



platelet P-selectin and tumor cell sialylated mucins that has been shown to be essential for metastatic spread in mouse models (17). Finally, the findings of Boucharaba et al. lay the groundwork to suggest that inhibition of platelet-derived LPA action on its cognate receptors expressed by tumor cells may be another promising therapeutic target, especially for bone metastasis. The development and clinical testing of this class of specific modulators of platelet function will be necessary before a verdict can be reached regarding the importance of platelets in the progression of disease in cancer patients.

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1. Rickles, F.R., and Falanga, A. 2001. Molecular basis for the relationship between thrombosis and can-

cer. *Thromb. Res.* **102**:V215–V224.
 2. Nash, G.F., Turner, L.F., Scully, M.F., and Kakkar, A.K. 2002. Platelets and cancer. *Lancet Oncol.* **3**:425–430.
 3. Karpatkin, S., and Pearlstein, E. 1981. Role of platelets in tumor cell metastases. *Ann. Intern. Med.* **95**:636–641.
 4. Nieswandt, B., Hafner, M., Echtenacher, B., and Mannel, D.N. 1999. Lysis of tumor cells by natural killer cells in mice is impeded by platelets. *Cancer Res.* **59**:1295–1300.
 5. Trikha, M., and Nakada, M.T. 2002. Platelets and cancer: implications for antiangiogenic therapy. *Semin. Thromb. Hemost.* **28**:39–44.
 6. Boucharaba, A., et al. 2004. Platelet-derived lysophosphatidic acid supports the progression of osteolytic bone metastases in breast cancer. *J. Clin. Invest.* **114**:1714–1725. doi:10.1172/JCI200422123.
 7. Jurasz, P., Alonso-Escolano, D., and Radomski, M.W. 2004. Platelet-cancer interactions: mechanisms and pharmacology of tumour cell-induced platelet aggregation. *Br. J. Pharmacol.* doi:10.1038/sj.bjp.0706013.
 8. FitzGerald, G.A. 1991. Mechanisms of platelet activation: thromboxane A2 as an amplifying signal for other agonists. *Am. J. Cardiol.* **68**:11B–15B.
 9. Brass, L.F. 2003. Thrombin and platelet activation. *Chest.* **124**:18S–25S.
 10. Mills, G.B., and Moolenaar, W.H. 2003. The emerging role of lysophosphatidic acid in cancer. *Nat. Rev. Cancer.* **3**:582–591.
 11. Ishii, I., Fukushima, N., Ye, X., and Chun, J. 2004.

Lysophospholipid receptors: signaling and biology. *Annu. Rev. Biochem.* **73**:321–354.
 12. van Corven, E.J., Groenink, A., Jalink, K., Eichholtz, T., and Moolenaar, W.H. 1989. Lysophosphatidate-induced cell proliferation: identification and dissection of signaling pathways mediated by G proteins. *Cell.* **59**:45–54.
 13. Mundy, G.R. 2002. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat. Rev. Cancer.* **2**:584–593.
 14. Yin, J.J., et al. 1999. TGF- β signaling blockade inhibits PTHrP secretion by breast cancer cells and bone metastases development. *J. Clin. Invest.* **103**:197–206.
 15. Kang, Y., et al. 2003. A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell.* **3**:537–549.
 16. Falanga, A. 2004. The effect of anticoagulant drugs on cancer. *J. Thromb. Haemost.* **2**:1263–1265.
 17. Varki, N.M., and Varki, A. 2002. Heparin inhibition of selectin-mediated interactions during the hematogenous phase of carcinoma metastasis: rationale for clinical studies in humans. *Semin. Thromb. Hemost.* **28**:53–66.
 18. Amirkhosravi, A., et al. 2003. Inhibition of tumor cell-induced platelet aggregation and lung metastasis by the oral GpIIb/IIIa antagonist XV454. *Thromb. Haemost.* **90**:549–554.
 19. Bakewell, S.J., et al. 2003. Platelet and osteoclast beta3 integrins are critical for bone metastasis. *Proc. Natl. Acad. Sci. U. S. A.* **100**:14205–14210.

