



The Role of Microbes in the Nutrition of Detritivorous Invertebrates: A Stoichiometric Analysis

Thomas R. Anderson^{1*}, David W. Pond² and Daniel J. Mayor^{1*}

¹ National Oceanography Centre, Southampton, UK, ² Scottish Association for Marine Science, Natural Environment Research Council, Oban, UK

OPEN ACCESS

Edited by:

James Joseph Elser,
University of Montana, USA

Reviewed by:

Andrew Decker Steen,
University of Tennessee, USA

Tom Andersen,
University of Oslo, Norway
Angela Peace,
Texas Tech University, USA

*Correspondence:

Thomas R. Anderson
tra@noc.ac.uk
Daniel J. Mayor
dan.mayor@noc.ac.uk

Specialty section:

This article was submitted to
Aquatic Microbiology,
a section of the journal
Frontiers in Microbiology

Received: 07 November 2016

Accepted: 14 December 2016

Published: 04 January 2017

Citation:

Anderson TR, Pond DW and Mayor DJ (2017) The Role of Microbes in the Nutrition of Detritivorous Invertebrates: A Stoichiometric Analysis. *Front. Microbiol.* 7:2113. doi: 10.3389/fmicb.2016.02113

Detritus represents an important pool in the global carbon cycle, providing a food source for detritivorous invertebrates that are conspicuous components of almost all ecosystems. Our knowledge of how these organisms meet their nutritional demands on a diet that is typically comprised of refractory, carbon-rich compounds nevertheless remains incomplete. “Trophic upgrading” of detritus by the attached microbial community (enhancement of zooplankton diet by the inclusion of heterotrophic protozoans) represents a potential source of nutrition for detritivores as both bacteria and their flagellated protistan predators are capable of biosynthesizing essential micronutrients such as polyunsaturated fatty acids (PUFAs). There is however a trade-off because although microbes enhance the substrate in terms of its micronutrient content, the quantity of organic carbon is diminished through metabolic losses as energy passes through the microbial food web. Here, we develop a simple stoichiometric model to examine this trade-off in the nutrition of detritivorous copepods inhabiting the mesopelagic zone of the ocean, focusing on their requirements for carbon and an essential PUFA, docosahexaenoic acid (DHA). Results indicate that feeding on microbes may be a highly favorable strategy for these invertebrates, although the potential for carbon to become limiting when consuming a microbial diet exists because of the inefficiencies of trophic transfer within the microbial food web. Our study highlights the need for improved knowledge at the detritus-microbe-metazoan interface, including interactions between the physiology and ecology of the associated organisms.

Keywords: detritus, microbial loop, stoichiometry, trophic upgrading, polyunsaturated fatty acids, mesopelagic zone

INTRODUCTION

The production of dead and decaying particulate organic matter (“detritus” hereafter) may account for as much as 56% of primary production when averaged across a range of ecosystems (Cebrián and Duarte, 1995). This flux of detritus thereby constitutes a significant term in the global carbon cycle (Ciais et al., 2013) and is a major conduit through which organic matter is transported both within and between ecosystems (Bartels et al., 2012). It also provides sustenance to countless detritivorous invertebrates, which we loosely interpret as any animal that has a trophic association with dead organic matter, including organismal egesta. Detritus-detritivore interactions influence the potential for carbon sequestration in both terrestrial and aquatic environments. Understanding

the interface between living and dead organic matter is therefore a prerequisite to improving predictions of global biogeochemical cycles and climate (Burd et al., 2016; Luo et al., 2016).

Detritus is mainly composed of refractory compounds such as structural polysaccharides (Mann, 1988; Kiem and Kögel-Knabner, 2003), but is depleted in micronutrients such as amino acids and fatty acids (Cowie and Hedges, 1996; Pokarzhevskii et al., 1997; Mayor et al., 2011) that are considered essential for the growth of metazoan animals (Müller-Navarra et al., 2000; Anderson et al., 2004; Sampedro et al., 2006; Larsen et al., 2016). The nutritional challenge facing detritivores may, however, be mitigated by the presence of microorganisms that colonize the detrital substrate (Moran and Hodson, 1989; Turley and Mackie, 1994). Detritivores actively ingest this detritus-associated microbial community which, unlike the basal substrate, is readily absorbed and provides a rich source of micronutrients (Bärlocher and Kendrick, 1975; Phillips, 1984; Lawrence et al., 1993; Koski et al., 2005). Indeed, a key functional characteristic of many detritivorous invertebrates is their propensity to shred or fragment detritus (Anderson and Sedell, 1979; Iversen and Poulsen, 2007), an activity that has been proposed to stimulate the production of microbial biomass by increasing the surface area of the substrate, so-called “microbial gardening” (Fenchel, 1970; Mayor et al., 2014). The resulting uplift in the nutritional content of detritus represents a form of “trophic upgrading,” a term which originates from the marine literature and refers to the enhancement of zooplankton growth by the inclusion of micronutrient-rich heterotrophic protozoans in an otherwise herbivorous diet (Klein Breteler et al., 1999). Relying on microbes as a primary source of nutrition does, however come at an energetic cost because their gross growth efficiencies are typically <30 % (Del Giorgio and Cole, 1998) and the majority of organic carbon in the detrital substrate is therefore lost during the trophic upgrading process. Detritivorous invertebrates thus face a trade-off between consuming a high quality, low quantity diet that is rich in microbes versus the low quality, high quantity detritus (Mayor et al., 2014).

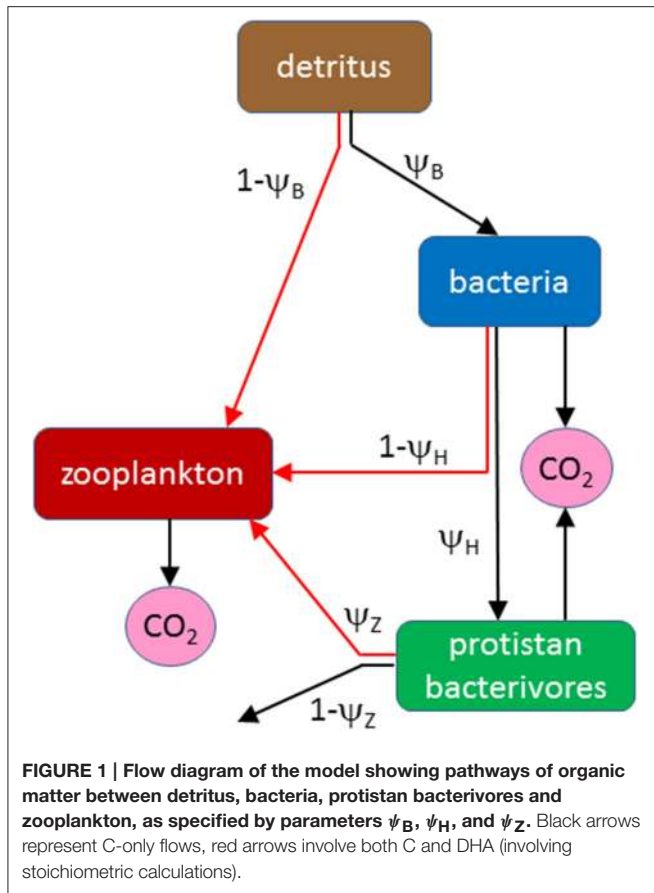
Here, we use a simple stoichiometric model to examine the extent to which invertebrates maximize growth by incorporating microbes into their diet, using detritivorous zooplankton in the mesopelagic zone (MPZ) of the ocean as a case study. The MPZ extends from the base of the sunlit (euphotic) zone down to ~1000 m and many of the resident organisms are primarily sustained by an estimated global detrital flux of 5–12 Gt C yr⁻¹ (Henson et al., 2011). The depth at which organic matter is remineralized within the MPZ influences the residence time of carbon in the oceans and hence global climate (Kwon et al., 2009). Sinking detrital particles in the MPZ exhibit the characteristic poor nutritional status described above, having undergone stripping of the most desirable compounds by bacteria and/or multiple ingestion events by zooplankton (Podgórska and Mundryk, 2003; Wilson et al., 2008). The resulting substrate is thus largely devoid of essential micronutrients such as amino or fatty acids (Wakeham et al., 1997; Fileman et al., 1998; Schneider et al., 2003). We suggest that the problem of obtaining sufficient nutrition may be felt acutely by detritivorous zooplankton that permanently reside in the MPZ, e.g., copepods

of the genus *Oithona* that are ubiquitous throughout the world ocean (Gallienne and Robins, 2001; Dahms et al., 2015). Members of this genus are well known to interact with detrital particles (Gonzalez and Smetacek, 1994; Iversen and Poulsen, 2007), particularly in the mesopelagic (Suzuki et al., 2003). Organisms inhabiting the MPZ experience high hydrostatic pressure and low temperatures, both of which negatively affect the functioning of cellular membranes (Hazel and Williams, 1990). Zooplankton overcome these difficulties by increasing the relative abundance of the essential polyunsaturated fatty acid, docosahexaenoic acid (DHA), in their membranes (Pond et al., 2014). Copepods and other highly motile zooplankton also possess myelin-like sheathes around their nerve axons to facilitate rapid escape responses (Raymont et al., 1974; Davis et al., 1999) and DHA has been suggested to be an important component of the associated sphingomyelin lipid pool (Scott et al., 2002). The model presented herein has C and DHA as currencies and is used to examine the trade-off for detritivorous zooplankton when consuming a high quantity, low DHA:C diet (detritus) versus a nutritionally-upgraded diet of microbial biomass present in low quantity, but with a high DHA:C ratio. Our analysis, which is underpinned by empirical data from a number of sources, highlights the need for improved understanding of food web processes in the mesopelagic, including the associated physiology of the resident organisms.

