MK-7 of 125 young Japanese women using a 3 d weighed food record. The MK-7 intake of this population was originally estimated at 57.4 (sd 83.7) µg/d and accounted for approximately 25% of the total intake of vitamin K. However, this estimate is mainly driven by natto consumption, as it accounted for 99% of the MK-7 intake and almost half of the population studied consumed natto. After stratifying by natto consumption, intake of MK-7 was estimated at 154.1 (sd 87.8) µg/d among natto consumers, but data on non-natto consumers were not provided.

Several Dutch studies investigating the associations of menaquinone intake with disease incidence obtained estimates for menaquinone intake<sup>(2,3,22,23)</sup> (Table 1) using FFQ. These intake estimates were based on direct measurements of menaquinones in foods in combination with published data<sup>(16,24)</sup>. The self-reported mean intake of menaquinones was approximately 31 µg/d<sup>(3,22)</sup>. In the EPIC-Netherlands cohort, cheese contributed 53% of menaquinone intake, while milk products and meat contributed 19 and 17%, respectively. The most prominent long-chain menaquinone reported in the diet was MK-9. In the Rotterdam Study, MK-5 to MK-10 contributed 23.1 (sd 16.3) µg/d for men and 20.7 ± 13.8 µg/d for women. These data on food contents of menaquinones have also been applied to a German cohort of approximately 12 000 men<sup>(19)</sup>. Similar estimates of 34.7 µg/d (interquartile range 25.7–45.7) were reported for all menaquinones and 18.0 µg/d (11.7–27.0) for MK-5 to MK-9, with cheese being the most important food source of menaquinones<sup>(19)</sup>.

Finally, the European Food Safety Authority reported data on UK intake of menaquinones based on the UK National Dietary and Nutrition Survey<sup>(25)</sup>. This study used weighed dietary records, albeit based on the menaquinone content from a limited number of food items<sup>(25)</sup>. The overall estimated intake of menaquinones ranged from 36 µg/d (female adults) to 54 µg/d (male teenagers). The mean estimated intake of menaquinones among male adults was 43 µg/d, which is similar to the intakes reported for other European countries.

It should be noted that estimates from the Dutch and German populations were obtained from FFQ that are designed to estimate the relative dietary intake of large populations, but not to estimate the absolute dietary intake. These limitations should be kept in mind when interpreting these data. In order to obtain more precise estimates of menaquinone intakes, studies using (weighed) food records and concentrations of individual menaquinones obtained from representative foods from different food supplies are required. Nonetheless, current studies have shown estimated intakes

**Table 1.** Menaquinone intake, arterial calcification and risk of CHD

Study	Study design	Participants	Location	Dietary assessment	Menaquinone intake (µg/d)	Endpoint	Results for highest v. lowest quartile
Coronary calcification Maas <i>et al.</i> <sup>(23)</sup>	Cross-sectional study	1689 women, aged 49–70 years	The Netherlands	FFQ	26.9 (BAC +) and 29.4 (BAC –)	BAC	0.7 (0.5–1.1)
Beulens <i>et al.</i> <sup>(2)</sup>	Cross-sectional study	564 postmenopausal women, aged 49–70 years	The Netherlands	FFQ	Mean: 31.6 (sd 12.3) µg/d Mean highest quartile: 48.5	CAC	0.80 (0.65–0.98)
Geleijnse <i>et al.</i> <sup>(3)</sup>	Cross-sectional study	4473 men and women, aged 55 years and over	The Netherlands	FFQ	Men: 30.8 (sd 18.0); women: 27.0 (sd 15.1) Highest tertile: > 32.7	CAC	0.48 (0.32–0.71)
CHD Geleijnse <i>et al.</i> <sup>(3)</sup>	Cohort study	4807 adults, aged 55 years and over	The Netherlands	FFQ	Men: 30.8 (sd 18.0) µg/d; women: 27.0 (sd 15.1) µg/d	CHD	0.59 (0.40–0.86)
Gast <i>et al.</i> <sup>(23)</sup>	Cohort study	16 057 women, aged 49–70 years	The Netherlands	FFQ	Highest quartile: > 32.7 Mean: 29.1 (sd 12.8) µg/d	CHD	0.91 (0.85–1.00) per 10 µg increment

BAC, breast arterial calcification; CAC, coronary artery calcification.

of menaquinones ranging between 30 and 50 µg/d, which account for up to 25% of intake of total vitamin K.

### Absorption and metabolism of menaquinones

Phylloquinone is primarily obtained from green, leafy vegetables in which it is tightly bound to the membranes of plant chloroplasts, and thus less bioavailable compared with phylloquinone obtained from plant oils and/or dietary supplements<sup>(11)</sup>. Menaquinones, which are primarily derived from animal-based sources, are consumed in food matrices containing more fat that may improve absorption and lead to higher bioavailability than phylloquinone<sup>(26)</sup>. However, this has yet to be systematically tested for all menaquinones.

Following intestinal absorption, all vitamin K forms are incorporated into TAG-rich lipoproteins and transported primarily to the liver, but also to other target tissues. Circulating TAG-bound forms of vitamin K peak at around 4–10 h after intake and the majority of phylloquinone and MK-4 are removed from the circulation by 24 h postprandially<sup>(13,16,27)</sup>. Currently, human data on the absorption of menaquinones from food sources are limited to MK-7. These MK-7 data show similar peaks at 4 h after intake, but MK-7 does not appear to be completely removed from the circulation after 72–96 h<sup>(13,16)</sup>. The different pharmacokinetics among various vitamin K forms also result in very different plasma half-life times. Whereas phylloquinone has a relatively short half-life time<sup>(28)</sup>, MK-7 has a reported half-life time of several days<sup>(13,27)</sup>. Available data indicate higher absorption and bioavailability of MK-7 than phylloquinone, which may facilitate its uptake by various target tissues.

Another difference between the short-chain forms of vitamin K (phylloquinone and MK-4) and the long-chain forms relates to tissue distribution. For example, one study showed that MK-9 is preferentially incorporated in LDL, which facilitates its transport to non-hepatic target tissues<sup>(29)</sup>. It is not known whether other long-chain menaquinones have similar transport differences. MK-4 is unique among the menaquinones in its tissue distribution, which relates to its non-bacterial origin. As recently shown in a rodent model using stable isotopes<sup>(30)</sup>, phylloquinone consumed in the form of leafy greens is converted to MK-4 in some, but not all, tissues. The labelled MK-4 was most abundant in the brain, kidney, fat and reproductive organs. In contrast to phylloquinone as the sole dietary source of vitamin K, there was no conversion of phylloquinone to MK-4 in the liver nor were there detectable amounts of labelled MK-4 in serum. These data confirm earlier rodent studies that have reported differences in tissue distribution between phylloquinone and MK-4<sup>(31,32)</sup>.

