

## CHAPTER 1

# The Tumor Microenvironment as Target for New Cancer Therapies

Reto A. Schwendener\* and Sibel Mete

*Institute of Molecular Cancer Research, University of Zurich, Zurich, Switzerland*

**Abstract:** Solid tumors grow within a complex microenvironment composed of diverse cell types such as fibroblasts, endothelial cells, mast cells, macrophages and immune cells that are attracted by tumor cell derived factors and embedded in an extracellular matrix. Molecular and cellular interactions between epithelial cells and cells surrounding the tumor stroma promote growth, invasion and spread of tumors. To delay or impede tumor growth, the tumor microenvironment (TME) is increasingly being explored as a potential therapeutic target for which novel strategies are developed.

This article reviews how key interactions between tumor cells and surrounding mesenchymal and immune cells in the TME can promote tumor progression and it highlights cellular and molecular elements that might represent novel therapeutic targets. Special emphasis is given on therapies targeted towards tumor-associated macrophages. As main class of drugs the bisphosphonates are covered with their properties to repolarize a pro-tumorigenic, immunosuppressive environment to a tumor growth inhibiting and immunocompetent microenvironment. Properties and advantages of liposome-encapsulated bisphosphonates as macrophage depleting or modulating agents as well as the latest developments towards clinical applications of compounds targeting cellular and molecular components of the TME are described and reviewed.

**Keywords:** Tumor microenvironment, stromal cells, macrophages, tumor associated macrophages, neutrophils, tumor associated neutrophils, myeloid derived suppressor cells, immune cells, fibroblasts, stromal interactions, bisphosphonates, clodronate, liposomes, macrophage depletion, therapeutic targets, repolarization, reprogramming, immunotherapy, adjuvant cancer therapy.

## INTRODUCTION

Cancer progression mostly depends on the ability of malignant cells to exploit physiological processes of the host. Solid tumors can only develop with a steady

---

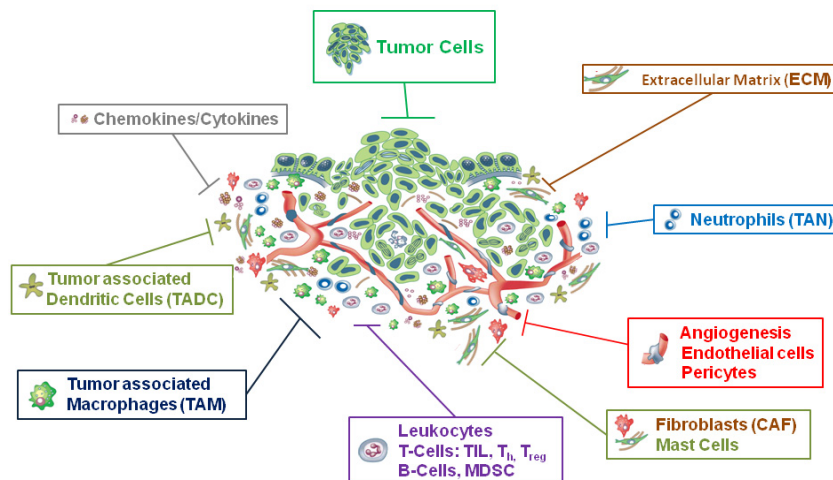
\*Address correspondence to Reto Schwendener: Institute of Molecular Cancer Research, University of Zurich, Winterthurerstrasse 190, CH-8057 Zurich, Switzerland; Email: [rschwendener@imcr.uzh.ch](mailto:rschwendener@imcr.uzh.ch)

supply of nutrients and oxygen, provided by blood and by support of cells, factors and conditions provided by the microenvironment [1-26]. Cells of the microenvironment become activated by communication with the tumor cells, consequently creating numerous conditions that promote cancer growth and ultimately lead to metastatic dissemination [5, 27-38].

First evidences about the effect of the host microenvironment on tumor growth were provided in the 1970s [39, 40] postulating that expansion from a single mutated cell to a solid tumor can only occur when the stromal environment is altered in a way to allow unrestrained tumor growth. Despite of continuous efforts, for many years cancer research largely focused on cancer-cell driven carcinogenesis and on understanding the mutations causing neoplastic cell transformations. This cancer cell centric view of tumor progression largely ignored the fact that complex interactions between cancer cells and stromal components tightly regulate and orchestrate tumor growth and metastatic dissemination. For this and other reasons, even after decades of implementing treatments that selectively target the tumor cell, survival of metastatic cancer patients is still disappointingly short. Therefore, novel strategies are urgently needed to complement the classical treatment modalities with new therapeutic approaches. In this regard, interactions between cancer cells and their host environment offer novel opportunities for therapies based on the improved understanding of the nature of these interactions and the mechanisms that govern them. Treatment modalities that target both cancer cells and components of the tumor microenvironment (TME) are likely to be more effective than those classically directed against cancer cells. A potential advantage in targeting the non-malignant cells of the TME is that these cells tend to be more genetically stable and are therefore less expected to develop resistance to therapies.

To provide new therapeutic strategies targeted at the immune components of the TME, it is critical to understand how these cells are altered during tumor progression and how they reciprocally influence tumor initiation, progression and metastasis. Here, we review the current understanding of the interactions of tumor cells with the microenvironment with a particular focus on tumor associated macrophages (TAM), tumor associated neutrophils (TAN) and myeloid derived suppressor cells (MDSC) (Fig. 1). Current therapeutic approaches aiming at the

TME, in particular cell-based therapies and therapies with bisphosphonates (BP), a class of drugs that show to have potential immunomodulatory properties on immune cells in cancer, are reviewed and discussed. Their pharmacological properties and anti-tumor activities are summarized with a special emphasis on the properties of clodronate encapsulated in liposomes, a drug formulation that has the ability to deplete tumor-associated macrophages. Together, all these properties point toward the significance of re-programming myeloid cell phenotypes to affect tumor growth and accordingly, suggest this concept as a promising strategy to complement the established anticancer treatment modalities.



**Figure 1:** The tumor microenvironment (TME) is composed of numerous different cell types that infiltrate a growing tumor. These cell types include vascular or lymphatic endothelial cells, endothelial cell supporting pericytes, fibroblasts, mast cells, and the cells of the innate and adaptive immune system, namely macrophages, dendritic cells, neutrophils, leukocytes (T cells, B cells) and myeloid derived suppressor cells (MDSC). In addition, the non-cellular components of the TME include components of the extracellular matrix (ECM) and soluble factors as chemokines and cytokines. The therapeutic strategies of targeting components of the TME include the tumor cells themselves by combining novel adjuvant therapy approaches targeted to cellular or molecular components of the TME with the current chemotherapy and radiotherapy. Novel and experimental therapies that aim at components of the TME include inhibitors of angiogenesis (*e.g.* anti-VEGF or VEGF-receptor antibodies), inhibitors of fibroblast functions, drugs aimed at macrophages and neutrophils (depletion, re-polarization), immune stimulating therapies (antibodies, cellular therapies, vaccines), inhibitors of EMC components (*e.g.* MMP inhibitors) and inhibitors of chronic inflammation.

## **Characteristics and Components of the Tumor Microenvironment**

### ***Angiogenesis, Hypoxia and Oxygen Regulation***

Angiogenesis is a key process for tumor development. Small colonies of malignant cells of 1-2 mm<sup>3</sup> size, the so-called “carcinoma-in-situ”, alter their phenotype to induce continuous proliferation of endothelial cells and development of new blood and lymph vessels. This “angiogenic switch” triggers the expansion of the tumor cells by growth of new vessels that provide nutrients, oxygen and removal of waste products, as well as an escape route for metastasizing tumor cells [22, 24, 41-47].

Although various studies demonstrated that tumor cells produce pro-angiogenic factors, angiogenesis is also stimulated by activated myeloid cells recruited into the neoplastic tissue. Production of vascular endothelial growth factor (VEGF) is an important mechanism by which tumor infiltrating myeloid cells trigger and enhance angiogenesis and foster tumor development [48, 49]. TAMs are a major source of VEGF as they accumulate in poorly vascularized hypoxic areas and respond to hypoxia by releasing VEGF and other angiogenic factors (see below and Fig. 2). Hypoxic conditions in tumors stimulate the expression of pro-angiogenic molecules by activating hypoxia-inducible factors (HIFs) in macrophages [19, 50-58]. Activated macrophages also release nitric oxide (NO), a molecule that provokes increased vascular flow [46, 59-64]. Myeloid derived suppressor cells (MDSC, see below) represent another cell population involved in tumor angiogenesis. Tumor cell educated MDSCs express elevated levels of the matrix degrading metalloproteinase MMP-9 that triggers VEGF release from the extracellular matrix (ECM) which induces proliferation of endothelial cells [65-68]. Despite of their low abundance, tumor associated neutrophils (TANs, see below) have also been reported to support tumor growth by producing pro-angiogenic factors such as VEGF, IL-8 and proteases including MMPs and elastase [65, 69-73]. In this context, it was found that Stat3 activation in tumor-associated myeloid cells is critical for tumor angiogenesis [74]. Last but not least, pericytes, responsible for the stabilization of endothelial cells of the vessel wall, play a crucial role in hem- and lymphangiogenesis where they closely interact with endothelial cells and vascular smooth muscle cells [75-78]. Although the importance of myeloid cells in promoting tumor angiogenesis has been

investigated carefully, the underlying molecular mechanisms as well as the individual contributions of the different cell types remain to be fully explored.

### ***The Extracellular Matrix (ECM) and Regulation of Invasion and Metastasis***

The ECM serves as a scaffold for the cellular components of normal tissues as well as of tumors and it also strongly influences cell growth, differentiation, adhesion, motility, invasion and viability. The ECM consists of proteins that possess multiple functions and that provide vital signals for tumor progression and metastatic spread [79-85]. The matrix metalloproteinases (MMPs) with their proteolytic activity are key modulators of the TME and the most prominent family of proteases associated with tumorigenesis. They play an important role in ECM turnover and remodeling and in tumor cell migration. MMPs also control signaling pathways that regulate cell growth, inflammation and angiogenesis [86-88]. The transmission of signals between the ECM and neighboring cells occurs mainly through the integrins. These proteins have the capability to transduce mechanical cues created by the ECM or the cell cytoskeleton into chemical signals that regulate many cellular processes such as proliferation, survival, migration, and invasion [80, 82, 84, 89, 90].

An important step in tumor progression is the acquisition of invasive properties by tumor cells. Epithelial-mesenchymal transition (EMT) is a well characterized mechanism, through which epithelial cells trans-differentiate and acquire an invasive, fibroblast-like phenotype [32, 91-95]. Although it is well established that the TME contains cytokines, growth factors and enzymes that induce EMT, the cellular sources of these factors remain to be fully identified. TAMs, cancer-associated fibroblasts, CAFs, mesenchymal stem cells (MSCs) and lymphocytes have all been shown to contribute to an EMT promoting tumor microenvironment [95, 96]. Pro-inflammatory macrophages have likewise been shown to induce EMT at the invasive front, but also in the core of tumors, mainly through stabilization of Snail and Smad3, key mediators of EMT [97-99].

The ability of a growing tumor to invade tissue and to metastasize to distant organs was thought to be strictly cancer cell intrinsic. However, it is now established that tumor infiltrating and resident myeloid cells significantly contribute to tumor progression. Myeloid cell subsets as macrophages, MDSCs,

neutrophils and mast cells as well as soluble factors play an important role in ECM remodeling, invasion and metastasis which will be discussed in the forthcoming paragraphs.

### ***Chemokines and Cytokines***

The TME is rich in chemokines and cytokines which are vital factors for the regulation of tumor growth, invasion and metastasis. Most of resident and infiltrating cellular components of the TME contribute to a dynamic chemokine/cytokine network which is spatially and temporally fluctuating, depending on the local conditions of the TME. Beyond activating tumor vascularization, infiltrating myeloid cells also promote tumor growth by creating a microenvironment that is rich in growth factors and pro-inflammatory cytokines that stimulate proliferation and survival of neoplastic cells [26, 36, 70, 100-110]. Myeloid cell-derived cytokines and growth factors secreted by TAMs and MDSCs such as epidermal growth factor (EGF), platelet-derived growth factor (PDGF), fibroblast growth factor (FGF) and hepatocyte growth factor (HGF) all contribute to tumor growth [111]. Besides directly promoting tumor cell proliferation, tumor-educated myeloid cells can also indirectly facilitate tumor growth through suppression of anti-tumor immune responses by secretion of immunosuppressive cytokines, generation of reactive oxygen species (ROSs) and increased activity of arginase and nitric oxide (NO). Another important immunosuppressive mediator, TGF- $\beta$  converts naive CD4<sup>+</sup> T cells to adaptive regulatory T cells [112, 113].

### ***Fibroblasts and Mast Cells***

Cancer associated fibroblasts (CAF) are a heterogeneous cell population. The main progenitors of activated fibroblasts in the TME are originating from resident fibroblasts. CAFs can also stem from pericytes, smooth muscle cells and from bone marrow derived mesenchymal cells [114, 115]. CAFs contribute to a pro-tumorigenic environment through interaction with other cells in the TME. They are regulators of tumorigenesis and they differ from tumor cells by being more genetically stable. CAFs have properties to enhance tumor angiogenesis by secretion of stromal cell-derived factor 1 (SDF-1), also known as CXCL12, which plays a central role in the promotion of tumor growth and angiogenesis [116]. Besides that they produce many growth

factors (HGF, VEGF, TGF- $\beta$ ), cytokines (IL-8, CXCL14, CCL7, IL-6, IL-1 $\alpha$ ), proteases (MMPs, uPA) and other enzymes [117]. The clinical relevance of CAFs tumor growth promoting role has also been recognized by exploiting CAF expressed factors as prognostic markers [114, 116, 118-122].

Mast cells (MC) are derived from the bone marrow and are also a heterogeneous cell population with many functions. Apart from their role in innate and adaptive immunity they influence tumor cell proliferation and invasion and modulate the immune responses to tumor cells [123]. The number of tumor infiltrating mast cells correlates with increased intratumoral microvessel density, enhanced tumor growth and invasion, and poor clinical outcome. MCs are predominantly located at the boundary between healthy tissues and the TME and are often found in close association with blood vessels. They support angiogenesis by expression of pro-angiogenic factors and by inhibition of ECM remodeling the MCs support tumor spread and metastasis. Tumor-associated mast cells are also regarded as potential therapeutic targets [124-128] and prognostic factor [129-131].

### ***Leukocytes***

Leukocyte infiltration into malignant tissue was first described by the pathologist Rudolf Virchow in 1863 [132]. Solid tumors contain various types and numbers of leukocytes that can represent up to 50% of the tumor mass. The major components of the leukocytic infiltrates in the TME are myeloid cells and B and T lymphocytes [38, 133] as well as regulatory T cells [134-138]. Specifically, myeloid cells are the major component of the leukocytic infiltrates found in tumors. Immune cell infiltration into tumors and the impact the immune cells have on cancer has been named cancer immunoediting or cancer immunosurveillance. This concept that describes the role the immune system plays in cancer development was considered and discussed throughout the last decades. The central principle is that the immune system can prevent tumor development but that it is also able to select tumor variants with reduced immunogenicity, and creating an inflammatory environment that provides tumors with mechanisms to escape immunodetection and elimination [139-146]. Initially, the presence of leukocytes in malignant neoplasms was thought to represent the host's immune response to a growing tumor [147]. Yet, solid tumors are mostly recognized as "self" and they do not evoke efficient immune responses

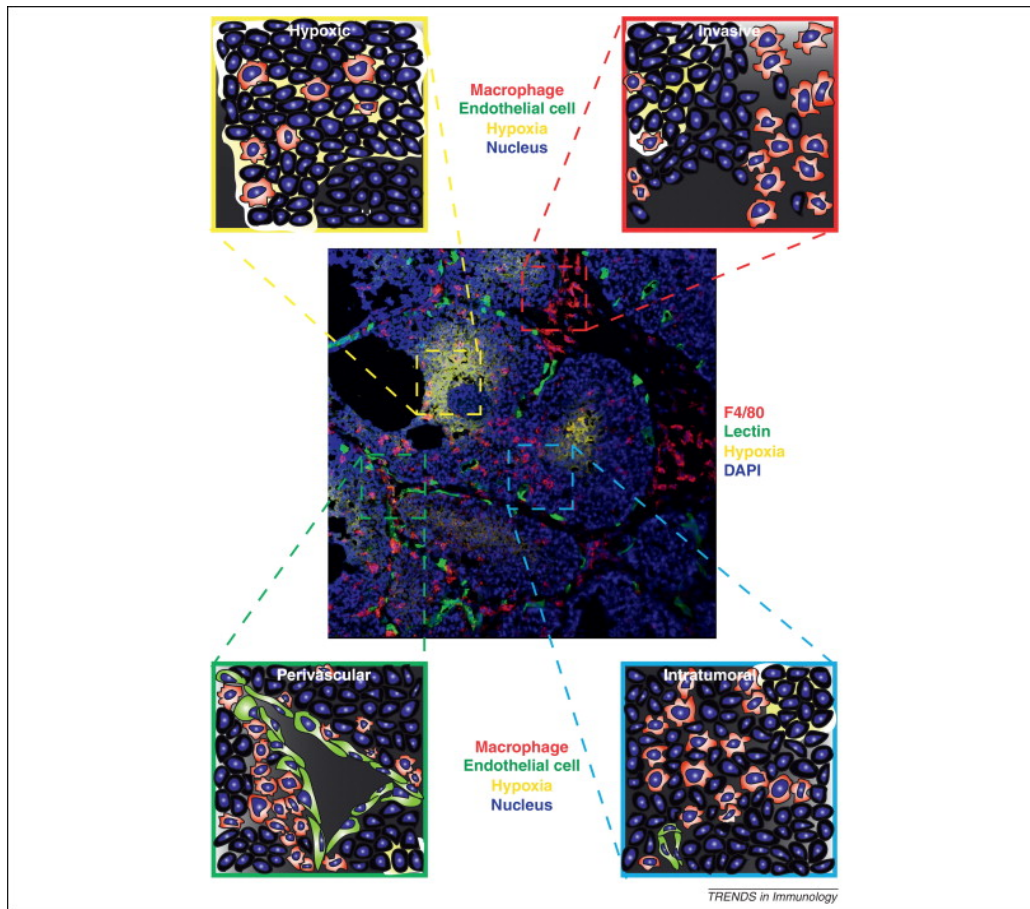
capable of eradicating tumors [148, 149]. In contrast, it was found that these cells are actively recruited to neoplastic tissues by tumor cells and that high numbers of several types of leukocytes are associated with tumor progression [38, 150-152]. Nevertheless in some cancers, the presence of leukocytes is associated with a favorable prognosis [153]. For example, enhanced infiltration of natural killer cells and cytotoxic T cells into tumors has been reported to correlate with a good prognosis in human ovarian, colorectal and gastric cancers [154, 155]. Similarly, cytotoxic activation of lymphocytes, particularly CD8<sup>+</sup> T cells in response to tumor growth result in regression [156]. In contrast, as described in more details below, tumor-activated myeloid leukocytes (TAMs, DCs, MDSCs) are known to restrain the protective function of these immune cells with anti-tumor activity and to promote tumor growth.

### ***Macrophages***

Macrophages belong to the mononuclear phagocyte system (MPS) which are cells involved in host defense functions, immune reactions, disposal of dead cells and cellular components and synthesis of biologically active compounds such as complement components and prostaglandins [157-159]. The MPS includes precursor cells in the bone marrow, blood monocytes, alveolar, peritoneal and splenic macrophages and Kupffer cells in the liver. Macrophages are extremely versatile cells that can adapt a particular phenotype depending on environmental stimuli. As most of the other cell types that populate the TME, they produce an assorted array of chemokines, cytokines, proteases, angiogenic and other growth factors. As unique property they possess the ability to phagozytose particular matter as dead cells, bacteria, viruses as well as artificial particles like liposomes, nanoparticles and other pharmaceutical drug carriers [157, 160-173]. Macrophages play a very important role in tumor development as they are a major component of the myeloid infiltrate in a tumor microenvironment.

Hence, of all cells of the myeloid lineage, they are among the most studied for their contribution to tumor development. Monocytes circulating in the blood are recruited to tumors by tumor-derived chemotactic factors such as the colony stimulating factors M-CSF and GM-CSF (macrophage and granulocyte-macrophage colony stimulating factor), CCL2 (chemokine C-C motif ligand 2,

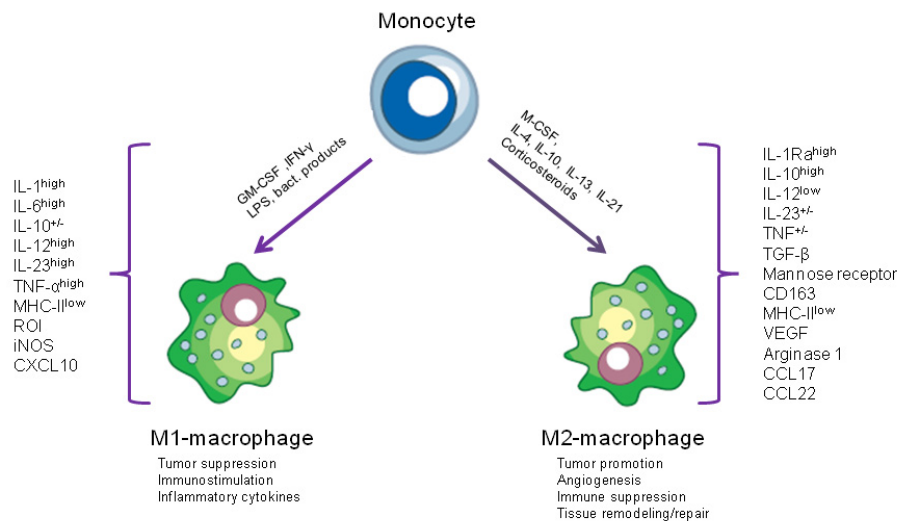




**Figure 2:** TAM can localize within unique tumor microenvironments. The immunofluorescent confocal micrograph in the center shows red stained F4/80<sup>+</sup> macrophages within a late-stage tumor of mammary carcinogenesis in the MMTV-PyMT mouse model. Areas of hypoxia are shown in yellow, functional vasculature is stained in green and all cell nuclei are stained in blue with DAPI. Insets display enlarged graphical representations of TAMs within a hypoxic region, at an invasive front, in a normoxic area within the tumor, and associated with the vasculature. Adapted from TRENDS in Immunology with permission [192].