MODEL DESCRIPTION

Equations

The model is a steady-state flow analysis of the detrital food web in the MPZ of the ocean, including colonization of detritus by microbes (particle-attached bacteria and protistan bacterivores) and their consumption by detritivorous zooplankton (**Figure 1**; lists of model variables and parameters are provided in **Tables 1, 2**). The main focus is the growth of zooplankton and its stoichiometric regulation by C and DHA. The baseline currency of the model is C from which flows are calculated throughout the food web as a whole. Zooplankton growth, on the other hand, is calculated from stoichiometric equations involving both C and DHA. Fixed ratios (model parameters) are specified for DHA:C in detritus, bacteria and bacterivores which, in conjunction with predicted C cycling throughout the food web, permits an assessment of the roles of C and DHA in limiting the growth of zooplankton (depending on the relative availability of each food type to their diet). It is thus possible to examine the potential trade-off between consuming a high quantity, low quality diet (detritus with a low DHA:C ratio) versus a low quantity, high quality diet (microbes with a high DHA:C ratio). In this context, it is useful to define the two end-members of the nutritional spectrum: a “detritivorous pathway” and a “microbial pathway.” The former represents consumption of the non-living detrital substrate, whereas the microbial pathway consists of a diet solely of microbes. Our default assumption is that detritivorous zooplankton selectively ingest protistan bacterivores on the basis of their motility. The microbial pathway therefore represents a diet consisting solely

**TABLE 1 |** Model variables.

Variable	Definition	Unit of measure
F_D	Entry flux of D into system	$\text{mol C m}^{-3} \text{d}^{-1}$
$A_{C,det}$	Absorption C: detrit. path	$\text{mol C m}^{-3} \text{d}^{-1}$
$A_{DHA,det}$	Absorption DHA: detrit. path	$\text{mmol DHA m}^{-3} \text{d}^{-1}$
$A_{C,mic}$	Absorption C: microb. path	$\text{mol C m}^{-3} \text{d}^{-1}$
$A_{DHA,mic}$	Absorption DHA: microb path	$\text{mmol DHA m}^{-3} \text{d}^{-1}$
G_B	Bacterial production	$\text{mol C m}^{-3} \text{d}^{-1}$
G_H	Bacterivore production	$\text{mol C m}^{-3} \text{d}^{-1}$
G_Z	Zooplankton production	$\text{mol C m}^{-3} \text{d}^{-1}$

of these organisms and excludes particle-attached bacteria. The sensitivity of predicted zooplankton growth to whether or not bacteria constitute a food source will nevertheless be investigated by including the possibility of ingesting bacteria in the model structure and parameterization.

The stoichiometric calculations of zooplankton growth assume that these animals are unable to synthesize DHA *de novo* (Bell et al., 2007) in which case this essential fatty acid can be treated in the same way as elements such as C, N and P when using theoretical stoichiometry to analyze limitation of growth (Anderson and Pond, 2000). Bacteria and bacterivores are, on the other hand capable of synthesizing essential acids, including DHA, *de novo* (Klein Breteler et al., 1999; Russell and Nichols,

TABLE 2 | Model parameters.

Parameter	Definition	Default value	Unit of measure
θ_D	DHA:C, detritus	0.21	$\text{mmol DHA mol C}^{-1}$
θ_Z	DHA:C, zooplankton	1.76	$\text{mmol DHA mol C}^{-1}$
θ_B	DHA:C, bacteria	0.08	$\text{mmol DHA mol C}^{-1}$
θ_H	DHA:C, bacterivores	1.40	$\text{mmol DHA mol C}^{-1}$
ω_B	bacteria GGE	0.12	dimensionless
β_H	AE, bacterivores on bacteria	0.72	dimensionless
k_H	max. NPE, bacterivores: C	0.44	dimensionless
β_{ZC}	AE, zooplankton on D: C	0.1	dimensionless
β_{ZDHA}	AE, zooplankton on D: DHA	0.1	dimensionless
β_{ZBH}	AE, zooplankton on B,H	0.72	dimensionless
k_{ZC}	max. NPE, zooplankton: C	0.36	dimensionless
k_{ZDHA}	max. NPE, zoopl.: DHA	0.9	dimensionless
ψ_B	partitioning D to bacteria	0.1	dimensionless
ψ_H	partitioning B to bacterivores	1.0	dimensionless
ψ_Z	partitioning H to zoopl.	0.8	dimensionless

1999; Fang et al., 2002) and so their growth is calculated assuming that limitation is by C.

Detritus provides the foundation of the mesopelagic food web, specified as an input flux to the model, F_D ($\text{mol C m}^{-3} \text{d}^{-1}$). The detrital substrate is acted on by either particle-attached bacteria (fraction ψ_B) or by zooplankton (fraction $1-\psi_B$). The latter gives rise to the detritivorous pathway, which we consider first. Ingested C and DHA following this pathway, i.e., from direct consumption of non-living detritus by zooplankton, are subject to absorption efficiencies (AEs) β_{ZC} and β_{ZDHA} in which case quantities of absorbed C and DHA, $A_{C,det}$ and $A_{DHA,det}$, are:

$$A_{C,det} = (1 - \psi_B)\beta_{ZC}F_D \quad (1)$$

$$A_{DHA,det} = (1 - \psi_B)\beta_{ZDHA}\theta_D F_D \quad (2)$$

where θ_D is the DHA:C ratio in detritus (excluding microbes within the detrital matrix).

The alternative is for detritivores to obtain nutrition by consuming microbes, the “microbial pathway,” which necessitates predicting the availability of bacteria and protistan bacterivores deriving from trophic transfer within the food web. Bacteria utilize detritus with growth efficiency ω_B , from which their growth, G_B , is:

$$G_B = \psi_B\omega_B F_D \quad (3)$$

The fate of bacteria in the model is either consumption by protistan bacterivores within the particle-attached food web (fraction ψ_H) or zooplankton (fraction $1-\psi_H$); note that our default assumption is that of zero consumption by zooplankton, i.e., $\psi_H = 1$. The growth of the bacterivores, G_H , is calculated as the product of ingestion ($\psi_H G_B$), absorption efficiency (for C; parameter β_H) and net production efficiency (NPE; the fraction of absorbed C allocated to growth; parameter k_H):

$$G_H = \psi_H\beta_H k_H G_B \quad (4)$$

Total ingestion of C by zooplankton via the microbial pathway is the sum of that on bacteria, $(1-\psi_H)G_B$, and protistan bacterivores, $\psi_Z G_H$ (fraction ψ_Z of bacterivore production is utilized by zooplankton), with corresponding intake of DHA calculated from the DHA:C ratios of these food sources (θ_B and θ_H for bacteria and protistan bacterivores, respectively). The resulting quantities of absorbed C and DHA following the microbial pathway, $A_{C,mic}$ and $A_{DHA,mic}$, are then:

$$A_{C,mic} = \beta_{ZBH}((1 - \psi_H)G_B + \psi_Z G_H) \quad (5)$$

$$A_{DHA,mic} = \beta_{ZBH}((1 - \psi_H)\theta_B G_B + \psi_Z \theta_H G_H) \quad (6)$$

where β_{ZBH} is absorption efficiency for zooplankton on bacterivores (applied equally to C and DHA).

Zooplankton growth can now be calculated using established stoichiometric equations (e.g., Anderson and Hessen, 1995) that compare the relative availability of C and DHA in absorbed substrates, as supplied by both the detritivorous and microbial pathways. If C is limiting then growth, G_Z ($\text{mol C m}^{-3} \text{d}^{-1}$), is:

$$G_Z(C) = k_{ZC}(A_{C,det} + A_{C,mic}) \quad (7)$$

where parameter k_{ZC} is the maximum NPE for C (maximum k_{ZC} occurs when C is limiting; realized k_{ZC} is lower when DHA is limiting growth because C is then in stoichiometric excess). The corresponding equation for G_Z when DHA is limiting is:

$$G_Z(DHA) = k_{ZDHA}(A_{DHA,det} + A_{DHA,mic})/\theta_Z \quad (8)$$

where k_{ZDHA} is maximum net production efficiency for DHA and θ_Z is the DHA:C ratio in zooplankton biomass. Realized growth is then the minimum of the calculated C- and DHA-limited rates:

$$G_Z = \text{MIN}[G_Z(C), G_Z(DHA)] \quad (9)$$

A threshold elemental ratio (TER) can be calculated, θ_A^* , which is the optimum ratio of DHA and C in absorbed substrates for growth:

$$\theta_A^* = \frac{k_{ZC}\theta_Z}{k_{ZDHA}} \quad (10)$$

With parameters as in **Table 2** ($k_{ZC} = 0.36$, $k_{ZDHA} = 0.9$ and $\theta_Z = 1.76$), calculated θ_A^* is 0.70 meaning that optimal growth requires that each mol of absorbed C is accompanied by 0.70 mmol of absorbed DHA.

Parameterization

Model parameters fall into three categories: those specifying trophic transfer (growth efficiencies), those that define the fractionation of C between the different flow pathways in the model, and the four parameters that define DHA:C ratios in biomass. Starting with the first category, the absorption efficiency of C for zooplankton grazing on detritus, parameter β_{ZC} , was assigned a low value of 0.1 because of the refractory nature of the substrate (Bärlocher and Kendrick, 1975). The same absorption efficiency was applied to DHA, i.e., $\beta_{ZDHA} = 0.1$, thereby

assuming that zooplankton are unable to selectively extract DHA from the detritus matrix; this parameter will be subject to sensitivity analysis. Living microbes are considerably more amenable to digestion by zooplankton and so the efficiencies with which ingested bacteria and protistan bacterivores are absorbed, parameter β_{ZBH} (applied equally to both groups), was assigned a value of 0.72 (Anderson and Tang, 2010). The net production efficiency with which absorbed C is used for growth is well below 1.0 because of the energetic costs of metabolism. We set $k_{ZC} = 0.36$ based on a mean gross growth efficiency (GGE) of 0.26 for copepods (Straile, 1997) from which NPE is calculated by dividing through by AE of 0.72 (GGE is the product of AE and NPE). The role of essential fatty acids such as DHA in metabolism is not well known. The simplest assumption is that they are not heavily involved in which case DHA may be utilized for growth with high NPE e.g., $k_{ZDHA} = 0.9$ (Anderson and Pond, 2000; Mayor et al., 2009).

Moving on to the microbial food web, a typical BGE for particle-attached bacteria is 0.24 (Anderson and Tang, 2010) but this does not take into account that as much as 50% of the substrate may be lost in dissolved form through solubilization by exoenzymes (Anderson and Tang, 2010; Mayor et al., 2014). The model here does not explicitly represent solubilization losses and therefore, in practical terms, the value of 0.24 should be halved, giving $\omega_B = 0.12$. The magnitude of BGE is not well understood in marine systems and so this parameter, which sets the inflow of carbon to the microbial pathway, will be the subject of sensitivity testing. Protistan bacterivores graze on the particle-attached bacteria. As for the zooplankton, an absorption efficiency of 0.72 was applied, along with a NPE for C of 0.44 (derived from a GGE of 0.32 for flagellates: Straile, 1997), parameters β_H and k_H , respectively.