The *Staphylococcus aureus* “superbug”

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There has been some debate about the disease-invoking potential of *Staphylococcus aureus* strains and whether invasive disease is associated with particularly virulent genotypes, or “superbugs.” A study in this issue of the *JCI* describes the genotyping of a large collection of nonclinical, commensal *S. aureus* strains from healthy individuals in a Dutch population. Extensive study of their genetic relatedness by amplified restriction fragment typing and comparison with strains that are associated with different types of infections revealed that the *S. aureus* population is clonal and that some strains have enhanced virulence (see the related article beginning on page 1732). This is discussed in the context of growing interest in the mechanisms of bacterial colonization, antibiotic resistance, and novel vaccines.

Nasal colonization

Staphylococcus aureus is a common commensal of humans and its primary habitat is the moist squamous epithelium of the anterior nares (1). About 20% of the population are always colonized with *S. aureus*, 60% are

intermittent carriers, and 20% never carry the organism. As there is considerable evidence that carriage is an important risk factor for invasive infection (1, 2), it is surprising that so little is known about the bacterial factors that promote colonization of squamous epithelial surfaces and the host factors that determine whether an individual can be colonized or not.

Methicillin-resistant *S. aureus*

Healthy individuals have a small but finite risk of contracting an invasive infection caused by *S. aureus*, and this risk is increased among carriers. Hospital patients who are

catheterized or who have been treated surgically have a significantly higher rate of infection. In some, but not all, developed countries, many nosocomial infections are caused by *S. aureus* strains that are multiply resistant to antibiotics — known as methicillin-resistant *Staphylococcus aureus* (MRSA) (3, 4) — although the acronym MRSA is somewhat misleading because the semisynthetic β -lactam methicillin is no longer used to treat *S. aureus* infections. In MRSA, the horizontally acquired *mecA* gene encodes a penicillin-binding protein, PBP2a, which is intrinsically insensitive to methicillin and all β -lactams that have been developed, including the isoxazolyl penicillins (e.g., oxacillin) that superseded methicillin, in addition to the broad spectrum β -lactams (third-generation cephalosporins, cefamycins, and carbapenems) that were introduced primarily to treat infections caused by Gram-negative bacteria (4) (Figure 1). In contrast to nosocomial MRSA strains, which are usually multidrug resistant, the recently emerged community-acquired MRSA (CA-MRSA) strains are susceptible to drugs other than β -lactams (5).

Nonstandard abbreviations used: AFLP, amplified fragment length polymorphism; CA-MRSA, community-acquired MRSA; MLST, multilocus sequence typing; MRSA, methicillin-resistant *Staphylococcus aureus*; PVL; Panton-Valentine leukocidin; WTA, wall teichoic acid.

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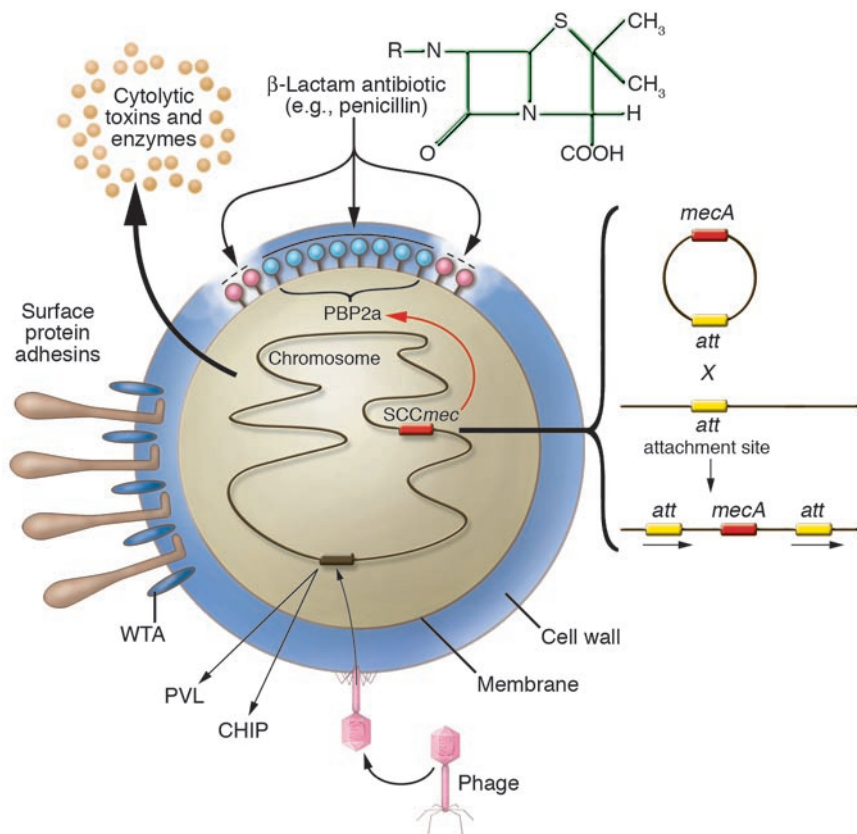


