



## The Role of the Gut-Liver Axis in Metabolic Dysfunction-Associated Fatty Liver Disease

Rosa Martín-Mateos and Agustín Albillos\*

Department of Gastroenterology and Hepatology, Hospital Universitario Ramón y Cajal, Universidad de Alcalá, Instituto Ramón y Cajal de Investigación Sanitaria (IRYCIS), Centro de Investigación Biomédica en Red de Enfermedades Hepáticas y Digestivas (CIBERehd), Instituto de Salud Carlos III, Madrid, Spain

The complex interplay between the gut microbiota, the intestinal barrier, the immune system and the liver is strongly influenced by environmental and genetic factors that can disrupt the homeostasis leading to disease. Among the modulable factors, diet has been identified as a key regulator of microbiota composition in patients with metabolic syndrome and related diseases, including the metabolic dysfunction-associated fatty liver disease (MAFLD). The altered microbiota disrupts the intestinal barrier at different levels inducing functional and structural changes at the mucus lining, the intercellular junctions on the epithelial layer, or at the recently characterized vascular barrier. Barrier disruption leads to an increased gut permeability to bacteria and derived products which challenge the immune system and promote inflammation. All these alterations contribute to the pathogenesis of MAFLD, and thus, therapeutic approaches targeting the gut-liver-axis are increasingly being explored. In addition, the specific changes induced in the intestinal flora may allow to characterize distinctive microbial signatures for non-invasive diagnosis, severity stratification and disease monitoring.

#### **OPEN ACCESS**

#### Edited by:

Javier Crespo, Marqués de Valdecilla Health Research Institute (IDIVAL), Spain

#### Reviewed by:

Javier González-Gallego, University of León, Spain Vanessa Figliuolo da Paz, University of Arizona, United States

\*Correspondence: Agustín Albillos agustin.albillos@uah.es

#### Specialty section:

This article was submitted to Inflammation, a section of the journal Frontiers in Immunology

Received: 28 January 2021 Accepted: 29 March 2021 Published: 16 April 2021

#### Citation:

Martín-Mateos R and Albillos A (2021) The Role of the Gut-Liver Axis in Metabolic Dysfunction-Associated Fatty Liver Disease. Front. Immunol. 12:660179. doi: 10.3389/fimmu.2021.660179 Keywords: gut abnormalities, steatosis, intestinal immunity, intestinal microbiota, metabolic liver disease

## INTRODUCTION

Metabolic dysfunction-associated fatty liver disease (MAFLD), previously known as non-alcoholic fatty liver disease (NAFLD), is a major cause of liver-related morbimortality, with an estimated global prevalence of 25.24% (1). The burden of MAFLD is expected to increase paralleling the growing incidence of obesity and diabetes mellitus (2). MAFLD represents the hepatic manifestation of an underlying multisystemic metabolic dysfunction, and, thus, patients are not only at risk of liver-related complications (cirrhosis and/or hepatocellular carcinoma), but also cardiometabolic-related events, which are the leading cause of mortality (1). The term reflects the wide spectrum of the disease, which ranges from simple steatosis, to liver inflammation (steatohepatitis), fibrosis and cirrhosis. The extent of fibrosis is the strongest predictor of overall and liver-specific mortality (3).

Recently, the diagnostic criteria for MAFLD have been reformulated, and now are based on the evidence of hepatic steatosis (demonstrated by biopsy, imaging or validated blood biomarkers), in addition to one of the following criteria: overweight/obesity, type 2 diabetes mellitus, or metabolic dysregulation defined by the presence of at least two metabolic risk factors (4). Therefore, currently

it is not mandatory to exclude alcohol consumption or other concomitant liver diseases to make a positive diagnosis of MAFLD, as it can frequently coexist with other conditions.

The interaction between the gut, its microbiome and the liver, is key to the pathogenesis of MAFLD, and thus, a significant number of reviews addressing the role of the gut-liver axis have been published over the past few years (5–7). This evolving field has recently achieved promising therapeutic advances based on microbiota composition manipulation. In addition, the characterization of distinctive microbiota signatures for disease diagnosis and stratification is increasingly been explored. In light of the new evidence, this review aims to summarize the characteristic alterations at the different components of the gut-liver axis, their contribution to the pathogenesis of MAFLD, and the most recent and clinically relevant advances involving the gut-liver axis as a potential therapeutic target and non-invasive diagnostic tool in patients with MAFLD.

# THE ROLE OF THE GUT-LIVER AXIS IN LIVER DISEASE

The term gut-liver-axis has been coined to highlight the close interaction between the intestine and the liver, which also involves a complex interplay with the gut microbiome and the immune system (8). The portal vein serves as the main functional link leading nutrients and gut-derived antigens toward the liver, which in turn provides bile acids, proteins, lipids and immune components. Although highly specialized epithelial and vascular barriers regulate the transport across the intestinal mucosa, a wide variety of microbial products and bacteria reach the liver through the portal circulation. Most of them are harmless dietary and commensal products, and thus, the immune system cells involved in surveillance remain tolerant upon recognition. However, when gut-derived bacterial pathogens reach the liver sinusoids, an effective immune response is activated in order to prevent their spread through the systemic circulation. The disruption of the gutliver axis alters the balance between immune activation and tolerance (9), and the subsequent immune dysfunction critically contributes to the pathogenesis and progression of liver diseases, and in particular, to MAFLD (5).

## Gut Microbiota as a Key Regulator of the Metabolic Profile

The gut microbiota comprises a diverse spectrum of microorganisms involved in metabolic, synthetic and regulatory functions, including fermentation of non-digestible dietary substrates, synthesis of vitamins, bile acids metabolism, regulation of epithelial cell proliferation or modulation of the inflammatory response (10). The gut microbiota also hinders pathogen colonization by competing for nutrients and space, and thus, it represents the first-line defense of the intestinal barrier.

Microbiota composition is critically influenced by genetic and environmental factors, such as diet, alcohol consumption, or certain drugs. These factors lead to quantitative and qualitative alterations that significantly impact the metabolic activity of the bacterial community and are involved in the pathogenesis of multiple diseases (11–13). In particular, dysbiosis and the alteration of the intestinal barrier are strongly associated with inflammation and metabolic disorders (14). Preclinical studies have shown that microbiome from obese mice has an increased capacity to harvest energy from the diet and also influences how this energy is used and stored (15). In humans, low bacterial richness and distinctive alterations of the metagenome allow to define subsets of individuals with high-risk metabolic profiles (16), and, also, identify specific microbiome signatures for related diseases such as type-2 diabetes (17) or obesity (18).