Parameters for the fractionation of C via the flow pathways in the food web, ψ_B , ψ_H , and ψ_Z , are not easy to estimate. The first of these, namely the partitioning of detritus usage between particle-attached bacteria (parameter ψ_B , leading to the microbial pathway) and detritivorous zooplankton ($1-\psi_B$; leading to the detritivorous pathway) was guesstimated at 0.75 by Anderson and Tang (2010) based on the data of Steinberg et al. (2008). An improved estimate of $\psi_B = 0.5$ was justified by Mayor et al. (2014), based on data from the North Atlantic. Most of our analysis of the model will focus on the two separate ends of the spectrum of this parameter, i.e., $\psi_B = 0,1$, in order to provide a theoretical comparison of the nutritional benefits of the detritivorous and microbial pathways in isolation to each other. Values of ψ_B that lead to optimal zooplankton nutrition are then calculated, which can be compared to the estimates above. The trophic linkages of the microbial food web on particles are not well known but it is reasonable to expect a tight coupling between bacteria and protistan bacterivores because of their close proximity (Grossart and Ploug, 2001), and thereby a high value of ψ_H . Moreover, it may be that the detritivorous zooplankton selectively ingest protistan bacterivores on the basis of their motility (Kjørboe, 2011), leaving the bacteria untouched, in which case $\psi_H = 1$ (the default value used in our analysis). The fate of flagellate biomass is even less certain. We tentatively assume that, without other obvious predators, the majority of

the flagellate loss term is available to support the growth of zooplankton and set $\psi_Z = 0.8$.

Data Sources

Studies that concurrently present data on the C and DHA content of marine seston and/or organisms are scarce, and almost non-existent for the MPZ. Parameter values for the DHA:C values in seston biomass, $\theta_D = 0.21 \text{ mmol mol}^{-1}$ (detritus), $\theta_B = 0.08$ (bacteria), $\theta_H = 1.4$ (protistan bacterivores) and $\theta_Z = 1.76$ (zooplankton) were therefore obtained from a variety of representative sources.

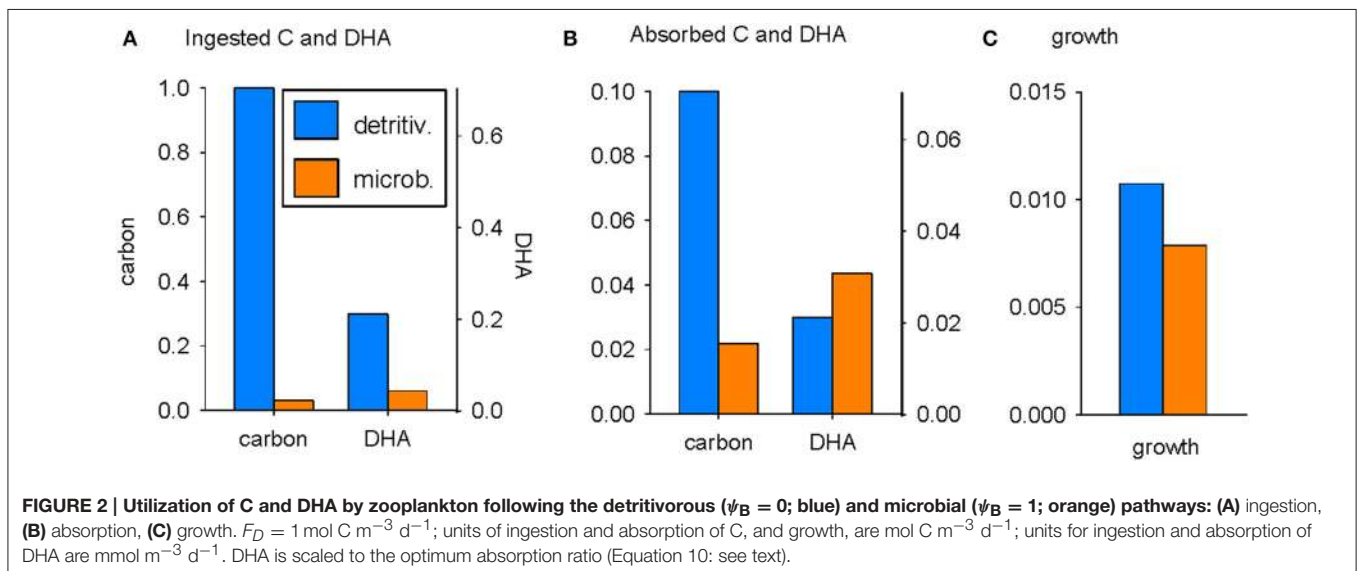
The DHA:C content of detritus ($\theta_D = 0.21 \text{ mmol mol}^{-1}$) is for seston collected on a pre-combusted GF/F filter ($0.7 \mu\text{m}$) at a depth of 215 m in the Bellingshausen Sea, Antarctica (Fileman et al., 1998). This likely represents an upper-estimate of this parameter because the sample came from the upper MPZ and the collection method made no attempt to distinguish between non-living detritus and (DHA-rich) organismal biomass. The DHA:C content of particle-attached bacteria ($\theta_B = 0.08 \text{ mmol mol}^{-1}$) represents an average value derived from various culture studies on deep-sea microbes ($\theta_B = 0.11, 0.11, 0.03$; Fang et al., 2002, 2003, 2004, respectively). The DHA:C content of protistan bacterivores ($\theta_H = 1.4 \text{ mmol mol}^{-1}$) is an average value for the heterotrophic dinoflagellate, *Oxyrrhis marina*, reared on the algae *Rhodomonas* sp. ($\theta_H = 1.54$) and *Dunaliella* sp. ($\theta_H = 1.32$) (Klein Breteler et al., 1999). An average value for the DHA:C content of zooplankton ($\theta_Z = 1.76 \text{ mmol mol}^{-1}$) was used based on published data for female copepods of the species *Oithona similis*, collected from between 400 m depth and the surface in Antarctic waters (Pond and Ward, 2011). Interested readers are guided to the relevant citations for further details of individual sample collection and analysis.

RESULTS

The main focus of the analysis presented herein is a theoretical examination of the two ends of the nutritional spectrum,

namely the detritivorous pathway ($\psi_B = 0$; zooplankton diet of non-living detritus) and the microbial pathway ($\psi_B = 1$; diet consisting solely of protistan bacterivores). This provides the most effective means of examining the trade-off between consuming a high quantity, low quality diet (detritus with a low DHA:C ratio) versus a low quantity, high quality diet (microbes with a high DHA:C ratio). The growth of zooplankton on a mixed diet incorporating both detritus and microbes will be investigated thereafter.

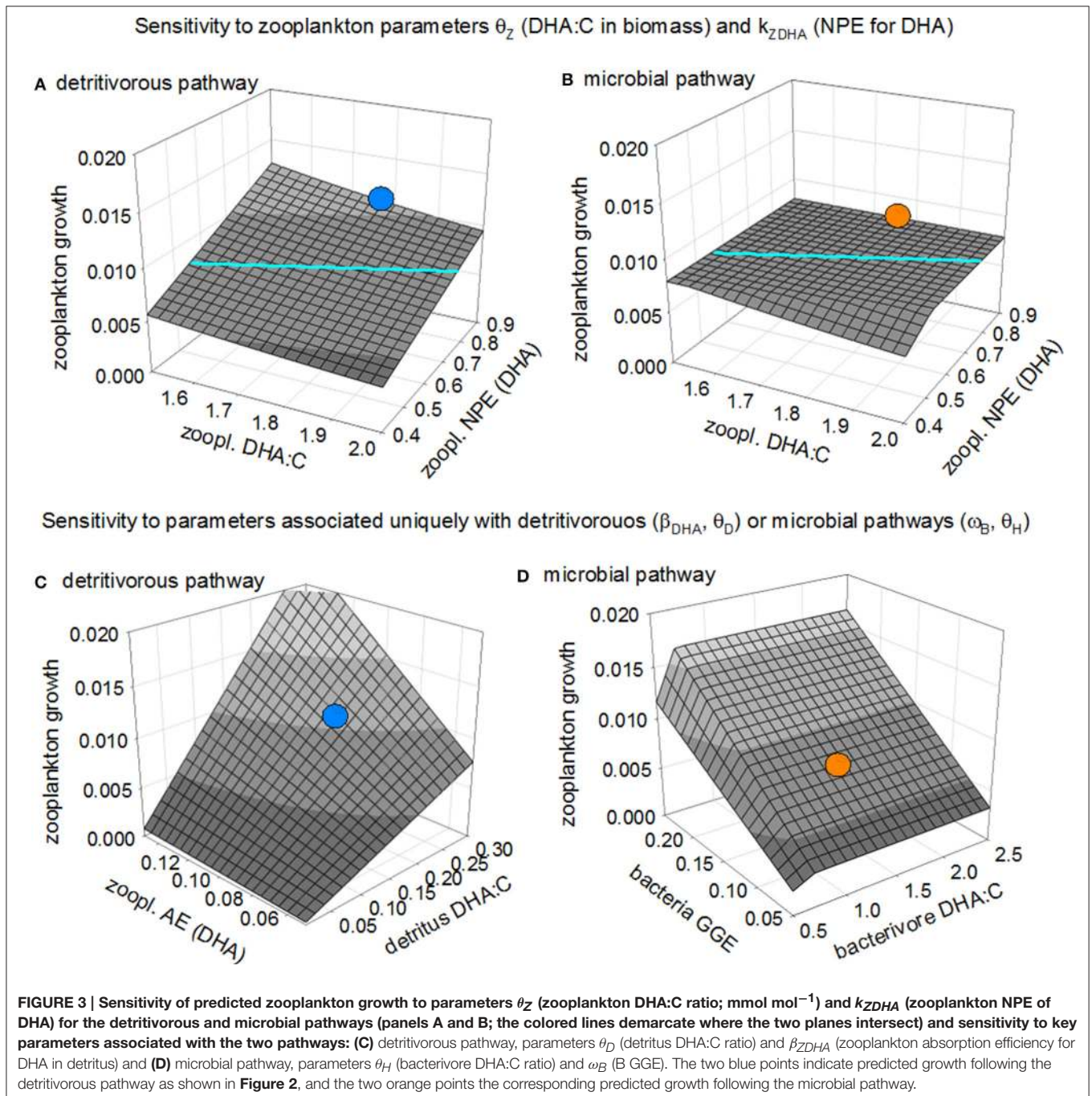
The utilization of C and DHA by zooplankton for growth, via ingestion and absorption, is compared for the detritivorous and microbial pathways in **Figure 2** (parameters as in **Table 2**). The detritus flux into the system, F_D , was nominally set at $1 \text{ mol C m}^{-3} \text{ d}^{-1}$, facilitating ease of analysis (everything is normalized to an input of 1; there is no need to use an observed value of F_D in order to compare the relative merits of the detritivorous and microbial pathways as a source of nutrition for zooplankton). The supply of C via the detritivorous pathway is plentiful whereas ingestion of C via the microbial pathway is reduced by 97% because of C losses in trophic transfer associated with the growth efficiencies of bacteria and bacterivores (**Figure 2A**). Perhaps surprisingly, detritus is also predicted to be the most plentiful source of DHA, with intake of $0.21 \text{ mmol m}^{-3} \text{ d}^{-1}$ compared to $0.043 \text{ mmol m}^{-3} \text{ d}^{-1}$ via the microbial pathway (**Figure 2A**). This is again a consequence of the much diminished stocks of bacterivore biomass compared to detritus and occurs despite the DHA:C ratio being more than six times higher in bacterivores (1.4 in bacterivores versus $0.21 \text{ mmol mol}^{-1}$ in detritus). Microbial biomass is, however, absorbed with much higher efficiency than detritus ($\beta_{ZBH} = 0.72$ versus $\beta_{ZC} = \beta_{ZDHA} = 0.1$) and so the difference in substrate supply between the two pathways is diminished post-absorption (**Figure 2B**). The absorbed quantity of DHA is greatest following the microbial pathway (0.031 vs. $0.021 \text{ mmol m}^{-3} \text{ d}^{-1}$) whereas the amount of absorbed C remains considerably lower than in the detritivorous pathway (0.022 vs. $0.1 \text{ mol C m}^{-3} \text{ d}^{-1}$).