Figure 1

Schematic diagram illustrating how *S. aureus* acquires resistance to methicillin and its ability to express different virulence factors. The bacterium expresses surface protein adhesins and WTA and also secretes many toxins and enzymes by activation of chromosomal genes. Adhesins and WTA have been implicated in nasal and skin colonization. Resistance to methicillin is acquired by insertion of a horizontally transferred DNA element called *SCCmec*. Five different *SCCmec* elements can integrate at the same site in the chromosome by a Campbell-type mechanism involving site-specific recombination. The *mecA* gene encodes a novel β -lactam-insensitive penicillin binding protein, PBP2a, which continues to synthesize new cell wall peptidoglycan even when the normal penicillin binding proteins are inhibited. Some virulence factors such as PVL and the chemotaxis inhibitory protein, CHIP, are encoded by genes located on lysogenic bacteriophages.

The term MRSA is synonymous with multidrug-resistant *S. aureus* because many nosocomial MRSA strains are resistant to most commonly used antibiotics. The glycopeptide vancomycin was the last available drug to which this organism had remained uniformly sensitive until recent reports of low-level glycopeptide resistance (6) and, very recently, the transfer of high-level vancomycin resistance from *Enterococcus* to *S. aureus* (7). Although new drugs, including linezolid and synergicid, have recently been introduced to treat MRSA infections (8), there is a worrying lack of novel drugs in the pipeline.

Virulence factors

S. aureus strains can express a wide array of potential virulence factors (Figure 1), including surface proteins that promote adher-

ence to damaged tissue (9), bind proteins in blood to help evade antibody-mediated immune responses (9), and promote iron uptake (10). The organism also expresses a number of membrane-damaging toxins and superantigen toxins that can cause tissue damage and the symptoms of septic shock, respectively (11). There is a growing realization that *S. aureus* has multiple mechanisms for evading both innate immunity mediated by polymorphonuclear leukocytes (12, 13) and induced immunity mediated by both B and T cells (11, 14). Some virulence factors are expressed by genes that are located on mobile genetic elements called pathogenicity islands (e.g., toxic shock syndrome toxin-1 and some enterotoxins; ref. 15) or lysogenic bacteriophages (e.g., Panton-Valentine leucocidin [PVL]; refs. 15, 16) and

factors associated with suppressing innate immunity such as the chemotaxis inhibitory protein and staphylokinase (ref. 13), which are integrated in the bacterial chromosome.