MCP-1) and VEGF. Upon migrating into the tumor the monocytes differentiate into tissue-resident macrophages termed tumor-associated macrophages (TAMs) [38, 46, 59-63, 98, 174-191]. The term TAM defines localization of macrophages at the tumor-stroma interface and in the tumor core. As depicted in Fig. 2, TAMs localize at different sites in a tumor where they assume different functions that are driven by signals they obtain from the particular microenvironment in which they are located [192]. In response to diverse stimulants in the TME, TAMs undergo

polarized activation. The activation states of macrophages, as well as of other myeloid cells, have been defined by a nomenclature adapted from the T<sub>H</sub>1 and T<sub>H</sub>2 cell response, referred to as M1 (classical) or M2 (alternative) activation, respectively (Fig. 3).



**Figure 3:** Tumor-associated macrophages (TAM) can either assume tumor-promoting or -suppressing functions. Monocytes are attracted to a growing tumor through a chemokine gradient and differentiate in the tumor stroma to tissue macrophages. Depending on the particular cytokine composition in the microenvironment macrophages differentiate into two major conditions, the M1- or M2-phenotype. M1-TAMs actively present tumor antigens to T cells to elicit an anti-tumor immune response. M1-macrophages also produce, among other factors, the interleukins IL-1, IL-6, IL-12 and IL-23, TNF- $\alpha$  and iNOS, ROI and CXCL10 that all contribute to a tumor-suppressive TME. Conversely, in a TME that contains high levels of immunosuppressive factors that promote tumor growth, such as IL-1R $\alpha$  and IL-10, TGF- $\beta$  and scavenger receptors (MR, CD163) as well as arginase 1, VEGF, CCL17 and CCL23, M2-macrophages assume a pro-tumor function by supplying factors that enhance tumor progression, angiogenesis, tissue remodeling and immune suppression.

The classically activated M1-macrophages are pro-inflammatory cells that, following exposure to interferon- $\gamma$  (IFN- $\gamma$ ) or microbial products (*e.g.* LPS) release inflammatory cytokines, reactive nitrogen and oxygen intermediates, and

therefore they are endowed with an enhanced ability to kill tumor cells. In contrast, when TAMs are exposed to anti-inflammatory molecules, such as the interleukins IL-4, IL-10, IL-13 or glucocorticoid hormones and other factors, they are polarized to the opposite extreme called M2. M2-TAMs are poor antigen presenting cells and they support tumor growth, angiogenesis, and metastasis.

Conversely, TAMs suppress the immune system by responding to anti-inflammatory cytokines, apoptotic cells and immune complexes. M1 macrophage activation is characterized by high levels of major histocompatibility complex class II (MHC-II) expression and antigen presenting capacity, high production of pro-inflammatory cytokines such as IL-1, IL-12, IL-23, TNF- $\alpha$  and of toxic inducible nitric oxide synthase (iNOS) and reactive oxygen intermediates (ROI). In contrast, the M2 activation state is characterized by an IL-10<sup>high</sup> and IL-12<sup>low</sup> phenotype, expression of low levels of MHC-II and increased production of angiogenic factors and anti-inflammatory cytokines like IL-10, arginase and TGF- $\beta$ . Furthermore, M1 macrophages express opsonic receptors (*e.g.* Fc $\gamma$ RIII), whereas M2 macrophages preferentially express non-opsonic scavenger receptors such as the mannose receptor (MR) and CD163 [193-195]. In the majority of solid tumors TAMs predominantly are of the M2-phenotype. They promote angiogenesis (see Fig. 3) and express high levels of M2-markers (IL-10, TGF- $\beta$ , ARG1, CD163, MR) and low levels of mediators of inflammation (IL-6, IL-12, iNOS and TNF- $\alpha$ ) [181, 185, 186, 196-198].

This discrimination between M1 and M2 macrophages is a rather simplified view of two extremes of polarization and it does not fully represent the continuum of functional states of macrophages in the TME. Not only the intratumoral macrophages, but also spleen and peritoneal macrophages of tumor-bearing individuals share these similar immunosuppressive properties and play an important role in tumorigenesis [199, 200]. TAMs were also shown to attract CD4<sup>+</sup>CD25<sup>+</sup>FOXP3<sup>+</sup> regulatory T cells [112] that are known to suppress the anti-tumor function of cytotoxic T cells. Accumulation of T<sub>reg</sub> in tumors is a common feature of human cancers and the abundance, as well as their suppressor activities are highly correlated with a poor disease prognosis. In ovarian carcinoma it was found that TAMs regulate T<sub>reg</sub> trafficking to tumors by producing CCL22, a chemokine that mediates regulatory T cells recruitment [201].

Numbers, polarization state and cytokine expression pattern of TAMs can be correlated in several cancer types with the clinical prognosis of the disease [38, 185, 202]. For example, high numbers of TAMs are, among others, indicative of bad prognosis in colorectal cancer [203, 204], non small cell lung cancer (NSCLC) [98, 205-207], Hodgkin's lymphoma [208], breast cancer [209, 210], liver cancer [211, 212] and prostate cancer [213].

Analysis of the molecular basis of the TAM phenotype identified components of the NF- $\kappa$ B signaling system as one of the main players in the modulation of macrophage function [214-216]. For example, NF- $\kappa$ B inhibition by targeted deletion of IKK- $\beta$  in TAMs increased their anti-tumor activity through reduced production of arginase-1, IL-10 and TNF- $\alpha$  with concomitant increased production of iNOS and IL-12, suggesting that IKK- $\beta$  signaling in macrophages maintains their alternative tumor-promoting phenotype [217]. On the contrary, in more advanced stage tumors, a therapeutic effect was achieved through the restoration of NF- $\kappa$ B activity in myeloid cells [218, 219]. These divergent results may be associated with progressive modulation of NF- $\kappa$ B activity in tumor-infiltrating macrophages. Other important modulators of macrophage polarization are members of the STAT family of transcription factors. Although earlier evidence indicated that the STAT1 activation regulates the M1 activation of macrophages, recent reports argue that activated STAT1 may induce TAM-mediated suppressive activity and tumor progression [220-222]. In addition, STAT3 and STAT6 activation were also shown to be associated with M2 macrophage polarization [223, 224]. The interplay of TAMs with immune cells (B-cells, T-cells, regulatory T-cells and neutrophils) will be described and summarized in the respective paragraphs below.

### ***Dendritic Cells***

The second cell type that belongs to the mononuclear phagocyte system (MPS) are the dendritic cells (DC). DCs are bone marrow-derived cells originating from both lymphoid and myeloid progenitors. They populate all lymphoid organs including the thymus, spleen, and lymph nodes, and comparable to the macrophages, nearly all non-lymphoid tissues and organs. DCs have potent antigen-presenting capacity for the stimulation of T cells and they also belong to

the innate immune system where they respond as immature cells to danger signals in the microenvironment by differentiating and acquiring the capacity to mount primary immune responses. DCs possess powerful adjuvant activity as they have the ability to stimulate specific CD4 and CD8 T cells [38, 180, 225-231]. This property has made them attractive targets in vaccine development strategies for the prevention and treatment of infections, allograft reactions, allergic and autoimmune diseases and cancer. A major use of DCs as immunotherapeutic vaccines consists in their *ex vivo* priming combined with adjuvant treatments that eliminate immunosuppressive mechanisms in the TME (see below).

Similar to TAMs, the dendritic cells are also infiltrating tumor tissue following chemokine signals released by the TME. These tumor-associated dendritic cells (TADC) share many properties with TAMs as they can also be polarized either to tumor-suppressive “M1-like” or to tumor-promoting “M2-like” phenotypes [38, 231-233].

### ***Myeloid Derived Suppressor Cells (MDSC)***

Myeloid derived suppressor cells (MDSCs) are another complex but well characterized population of tumor-infiltrating myeloid cells that negatively affect the anti-tumor immune response. MDSCs are a heterogeneous population of cells comprised of monocyte, granulocyte and dendritic cell precursors and myeloid cells at an early stage of differentiation [67, 234-241]. These cells are defined by the co-expression of the monocytic marker CD11b and the granulocyte differentiation antigen Gr1 (constituted by the epitopes Ly6C and Ly6G in mice). In recent studies MDSCs were broadly classified as two major subsets, namely cells of granulocytic (CD11b<sup>+</sup>Ly6G<sup>+</sup>Ly6C<sup>low</sup>) and monocytic (CD11b<sup>+</sup>Ly6G<sup>-</sup>Ly6C<sup>high</sup>) phenotype [242, 243].

It has been well established that the frequency of these cells significantly increases in the spleen and bone marrow of tumor-bearing mice, as well as in the peripheral blood and tumors of cancer patients [241]. In naive tumor-free mice, MDSCs constitute approximately 30% of all bone marrow cells and 3% of all nucleated splenocytes. However, in tumor bearing mice, they may represent more than 20% of all splenocytes [238]. In both patients and experimental animals,

MDSCs have been shown to be mobilized from bone marrow in response to multiple tumor-derived factors such as Bv8 and endocrine gland-derived VEGF [244, 245]. Their recruitment to tumors is mediated by chemotactic factors like CCL2/MCP-1, CXCL12/SDF-1 $\alpha$ , CXCL5 and KIT ligand [246]. Although MDSCs are able to differentiate into mature myeloid cells upon exposure to appropriate stimuli, their differentiation is blocked by tumor cell conditioned media *in vitro* or in a tumor-bearing host *in vivo* [247]. These immature myeloid cells potently suppress maturation and anti-tumor activation of dendritic cells, T cells and natural killer cells, a phenotype that provides the most effective way of identifying MDSC [248]. Hence, injection of tumor cells in combination with CD11b<sup>+</sup>Gr1<sup>+</sup> cells in mice prompt tumor growth [249]. Accordingly, depletion of Gr1<sup>+</sup> cells in tumor-bearing mice leads to delayed tumor growth, suggesting MDSC as potential targets for anti-cancer therapy [250-256]. A report by Youn and colleagues indicated that CD11b<sup>+</sup>Gr1<sup>+</sup> cells from naïve tumor-free mice are not immune suppressive [243]. However, it is not yet fully known why CD11b<sup>+</sup>Gr1<sup>+</sup> cells isolated from tumor-free and tumor-bearing animals exhibit different functions. A recent study suggested a HIF-1 $\alpha$  mediated regulatory mechanism for the biological dichotomy displayed by MDSCs within the TME. These researchers demonstrated that splenic MDSCs of tumor bearing animals cause ROS mediated antigen-specific T cell unresponsiveness, whereas intratumoral MDSCs with similar morphology and phenotype suppress both antigen specific and nonspecific T cell function through elevated NO levels and arginase I production [257].

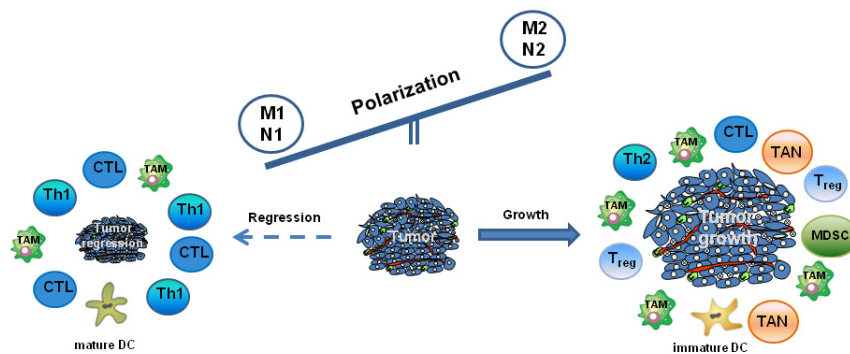
### ***Neutrophils***

Neutrophils are short-lived white blood cells derived from bone marrow myeloid progenitors. During infection-related immune responses neutrophils are among the first cells to arrive at the site of infection where they release chemokines and proteases that trigger the recruitment of both innate and adaptive immune effector cells. Neutrophils also release cytotoxic mediators, including reactive oxygen species, membrane-perforating agents, proteases and soluble mediators such as interferons, TNF- $\alpha$  and IL-1 $\beta$ , suggesting their potential anti-tumor activity [72, 258, 259]. Generally, in most tumors low numbers of neutrophils are found. Both cancer cells and cells of the TME actively recruit neutrophils by means of

secreted chemotactic factors, in particular G-CSF, GM-CSF, CXCL2/MIP-2 $\alpha$ , CCL3/MIP-1 $\alpha$ , CXCL5/LIX and CXCL1/KC. Upon recruitment to the tumor site, neutrophils can assume tumor growth-stimulatory or -inhibitory functions [71]. In human tumors, an increased density of tumor-infiltrating neutrophils was found to correlate with a poor prognosis in patients with adenocarcinoma and metastatic melanoma, whereas in few cases like gastric carcinoma neutrophil infiltration was linked to beneficial disease outcome [260-262]. This discrepancy is probably related with the degree of neutrophil recruitment and their differential activation, depending on the intratumoral cytokine microenvironment in which they reside. Similar to TAMs, the functional status of tumor associated neutrophils (TANs) regulates their ability to express an anti-tumor potential. Accumulating experimental and clinical evidence also confirms that neutrophils can polarize in a type I or type II direction in tumors. Recently, Fridlender and colleagues characterized N1- and N2-polarized phenotypes of TANs, similar as described for TAMs [263]. In lung and mesothelioma tumor models, TANs were shown to acquire a N2-phenotype. The pro-tumorigenic activities of N2-TANs include increased production of immunosuppressive cytokines and reduced cytotoxic activity. This pro-tumor phenotype of neutrophils was found to be induced and maintained by TGF- $\beta$  [264]. N1-polarized neutrophils exert anti-tumor activities indirectly as well by promoting recruitment and activation of CD8<sup>+</sup> T cells. In addition to induction of the anti-tumor N1-polarization, blocking of the TGF- $\beta$  pathway caused increased recruitment of Ly6G<sup>+</sup> neutrophils in tumors [263]. This finding is consistent with studies that demonstrated an enhanced influx of myeloid cells into mammary carcinomas deficient in type-II TGF- $\beta$  receptor [249]. Further, it was shown that abrogation of TGF- $\beta$  signaling in human breast cancer cells enhanced the production of the neutrophil chemoattractants CXCL1 and CXCL5 [265]. Apparently, TGF- $\beta$  is one of the major players in regulating neutrophil recruitment and activation in the TME. A recent study suggested that constitutive expression of IFN- $\beta$  counteracts the cancer-supportive function of neutrophils by inhibiting expression of genes encoding pro-angiogenic and homing factors in these cells [266].

In summary, as shown in Fig. 4, the types of cells infiltrating a tumor microenvironment and their state of polarization control the fate of a growing

tumor. Type-1 polarized macrophages and neutrophils, mature DCs and mature T cells with  $T_H1$  activity create a tumor growth inhibitory environment. At the opposite, type-2 polarized macrophages and neutrophils, immature DCs, MDSC, regulatory T cells and  $T_H2$  T cells promote angiogenesis and tumor growth.



**Figure 4:** Immune cells infiltrating a tumor regulate tumor growth, progression and metastatic dissemination. Depending on the state of polarization of tumor associated immune cells tumor development is suppressed or enhanced. Tumor regression is associated with M1-macrophages, N1-neutrophils, mature dendritic cells (DCs) and mature T cells with  $T_H1$ -activity. In contrast, tumor growth is facilitated *via* immune-suppression and induced angiogenesis, M2-macrophages, N2-neutrophils, immature DCs and plasmacytoid DCs, myeloid-derived suppressor cells (MDSCs), regulatory T cells and a low frequency of  $T_H2$  activated CD4 and CD8 effector T cells.

## Therapies Aiming at Components of the TME

### *Cell-based Therapies*

Based on a vast amount of clinical and pre-clinical evidence, our current knowledge suggests that therapeutic targeting should not only be aimed at the malignant cancer cells, but also at the components of the TME to effectively inhibit tumor growth. Thus, interference with microenvironmental growth support is becoming appreciated as an attractive therapeutic strategy [267, 268]. As a key



component of the TME, the tumor promoting properties of myeloid cells render these cell types as valuable tools and targets for therapeutic interventions. One of the first strategies that have been explored since many years is the adoptive immunotherapy which consists in the transfusion of host derived and *in vitro* activated or engineered lymphoid cells. Transfer of tumor infiltrating leukocytes (TIL) to tumor bearing hosts mediates antitumor responses and several myeloid cell subpopulations were found to be suitable for use in adoptive immunotherapy. Lymphocytes treated with IL-2 give rise to lymphokine activated killer (LAK) cells that have the ability to lyse malignant but not normal cells. Clinical studies in patients with advanced cancer revealed that treatment with IL-2 alone or in combination with LAK cells mediate complete or partial regression of cancer, predominantly melanomas [269-273]. Other methodologies either used combinations of lymphokines, such as TNF- $\alpha$  or interferons in conjunction with IL-2 or gene therapy approaches to further improve the effects of adaptive immunotherapy [274-278]. Although the significance of MHC class I-restricted cytotoxic T lymphocytes (CTLs) as effectors of anti-tumor immunity has widely been demonstrated, most human tumors lack MHC-I expression or are inadequately differentiated and poorly immunogenic, a culprit that limits successful T-cell based tumor-specific immunotherapy [279]. In another cell-based therapy approach efficient tumor-specific effector and memory T cells are induced through therapeutic vaccination. Such vaccines follow two purposes, namely priming antigen-specific T cells and reprogramming memory T cells by transforming them from the immunosuppressive to the immunostimulating and cytotoxic phenotype. Dendritic cells (DCs) are very potent antigen presenting cells and thus essential in generation of immune responses, and they therefore represent valuable targets and vectors for cancer vaccination [280-293].

### ***Therapies Aimed at TAMs***

Based on the M1 *versus* M2 paradigm of macrophage polarization, inhibition of M2- and activation of M1-inducing signals was suggested as a potential strategy to re-establish the anti-tumor function of macrophages [294]. Indeed, pharmacological skewing of macrophage polarization from the M2- to M1-phenotype is able to induce an anti-tumor activity. Co-administration of the macrophage chemoattractant CCL16 with a CpG oligonucleotide and an anti-IL-

10 receptor antibody was shown to skew M2-TAMs to M1-TAMs that triggered an innate response resulting in the regression of pre-established tumors [295]. Similarly, combination of an anti-CD40 antibody with CpG oligonucleotides and multidrug chemotherapy induced antitumor effects by TAM polarization [296]. Considering the central role the statins play in myeloid cell polarization, members of the STAT family of transcription factors are valuable targets for the modulation of myeloid cells. To this end, tumor bearing STAT6<sup>-/-</sup> mice were shown to display an M1-TAM phenotype and to reject a spontaneously growing mammary carcinoma [297, 298]. Accordingly, it was found that the SHIP1 phosphatase plays an important role in macrophage re-programming. Mice deficient in SHIP1 displayed a skewed development toward M2-TAM and thus pharmacological modulators of this phosphatase could be developed [299]. More recently, a host-derived factor, histidine-rich glycoprotein (HRG) was reported to promote M1 polarization of TAMs by downregulation of PLGF [300].

Other approaches aim at the depletion of TAMs, either by blocking vital functions of the cells or by their physical depletion. In various models it was shown that blockade of the macrophage specific colony-stimulating factor 1 (CSF-1) or its receptor CSF-1R suppresses macrophage infiltration and tumor growth [301-303].

The physical (pharmacological) depletion of macrophages from organs of the MPS using the bisphosphonate clodronate encapsulated in liposomes (Clodrolip) has become an important, reliable and widely used method to study not only the role of macrophages in the immune system and in inflammatory processes but also in tumor growth and metastasis [304-311].

### **Bisphosphonates**

Bisphosphonates (BPs) are inorganic pyrophosphate analogs (PPi) that effectively inhibit osteoclastic bone resorption and are widely used to treat metabolic bone diseases, such as postmenopausal osteoporosis [312], Paget's disease [313], tumor associated osteolysis [314] and to prevent bone metastasis [315]. The high affinity of the BPs for the calcium component of the bone matrix hydroxyapatite is the cause of the bone-specificity of these compounds. Organ distribution studies demonstrated that BPs are mainly localized in newly formed bones and internalized by the bone resorbing osteoclasts where they inhibit their activity [316]. Due to their high affinity

for bone matrix, systemic availability of BPs is rather low with the exception of a transient raise of plasma levels in the post-administration period [317].