The growth of zooplankton depends not only on quantities of absorbed substrates, but also on the net production efficiencies for DHA and C, k_{ZDHA} and k_{ZC} respectively, as well as the DHA:C ratio in biomass, θ_Z (Equations 7, 8). Note that the DHA axes in **Figure 2** are scaled to the optimal DHA:C ratio in absorbed substrates ($\theta_A^* = 0.70$; Equation 10) so that the potential for growth limitation by C or DHA can be determined by visual comparison of the bar heights for a given trophic pathway. It can be seen that predicted zooplankton growth following the detritivorous pathway is limited by DHA (the blue bar for DHA

is lower than that for C in **Figure 2B**) whereas growth following the microbial pathway is limited by C (the orange bar for C is lower than that for DHA). Overall, the assembled parameter set indicates that growth is greatest following the detritivorous pathway, although the margin is small (0.011 vs. $0.008 \text{ mol C m}^{-3} \text{ d}^{-1}$; **Figure 2C**).

We used parameter sensitivity analysis to investigate the circumstances under which predicted zooplankton growth is greatest following the microbial pathway. **Figures 3A,B** illustrate how chosen parameter values for zooplankton net production



efficiency for DHA (k_{ZDHA}) and the DHA:C in zooplankton biomass (θ_Z) influence growth following the two pathways. Zooplankton are DHA-limited in the detritivorous pathway throughout the parameter domain (**Figure 3A**). Recent work has shown that a range of aquatic invertebrates, including marine zooplankton, catabolize essential PUFAs at high rates (Mezek et al., 2010; Mayor et al., 2011, 2015; Maity et al., 2012) in which case our default zooplankton NPE for DHA of 0.9 (Anderson and Pond, 2000; Mayor et al., 2009) may be too high. Reducing the value of this parameter results in a proportional lowering of predicted zooplankton growth, to the extent that the detritivorous pathway becomes an inferior source of nutrition relative to the microbial pathway (in areas of the plane shown in **Figure 3A** that are lower than those of the corresponding parameter space shown in **Figure 3B**). Increasing the DHA:C ratio in the biomass of zooplankton, thereby increasing the demand for DHA, likewise causes a decrease in predicted growth following the detritivorous pathway. Growth following the microbial pathway is, in contrast, relatively insensitive to changing either k_{ZDHA} or θ_Z throughout most of the parameter space because limitation is by C (**Figure 3B**).

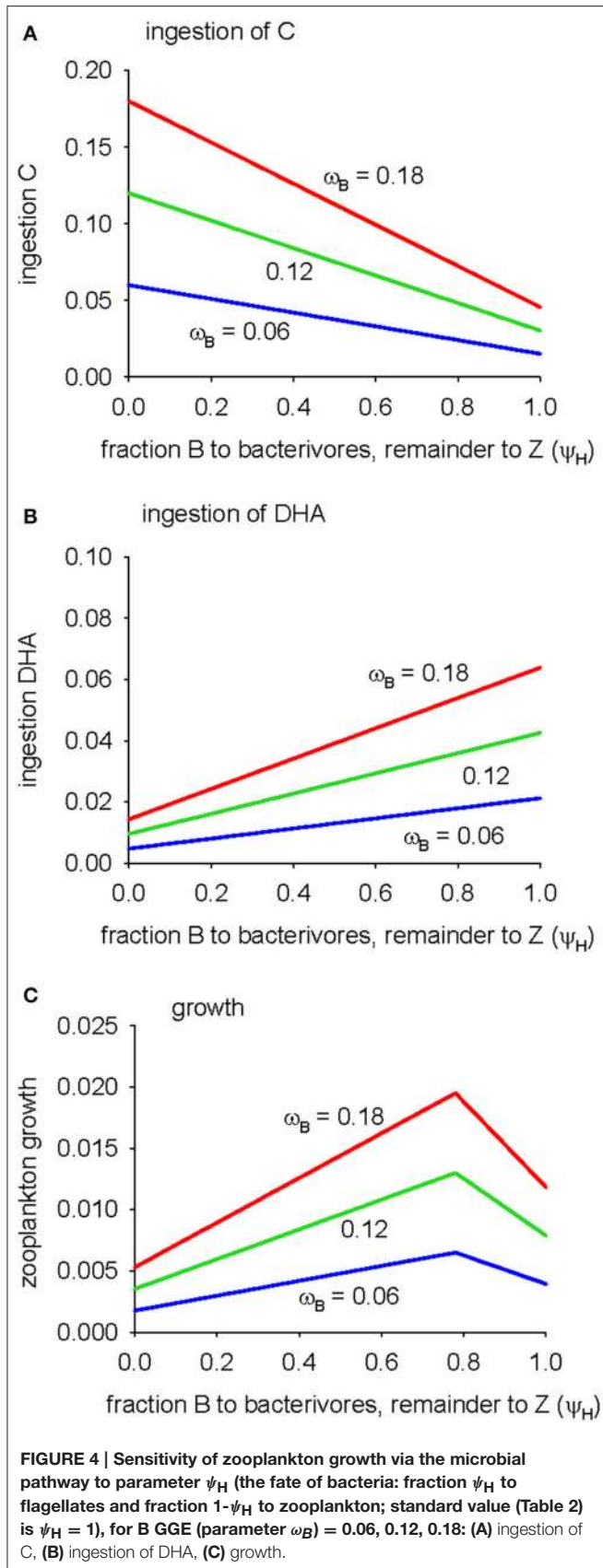
Figures 3C,D show the sensitivity of zooplankton growth to the absorption efficiency for DHA (β_{ZDHA}) and the detritus DHA:C ratio (θ_D) for the detritivorous pathway, and bacterial gross growth efficiency (ω_B) and DHA:C ratio in protistan bacterivores (θ_H) for the microbial pathway. Predicted growth following the detritivorous pathway is limited by DHA and so declines as this micronutrient becomes less available, either due to decreased absorption efficiency and/or reduced availability in detritus (**Figure 3C**). Our default value for the DHA:C of detritus ($\theta_D = 0.21$ mmol DHA mol C⁻¹) is likely too high because the samples upon which it is based were from a relatively shallow depth and did not exclude microbes from the detrital matter (see “Data sources” section), leading to overestimated growth following the detritivorous pathway. We assumed that C and DHA within detritus are absorbed by zooplankton with the same efficiency ($\beta_{ZC} = \beta_{ZDHA} = 0.1$), i.e., these animals are unable to selectively extract DHA from the detritus matrix. If they were able to do so, which is achieved in the model by increasing parameter β_{ZDHA} while keeping β_{ZC} at 0.1, the detritivorous pathway then becomes more profitable as a source of nutrition (**Figure 3C**). Growth of zooplankton following the microbial pathway shows no sensitivity to the DHA:C ratio in protistan bacterivores, except when this ratio is very low (<0.7; **Figure 3D**) because, although the bacterivores are a plentiful supply of DHA, limitation is by C. Growth does, however, increase with increasing bacterial growth efficiency because this results in more C being incorporated into the microbial food web.

In summary, the sensitivity analysis presented in **Figure 3** confirms the findings of **Figure 2**, showing the basic trade-off facing detritivorous zooplankton: a choice between consuming high quantity, low quality detritus via the detritivorous pathway which leads to limitation by DHA, or a low quantity, high quality protistan diet via the microbial pathway, with limitation by C. The analysis of **Figure 2** showed that, with the default parameter set, the growth of zooplankton was greatest following the detritivorous pathway. The trade-off choice of opting for

DHA-rich microbes (the microbial pathway) was less favorable in this instance because the losses of C due to trophic transfer in the microbial food web overrode the gains in greater DHA availability. The sensitivity analysis showed that this situation can easily be reversed by alteration of various parameter values, leading to the microbial pathway being the superior source of nutrition for zooplankton: predicted growth via the detritivorous pathway decreased when the net production efficiency for DHA (k_{ZDHA}) or the DHA:C in detritus (θ_D) are lowered, or when the DHA:C of zooplankton biomass (θ_Z) was increased. Increasing bacterial gross growth efficiency (ω_B), which promotes protistan growth, also reduced the relative effectiveness of the detrital pathway. On the other hand, the detritivorous pathway became a better source of nutrition if zooplankton were assumed to selectively absorb DHA from detritus (increase in β_{ZDHA} relative to β_{ZC}). We conclude that, given uncertainty associated with these various parameters, it is currently impossible to say with any certainty that either pathway will necessarily provide the best source of nutrition for detritivorous zooplankton in the MPZ of the ocean. The analysis has nevertheless highlighted that the microbial pathway, i.e., trophic upgrading, has the potential to be the best source of nutrition in many instances, based on results for the combinations of parameters investigated in the sensitivity analysis.