The *S. aureus* population is clonal

The study by Melles et al. reported in this issue of the *JCI* (17) examines the major questions about natural populations of *S. aureus* concerning clonality and virulence. The authors examined 829 *S. aureus* strains from healthy donors from the city of Rotterdam in The Netherlands. Selective amplified fragment length polymorphism (AFLP) amplification analysis was used to compare genetic relatedness of strains, and this analysis revealed the existence of 3 major and 2 minor genetic clusters, subsequently confirmed by multilocus sequence typing (MLST) (Figure 2). The authors therefore concluded that the *S. aureus* population is clonal. These clusters corresponded to the predominant groups identified in a recent study of carriage isolates from the county of Oxfordshire in the United Kingdom using MLST (18). Thus the same clonal lineages appear to be dominant in 2 distinct geographic locations.

Evidence for clones with enhanced virulence

Melles et al. (17) also examined a smaller number of isolates from individuals with invasive disease (bacteremia and deep-seated abscesses) as well as those from individuals with severe impetigo. There was clear evidence that some clonal types are more virulent than others in that they appeared more frequently among disease isolates than among carriage isolates (Figure 2). This sharply contrasts with the earlier Oxfordshire study, in which no evidence for hypervirulent clones was found (18). In the Melles et al. study, bacteremia in elderly patients was significantly more frequently caused by 1 strain in cluster IVa (17). In addition, strains causing severe skin infections were significantly more frequently found to be members of cluster IVb. This could be due to lysogenization of a progenitor IVb strain with the bacteriophage that encodes PVL (16), a toxin that is strongly associated with severe skin infections (19).

The question remains: Why did this study find evidence for virulent clones, whereas the Oxfordshire study did not? Perhaps this discrepancy can be attributed to the fact that Melles et al. tested a larger number of strains (17). Furthermore, the Oxfordshire study (18) was confined to isolates obtained from patients with invasive infec-

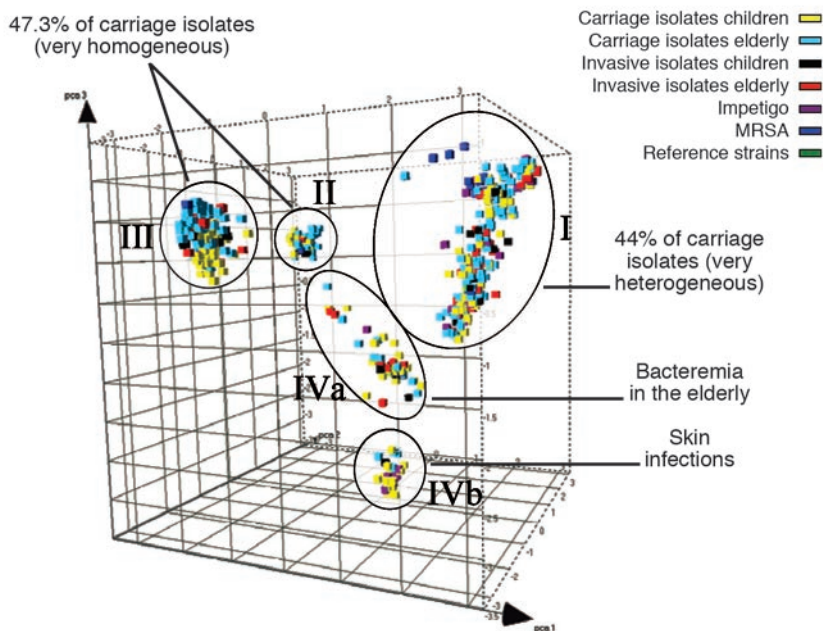


Figure 2

Principal component analysis of 1,056 *S. aureus* strains reveals genetic clusters of hypervirulent clones (17). The different cubes, plotted here in a 3D space and colored according to their source, represent each *S. aureus* strain analyzed in the Melles et al. study. The 5 circles indicate the 3 major (I, II, and III) and 2 minor (IVa and IVb) different phylogenetic clusters identified by AFLP. While strains from each of the genetic clusters are essentially able to cause invasive disease, some clusters contain proportionally more invasive isolates.

tions requiring admission to hospital and thus excluded impetigo and skin infections. Another factor might have been the prevalence of MRSA strains present in the nosocomial invasive isolates from individuals in the UK; in contrast, of the invasive strains isolated from Dutch individuals, none were identified as MRSA. This is consistent with the conclusion that all strains of *S. aureus* have the potential to cause infection and that some are more virulent than others.