## The Impact of Gut Microbiome in the Pathogenesis of MAFLD

Consistent with the growing evidence that suggest an strong correlation between the microbiome and the metabolic dysfunction, the role of the microbiota in the pathogenesis of MAFLD has also been explored (19). In this regard, germ-free mice have shown a significant decreased susceptibility to diet induced hepatic steatosis (20). This pathogenic role is further highlighted by the fact that microbiota composition can determine the response to the administration of high-fat diet, leading to hyperglycemia and hepatic steatosis depending on the bacterial host profile. Interestingly, the propensity to develop MAFLD in response to high-fat diet may be transmissible by gut microbiota transplantation (21).

Taxonomic studies have proposed distinctive microbiome signatures based on the flora modifications associated to the different stages of the disease (22). On one hand, some authors have shown that, compared with healthy subjects, the Proteobacteria and Fusobacteria phyla are more abundant in patients with MAFLD (23). Conversely, a lower abundance of Bacteroidetes, and in particular, the genera *Prevotella*, has been found in this group. With regards to steatohepatitis, increased abundance of *Bacteroides* and decreased concentrations of *Prevotella* were independently associated with liver inflammation in a well-characterized population of adult MAFLD (24). These quantitative changes were accompanied by significant shifts in the modulatory functions of the microbiota and the metabolism of carbohydrates, lipids, and amino acids (24).

As MAFLD progresses from liver steatosis and inflammation to fibrosis, other distinctive signatures have been found. Metagenomic sequencing has identified an association between advanced fibrosis and an increased abundance of Escherichia coli and Bacteriodes vulgatus (25). Of note, both E. Coli and B. vulgatus are also increased in other metabolic-syndrome related diseases and, as such, their abundance parallels the increasing body mass index, hemoglobin A1c levels (26), and insulin resistance (27). Other authors have found that the genera Ruminococcus is independently associated with significant fibrosis (24), and, in non-obese MAFLD patients, Ruminococcaceae and Veillonellaceae families were associated with fibrosis severity (28). Interestingly, a recent study proposes an "extra-hepatic" signature that combines the association of muscle fat infiltration (myosteatosis) and a reduction of fecal Clostridium sensu stricto for identifying obese patients at higher risk of liver fibrosis (29).

There is, however, substantial heterogeneity and conflicting results among studies aiming to characterize specific signatures associated to the different clinical phenotypes. These discrepancies are probably related to the underlying complex interplay among the different pathogenic elements and the effect of potential confounders (22). For instance, multiple factors such as demographic features (ethnicity, sex, geographical location, etc.), drug consumption, or the circadian oscillations (30) have been shown to impact gut microbiota in terms of proliferation, composition, function and metabolite production, determining the selection of specific genera independently of the disease stage.

Other authors have proposed disease classification signatures based on the metabolomic analysis of substrates derived from gut flora metabolism. Among these products, 3-(4-hydroxyphenyl) lactate, which is derived from aromatic amino acid metabolism, is associated with advanced fibrosis in patients with MAFLD (31). Similarly, circulating levels of trimethylamine N-oxide, which results from choline metabolism, has been shown to correlate with the severity of MAFLD (32).

Experimental and translational studies also support the contribution of small intestine bacterial overgrowth (SIBO) to hepatic steatosis (33). In this regard, patients with MAFLD present an increased prevalence of SIBO, which correlates with the severity of steatosis (34) and hepatic inflammation (35). Of interest, SIBO is associated with higher serum endotoxin levels and higher expression of toll-like receptor 4, CD14 and NF $\kappa\beta$  (35), a well-known proinflammatory signaling pathway.

The gut microbiota also plays a critical role as modulator of bile acids (BAs) pool size and composition, by deconjugating primary into secondary BAs. The BAs also regulate microbiome diversity by inducing the production of antimicrobial peptides and modulating the expression of genes related with the innate immunity response, intestinal tight junctions, and lipid metabolism (36). The altered interplay among the circulating BAs and the gut microbiota decreases Farnesoid X receptor (FXR) intestinal signaling, which contributes to the development and progression of the metabolic abnormalities present in MAFLD (37). Conversely, experiments with germ-free mice transplanted with protective gut microbiota against MAFLD, led to increased specific secondary BA, the induction of hepatic BAs transporters, and the repression of hepatic lipogenic genes (38).

Most of the studies targeting the microbiota are focused on the bacterial components, however, fungi and viruses are also present in the intestinal flora. Recently, the analysis of fecal viromes from MAFLD patients has identified a decreased viral diversity compared to healthy volunteers (39). The severity of MAFLD (considered as a NAFLD Activity Score 5-8 or liver cirrhosis) was associated with a significant reduction in the proportion of bacteriophages (viruses that specifically infect and kill bacteria). The addition of viral diversity data to clinical variables in the multivariate analysis, significantly improved model accuracy to identify patients at increased risk of severe disease. The potential infection of the gut by the recently described SARS CoV-2, may exacerbate an existing state of increased intestinal permeability and mucosal inflammation, thereby contributing to the systemic immune dysfunction characteristic of severe COVID-19 (40).

In summary, microbiota dysbiosis and decreased diversity are present in patients with MAFLD as in other metabolic syndrome-related diseases. These alterations are linked to the pathogenesis, but the exact mechanisms underlying disease initiation and progression are partially unknown. Aiming to characterize different MAFLD phenotypes, significant efforts are being made to identify specific microbiome signatures. The multifactorial pathogenesis of the disease hinders these approaches, which, however, may contribute to non-invasive diagnosis, severity stratification, and monitoring in these patients.

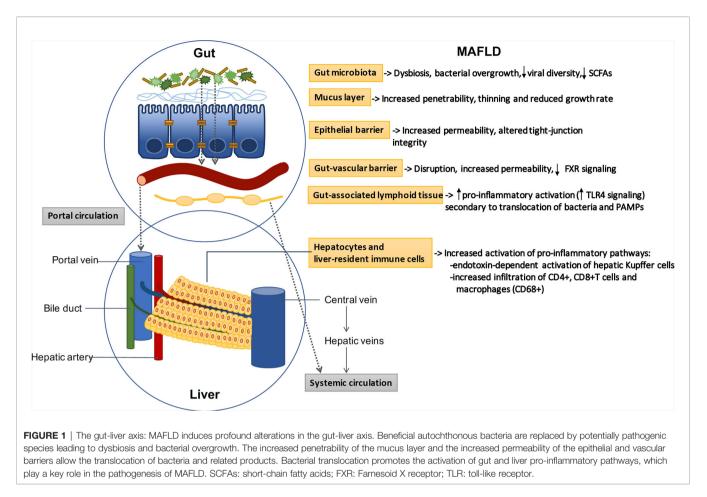
### Intestinal Barrier Integrity Regulates Cross-Talk Between the Gut and the Liver

The intestinal barrier controls the transport from the gut to the liver and systemic circulation, promoting the absorption of nutrients and water while preventing translocation of pathogens and derived products (pathogen associated molecular patterns or PAMPs) (41). The barrier is composed by a mucus layer and an epithelial monolayer of specialized cells connected by intercellular junctions that seal the space and control the paracellular passage. The mucus prevents the direct contact between the bacteria and the epithelial lining, and is mainly composed by large glycosylated proteins (mucins) secreted by intestinal globet cells (42). In addition to the mucus and the epithelial layer, recent evidence has characterized the gut-vascular barrier, which prevents translocation of bacteria and PAMPs directly into the portal circulation (43) (**Figure 1**).