Based on their chemical structure BPs can be divided into two distinct pharmacological classes; the nitrogen-containing bisphosphonates, (N-BPs, *e.g.* zoledronate) and the first-generation BPs, the non-nitrogen-containing bisphosphonates (non-N-BPs, *e.g.* clodronate, see Fig. 5) that chemically resemble pyrophosphate (PPi). Pyrophosphate has a P–O–P structure, whereas the BPs have a P–C–P structure where the central oxygen atom is replaced by a carbon atom. The most important first generation non-N-BP bisphosphonate, clodronate has R<sup>1</sup> and R<sup>2</sup> side chains with two chlorine atoms, whereas the N-BP zoledronate carries a hydroxyl group on R<sup>1</sup> and an imidazolyl group on R<sup>2</sup>. BPs containing nitrogen atoms in the R<sup>2</sup> side-chain like zoledronate are significantly more potent than non-N-BPs [317]. The mechanism of action of the BPs differs according to their chemical structure. After cellular uptake, non-N-BPs are metabolized to cytotoxic analogs of adenosine triphosphate (ATP) causing cell death by apoptosis [318]. The N-BPs exert their effects mainly by inhibiting a key enzyme in the mevalonate pathway, the farnesyl pyrophosphate synthase (FPP synthase), thereby preventing the synthesis of isoprenoid compounds that are essential for the post-translational modification of small guanosine triphosphate (GTP)-binding proteins such as Rab, Rho, and Rac [319]. Recent studies revealed that N-BPs can also induce formation of a new pro-apoptotic ATP analog that induces mitochondria-mediated apoptosis [320].

Although the most effective N-BP zoledronate has originally been developed to inhibit osteoclast mediated bone resorption, the anti-cancer effects of this compound are currently being evaluated. In this context zoledronate is used as adjuvant therapy to inhibit local bone destruction by tumors and to prevent or delay metastasis to bone [321-323]. Moreover, zoledronate has demonstrated significant clinical benefits in patients with metastatic prostate and lung cancer [322, 324, 325]. Zoledronate exerts these anti-tumorigenic activities directly on cancer cells by modulating their tumorigenic properties and indirectly on stromal cells by changing their tumor-promoting properties. One of the major anti-tumor effects of zoledronate is the induction of apoptosis but the drug also interferes with migratory and invasive properties of tumor cells [326-329] and with angiogenesis [330-332]. In addition to

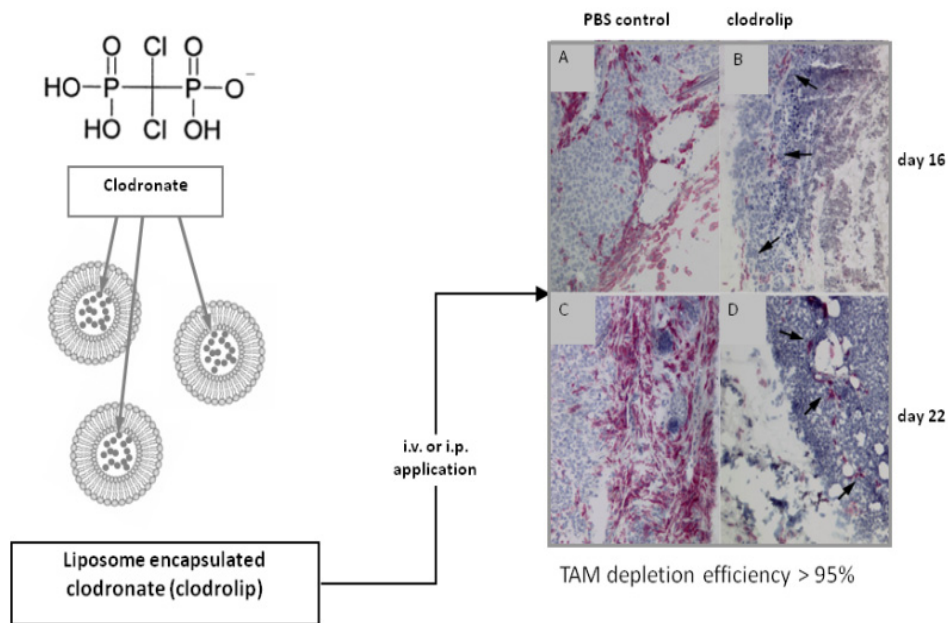
its pharmacological effects, zoledronate has immune modulatory activities that include stimulation of proliferation and activation of the V $\gamma$ 9V $\delta$ 2 subset of  $\gamma\delta$ T cells. T cells expressing the V $\gamma$ 9V $\delta$ 2 T cell receptor play a significant role in immune surveillance and defense [333-335]. These cells have the ability to recognize and kill tumor cells in an MHC-independent manner, suggesting their potential utility in the elimination of cancer cells with poor antigen presentation capacity [336]. Several pre-clinical studies have shown that V $\gamma$ 9V $\delta$ 2 T cells expanded *in vitro* sustain their anti-cancer activity upon adoptive transfer into mice transplanted with various human cancer cells along with zoledronate treatment [337-340]. Clinical studies also demonstrated expansion and activation of V $\gamma$ 9V $\delta$ 2 T cells to a subset of IFN- $\gamma$  producing effector T cells in patients treated with zoledronate, either alone or in combination with IL-2 [337, 341, 342]. Besides cancer cells, monocytes treated with zoledronate were also shown to stimulate proliferation and cytotoxic activation of human V $\gamma$ 9V $\delta$ 2 T cells. Notably, activation of  $\gamma\delta$ T cells requires cell-to-cell contact with zoledronate treated tumor cells or monocytes [343]. Among several growth factors, TGF- $\beta$  is known as the most abundant cytokine in bone and considered as the main bone-derived factor responsible for driving this vicious cycle of bone metastasis [344]. Activated TGF- $\beta$  is released from mineralized bone matrix and in turn it induces production of tumor-derived osteolytic factors [345-349].

These and other data suggest that modulation of bone derived factors like TGF- $\beta$  might also be a possible mechanism responsible for the anti-tumor activity of zoledronate. All together, pre-clinical and clinical studies suggest multifaceted anti-cancer effects of zoledronate in different tumor types. In addition, clinical studies showed that zoledronate prolongs disease-free survival in cancer patients [350]. However, the identification of new cellular targets and further elucidation of the cellular and molecular mechanisms by which zoledronate mediates anti-tumor effects will be useful in the design of new therapeutic strategies to modulate and potentiate the anti-tumor effects of this compound.

### **Depletion of TAMs with Clodronate-Liposomes (Clodrolip)**

Exploiting the anti-tumor properties of the bisphosphonates and in particular clodronate-liposomes, we examined the possibility whether depletion of TAMs would inhibit tumor angiogenesis and tumor growth. In our experiments, we showed for the first time that treatment of tumor bearing mice with Clodrolip as

single therapy in comparison to free clodronate and in combination with anti-VEGF single chain fragment antibodies, resulted in drastic tumor growth inhibition and exhaustion of TAM cell populations [351] (Fig. 5).



**Figure 5:** Scheme of clodronate and encapsulated clodronate (Clodrolip) in small unilamellar liposomes. Example of macrophage depletion efficiency of Clodrolip given by the i.p. or i.v. route in A673 rhabdomyosarcoma tumors and analyzed by immunohistochemistry at day16 and 22 after tumor cell inoculation. Cells stained in red are F4/80<sup>+</sup> positive TAM. Adapted from [351].

Based on our findings, summarized in Table 1, several follow-up studies using Clodrolip: or other clodronate-liposomes confirmed the therapeutic validity of the TAM depletion method. In fact, clodronate- or other bisphosphonate liposome-mediated macrophage depletion or modulation opens new opportunities to study the role of tumor infiltrating cells and combined with anti-angiogenic or cytotoxic therapies TAM depletion represents a promising new approach of high clinical potential.

**Table 1:** Effects on Tumor Growth by Clodrolip mediated Depletion of TAMs in select preclinical Tumor Models

Models (Tumor cells, treatments)	Effects of TAM Depletion	Notes	Ref.
Breast cancer (MDA-MD-231, MVT-1) overexpr. S100A7	Inhibition of the effects of S100A7 induction on tumor growth and angiogenesis in orthotopic models.	S100A7 is overexpressed in invasive estrogen receptor $\alpha$ -negative breast cancer and activates pro-inflammatory pathways.	[363]
F9 teratocarcinoma in Sv129 mice	Depletion correlated positively between TAM-densities and mesenchymal marker expression.	TAMs induce EMT through TGF- $\beta$ signaling and $\beta$ -catenin activation. Clinical relevance is shown in non-small cell lung cancer (NSCLC).	[98]
Lung cancer induced by urethane in FVB mice	Alveolar macrophage depletion reduced number and size of lung tumors and inhibited angiogenesis.	Urethane treatment induced M1 macrophages (first 2-3 wks) followed by M2 macrophages by week 6.	[364]
Bladder cancer (MBT-2)	TAM depletion by Clodrolip or VEGF block inhibited lymphangiogenesis and lymph node metastases but not growth of orthotopic primary tumors.	Massive lymphangiogenesis and TAM infiltration in primary tumor and metastasis in lymph nodes.	[365]
Liver cancer (Hepa 1-6)	TAM depletion reduced tumor growth in s.c. and orthotopic liver tumors.	TAMs express MHC-II <sup>high</sup> at early stages and pro-tumorigenic MHC-II <sup>low</sup> during tumor growth.	[366]
Hepatocellular cancer xenografts, sorafenib treatm.	TAM depletion or zoledronate (zol) + sorafenib inhibited tumor progression, angiogenesis and lung metastasis.	Combined therapy with zol or TAM depletion enhanced the effect of sorafenib. Zol was more effective than Clodrolip.	[354]
Melanomas in C57BL/6 and TNFR1,2 <sup>-/-</sup> , TNF <sup>-/-</sup> mice, local radiation therapy	TAM depletion before radiotherapy increased antitumor effects of ionizing radiation in a TNF $\alpha$ dependent way.	Treatment with a TNF receptor fusion protein (Enbrel) showed that macrophage mediated radioresistance required intact TNF $\alpha$ signaling.	[367]
Colon adenocarcinoma (MC38), mammary tumors (AT-3, 4T1.2) targeting DR4 and DR5 with mab MD5-1	MD5-1 mab treatment inhibited tumor growth by TRAIL-R dependent tumor cell apoptosis. Clodrolip treatment enhanced efficacy of MD5-1.	Ab-mediated targeting of DR5 triggers tumor cell apoptosis in a B cell-dependent manner. Contribution of NK cells, CD11b <sup>+</sup> cells, and macrophages to the antitumor effects of MD5-1.	[368]
Colon adenocarcinoma (MC38), renal cell carcinoma (Renca) combination of CpG 1826 with a CD137 specific T-cell antibody	CpG plus anti-CD137 caused tumor regression. TAM depletion enhanced therapy leading to tumor rejection in 100% of mice.	This study provides support for the use of a novel combination of immunomodulatory agents stimulating multiple facets of immunity for the effective immunotherapy of cancer.	[369]
HPV16 E6- and E7-expressing TC-1 mouse tumor model	TAM depletion inhibited tumor growth and stimulated HPV16 tumor infiltration by virus-specific CD8 lymphocytes.	M2-like macrophages infiltrate HPV16-associated tumors causing suppression of antitumor T-cell response.	[370]

Table 1: contd.....

Ovarian carcinoma (MDAH-2774, SKOV-3, OVCAR3) in nude mice	Depletion of macrophages by Clodrolip markedly reduced lymph-angiogenesis.	Blockade of VEGF/VEGFR signaling or depletion of macrophages reduced lymph-angiogenesis.	[371]
Lung cancer (HARA-B) injected into the left cardiac ventricle of mice	Clodrolip significantly reduced the number of macrophages in tumors and osteoclasts in bone marrow.	Clodrolip exerted antimetastatic effects in both bone and muscle.	[372]
Rat glioma (D74/HveC), oncolytic viruses (OV) injected into intracranial gliomas	Depletion of TAMs enhanced intratumoral OV spread.	CD163 <sup>+</sup> macrophages infiltrated the tumor. TAM depletion during OV delivery helps intratumoral propagation and persistence of virus, rendering more efficient therapy.	[373]
Murine teratocarcinoma (F9) and human rhabdomyo-sarcoma (A673)	75 - >92% TAM depletion with Clodrolip. Combination therapy of Clodrolip plus a VEGF-neutralizing antibody was most effective.	First demonstration of TAM depletion. Tumor inhibition was accompanied by drastic anti-angiogenic effects. CD11c <sup>+</sup> TADCs were also depleted by Clodrolip or antibody treatment.	[351]

Zoledronate has also been encapsulated in liposomes either targeted to the folate receptor expressed on tumor cells showing cytotoxic activity [352] or in (polyethylene)glycol liposomes tested in murine models of human prostate cancer and multiple myeloma where the liposomal formulation proved to be more cytotoxic compared to the free drug [353]. Another study showed that treatment of hepatocellular carcinoma xenografts with sorafenib, a multikinase inhibitor, was markedly enhanced by concomitant depletion of macrophages by clodrolip or free zoledronate [354].

Drug formulations with liposomes are also used to target other cell types in the TME including the tumor cells themselves [355-357]. The high vascular permeability of tumor blood vessels can be exploited for increased accumulation and retention of macromolecules and liposomes in the tumor tissue. Passive targeting of long circulating liposomes to tumors with liposomal doxorubicin was one of the first clinically approved drug application with enhanced activity and reduced toxicity [358] and several other drugs are currently being evaluated as liposome formulations.

Extravasation and accumulation of liposomal drugs within the TME occurs because small liposomes are able to penetrate through the leaky vasculature into

the tumor tissue where they are taken up by cells such as macrophages or dendritic cells or where they release the encapsulated payload into the ECM. In an earlier mouse tumor model study, we demonstrated the specificity and cytotoxicity of immunoliposomes that were targeted against the ED-B isoform of fibronectin which is uniquely expressed in the ECM of solid tumors [359].

Other examples of target-specific immunoliposomes are doxorubicin loaded anti-HER2 immunoliposomes [360] or epidermal growth factor receptor (EGFR)-targeted immunoliposomes [361]. In summary, nanocarriers, most notably liposomes, possess a great potential for the delivery of cytotoxic drugs or immunomodulating agents to the TME and to metastases [362].

### ***Therapies Aimed at TANs***

To date, the anti-tumor potential of neutrophils has received scarce attention and their functions as effective weapons against cancer are still not fully exploited. Yet, recently gathered evidence indicates that under appropriate stimulation neutrophils reveal very powerful tumor-inhibitory properties. As neutrophils in tumor-bearing hosts have impaired cytotoxic activity, the development of methods that stimulate recruitment and anti-tumorigenic activation within a TME can be exploited as new therapeutic opportunities.

Early studies with cytokine or chemokine gene transfected mammary adenocarcinomas in syngeneic tumor models indicated that nonspecific mechanisms, mostly supported by neutrophil functions, had much greater therapeutic power than those elicited by specific immunity [374-376]. For example, local or systemic administration of rIL-12 in mice bearing subcutaneous mammary carcinoma resulted in a rapid influx of neutrophils with high cytotoxic potential and anti-angiogenic function [377]. TGF- $\beta$  has been defined as a major functional regulator of neutrophils. Specifically in tumors, TGF- $\beta$  has been found to drive the pro-tumorigenic polarization of neutrophils. Thus, inhibition of TGF- $\beta$  signaling offers a means to manipulate neutrophil polarization by shifting N2-TANs to tumor growth inhibiting N1-neutrophils. Additionally, TGF- $\beta$  receptor blockage in tumor bearing mice was shown to induce the activation of CD11b<sup>+</sup>Ly6G<sup>+</sup> neutrophils that resulted in a significant tumor growth delay [263].



### ***Therapies Aimed at CAFs and Mast Cells***

CAFs represent another therapeutic target within the TME. However, due to a lack of compounds that specifically target this cell population, such strategies have not been widely used in the clinical setting. The most studied target molecule is the fibroblast activation protein (FAP) that is selectively expressed on stromal fibroblasts or on CAFs. FAP is a membrane-bound serine protease of the prolyl oligopeptidase family with distinctive endopeptidase activity and with low or undetectable expression in fibroblasts of normal tissues [378, 379]. In a preclinical vaccine approach, it was shown that immunological targeting of FAP can elicit protective immunity. A DNA vaccine directed against FAP suppressed primary tumor growth and pulmonary metastases primarily through CD8<sup>+</sup> T-cell-mediated killing in tumor-bearing mice [380].

Mast cells (MC) play an essential role as effector cells in allergy but they also contribute to tumor development. Activated MC located in the TME release angiogenic and tumor growth stimulating factors [124, 125, 381]. Recent findings indicate that tumor-associated mast cells might represent valuable targets for therapeutic interventions, most notably to kinase inhibitors as c-Kit [128, 382].

### ***Other Therapies: Anti-angiogenic Therapies, Antibodies, Antibody-drug Conjugates, Cytokines, Gene Therapeutics***

The major non-cellular therapies aiming at specific targets in the TME including antibodies and small molecule inhibitors are summarized in the following section.

#### **Anti-Angiogenic Therapies**

The vascular endothelial growth factor (VEGF) proteins are key regulators of normal and tumor angiogenesis and they are therefore extensively studied as therapeutic targets [383]. Antibodies and fusion proteins targeting VEGF are the clinically approved bevacizumab (Avastin, Genentech) [384-386], r84 (AT001, Affitech AS), a human antibody which inhibits VEGF from binding to the VEGF-receptor-2 and VEGF-trap which is a fusion protein containing the binding domains of the VEGF-receptors 1 and 2 fused to the human IgG Fc region [383]. However, anti-angiogenic therapies may be compromised by the finding that myeloid CD11b<sup>+</sup>Gr1<sup>+</sup> cells which contribute to tumor angiogenesis render tumors

refractory to angiogenic blockade by VEGF antibodies. This CD11b<sup>+</sup>Gr1<sup>+</sup>-mediated effect is driven by the protein Bv8 which, in turn, is up-regulated by G-CSF. Thus, G-CSF may contribute to tumor refractoriness to anti-angiogenic therapies [66, 245, 387]. Different anti-angiogenic compounds such as small molecule tyrosine kinase inhibitors TKIs (*e.g.* sunitinib, sorafenib, imatinib, dasatinib, nilotinib and the proteasome inhibitor bortezomib) and other immunomodulatory drugs targeting VEGF or other pathways seem to be capable of modulating immune responses, in a positive as well as a harmful manner. Recent studies focused not only on their direct anti-tumor responses, but also on their influence on the TME, as well as on their effects on malignant and healthy cells. Thus, for an optimal clinical anti-cancer treatment, a better understanding of these immunomodulatory effects is essential [388].

Unfortunately, the initial expectations and optimism for therapies with anti-angiogenic drugs targeting the VEGF signaling pathway were impeded by the limited clinical benefits. New data indicate that the unique characteristics of the tumor vasculature within the TME may hold the key for successful novel anti-angiogenic therapies. The molecular and cellular alterations that maintain aberrant tumor angiogenesis represents novel targets for improving current anti-angiogenic strategies [389]. This so-called "vascular normalization" is characterized by attenuation of hyperpermeability, increased vascular pericyte coverage and a normalized basement membrane, resulting in the reduction of tumor hypoxia and interstitial fluid pressure. This improves the metabolic profile of the TME and the delivery and efficacy of therapeutics. Novel genetic and pharmacological approaches characterized key regulators of vascular normalization such as proteins that regulate tissue oxygen sensing and vessel maturation [45, 390-392].

### **Antibodies**

Monoclonal antibodies have considerably modified the therapy concepts in clinical oncology. Antibodies and smaller fragments such as antigen-binding fragments (Fab), single chain variable fragments (scFv) and smaller molecules are produced by recombinant technologies [393]. Antibodies possess several clinically relevant mechanisms of action. They can manipulate tumor-related signaling and various antibodies show immunomodulatory properties and, by

activation or inhibition of the immune system, they can induce antitumor immune responses [394, 395]. Specifically, Fc-receptor expressing immune cells mediate the killing of tumor cells by mAbs. Stimulation of these immune effector cells therefore represents an interesting strategy to improve the therapeutic efficacy of mAbs. The stimulation of natural killer cells,  $\gamma\delta$ T cells, macrophages, or dendritic cells can be used to enhance antibody-dependent cellular cytotoxicity, phagocytosis or tumor vaccine effects [396]. Besides supporting development and strengthening of the adaptive immunity, therapeutic antibodies are able to trigger early anti-tumor events such as receptor blockade, cytostasis, apoptosis, complement-dependent cytotoxicity and/or antibody-dependent cytotoxicity [397-399]. Bispecific antibodies are used to mount and sustain tumor-specific cellular responses or in radioimmunotherapy to improve target binding, selectivity, and efficacy [400-403]. A widely studied target is the cytotoxic T-lymphocyte-associated antigen CTLA-4, also called CD152, which regulates T-cell activation. Antibodies that block the interaction of CTLA-4 with its ligands B7.1 and B7.2 enhance immune responses, including antitumor immunity. The recently FDA-approved anti-CTLA-4 antibody ipilimumab (Yervoy) and tremelimumab are the most advanced antibodies for the treatment of metastatic melanoma [404-408].