The analysis of the microbial pathway has thus far assumed that 100% of bacterial losses are due to grazing by protistan bacterivores ($\psi_H = 1$) and that bacteria do not therefore contribute to the diet of detritivorous zooplankton. Decreasing this parameter short-circuits the microbial food chain as fraction ($1 - \psi_H$) of bacteria are then consumed directly by zooplankton. Taken to the extreme ($\psi_H = 0$), all bacteria go to zooplankton. The effects of increasing the proportion of bacteria directly ingested by zooplankton ($0 \leq \psi_H \leq 1$) on predicted ingestion of C and DHA following the microbial pathway, and the resulting zooplankton growth, are shown in **Figure 4**. Bacteria constitute the base of the microbial food web and so direct access to this food source (low values of ψ_H), rather than the bacterivores one trophic level above, increases the C available to zooplankton (**Figure 4A**). On the other hand, bacterial biomass has a low DHA:C ratio and so the quantity of ingested DHA decreases as the proportion of bacteria ingested by zooplankton increases (low ψ_H ; **Figure 4B**). A point is reached, $\psi_H = 0.78$, where the supply of C and DHA is optimal and growth is maximized (**Figure 4C**). Growth is limited by C for $\psi_H > 0.78$ and by DHA for $\psi_H < 0.78$, respectively. Increasing bacterial gross growth efficiency (parameter ω_B) supplies extra DHA and C via the microbial pathway but does not influence the ratio of bacterial growth to bacterivore growth in the microbial food web and therefore has no effect on the optimum dietary intake of bacterial biomass (ψ_H). Overall, the analysis of **Figure 4** shows that C-limitation of zooplankton growth via the microbial pathway can be alleviated if these animals are able to access bacteria directly as a food source.

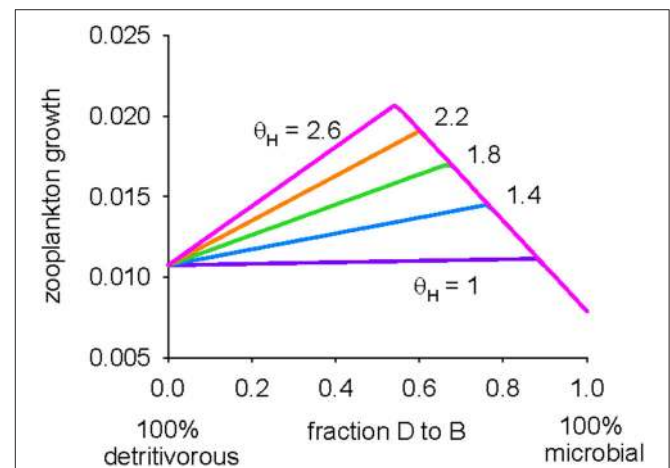
We conclude our analysis of the model by moving away from examining the detritivorous and microbial pathways in isolation from each other, and look at zooplankton growth when the two pathways are utilized simultaneously. In other words, rather than examining the two end members, the detrital pathway ($\psi_B = 0$)



and microbial pathway ($\psi_B = 1$), growth is now shown for the full range, $0 \leq \psi_B \leq 1$ (Figure 5). The growth of zooplankton is maximized when the diet consists of a mix of detritus and protistan bacterivores, irrespective of the bacterivore DHA:C ratio (θ_H). The growth of these copepods is limited by C to the right of the optimum because of C losses in the microbial food web, whereas limitation is by DHA to the left because of the low DHA content in detritus. Increasing the bacterivore DHA:C ratio offsets DHA limitation and thus increases the requirement for C in detritus in order to achieve optimal nutrition (and so the optimum ψ_B shifts to the left). Assuming that the DHA:C ratio in protistan bacterivores (θ_H) is 1.4 (Table 2), growth is maximized when ψ_B is 0.76, indicating that the optimal diet is primarily microbial.

DISCUSSION

A new model is presented and used herein to investigate the nutrition of metazoan detritivores, specifically the trade-off between consuming a diet of high-quantity, low-quality detritus versus a low-quantity, high quality diet that is rich in nutritious microbial biomass. The study focuses on the MPZ of the open ocean and involves a stoichiometric analysis of the growth of metazoan zooplankton with model currencies of C, because of its role in structural biomass and energy provisioning, and DHA, which is central to physiological adaptations to the cold temperatures and high pressures typical of the MPZ (Hazel and Williams, 1990). The model extends our previous C-only flow analysis (Mayor et al., 2014) that examined the potential gains that mesopelagic zooplankton stand to make from promoting and subsequently harvesting microbial growth via the fragmentation of large detrital particles, so-called “microbial gardening” (Fenchel, 1970). The model here was first used to compare the growth of zooplankton when consuming a diet consisting solely of non-living detritus (the “detritivorous pathway”) versus growth when consuming a



purely microbial diet (the “microbial pathway”). The microbial pathway represents “trophic upgrading” (Klein Breteler et al., 1999) of the non-living detrital substrate, i.e., consumption of the community of micronutrient-rich protistan bacterivores that colonize detritus, but which are present in low biomass because of losses in trophic transfer within the microbial food web. The conditions which maximize the growth of zooplankton were subsequently examined, where both detritus and microbes are utilized simultaneously in a mixed diet.

Our initial comparison of the two pathways, detritivorous and microbial, showed that predicted zooplankton growth could, at least in theory, be higher on the former (**Figure 2**). The nutritional benefits of consuming microbes were offset by the increased potential for zooplankton to be limited by food quantity (C). We assumed that zooplankton only had access to the protistan bacterivores in our baseline calculations, with no consumption of bacteria. The movements of motile protists, such as the myriad flagellates that colonize sinking marine detritus (Patterson et al., 1993; Turner, 2002), indicate that they should be readily detected by mechanoreceptors that are typical to copepods (Kiørboe, 2011). If zooplankton consume a diet consisting of protistan bacterivores, much of the detrital C is lost to bacterial and protistan respiration within the particle-attached microbial loop (Azam et al., 1983). This facet of the model underscores the need to understand the dynamics of microbial food webs and their interaction with higher trophic levels.

The limitation of zooplankton growth by food quantity (C) following the microbial pathway can be alleviated if direct ingestion of bacteria is possible. This short-circuits the microbial loop, removing losses of C through protistan respiration, but also lowering the DHA content of the ingested ration because the DHA:C content of bacterial biomass is considerably lower than that of their protistan predators (see Data Sources section). The potential for limitation by DHA therefore becomes more acute under this scenario, although the optimum ratio between the size of copepods of their prey (18:1; Hansen et al., 1994) suggests that direct and deliberate ingestion of bacteria by zooplankton (0.1–1 mm) is unlikely. Another possible short circuit of the microbial pathway occurs if the protists in our model are allowed to directly consume detritus (e.g., Poulsen et al., 2011). This shortening of the food chain between detritus and zooplankton via the microbial pathway is more favorable for zooplankton growth, relative to the bacteria short circuit, because the protists are rich in DHA. It follows that understanding the efficiency and structure of the microbial loop, and the trophic level at which detritivorous consumers interact with this food web, are both crucial for the development of quantitative models to explore the biogeochemistry of detrital ecosystems.

Further exploration of the model involving parameter sensitivity analysis highlighted a range of conditions where the microbial pathway is more favorable than the detritivorous pathway as a source of zooplankton nutrition. Increasing bacterial growth efficiency beyond its standard value of 0.12 is perhaps the most obvious way to achieve this, thereby directly increasing the flow of C into the microbial food web. Reported BGEs are highly variable and often very low (Steinberg et al., 2008). The stoichiometric prediction of zooplankton

growth also depends heavily on the DHA:C ratios in seston used in the calculation. These are not well known for the MPZ. Our default value for the ratio in detritus may be somewhat high because the underlying data were derived from measurements in the upper MPZ using methods that did not distinguish between detritus and the associated detrital community (see Data Sources section). Decreasing this ratio, or increasing the DHA:C ratio in zooplankton biomass, both lead to the microbial pathway becoming more favorable than the detritivorous pathway. A further assumption in the model parameterization is that zooplankton can utilize DHA with high efficiency ($k_{ZDHA} = 0.9$; **Table 2**), i.e., this essential micronutrient is solely required for physiological adaptations and is not used for energy generation (Anderson and Pond, 2000; Mayor et al., 2009). Recent observations suggest, however, that at least some marine copepods have high metabolic demands for DHA and other PUFAs (Mayor et al., 2011, 2015) and thus utilize these compounds with relatively low efficiency. Lowering the assumed efficiency with which DHA is utilized increases the demand for this essential fatty acid and so is another way of increasing the potential for the microbial pathway to be a superior source of nutrition to the detritivorous pathway. We are unaware of any data that specifically relates to the demands for DHA or other micronutrients in mesopelagic copepods and call for observations and experiments that may generate such information.

The idea that microbes support the growth of higher trophic levels is not new. An early study found that a detritus-consuming amphipod, *Parhyalella whelpleyi*, obtains its nutrition from the associated microbial communities, the non-living plant residue passing undigested through the gut (Fenchel, 1970). Stream invertebrates have also been observed to preferentially feed on leaves that have been colonized and “conditioned” by microorganisms (Kaushik and Hynes, 1971; Bärlocher and Kendrick, 1975). The nutritional environment facing detritivores has been likened to humans eating peanut butter and crackers (Cummins, 1974), microbial biomass being akin to the nutritious peanut butter spread on the indigestible crackers. Following on from this early work, a number of studies have since shown microbial biomass to be a potentially important source of nutrition for invertebrates in a range of systems including deposit-feeding mayflies (Edwards and Meyer, 1990; Hall and Meyer, 1998), leaf shredders (Connolly and Pearson, 2013), benthic polychaetes (Gontikaki et al., 2011), earthworms (Larsen et al., 2016) and other soil animals including collembolans, mites, woodlice and centipedes (Pollierer et al., 2012; Lemanski and Scheu, 2014). Recent observations have even revealed potentially important trophic linkages between detritus-associated microbes and vertebrates such as fish (e.g., Choy et al., 2015). Given the global importance of heterotrophic protists in the MPZ of the ocean (Pernice et al., 2015) and their role in biosynthesizing essential micronutrients such as DHA (Zhukova and Kharlamenko, 1999), we suggest that these organisms are highly likely to feature in the diets of metazoans that reside in this habitat.