A caveat to these conclusions is that the data for phylloquinone are much more comprehensive than those for menaquinones. Many investigators have studied phylloquinone pharmacokinetics using different study designs, including stable isotopes<sup>(33–35)</sup>. In contrast, menaquinone pharmacokinetic data are limited: (1) in the forms studied (mainly limited to MK-7); (2) from a lack of replication by independent laboratories; (3) by an absence of using stable isotope technology. More research is clearly required to quantify the differences

in absorption and bioavailability among the various forms of vitamin K in order to set nutrient requirements.

### Microbiotic production of menaquinones

Most aerobic Gram-positive bacteria and the majority of anaerobic bacteria produced by the gut use menaquinones in their electron transport pathways. The length of the side chain, as indicated by different menaquinones, is controlled by specific bacteria<sup>(10)</sup>. The reasons for this are not entirely clear, but the length and degree of saturation of the menaquinone side chain are often dependent on the growth temperature of a given species<sup>(36)</sup>. Based on qualitative bacteriological analyses, several bacteria have been identified to produce specific menaquinones. Menaquinones produced by the gut flora have been tabulated by previous studies<sup>(37–39)</sup>. For example, *Bacteroides fragilis* produces MK-10 to MK-12, while *Eubacterium lentum* produces MK-6. Likewise, bacteria used as starter cultures for the production of foods such as cheese may also produce specific menaquinones. For example, the lactic acid bacteria *Lactococcus lactis* ssp. *lactis* and *L. lactis* ssp. *cremoris* produce mainly MK-8 and MK-9<sup>(40)</sup>, while propionibacteria produce mainly MK-9<sup>(41)</sup>. The implications for the relative bioavailability of dietary menaquinones produced by bacteria in the food supply need to be considered relative to the bioavailability of menaquinones produced by bacteria in the human intestine.

It was once stated that up to 50% of the human requirement for vitamin K was fulfilled by the intestinal production of menaquinones<sup>(42,43)</sup>. The 50% estimate was based on semi-quantitative measurements of the vitamin K content of the human liver, in which one-half of the vitamin K content was phylloquinone and the other half was a mixture of long-chain menaquinones<sup>(44)</sup>. However, subsequent studies indicated that phylloquinone accounted for less than 10% of the vitamin K content in the human liver, with a greater preponderance of MK-10, MK-11 and MK-12 than previously assumed<sup>(43,45)</sup>. Based on these hepatic menaquinone concentrations, one would predict that circulating long-chain menaquinones would be in much higher concentrations than phylloquinone should these menaquinones have a major contribution to the human requirement for vitamin K. This, however, does not appear to be the case, and the route of absorption of bacterially produced menaquinones is still unclear. The absorption of all vitamin K forms takes place in the small intestine via a process requiring bile salts<sup>(46)</sup>. However, bile salts are absent in the colon where the majority of menaquinones are produced, suggesting a low absorption of these vitamin K forms<sup>(47)</sup>. This was confirmed by Ichihashi *et al.*<sup>(48)</sup>, who showed that the absorption of intestinally produced menaquinones in rats is low and that the absorption rates decrease markedly with the length of the side chain. A study in infants also indicated that intestinally produced menaquinones are not well absorbed<sup>(49)</sup>. This study compared faecal and serum concentrations of phylloquinone and menaquinones of formula-fed infants with breast-fed infants. Formula-fed infants had higher serum and faecal phylloquinone concentrations as well as a higher MK-5 to MK-9 faecal concentration

compared with breast-fed infants. Serum menaquinones were undetected in most formula-fed infants, suggestive of poor absorption<sup>(49)</sup>.

Another consideration is that most bacterially produced menaquinones are within the bacterial membranes, hence not readily bioavailable. It has been postulated that these bacterially synthesised menaquinones may be important in maintaining normal coagulation among severely ill patients with prolonged vitamin K deficiency<sup>(50)</sup>; however, current data are inconclusive regarding the relative contribution of menaquinones to fulfilling the dietary requirements for vitamin K.

### The amount of menaquinones required to prevent deficiency and maintain optimal body stores

To understand the impact of menaquinones on health, it is necessary to demonstrate the link between the intakes of menaquinones and the nutritional status of vitamin K. Several biochemical markers of vitamin K status are available and all have their strengths and weaknesses, as detailed elsewhere<sup>(51)</sup>. However, measures of plasma or tissue menaquinone concentrations are needed to isolate the effects of menaquinones from those of phylloquinone on human health. Other markers of vitamin K status, which include urinary metabolites of vitamin K<sup>(52,53)</sup>, coagulation times and uncarboxylated Gla proteins, cannot differentiate the effects of menaquinones from phylloquinone. Therefore, differences between menaquinones and phylloquinone can only be determined through the use of study designs that directly compare the response of individual biomarkers with the intakes of individual forms of vitamin K.

Under controlled conditions of dietary intake, circulating menaquinone concentrations increase in response to the high intake of menaquinones and decline over time when the dietary source of menaquinones is removed<sup>(13,26)</sup>. However, data are limited, since the HPLC and MS techniques are limited to a few qualified laboratories and the long-chain menaquinones are often below the detection limit in the circulation when measured in the general population. Only a few studies have measured plasma menaquinones in response to the intake of individual menaquinones, and these have been limited to MK-7 or MK-4 supplementation<sup>(13,27,54)</sup>. For plasma MK-7, two studies showed a clear dose–response effect on circulating MK-7 concentrations after supplementation with doses ranging between 45 and 420  $\mu\text{g}$ <sup>(13,54)</sup>. In contrast, MK-4 was not detected in the circulation following a single dose of 420  $\mu\text{g}$ <sup>(27)</sup>.

Only two studies have investigated the response of vitamin K urinary metabolites to single oral doses of menaquinones. Both studies showed a good response of urinary menadiolone<sup>(12)</sup> or side-chain catabolite<sup>(55)</sup> excretion to relatively high doses of 15 or 45 mg of MK-4 or 1 mg of MK-7. Both studies included a direct comparison of menaquinones with phylloquinone, and showed similar results for both vitamin K forms. Harrington *et al.*<sup>(53)</sup> showed that the excretion of the 5- and 7-carbon side-chain metabolites responds to the depletion and repletion of phylloquinone. A similar response

to menaquinones would be expected, but no studies of similar design have been conducted.

Prothrombin time (PT), also expressed as an international normalised ratio, is a test of coagulation that can reflect clinical deficiency of vitamin K due to frank deficiency or the antagonism of vitamin K. However, PT is non-specific because abnormal values are also indicative of diseases unrelated to vitamin K deficiency. PT changes only when prothrombin concentrations drop below 50% of normal, demonstrating its low sensitivity for detecting the deficiency of vitamin K<sup>(56)</sup>. To date, only two studies<sup>(13,57)</sup> have reported the effects of MK-7 and MK-9 on coagulation parameters. In these studies, the antidotal effect of single doses of MK-7 and MK-9 was studied in volunteers stabilised on oral vitamin K antagonists. Both studies showed that MK-7 and MK-9 decreased the international normalised ratio and the concentrations of coagulation factors, and this effect was stronger for MK-7 than phylloquinone<sup>(13,57)</sup>. However, in one study<sup>(57)</sup>, menaquinones were provided as different food sources with differing doses and bioavailability, which may have influenced the results. Although these studies are informative in the clinical context of the reversal of oral anticoagulation, they are not suitable for determining the amount of menaquinones required to prevent deficiency or maintain optimal body stores. Although the effects of coagulation factors on depletion or repletion with menaquinones have not been investigated to date, sustained intakes as low as 10  $\mu\text{g}/\text{d}$  of phylloquinone for several weeks do not prolong PT in otherwise healthy adults<sup>(7,58–60)</sup>.