Evolution of MRSA

MRSA strains have emerged by acquisition of mobile genetic elements called *SCCmec* cassettes, which carry the *mecA* gene that encodes PBP2a. There are 5 different cassettes (*SCCmec* types I–V; refs. 3, 20, and 21). It is now clear that major MRSA clones were created on multiple occasions by acquisition of *SCCmec* by prevalent strains that have continued to flourish (22). None of the Dutch carriage or invasive disease isolates were found to harbor the *mecA* gene (17). Nevertheless, the authors examined a variety of MRSA strains from other sources and found *SCCmec*-containing strains in each of their major genetic clusters. This is consistent with previous studies (22) that

established that MRSA strains have arisen many times by transfer of *SCCmec* cassettes into susceptible host strains.

PVL is a 2-component cytolytic toxin with high affinity for human leukocytes (11). It has been associated with *S. aureus* strains causing severe skin infections (19) and with necrotizing pneumonia in previously healthy youths (23). In the Melles et al. study (17), PVL was rarely found in the carriage isolates (0.6%) but was present in a significantly high number of strains that caused abscesses and arthritis. PVL is also expressed by the newly emerged CA-MRSA strains, which appear to have enhanced virulence (5, 23). However, Melles et al. did not examine CA-MRSA strains in this study.

Future prospects for combating *S. aureus*

Given the problems caused by the development of antibiotic-resistant *S. aureus*, vaccination may well have a significant role to play in controlling this organism in the future. A number of companies are developing products intended for active or passive immunization against *S. aureus* infections, including a capsular polysaccharide vaccine that has been subjected to a clinical trial

with hemodialysis patients (24), a monoclonal antibody (25), and human immunoglobulin that is enriched for antibodies that recognize clumping factor A (26).

An increased understanding of how *S. aureus* colonizes the nares could allow improved methods for controlling nasal and skin carriage. Recent studies of mutant strains defective in wall teichoic acid (WTA) in a rat model of nasal colonization implicated WTA in colonization (27). Also, several different surface proteins can promote adherence of *S. aureus* to squamous epithelial cells isolated from the nares (28, 29) and could act as adhesins involved in nasal colonization. Another mystery that deserves greater attention is the question of why some members of the population never carry *S. aureus*, while others are persistent carriers.

The study by Melles et al. (17), in combination with MLST analysis (18), provides a solid foundation for analysis of novel hypervirulent or epidemic drug-resistant *S. aureus* clones that might arise in the future. One thing seems certain: *S. aureus* will continue to respond to challenges imposed by humans' continued attempts to combat its carriage and development of related disease.

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1. Peacock, S.J., de Silva, I., and Lowy, F.D. 2001. What determines nasal carriage of *Staphylococcus aureus*? *Trends Microbiol.* **9**:605–610.
2. von Eiff, C., Becker, K., Machka, K., Stammer, H., and Peters, G. 2001. Nasal carriage as a source of *Staphylococcus aureus* bacteremia. Study Group. *N. Engl. J. Med.* **344**:11–16.
3. Hiramatsu, K., Cui, L., Kuroda, M., and Ito, T. 2001. The emergence and evolution of methicillin-resistant *Staphylococcus aureus*. *Trends Microbiol.* **9**:486–493.
4. Chambers, H.F. 1997. Methicillin resistance in staphylococci: molecular and biochemical basis and clinical implications. *Clin. Microbiol. Rev.* **10**:781–791.
5. Vandenesch, F., et al. 2003. Community-acquired methicillin-resistant *Staphylococcus aureus* carrying Panton-Valentine leukocidin genes: worldwide emergence. *Emerging Infect. Dis.* **9**:978–984.
6. Hiramatsu, K. 2001. Vancomycin-resistant *Staphylococcus aureus*: a new model of antibiotic resistance. *Lancet Infect. Dis.* **1**:147–155.
7. Weigel, L.M., et al. 2003. Genetic analysis of a high-level vancomycin-resistant isolate of *Staphylococcus aureus*. *Science*. **28**:1569–1571.
8. Eliopoulos, G.M. 2003. Quinupristin-dalfopristin and linezolid: evidence and opinion. *Clin. Infect. Dis.* **36**:473–481.
9. Foster, T.J., and Höök, M. 1998. Surface protein adhesins of *Staphylococcus aureus*. *Trends Microbiol.* **6**:484–488.