### **Antibody-Drug Conjugates**

The development of antibody-drug or antibody-enzyme immunoconjugates for a more specifically targeted and efficient delivery of active compounds to target tumor cells has been followed since more than three decades. Several immunoconjugates, particularly those that incorporate internalizing antibodies and tumor-selective linkers have demonstrated impressive activity in preclinical models. Immunoconjugates that deliver doxorubicin, maytansine and calicheamicin were among the first to be evaluated in clinical trials [409,410]. The immunoconjugate gemtuzumab ozogamicin (Mylotarg, CMA-676), a calicheamicin conjugate that targets CD33, has been approved by the Food and Drug Administration (FDA) in 2000 for treatment of acute myelogenous leukemia (AML). Although gemtuzumab ozogamicin improved survival in a subset of AML patients when combined with standard chemotherapy, the drug was recently withdrawn by the FDA due to safety concerns [411]. However, the cytotoxic activity of the immunoconjugate confirms that CD33 remains a possible

therapeutic target for AML. In recent years, significant progress owing to the optimization of several parameters, including mAb specificity, drug potency, linker technology, and the stoichiometry and molecular sites of attachment of conjugated drugs has been made. These developments have led to an increase of conjugates being tested clinically, three of which are currently in late stage clinical trials: brentuximab vedotin (SGN-35) for Hodgkin lymphoma; trastuzumab-DM1 for breast cancer and inotuzumab ozogamicin for non-Hodgkin lymphoma [412]. The immunoconjugate trastuzumab emtansine (T-DM1) is a tumor-activated prodrug resulting from the conjugation of the cytotoxic and antimitotic maytansine derivative DM1 with the humanized anti-HER2 mAb trastuzumab which has been used for the treatment of breast cancer for over 10 years. The maytansinoids bind microtubules in a manner similar to the vinca alkaloids, but they block mitosis 20 to 100-fold more potently. Clinically, trastuzumab emtansine exhibited efficacy in patients with HER2<sup>+</sup> metastatic breast cancer. Furthermore, preclinical studies have reported that trastuzumab emtansine potentiates the effect of several chemotherapeutic agents (carboplatin, 5-fluorouracil and docetaxel), other antibodies as well as receptor tyrosine kinase and PI3K inhibitors. Many of these combinations are currently investigated in humans [413]. Phase I and II trials of T-DM1 as single agent and in combination with paclitaxel, docetaxel and pertuzumab have shown clinical activity and favorable safety profiles in HER2<sup>+</sup> metastatic breast cancer patients. Additional combinations of T-DM1 with antitumor drugs and additional disease settings such as early-stage HER2<sup>+</sup> breast cancer are also under investigation [414, 415]. Brentuximab vedotin (SGN-35) is a novel antibody-drug conjugate consisting of the anti-CD30 antibody cAC10 conjugated by a protease-cleavable linker to monomethyl-auristatin E, a potent microtubule blocking agent. In phase II trials, response rates of 75% in relapsed/refractory Hodgkin's lymphoma and 87% in relapsed/refractory systemic anaplastic large-cell lymphoma were recently reported. The impressive response rates and limited toxicity of brentuximab vedotin (SGN-35) are very promising for relapsed/refractory patients with few treatment options. In 2011, brentuximab vedotin was approved in the US for the treatment of Hodgkin lymphoma after failure of autologous stem cell transplant (ASCT) or after failure of multiagent chemotherapy regimens in ASCT-ineligible candidates and for the treatment of systemic anaplastic large-cell lymphoma after

failure of prior multiagent chemotherapy regimens [416]. The efficacy of brentuximab vedotin in other CD30 positive lymphomas is currently under investigation [417-419].

Radioimmunotherapy (RAIT) of non-Hodgkin lymphoma (NHL), a disease that is radiosensitive as well as readily accessible to the antibody conjugates using directly labeled MAbs is of current interest after approval of the radiolabeled anti-CD20 MAbs  $^{131}\text{I}$ -tositumomab and  $^{90}\text{Y}$ -ibritumomab tiuxetan [420]. The high efficacy of RAIT was illustrated with the nearly 100% overall response rate obtained in a clinical trial using an investigational radiolabeled anti-CD22 MAb,  $^{90}\text{Y}$ -epratuzumab. The advantage of pretargeted RAIT over directly labeled MAbs is continuing to be validated in preclinical models of lymphomas and solid tumors. The advantages of combining RAIT with radiation sensitizers, with immunotherapy or drug conjugates targeting different antigens are being studied clinically and preclinically [421].

Comprehensive and updated lists of therapeutic antibodies and conjugates including their status of clinical use can be found at the website of the international ImMunoGeneTics information system (IMGT) (<http://www.imgt.org/mAb-DB/index>) and in the "Marketed therapeutic antibodies compendium" [422].

Antibody-enzyme conjugates are directed at tumor-associated antigens to achieve site-specific activation of prodrugs to potent cytotoxic drugs. This "antibody-directed enzyme prodrug therapy" (ADEPT) technology has attracted considerable interest since the concept was first described in 1987 [423]. A particular advantage of the ADEPT approach is that it may allow the use of extremely toxic and potent agents at very low concentrations. The principle of ADEPT therapy is to use a tumor-associated antigen specific antibody to target an enzyme to tumor cells. The enzyme should be retained in the tumor after clearance from blood and normal tissues. A nontoxic prodrug, which is a substrate for the enzyme is then applied and by cleaving of the enzyme-prodrug complex a potent cytotoxic agent is generated in the tumor tissue [424-426]. More recently, complementing the ADEPT technology, the promising approaches GDEPT (gene-directed enzyme prodrug therapy) and PMT (prodrug monotherapy) have been

developed. GDEPT and PMT allow a selective release of cytotoxic agents from non-toxic prodrugs at the tumor site either by enzyme encoding genes or by exploiting physiological and metabolic aberrations in cancerous tissue [427].

### **Chemokines and Cytokines**

As mentioned, the TME contains chemokines and cytokines which are vital factors for the regulation of tumor growth, invasion and metastasis. Beyond activating tumor vascularization, infiltrating myeloid cells also promote tumor growth by creating a microenvironment that is rich in growth factors and pro-inflammatory cytokines that stimulate proliferation and survival of neoplastic cells [26, 108, 428]. Chemokines/cytokines and their receptors represent potential targets for therapeutic intervention, either with antibodies or small molecule antagonists. On the other hand, due to the complexity of the TME, and the large number of chemokines/cytokines and receptors that are also expressed by normal cells, issues remain regarding the targetability of inhibitors and whether the redundancy of the system will compensate an inactivated chemokine/cytokine or its receptor [429, 430].

The most studied cytokines for cancer immunotherapy are the interleukins (IL). *Ex vivo* treatment of lymphocytes with IL-2 gives rise to lymphokine activated killer (LAK) cells and clinical studies in patients with advanced cancer showed that treatment with IL-2 alone or in combination with LAK cells mediate complete or partial regression of cancer, predominantly melanomas and renal cell carcinoma [269-273, 431]. More recently, several new interleukins, namely IL-12 in ovarian cancer [432, 433], IL-15 in various experimental tumor models [434], IL-18 in metastatic melanoma [435] and IL-21 in early phase renal cell carcinoma and melanoma clinical trials [436] have been characterized that have considerable promise for future immunotherapy [437]. IL-15 binds to its specific receptor, IL-15R $\alpha$ , which is expressed on dendritic cells, monocytes and macrophages. IL-15 induces differentiation and proliferation of T, B and natural killer cells. It also enhances the cytolytic activity of CD8<sup>+</sup> T cells and induces CD8<sup>+</sup>CD44<sup>high</sup> memory T cells. Furthermore, IL-15 stimulates cell differentiation and immunoglobulin synthesis by B cells and induces maturation of dendritic cells [438]. IL-18 functions mainly as a co-stimulatory cytokine and its optimal

efficacy may be obtained in combination with other immunostimulatory therapeutics. Finally, IL-27 which is a member of the IL-6/IL-12 heterodimeric cytokine family acts on naive CD4<sup>+</sup> T cells and plays pivotal roles as a proinflammatory cytokine and generation of CTLs. Recent studies revealed that IL-27 plays an important role in CD8<sup>+</sup> T cells as well [439].

Lastly, the interferons (IFN) are cytokines with a long history of use as immunotherapeutic drugs. The initial use of interferons in cancer therapy was based on their growth inhibitory and immunomodulatory effects, and more recently they have been shown to possess cytotoxic and anti-angiogenic properties. However, the availability of novel alternative therapies have replaced IFN therapy in many cancers [440]. Interferon- $\alpha$  (IFN- $\alpha$ ) is a type-I interferon which exerts multiple biological effects, including antiviral and antitumor activities in patients with defined types of cancer and viral diseases. A combined antiviral and antitumor effect of interferon is assumed to occur after surgical resection of hepatocellular carcinoma (HCC). Thus, IFN has a significant beneficial effect after curative treatment of HCC in terms of both survival and tumor recurrence [441]. Early preclinical studies demonstrated the importance of host immune mechanisms in the generation of long-lasting antitumor responses after type-I IFN treatment. More recent studies have revealed new immunomodulatory effects of IFN- $\alpha$ , including activities on T cells and dendritic cells. Overall, therapeutic strategies based on IFN- $\alpha$  include the use of these cytokines *in vivo* as immune adjuvants of cancer vaccines or their use *ex vivo* to generate DC-based vaccines and the combination of certain chemotherapy regimens with IFN- $\alpha$  [442-444]. Interferon- $\gamma$  (IFN- $\gamma$ ) is a cytokine that acts on cell-surface receptors, activating transcription of genes that increase tumor immunogenicity, disrupt proliferative mechanisms and inhibit tumor angiogenesis. Current investigations of IFN- $\gamma$  suggest that the cytokine has the potential to be used clinically in the treatment of brain tumors and as an adjuvant to other immunotherapeutic modalities [445]. The discovery of the interferon- $\lambda$  (IFN- $\lambda$ ) family has considerably contributed the understanding of the role interferons play in viral infections and in cancer. The IFN- $\lambda$  proteins, also termed interleukin-28 and -29, belong to the new type-III interferons. Type-III interferons are structurally similar to type-II IFN (IFN- $\gamma$ ) but functionally they are identical to

type-I IFN (IFN- $\alpha/\beta$ ). The IFN- $\lambda$ , have similar signaling pathways as IFN- $\alpha/\beta$  and they inhibit proliferation of tumor cells through cell cycle arrest or apoptosis. However, in contrast to type-I or -II IFNs, the response to type-III interferons is highly cell-type specific. Only epithelial cells and some immune cells respond to IFN- $\lambda$ . This particular pattern of response is controlled by the differential expression of the IFN- $\lambda$  receptor. Recently, the potent antitumor effects of IFN- $\lambda$  were demonstrated, opening new opportunities for IFN therapy [446, 447].

### **Gene Therapeutics**

Although the significance of MHC class I-restricted cytotoxic T lymphocytes (CTLs) as effectors of anti-tumor immunity has widely been demonstrated, most human tumors lack MHC-I expression or are inadequately differentiated and poorly immunogenic, a culprit that limits successful T-cell based tumor-specific immunotherapy. To overcome these disadvantages, the genetic modulation of T-lymphocytes using T cell receptor (TCR) transfer with tumor-specific TCR genes is an attractive strategy to generate anti-tumor responses, especially in large solid tumors. In this approach, the genes encoding a TCR specific for a defined antigen can be isolated from a T-cell clone and transduced to stimulated normal peripheral T-lymphocytes. This approach enables the redirection of the adaptive immune response against antigens of choice [448, 449]. A first demonstration of the feasibility of this method was given by Morgan and coworkers who demonstrated that it is possible to transduce normal autologous PBLs from metastatic melanoma patients with a MART1-specific TCR and generate large numbers of MART1-specific cells to be infused back to the patients [450]. However, several factors may hold back the clinical benefit of this approach, such as the type of cells to modulate, the vector configuration or the safety of the procedure.

The novel technique of RNA interference (RNAi), including small interfering RNA (siRNA), short hairpin RNA (shRNA) and microRNA (miRNA), mediate RNAi effects through the RNA inducible silencing complex RISC and represent attractive systems to be utilized as therapeutic tools [451]. Synthetic RNAs are nowadays widely used as tools for target validation and gene knock-down or knock-in. Presently, there is considerable interest for therapeutic applications of RNAi, particularly in areas of infectious disease and cancer. Preclinical data



demonstrate the efficacy of RNAi, for example knock-down of gene messages that are essential for tumor cell growth, metastasis, angiogenesis and chemoresistance, leading to anti-tumor effects. All types of RNA used for RNAi possess pharmacokinetic properties similar to single-stranded antisense oligonucleotides, but they are generally more robust than the latter [452, 453]. Despite all the potential of RNAi as a novel class of therapeutics, limited cellular uptake, low biological stability and unfavorable pharmacokinetic profiles are hampering their successful application in the clinic. Therefore, the translation of RNAi to the clinical setting is crucially dependent on the development of suitable delivery systems that improve their pharmacokinetic and biodistribution properties. Thus, delivery strategies for RNAi become the main hurdle that must be resolved prior to the full-scale clinical development of siRNA therapeutics [454-458]. As some examples, oncolytic adenoviral delivery of siRNA offers the potential benefits of restricted and renewable siRNA expression within the tumor microenvironment with an additive antitumor effect through viral oncolysis and siRNA-mediated oncogene silencing [459, 460]. Significant advances have been achieved with sterically stabilized lipid-based nanocarriers such as the stabilized nucleic acid lipid particles (SNALP). However, stabilization of nanocarriers with poly(ethylene glycol) (PEG) has not solved all problems associated with delivery of RNAi molecules. PEG modification weakens the internalization of the RNA molecules into the target cell and its subsequent escape from the endocytic pathway which reduces biological activity. To overcome such limitations novel exchangeable PEG-derivatized lipids can be used. After systemic administration, these lipids can be released from the nanoparticle surface. Additionally, the design and synthesis of cationic lipids that are more fusogenic and the use of internalizing targeting ligands have contributed to the emergence of novel lipid-based nanoparticles with remarkable transfection efficiency [461]. Finally, a nanoparticle formulation consisting of liposome-protamine-hyaluronic acid nanoparticles (LPH-NP) for systemic delivery of siRNA to tumors has been developed in a self-assembling process. The LPH-NP was further modified by PEG or PEG-anisamide lipids. Anisamide is a targeting ligand for the sigma receptor over-expressed in B16F10 melanoma cells. The targeted LPH-NP silenced 80% of luciferase activity in metastatic B16F10 lung tumors after a single i.v. injection and also showed very little immunotoxicity [462].

### **TME Mediated Drug Resistance**

Resistance against antitumor drugs and therapeutic radiation represents a tremendous challenge for most cancer therapies. It has been demonstrated by various experimental approaches that the mesenchymal TME provides a protective environment that obstructs drug or radiation access to the tumor cells or creates a permissive environment that supports for example the existence of cancer stem cell (CSC) niches where tumor cells overcome treatment- and cancer-induced stresses [463-471]. Resistance of tumors to anticancer drugs is mostly attributed to gene mutations, amplification of the multidrug resistance genes, epigenetic changes that influence drug uptake and metabolism, or export of drugs from cells [472-474]. An important advance in the understanding of tumor multidrug resistance (MDR) came with the identification of the P-glycoproteins (ABC transporter family) and other related transporters that are expressed in cancer cells and orchestrate the efflux of drugs from cells [475-477]. Tumor cells can also undergo physiologic changes in response to extracellular acidosis, a consequence of high glycolytic flux and poor vascular perfusion, both of which contribute to drug resistance including reduced apoptotic potential, genetic alterations, and elevated P-glycoprotein levels. A low extracellular pH creates a physiological drug barrier described by an "ion trapping" phenomenon [478, 479]. In addition, unfavorable pharmacokinetics and -dynamics and the limited ability of cancer drugs to diffuse deeply into hypoxic tumor tissue and to accumulate in tumor cells at lethal concentrations contributes to the unsatisfactory efficacy of cancer therapy [480, 481].

Regarding the contribution of stromal cells in the induction of drug resistance, increased infiltration of macrophages and high cathepsin protease levels in TAM were found in tumors following chemotherapy with paclitaxel, etoposide and doxorubicin, suggesting that cathepsin-expressing macrophages protected tumor cells against drug-induced tumor cell death [482]. It was also reported that TAM and their expression of milk-fat globule-epidermal growth factor-VIII (MFG-E8) play a role in the regulation of CSC. MFG-E8 activates Stat3 and Sonic Hedgehog pathways in CSC and further amplifies their anticancer drug resistance in cooperation with IL-6 [483]. The contribution of cancer associated fibroblasts (CAF) in induction of drug resistance was recently demonstrated in a co-culture

study of estrogen receptor positive MCF7 breast cancer cells with fibroblasts showing that tamoxifen resistance was induced by CAF. The fibroblasts also protected MCF7 cells against apoptosis induced by other anticancer agents, such as doxorubicin and the PARP-1 inhibitor ABT-888 [484].

Work in several different cancers has suggested that the CSC population serves as a source of chemotherapy and radiation-therapy resistance within tumors. Several resistance mechanisms have been proposed, including amplified checkpoint activation and DNA damage repair as well as increased Wnt/ $\beta$ -catenin and Notch signaling. Targeted therapies against the DNA damage checkpoint or stem-cell maintenance pathways may sensitize CSC to radiation or other therapies. CSC may also play a role in the induction of angiogenesis as well as in the mechanisms of resistance towards anti-angiogenic agents [485]. The dynamics of cancer cell death in response to therapy was recently investigated by intravital microscopy of chemotherapy-treated mouse tumors allowing a dynamic analysis of drug distribution, cell death and tumor-stroma interactions. Thereby, associations between vascular leakage and response to doxorubicin, including improved response in MMP-9 knockout mice that had increased vascular leakage were observed. Furthermore, CCR2-dependent infiltration of myeloid cells after treatment and better response of Ccr2 null host mice to doxorubicin and cisplatin treatment was demonstrated [486].

In respect to anti-angiogenic therapies, inhibitors targeting the VEGF signaling pathways have demonstrated, in both preclinical and clinical settings, that the benefits are at best transitory and often followed by re-establishment of tumor growth and progression. Several findings support the notion that two modes of unconventional resistance underlie such results; either the mode of evasive resistance, which is an adaptation to circumvent the specific angiogenic blockade, or an intrinsic or pre-existing indifference towards anti-angiogenic drugs [487-490]. Emerging evidence indicates that anti-angiogenic agents may increase intratumor hypoxia by promoting vessel pruning and inhibiting neo-angiogenesis. Indeed, several studies have highlighted the possibility that VEGF and VEGF-receptor inhibition can promote an invasive metastatic switch, in part by creating an increasingly hypoxic tumor microenvironment. As a potential remedy, a number of therapeutic approaches have been investigated that target the hypoxic tumor compartment to improve the clinical outcome of anti-angiogenic therapy [491-493].

Novel approaches to control drug resistance include functional genomics and proteomics [494,495]. RNA interference based screening provides a valuable opportunity for the examination of intrinsic and acquired resistance mechanisms. The availability of short interfering RNA libraries targeting genes allows performing large-scale screens to identify molecules that are involved in multidrug resistance pathways [496]. The emerging role of microRNAs as key gene expression regulators is also being explored in drug resistance research [497]. Finally, immunotherapy could represent an important adjuvant to treat MDR, as resistance to immunotherapy generally is unrelated to the classical mechanisms of resistance to cytotoxic agents. Immunotherapy to combat MDR could consist of direct immune attack against MDR positive cells, using MDR as an immune target to deliver cytotoxic drugs, taking advantage of other immune properties of MDR positive cells or application of immunotoxins expressed under MDR control [498, 499]. Regarding therapeutic approaches against drug resistance, nanodrug carriers, in particular liposomes, are widely explored [500-502]. Nanocarrier strategies for the reversal of resistance involve the alteration of drug efflux pumps and other resistance mechanisms. The methodologies involved include specific targeting of drugs and nucleotide therapeutics, improvement of cellular uptake and bioavailability of drugs with poor physicochemical characteristics. Multifunctional nanoparticulate systems consisting of a targeting moiety, encapsulated cytotoxic drugs and an element responsive to the TME to release the encapsulated therapeutics hold promise toward ways to improve cancer treatment [503-505].

## **CONCLUSIONS**

The cancer cell centric view of tumor progression largely ignored for a long time the fact that complex interactions between cancer cells and the cellular and molecular components of the tumor microenvironment tightly regulate and orchestrate tumor growth, metastatic dissemination and in many instances also the outcome of cancer therapies. Despite of continuous efforts, for many years cancer research largely focused on cancer-cell driven carcinogenesis and on understanding the mutations causing neoplastic cell transformations. But to provide new therapeutic strategies targeted at the immune components of the TME, it is critical to understand how these cells are altered during tumor progression and how they reciprocally influence tumor initiation, progression and

metastasis. The three mainstays of cancer therapy, surgical removal of tumor tissue, chemotherapy and radiotherapy will be complemented in the future by a fourth pillar, namely tumor immunotherapy and novel treatments aimed at the cellular and molecular components of the tumor microenvironment. Such novel strategies are urgently needed to complement the classical treatment modalities with more effective and patient tailored therapeutic approaches.