Thus far, conformation-specific tests have been developed for prothrombin, OC and MGP to evaluate the extent to which the various Gla proteins are carboxylated in healthy subjects. Advantages of measuring uncarboxylated vitamin K-dependent proteins are that insufficiencies measured in circulating forms theoretically reflect what occurs at the tissue level. Undercarboxylated prothrombin, also known as PIVKA-II, detects abnormalities in prothrombin before the prolongation of PT, but does not have the sensitivity to detect the variability of usual vitamin K intakes observed in normal healthy populations. PIVKA-II has been used as a marker of vitamin K status in healthy people and has been shown to respond to both dietary depletion and subsequent repletion with phylloquinone<sup>(61,62)</sup>. However, only one study<sup>(63)</sup> investigated the effect of a single intravenous dose of 10 mg MK-4 in vitamin K-deficient cancer patients, and showed a decrease in PIVKA-II levels 1–3 d after ingestion.

The effect of menaquinones on the proportion of OC that is uncarboxylated (ucOC) has been more frequently studied. ucOC is highly responsive to supplementation with either MK-4 or MK-7 in doses ranging from 45  $\mu\text{g}/\text{d}$  to 45 mg/d (MK-4 only)<sup>(13,54,64–66)</sup>. Only a low dose of 45  $\mu\text{g}$  MK-7/d did not lead to a significant reduction in ucOC<sup>(54)</sup>. A direct comparison of phylloquinone with MK-7 supplementation indicated that MK-7 is more effective in carboxylating OC than phylloquinone<sup>(13)</sup>. Assays to measure desphospho-uncarboxylated MGP only recently became available. Since that time, several intervention studies have shown clear dose–response effects of desphospho-uncarboxylated MGP



to MK-7 supplementation with doses ranging between 10 and 360  $\mu\text{g}/\text{d}$ <sup>(67–70)</sup>. However, a direct comparison between menaquinones and phylloquinone in the response of desphospho-uncarboxylated MGP to supplementation by the individual vitamin K forms has not been made.

## The amount of menaquinones required to prevent chronic diseases

### *Menaquinones, coronary calcification and CVD*

Coronary artery calcification is an important predictor of CVD<sup>(71)</sup>. MGP is an inhibitor of vascular calcification<sup>(72)</sup>. Through carboxylation of MGP, vitamin K may help reduce coronary calcification and thereby reduce the risk of CVD. Observational studies have indeed shown that a high intake of vitamin K is associated with reduced coronary calcification and a reduced risk of CVD<sup>(2,3,23)</sup>. The results from some studies suggest that this is mainly due to menaquinones<sup>(2–4,23)</sup>. Thus far, three cross-sectional studies<sup>(2,3,73)</sup> investigated the associations of menaquinone intake and coronary calcification, as summarised in Table 1. In the Rotterdam Study, intakes of menaquinones were lower in participants with severe aortic calcifications (25.6  $\mu\text{g}/\text{d}$ ) than in participants with moderate or mild calcifications (28.6 and 28.8  $\mu\text{g}/\text{d}$ , respectively;  $P=0.001$ )<sup>(3)</sup>. A strong inverse relationship between menaquinone intake and severe calcification was found in the mid and upper tertiles of menaquinone intake compared with the lowest tertile, reaching significance in the highest tertile with a menaquinone intake of more than 32.7  $\mu\text{g}/\text{d}$ . Using breast arterial calcification as a measure of arterial calcification, the prevalence of breast arterial calcification was less common in the highest (9%) quartile of menaquinone intakes, compared with the lowest quartile (13%)<sup>(73)</sup>. This study showed a similar association to that of Geleijnse *et al.*<sup>(3)</sup> with an OR of 0.7 (95% CI 0.5, 1.1), although it did not reach significance. Similarly, a high menaquinone intake over 48  $\mu\text{g}/\text{d}$  was associated with reduced coronary calcification among 600 middle-aged women<sup>(2)</sup>. We are not aware of any randomised trials to date that investigated the effect of menaquinones on the progression of arterial calcification.

Also, two of the previously mentioned cohort studies investigated the relationship between menaquinone intake and the risk of CHD (Table 1). In the Rotterdam cohort<sup>(3)</sup>, the relative risk of incident CHD was reduced in the upper tertile of menaquinone intake compared with the lowest tertile (0.43; 95% CI 0.24, 0.77). In the Prospect-EPIC cohort<sup>(23)</sup>, the investigators also observed an inverse association between the intake of menaquinones and the risk of CHD with a hazard ratio of 0.91 (95% CI 0.85, 1.00,  $P=0.08$ ) per 10  $\mu\text{g}/\text{d}$  of menaquinone intake. In order to compare these results with previous studies using categories, a menaquinone intake of 35  $\mu\text{g}/\text{d}$  would lead to a hazard ratio of about 0.7, which compares nicely with previous studies. The association between menaquinone intake and the incidence of stroke has not been investigated to date. Of note, several of these studies also investigated the relationship between phylloquinone intake and coronary calcification or the risk of CHD, but could not detect

significant associations<sup>(2,3,23)</sup>. Whether this is due to biological differences between menaquinones and phylloquinone or perhaps lower validity of the FFQ to estimate phylloquinone intake is currently unclear<sup>(2)</sup>. Finally, these associations have only been investigated in Dutch populations and generalisability of these results should be studied in different populations.

### *Menaquinones and bone*

In bone, three vitamin K-dependent proteins have been isolated: protein S; MGP; OC. The anticoagulant protein S is synthesised by osteoblasts (bone-forming cells), but its role in bone metabolism is unclear. MGP has been found in bone, dentine, cartilage and soft tissue, including blood vessels, and is associated with the organic matrix and mobilisation of bone Ca. The results of animal studies suggest that MGP prevents the calcification of soft tissue and cartilage, while facilitating normal bone growth and development<sup>(74)</sup>. OC is a protein synthesised by osteoblasts. The synthesis of both OC and MGP is regulated by calcitriol and retinoic acid. Higher ucOC concentrations, indicating a low vitamin K status, were associated with a higher hip fracture risk and lower bone mineral density (BMD) in adults<sup>(75–85)</sup> and children<sup>(86–89)</sup>. Unfortunately, the proportion of ucOC does not differ between phylloquinone and menaquinones in terms of the form responsible as an enzyme cofactor. However, serum menaquinone levels were lower in patients with osteoporosis, osteopenia and osteoporotic fractures compared with controls<sup>(90–93)</sup>. In addition, an inverse association was found between circulating MK-7 levels and the incidence of vertebral fractures in Japanese women, although this association was stronger for phylloquinone and fracture risk<sup>(94)</sup>.