#### Increased Intestinal Permeability in MAFLD

Increased intestinal permeability has been demonstrated in preclinical models and in patients with MAFLD. Specifically, F11r-knockout mice (a gene which encodes the junctional adhesion molecule A), develop more severe steatohepatitis than control mice following a diet high in saturated fat, fructose and cholesterol (44). In humans, Miele L. et al. investigated intestinal permeability in patients with MAFLD by measuring the urinary excretion of <sup>51</sup>Cr-ethylene diamine tetraacetate (<sup>51</sup>Cr-EDTA) and analyzing tight junctions integrity by immunohistochemical expression of zona occludens-1 in duodenal biopsy specimens (34). They found that, compared with the values observed in healthy volunteers, <sup>51</sup>Cr-EDTA excretion was significantly increased, especially in those with moderate/severe steatosis. Further, zonula occludens-1 expression was reduced in intestinal crypts and villi as compared to healthy subjects. Together these data support that disruption of the intercellular tight junctions contributes to increase intestinal permeability and has an important role in the pathogenesis of hepatic steatosis.

Alterations at the mucus layer have been also linked to the pathogenesis of MAFLD. For instance, Mucin-2 deficient mice were protected from high-fat diet induced weight gain, fatty liver, and insulin resistance (45). On the other hand, decreased abundance of *Akkermansia muciniphila*, a mucin-degrading bacteria which represents more than 1% of the microbial community in healthy humans (46), has been associated with



thinning of the mucus layer, and a subsequent increase in gut permeability and inflammation (47). Depletion of this gram negative bacteria has been linked to obesity, insulin resistance, hypertension and liver inflammation (48, 49). In line with this findings, oral supplementation with *A. muciniphila* in overweight/obese insulin-resistant volunteers, improved insulin sensitivity, and reduced insulinemia, total cholesterol, markers of liver dysfunction and inflammation (50). On addition, dietary emulsifiers such as carboxymethylcellulose and polysorbate-80 are widely used as components of processed foods. These additives induce a reduction of mucus thickness, which leads to a higher contact of bacteria with the epithelium, and an increased permeability to dextrans, contributing to low grade inflammation and metabolic syndrome (51).

#### Diet Impacts Intestinal Barrier Integrity by Modulating Gut Microbiota

Microbial fermentation of dietary indigestible fibers produces short-chain fatty acids (SCFAs), mainly, acetate, propionate, and butyrate (52). SCFAs serve as energy substrates to the colonic epithelium, modulate cell functions such as histone deacetylation (53), and contribute to control gut inflammation (54), immune homeostasis (55) and appetite (56). Western-style diet (WSD) is rich in saturated fats and simple carbohydrates but poor in dietary fiber, and thus, lower concentrations of SCFAs are found in mice fed with WSD (57). These mice also showed functional defects in the intestinal barrier, specifically, an increased mucus penetrability and a reduced growth rate of the inner mucus layer. Both defects were prevented by fecal microbiota transplantation from chow-fed donors. Of note, administration of Bifidobacterium longum restored mucus growth, but not the increased penetrability, which was instead prevented by fiber supplementation (57). Likewise, microbiota transplantation and supplementation with Quercetin (a flavonoid with antioxidant and anti-inflammatory properties) restored SCFAs production in mice fed with high-fat diet (58, 59). Quercetin and other prebiotic compounds, such as polyphenol-rich extracts or some dietary fibers as inulin, may also counteract lipid metabolism dysregulation, and liver and systemic inflammation in preclinical models (60-62). In addition, probiotics, such as wild rice, which is a genus of grasses (Zizania), have shown to reduce body weight, liver steatosis and inflammation in high-fat diet-fed mice (63).

The above preclinical evidences are in line with clinical studies in patients with type 2 diabetes mellitus who received high-fiber diet vs standard of care. Increased availability of nondigestible but fermentable carbohydrates was sufficient to induce a clinically relevant improvement in hemoglobin A1c levels. Fiber supplementation also induced selective promotion of SCFA-producing bacterial strains. This coincided with a significant increase in postprandial glucagon-like peptide-1 and higher levels of fasting peptide YY, which favors insulin secretion and glucose regulation (64). Vitamin D supplementation, that has shown potential anti-fibrotic, anti-inflammatory, and insulin-sensitizing properties, is under investigation following conflicting results about its beneficial effects in MAFLD (65).

All these evidences indicate that the alterations in the intestinal barrier are potentially reversible by modifying the composition of the microbiota with fecal transplantation or dietary approaches.

## Emerging Data on Gut Vascular Barrier Contribution to MAFLD

The role of the gut vascular barrier in the pathogenesis of MAFLD has also been explored. Data from patients with celiac disease and altered serum transaminases suggest that the disruption of the endothelial barrier may also contribute to liver inflammation and damage (43). Recently, studies have found that high-fat diet modifies the microbiota leading to an early disruption of the epithelial barrier and an increased vascular permeability. The disruption of the endothelial barrier induced by dysbiosis allows bacterial translocation to the liver, and has been found to be an early and required event in MAFLD development (66). Mechanistically, the alteration of the vascular barrier involved the WNT/b-catenin signaling pathway, and, consistent with these findings, obeticholic acid, a FXR receptor agonist that drives b-catenin activation in endothelial cells, protected against barrier disruption and limited steatohepatitis development in mice fed with a high-fat diet. This suggests that protection against vascular barrier disruption could be an additional mechanism by which obeticholic acid may improve outcomes in MALFD.

# The Role of the Immune System in the Pathogenesis of MAFLD

The immune system plays a critical role in the homeostasis and regulation of the gut-liver axis. The accurate balance between immune activation and tolerance is controlled by a network of innate and adaptive immune cells present not only in the liver, but also in the intestine and the adipose tissue (67). The first defense against intestinal pathogens is the gut-associated lymphoid tissue (GALT), which is composed by intraepithelial lymphocytes, Peyer's patches, the mesenteric lymph nodes and the immune effector cells within the lamina propria (68). The GALT also extends to the large intestine which contains cryptopatches and isolated lymphoid follicles. On the other hand, the liver displays a number of specialized innate and adaptive immune cells responsible for detecting and clearing pathogens that reach the sinusoids. Liver-resident immune cells include antigen presenting cells (APCs) (Kupffer, dendritic and liver sinusoidal endothelial cells), T cells, B cells, natural killer cells and monocytes (69). In addition, the hepatocytes produce essential immune components, and may also act as APCs by expressing MHC-I and II and costimulatory molecules (70).