### **ABBREVIATIONS**

TME	= tumor microenvironment
TAM	= tumor-associated macrophage
M1	= classically activated macrophages
M2	= alternatively activated macrophages
TAN	= tumor-associated neutrophil
N1	= classically activated neutrophil
N2	= alternatively activated neutrophil
MDSC	= myeloid derived suppressor cell
DC	= dendritic cell
TADC	= tumor-associated dendritic cell
TIL	= tumor infiltrating leucocyte
LAK	= lymphokine activated killer cell
T <sub>reg</sub>	= regulatory T-cell
T <sub>h</sub>	= helper T-cell
CTL	= cytotoxic T-lymphocyte

MSC	= mesenchymal stem cell
CAF	= cancer-associated fibroblast
EC	= endothelial cell
MC	= mast cell
CSC	= cancer stem cell
ECM	= extracellular matrix
EMT	= epithelial-mesenchymal transition
MR	= mannose receptor
MHC	= major histocompatibility complex
Gr1	= granulocyte differentiation antigen
MMP	= matrix metalloproteinase
Arg	= arginase
HIF-1	= hypoxia inducible factor 1
ROS	= reactive oxygen species
ROI	= reactive oxygen intermediate
iNOS	= inducible nitric oxide synthase
NO	= nitric oxide
VEGF	= vascular endothelial growth factor
EGF	= epidermal growth factor
PDGF	= platelet-derived growth factor

FGF	=	fibroblast growth factor
HGF	=	hepatocyte growth factor
TNF- $\alpha$	=	tumor necrosis factor- $\alpha$
CCL	=	chemokine (C-C motif) ligand
CCR	=	chemokine (C-C motif) receptor
CXCL	=	chemokine (C-X-C motif) ligand
CSF-1	=	colony-stimulating factor 1
GM-CSF	=	granulocyte-macrophage colony-stimulating factor
IFN- $\gamma$	=	interferon $\gamma$
IL	=	interleukin
TGF- $\alpha$	=	transforming growth factor $\alpha$
TNF- $\alpha$	=	tumor necrosis factor $\alpha$
TGF- $\beta$	=	transforming growth factor- $\beta$
NF- $\kappa$ B	=	nuclear factor kappaB
LPS	=	lipopolysaccharide
STAT	=	signal transducer and activator of transcription
PDGF	=	platelet derived growth factor
HGF	=	hepatocyte growth factor
EGF	=	epidermal growth factor
bFGF	=	basic fibroblast growth factor

uPA	=	urokinase-type plasminogen activator
BP	=	bisphosphonate
N-BP	=	nitrogen-containing bisphosphonate
non-N-BP	=	non-nitrogen containing bisphosphonate
RAIT	=	radioimmunotherapy
ADEPT	=	antibody-directed enzyme prodrug therapy
GDEPT	=	gene-directed enzyme prodrug therapy
PMT	=	prodrug monotherapy
RNAi	=	RNA interference
siRNA	=	small interfering RNA
shRNA	=	short hairpin RNA
miRNA	=	microRNA.

#### **ACKNOWLEDGEMENTS**

The authors thank the Stiftung zur Krebsforschung, Zurich for financial support and S. Kumar for helpful discussions.

#### **CONFLICT OF INTEREST**

The author(s) confirm that this chapter content has no conflict of interest.

#### **DISCLOSURE**

Declared none.

#### **REFERENCES**

- [1] Vaupel P, Kallinowski F, Okunieff P. Blood flow, oxygen consumption and tissue oxygenation of human tumors. *Adv Exp Med Biol.* 1990;277:895-905.



- [2] Nicolson GL. Tumor microenvironment: paracrine and autocrine growth mechanisms and metastasis to specific sites. *Front Radiat Ther Oncol.* 1994;28:11-24.
- [3] Yuan J, Glazer PM. Mutagenesis induced by the tumor microenvironment. *Mutat Res.* 1998;400(1-2):439-446.
- [4] Park CC, Bissell MJ, Barcellos-Hoff MH. The influence of the microenvironment on the malignant phenotype. *Mol Med Today.* 2000;6(8):324-329.
- [5] Cairns RA, Khokha R, Hill RP. Molecular mechanisms of tumor invasion and metastasis: an integrated view. *Curr Mol Med.* 2003;3(7):659-671.
- [6] van Kempen LC, Ruiter DJ, van Muijen GN, Coussens LM. The tumor microenvironment: a critical determinant of neoplastic evolution. *Eur J Cell Biol.* 2003;82(11):539-548.
- [7] Leo C, Giaccia AJ, Denko NC. The hypoxic tumor microenvironment and gene expression. *Semin Radiat Oncol.* 2004;14(3):207-214.
- [8] Ben-Baruch A. The multifaceted roles of chemokines in malignancy. *Cancer Metast Rev.* 2006;25(3):357-371.
- [9] Polyak K, Haviv I, Campbell IG. Co-evolution of tumor cells and their microenvironment. *Trends Genet.* 2009;25(1):30-38.
- [10] Le Bitoux MA, Stamenkovic I. Tumor-host interactions: the role of inflammation. *Histochem Cell Biol.* 2008;130(6):1079-1090.
- [11] Mbeunkui F, Johann DJ, Jr. Cancer and the tumor microenvironment: a review of an essential relationship. *Cancer Chemother Pharmacol.* 2009;63(4):571-582.
- [12] Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A. Cancer-related inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis.* 2009;30(7):1073-1081.
- [13] Kim Y, Lin Q, Glazer PM, Yun Z. Hypoxic tumor microenvironment and cancer cell differentiation. *Curr Mol Med.* 2009;9(4):425-434.
- [14] Zhang X, Nie D, Chakrabarty S. Growth factors in tumor microenvironment. *Front Biosci.* 2010;15:151-165.
- [15] Alphonso A, Alahari SK. Stromal cells and integrins: conforming to the needs of the tumor microenvironment. *Neoplasia.* 2009;11(12):1264-1271.
- [16] Pietras K, Ostman A. Hallmarks of cancer: interactions with the tumor stroma. *Exp Cell Res.* 2010;316(8):1324-1331.
- [17] Kessenbrock K, Plaks V, Werb Z. Matrix metalloproteinases: regulators of the tumor microenvironment. *Cell.* 2010;141(1):52-67.
- [18] McAllister SS, Weinberg RA. Tumor-host interactions: a far-reaching relationship. *J Clin Oncol.* 2010;28(26):4022-4028.
- [19] Noman MZ, Messai Y, Carre T, Akalay I, Meron M, *et al.* Microenvironmental hypoxia orchestrating the cell stroma cross talk, tumor progression and antitumor response. *Crit Rev Immunol.* 2011;31(5):357-377.
- [20] Chouaib S, Kieda C, Benlalam H, Noman MZ, Mami-Chouaib F, *et al.* Endothelial cells as key determinants of the tumor microenvironment: interaction with tumor cells, extracellular matrix and immune killer cells. *Crit Rev Immunol.* 2010;30(6):529-545.
- [21] Zhang W, Huang P. Cancer-stromal interactions: role in cell survival, metabolism and drug sensitivity. *Cancer Biol Ther.* 2011;11(2):150-156.
- [22] Onimaru M, Yonemitsu Y. Angiogenic and lymphangiogenic cascades in the tumor microenvironment. *Front Biosci (Schol Ed).* 2011;3:216-225.
- [23] Mason SD, Joyce JA. Proteolytic networks in cancer. *Trends Cell Biol.* 2011;21(4):228-237.

- [24] Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674.
- [25] Bissell MJ, Hines WC. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. *Nat Med*. 2011;17(3):320-329.
- [26] Balkwill FR. The chemokine system and cancer. *J Pathol*. 2012;226(2):148-157.
- [27] Fidler IJ. Seed and soil revisited: contribution of the organ microenvironment to cancer metastasis. *Surg Oncol Clin N Am*. 2001;10(2):257-269.
- [28] Fodstad O, Kjønniksen I. Microenvironment revisited: time for reappraisal of some prevailing concepts of cancer metastasis. *J Cell Biochem*. 1994;56(1):23-28.
- [29] Quaranta V. Motility cues in the tumor microenvironment. *Differentiation*. 2002;70(9-10):590-598.
- [30] Karnoub AE, Weinberg RA. Chemokine networks and breast cancer metastasis. *Breast Dis*. 2006;26:75-85.
- [31] Yilmaz M, Christofori G, Lehenbre F. Distinct mechanisms of tumor invasion and metastasis. *Trends Mol Med*. 2007;13(12):535-541.
- [32] Tse JC, Kalluri R. Mechanisms of metastasis: epithelial-to-mesenchymal transition and contribution of tumor microenvironment. *J Cell Biochem*. 2007;101(4):816-829.
- [33] Langley RR, Fidler IJ. Tumor cell-organ microenvironment interactions in the pathogenesis of cancer metastasis. *Endocr Rev*. 2007;28(3):297-321.
- [34] Lunt SJ, Chaudary N, Hill RP. The tumor microenvironment and metastatic disease. *Clin Exp Metast*. 2009;26(1):19-34.
- [35] Finger EC, Giaccia AJ. Hypoxia, inflammation, and the tumor microenvironment in metastatic disease. *Cancer Metast Rev*. 2010;29(2):285-293.
- [36] Keeley EC, Mehrad B, Strieter RM. CXC chemokines in cancer angiogenesis and metastases. *Adv Cancer Res*. 2010;106:91-111.
- [37] Guise T. Examining the metastatic niche: targeting the microenvironment. *Semin Oncol*. 2010;37 Suppl 2:S2-14.
- [38] Talmadge JE. Immune cell infiltration of primary and metastatic lesions: mechanisms and clinical impact. *Semin Cancer Biol*. 2011;21(2):131-138.
- [39] Farber E. Carcinogenesis--cellular evolution as a unifying thread: Presidential address. *Cancer Res*. 1973;33(11):2537-2550.
- [40] Vaupel P, Kallinowski F, Okunieff P. Blood flow, oxygen and nutrient supply, and metabolic microenvironment of human tumors: a review. *Cancer Res*. 1989;49(23):6449-6465.
- [41] Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med*. 1971;285(21):1182-1186.
- [42] Folkman J. Tumor angiogenesis. *Adv Cancer Res*. 1985;43:175-203.
- [43] Kerbel RS. Tumor angiogenesis: past, present and the near future. *Carcinogenesis*. 2000;21(3):505-515.
- [44] Jung YD, Ahmad SA, Liu W, Reinmuth N, Parikh A, *et al*. The role of the microenvironment and intercellular cross-talk in tumor angiogenesis. *Semin Cancer Biol*. 2002;12(2):105-112.
- [45] Jain RK. Antiangiogenic therapy for cancer: current and emerging concepts. *Oncology (Williston Park)*. 2005;19(4 Suppl 3):7-16.
- [46] Nyberg P, Salo T, Kalluri R. Tumor microenvironment and angiogenesis. *Front Biosci*. 2008;13:6537-6553.
- [47] Baeriswyl V, Christofori G. The angiogenic switch in carcinogenesis. *Semin Cancer Biol*. 2009;19(5):329-337.

- [48] Takahashi H, Shibuya M. The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and pathological conditions. *Clin Sci (Lond)*. 2005;109(3):227-241.
- [49] Nucera S, Biziato D, De Palma M. The interplay between macrophages and angiogenesis in development, tissue injury and regeneration. *Int J Dev Biol*. 2011;55(4-5):495-503.
- [50] Michiels C, Arnould T, Remalec J. Endothelial cell responses to hypoxia: initiation of a cascade of cellular interactions. *Biochim Biophys Acta*. 2000;1497(1):1-10.
- [51] Hockel M, Vaupel P. Biological consequences of tumor hypoxia. *Semin Oncol*. 2001;28(2 Suppl 8):36-41.
- [52] Shannon AM, Bouchier-Hayes DJ, Condrón CM, Toomey D. Tumour hypoxia, chemotherapeutic resistance and hypoxia-related therapies. *Cancer Treat Rev*. 2003;29(4):297-307.
- [53] Acker T, Plate KH. Hypoxia and hypoxia inducible factors (HIF) as important regulators of tumor physiology. *Cancer Treat Res*. 2004;117:219-248.
- [54] Dayan F, Mazure NM, Brahimi-Horn MC, Pouyssegur J. A dialogue between the hypoxia-inducible factor and the tumor microenvironment. *Cancer Microenviron*. 2008;1(1):53-68.
- [55] Ruan K, Song G, Ouyang G. Role of hypoxia in the hallmarks of human cancer. *J Cell Biochem*. 2009;107(6):1053-1062.
- [56] Ziche M, Morbidelli L. Molecular regulation of tumour angiogenesis by nitric oxide. *Eur Cytokine Netw*. 2009;20(4):164-170.
- [57] Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, *et al*. HIF-1 $\alpha$  regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. *J Exp Med*. 2010;207(11):2439-2453.
- [58] Takenaga K. Angiogenic signaling aberrantly induced by tumor hypoxia. *Front Biosci*. 2011;16:31-48.
- [59] Murdoch C, Lewis CE. Macrophage migration and gene expression in response to tumor hypoxia. *Int J Cancer*. 2005;117(5):701-708.
- [60] Lewis C, Murdoch C. Macrophage responses to hypoxia: implications for tumor progression and anti-cancer therapies. *Am J Pathol*. 2005;167(3):627-635.
- [61] Lamagna C, Aurrand-Lions M, Imhof BA. Dual role of macrophages in tumor growth and angiogenesis. *J Leukoc Biol*. 2006;80(4):705-713.
- [62] Coffelt SB, Hughes R, Lewis CE. Tumor-associated macrophages: effectors of angiogenesis and tumor progression. *Biochim Biophys Acta*. 2009;1796(1):11-18.
- [63] Sica A. Role of tumour-associated macrophages in cancer-related inflammation. *Exp Oncol*. 2010;32(3):153-158.
- [64] Tartour E, Pere H, Maillere B, Terme M, Merillon N, *et al*. Angiogenesis and immunity: a bidirectional link potentially relevant for the monitoring of antiangiogenic therapy and the development of novel therapeutic combination with immunotherapy. *Cancer Metast Rev*. 2011;30(1):83-95.
- [65] Noonan DM, De Lerma Barbaro A, Vannini N, Mortara L, Albini A. Inflammation, inflammatory cells and angiogenesis: decisions and indecisions. *Cancer Metast Rev*. 2008;27(1):31-40.
- [66] Shojaei F, Ferrara N. Refractoriness to antivascular endothelial growth factor treatment: role of myeloid cells. *Cancer Res*. 2008;68(14):5501-5504.
- [67] Mantovani A, Sica A, Allavena P, Garlanda C, Locati M. Tumor-associated macrophages and the related myeloid-derived suppressor cells as a paradigm of the diversity of macrophage activation. *Hum Immunol*. 2009;70(5):325-330.

- [68] Gerber HP, Olazoglu E, Grewal IS. Targeting inflammatory cells to improve anti-VEGF therapies in oncology. *Recent Results Cancer Res.* 2010;180:185-200.
- [69] Benelli R, Albini A, Noonan D. Neutrophils and angiogenesis: potential initiators of the angiogenic cascade. *Chem Immunol Allergy.* 2003;83:167-181.
- [70] Yuan A, Chen JJ, Yao PL, Yang PC. The role of interleukin-8 in cancer cells and microenvironment interaction. *Front Biosci.* 2005;10:853-865.
- [71] Tazzyman S, Lewis CE, Murdoch C. Neutrophils: key mediators of tumour angiogenesis. *Int J Exp Pathol.* 2009;90(3):222-231.
- [72] Gregory AD, Houghton AM. Tumor-associated neutrophils: new targets for cancer therapy. *Cancer Res.* 2011;71(7):2411-2416.
- [73] Ferrara N. Role of myeloid cells in vascular endothelial growth factor-independent tumor angiogenesis. *Curr Opin Hematol.* 2010;17(3):219-224.
- [74] Kujawski M, Kortylewski M, Lee H, Herrmann A, Kay H, *et al.* Stat3 mediates myeloid cell-dependent tumor angiogenesis in mice. *J Clin Invest.* 2008;118(10):3367-3377.
- [75] Diaz-Flores L, Gutierrez R, Varela H. Angiogenesis: an update. *Histol Histopathol.* 1994;9(4):807-843.
- [76] Armulik A, Abramsson A, Betsholtz C. Endothelial/pericyte interactions. *Circ Res.* 2005;97(6):512-523.
- [77] Diaz-Flores L, Gutierrez R, Madrid JF, Varela H, Valladares F, *et al.* Pericytes. Morphofunction, interactions and pathology in a quiescent and activated mesenchymal cell niche. *Histol Histopathol.* 2009;24(7):909-969.
- [78] Fakhrejehani E, Toi M. Tumor angiogenesis: pericytes and maturation are not to be ignored. *J Oncol.* 2012;2012:261750.
- [79] Ingber DE. Extracellular matrix as a solid-state regulator in angiogenesis: identification of new targets for anti-cancer therapy. *Semin Cancer Biol.* 1992;3(2):57-63.
- [80] Boudreau N, Bissell MJ. Extracellular matrix signaling: integration of form and function in normal and malignant cells. *Curr Opin Cell Biol.* 1998;10(5):640-646.
- [81] Hornebeck W, Emonard H, Monboisse JC, Bellon G. Matrix-directed regulation of pericellular proteolysis and tumor progression. *Semin Cancer Biol.* 2002;12(3):231-241.
- [82] Cretu A, Brooks PC. Impact of the non-cellular tumor microenvironment on metastasis: potential therapeutic and imaging opportunities. *J Cell Physiol.* 2007;213(2):391-402.
- [83] Marastoni S, Ligresti G, Lorenzon E, Colombatti A, Mongiat M. Extracellular matrix: a matter of life and death. *Connect Tissue Res.* 2008;49(3):203-206.
- [84] Barkan D, Green JE, Chambers AF. Extracellular matrix: a gatekeeper in the transition from dormancy to metastatic growth. *Eur J Cancer.* 2010;46(7):1181-1188.
- [85] Kim SH, Turnbull J, Guimond S. Extracellular matrix and cell signalling: the dynamic cooperation of integrin, proteoglycan and growth factor receptor. *J Endocrinol.* 2011;209(2):139-151.
- [86] Lynch CC, Matrisian LM. Matrix metalloproteinases in tumor-host cell communication. *Differentiation.* 2002;70(9-10):561-573.
- [87] Jodele S, Blavier L, Yoon JM, DeClerck YA. Modifying the soil to affect the seed: role of stromal-derived matrix metalloproteinases in cancer progression. *Cancer Metast Rev.* 2006;25(1):35-43.
- [88] Gialeli C, Theocharis AD, Karamanos NK. Roles of matrix metalloproteinases in cancer progression and their pharmacological targeting. *FEBS J.* 2011;278(1):16-27.
- [89] Pupa SM, Menard S, Forti S, Tagliabue E. New insights into the role of extracellular matrix during tumor onset and progression. *J Cell Physiol.* 2002;192(3):259-267.

- [90] Kumar S, Weaver VM. Mechanics, malignancy, and metastasis: the force journey of a tumor cell. *Cancer Metast Rev.* 2009;28(1-2):113-127.
- [91] El-Bahrawy MA, Pignatelli M. E-cadherin and catenins: molecules with versatile roles in normal and neoplastic epithelial cell biology. *Microsc Res Tech.* 1998;43(3):224-232.
- [92] Vincent-Salomon A, Thiery JP. Host microenvironment in breast cancer development: epithelial-mesenchymal transition in breast cancer development. *Breast Cancer Res.* 2003;5(2):101-106.
- [93] Klymkowsky MW, Savagner P. Epithelial-mesenchymal transition: a cancer researcher's conceptual friend and foe. *Am J Pathol.* 2009;174(5):1588-1593.
- [94] Iwatsuki M, Mimori K, Yokobori T, Ishi H, Beppu T, *et al.* Epithelial-mesenchymal transition in cancer development and its clinical significance. *Cancer Sci.* 2010;101(2):293-299.
- [95] Jing Y, Han Z, Zhang S, Liu Y, Wei L. Epithelial-Mesenchymal Transition in tumor microenvironment. *Cell Biosci.* 2011;1:29.
- [96] Ungefroren H, Sebens S, Seidl D, Lehnert H, Hass R. Interaction of tumor cells with the microenvironment. *Cell Commun Signal.* 2011;9:18.
- [97] Roberts AB, Tian F, Byfield SD, Stuelten C, Ooshima A, *et al.* Smad3 is key to TGF-beta-mediated epithelial-to-mesenchymal transition, fibrosis, tumor suppression and metastasis. *Cytokine Growth Factor Rev.* 2006;17(1-2):19-27.
- [98] Bonde AK, Tischler V, Kumar S, Soltermann A, Schwendener R. Intratumoral macrophages contribute to epithelial-mesenchymal transition in solid tumors. *BMC Cancer.* 2012;12(1):35.
- [99] Giannoni E, Parri M, Chiarugi P. EMT and oxidative stress: a bidirectional interplay affecting tumor malignancy. *Antioxid Redox Signal.* 2012;16(11):1248-1263.
- [100] Vicari AP, Caux C. Chemokines in cancer. *Cytokine Growth Factor Rev.* 2002;13(2):143-154.
- [101] Wilson J, Balkwill F. The role of cytokines in the epithelial cancer microenvironment. *Semin Cancer Biol.* 2002;12(2):113-120.
- [102] Ben-Baruch A. Host microenvironment in breast cancer development: inflammatory cells, cytokines and chemokines in breast cancer progression: reciprocal tumor-microenvironment interactions. *Breast Cancer Res.* 2003;5(1):31-36.
- [103] Shurin MR, Shurin GV, Lokshin A, Yurkovetsky ZR, Gutkin DW, *et al.* Intratumoral cytokines/chemokines/growth factors and tumor infiltrating dendritic cells: friends or enemies? *Cancer Metast Rev.* 2006;25(3):333-356.
- [104] Desiderio MA. Hepatocyte growth factor in invasive growth of carcinomas. *Cell Mol Life Sci.* 2007;64(11):1341-1354.
- [105] Raman D, Baugher PJ, Thu YM, Richmond A. Role of chemokines in tumor growth. *Cancer Lett.* 2007;256(2):137-165.
- [106] Allavena P, Garlanda C, Borrello MG, Sica A, Mantovani A. Pathways connecting inflammation and cancer. *Curr Opin Genet Dev.* 2008;18(1):3-10.
- [107] Keibel A, Singh V, Sharma MC. Inflammation, microenvironment, and the immune system in cancer progression. *Curr Pharm Des.* 2009;15(17):1949-1955.
- [108] Mantovani A, Savino B, Locati M, Zampataro L, Allavena P, *et al.* The chemokine system in cancer biology and therapy. *Cytokine Growth Factor Rev.* 2010;21(1):27-39.
- [109] Sheng KC, Wright MD, Apostolopoulos V. Inflammatory mediators hold the key to dendritic cell suppression and tumor progression. *Curr Med Chem.* 2011;18(36):5507-5518.