Intervention studies using pharmacological doses of MK-4 showed beneficial effects on bone parameters<sup>(1)</sup>. However, these intervention studies used very high doses of MK-4 (generally 45 mg) that cannot be obtained from the habitual diet. Since this paper is focused on dietary doses that are relevant for nutritional requirements, we will focus on intervention studies that used doses that can be nutritionally obtained. Although natto contains more than 100 times as much menaquinones as various cheeses, studies on natto or its equivalent amount of MK-7 are considered within the dietary intake range.

Intervention studies investigating the effect of menaquinone supplementation on bone markers are shown in Table 2. Whereas MK-7 at low doses did not affect bone formation<sup>(64,65)</sup>, intake of natto three times per week increased bone-specific alkaline phosphatase when compared with once-per-week natto intake<sup>(95)</sup>. Only one study showed decreased bone resorption due to a combination of vitamin K forms, vitamin D<sub>3</sub>, Ca and lifestyle recommendations<sup>(96)</sup>. This finding was independent of the form of vitamin K taken, although on a molecular basis, the daily intake of MK-7 (0.154  $\mu\text{mol}$ ) in this study was about 30% less than that of phylloquinone (0.221  $\mu\text{mol}$ ).

A few cross-sectional studies investigated the association between menaquinone intake and bone maintenance. For



**Table 2.** Intervention studies on the dietary levels of vitamin K, bone markers and bone mineral density (BMD)

Study	Study design	Participants	Location	Duration (months)	Intervention	Menaquinone dose	Bone markers	BMD
Bone markers Kanelakis <i>et al.</i> <sup>(96)</sup>	DBPC trial	173 women aged 55–65 years	Greece	12	Four groups: (1) control group; (2) dairy product with Ca and vitamin D <sub>3</sub> ; (3) dairy product with vitamin K <sub>1</sub> ; (4) dairy product with MK-7	100 µg phylloquinone/d and 100 µg MK-7	Decreased in groups (3) and (4) com- pared with the other groups*	Reduced bone loss of total body in groups (2)–(4), and of lumbar spine in groups (3) and (4) compared with group (1) NA
van Summeren <i>et al.</i> <sup>(66)</sup>	DBPC trial	Fifty-five Dutch children aged 6–10 years	The Netherlands	2	Placebo and MK-7	45 µg MK-7	Unaffected††	NA
Emaus <i>et al.</i> <sup>(64)</sup>	DBPC trial	334 postmenopausal women aged 50–60 years	The Netherlands	12	Placebo and MK-7	360 µg MK-7	Unaffected††	No difference
Katsuyama <i>et al.</i> <sup>(95)</sup>	DBPC trial	Seventy-three premenopausal women	Japan	12	Four groups: (1) no natto intake; (2) once per month; (3) once per week; (4) three times per week natto intake at lunch	MK-7 (not specified)	Significant higher in group (4) compared with group (3)†	No difference

DBPC, double-blind placebo-controlled; MK-7, menaquinone-7; NA, not applicable.

\* Urinary deoxypyridinoline.

† Bone-specific alkaline phosphatase.

‡ Degradation products of C- or N-terminal telopeptides of type I collagen.

example, two Japanese studies showed that the usual dietary intake of natto was effective in maintaining bone stiffness<sup>(97)</sup> and was positively associated with a 3-year change in BMD at the femoral neck<sup>(98)</sup>. Within Norwegian individuals, no linear association was found between dietary menaquinones and BMD of the total hip; however, fractional polynomial regression analyses for the detection of non-linear associations showed a small, positive association between dietary menaquinones and BMD among women<sup>(99)</sup>. Of note, these studies described a proposed effect of dietary menaquinones via natto consumption. Although MK-7 is an important nutrient of natto, natto also contains other ingredients that have been postulated to promote bone health. Hence MK-7 may be a surrogate marker for other bone-promoting ingredients in these studies.

As shown in Table 2, three intervention studies on menaquinones and BMD used doses of menaquinones that are attainable with the diet. Only one study<sup>(96)</sup> observed a beneficial effect of vitamin K on BMD of the lumbar spine. The two other studies did not show an effect on bone stiffness or the rate of bone loss. The main differences between these three studies are the inclusion of vitamin D as part of the treatment and the regional differences in the prevalence of vitamin D deficiency<sup>(100)</sup>. These vitamin D-related disparities in study designs may have influenced the effect of menaquinones.

Only one observational study specifically examined the association between dietary menaquinone intake and fracture risk, reporting that a low intake of phylloquinone, but not menaquinones, was associated with an increased risk of hip fractures in Norwegian individuals<sup>(101)</sup>. To date, no intervention study has evaluated the efficacy of menaquinones in doses attainable in the diet in reducing fracture risk.

Only a few studies have made a direct comparison between menaquinones and phylloquinone for bone health. These studies indicated no differences<sup>(96)</sup> or showed stronger effects for phylloquinone than menaquinones<sup>(94,101)</sup>.

### Requirements across the life cycle

Dietary intake has historically been considered the primary determinant of vitamin K status<sup>(11)</sup>. However, other non-dietary factors are emerging as determinants, such as age and ethnicity<sup>(51)</sup>. To develop recommendations for dietary intakes<sup>(8)</sup>, sufficient data are needed to evaluate requirements across the life cycle. The data for phylloquinone are sparse<sup>(7)</sup>, but there are even less data for menaquinones. Currently, data on the intake or supplementation of menaquinones are limited to the assessment in children and teenagers in the UK National Dietary and Nutrition Survey<sup>(25)</sup>.

The only clinically indicated use of vitamin K is as a prophylactic against vitamin K deficiency bleeding in otherwise healthy-appearing neonates<sup>(102)</sup>. The low content of vitamin K in breast milk, low placental transfer of vitamin K, low levels of clotting factors at birth and a sterile gut are all contributing factors to the risk of vitamin K deficiency bleeding in the first few months of life. Prevention of vitamin K deficiency bleeding by oral or intramuscular administration of vitamin K at birth is standard practice in many countries. Whereas most countries use phylloquinone, certain Asian countries,

including Japan, use MK-4 prophylactically<sup>(102)</sup>. At no other point in the life cycle is frank deficiency of vitamin K a concern among an otherwise healthy population.

### Safety of high vitamin K intake

There is no documented case of toxicity for phylloquinone or menaquinones<sup>(7,25)</sup>. The European Food Safety Authority's safety assessment of menaquinones as a source of vitamin K added for nutritional purposes concluded that low doses of menaquinones presented no safety concerns<sup>(25)</sup>. Similarly, an animal study reported no toxicity associated with synthetic MK-7 administered in a single oral dose up to 2000 mg/kg or for 90 d of oral administration of 10 mg/kg per d<sup>(103)</sup>.