Despite the continuous challenge, in basal conditions, the gut and the liver immune system act coordinately screening and classifying gut-derived antigens as harmless dietary elements or bacterial pathogens, balancing tolerance and immune activation. However, specific pro-inflammatory pathways are triggered in patients with MAFLD (71). In particular, TLR4 signaling has been identified as a key driver of inflammation and steatohepatitis in diverse experimental animal models of MAFLD (72, 73). On the other hand, NLRP3 inflammasome deficiency attenuates dysbiosis, systemic inflammation and the total cholesterol increase induced in mice exposed to high fat diet (74). Fructoseinduced MAFLD is also associated with intestinal bacterial overgrowth and increased intestinal permeability, subsequently leading to an endotoxin-dependent activation of hepatic Kupffer cells (75). Additionally, increased visceral adipose tissue is infiltrated by pro-inflammatory macrophages that induce the release of chemokines and cytokines involved in liver inflammation and hepatic insulin resistance (76).

Immune mechanisms have also been involved in the progression from steatosis to fibrosis. Emerging data highlight the important role of CD4+memory T cells, specifically IL-17A-secreting Th17 and IFN $\gamma$ -secreting Th1 subsets. Humanized mouse models of dietinduced MAFLD showed a remarkable liver infiltration of CD4+, CD8+T cells and macrophages (CD68+), which were largely confined to fibrotic regions. Conversely, depletion of CD4+T cells abrogated diet-induced inflammatory response and liver fibrosis development (77). Further studies have shown that blocking integrin receptor  $\alpha$ 4 $\beta$ 7-mediated recruitment of CD4+ T cells to the intestine and liver, not only attenuates hepatic inflammation and fibrosis, but also improves metabolic alterations associated with MAFLD (78).

# Therapeutic Interventions Targeting the Gut-Liver-Axis in Patients With MAFLD

Taken together, all the above evidences show that disruption of the gut-liver-axis contributes to the pathogenesis of MAFLD, and thus, there is a great interest in the modulation of their components for therapeutic purposes. These strategies have shown promising results in preclinical studies, however, evidence in human clinical trials is limited so far. Below we have selected emerging therapeutic approaches targeting the gutliver axis on MAFLD, whose results highlight the complex pathophysiology underlying the disease (**Table 1**).

Fecal microbiota transplantation (FMT) can potentially restore microbial diversity and function. Therefore, Craven L. and colleagues hypothesized that fecal transplantation from healthy lean donors to MAFLD patients could have metabolic benefits by restoring the integrity of the intestinal barrier. Allogenic FMT failed to decrease insulin resistance or the percentage of hepatic fat assessed by magnetic resonance, however, it did improve intestinal permeability evaluated by the lactulose/mannitol ratio urine test (79). These results are in contrast with the findings of Vrieze et al. that found that infusion of microbiota from lean donors, increased insulin sensitivity in recipients with metabolic syndrome (80). Another interesting study has demonstrated that FMT from metabolically compromised obese donors temporarily worsens insulin sensitivity in recipients with metabolic syndrome, whereas a non-significant increase in insulin sensitivity was observed in

#### TABLE 1 | Selected therapeutic interventions targeting the gut-liver axis.

Study	Identification	Main results
Safety and efficacy of hydrothermal duodenal mucosal resurfacing in patients with type 2 diabetes: the randomized, double-blind, sham-controlled, multicenter REVITA-2 feasibility trial	NCT02879383	DMR is safe and exerts beneficial disease-modifying metabolic effects in T2D with or without non-alcoholic liver disease (Phase 2)
Effect of duodenal mucosal resurfacing in the treatment of NASH (DMR_NASH_001)	NCT03536650	Completed, no results posted yet
Cenicriviroc treatment for adults with nonalcoholic steatohepatitis and fibrosis: final analysis of the phase 2b CENTAUR study	NCT02217475	A similar proportion on CVC or placebo achieved ≥1-stage fibrosis improvement and no worsening of NASH (Phase 2b)
Cenicriviroc for the treatment of liver fibrosis in adults with nonalcoholic steatohepatitis: AURORA phase 3 study design	NCT03028740	Recruiting (Phase 3)
The fatty acid-bile acid conjugate aramchol reduces liver fat content in patients with nonalcoholic fatty liver disease	NCT01094158	Aramchol is safe, tolerable, and significantly reduces liver fat content in patients with NAFLD (Phase 2)
A Phase 2b, randomized, double-blind, placebo-controlled study evaluating the safety and efficacy of efruxifermin in non-cirrhotic subjects with nonalcoholic steatohepatitis	NCT04767529	Recruiting. Preliminary results indicate a high percentage of fibrosis resolution (Phase 2)
Resmetirom (MGL-3196) for the treatment of non-alcoholic steatohepatitis: a multicenter, randomized, double-blind, placebo-controlled, phase 2 trial	NCT02912260	Significant reduction in hepatic fat in patients with NASH (Phase 2)
Investigation of efficacy and safety of three dose levels of subcutaneous semaglutide once daily versus placebo in subjects with non-alcoholic steatohepatitis.	NCT02970942	Semaglutide resulted in a significantly higher percentage of patients with NASH resolution than placebo. It did not show a significant difference in fibrosis stage (Phase 2)
Researching an effect of GLP-1 agonist on liver steatosis (REALIST)	NCT03648554	Completed, no results posted yet (Phase 4)
Randomized global phase 3 study to evaluate the impact on NASH with fibrosis of obeticholic acid treatment (REGENERATE)	NCT02548351	Interim analysis showed that obeticholic acid 25 mg significant improved fibrosis and key components of NASH (Phase 3)
Liraglutide safety and efficacy in patients with non-alcoholic steatohepatitis (LEAN): a multicenter, double-blind, randomized, placebo-controlled phase 2 study	NCT01237119	Liraglutide was safe, well tolerated, and led to histological resolution of NASH (Phase 2)
Safety and tolerability of yaq-001 in patients with non-alcoholic steatohepatitis	NCT03962608	Not yet recruiting
The effect of consecutive fecal microbiota transplantation on non-alcoholic fatty liver disease	NCT04465032	Recruiting (Phase 4)
The effect of probiotics on the clinical outcomes and gut microenvironment in patients with fatty liver	NCT04074889	Recruiting
An adaptive design study for the assessment of the safety, tolerability, and pharmacokinetics of RYI-018 (Anti-CB1 monoclonal antibody) after repeat dosing in subjects with non-alcoholic fatty liver disease	NCT03261739	Completed, no results posted yet (Phase 1)
Polyunsaturated fatty acids in patients with NAFLD An investigator initiated prospective, four arms randomized comparative study of efficacy and safety of saroglitazar, vitamin e and life style modification in patients with nonalcoholic fatty liver disease/non-alcoholic steatohepatitis	NCT02647294 NCT04193982	
A double-blind, randomized, placebo-controlled, phase 2 study to explore the efficacy and safety of elobixibat in adults with nonalcoholic fatty liver disease or nonalcoholic steatohepatitis	NCT04006145	Completed, no results posted yet (Phase 2)
Effect of silymarin in patients with non-alcoholic fatty liver disease Study to assess the efficacy, safety and tolerability of LCQ908 (Pradigastat) in NAFLD patients	NCT03749070 NCT01811472	Recruiting Completed. Preliminary results suggest that LCQ908 was effective, safe and well tolerated (Phase 2)

DMR: duodenal mucosal resurfacing; T2D: type 2 diabetes; NASH: non-alcoholic steatohepatitis; CVC: cenicriviroc; NAFLD: non-alcoholic fatty liver disease; GLP-1: Glucagon-like peptide 1.

recipients of FMT from healthy postgastric bypass donors (81). These results suggest that the response to FMT is modulated by differences in fecal microbiota composition and could explain in part the heterogeneity of the results obtained so far.