- [110] Klampfer L. Cytokines, inflammation and colon cancer. *Curr Cancer Drug Targets*. 2011;11(4):451-464.
- [111] Sheu BC, Chang WC, Cheng CY, Lin HH, Chang DY, *et al*. Cytokine regulation networks in the cancer microenvironment. *Front Biosci*. 2008;13:6255-6268.
- [112] Kischel P, Waltregny D, Dumont B, Turtoi A, Greffe Y, *et al*. Versican overexpression in human breast cancer lesions: known and new isoforms for stromal tumor targeting. *Int J Cancer*. 2010;126(3):640-650.
- [113] Bierie B, Moses HL. Transforming growth factor beta (TGF-beta) and inflammation in cancer. *Cytokine Growth Factor Rev*. 2010;21(1):49-59.
- [114] Rasanen K, Vaheri A. Activation of fibroblasts in cancer stroma. *Exp Cell Res*. 2010;316(17):2713-2722.
- [115] Hughes CC. Endothelial-stromal interactions in angiogenesis. *Curr Opin Hematol*. 2008;15(3):204-209.
- [116] Orimo A, Weinberg RA. Stromal fibroblasts in cancer: a novel tumor-promoting cell type. *Cell Cycle*. 2006;5(15):1597-1601.
- [117] Hugo HJ, Lebet S, Tomaskovic-Crook E, Ahmed N, Blick T, *et al*. Contribution of Fibroblast and Mast Cell (Afferent) and Tumor (Efferent) IL-6 Effects within the Tumor Microenvironment. *Cancer Microenviron*. 2012. [Epub ahead of print]
- [118] Elenbaas B, Weinberg RA. Heterotypic signaling between epithelial tumor cells and fibroblasts in carcinoma formation. *Exp Cell Res*. 2001;264(1):169-184.
- [119] Kataoka H, Tanaka H, Nagaika K, Uchiyama S, Itoh H. Role of cancer cell-stroma interaction in invasive growth of cancer cells. *Hum Cell*. 2003;16(1):1-14.
- [120] Mueller L, Goumas FA, Affeldt M, Sandtner S, Gehling UM, *et al*. Stromal fibroblasts in colorectal liver metastases originate from resident fibroblasts and generate an inflammatory microenvironment. *Am J Pathol*. 2007;171(5):1608-1618.
- [121] Mishra PJ, Glod JW, Banerjee D. Mesenchymal stem cells: flip side of the coin. *Cancer Res*. 2009;69(4):1255-1258.
- [122] Liao D, Luo Y, Markowitz D, Xiang R, Reisfeld RA. Cancer associated fibroblasts promote tumor growth and metastasis by modulating the tumor immune microenvironment in a 4T1 murine breast cancer model. *PLoS One*. 2009;4(11):e7965.
- [123] Shea-Donohue T, Stiltz J, Zhao A, Notari L. Mast cells. *Curr Gastroenterol Rep*. 2010;12(5):349-357.
- [124] Liu J, Zhang Y, Zhao J, Yang Z, Li D, *et al*. Mast cell: insight into remodeling a tumor microenvironment. *Cancer Metast Rev*. 2011;30(2):177-184.
- [125] Khazaie K, Blatner NR, Khan MW, Gounari F, Gounaris E, *et al*. The significant role of mast cells in cancer. *Cancer Metast Rev*. 2011;30(1):45-60.
- [126] Maltby S, Khazaie K, McNagny KM. Mast cells in tumor growth: angiogenesis, tissue remodelling and immune-modulation. *Biochim Biophys Acta*. 2009;1796(1):19-26.
- [127] Wensman H, Kamgari N, Johansson A, Grujic M, Calounova G, *et al*. Tumor-mast cell interactions: induction of pro-tumorigenic genes and anti-tumorigenic 4-1BB in MCs in response to Lewis Lung Carcinoma. *Mol Immunol*. 2012;50(4):210-219.
- [128] Groot Kormelink T, Abudukelimu A, Redegeld FA. Mast cells as target in cancer therapy. *Curr Pharm Des*. 2009;15(16):1868-1878.
- [129] Strouch MJ, Cheon EC, Salabat MR, Krantz SB, Gounaris E, *et al*. Crosstalk between mast cells and pancreatic cancer cells contributes to pancreatic tumor progression. *Clin Cancer Res*. 2010;16(8):2257-2265.

- [130] Takanami I, Takeuchi K, Naruke M. Mast cell density is associated with angiogenesis and poor prognosis in pulmonary adenocarcinoma. *Cancer*. 2000;88(12):2686-2692.
- [131] Rajput AB, Turbin DA, Cheang MC, Voduc DK, Leung S, *et al*. Stromal mast cells in invasive breast cancer are a marker of favourable prognosis: a study of 4,444 cases. *Breast Cancer Res Treat*. 2008;107(2):249-257.
- [132] Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet*. 2001;357(9255):539-545.
- [133] Sica A, Allavena P, Mantovani A. Cancer related inflammation: The macrophage connection. *Cancer Lett*. 2008;267(2):204-215.
- [134] Wang Y, Ma Y, Fang Y, Wu S, Liu L, *et al*. Regulatory T cell: a protection for tumour cells. *J Cell Mol Med*. 2012;16(3):425-436.
- [135] Byrne WL, Mills KH, Lederer JA, O'Sullivan GC. Targeting regulatory T cells in cancer. *Cancer Res*. 2011;71(22):6915-6920.
- [136] Elkord E, Alcantar-Orozco EM, Dovedi SJ, Tran DQ, Hawkins RE, *et al*. T regulatory cells in cancer: recent advances and therapeutic potential. *Expert Opin Biol Ther*. 2010;10(11):1573-1586.
- [137] Beyer M, Schultze JL. Regulatory T cells: major players in the tumor microenvironment. *Curr Pharm Des*. 2009;15(16):1879-1892.
- [138] Wang HY, Wang RF. Regulatory T cells and cancer. *Curr Opin Immunol*. 2007;19(2):217-223.
- [139] Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol*. 2002;3(11):991-998.
- [140] Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoediting. *Annu Rev Immunol*. 2004;22:329-360.
- [141] Smyth MJ, Dunn GP, Schreiber RD. Cancer immunosurveillance and immunoediting: the roles of immunity in suppressing tumor development and shaping tumor immunogenicity. *Adv Immunol*. 2006;90:1-50.
- [142] Kim R, Emi M, Tanabe K. Cancer immunoediting from immune surveillance to immune escape. *Immunology*. 2007;121(1):1-14.
- [143] Ostrand-Rosenberg S. Immune surveillance: a balance between protumor and antitumor immunity. *Curr Opin Genet Dev*. 2008;18(1):11-18.
- [144] Vesely MD, Kershaw MH, Schreiber RD, Smyth MJ. Natural innate and adaptive immunity to cancer. *Annu Rev Immunol*. 2011;29:235-271.
- [145] Schreiber RD, Old LJ, Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science*. 2011;331(6024):1565-1570.
- [146] Chow MT, Moller A, Smyth MJ. Inflammation and immune surveillance in cancer. *Semin Cancer Biol*. 2012;22(1):23-32.
- [147] Kreider JW, Bartlett GL, Butkiewicz BL. Relationship of tumor leucocytic infiltration to host defense mechanisms and prognosis. *Cancer Metast Rev*. 1984;3(1):53-74.
- [148] Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoediting. *Immunity*. 2004;21(2):137-148.
- [149] Adams TE, Alpert S, Hanahan D. Nontolerance and autoantibodies to a transgenic self antigen expressed in pancreatic beta-cells. *Nature*. 1987;325(6101):223-228.
- [150] Lin EY, Pollard JW. Role of infiltrated leucocytes in tumour growth and spread. *Br J Cancer*. 2004;90(11):2053-2058.
- [151] Whiteside TL. The role of immune cells in the tumor microenvironment. *Cancer Treat Res*. 2006;130:103-124.

- [152] Coussens LM, Werb Z. Inflammation and cancer. *Nature*. 2002;420(6917):860-867.
- [153] Fridman WH, Galon JP, F., Tartour E, Sautès-Fridman C, Kroemer G. Prognostic and predictive impact of intra- and peritumoral immune infiltrates. *Cancer Res*. 2011;71(17):5601-5605.
- [154] Coca S, PerezPiqueras J, Martinez D, Colmenarejo A, Saez MA, *et al*. The prognostic significance of intratumoral natural killer cells in patients with colorectal carcinoma. *Cancer*. 1997;79(12):2320-2328.
- [155] Ishigami S, Natsugoe S, Tokuda K, Nakajo A, Che XM, *et al*. Prognostic value of intratumoral natural killer cells in gastric carcinoma. *Cancer*. 2000;88(3):577-583.
- [156] Yamada N, Oizumi S, Kikuchi E, Shinagawa N, Konishi-Sakakibara J, *et al*. CD8(+) tumor-infiltrating lymphocytes predict favorable prognosis in malignant pleural mesothelioma after resection. *Cancer Immunol Immunother*. 2010; 59(10):1543-1549.
- [157] Hume DA. The mononuclear phagocyte system. *Curr Opin Immunol*. 2006;18(1):49-53.
- [158] Chow A, Brown BD, Merad M. Studying the mononuclear phagocyte system in the molecular age. *Nat Rev Immunol*. 2011;11(11):788-798.
- [159] Chang ZL. Recent development of the mononuclear phagocyte system: in memory of Metchnikoff and Ehrlich on the 100th Anniversary of the 1908 Nobel Prize in Physiology or Medicine. *Biol Cell*. 2009;101(12):709-721.
- [160] van Furth R. Macrophage activity and clinical immunology. Origin and kinetics of mononuclear phagocytes. *Ann N Y Acad Sci*. 1976;278:161-175.
- [161] Kende M. Role of macrophages in the expression of immune responses. *J Am Vet Med Assoc*. 1982;181(10):1037-1042.
- [162] van Furth R. Current view on the mononuclear phagocyte system. *Immunobiology*. 1982;161(3-4):178-185.
- [163] Lasser A. The mononuclear phagocytic system: a review. *Hum Pathol*. 1983;14(2):108-126.
- [164] Gordon S, Keshav S, Chung LP. Mononuclear phagocytes: tissue distribution and functional heterogeneity. *Curr Opin Immunol*. 1988;1(1):26-35.
- [165] Hume DA, Ross IL, Himes SR, Sasmono RT, Wells CA, *et al*. The mononuclear phagocyte system revisited. *J Leukoc Biol*. 2002;72(4):621-627.
- [166] Mantovani A, Sozzani S, Locati M, Allavena P, Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol*. 2002;23(11):549-555.
- [167] Gordon S, Mantovani A. Diversity and plasticity of mononuclear phagocytes. *Eur J Immunol*. 2011;41(9):2470-2472.
- [168] Sou K, Goins B, Oyajobi BO, Travi BL, Phillips WT. Bone marrow-targeted liposomal carriers. *Expert Opin Drug Deliv*. 2011;8(3):317-328.
- [169] Areschoug T, Gordon S. Scavenger receptors: role in innate immunity and microbial pathogenesis. *Cell Microbiol*. 2009;11(8):1160-1169.
- [170] Owens DE, 3rd, Peppas NA. Opsonization, biodistribution, and pharmacokinetics of polymeric nanoparticles. *Int J Pharm*. 2006;307(1):93-102.
- [171] Linehan SA, Martinez-Pomares L, Gordon S. Mannose receptor and scavenger receptor: two macrophage pattern recognition receptors with diverse functions in tissue homeostasis and host defense. *Adv Exp Med Biol*. 2000;479:1-14.
- [172] Vuarchey CK, S. Schwendener R. Albumin coated liposomes: a novel platform for macrophage specific drug delivery. *Nanotechnol Development*. 2011;1:e2.



- [173] Schwendener RA, Lagocki PA, Rahman YE. The effects of charge and size on the interaction of unilamellar liposomes with macrophages. *Biochim Biophys Acta*. 1984;772(1):93-101.
- [174] Key ME. Macrophages in cancer metastases and their relevance to metastatic growth. *Cancer Metast Rev*. 1983;2(1):75-88.
- [175] Walter S, Govoni D, Bottazzi B, Mantovani A. The role of macrophages in the regulation of primary tumor growth. *Pathobiology*. 1991;59(4):239-242.
- [176] Mantovani A, Bottazzi B, Colotta F, Sozzani S, Ruco L. The origin and function of tumor-associated macrophages. *Immunol Today*. 1992;13(7):265-270.
- [177] Lewis CE, Leek R, Harris A, McGee JO. Cytokine regulation of angiogenesis in breast cancer: the role of tumor-associated macrophages. *J Leukoc Biol*. 1995;57(5):747-751.
- [178] Elgert KD, Alleva DG, Mullins DW. Tumor-induced immune dysfunction: the macrophage connection. *J Leukoc Biol*. 1998;64(3):275-290.
- [179] Sica A, Saccani A, Mantovani A. Tumor-associated macrophages: a molecular perspective. *Int Immunopharmacol*. 2002;2(8):1045-1054.
- [180] Mantovani A, Schioppa T, Biswas SK, Marchesi F, Allavena P, *et al*. Tumor-associated macrophages and dendritic cells as prototypic type II polarized myeloid populations. *Tumori*. 2003;89(5):459-468.
- [181] Porta C, Subhra Kumar B, Larghi P, Rubino L, Mancino A, *et al*. Tumor promotion by tumor-associated macrophages. *Adv Exp Med Biol*. 2007;604:67-86.
- [182] Guruvayoorappan C. Tumor *versus* tumor-associated macrophages: how hot is the link? *Integr Cancer Ther*. 2008;7(2):90-95.
- [183] Martinez FO, Sica A, Mantovani A, Locati M. Macrophage activation and polarization. *Front Biosci*. 2008;13:453-461.
- [184] Fukuda K, Kobayashi A, Watabe K. The role of tumor-associated macrophage in tumor progression. *Front Biosci (Schol Ed)*. 2012;4:787-798.
- [185] Allavena P, Mantovani A. Immunology in the clinic review series; focus on cancer: tumour-associated macrophages: undisputed stars of the inflammatory tumour microenvironment. *Clin Exp Immunol*. 2012;167(2):195-205.
- [186] Mantovani A, Germano G, Marchesi F, Locatelli M, Biswas SK. Cancer-promoting tumor-associated macrophages: new vistas and open questions. *Eur J Immunol*. 2011;41(9):2522-2525.
- [187] Ueha S, Shand FH, Matsushima K. Myeloid cell population dynamics in healthy and tumor-bearing mice. *Int Immunopharmacol*. 2011;11(7):783-788.
- [188] Laoui D, Van Overmeire E, Movahedi K, Van den Bossche J, Schoupe E, *et al*. Mononuclear phagocyte heterogeneity in cancer: different subsets and activation states reaching out at the tumor site. *Immunobiology*. 2011;216(11):1192-1202.
- [189] Mantovani A, Sica A. Macrophages, innate immunity and cancer: balance, tolerance, and diversity. *Curr Opin Immunol*. 2010;22(2):231-237.
- [190] Stout RD, Watkins SK, Suttles J. Functional plasticity of macrophages: *in situ* reprogramming of tumor-associated macrophages. *J Leukoc Biol*. 2009;86(5):1105-1109.
- [191] Siveen KS, Kuttan G. Role of macrophages in tumour progression. *Immunol Lett*. 2009;123(2):97-102.
- [192] Ruffell B, Affara NI, Coussens LM. Differential macrophage programming in the tumor microenvironment. *Trends Immunol*. 2012;33(3):119-126.
- [193] Taylor PR, Martinez-Pomares L, Stacey M, Lin HH, Brown GD, *et al*. Macrophage receptors and immune recognition. *Annu Rev Immunol*. 2005;23:901-944.

- [194] Van Gorp H, Delputte PL, Nauwynck HJ. Scavenger receptor CD163, a Jack-of-all-trades and potential target for cell-directed therapy. *Mol Immunol.* 2010;47(7-8):1650-1660.
- [195] Greaves DR, Gordon S. The macrophage scavenger receptor at 30 years of age: current knowledge and future challenges. *J Lipid Res.* 2009;50 Suppl:S282-286.
- [196] Mantovani A, Sozzanic S, Locatie M, Allavenaf P, Sicaf A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* 2002;23(11):549-555.
- [197] Schmieder A, Michel J, Schonhaar K, Goerdts S, Schledzewski K. Differentiation and gene expression profile of tumor-associated macrophages. *Semin Cancer Biol.* 2012.
- [198] Ruffell B, Au A, Rugo HS, Esserman LJ, Hwang ES, *et al.* Leukocyte composition of human breast cancer. *Proc Natl Acad Sci U S A.* 2012;109(8):2796-2801.
- [199] Torroella-Kouri M, Silvera R, Rodriguez D, Caso R, Shatry A, *et al.* Identification of a subpopulation of macrophages in mammary tumor-bearing mice that are neither M1 nor M2 and are less differentiated. *Cancer Res.* 2009;69(11):4800-4809.
- [200] Cortez-Retamozo V, Etzrodt M, Newton A, Rauch PJ, Chudnovskiy A, *et al.* Origins of tumor-associated macrophages and neutrophils. *Proc Natl Acad Sci U S A.* 2012;109(7):2491-2496.
- [201] Curiel TJ, Coukos G, Zou LH, Alvarez X, Cheng P, *et al.* Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med.* 2004;10(9):942-949.
- [202] Mlecnik B, Bindea G, Pages F, Galon J. Tumor immunosurveillance in human cancers. *Cancer Metast Rev.* 2011;30(1):5-12.
- [203] Tan SY, Fan Y, Luo HS, Shen ZX, Guo Y, *et al.* Prognostic significance of cell infiltrations of immunosurveillance in colorectal cancer. *World J Gastroenterol.* 2005;11(8):1210-1214.
- [204] Erreni M, Mantovani A, Allavena P. Tumor-associated Macrophages (TAM) and Inflammation in Colorectal Cancer. *Cancer Microenviron.* 2011;4(2):141-154.
- [205] Dai F, Liu L, Che G, Yu N, Pu Q, *et al.* The number and microlocalization of tumor-associated immune cells are associated with patient's survival time in non-small cell lung cancer. *BMC Cancer.* 2010;10:220.
- [206] Ho CC, Liao WY, Wang CY, Lu YH, Huang HY, *et al.* TREM-1 expression in tumor-associated macrophages and clinical outcome in lung cancer. *Am J Respir Crit Care Med.* 2008;177(7):763-770.
- [207] Chen JJ, Lin YC, Yao PL, Yuan A, Chen HY, *et al.* Tumor-associated macrophages: the double-edged sword in cancer progression. *J Clin Oncol.* 2005;23(5):953-964.
- [208] Jakovic LR, Mihaljevic BS, Perunicic Jovanovic MD, Bogdanovic AD, Andjelic BM, *et al.* The prognostic relevance of tumor associated macrophages in advanced stage classical Hodgkin lymphoma. *Leuk Lymphoma.* 2011;52(10):1913-1919.
- [209] Campbell MJ, Tonlaar NY, Garwood ER, Huo D, Moore DH, *et al.* Proliferating macrophages associated with high grade, hormone receptor negative breast cancer and poor clinical outcome. *Breast Cancer Res Treat.* 2011;128(3):703-711.
- [210] Mantovani A, Marchesi F, Porta C, Sica A, Allavena P. Inflammation and cancer: breast cancer as a prototype. *Breast.* 2007;16 Suppl 2:S27-33.
- [211] Jia JB, Wang WQ, Sun HC, Zhu XD, Liu L, *et al.* High expression of macrophage colony-stimulating factor-1 receptor in peritumoral liver tissue is associated with poor outcome in hepatocellular carcinoma after curative resection. *Oncologist.* 2010;15(7):732-743.
- [212] Yang JD, Nakamura I, Roberts LR. The tumor microenvironment in hepatocellular carcinoma: current status and therapeutic targets. *Semin Cancer Biol.* 2011;21(1):35-43.