It is often postulated that excessive vitamin K may result in overcoagulation, i.e. increased thrombosis risk. However, vitamin K-dependent proteins have a limited number of Glu residues capable of  $\gamma$ -carboxylation per molecule, beyond which there can be no further  $\gamma$ -carboxylation or excessive coagulation. Despite this, it is critical to demonstrate that a high intake of menaquinones does not increase thrombosis risk. It was shown in rats that thrombosis risk is not increased at doses up to 250 mg/kg of MK-4<sup>(104)</sup>. In human subjects, the endogenous thrombin potential, which is the most sensitive marker to evaluate thrombosis risk in plasma<sup>(105)</sup>, was not affected by MK-7 intakes as high as 360  $\mu$ g/d for 6 weeks<sup>(70)</sup>. The only exception to this is observed in individuals on coumarin-based oral anticoagulants, for whom dietary supplementation with vitamin K can influence the stability of the international normalised ratio<sup>(59,106)</sup>. MK-7 has the potential to interfere with oral anticoagulants at doses greater than 50  $\mu$ g/d<sup>(13)</sup>. However, there is little collective experience on the potential toxicity or adverse events associated with sustained menaquinone supplementation among individuals with normal coagulation.

In Asia, MK-4 is routinely used for osteoporosis treatment in doses of 45 mg/d without reported toxicity. Reported adverse effects associated with these high doses are limited to skin rashes that subside with cessation of the MK-4 dosing<sup>(107)</sup>. As concluded by the European Food Safety Authority<sup>(25)</sup>, phase I clinical trials have not yet been designed to test the safety of menaquinones nor has any form of vitamin K been adequately tested for mutagenicity. However, it is biologically implausible to attain such high levels of intake and sufficient bioavailability from menaquinones obtained from food sources to present a risk to health.

### Conclusions

There is growing speculation that certain dietary menaquinones, while consumed in lower quantities than phylloquinone, may have unique and important contributions to the role of vitamin K on human health. However, present DRV for vitamin K are exclusively based on phylloquinone. In recognition of this emerging paradigm shift in vitamin K nutrition research, we have reviewed existing literature to evaluate the current state of knowledge on menaquinones that would be needed for inclusion in the DRV for vitamin K. It was concluded that differences in the chemical structure of menaquinones compared with phylloquinone may lead to differences in absorption and bioavailability (Table 3). Several studies have shown that certain forms of menaquinones may be more bioavailable and effective in carboxylating particular extra-hepatic Gla proteins than phylloquinone. The intake of menaquinones accounts for up to 25% of the total intake of vitamin K, and should there be a higher bioavailability, menaquinones would be important to consider in their contribution to human health. Indeed, certain observational studies have indicated that high intakes of menaquinones may be associated with greater reductions of vascular calcification and the risk of CVD than comparable amounts of phylloquinone.

**Table 3.** Summary and recommendations for future research

Criterion	Difference with phylloquinone	Recommendation for future research
Structure	Length and degree of saturation of the side chain	Do long-chain menaquinones convert to MK-4? Determine the location and biochemical pathways required for the conversion of phylloquinone to MK-4
Function	MK-4 has functions that may be unrelated to the role as an enzyme cofactor	
Dietary intake	10–25% of total intake of vitamin K	More accurate data on the food content of menaquinones More accurate data on the intake of menaquinones over more countries
Absorption	Absorption of certain menaquinones higher than phylloquinone	Stable isotope studies to quantify relative differences in absorption among different menaquinones and with phylloquinone
Metabolism	Distribution to extra-hepatic tissue may differ with phylloquinone	Stable isotope studies to quantify relative differences with phylloquinone
Bioavailability	Longer half-life time of certain menaquinones	Stable isotope studies to quantify differences with phylloquinone
Effect on the status of vitamin K	None reported	More elaborate validation of biomarkers for the status of vitamin K
Effect on health outcomes	Stronger associations with coronary calcification and the risk of CHD	Studies with clinical endpoints to isolate the putative effects of individual menaquinones
Requirement across the life cycle	Both forms used prophylactically	
Safety	None reported	

MK-4, menaquinone-4.





Such effects have not been observed for bone health. However, these data are limited to observational studies conducted among Dutch or Japanese populations. Moreover, food composition tables for menquinones are limited and available only for a few countries. In addition, studies investigating the bioavailability of menquinones using stable isotope techniques are lacking. Therefore, research is warranted to compile more elaborate food composition data of menquinones and more accurate data on the intake of menquinones, at different stages of the life cycle (Table 3). These data should be used to investigate the relationship with disease incidence in populations other than the Dutch or Japanese. Stable isotope studies are required to quantify differences in absorption, bioavailability and distribution over the body between individual menquinone forms and phyloquinone. Finally, intervention studies with clinical endpoints and a more elaborate validation of biomarkers for vitamin K status are required to quantify how the bioavailability and tissue distribution of menquinones affect vitamin K status and health outcomes.

Clearly, significant gaps in the current knowledge on menquinones exist. However, there is merit for considering both menquinones and phyloquinone when developing future recommendations for vitamin K intake.

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### References

- Cockayne S, Adamson J, Lanham-New S, *et al.* (2006) Vitamin K and the prevention of fractures: systematic review and meta-analysis of randomized controlled trials. *Arch Intern Med* **166**, 1256–1261.
- Beulens JW, Bots ML, Atsma F, *et al.* (2009) High dietary menquinone intake is associated with reduced coronary calcification. *Atherosclerosis* **203**, 489–493.
- Geleijnse JM, Vermeer C, Grobbee DE, *et al.* (2004) Dietary intake of menquinone is associated with a reduced risk of coronary heart disease: the Rotterdam Study. *J Nutr* **134**, 3100–3105.
- Spronk HM, Soute BA, Schurgers LJ, *et al.* (2003) Tissue-specific utilization of menquinone-4 results in the prevention of arterial calcification in warfarin-treated rats. *J Vasc Res* **40**, 531–537.
- Australian National Health and Medical Research Council, New Zealand Ministry of Health (2005) *Vitamin K. Nutrient Reference Values for Australia and New Zealand*, pp. 147–151. Canberra: Commonwealth of Australia.
- FAO Rome Food and Nutrition Division (2001) *Vitamin K. Human Vitamin and Mineral Requirements: Report of a Joint FAO/WHO Expert Consultation, Bangkok, Thailand*, pp. 133–150. Rome: FAO.
- National Research Council (2000) *Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc*. Washington, DC: National Academy Press.
- EFSA Panel on Dietetic Products, Nutrition and Allergies (NDA) (2010) Scientific Opinion on principles for deriving and applying Dietary Reference Values. *EFSA J* **8**, 1458.
- Shearer MJ & Newman P (2008) Metabolism and cell biology of vitamin K. *Thromb Haemost* **100**, 530–547.
- Collins MD & Jones D (1981) Distribution of isoprenoid quinone structural types in bacteria and their taxonomic implication. *Microbiol Rev* **45**, 316–354.
- Booth SL & Suttie JW (1998) Dietary intake and adequacy of vitamin K. *J Nutr* **128**, 785–788.
- Thijssen HH, Vervoort LM, Schurgers LJ, *et al.* (2006) Menadione is a metabolite of oral vitamin K. *Br J Nutr* **95**, 260–266.
- Schurgers LJ, Teunissen KJ, Hamulyak K, *et al.* (2007) Vitamin K-containing dietary supplements: comparison of synthetic vitamin K<sub>1</sub> and natto-derived menquinone-7. *Blood* **109**, 3279–3283.
- Rishavy MA & Berkner KL (2012) Vitamin K oxygenation, glutamate carboxylation, and processivity: defining the three critical facets of catalysis by the vitamin K-dependent carboxylase. *Adv Nutr* **3**, 135–148.
- McCann JC & Ames BN (2009) Vitamin K, an example of triage theory: is micronutrient inadequacy linked to diseases of aging? *Am J Clin Nutr* **90**, 889–907.
- Schurgers LJ & Vermeer C (2000) Determination of phyloquinone and menquinones in food. Effect of food matrix on circulating vitamin K concentrations. *Haemostasis* **30**, 298–307.
- Bolton-Smith C, Price RJ, Fenton ST, *et al.* (2000) Compilation of a provisional UK database for the phyloquinone (vitamin K<sub>1</sub>) content of foods. *Br J Nutr* **83**, 389–399.
- Shearer MJ & Bolton-Smith C (2000) The UK food data-base for vitamin K and why we need it. *Food Chem* **68**, 213–218.
- Nimptsch K, Rohrmann S & Linseisen J (2008) Dietary intake of vitamin K and risk of prostate cancer in the Heidelberg cohort of the European Prospective Investi-