Non-absorbable carbons of controlled porosity (Yaq-001) have shown to selectively modulate the composition and function of stool microbiome (82), and thus, safety and tolerability of Yaq-001 is currently under investigation (NCT03962608). Other ongoing study aims to assess the feasibility and safety of endoscopic duodenal mucosal resurfacing in this population (NCT03536650), which can potentially mimic the beneficial metabolic effects on glucose metabolism achieved by surgical upper-intestinal bypass (83). Other strategies aiming to improve insulin resistance and metabolic dysfunction by Glucagon-like peptide-1 analogues have shown positive results in terms of steatohepatitis resolution (84, 85), however, no significant differences in fibrosis regression have yet been achieved. In this regard, an interim analysis from a multicenter, randomized, placebo-controlled trial with obeticholic acid has demonstrated to be the only strategy that significantly improves fibrosis as well as other key components of MAFLD, and warrants further evaluation (86).

### CONCLUSIONS

MAFLD is the most prevalent liver disease worldwide. The disruption of the gut-liver axis drives its pathogenesis and progression. Dietary factors may induce profound qualitative and quantitative changes in the gut microbiota, which subsequently impairs the integrity of the epithelial and vascular barriers. The increased translocation of bacteria and PAMPs induce a persistent immune activation that promotes gut and liver inflammation. A better understanding of the different mechanisms involved in the pathogenesis of MAFLD will contribute to develop better diagnostic and therapeutic approaches.

### **AUTHOR CONTRIBUTIONS**

RM drafted the manuscript. RM and AA: critical revision for important intellectual content. All authors contributed to the article and approved the submitted version.

### REFERENCES

- Younossi ZM, Koenig AB, Abdelatif D, Fazel Y, Henry L, Wymer M. Global epidemiology of nonalcoholic fatty liver disease—Meta-analytic assessment of prevalence, incidence, and outcomes. *Hepatology* (2016) 64:73–84. doi: 10.1002/hep.28431
- Estes C, Razavi H, Loomba R, Younossi Z, Sanyal AJ. Modeling the epidemic of nonalcoholic fatty liver disease demonstrates an exponential increase in burden of disease. *Hepatology* (2018) 67:123–33. doi: 10.1002/hep.29466
- Angulo P, Kleiner DE, Dam-Larsen S, Adams LA, Bjornsson ES, Charatcharoenwitthaya P, et al. Liver fibrosis, but no other histologic features, is associated with long-term outcomes of patients with nonalcoholic fatty liver disease. *Gastroenterology* (2015) 149:389–97. doi: 10.1053/j.gastro.2015.04.043
- Eslam M, Newsome PN, Sarin SK, Anstee QM, Targher G, Romero-Gomez M, et al. A new definition for metabolic dysfunction-associated fatty liver disease: An international expert consensus statement. J Hepatol (2020) 73:202–9. doi: 10.1016/j.jhep.2020.03.039
- Albillos A, de Gottardi A, Rescigno M. The gut-liver axis in liver disease: Pathophysiological basis for therapy. J Hepatol (2020) 72:558–77. doi: 10.1016/j.jhep.2019.10.003
- Leung C, Rivera L, Furness JB, Angus PW. The role of the gut microbiota in NAFLD. Nat Rev Gastroenterol Hepatol (2016) 13:412–25. doi: 10.1038/ nrgastro.2016.85
- Parthasarathy G, Revelo X, Malhi H. Pathogenesis of nonalcoholic steatohepatitis: an overview. *Hepatol Commun* (2020) 4:478–92. doi: 10.1002/hep4.1479
- Wiest R, Albillos A, Trauner M, Bajaj JS, Jalan R. Targeting the gut-liver axis in liver disease. J Hepatol (2017) 67:1084–103. doi: 10.1016/j.jhep.2017.05.007
- Albillos A, Lario M, Álvarez-Mon M. Cirrhosis-associated immune dysfunction: distinctive features and clinical relevance. J Hepatol (2014) 61:1385–96. doi: 10.1016/j.jhep.2014.08.010
- 10. Guarner F, Malagelada J-R. Gut flora in health and disease. Lancet (2003) 361:512-9. doi: 10.1016/S0140-6736(03)12489-0
- Konjevod M, Nikolac Perkovic M, Sáiz J, Svob Strac D, Barbas C, Rojo D. Metabolomics analysis of microbiota-gut-brain axis in neurodegenerative and psychiatric diseases. J Pharm BioMed Anal (2020) 194:113681. doi: 10.1016/ j.jpba.2020.113681
- Docimo G, Cangiano A, Romano RM, Pignatelli MF, Offi C, Paglionico VA, et al. The human microbiota in endocrinology: implications for pathophysiology, treatment, and prognosis in thyroid diseases. *Front Endocrinol (Lausanne)* (2020) 11:586529. doi: 10.3389/fendo.2020.586529
- Cheng Y, Ling Z, Li L. The intestinal microbiota and colorectal cancer. Front Immunol (2020) 11:615056. doi: 10.3389/fimmu.2020.615056
- Tilg H, Zmora N, Adolph TE, Elinav E. The intestinal microbiota fuelling metabolic inflammation. *Nat Rev Immunol* (2020) 20:40–54. doi: 10.1038/ s41577-019-0198-4
- Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated gut microbiome with increased capacity for energy harvest. *Nature* (2006) 444:1027–31. doi: 10.1038/nature05414
- Le Chatelier E, Nielsen T, Qin J, Prifti E, Hildebrand F, Falony G, et al. Richness of human gut microbiome correlates with metabolic markers. *Nature* (2013) 500:541–6. doi: 10.1038/nature12506
- Qin J, Li Y, Cai Z, Li S, Zhu J, Zhang F, et al. A metagenome-wide association study of gut microbiota in type 2 diabetes. *Nature* (2012) 490:55–60. doi: 10.1038/nature11450

#### FUNDING

This work was financed by grants from the Spanish Ministerio de Ciencia e Innovación (SAF 2017-86343-R, and Instituto de Salud Carlos III PI20/01302 awarded to A.A). CIBEREHD is funded by the Instituto de Salud Carlos III. Grants cofinanced by the European Development Regional Fund "A way to achieve Europe" (EDRF).

- Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. *Nature* (2009) 457:480– 4. doi: 10.1038/nature07540
- Sharpton SR, Schnabl B, Knight R, Loomba R. Current concepts, opportunities, and challenges of gut microbiome-based personalized medicine in nonalcoholic fatty liver disease. *Cell Metab* (2021) 33:21–32. doi: 10.1016/j.cmet.2020.11.010
- Bäckhed F, Ding H, Wang T, Hooper LV, Gou YK, Nagy A, et al. The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* (2004) 101:15718–23. doi: 10.1073/pnas.0407076101
- Le Roy T, Llopis M, Lepage P, Bruneau A, Rabot S, Bevilacqua C, et al. Intestinal microbiota determines development of non-alcoholic fatty liver disease in mice. *Gut* (2013) 62:1787–94. doi: 10.1136/gutjnl-2012-303816
- Aron-Wisnewsky J, Vigliotti C, Witjes J, Le P, Holleboom AG, Verheij J, et al. Gut microbiota and human NAFLD: disentangling microbial signatures from metabolic disorders. *Nat Rev Gastroenterol Hepatol* (2020) 17:279–97. doi: 10.1038/s41575-020-0269-9
- Shen F, Zheng R-D, Sun X-Q, Ding W-J, Wang X-Y, Fan J-G. Gut microbiota dysbiosis in patients with non-alcoholic fatty liver disease. *Hepatobiliary Pancreat Dis Int* (2017) 16:375–81. doi: 10.1016/S1499-3872(17)60019-5
- Boursier J, Mueller O, Barret M, Machado M, Fizanne L, Araujo-Perez F, et al. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. *Hepatology* (2016) 63:764–75. doi: 10.1002/hep.28356
- Loomba R, Seguritan V, Li W, Long T, Klitgord N, Bhatt A, et al. Gut microbiome-based metagenomic signature for non-invasive detection of advanced fibrosis in human nonalcoholic fatty liver disease. *Cell Metab* (2017) 25:1054–62. doi: 10.1016/j.cmet.2017.04.001
- Aron-Wisnewsky J, Prifti E, Belda E, Ichou F, Kayser BD, Dao MC, et al. Major microbiota dysbiosis in severe obesity: fate after bariatric surgery. *Gut* (2019) 68:70–82. doi: 10.1136/gutjnl-2018-316103
- Pedersen HK, Gudmundsdottir V, Nielsen HB, Hyotylainen T, Nielsen T, Jensen BAH, et al. Human gut microbes impact host serum metabolome and insulin sensitivity. *Nature* (2016) 535:376–81. doi: 10.1038/nature18646
- Lee G, You HJ, Bajaj JS, Joo SK, Yu J, Park S, et al. Distinct signatures of gut microbiome and metabolites associated with significant fibrosis in non-obese NAFLD. *Nat Commun* (2020) 11:4982. doi: 10.1038/s41467-020-18754-5
- Lanthier N, Rodriguez J, Nachit M, Hiel S, Trefois P, Neyrinck AM, et al. Microbiota analysis and transient elastography reveal new extra-hepatic components of liver steatosis and fibrosis in obese patients. *Sci Rep* (2021) 11:1–14. doi: 10.1038/s41598-020-79718-9
- Thaiss CA, Levy M, Korem T, Dohnalová L, Shapiro H, Jaitin DA, et al. Microbiota diurnal rhythmicity programs host transcriptome oscillations. *Cell* (2016) 167:1495–510. doi: 10.1016/j.cell.2016.11.003
- Caussy C, Hsu C, Lo M-T, Liu A, Bettencourt R, Ajmera VH, et al. Link between gut-microbiome derived metabolite and shared gene-effects with hepatic steatosis and fibrosis in NAFLD. *Hepatology* (2018) 68:918–32. doi: 10.1002/hep.29892
- 32. Chen Y, Liu Y, Zhou R, Chen X, Wang C, Tan X, et al. Associations of gutflora-dependent metabolite trimethylamine-N-oxide, betaine and choline with non-alcoholic fatty liver disease in adults. *Sci Rep* (2016) 6:19076. doi: 10.1038/srep19076
- 33. Wijarnpreecha K, Lou S, Watthanasuntorn K, Kroner PT, Cheungpasitporn W, Lukens FJ, et al. Small intestinal bacterial overgrowth and nonalcoholic fatty liver disease: a systematic review and meta-analysis. *Eur J Gastroenterol Hepatol* (2020) 32:601–8. doi: 10.1097/MEG.000000000001541