- [213] Lissbrant IF, Stattin P, Wikstrom P, Damber JE, Egevad L, *et al.* Tumor associated macrophages in human prostate cancer: relation to clinicopathological variables and survival. *Int J Oncol.* 2000;17(3):445-451.
- [214] Ditsworth D, Zong WX. NF-kappaB: key mediator of inflammation-associated cancer. *Cancer Biol Ther.* 2004;3(12):1214-1216.
- [215] Inoue J, Gohda J, Akiyama T, Semba K. NF-kappaB activation in development and progression of cancer. *Cancer Sci.* 2007;98(3):268-274.
- [216] Grivennikov SI, Karin M. Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev.* 2010;21(1):11-19.
- [217] Hagemann T, Lawrence T, McNeish I, Charles KA, Kulbe H, *et al.* "Re-educating" tumor-associated macrophages by targeting NF-kappa B. *J Exp Med.* 2008;205(6):1261-1268.
- [218] Colombo MP, Mantovani A. Targeting myelomonocytic cells to revert inflammation-dependent cancer promotion. *Cancer Res.* 2005;65(20):9113-9116.
- [219] Saccani A, Schioppa T, Porta C, Biswas SK, Nebuloni M, *et al.* P50 nuclear factor-kappa B overexpression in tumor-associated macrophages inhibits M1 inflammatory responses and antitumor resistance. *Cancer Res.* 2006;66(23):11432-11440.
- [220] Kusmartsev S, Gabrilovich DI. STAT1 signaling regulates tumor-associated macrophage-mediated T cell deletion. *J Immunol.* 2005;174(8):4880-4891.
- [221] Torrero MN XX, Henk W, Yu S, Li S. Stat1 deficiency in the host enhances interleukin-12-mediated tumor regression. *Cancer Res.* 2006;66(8).
- [222] Hanada T, Kobayashi T, Chinen T. IFNgamma-dependent, spontaneous development of colorectal carcinomas in SOCS1-deficient mice. *J Exp Med.* 2006;203(6):1391-1397.
- [223] Kortylewski M, Kujawsk M, Wang T. Inhibiting Stat3 signaling in the hematopoietic system elicits multicomponent antitumor immunity. *Nat Med.* 2005;11(13):14-21.
- [224] Sinha P, Clements VK, Miller S, Ostrand-Rosenberg S. Tumor immunity: a balancing act between T cell activation, macrophage activation and tumor-induced immune suppression. *Cancer Immunol Immunother.* 2005;54(11):1137-1142.
- [225] Knight SC, Stagg A, Hill S, Fryer P, Griffiths S. Development and function of dendritic cells in health and disease. *J Invest Dermatol.* 1992;99(5):33S-38S.
- [226] Lipscomb MF, Masten BJ. Dendritic cells: immune regulators in health and disease. *Physiol Rev.* 2002;82(1):97-130.
- [227] Kim R, Emi M, Tanabe K. Functional roles of immature dendritic cells in impaired immunity of solid tumour and their targeted strategies for provoking tumour immunity. *Clin Exp Immunol.* 2006;146(2):189-196.
- [228] Fricke I, Gabrilovich DI. Dendritic cells and tumor microenvironment: a dangerous liaison. *Immunol Invest.* 2006;35(3-4):459-483.
- [229] Ullrich E, Menard C, Flament C, Terme M, Mignot G, *et al.* Dendritic cells and innate defense against tumor cells. *Cytokine Growth Factor Rev.* 2008;19(1):79-92.
- [230] Ma Y, Aymeric L, Locher C, Kroemer G, Zitvogel L. The dendritic cell-tumor cross-talk in cancer. *Curr Opin Immunol.* 2011;23(1):146-152.
- [231] Hurwitz AA, Watkins SK. Immune suppression in the tumor microenvironment: a role for dendritic cell-mediated tolerization of T cells. *Cancer Immunol Immunother.* 2012;61(2):289-293.
- [232] Ma Y, Shurin GV, Gutkin DW, Shurin MR. Tumor associated regulatory dendritic cells. *Semin Cancer Biol.* 2012.

- [233] Lin A, Schildknecht A, Nguyen LT, Ohashi PS. Dendritic cells integrate signals from the tumor microenvironment to modulate immunity and tumor growth. *Immunol Lett.* 2010;127(2):77-84.
- [234] Kusmartsev S, Gabrilovich DI. Role of immature myeloid cells in mechanisms of immune evasion in cancer. *Cancer Immunol Immunother.* 2006;55(3):237-245.
- [235] Gabriel A, Rabinovich DG, Eduardo M, Sotomayor. Immunosuppressive Strategies that are Mediated by Tumor Cells. *Ann Rev Immunol.* 2007;25::267–296.
- [236] Talmadge JE, Donkor M, Scholar E. Inflammatory cell infiltration of tumors: Jekyll or Hyde. *Cancer Metast Rev.* 2007;26(3-4):373-400.
- [237] Marigo I, Dolcetti L, Serafini P, Zanovello P, Bronte V. Tumor-induced tolerance and immune suppression by myeloid derived suppressor cells. *Immunol Rev.* 2008;222:162-179.
- [238] Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol.* 2009;9(3):162-174.
- [239] Ribechini E, Greifengberg V, Sandwick S, Lutz MB. Subsets, expansion and activation of myeloid-derived suppressor cells. *Med Microbiol Immunol.* 2010;199(3):273-281.
- [240] Pastula A, Marcinkiewicz J. Myeloid-derived suppressor cells: a double-edged sword? *Int J Exp Pathol.* 2011;92(2):73-78.
- [241] Filipazzi P, Huber V, Rivoltini L. Phenotype, function and clinical implications of myeloid-derived suppressor cells in cancer patients. *Cancer Immunol Immunother.* 2012;61(2):255-263.
- [242] Kiavash M, Damyra L, Conny G. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer Res.* 2010;70(14):5728-5739.
- [243] Youn JI, Nagaraj S, Collazo M, Gabrilovich DI. Subsets of myeloid-derived suppressor cells in tumor-bearing mice. *J Immunol.* 2008;181(8):5791-5802.
- [244] LeCouter C, Zlot C, Tejada M, Peale F. MDSC are generally defined as CD14- CD11b+ cells or as cells that express the common myeloid marker CD33 but lack mature myeloid and lymphoid cells. *Proc Nat Acad Sci USA.* 2004;101(48):16813-16818
- [245] Shojaei F, Singh M, Thompson JD, Ferrara N. Role of Bv8 in neutrophil-dependent angiogenesis in a transgenic model of cancer progression. *Proc Nat Acad Sci USA* 2008;105(7):2640-2645.
- [246] Lewis C, Murdoch C. Macrophage responses to hypoxia - Implications for tumor progression and anti-cancer therapies. *Am J Pathol.* 2005;167(3):627-635.
- [247] Talmadge JE. Pathways mediating the expansion and immuno-suppressive activity of myeloid derived suppressor cells and their relevance. *Clin Cancer Res.* 2007 13(18):5243-5248.
- [248] Bronte V, Apolloni E, Cabrelle A, Ronca R, Serafini P, *et al.* Identification of a CD11b(+)/Gr-1(+)/CD31(+) myeloid progenitor capable of activating or suppressing CD8(+) T cells. *Blood.* 2000;96(12):3838-3846.
- [249] Yang L, Huang J, Ren X, Gorska AE, Chytil A, *et al.* Abrogation of TGF beta signaling in mammary carcinomas recruits Gr-1+CD11b+ myeloid cells that promote metastasis. *Cancer Cell.* 2008;13(1):23-35.
- [250] Pekarek LA, Starr BA, Toledano AY, Schreiber H. Inhibition of Tumor-Growth by Elimination of Granulocytes. *J Exp Med.* 1995;181(1):435-440.
- [251] Michels T, Shurin GV, Naiditch H, Sevko A, Umansky V, *et al.* Paclitaxel promotes differentiation of myeloid-derived suppressor cells into dendritic cells *in vitro* in a TLR4-independent manner. *J Immunotoxicol.* 2012.

- [252] Shirota Y, Shirota H, Klinman DM. Intratumoral injection of CpG oligonucleotides induces the differentiation and reduces the immunosuppressive activity of myeloid-derived suppressor cells. *J Immunol.* 2012;188(4):1592-1599.
- [253] Kao J, Ko EC, Eisenstein S, Sikora AG, Fu S, *et al.* Targeting immune suppressing myeloid-derived suppressor cells in oncology. *Crit Rev Oncol Hematol.* 2011;77(1):12-19.
- [254] Apetoh L, Vegran F, Ladoire S, Ghiringhelli F. Restoration of antitumor immunity through selective inhibition of myeloid derived suppressor cells by anticancer therapies. *Curr Mol Med.* 2011;11(5):365-372.
- [255] Ko JS, Rayman P, Ireland J, Swaidani S, Li G, *et al.* Direct and differential suppression of myeloid-derived suppressor cell subsets by sunitinib is compartmentally constrained. *Cancer Res.* 2010;70(9):3526-3536.
- [256] Ko JS, Bukowski RM, Fincke JH. Myeloid-derived suppressor cells: a novel therapeutic target. *Curr Oncol Rep.* 2009;11(2):87-93.
- [257] Corzo CA, Condamine T, Lu L, Cotter MJ, Youn JI, *et al.* HIF-1 alpha regulates function and differentiation of myeloid-derived suppressor cells in the tumor microenvironment. *J Exp Med.* 2010;207(11):2439-2453.
- [258] Di Carlo E, Forni G, Lollini P. The intriguing role of polymorphonuclear neutrophils in antitumor reactions. *Blood.* 2001;97(2):339-345.
- [259] Houghton AM. The paradox of tumor-associated neutrophils: fueling tumor growth with cytotoxic substances. *Cell Cycle.* 2010;9(9):1732-1737.
- [260] Bellocq A, Antoine M, Flahault A, Philippe C, Crestani B, *et al.* Neutrophil alveolitis in bronchioloalveolar carcinoma - Induction by tumor-derived interleukin-8 and relation to clinical outcome. *Am J Pathol.* 1998;152(1):83-92.
- [261] Caruso RA, Bellocco R, Pagano M. Prognostic value of intratumoral neutrophils in advanced gastric carcinoma in a high-risk area in northern Italy. *Modern Pathol.* 2002;15(8):831-837.
- [262] Schmidt H, Suci S, Punt CJA, Gore M, Kruit W, *et al.* Pretreatment levels of peripheral neutrophils and leukocytes as independent predictors of overall survival in patients with American joint committee on cancer stage IV melanoma: Results of the EORTC 18951 biochemotherapy trial. *J Clin Oncol.* 2007;25(12):1562-1569.
- [263] Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng GJ, *et al.* Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell.* 2009;16(3):183-194.
- [264] Shen L, Smith JM, Shen Z, Eriksson M, Sentman C, *et al.* Inhibition of human neutrophil degranulation by transforming growth factor-beta 1. *Clin Exp Immunol.* 2007;149(1):155-161.
- [265] Bierie B, Chung CH, Parker JS. Abrogation of TGF-beta signaling enhances chemokine production and correlates with prognosis in human breast cancer. *J Clin Invest.* 2009;119(6):1571-1582.
- [266] Jablonska J, Leschner S, Westphal K, Lienenklaus S, Weiss S. Neutrophils responsive to endogenous IFN-beta regulate tumor angiogenesis and growth in a mouse tumor model. *J Clin Invest.* 2010;120(4):1151-1164.
- [267] Lesterhuis WJ, Haanen JB, Punt CJ. Cancer immunotherapy--revisited. *Nat Rev Drug Discov.* 2011;10(8):591-600.
- [268] Stewart TJ, Smyth MJ. Improving cancer immunotherapy by targeting tumor-induced immune suppression. *Cancer Metast Rev.* 2011;30(1):125-140.
- [269] Vose BM, Moore M. Human tumor-infiltrating lymphocytes: a marker of host response. *Semin Hematol.* 1985;22(1):27-40.

- [270] Rosenberg SA. The development of new immunotherapies for the treatment of cancer using interleukin-2. A review. *Ann Surg.* 1988;208(2):121-135.
- [271] Parkinson DR. Lessons from the clinical trials of interleukin-2. *Nat Immun Cell Growth Regul.* 1990;9(4):242-252.
- [272] Ioannides CG, Whiteside TL. T cell recognition of human tumors: implications for molecular immunotherapy of cancer. *Clin Immunol Immunopathol.* 1993;66(2):91-106.
- [273] Hershkovitz L, Schachter J, Treves AJ, Besser MJ. Focus on adoptive T cell transfer trials in melanoma. *Clin Dev Immunol.* 2010;2010:260267.
- [274] Chang AE, Shu S. Current status of adoptive immunotherapy of cancer. *Crit Rev Oncol Hematol.* 1996;22(3):213-228.
- [275] Rosenberg SA. Development of cancer immunotherapies based on identification of the genes encoding cancer regression antigens. *J Natl Cancer Inst.* 1996;88(22):1635-1644.
- [276] Melief CJ, Toes RE, Medema JP, van der Burg SH, Ossendorp F, *et al.* Strategies for immunotherapy of cancer. *Adv Immunol.* 2000;75:235-282.
- [277] Paul S, Calmels B, Acres RB. Improvement of adoptive cellular immunotherapy of human cancer using ex-vivo gene transfer. *Curr Gene Ther.* 2002;2(1):91-100.
- [278] McKee MD, Fichera A, Nishimura MI. T cell immunotherapy. *Front Biosci.* 2007;12:919-932.
- [279] Lampen MH, van Hall T. Strategies to counteract MHC-I defects in tumors. *Curr Opin Immunol.* 2011;23(2):293-298.
- [280] Engleman EG. Dendritic cells: potential role in cancer therapy. *Cytotechnology.* 1997;25(1-3):1-8.
- [281] Ada G. The coming of age of tumour immunotherapy. *Immunol Cell Biol.* 1999;77(2):180-185.
- [282] Meidenbauer N, Andreesen R, Mackensen A. Dendritic cells for specific cancer immunotherapy. *Biol Chem.* 2001;382(4):507-520.
- [283] Gabrilovich DI. Dendritic cell vaccines for cancer treatment. *Curr Opin Mol Ther.* 2002;4(5):452-458.
- [284] Mocellin S, Rossi CR, Nitti D. Cancer vaccine development: on the way to break immune tolerance to malignant cells. *Exp Cell Res.* 2004;299(2):267-278.
- [285] Gilboa E. DC-based cancer vaccines. *J Clin Invest.* 2007;117(5):1195-1203.
- [286] Nencioni A, Grunebach F, Schmidt SM, Muller MR, Boy D, *et al.* The use of dendritic cells in cancer immunotherapy. *Crit Rev Oncol Hematol.* 2008;65(3):191-199.
- [287] Sabado RL, Bhardwaj N. Directing dendritic cell immunotherapy towards successful cancer treatment. *Immunotherapy.* 2010;2(1):37-56.
- [288] Ilett EJ, Prestwich RJ, Melcher AA. The evolving role of dendritic cells in cancer therapy. *Expert Opin Biol Ther.* 2010;10(3):369-379.
- [289] Speiser DE, Romero P. Molecularly defined vaccines for cancer immunotherapy, and protective T cell immunity. *Semin Immunol.* 2010;22(3):144-154.
- [290] Apetoh L, Locher C, Ghiringhelli F, Kroemer G, Zitvogel L. Harnessing dendritic cells in cancer. *Semin Immunol.* 2011;23(1):42-49.
- [291] Palucka K, Ueno H, Roberts L, Fay J, Banchereau J. Dendritic cell subsets as vectors and targets for improved cancer therapy. *Curr Top Microbiol Immunol.* 2011;344:173-192.
- [292] Hammerstrom AE, Cauley DH, Atkinson BJ, Sharma P. Cancer immunotherapy: sipuleucel-T and beyond. *Pharmacotherapy.* 2011;31(8):813-828.
- [293] Jahnisch H, Fussel S, Kiessling A, Wehner R, Zastrow S, *et al.* Dendritic cell-based immunotherapy for prostate cancer. *Clin Dev Immunol.* 2010;2010:517493.

- [294] Sica A, Bronte V. Altered macrophage differentiation and immune dysfunction in tumor development. *J Clin Invest.* 2007;117(5):1155-1166.
- [295] Guiducci C, Vicari AP, Sangaletti S. Redirecting *in vivo* elicited tumor infiltrating macrophages and dendritic cells towards tumor rejection. *Cancer Res.* 2005;65(8):3437-3446.
- [296] Buhtoiarov IN, Sondel PM, Wigginton JM, Buhtoiarova TN, Yanke EM, *et al.* Anti-tumour synergy of cytotoxic chemotherapy and anti-CD40 plus CpG-ODN immunotherapy through repolarization of tumour-associated macrophages. *Immunology.* 2011;132(2):226-239.
- [297] Sinha P, Clements VK, Ostrand-Rosenberg S. Reduction of myeloid-derived suppressor cells and induction of M1 macrophages facilitate the rejection of established metastatic disease. *J Immunol.* 2005;174(2):636-645.
- [298] Ostrand-Rosenberg S, Clements VK, Terabe M. Resistance to metastatic disease in STAT6-Deficient mice requires hemopoietic and nonhemopoietic cells and is IFN-gamma Dependent(1). *J Immunol.* 2002;169(10):5796-5804.
- [299] Rauh MJ, Sly LM, Kalesnikoff J, Hughes MR, Cao LP, *et al.* The role of SHIP1 in macrophage programming and activation. *Biochem Soc Transact.* 2004;32:785-788.
- [300] Rolny C, Mazzone M, Tugues S. HRG inhibits tumor growth and metastasis by inducing macrophage polarization and vessel normalization through downregulation of PlGF. *Cancer Cell.* 2011;19(1):31-44.
- [301] Aharinejad S, Paulus P, Sioud M, Hofmann M, Zins K, *et al.* Colony-stimulating factor-1 blockade by antisense oligonucleotides and small interfering RNAs suppresses growth of human mammary tumor xenografts in mice. *Cancer Res.* 2004;64(15):5378-5384.
- [302] Denardo DG, Brennan DJ, Rexhepaj E, Ruffell B, Shiao SL, *et al.* Leukocyte complexity predicts breast cancer survival and functionally regulates response to chemotherapy. *Cancer Discov.* 2011;1:54-67.
- [303] Lin EY, Li JF, Gnatovskiy L, Deng Y, Zhu L, *et al.* Macrophages regulate the angiogenic switch in a mouse model of breast cancer. *Cancer Res.* 2006;66(23):11238-11246.
- [304] Camilleri JP, Williams AS, Amos N, Douglas-Jones AG, Love WG, *et al.* Methods for assessing splenic macrophage depletion by liposome encapsulated clodronate. *Inflamm Res.* 1995;44(4):152-157.
- [305] Seiler P, Aichele P, Odermatt B, Hengartner H, Zinkernagel RM, *et al.* Crucial role of marginal zone macrophages and marginal zone metallophilic cells in the clearance of lymphocytic choriomeningitis virus infection. *Eur J Immunol.* 1997;27(10):2626-2633.
- [306] Roscic-Mrkic B, Schwendener RA, Odermatt B, Zuniga A, Pavlovic J, *et al.* Roles of macrophages in measles virus infection of genetically modified mice. *J Virol.* 2001;75(7):3343-3351.
- [307] Schwendener RA. Liposomes in biology and medicine. *Adv Exp Med Biol.* 2007;620:117-128.
- [308] Aichele P, Zinke J, Grode L, Schwendener RA, Kaufmann SH, *et al.* Macrophages of the splenic marginal zone are essential for trapping of blood-borne particulate antigen but dispensable for induction of specific T cell responses. *J Immunol.* 2003;171(3):1148-1155.
- [309] Beck-Schimmer B, Schwendener R, Pasch T, Reyes L, Booy C, *et al.* Alveolar macrophages regulate neutrophil recruitment in endotoxin-induced lung injury. *Respir Res.* 2005;6:61.
- [310] Zattoni M, Mura ML, Deprez F, Schwendener RA, Engelhardt B, *et al.* Brain infiltration of leukocytes contributes to the pathophysiology of temporal lobe epilepsy. *J Neurosci.* 2011;31(11):4037-4050.

- [311] Monkkonen J, Urtti A, Paronen P, Elo HA, Ylitalo P. The uptake of clodronate (dichloromethylene bisphosphonate) by macrophages *in vivo* and *in vitro*. *Drug Metab Dispos.* 1989;17(6):690-693.
- [312] Chapurlat RD, Delmas PD. Drug insight: bisphosphonates for postmenopausal osteoporosis. *Nat Clin Pract Endocrinol Metabol.* 2006;2(4):211-219.
- [313] Dougados RaM. Treatment of patients with Paget's disease of bone. *Drugs.* 1999;58(5):823-830.
- [314] Coleman RE. The role of bisphosphonates in breast cancer. *Breast.* 2004;13(S1):19-28.
- [315] Rosen LS, Gordon D, Tchekmedyian S. Zoledronic acid *versus* placebo in the treatment of skeletal metastases in patients with lung cancer and other solid tumors: a phase III, double-blind, randomized trial--the Zoledronic Acid Lung Cancer and Other Solid Tumors Study Group. *J Clin Oncol* 2003;21(16):3150-3157.
- [316] Russell RGG, Rogers MJ. Bisphosphonates: From the laboratory to the clinic and back again. *Bone.* 1999;25(1):97-106.
- [317] Roelofs AJ, Thompson K, Gordon S, Rogers MJ. Molecular mechanisms of action of bisphosphonates: Current status. *Clin Cancer Res.* 2006;12(20):6222S-6230S.
- [318] Frith JC, Monkkonen J, Blackburn GM. Clodronate and liposome-encapsulated clodronate are metabolized to a toxic ATP analog, adenosine 5'-(beta,gamma-dichloromethylene) triphosphate, by mammalian cells *in vitro*. *J Bone Mineral Res.* 1997;12(9):1358-1367.
- [319] Rogers MJ. From molds and macrophages to mevalonate: A decade of progress in understanding the molecular mode of action of bisphosphonates. *Calcified Tissue Int.* 2004;75(6):451-461.
- [320] Monkkonen H, Ottewell PD, Kuokkanen J. Zoledronic acid-induced IPP/ApppI production *in vivo*. *Life Sci.* 2007;81:1066-1070.
- [321] Galvez-Munoz E, Rodriguez-Lescure A. The role of bisphosphonates of adjuvant therapy in breast cancer. *Med Clin (Barc).* 2010;135(2):70-4.
- [322] Rosen LS, Gordon D, Dugan W. Zoledronic acid is superior to pamidronate for the treatment of bone metastases in breast carcinoma patients with at least one osteolytic lesion. *Cancer.* 2004;100(1):36-43.
- [323] Benford HL, Frith JC, Auriola S, Monkkonen J, Rogers MJ. Farnesol and geranylgeraniol prevent activation of caspases by aminobisphosphonates: Biochemical evidence for two distinct pharmacological classes of bisphosphonate drugs. *Mol Pharmacol.* 1999;56(1):131-140.
- [324] Lipton A, Colombo-Berra A, Bukowski RM, Rosen L, Zheng M, *et al.* Skeletal complications in patients with bone metastases from renal cell carcinoma and therapeutic benefits of zoledronic acid. *Clin Cancer Res.* 2004;10(18):6397S-6403S.
- [325] Ibrahim A, Scher N, Williams G. Approval summary for zoledronic acid for treatment of multiple myeloma and cancer bone metastasis. *Clin Cancer Res.* 2003;9(7):2394-2399.
- [326] Jagdev SP, Coleman RE, Shipman CM. The bisphosphonate, zoledronic acid, induces apoptosis of breast cancer cells: evidence for synergy with paclitaxel. *Br J Cancer.* 2001;84(8):1126-1134.
- [327] Lee MV, Fong EM, Singer FR. Bisphosphonate treatment inhibits the growth of prostate cancer cells. *Cancer Res.* 2001;61(6):2602-2608.
- [328] Mackle PS, Fisher JS, Zhou H, Choong PFM. Bisphosphonates regulate cell growth and gene expression in the UMR 106-01 clonal rat osteosarcoma cell line. *Br J Cancer.* 2001;84(7):951-958.
- [329] Green J, Lipton A. Anticancer properties of zoledronic acid. *Cancer Invest.* 2010;28(9):944-957.