- gation into Cancer and Nutrition (EPIC-Heidelberg). *Am J Clin Nutr* **87**, 985–992.
20. Kamao M, Suhara Y, Tsugawa N, *et al.* (2007) Vitamin K content of foods and dietary vitamin K intake in Japanese young women. *J Nutr Sci Vitaminol (Tokyo)* **53**, 464–470.
  21. Fox PF & McSweeney PLH (2004) Cheese: an overview. In *Cheese Chemistry, Physics and Microbiology*, 1st vol., 3rd ed., pp. 1–18 [PF Fox, PLH McSweeney, TM Cogan and TP Guinee, editors]. London: Elsevier Academic Press.
  22. Beulens JW, Van der A D, Grobbee DE, *et al.* (2010) Dietary phyloquinone and menaquinones intakes and risk of type 2 diabetes. *Diabetes Care* **33**, 1699–1705.
  23. Gast GC, de Roos NM, Sluijs I, *et al.* (2009) A high menaquinone intake reduces the incidence of coronary heart disease. *Nutr Metab Cardiovasc Dis* **19**, 504–510.
  24. Elder SJ, Haytowitz DB, Howe J, *et al.* (2006) Vitamin k contents of meat, dairy, and fast food in the U.S. *Diet. J Agric Food Chem* **54**, 463–467.
  25. Bresson JL, Flynn A, Heinonen M, *et al.* (2008) Vitamin K<sub>2</sub> added for nutritional purpose in foods for particular nutritional uses, food supplements and foods intended for the general population. *EFSA J* **822**, 1–31.
  26. Gijsbers BL, Jie KS & Vermeer C (1996) Effect of food composition on vitamin K absorption in human volunteers. *Br J Nutr* **76**, 223–229.
  27. Sato T, Schurgers LJ & Uenishi K (2012) Comparison of menaquinone-4 and menaquinone-7 bioavailability in healthy women. *Nutr J* **11**, 93.
  28. Novotny JA, Kurilich AC, Britz SJ, *et al.* (2010) Vitamin K absorption and kinetics in human subjects after consumption of <sup>13</sup>C-labelled phyloquinone from kale. *Br J Nutr* **104**, 858–862.
  29. Schurgers LJ & Vermeer C (2002) Differential lipoprotein transport pathways of K-vitamins in healthy subjects. *Biochim Biophys Acta* **1570**, 27–32.
  30. AL Rajaba A, Booth SL, Peterson JW, *et al.* (2012) Deuterium-labeled phyloquinone has tissue-specific conversion to menaquinone-4 among Fischer 344 male rats. *J Nutr* **142**, 841–845.
  31. Ronden JE, Thijssen HH & Vermeer C (1998) Tissue distribution of K-vitamins under different nutritional regimens in the rat. *Biochim Biophys Acta* **1379**, 16–22.
  32. Thijssen HH & Driessens FJ (1994) Vitamin K distribution in rat tissues: dietary phyloquinone is a source of tissue menaquinone-4. *Br J Nutr* **72**, 415–425.
  33. Fu X, Peterson JW, Hdeib M, *et al.* (2009) Measurement of deuterium-labeled phyloquinone in plasma by high-performance liquid chromatography/mass spectrometry. *Anal Chem* **81**, 5421–5425.
  34. Jones KS, Bluck LJ, Wang LY, *et al.* (2008) A stable isotope method for the simultaneous measurement of vitamin K<sub>1</sub> (phyloquinone) kinetics and absorption. *Eur J Clin Nutr* **62**, 1273–1281.
  35. Jones KS, Bluck LJ, Wang LY, *et al.* (2009) The effect of different meals on the absorption of stable isotope-labelled phyloquinone. *Br J Nutr* **102**, 1195–1202.
  36. Nowicka B & Kruk J (2010) Occurrence, biosynthesis and function of isoprenoid quinones. *Biochim Biophys Acta* **1797**, 1587–1605.
  37. Fernandez F & Collins MD (1987) Vitamin-K composition of anaerobic gut bacteria. *FEMS Microbiol Lett* **63**, 175–180.
  38. Hill MJ (1997) Intestinal flora and endogenous vitamin synthesis. *Eur J Cancer Prev* **6**, Suppl. 1, S43–S45.
  39. Mathers JC, Fernandez F, Hill MJ, *et al.* (1990) Dietary modification of potential vitamin K supply from enteric bacterial menaquinones in rats. *Br J Nutr* **63**, 639–652.
  40. Morishita T, Tamura N, Makino T, *et al.* (1999) Production of menaquinones by lactic acid bacteria. *J Dairy Sci* **82**, 1897–1903.
  41. Hojo K, Watanabe R, Mori T, *et al.* (2007) Quantitative measurement of tetrahydromenaquinone-9 in cheese fermented by propionibacteria. *J Dairy Sci* **90**, 4078–4083.
  42. Conly JM & Stein K (1992) The production of menaquinones (vitamin K<sub>2</sub>) by intestinal bacteria and their role in maintaining coagulation homeostasis. *Prog Food Nutr Sci* **16**, 307–343.
  43. Suttie JW (1995) The importance of menaquinones in human nutrition. *Annu Rev Nutr* **15**, 399–417.
  44. Duello TJ & Matschner JT (1972) Characterization of vitamin K from human liver. *J Nutr* **102**, 331–335.
  45. Usui Y, Tanimura H, Nishimura N, *et al.* (1990) Vitamin K concentrations in the plasma and liver of surgical patients. *Am J Clin Nutr* **51**, 846–852.
  46. Olson RE (1984) The function and metabolism of vitamin K. *Annu Rev Nutr* **4**, 281–337.
  47. Conly JM & Stein K (1992) Quantitative and qualitative measurements of K vitamins in human intestinal contents. *Am J Gastroenterol* **87**, 311–316.
  48. Ichihashi T, Takagishi Y, Uchida K, *et al.* (1992) Colonic absorption of menaquinone-4 and menaquinone-9 in rats. *J Nutr* **122**, 506–512.
  49. Fujita K, Kakuya F & Ito S (1993) Vitamin K<sub>1</sub> and K<sub>2</sub> status and faecal flora in breast fed and formula fed 1-month-old infants. *Eur J Pediatr* **152**, 852–855.
  50. Ramotar K, Conly JM, Chubb H, *et al.* (1984) Production of menaquinones by intestinal anaerobes. *J Infect Dis* **150**, 213–218.
  51. Booth SL & AL Rajaba A (2008) Determinants of vitamin K status in humans. *Vitam Horm* **78**, 1–22.
  52. AL Rajaba A, Peterson J, Choi SW, *et al.* (2010) Measurement of menadiene in urine by HPLC. *J Chromatogr B Analyt Technol Biomed Life Sci* **878**, 2457–2460.
  53. Harrington DJ, Booth SL, Card DJ, *et al.* (2007) Excretion of the urinary 5C- and 7C-aglycone metabolites of vitamin K by young adults responds to changes in dietary phyloquinone and dihydrophyloquinone intakes. *J Nutr* **137**, 1763–1768.
  54. Bruge F, Bacchetti T, Principi F, *et al.* (2011) Olive oil supplemented with menaquinone-7 significantly affects osteocalcin carboxylation. *Br J Nutr* **106**, 1058–1062.
  55. Harrington DJ, Soper R, Edwards C, *et al.* (2005) Determination of the urinary aglycone metabolites of vitamin K by HPLC with redox-mode electrochemical detection. *J Lipid Res* **46**, 1053–1060.
  56. Suttie JW (1992) Vitamin K and human nutrition. *J Am Diet Assoc* **92**, 585–590.
  57. Schurgers LJ, Shearer MJ, Hamulyak K, *et al.* (2004) Effect of vitamin K intake on the stability of oral anticoagulant treatment: dose–response relationships in healthy subjects. *Blood* **104**, 2682–2689.
  58. Frick PG, Riedler G & Brogli H (1967) Dose response and minimal daily requirement for vitamin K in man. *J Appl Physiol* **23**, 387–389.
  59. Suttie JW, Mummah-Schendel LL, Shah DV, *et al.* (1988) Vitamin K deficiency from dietary vitamin K restriction in humans. *Am J Clin Nutr* **47**, 475–480.
  60. Udall JA (1965) Human sources and absorption of vitamin K in relation to anticoagulation stability. *JAMA* **194**, 127–129.
  61. Booth SL, Lichtenstein AH, O'Brien-Morse M, *et al.* (2001) Effects of a hydrogenated form of vitamin K on bone formation and resorption. *Am J Clin Nutr* **74**, 783–790.