Analysis of zooplankton ingesting a mixture of pure detritus and protistan biomass (**Figure 5**) showed that it may be that

the optimal diet involves utilization of both the detritivorous and microbial pathways in combination, with C supplied by the former balanced by DHA from the microbes. The predicted optimal diet using the standard parameter set (Table 2) contained a strong microbial component (the detritivorous and microbial pathways contributed 24 and 76% respectively to nutrition; $\psi_B = 0.76$). The analysis thus demonstrates the potential for protistan biomass to be the primary, if not the sole, part of the diet of metazoan zooplankton (Mayor et al., 2014), although this result is of course subject to the uncertainties in predicted growth highlighted by the parameter sensitivity analyses shown in Figures 3, 4. Both our study and that of Mayor et al. (2014) achieve this result, at least in part, because they are underpinned by the assumption that energy and nutrients within detritus are absorbed with much lower efficiencies than those in microbial biomass, i.e., flagellates and other soft bodied protists are more easily digested than detrital particles consisting of refractory compounds such as cellulose and chitin. We are unaware of any empirical data to directly verify this assumption, but it is supported by the conspicuous absence of flagellate remains in the guts and feces of zooplankton (reviewed by Turner, 2002), despite their long-since acknowledged significance as prey items (Stoecker and Capuzzo, 1990). We further reason that it is likely harder for zooplankton to digest and absorb detrital material, particularly as particles sink deeper into the oceans interior, because it is continuously reworked and repackaged by heterotrophic organisms that strip out anything of energetic or nutritional value (Podgórska and Mundryk, 2003; Wilson et al., 2008). The effects of this stripping are manifest as declining particulate concentrations of nitrogen and micronutrients such as fatty acids and amino acids with increasing water depth (Wakeham et al., 1997; Fileman et al., 1998; Schneider et al., 2003). An improved knowledge of the efficiencies with which mesopelagic zooplankton process different food items is required in order to further our quantitative understanding of the flows of energy and organic matter in detrital food webs. This is a particularly challenging task, potentially requiring the need for *in situ* experiments that determine absorption efficiencies and food preferences for a range of detritivorous invertebrates.

Evolving the means for internal digestion of recalcitrant organic compounds represents a stark alternative to encouraging, or even allowing, microbial growth on external particles of detritus. Recent work on terrestrial detritivores has highlighted a plethora of intricate relationships between invertebrates and their microbiome that facilitate the internal digestion of lignocellulose and other refractory molecules (König and Varma, 2006). In termites, for example, digestion of refractory material is achieved through symbiotic relationships with both bacteria and flagellates (Bignell et al., 2011; Brune, 2014). Relationships of this kind typically require the presence of one or more enlarged gut compartments to house specific microbial communities that carry out fermentation under anoxic conditions (Plante et al., 1990), such as the voluminous hindgut paunch observed in termites (Brune and Dietrich, 2015). The apparent absence of specialized gut structures in copepods commonly found in the mesopelagic, e.g., *Oithona* spp. and *Oncaea* spp., and their small size (≤ 1 mm) relative to typical detritivorous

invertebrates on land (> 10 mm), suggest that internal digestive symbioses are not particularly prevalent in midwater crustaceans. Indeed, the conspicuous difference in size between detritivorous invertebrates in terrestrial and mesopelagic ecosystems may arise because the evolutionary pressures to remain small (Kjørboe, 2011) outweigh the need for internal microbially-mediated fermentation in particle-collecting marine zooplankton. More effort is required to identify the internal microbiome of mesopelagic copepods and understand its physiological roles.

Marine detritivorous zooplankton, including *Oithona*, contain significant levels of DHA (Kattner et al., 2003; Pond and Ward, 2011) and numerous studies have highlighted the physiological roles of unsaturated fatty acids in adaptations to temperature and pressure (Cossins and Macdonald, 1989; Hazel and Williams, 1990; Pond et al., 2014). It was assumed that detritivorous invertebrates in our model have physiological requirements for DHA that cannot be met by endogenous biosynthesis, either by the copepods or their internal microbiome, i.e., DHA is an essential micronutrient. The potential for endogenous DHA biosynthesis in detritivorous copepods, by contrast, remains equivocal. Work on benthic copepods suggests that these animals may be capable of elongating shorter-chain PUFA [e.g., 18:3(n-3)] into DHA (Norsker and Støttrup, 1994; Nanton and Castell, 1998; De Troch et al., 2012), but this is not the case for epipelagic zooplankton (Bell et al., 2007). Terrestrial invertebrates are reported to obtain essential micronutrients such as amino acids and fatty acids via their biosynthesis by gut microbes (e.g., Sampedro et al., 2006; Brune, 2014) but the extent to which this occurs in marine invertebrates remains unclear (Plante et al., 1990; Harris, 1993). The guts of marine copepods are known to harbor bacteria (Sochard et al., 1979), some of which show potential for PUFA biosynthesis (Jøstensen and Landfald, 1997), but their actual role(s) within these organisms remains poorly understood. Indeed, we can find no clear evidence that marine copepods are capable of endogenous DHA biosynthesis in the absence of pre-cursor PUFAs, as we propose would be necessary for mesopelagic copepods consuming refractory detritus alone. New information on the source(s) of DHA and other micronutrients in mesopelagic detritivores will provide useful insight into the ecology and biogeochemistry of their habitat. Advances in this area may arise from examining the isotopic signatures of specific micronutrient compounds in detritivores and comparing these to the values found in autotrophic producers and mesopelagic detritus. Improved understanding of the biosynthetic capabilities of animals from the mesopelagic and the significance of internal microorganisms, potentially arising through the application of genomic, transcriptomic and metabolomic techniques, will further help resolve this knowledge gap.

In conclusion, our results indicate that ingesting nutrient-rich microbial biomass potentially represents a beneficial strategy relative to consuming refractory detritus, despite the considerable losses of C due to the inefficiency of the microbial loop. Overall, our work has highlighted how little we know about the physiology of the organisms within detritivorous food webs and hence how and why they interact with organic matter and the wider ecosystem. “Despite their global distribution and essential

roles in nutrient cycling, microbial decomposers are among the least known organisms in terms of elemental concentrations and stoichiometric relationships” (Danger et al., 2016). We suggest that better understanding the ecology and physiology of organisms in the mesopelagic is urgently required if we are to develop mechanistic biogeochemical models of this important ecosystem.

AUTHOR CONTRIBUTIONS

TA led the work, developing the model and generating the results, with a major contribution from DM in terms of advising on parameterizations, data, and in the analysis and writing of the

manuscript. DP provided additional advice, especially regarding the fatty acid work described in the manuscript.

ACKNOWLEDGMENTS

TA, DP, and DM are funded by the Natural Environment Research Council (NERC), UK. This work contributes to the NERC-funded programme “Controls over Ocean Mesopelagic Interior Carbon Storage” (COMICS), NE/M020835/1 and the “Culture Collection of Algae and Protozoa” (CCAP) National Capability. We wish to thank three reviewers for their constructive critique of the manuscript.