10. Mazmanian, S.K., et al. 2003. Passage of heme-iron across the envelope of *Staphylococcus aureus*. *Science*. **299**:906–909.
11. Bohach, G.A., and Foster, T.J. 1999. *Staphylococcus aureus* exotoxins. In *Gram positive bacterial pathogens*. V.A. Fischetti, R.P. Novick, J.J. Ferretti, and J.I. Rood, editors. American Society for Microbiology. Washington, D.C., USA. 367–378.
12. Fedtke, I., Gotz, F., and Peschel, A. 2004. Bacterial evasion of innate host defenses—the *Staphylococcus aureus* lesson. *Int. J. Med. Microbiol.* **294**:189–194.
13. de Haas, C.J., et al. 2004. Chemotaxis inhibitory protein of *Staphylococcus aureus*, a bacterial anti-inflammatory agent. *J. Exp. Med.* **199**:687–695.
14. Goodyear, C.S., and Silverman, G.J. 2003. Death by a B cell superantigen: in vivo VH-targeted apoptotic supraclonal B cell deletion by a staphylococcal Toxin. *J. Exp. Med.* **197**:1125–1139.
15. Novick, R.P. 2003. Mobile genetic elements and bacterial toxinoses: the superantigen-encoding pathogenicity islands of *Staphylococcus aureus*. *Plasmid*. **49**:93–105.
16. Narita, S., et al. 2001. Phage conversion of Panton-Valentine leukocidin in *Staphylococcus aureus*: molecular analysis of a PVL-converting phage, phiSLT. *Gene*. **268**:195–206.
17. Melles, D.C., et al. 2004. Natural population dynamics and expansion of pathogenic clones of *Staphylococcus aureus*. *J. Clin. Invest.* **114**:1732–1740. doi:10.1172/JCI200423083.
18. Feil, E.J., et al. 2003. How clonal is *Staphylococcus aureus*? *J. Bacteriol.* **185**:3307–3316.
19. Lina, G., et al. 1999. Involvement of Panton-Valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. *Clin. Infect. Dis.* **29**:1128–1132.
20. Ito, T., et al. 2004. Novel type V staphylococcal cassette chromosome mec driven by a novel cassette chromosome recombinase, ccrC. *Antimicrob. Agents Chemother.* **48**:2637–2651.
21. Ma, X.X., et al. 2002. Novel type of staphylococcal cassette chromosome mec identified in community-acquired methicillin-resistant *Staphylococcus aureus* strains. *Antimicrob. Agents Chemother.* **46**:1147–1152.
22. Enright, M.C., et al. 2002. The evolutionary history of methicillin-resistant *Staphylococcus aureus* (MRSA). *Proc. Natl. Acad. Sci. U. S. A.* **99**:7687–7692.
23. Gillet, Y., et al. 2002. Association between *Staphylococcus aureus* strains carrying gene for Pantone-Valentine leukocidin and highly lethal necrotising pneumonia in young immunocompetent patients. *Lancet*. **359**:753–759.
24. Fattom, A.I., Horwith, G., Fuller, S., Propst, M., and Naso, R. 2004. Development of StaphVAX, a polysaccharide conjugate vaccine against *S. aureus* infection: from the lab bench to phase III clinical trials. *Vaccine*. **17**:880–887.
25. Hall, A.E., et al. 2003. Characterization of a protective monoclonal antibody recognizing *Staphylococcus aureus* MSCRAMM protein clumping factor A. *Infect. Immun.* **71**:6864–6870.
26. Vernachio, J., et al. 2003. Anti-clumping factor A immunoglobulin reduces the duration of methicillin-resistant *Staphylococcus aureus* bacteremia in an experimental model of infective endocarditis. *Antimicrob. Agents Chemother.* **47**:3400–3406.
27. Weidenmaier, C., et al. 2004. Role of teichoic acids in *Staphylococcus aureus* nasal colonization, a major risk factor in nosocomial infections. *Nat. Med.* **10**:243–245.
28. Roche, F.M., Meehan, M., and Foster, T.J. 2003. The *Staphylococcus aureus* surface protein SasG and its homologues promote bacterial adherence to human squamous nasal epithelial cells. *Microbiol. J.* **149**:2759–2767.
29. O'Brien, L.M., Walsh, E.J., Massey, R.C., Peacock, S.J., and Foster, T.J. 2002. *Staphylococcus aureus* clumping factor B (ClfB) promotes adherence to human type I cytokeratin 10: implications for nasal colonization. *Cell. Microbiol.* **4**:759–770.