62. Booth SL, Martini L, Peterson JW, *et al.* (2003) Dietary phyloquinone depletion and repletion in older women. *J Nutr* **133**, 2565–2569.
63. Furukawa M, Nakanishi T, Okuda H, *et al.* (1992) Changes of plasma des-gamma-carboxy prothrombin levels in patients with hepatocellular carcinoma in response to vitamin K. *Cancer* **69**, 31–38.
64. Emaus N, Gjesdal CG, Almas B, *et al.* (2010) Vitamin K<sub>2</sub> supplementation does not influence bone loss in early menopausal women: a randomised double-blind placebo-controlled trial. *Osteoporos Int* **21**, 1731–1740.
65. Tsukamoto Y, Ichise H, Kakuda H, *et al.* (2000) Intake of fermented soybean (natto) increases circulating vitamin K<sub>2</sub> (menaquinone-7) and gamma-carboxylated osteocalcin concentration in normal individuals. *J Bone Miner Metab* **18**, 216–222.
66. van Summeren MJ, Braam LA, Lilien MR, *et al.* (2009) The effect of menaquinone-7 (vitamin K<sub>2</sub>) supplementation on osteocalcin carboxylation in healthy prepubertal children. *Br J Nutr* **102**, 1171–1178.
67. Cranenburg EC, Koos R, Schurgers LJ, *et al.* (2010) Characterisation and potential diagnostic value of circulating matrix Gla protein (MGP) species. *Thromb Haemost* **104**, 811–822.
68. Dalmeijer GW, van der Schouw YT, Magdeleyns E, *et al.* (2012) The effect of menaquinone-7 supplementation on circulating species of matrix Gla protein. *Atherosclerosis* **225**, 397–402.
69. Theuwissen E, Cranenburg EC, Knapen MH, *et al.* (2012) Low-dose menaquinone-7 supplementation improved extra-hepatic vitamin K status, but had no effect on thrombin generation in healthy subjects. *Br J Nutr* **108**, 1652–1657.
70. Westenfeld R, Krueger T, Schlieper G, *et al.* (2012) Effect of vitamin K<sub>2</sub> supplementation on functional vitamin K deficiency in hemodialysis patients: a randomized trial. *Am J Kidney Dis* **59**, 186–195.
71. Greenland P, Bonow RO, Brundage BH, *et al.* (2007) ACCF/AHA clinical expert consensus document on coronary artery calcium scoring by computed tomography in global cardiovascular risk assessment and in evaluation of patients with chest pain: a report of the American College of Cardiology Foundation Clinical Expert Consensus Task Force (ACCF/AHA Writing Committee to Update the 2000 Expert Consensus Document on Electron Beam Computed Tomography) developed in collaboration with the Society of Atherosclerosis Imaging and Prevention and the Society of Cardiovascular Computed Tomography. *J Am Coll Cardiol* **49**, 378–402.
72. Shanahan CM, Proudfoot D, Farzaneh-Far A, *et al.* (1998) The role of Gla proteins in vascular calcification. *Crit Rev Eukaryot Gene Expr* **8**, 357–375.
73. Maas AH, van der Schouw YT, Beijerinck D, *et al.* (2007) Vitamin K intake and calcifications in breast arteries. *Maturitas* **56**, 273–279.
74. Booth SL (1997) Skeletal functions of vitamin K-dependent proteins: not just for clotting anymore. *Nutr Rev* **55**, 282–284.
75. Booth SL, Tucker KL, Chen H, *et al.* (2000) Dietary vitamin K intakes are associated with hip fracture but not with bone mineral density in elderly men and women. *Am J Clin Nutr* **71**, 1201–1208.
76. Jie KG, Bots ML, Vermeer C, *et al.* (1996) Vitamin K status and bone mass in women with and without aortic atherosclerosis: a population-based study. *Calcif Tissue Int* **59**, 352–356.
77. Kaneki M, Hodges SJ, Hosoi T, *et al.* (2001) Japanese fermented soybean food as the major determinant of the large geographic difference in circulating levels of vitamin K<sub>2</sub>: possible implications for hip-fracture risk. *Nutrition* **17**, 315–321.
78. Knapen MH, Nieuwenhuijzen Kruseman AC, Wouters RS, *et al.* (1998) Correlation of serum osteocalcin fractions with bone mineral density in women during the first 10 years after menopause. *Calcif Tissue Int* **63**, 375–379.
79. Luukinen H, Kakonen SM, Pettersson K, *et al.* (2000) Strong prediction of fractures among older adults by the ratio of carboxylated to total serum osteocalcin. *J Bone Miner Res* **15**, 2473–2478.
80. Schaafsma A, Muskiet FA, Storm H, *et al.* (2000) Vitamin D(3) and vitamin K(1) supplementation of Dutch postmenopausal women with normal and low bone mineral densities: effects on serum 25-hydroxyvitamin D and carboxylated osteocalcin. *Eur J Clin Nutr* **54**, 626–631.
81. Sugiyama T & Kawai S (2001) Carboxylation of osteocalcin may be related to bone quality: a possible mechanism of bone fracture prevention by vitamin K. *J Bone Miner Metab* **19**, 146–149.
82. Szulc P, Chapuy MC, Meunier PJ, *et al.* (1993) Serum undercarboxylated osteocalcin is a marker of the risk of hip fracture in elderly women. *J Clin Invest* **91**, 1769–1774.
83. Szulc P, Arlot M, Chapuy MC, *et al.* (1994) Serum undercarboxylated osteocalcin correlates with hip bone mineral density in elderly women. *J Bone Miner Res* **9**, 1591–1595.
84. Szulc P, Chapuy MC, Meunier PJ, *et al.* (1996) Serum undercarboxylated osteocalcin is a marker of the risk of hip fracture: a three year follow-up study. *Bone* **18**, 487–488.
85. Vergnaud P, Garnero P, Meunier PJ, *et al.* (1997) Undercarboxylated osteocalcin measured with a specific immunoassay predicts hip fracture in elderly women: the EPIDOS Study. *J Clin Endocrinol Metab* **82**, 719–724.
86. Kalkwarf HJ, Khoury JC, Bean J, *et al.* (2004) Vitamin K, bone turnover, and bone mass in girls. *Am J Clin Nutr* **80**, 1075–1080.
87. O'Connor E, Molgaard C, Michaelsen KF, *et al.* (2007) Serum percentage undercarboxylated osteocalcin, a sensitive measure of vitamin K status, and its relationship to bone health indices in Danish girls. *Br J Nutr* **97**, 661–666.
88. van Summeren M, van Coeverden SC, Schurgers LJ, *et al.* (2008) Vitamin K status is associated with childhood bone mineral content. *Br J Nutr* **100**, 852–858.
89. van SM, Braam L, Noirt F, *et al.* (2007) Pronounced elevation of undercarboxylated osteocalcin in healthy children. *Pediatr Res* **61**, 366–370.
90. Hodges SJ, Pilkington MJ, Stamp TC, *et al.* (1991) Depressed levels of circulating menaquinones in patients with osteoporotic fractures of the spine and femoral neck. *Bone* **12**, 387–389.
91. Hodges SJ, Akesson K, Vergnaud P, *et al.* (1993) Circulating levels of vitamins K<sub>1</sub> and K<sub>2</sub> decreased in elderly women with hip fracture. *J Bone Miner Res* **8**, 1241–1245.
92. Kanai T, Takagi T, Masuhiro K, *et al.* (1997) Serum vitamin K level and bone mineral density in post-menopausal women. *Int J Gynaecol Obstet* **56**, 25–30.
93. Tamatani M, Morimoto S, Nakajima M, *et al.* (1998) Decreased circulating levels of vitamin K and 25-hydroxyvitamin D in osteopenic elderly men. *Metabolism* **47**, 195–199.
94. Tsugawa N, Shiraki M, Suhara Y, *et al.* (2008) Low plasma phyloquinone concentration is associated with high incidence of vertebral fracture in Japanese women. *J Bone Miner Metab* **26**, 79–85.