REFERENCES

- Anderson, N. H., and Sedell, J. R. (1979). Detritus processing by macroinvertebrates in stream ecosystems. *Ann. Rev. Entomol.* 24, 351–377. doi: 10.1146/annurev.en.24.010179.002031
- Anderson, T. R., Boersma, M., and Raubenheimer, D. (2004). Stoichiometry: linking elements to biochemicals. *Ecology* 85, 1193–1202. doi: 10.1890/02-0252
- Anderson, T. R., and Hessen, D. O. (1995). Carbon or nitrogen limitation of marine copepods? *J. Plankton Res.* 19, 317–331. doi: 10.1093/plankt/17.2.317
- Anderson, T. R., and Pond, D. W. (2000). Stoichiometric theory extended to micronutrients: comparison of the roles of essential fatty acids, carbon, and nitrogen in the nutrition of marine copepods. *Limnol. Oceanogr.* 45, 1162–1167. doi: 10.4319/lo.2000.45.5.1162
- Anderson, T. R., and Tang, K. W. (2010). Carbon cycling and POC turnover in the mesopelagic zone of the ocean: insights from a simple model. *Deep-Sea Res. Part II Topic. Stud. Oceanogr.* 57, 1581–1592. doi: 10.1016/j.dsr2.2010.02.024
- Azam, F., Fenchel, T., Field, J. G., Gray, J. S., Meyer-Reil, L. A., and Thingstad, F. (1983). The ecological role of water-column microbes in the sea. *Mar. Ecol. Prog. Ser.* 10, 257–263. doi: 10.3354/meps010257
- Bärlocher, F., and Kendrick, B. (1975). Leaf-conditioning by microorganisms. *Oecologia* 20, 359–362. doi: 10.1007/BF00345526
- Bartels, P., Cucherousset, J., Steger, K., Eklöv, P., Tranvik, L. J., and Hillebrand, H. (2012). Reciprocal subsidies between freshwater and terrestrial ecosystems structure consumer resource dynamics. *Ecology* 93, 1173–1182. doi: 10.1890/11-1210.1
- Bell, M. V., Dick, J. R., Anderson, T. R., and Pond, D. W. (2007). Application of liposome and stable isotope tracer techniques to study polyunsaturated fatty acid biosynthesis in marine zooplankton. *J. Plankton Res.* 29, 417–422. doi: 10.1093/plankt/fbm025
- Bignell, D. E., Roisin, Y., and Lo, N. (eds.). (2011). *Biology of Termites: A Modern Synthesis*. Dordrecht: Springer.
- Brune, A. (2014). Symbiotic digestion of lignocellulose in termite guts. *Nat. Rev.* 12, 168–180. doi: 10.1038/nrmicro3182
- Brune, A., and Dietrich, C. (2015). The gut microbiota of termites: digesting the diversity in the light of ecology and evolution. *Annu. Rev. Microbiol.* 69, 145–166. doi: 10.1146/annurev-micro-092412-155715
- Burd, A. B., Frey, S., Cabre, A., Ito, T., Levine, N. M., Lønborg, C., et al. (2016). Terrestrial and marine perspectives on modeling organic matter degradation pathways. *Glob. Change Biol.* 22, 121–136. doi: 10.1111/gcb.12987
- Cebrián, J., and Duarte, C. M. (1995). Plant growth-rate dependence of detrital carbon storage in ecosystems. *Science* 268, 1606–1608. doi: 10.1126/science.268.5217.1606
- Choy, C. A., Popp, B. N., Hannides, C. C. S., and Drazen, J. C. (2015). Trophic structure and food resources of epipelagic and mesopelagic fishes in the North Pacific Subtropical Gyre ecosystem inferred from nitrogen isotopic compositions. *Limnol. Oceanogr.* 60, 1156–1171. doi: 10.1002/lno.10085
- Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., et al. (2013). “Carbon and other biogeochemical cycles,” in *Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*, eds T. F. Stocker, D. Qin, G. K. Plattner, M. Tignor, S. K. Allen, J. Boschung, A. Nauels, Y. Xia, V. Bex, and P. M. Midgley (Cambridge: Cambridge University Press).
- Connolly, N. M., and Pearson, R. G. (2013). Nutrient enrichment of a heterotrophic stream alters leaf litter nutritional quality and shredder physiological condition via the microbial pathway. *Hydrobiologia* 718, 85–92. doi: 10.1007/s10750-013-1605-7
- Cossins, A. R., and Macdonald, A. G. (1989). The adaptation of biological membranes to temperature and pressure: fish from the deep and cold. *J. Bioenerg. Biomembr.* 21, 115–135. doi: 10.1007/BF00762215
- Cowie, G. L., and Hedges, J. I. (1996). Digestion and alteration of the biochemical constituents of a diatom (*Thalassiosira weissflogii*) ingested by an herbivorous zooplankton (*Calanus pacificus*). *Limnol. Oceanogr.* 41, 581–594. doi: 10.4319/lo.1996.41.4.0581
- Cummins, K. W. (1974). Structure and function of stream ecosystems. *BioScience* 24, 631–641. doi: 10.2307/1296676
- Dahms, H.-U., Tseng, L. C., and Hwang, J. S. (2015). Biogeographic distribution of the cyclopoid copepod genus *Oithona* - from mesoscales to global scales. *J. Exp. Mar. Biol. Ecol.* 467, 26–32. doi: 10.1016/j.jembe.2015.02.009
- Danger, M., Gessner, M. O., and Bärlocher, F. (2016). Ecological stoichiometry of aquatic fungi: current knowledge and perspectives. *Fungal Ecol.* 19, 100–111. doi: 10.1016/j.funeco.2015.09.004
- Davis, A. D., Weatherby, T. M., Hartline, D. K., and Lenz, P. H. (1999). Myelin-like sheaths in copepod axons. *Nature* 398:571. doi: 10.1038/19212
- Del Giorgio, P. A., and Cole, J. J. (1998). Bacterial growth efficiency in natural aquatic systems. *Annu. Rev. Ecol. Syst.* 29, 503–541. doi: 10.1146/annurev.ecolsys.29.1.503
- De Troch, M., Boeckx, P., Cnudde, C., Van Gansbeke, D., Vanreusel, A., Vincx, M., and Caramujo, M. J. (2012). Bioconversion of fatty acids at the basis of marine food webs: insights from a compound-specific stable isotope analysis. *Mar. Ecol. Prog. Ser.* 465, 53–67. doi: 10.3354/meps09920
- Edwards, R. T., and Meyer, J. L. (1990). Bacterivory by deposit-feeding may fly larvae (*Stenonema* spp.). *Freshwater Biol.* 24, 453–462. doi: 10.1111/j.1365-2427.1990.tb00724.x
- Fang, J., Barcelona, M. J., Abrajano, T., Nogi, Y., and Kato, C. (2002). Isotopic composition of fatty acids of extremely piezophilic bacteria from the Mariana Trench at 11,000 m. *Mar. Chem.* 80, 1–9. doi: 10.1016/S0304-4203(02)00069-5
- Fang, J., Chan, O., Kato, C., Sato, T., Peeples, T., and Niggemeyer, K. (2003). Phospholipid FA of piezophilic bacteria from the Deep Sea. *Lipids* 38, 885–887. doi: 10.1007/s11745-003-1140-7
- Fang, J., Kato, C., Sato, T., Chan, O., and McKay, D. (2004). Biosynthesis and dietary uptake of polyunsaturated fatty acids by piezophilic bacteria. *Comp. Biochem. Physiol. B. Biochem. Mol. Biol.* 137, 455–461. doi: 10.1016/j.cbpc.2004.01.009
- Fenchel, T. (1970). Studies on the decomposition of organic detritus derived from the turtle grass *Thalassia testudinum*. *Limnol. Oceanogr.* 15, 14–20. doi: 10.4319/lo.1970.15.1.0014
- Fileman, T. W., Pond, D. W., Barlow, R. G., and Mantoura, R. F. C. (1998). Vertical profiles of pigments, fatty acids and amino acids: evidence for

- undegraded diatomaceous material sedimenting to the deep ocean in the Bellingshausen Sea, Antarctica. *Deep Sea Res. I Oceanogr. Res. Papers* 45, 333–346. doi: 10.1016/S0967-0637(97)00824-8
- Gallienne, C. P., and Robins, D. B. (2001). Is *Oithona* the most important copepod in the world's oceans? *J. Plankton Res.* 23, 1421–1432. doi: 10.1093/plankt/23.12.1421
- Gontikaki, E., van Oevelen, D., Soetaert, K., and Witte, U. (2011). Food web flows through a sub-arctic deep-sea benthic community. *Prog. Oceanogr.* 91, 245–259. doi: 10.1016/j.pcean.2010.12.014
- Gonzalez, H. E., and Smetacek, V. (1994). The possible role of the cyclopoid copepod *Oithona* in retarding vertical flux of zooplankton faecal material. *Mar. Ecol. Prog. Ser.* 113, 233–246. doi: 10.3354/meps113233
- Grossart, H.-P., and Ploug, H. (2001). Microbial degradation of organic carbon and nitrogen on diatom aggregates. *Limnol. Oceanogr.* 46, 267–277. doi: 10.4319/lo.2001.46.2.0267
- Hall, R. O. Jr., and Meyer, J. L. (1998). The trophic significance of bacteria in a detritus-based stream food web. *Ecology* 79, 1995–2012. doi: 10.1890/0012-9658(1998)079[1995:TTSOBI]2.0.CO;2
- Hansen, B., Bjørnsen, P. K., and Hansen, P. J. (1994). The size ratio between planktonic predators and their prey. *Limnol. Oceanogr.* 39, 395–403. doi: 10.4319/lo.1994.39.2.0395
- Harris, J. M. (1993). The presence, nature, and role of gut microflora in aquatic invertebrates: a synthesis. *Microb. Ecol.* 25, 195–231. doi: 10.1007/BF00171889
- Hazel, J. R., and Williams, E. E. (1990). The role of alterations in membrane lipid composition in enabling physiological adaptation of organisms to their physical environment. *Prog. Lipid Res.* 29, 167–227. doi: 10.1016/0163-7827(90)90002-3
- Henson, S. A., Sanders, R., Madsen, E., Morris, P. J., Le Moigne, F., and Quartly, G. D. (2011). A reduced estimate of the strength of the ocean's biological carbon pump. *Geophys. Res. Lett.* 38:L04606. doi: 10.1029/2011GL046735
- Iversen, M. H., and Poulsen, L. (2007). Coprophagy, coprophagy, and coprochaly in the copepods *Calanus helgolandicus*, *Pseudocalanus elongatus*, and *Oithona similis*. *Mar. Ecol. Prog. Ser.* 350, 79–89. doi: 10.3354/meps07095
- Jøstensen, J. P., and Landfald, B. (1997). High prevalence of polyunsaturated fatty acid producing bacteria in Arctic invertebrates. *FEMS Microbiol. Lett.* 151, 95–101. doi: 10.1016/S0378-1097(97)00148-1
- Kattner, G., Albers, C., Graeve, M., and Schnack-Schiel, S. B. (2003). Fatty acid and alcohol composition of the small polar copepods, *Oithona* and *Oncaea*: indication on feeding modes. *Polar Biol.* 26, 666–671. doi: 10.1007/s00300-003-0540-x
- Kaushik, N. K., and Hynes, H. B. N. (1971). The fate of dead leaves that fall into streams. *Arch. Hydrobiol.* 68, 465–515.
- Kiem, R., and Kögel-Knabner, I. (2003). Contribution of lignin and polysaccharides to the refractory carbon pool in C-depleted arable soils. *Soil Biol. Biochem.* 35, 101–118. doi: 10.1016/S0038-0717(02)00242-0
- Kjørboe, T. (2011). How zooplankton feed: mechanisms, traits and tradeoffs. *Biol. Rev.* 86, 311–340. doi: 10.1111/j.1469-185X.2010.00148.x
- Klein Breteler, W. C. M., Schogt, N., Baas, M., Schouten, S., and Kraay, G. W. (1999). Dynamics of bacterial community composition during degradation of copepod fecal pellets. *Mar. Biol.* 135, 191–198. doi: 10.1007/s002270050616
- König, H., and Varma, A. (eds.). (2006). *Intestinal Microorganisms of Termites and Other Invertebrates*. New York, NY: Springer.
- Koski, M., Kjørboe, T., and Takahashi, K. (2005). Benthic life in the pelagic: aggregate encounter and degradation rates by pelagic harpacticoid copepods. *Limnol. Oceanogr.* 50, 1254–1263. doi: 10.4319/lo.2005.50.4.1254
- Kwon, E. Y., Primeau, F., and Sarmiento, J. L. (2009). The impact of remineralization depth on the air-sea carbon balance. *Nat. Geosci.* 2, 630–635. doi: 10.1038/ngeo612
- Larsen, T., Pollierer, M. M., Holmstrup, M., D'Annibale, A., Maraldo, K., Andersen, N., et al. (2016). Substantial nutritional contribution of bacterial amino acids to earthworms and enchytraeids: a case study from organic grasslands. *Soil Biol. Biochem.* 99, 21–27. doi: 10.1016/j.soilbio.2016.03.018
- Lawrence, S. G., Ahmad, A., and Azam, F. (1993). Fate of particle bound bacteria ingested by *Calanus pacificus*. *Mar. Ecol. Prog. Ser.* 97, 299–307. doi: 10.3354/meps097299
- Lemanski, K., and Scheu, S. (2014). Fertilizer addition lessens the flux of microbial carbon to higher trophic levels in soil food webs of grassland. *Oecologia* 176, 487–496. doi: 10.1007/s00442-014-3037-0
- Luo, Y., Ahlstrom, A., Allison, S. D., Batjes, N. H., Brovkin, V., Carvalhais, N., et al. (2016). Toward more realistic projections of soil carbon dynamics by Earth system models. *Global Biogeochem. Cycles* 30, 40–56. doi: 10.1002/2015GB005239
- Maity, S., Jannasch, A., Adamec, J., Gribskov, M., Nalepa, T., Höök, T. O., et al. (2012). Metabolite profiles in starved *Diporeia* spp. using liquid chromatography-mass spectrometry (LC-MS) based metabolomics. *J. Crustacean Biol.* 32, 239–248. doi: 10.1163/193724011X615578
- Mann, K. H. (1988). Production and use of detritus in various freshwater, estuarine, and coastal marine systems. *Limnol. Oceanogr.* 33, 910–930.
- Mayor, D. J., Anderson, T. R., Pond, D. W., and Irigoien, X. (2009). Limitation of egg production in *Calanus finmarchicus* in the field: a stoichiometric analysis. *J. Mar. Syst.* 78, 511–517. doi: 10.1016/j.jmarsys.2008.12.020
- Mayor, D. J., Cook, K., Thornton, B., Walsham, P., Witte, U. F. M., Zuur, A. F., et al. (2011). Absorption efficiencies and basal turnover of C, N and fatty acids in a marine Calanoid copepod. *Funct. Ecol.* 25, 509–518. doi: 10.1111/j.1365-2435.2010.01791.x
- Mayor, D. J., Sanders, R., Giering, S. L. C., and Anderson, T. R. (2014). Microbial gardening in the ocean's twilight zone: detritivorous metazoans benefit from fragmenting, rather than ingesting, sinking detritus. *BioEssays* 36, 1132–1137. doi: 10.1002/bies.201400100
- Mayor, D. J., Sommer, U., Cook, K. B., and Viant, M. R. (2015). The metabolic response of marine copepods to environmental warming and ocean acidification in the absence of food. *Nat. Sci. Rep.* 5:13690. doi: 10.1038/srep13690
- Mezek, T., Simčič, T., Arts, M. T., and Brancelj, A. (2010). Effect of fasting on hypogean (*Niphargus stygius*) and epigean (*Gammarus fossarum*) amphipods: a laboratory study. *Aquat. Ecol.* 44, 397–408. doi: 10.1007/s10452-009-9299-7
- Moran, M. A., and Hodson, R. E. (1989). Bacterial secondary production on vascular plant detritus: relationships to detritus composition and degradation rate. *Appl. Environ. Microbiol.* 55, 2178–2189.
- Müller-Navarra, D. C., Brett, M. T., Liston, A. M., and Goldman, C. R. (2000). A highly unsaturated fatty acid predicts carbon transfer between primary producers and consumers. *Nature* 403, 74–77. doi: 10.1038/47469
- Nanton, D. A., and Castell, J. D. (1998). The effect of dietary fatty acids on the fatty acid composition of the harpacticoid copepod, *Tisbe* sp. for use as a live food for marine fish larvae. *Aquaculture* 163, 251–261. doi: 10.1016/S0044-8486(98)00236-1
- Norsker, N.-H., and Støttrup, J. G. (1994). The importance of dietary HUFAs for fecundity and HUFA content in the harpacticoid, *Tisbe holothuriae* Humes. *Aquaculture* 125, 155–166. doi: 10.1016/0044-8486(94)90292-5
- Patterson, D. J., Nygaard, K., Steinberg, G., and Turley, C. M. (1993). Heterotrophic flagellates and other protists associated with oceanic detritus throughout the water column in the mid North Atlantic. *J. Mar. Biol. Assoc. U.K.* 73, 67–95. doi: 10.1017/S0025315400032653
- Pernice, M. C., Forn, I., Gomes, A., Lara, E., Alonso-Saez, L., Arrieta, J. M., et al. (2015). Global abundance of planktonic heterotrophic protists in the deep ocean. *ISME J.* 9, 782–792. doi: 10.1038/ismej.2014.168
- Phillips, N. W. (1984). Role of different microbes and substrates as potential supplies of specific, essential nutrients to marine detritivores. *Bull. Mar. Sci.* 35, 283–298.
- Plante, C. J., Jumars, P. A., and Baross, J. A. (1990). Digestive associations between marine detritivores and bacteria. *Annu. Rev. Ecol. Syst.* 21, 93–127. doi: 10.1146/annurev.es.21.110190.000521
- Podgórska, B., and Mundryk, Z. J. (2003). Distribution and enzymatic activity of heterotrophic bacteria decomposing selected macromolecular compounds in a Baltic Sea sandy beach. *Est. Coast. Shelf Sci.* 56, 539–546. doi: 10.1016/S0272-7714(02)00204-4
- Pokarzhenskii, A. D., Zaboyev, D. P., Ganin, G. N., and Gordienko, S. A. (1997). Amino acids in earthworms: are earthworms ecosystemivorous. *Soil Biol. Biochem.* 29, 559–567. doi: 10.1016/S0038-0717(96)00180-0
- Pollierer, M. M., Dyckmans, J., Scheu, S., and Haubert, D. (2012). Carbon flux through fungi and bacteria into the forest soil animal food web as indicated by compound-specific ¹³C fatty acid analysis. *Funct. Ecol.* 26, 978–990. doi: 10.1111/j.1365-2435.2012.02005.x
- Pond, D. W., Tarling, G. A., and Mayor, D. J. (2014). Hydrostatic pressure and temperature effects on the membranes of a seasonally migrating marine copepod. *PLoS ONE* 9:e111043. doi: 10.1371/journal.pone.0111043