- 34. Miele L, Valenza V, La Torre G, Montalto M, Cammarota G, Ricci R, et al. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. *Hepatology* (2009) 49:1877–87. doi: 10.1002/ hep.22848
- 35. Kapil S, Duseja A, Sharma BK, Singla B, Chakraborti A, Das A, et al. Small intestinal bacterial overgrowth and toll-like receptor signaling in patients with non-alcoholic fatty liver disease. J Gastroenterol Hepatol (2016) 31:213–21. doi: 10.1111/jgh.13058
- Wang W, Zhao J, Gui W, Sun D, Dai H, Xiao L, et al. Tauroursodeoxycholic acid inhibits intestinal inflammation and barrier disruption in mice with non-alcoholic fatty liver disease. *Br J Pharmacol* (2018) 175:469–84. doi: 10.1111/bph.14095
- Chen J, Vitetta L. Gut microbiota metabolites in nafld pathogenesis and therapeutic implications. *Int J Mol Sci* (2020) 21:1–19. doi: 10.3390/ijms21155214
- Petrov PD, García-Mediavilla MV, Guzmán C, Porras D, Nistal E, Martínez-Flórez S, et al. A network involving gut microbiota, circulating bile acids, and hepatic metabolism genes that protects against non-alcoholic fatty liver disease. *Mol Nutr Food Res* (2019) 63:1900487. doi: 10.1002/mnfr.201900487
- Lang S, Demir M, Martin A, Jiang L, Zhang X, Duan Y, et al. Intestinal virome signature associated with severity of nonalcoholic fatty liver disease. *Gastroenterology* (2020) 159:1839–52. doi: 10.1053/j.gastro.2020.07.005
- Assante G, Williams R, Youngson NA. Is the increased risk for MAFLD patients to develop severe COVID-19 linked to perturbation of the gut-liver axis? J Hepatol (2021) 74:487–8. doi: 10.1016/j.jhep.2020.05.051
- Arab JP, Martin-Mateos RM, Shah VH. Gut–liver axis, cirrhosis and portal hypertension: the chicken and the egg. *Hepatol Int* (2018) 12:24–33. doi: 10.1007/s12072-017-9798-x
- Johansson MEV, Sjövall H, Hansson GC. The gastrointestinal mucus system in health and disease. Nat Rev Gastroenterol Hepatol (2013) 10:352–61. doi: 10.1038/nrgastro.2013.35
- Spadoni I, Zagato E, Bertocchi A, Paolinelli R, Hot E, Di Sabatino A, et al. A gut-vascular barrier controls the systemic dissemination of bacteria. *Sci (80-)* (2015) 350:830–4. doi: 10.1126/science.aad0135
- 44. Rahman K, Desai C, Iyer SS, Thorn NE, Kumar P, Liu Y, et al. Loss of junctional adhesion molecule a promotes severe steatohepatitis in mice on a diet high in saturated fat, fructose, and cholesterol. *Gastroenterology* (2016) 151:733–46. doi: 10.1053/j.gastro.2016.06.022
- Hartmann P, Seebauer CT, Mazagova M, Horvath A, Wang L, Llorente C, et al. Deficiency of intestinal mucin-2 protects mice from diet-induced fatty liver disease and obesity. *Am J Physiol - Gastrointest Liver Physiol* (2016) 310:310–22. doi: 10.1152/ajpgi.00094.2015
- Derrien M, Collado MC, Ben-Amor K, Salminen S, de Vos WM. The mucin degrader akkermansia muciniphila is an abundant resident of the human intestinal tract. *Appl Environ Microbiol* (2008) 74:1646–8. doi: 10.1128/AEM.01226-07
- 47. Derrien M, Van Baarlen P, Hooiveld G, Norin E, Müller M, de Vos WM. Modulation of mucosal immune response, tolerance, and proliferation in mice colonized by the mucin-degrader Akkermansia muciniphila. *Front Microbiol* (2011) 2:166 doi: 10.3389/fmicb.2011.00166
- Everard A, Belzer C, Geurts L, Ouwerkerk JP, Druart C, Bindels LB, et al. Cross-talk between Akkermansia muciniphila and intestinal epithelium controls diet-induced obesity. *Proc Natl Acad Sci* (2013) 110:9066–71. doi: 10.1073/pnas.1219451110
- Plovier H, Everard A, Druart C, Depommier C, Van Hul M, Geurts L, et al. A purified membrane protein from Akkermansia muciniphila or the pasteurized bacterium improves metabolism in obese and diabetic mice. *Nat Med* (2017) 23:107–13. doi: 10.1038/nm.4236
- Depommier C, Everard A, Druart C, Plovier H, Van Hul M, Vieira-Silva S, et al. Supplementation with Akkermansia muciniphila in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med* (2019) 25:1096–103. doi: 10.1038/s41591-019-0495-2
- Chassaing B, Koren O, Goodrich JK, Poole AC, Srinivasan S, Ley RE, et al. Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. *Nature* (2015) 519:92–6. doi: 10.1038/nature14232
- Koh A, De Vadder F, Kovatcheva-Datchary P, Bäckhed F. From dietary fiber to host physiology: Short-chain fatty acids as key bacterial metabolites. *Cell* (2016) 165:1332–45. doi: 10.1016/j.cell.2016.05.041
- Donohoe DR, Collins LB, Wali A, Bigler R, Sun W, Bultman SJ. The Warburg effect dictates the mechanism of butyrate-mediated histone acetylation and cell proliferation. *Mol Cell* (2012) 48:612–26. doi: 10.1016/j.molcel.2012.08.033

- Macia L, Tan J, Vieira AT, Leach K, Stanley D, Luong S, et al. Metabolitesensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nat Commun* (2015) 6:6734. doi: 10.1038/ncomms7734
- Belkaid Y, Harrison OJ. Homeostatic immunity and the microbiota. *Immunity* (2017) 46:562–76. doi: 10.1016/j.immuni.2017.04.008
- Frost G, Sleeth ML, Sahuri-Arisoylu M, Lizarbe B, Cerdan S, Brody L, et al. The short-chain fatty acid acetate reduces appetite *via a* central homeostatic mechanism. *Nat Commun* (2014) 5:3611. doi: 10.1038/ncomms4611
- Schroeder BO, Birchenough GMH, Ståhlman M, Arike L, Johansson MEV, Hansson GC, et al. Bifidobacteria or fiber protects against diet-induced microbiota-mediated colonic mucus deterioration. *Cell Host Microbe* (2018) 23:27–40. doi: 10.1016/j.chom.2017.11.004
- 58. Porras D, Nistal E, Martínez-Flórez S, Olcoz JL, Jover R, Jorquera F, et al. Functional interactions between gut microbiota transplantation, quercetin, and high-fat diet determine non-alcoholic fatty liver disease development in germ-free mice. *Mol Nutr Food Res* (2019) 63:e1800930. doi: 10.1002/mnfr.201800930
- Porras D, Nistal E, Martínez-Flórez S, Pisonero-Vaquero S, Olcoz JL, Jover R, et al. Protective effect of quercetin on high-fat diet-induced non-alcoholic fatty liver disease in mice is mediated by modulating intestinal microbiota imbalance and related gut-liver axis activation. *Free Radic Biol Med* (2017) 102:188–202. doi: 10.1016/j.freeradbiomed.2016.11.037
- 60. Yang Z, Liu G, Wang Y, Yin J, Wang J, Xia B, et al. Fucoidan A2 from the brown seaweed Ascophyllum nodosum lowers lipid by improving reverse cholesterol transport in C57BL/6J mice fed a high-fat diet. J Agric Food Chem (2019) 67:5782–91. doi: 10.1021/acs.jafc.9b01321
- Wu S, Hu R, Nakano H, Chen K, Liu M, He X, et al. Modulation of gut microbiota by Lonicera caerulea L. Berry polyphenols in a mouse model of fatty liver induced by high fat diet. *Molecules* (2018) 23:3213. doi: 10.3390/molecules23123213
- 62. Bao T, He F, Zhang X, Zhu L, Wang Z, Lu H, et al. Inulin exerts beneficial effects on non-alcoholic fatty liver disease *via* modulating gut microbiome and suppressing the Lipopolysaccharide-toll-like receptor 4-M\u03c8-nuclear factorkB-nod-like receptor protein 3 pathway *via* gut-Liver axis in mice. *Front Pharmacol* (2020) 11:558525. doi:. 10.3389/fphar.2020.558525
- Hou XD, Yan N, Du YM, Liang H, Zhang ZF, Yuan XL. Consumption of wild rice (zizania latifolia) prevents metabolic associated fatty liver disease through the modulation of the gut microbiota in mice model. *Int J Mol Sci* (2020) 21:1– 15. doi: 10.3390/ijms21155375
- Zhao L, Zhang F, Ding X, Wu G, Lam YY, Wang X, et al. Gut bacteria selectively promoted by dietary fibers alleviate type 2 diabetes. *Sci (80-)* (2018) 359:1151–6. doi: 10.1126/science.aao5774
- Barchetta I, Cimini FA , Cavallo MG. Vitamin D and metabolic dysfunctionassociated fatty liver disease (MAFLD): An update. *Nutrients* (2020) 12:3302. doi: 10.3390/nu12113302
- 66. Mouries J, Brescia P, Silvestri A, Spadoni I, Sorribas M, Wiest R, et al. Microbiota-driven gut vascular barrier disruption is a prerequisite for nonalcoholic steatohepatitis development. J Hepatol (2019) 71:1216–28. doi: 10.1016/j.jhep.2019.08.005
- Jenne CN, Kubes P. Immune surveillance by the liver. Nat Immunol (2013) 14:996–1006. doi: 10.1038/ni.2691
- Mowat AM. Anatomical basis of tolerance and immunity to intestinal antigens. Nat Rev Immunol (2003) 3:331–41. doi: 10.1038/nri1057
- Zhao J, Zhao S, Liu Y, He X, Qu M, Xu G, et al. Single-cell RNA sequencing reveals the heterogeneity of liver-resident immune cells in human. *Cell Discovery* (2020) 6:22. doi: 10.1038/s41421-020-0157-z
- Franco A, Barnaba V, Natali P, Balsano C, Musca A, Balsano F. Expression of class I and class II major histocompatibility complex antigens on human hepatocytes. *Hepatology* (1988) 8:449–54. doi: 10.1002/hep.1840080302
- Schuster S, Cabrera D, Arrese M, Feldstein AE. Triggering and resolution of inflammation in NASH. Nat Rev Gastroenterol Hepatol (2018) 15:349–64. doi: 10.1038/s41575-018-0009-6
- 72. Liu J, Zhuang Z, Bian D, Ma X, Xun Y, Yang W, et al. Toll-like receptor-4 signalling in the progression of non-alcoholic fatty liver disease induced by high-fat and high-fructose diet in mice. *Clin Exp Pharmacol Physiol* (2014) 41:482–8. doi: 10.1111/1440-1681.12241
- 73. Longo L, Tonin Ferrari J, Rampelotto PH, Hirata Dellavia G, Pasqualotto A, P Oliveira C, et al. Gut dysbiosis and increased intestinal permeability drive microRNAs, NLRP-3 inflammasome and liver fibrosis in a nutritional model