95. Katsuyama H, Ideguchi S, Fukunaga M, *et al.* (2004) Promotion of bone formation by fermented soybean (Natto) intake in premenopausal women. *J Nutr Sci Vitaminol (Tokyo)* **50**, 114–120.
96. Kanellakis S, Moschonis G, Tenta R, *et al.* (2012) Changes in parameters of bone metabolism in postmenopausal women following a 12-month intervention period using dairy products enriched with calcium, vitamin D, and phyloquinone (vitamin K(1)) or menaquinone-7 (vitamin K (2)): the Postmenopausal Health Study II. *Calcif Tissue Int* **90**, 251–262.
97. Katsuyama H, Ideguchi S, Fukunaga M, *et al.* (2002) Usual dietary intake of fermented soybeans (Natto) is associated with bone mineral density in premenopausal women. *J Nutr Sci Vitaminol (Tokyo)* **48**, 207–215.
98. Ikeda Y, Iki M, Morita A, *et al.* (2006) Intake of fermented soybeans, natto, is associated with reduced bone loss in postmenopausal women: Japanese Population-Based Osteoporosis (JPOS) Study. *J Nutr* **136**, 1323–1328.
99. Apalset EM, Gjesdal CG, Eide GE, *et al.* (2010) Dietary vitamins K<sub>1</sub>, K<sub>2</sub> and bone mineral density: The Hordaland Health Study. *Arch Osteopor* **5**, 73–81.
100. Lips P (2010) Worldwide status of vitamin D nutrition. *J Steroid Biochem Mol Biol* **121**, 297–300.
101. Apalset EM, Gjesdal CG, Eide GE, *et al.* (2011) Intake of vitamin K<sub>1</sub> and K<sub>2</sub> and risk of hip fractures: The Hordaland Health Study. *Bone* **49**, 990–995.
102. Shearer MJ (2009) Vitamin K deficiency bleeding (VKDB) in early infancy. *Blood Rev* **23**, 49–59.
103. Pucaj K, Rasmussen H, Moller M, *et al.* (2011) Safety and toxicological evaluation of a synthetic vitamin K<sub>2</sub>, menaquinone-7. *Toxicol Mech Methods* **21**, 520–532.
104. Ronden JE, Groenen-van Dooren MM, Hornstra G, *et al.* (1997) Modulation of arterial thrombosis tendency in rats by vitamin K and its side chains. *Atherosclerosis* **132**, 61–67.
105. Hemker HC, AL Dieri R, De Smeat E, *et al.* (2006) Thrombin generation, a function test of the haemostatic–thrombotic system. *Thromb Haemost* **96**, 553–561.
106. Holmes MV, Hunt BJ & Shearer MJ (2012) The role of dietary vitamin K in the management of oral vitamin K antagonists. *Blood Rev* **26**, 1–14.
107. Bunyaratavej N, Penkitti P, Kittimanon N, *et al.* (2001) Efficacy and safety of menatetrenone-4 postmenopausal Thai women. *J Med Assoc Thai* **84**, Suppl. 2, S553–S559.