- Pond, D. W., and Ward, P. (2011). Importance of diatoms for *Oithona* in Antarctic waters. *J. Plankton Res.* 33, 105–1181. doi: 10.1093/plankt/fbq089
- Poulsen, L. K., Moldrup, M., Berge, T., and Hansen, P. J. (2011). Feeding on copepod fecal pellets: a new trophic role of dinoflagellates as detritivores. *Mar. Ecol. Prog. Ser.* 441, 65–78. doi: 10.3354/meps09357
- Raymont, J. E. G., Krishnaswamy, S., Woodhouse, M. A., and Griffin, R. L. (1974). Studies on the fine structure of Copepoda: observations on *Calanus finmarchicus* (Gunnerus). *Proc. R. Soc. Lond. B* 185, 409–424. doi: 10.1098/rspb.1974.0026
- Russell, N. J., and Nichols, D. S. (1999). Polyunsaturated fatty acids in marine bacteria – a dogma rewritten. *Microbiology* 145, 767–779. doi: 10.1099/13500872-145-4-767
- Sampedro, L., Jeannotte, R., and Whalen, J. K. (2006). Trophic transfer of fatty acids from gut microbiota to the earthworm *Lumbricus terrestris* L. *Soil Biol. Biochem.* 38, 2188–2198. doi: 10.1016/j.soilbio.2006.02.001
- Schneider, B., Schlitzer, R., Fischer, G., and Nöthig, E.-M. (2003). Depth-dependent elemental compositions of particulate organic matter (POM) in the ocean. *Global Biogeochem. Cycles* 2:1032. doi: 10.1029/2002gb001871
- Scott, C. L., Kwasniewski, S., Falk-Petersen, S., and Sargent, J. R. (2002). Species differences, origins and functions of fatty alcohols and fatty acids in the wax esters and phospholipids of *Calanus hyperboreus*, *C. glacialis* and *C. finmarchicus* from Arctic waters. *Mar. Ecol. Prog. Ser.* 235, 127–134. doi: 10.3354/meps235127
- Sochard, M. R., Wilson, D. F., Austin, B., and Colwell, R. R. (1979). Bacteria associated with the surface and gut of marine copepods. *Appl. Environ. Microbiol.* 37, 570–579.
- Steinberg, D. K., Van Mooy, B. A. S., Buesseler, K. O., Boyd, P. W., Kobari, T., and Karl, D. M. (2008). Bacterial vs. zooplankton control of sinking particle flux in the ocean's twilight zone. *Limnol. Oceanogr.* 53, 1327–1338. doi: 10.4319/lo.2008.53.4.1327
- Stoecker, D. K., and Capuzzo, J. M. (1990). Predation on protozoa: its importance to zooplankton. *J. Plankton Res.* 12, 891–908. doi: 10.1093/plankt/12.5.891
- Straile, D. (1997). Gross growth efficiencies of protozoan and metazoan zooplankton and their dependence on food concentration, predator-prey weight ratio, and taxonomic group. *Limnol. Oceanogr.* 42, 1375–1385. doi: 10.4319/lo.1997.42.6.1375
- Suzuki, H., Sasaki, H., and Fukuchi, M. (2003). Loss processes of sinking fecal pellets of zooplankton in the mesopelagic layers of the Antarctic marginal ice zone. *J. Oceanogr.* 59, 809–818. doi: 10.1023/B:JOCE.0000009572.08048.0d
- Turley, C. M., and Mackie, P. J. (1994). Biogeochemical significance of attached and free-living bacteria and the flux of particles in the NE Atlantic Ocean. *Mar. Ecol. Prog. Ser.* 115, 191–203. doi: 10.3354/meps115191
- Turner, J. T. (2002). Zooplankton fecal pellets, marine snow and sinking phytoplankton blooms. *Aquat. Microb. Ecol.* 27, 57–102. doi: 10.3354/ame027057
- Wakeham, S. G., Hedges, J. I., Lee, C., Peterson, M. L., and Hernes, P. J. (1997). Compositions and transport of lipid biomarkers through the water column and surficial sediments of the equatorial Pacific ocean. *Deep-Sea Res. Part II Topic. Stud. Oceanogr.* 44, 2131–2162. doi: 10.1016/S0967-0645(97)00035-0
- Wilson, S. E., Steinberg, D. K., and Buesseler, K. O. (2008). Changes in fecal pellet characteristics with depth as indicators of zooplankton repackaging of particles in the mesopelagic zone of the subtropical and subarctic North Pacific Ocean. *Deep-Sea Res. Part II* 55, 1636–1647. doi: 10.1016/j.dsr2.2008.04.019
- Zhukova, N. V., and Kharlamenko, V. I. (1999). Source of essential fatty acids in the marine microbial loop. *Aquat. Microb. Ecol.* 17, 153–157. doi: 10.3354/ame017153

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Anderson, Pond and Mayor. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.