of non-alcoholic steatohepatitis in adult male sprague dawley rats. *Clin Exp Gastroenterol* (2020) 13:351-68. doi: 10.2147/CEG.S262879

- 74. Sokolova M, Yang K, Hansen SH, Louwe MC, Kummen M, Hov JER, et al. NLRP3 inflammasome deficiency attenuates metabolic disturbances involving alterations in the gut microbial profile in mice exposed to high fat diet. *Sci Rep* (2020) 10:1–16. doi: 10.1038/s41598-020-76497-1
- Spruss A, Kanuri G, Wagnerberger S, Haub S, Bischoff SC, Bergheim I. Tolllike receptor 4 is involved in the development of fructose-induced hepatic steatosis in mice. *Hepatology* (2009) 50:1094–104. doi: 10.1002/hep.23122
- Stanton MC, Chen S-C, Jackson JV, Rojas-Triana A, Kinsley D, Cui L, et al. Inflammatory signals shift from adipose to liver during high fat feeding and influence the development of steatohepatitis in mice. *J Inflammation* (2011) 8:8. doi: 10.1186/1476-9255-8-8
- Her Z, Tan JHL, Lim Y-S, Tan SY, Chan XY, Tan WWS, et al. CD4+ T cells mediate the development of liver fibrosis in high fat diet-induced NAFLD in humanized mice. *Front Immunol* (2020) 11:580968. doi: 10.3389/fimmu.2020.580968
- Rai RP, Liu Y, Iyer SS, Liu S, Gupta B, Desai C, et al. Blocking integrin α4β7mediated CD4 T cell recruitment to the intestine and liver protects mice from western diet-induced non-alcoholic steatohepatitis. *J Hepatol* (2020) 73:1013– 22. doi: 10.1016/j.jhep.2020.05.047
- 79. Craven L, Rahman A, Nair Parvathy S, Beaton M, Silverman J, Qumosani K, et al. Allogenic fecal microbiota transplantation in patients with nonalcoholic fatty liver disease improves abnormal small intestinal permeability. *Am J Gastroenterol* (2020) 115:1055–65. doi: 10.14309/ajg.00000000000661
- Vrieze A, Van Nood E, Holleman F, Salojärvi J, Kootte RS, Bartelsman JFWM, et al. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* (2012) 143:913–6. doi: 10.1053/j.gastro.2012.06.031
- 81. de Groot P, Scheithauer T, Bakker guido J, Prodan A, Levin E, Khan MT, et al. Donor metabolic characteristics drive effects of faecal microbiota transplantation on recipient insulin sensitivity, energy expenditure and intestinal transit time. *Gut* (2020) 69:502–12. doi: 10.1136/gutjnl-2019-318320
- Macnaughtan J, Ranchal I, Soeda J, Sawhney R, Oben J, Davies N, et al. Oral therapy with non-absorbable carbons of controlled porosity (YAQ-001)

selectively modulates stool microbiome and its function and this is associated with restoration of immune function and inflammasome activation. *J Hepatol* (2015) 62:S240. doi: 10.1016/S0168-8278(15)30110-0

- 83. Zervos EE, Agle SC, Warren AJ, Lang CG, Fitzgerald TL, Dar M, et al. Amelioration of insulin requirement in patients undergoing duodenal bypass for reasons other than obesity implicates foregut factors in the pathophysiology of type II diabetes. J Am Coll Surg (2010) 210:564–72. doi: 10.1016/j.jamcollsurg.2009.12.025
- Newsome PN, Buchholtz K, Cusi K, Linder M, Okanoue T, Ratziu V, et al. NN9931-4296 Investigators. A placebo-controlled trial of subcutaneous semaglutide in nonalcoholic steatohepatitis. N Engl J Med (2021) 384:1113– 24. doi: 10.1056/NEJMoa2028395
- 85. Armstrong MJ, Gaunt P, Aithal GP, Barton D, Hull D, Parker R, et al. Liraglutide safety and efficacy in patients with non-alcoholic steatohepatitis (LEAN): a multicentre, double-blind, randomised, placebo-controlled phase 2 study. *Lancet* (2016) 387:679–90. doi: 10.1016/S0140-6736(15)00803-X
- Younossi ZM, Ratziu V, Loomba R, Rinella M, Anstee QM, Goodman Z, et al. Obeticholic acid for the treatment of non-alcoholic steatohepatitis: interim analysis from a multicentre, randomised, placebo-controlled phase 3 trial. *Lancet* (2019) 394:2184–96. doi: 10.1016/S0140-6736(19)33041-7

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The reviewer JG-G declared a shared affiliation with the authors to the handling editor at the time of review.

Copyright © 2021 Martín-Mateos and Albillos. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.