Acid sensing in renal epithelial cells

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The kidney adjusts net acid excretion to match production with exquisite precision, despite little or no change in the plasma bicarbonate concentration. The acid-sensing pathway that signals the kidney to increase acid secretion involves activation of the proto-oncogene c-Src. A new study in this issue shows that proline-rich tyrosine kinase 2 (Pyk2) is responsible for acid-induced activation of c-Src and is essential for acid sensing in renal epithelial cells (see the related article beginning on page 1782). The findings implicate a broader role for Pyk2 in acid-base homeostasis in bone and other tissues beyond the kidney.

Although the principal product of metabolism in mammalian cells is the volatile acid carbon dioxide, humans on a typical Western diet produce about 70 millimoles of nonvolatile acid per day. Remarkably, varying metabolic acid production over a range of 0–150 millimoles is accompanied by a matching increase in net acid excretion by the kidney with a change of only 1 mM in plasma bicarbonate concentration (1). The adaptive responses that enable the kidney to increase net acid excretion in response to

increased acid generation have been studied extensively in animal models of metabolic acidosis. In the proximal tubule, acidosis increases the activity of luminal and basolateral proteins involved in bicarbonate transport (2, 3), ammonia generation (4), and the reabsorption and metabolism of citrate (5). In the collecting duct, acidosis suppresses bicarbonate secretion (6) and stimulates recruitment of proton pumps to the luminal membrane of intercalated cells (7). Of the acid-base transporters in the proximal tubule, the luminal sodium/hydrogen exchanger 3 (NHE3) has a prominent role, and the mechanism by which its activity increases during metabolic acidosis has been examined in some detail. Metabolic acidosis acutely increases the kinetic activity of NHE3 through direct pH effects and by phosphorylation (8), while chronic

acidosis increases the number of NHE3 transporters (9).

Acid-base transporter kinetics cannot account for precise pH sensing

How does the kidney “know” to adjust net acid excretion with such precision with only minimal changes in plasma bicarbonate concentration? Available data in the physiology literature suggests that transporter kinetics alone cannot account for this degree of sensitivity. In the proximal tubule, a reduction in extracellular bicarbonate induces a fall in intracellular pH, which directly activates the sodium/hydrogen exchanger through an intracellular pH regulatory site (10). This requires a change in intracellular pH of about 0.1 to achieve a 50% increase in the rate of transport or an approximately 5% change in the rate of transport in response to a change in extracellular bicarbonate concentration of 1 mM. Both the luminal vacuolar H⁺-ATPase and the basolateral sodium bicarbonate cotransporter in the proximal tubule are even less responsive to changes in intracellular pH (11–13). This suggests that a bicarbonate (or pH) sensor that can amplify luminal proton secretion must be present.

Nonstandard abbreviations used: FAK, focal adhesion kinase; NHE3, sodium/hydrogen exchanger 3; OKP, opossum kidney clone P; Pyk2, proline-rich tyrosine kinase 2.

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