- [330] Giraud E, Inoue M, Hanahan D. An amino-bisphosphonate targets MMP-9-expressing macrophages and angiogenesis to impair cervical carcinogenesis. *J Clin Invest.* 2004;114(5):623-633.
- [331] Wakchoure S, Merrell MA, Aldrich W, Millender-Swain T, Harris KW, *et al.* Bisphosphonates inhibit the growth of mesothelioma cells *in vitro* and *in vivo*. *Clin Cancer Res.* 2006;12(9):2862-2868.
- [332] Coscia M, Quaglino E, Iezzi M. Zoledronic acid repolarizes tumour-associated macrophages and inhibits mammary carcinogenesis by targeting the mevalonate pathway. *J Cell Mol Med.* 2010;14(12):2803-2815.
- [333] Tanaka Y, Morita CT, Tanaka Y, Nieves E, Brenner MB, *et al.* Natural and synthetic nonpeptide antigens recognized by human gamma-delta T-cells. *Nature.* 1995;375(6527):155-158.
- [334] Castella B, Vitale C, Coscia M, Massaia M. Vgamma9Vdelta2 T cell-based immunotherapy in hematological malignancies: from bench to bedside. *Cell Mol Life Sci.* 2011;68(14):2419-2432.
- [335] Gomes AQ, Martins DS, Silva-Santos B. Targeting gammadelta T lymphocytes for cancer immunotherapy: from novel mechanistic insight to clinical application. *Cancer Res.* 2010;70(24):10024-10027.
- [336] Zocchi MR, Poggi A. Role of gammadelta T lymphocytes in tumor defense. *Front Bioscience.* 2004;9:2588-2604.
- [337] Sato K, Kimura S, Segawa H, Yokota A, Matsumoto S, *et al.* Cytotoxic effects of gamma delta T cells expanded *ex vivo* by a third generation bisphosphonate for cancer immunotherapy. *Int J Cancer.* 2005;116(1):94-99.
- [338] Wesch D, Pitters E, Zoller M, Kebelitz D. Characterization of tumor reactivity of human V gamma 9V delta 2 gamma delta T-cells *in vitro* and in severe combined immunodeficiency mice *in vivo*. *Immunobiology.* 2004;209(4-6):316-317.
- [339] Zheng BJ, Chan KW, Im S, Chua D, Sham JST, *et al.* Anti-tumor effects of human peripheral gamma delta T cells in a mouse tumor model. *International J Cancer.* 2001;92(3):421-425.
- [340] Clezardin P, Massaia M. Nitrogen-containing bisphosphonates and cancer immunotherapy. *Curr Pharm Des.* 2010;16(27):3007-2014.
- [341] Dieli F, Gebbia N, Poccia F, Caccamo N, Montesano C, *et al.* Induction of gamma delta T-lymphocyte effector functions by bisphosphonate zoledronic acid in cancer patients *in vivo*. *Blood.* 2003;102(6):2310-2311.
- [342] Mattarollo SR, Kenna T, Nieda M, Nicol AJ. Chemotherapy and zoledronate sensitize solid tumour cells to V gamma 9V delta 2 T cell cytotoxicity. *Cancer Immunol Immunother.* 2007;56(8):1285-1297.
- [343] Green AE, Lissina A, Hutchinson SL, Hewitt RE, Temple B, *et al.* Recognition of nonpeptide antigens by human V gamma 9V delta 2 T cells requires contact with cells of human origin. *Clin Exp Immunol.* 2004;136(3):472-482.
- [344] Pfeilschifter J, Bonewald L, Mundy GR. Characterization of the latent transforming growth factor-beta complex in bone. *J Bone Mineral Res.* 1990;5(1):49-58.
- [345] Massague J. TGF beta in cancer. *Cell.* 2008;134(2):215-230.
- [346] Chirgwin JM, Guise TA. Skeletal metastases: Decreasing tumor burden by targeting the bone microenvironment. *J Cell Biochem.* 2007;102(6):1333-1342.

- [347] Shim KS, Kim KH, Han WS, Park EB. Elevated serum levels of transforming growth factor-beta 1 in patients with colorectal carcinoma - Its association with tumor progression and its significant decrease after curative surgical resection. *Cancer*. 1999;85(3):554-561.
- [348] Barthelemy-Brichant N, David JL, Bosquee L, Bury T, Seidel L, *et al.* Increased TGF beta(1) plasma level in patients with lung cancer: potential mechanisms. *Eur J Clin Invest*. 2002;32(3):193-198.
- [349] Krasagakis K, Tholke D, Farthmann B, Eberle J, Mansmann U, *et al.* Elevated plasma levels of transforming growth factor (TGF)-beta 1 and TGF-beta 2 in patients with disseminated malignant melanoma. *Br J Cancer*. 1998;77(9):1492-1494.
- [350] Gnant M, Mlineritsch B, Schippinger W, Luschin-Ebengreuth G, Postlberger S, *et al.* Endocrine Therapy plus Zoledronic Acid in Premenopausal Breast Cancer (vol 360, pg 679, 2009). *New Engl J Med*. 2009;360(22):2379-2379.
- [351] Zeisberger SM, Odermatt B, Marty C, Zehnder-Fjallman AH, Ballmer-Hofer K, *et al.* Clodronate-liposome-mediated depletion of tumour-associated macrophages: a new and highly effective antiangiogenic therapy approach. *Br J Cancer*. 2006;95(3):272-281.
- [352] Shmeeda H, Amitay Y, Gorin J, Tzemach D, Mak L, *et al.* Delivery of zoledronic acid encapsulated in folate-targeted liposome results in potent *in vitro* cytotoxic activity on tumor cells. *J Control Release*. 2010;146(1):76-83.
- [353] Marra M, Salzano G, Leonetti C, Tassone P, Scarsella M, *et al.* Nanotechnologies to use bisphosphonates as potent anticancer agents: the effects of zoledronic acid encapsulated into liposomes. *Nanomedicine*. 2011;7(6):955-964.
- [354] Zhang W, Zhu XD, Sun HC, Xiong YQ, Zhuang PY, *et al.* Depletion of tumor-associated macrophages enhances the effect of sorafenib in metastatic liver cancer models by antimetastatic and antiangiogenic effects. *Clin Cancer Res*. 2010;16(13):3420-3430.
- [355] Kuijpers SA, Coimbra MJ, Storm G, Schiffelers RM. Liposomes targeting tumour stromal cells. *Mol Membr Biol*. 2010;27(7):328-340.
- [356] Coimbra M, Crielaard BJ, Storm G, Schiffelers RM. Critical factors in the development of tumor-targeted anti-inflammatory nanomedicines. *J Control Release*. 2012;160(2):232-238.
- [357] Kaasgaard T, Andresen TL. Liposomal cancer therapy: exploiting tumor characteristics. *Expert Opin Drug Deliv*. 2010;7(2):225-243.
- [358] Gabizon AA. Pegylated liposomal doxorubicin: metamorphosis of an old drug into a new form of chemotherapy. *Cancer Invest*. 2001;19(4):424-436.
- [359] Marty C, Odermatt B, Schott H, Neri D, Ballmer-Hofer K, *et al.* Cytotoxic targeting of F9 teratocarcinoma tumours with anti-ED-B fibronectin scFv antibody modified liposomes. *Br J Cancer*. 2002;87(1):106-112.
- [360] Fall BI, Niessner R. Detection of known allergen-specific IgE antibodies by immunological methods. *Methods Mol Biol*. 2009;509:107-122.
- [361] Mamot C, Drummond DC, Noble CO, Kallab V, Guo Z, *et al.* Epidermal growth factor receptor-targeted immunoliposomes significantly enhance the efficacy of multiple anticancer drugs *in vivo*. *Cancer Res*. 2005;65(24):11631-11638.
- [362] Schroeder A, Heller DA, Winslow MM, Dahlman JE, Pratt GW, *et al.* Treating metastatic cancer with nanotechnology. *Nat Rev Cancer*. 2012;12(1):39-50.
- [363] Nasser MW, Qamri Z, Deol YS, Ravi J, Powell CA, *et al.* S100A7 Enhances Mammary Tumorigenesis through Upregulation of Inflammatory Pathways. *Cancer Res*. 2012.
- [364] Zaynagetdinov R, Sherrill TP, Polosukhin VV, Han W, Ausborn JA, *et al.* A critical role for macrophages in promotion of urethane-induced lung carcinogenesis. *J Immunol*. 2011;187(11):5703-5711.

- [365] Yang H, Kim C, Kim MJ, Schwendener RA, Alitalo K, *et al.* Soluble vascular endothelial growth factor receptor-3 suppresses lymphangiogenesis and lymphatic metastasis in bladder cancer. *Mol Cancer*. 2011;10:36.
- [366] Wang B, Li Q, Qin L, Zhao S, Wang J, *et al.* Transition of tumor-associated macrophages from MHC class II(hi) to MHC class II(low) mediates tumor progression in mice. *BMC Immunol*. 2011;12:43.
- [367] Meng Y, Beckett MA, Liang H, Mauceri HJ, van Rooijen N, *et al.* Blockade of tumor necrosis factor alpha signaling in tumor-associated macrophages as a radiosensitizing strategy. *Cancer Res*. 2010;70(4):1534-1543.
- [368] Haynes NM, Hawkins ED, Li M, McLaughlin NM, Hammerling GJ, *et al.* CD11c+ dendritic cells and B cells contribute to the tumoricidal activity of anti-DR5 antibody therapy in established tumors. *J Immunol*. 2010;185(1):532-541.
- [369] Westwood JA, Haynes NM, Sharkey J, McLaughlin N, Pegram HJ, *et al.* Toll-Like Receptor Triggering and T-Cell Costimulation Induce Potent Antitumor Immunity in Mice. *Clin Cancer Res*. 2009;15(24):7624-7633.
- [370] Lepique AP, Daghestanli KR, Cuccovia IM, Villa LL. HPV16 tumor associated macrophages suppress antitumor T cell responses. *Clin Cancer Res*. 2009;15(13):4391-4400.
- [371] Jeon BH, Jang C, Han J, Kataru RP, Piao L, *et al.* Profound but dysfunctional lymphangiogenesis *via* vascular endothelial growth factor ligands from CD11b+ macrophages in advanced ovarian cancer. *Cancer Res*. 2008;68(4):1100-1109.
- [372] Hiraoka K, Zenmyo M, Watari K, Iguchi H, Fotovati A, *et al.* Inhibition of bone and muscle metastases of lung cancer cells by a decrease in the number of monocytes/macrophages. *Cancer Sci*. 2008;99(8):1595-1602.
- [373] Lichtenberger BM, Tan PK, Niederleithner H, Ferrara N, Petzelbauer P, *et al.* Autocrine VEGF signaling synergizes with EGFR in tumor cells to promote epithelial cancer development. *Cell*. 2010;140(2):268-279.
- [374] Cavallo F, Giovarelli M, Gulino A, Vacca A, Stoppacciaro A, *et al.* Role of Neutrophils and Cd4+ Lymphocytes-T in the Primary and Memory Response to Nonimmunogenic Murine Mammary Adenocarcinoma Made Immunogenic by Il-2 Gene. *J Immunol*. 1992;149(11):3627-3635.
- [375] Di Carlo E, Forni G, Lollini P. The intriguing role of polymorphonuclear neutrophils in antitumor reactions. *Blood*. 2001;97(2):339-345.
- [376] Musiani P, Allione A, Modica A, Lollini PL, Giovarelli M, *et al.* Role of neutrophils and lymphocytes in inhibition of a mouse mammary adenocarcinoma engineered to release IL-2, IL-4, IL-7, IL-10, IFN-alpha, IFN-gamma, and TNF-alpha. *Lab Invest*. 1996;74(1):146-157.
- [377] Cavallo F, Di Carlo E, Butera M. Immune events associated with the cure of established tumors and spontaneous metastases by local and systemic interleukin 12. *Cancer Res*. 1999;59(2):414-421.
- [378] Brennen WN, Isaacs JT, Denmeade SR. Rationale behind targeting fibroblast activation protein-expressing carcinoma-associated fibroblasts as a novel chemotherapeutic strategy. *Mol Cancer Ther*. 2012;11(2):257-266.
- [379] Bonmort M, Ullrich E, Mignot G, Jacobs B, Chaput N, *et al.* Interferon-gamma is produced by another player of innate immune responses: the interferon-producing killer dendritic cell (IKDC). *Biochimie*. 2007;89(6-7):872-877.

- [380] Watanabe T. Treatment strategies for nodal and gastrointestinal follicular lymphoma: current status and future development. *World J Gastroenterol.* 2010;16(44):5543-5554.
- [381] Colombo R, Moll J. Target validation and biomarker identification in oncology : the example of aurora kinases. *Mol Diagn Ther.* 2008;12(2):71-76.
- [382] Galinsky DS, Nechushtan H. Mast cells and cancer--no longer just basic science. *Crit Rev Oncol Hematol.* 2008;68(2):115-130.
- [383] Sullivan LA, Brekken RA. The VEGF family in cancer and antibody-based strategies for their inhibition. *MAbs.* 2010;2(2):165-175.
- [384] Ferrara N. Role of vascular endothelial growth factor in physiologic and pathologic angiogenesis: therapeutic implications. *Semin Oncol.* 2002;29(6 Suppl 16):10-14.
- [385] Ferrara N, Hillan KJ, Novotny W. Bevacizumab (Avastin), a humanized anti-VEGF monoclonal antibody for cancer therapy. *Biochem Biophys Res Commun.* 2005;333(2):328-335.
- [386] Ferrara N. From the discovery of vascular endothelial growth factor to the introduction of avastin in clinical trials - an interview with Napoleone Ferrara by Domenico Ribatti. *Int J Dev Biol.* 2011;55(4-5):383-388.
- [387] Kowanz M, Wu X, Lee J, Tan M, Hagenbeek T, *et al.* Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes. *Proc Natl Acad Sci U S A.* 2010;107(50):21248-21255.
- [388] Heine A, Held SA, Bringmann A, Holderried TA, Brossart P. Immunomodulatory effects of anti-angiogenic drugs. *Leukemia.* 2011;25(6):899-905.
- [389] Cai J, Han S, Qing R, Liao D, Law B, *et al.* In pursuit of new anti-angiogenic therapies for cancer treatment. *Front Biosci.* 2011;16:803-814.
- [390] Goel S, Duda DG, Xu L, Munn LL, Boucher Y, *et al.* Normalization of the vasculature for treatment of cancer and other diseases. *Physiol Rev.* 2011;91(3):1071-1121.
- [391] Fukumura D, Jain RK. Tumor microvasculature and microenvironment: targets for anti-angiogenesis and normalization. *Microvasc Res.* 2007;74(2-3):72-84.
- [392] Fukumura D, Duda DG, Munn LL, Jain RK. Tumor microvasculature and microenvironment: novel insights through intravital imaging in pre-clinical models. *Microcirculation.* 2010;17(3):206-225.
- [393] Nelson AL. Antibody fragments: hope and hype. *MAbs.* 2010;2(1):77-83.
- [394] Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. *Nat Rev Immunol.* 2010;10(5):317-327.
- [395] Vasievich EA, Huang L. The suppressive tumor microenvironment: a challenge in cancer immunotherapy. *Mol Pharm.* 2011;8(3):635-641.
- [396] Houot R, Kohrt HE, Marabelle A, Levy R. Targeting immune effector cells to promote antibody-induced cytotoxicity in cancer immunotherapy. *Trends Immunol.* 2011;32(11):510-516.
- [397] Abes R, Teillaud JL. Modulation of tumor immunity by therapeutic monoclonal antibodies. *Cancer Metastasis Rev.* 2011;30(1):111-124.
- [398] Dalle S, Thieblemont C, Thomas L, Dumontet C. Monoclonal antibodies in clinical oncology. *Anticancer Agents Med Chem.* 2008;8(5):523-532.
- [399] Karyampudi L, Knutson KL. Antibodies in cancer immunotherapy. *Cancer Biomark.* 2010;6(5-6):291-305.
- [400] Muller D, Kontermann RE. Bispecific antibodies for cancer immunotherapy: Current perspectives. *BioDrugs.* 2010;24(2):89-98.

- [401] Chames P, Baty D. Bispecific antibodies for cancer therapy. *Curr Opin Drug Discov Devel.* 2009;12(2):276-283.
- [402] Sharkey RM, Goldenberg DM. Cancer radioimmunotherapy. *Immunotherapy.* 2011;3(3):349-370.
- [403] Choi BD, Cai M, Bigner DD, Mehta AI, Kuan CT, *et al.* Bispecific antibodies engage T cells for antitumor immunotherapy. *Expert Opin Biol Ther.* 2011;11(7):843-853.
- [404] Callahan MK, Wolchok JD, Allison JP. Anti-CTLA-4 antibody therapy: immune monitoring during clinical development of a novel immunotherapy. *Semin Oncol.* 2010;37(5):473-484.
- [405] Agarwala SS, O'Day SJ. Current and future adjuvant immunotherapies for melanoma: blockade of cytotoxic T-lymphocyte antigen-4 as a novel approach. *Cancer Treat Rev.* 2011;37(2):133-142.
- [406] Calabro L, Danielli R, Sigalotti L, Maio M. Clinical studies with anti-CTLA-4 antibodies in non-melanoma indications. *Semin Oncol.* 2010;37(5):460-467.
- [407] Sapoznik S, Hammer O, Ortenberg R, Besser MJ, Ben-Moshe T, *et al.* Novel anti-melanoma immunotherapies: disarming tumor escape mechanisms. *Clin Dev Immunol.* 2012;2012:818214.
- [408] McNeel DG, Smith HA, Eickhoff JC, Lang JM, Staab MJ, *et al.* Phase I trial of tremelimumab in combination with short-term androgen deprivation in patients with PSA-recurrent prostate cancer. *Cancer Immunol Immunother.* 2012;61(7):1137-1147.
- [409] Trail PA, King HD, Dubowchik GM. Monoclonal antibody drug immunoconjugates for targeted treatment of cancer. *Cancer Immunol Immunother.* 2003;52(5):328-337.
- [410] Chen J, Jaracz S, Zhao X, Chen S, Ojima I. Antibody-cytotoxic agent conjugates for cancer therapy. *Expert Opin Drug Deliv.* 2005;2(5):873-890.
- [411] Jurcic JG. What happened to anti-CD33 therapy for acute myeloid leukemia? *Curr Hematol Malig Rep.* 2012;7(1):65-73.
- [412] Alley SC, Okeley NM, Senter PD. Antibody-drug conjugates: targeted drug delivery for cancer. *Curr Opin Chem Biol.* 2010;14(4):529-537.
- [413] Niculescu-Duvaz I. Trastuzumab emtansine, an antibody-drug conjugate for the treatment of HER2+ metastatic breast cancer. *Curr Opin Mol Ther.* 2010;12(3):350-360.
- [414] LoRusso PM, Weiss D, Guardino E, Girish S, Sliwkowski MX. Trastuzumab emtansine: a unique antibody-drug conjugate in development for human epidermal growth factor receptor 2-positive cancer. *Clin Cancer Res.* 2011;17(20):6437-6447.
- [415] Mathew J, Perez EA. Trastuzumab emtansine in human epidermal growth factor receptor 2-positive breast cancer: a review. *Curr Opin Oncol.* 2011;23(6):594-600.
- [416] Foyil KV, Bartlett NL. Brentuximab vedotin for the treatment of CD30+ lymphomas. *Immunotherapy.* 2011;3(4):475-485.
- [417] Oki Y, Younes A. Brentuximab vedotin in systemic T-cell lymphoma. *Expert Opin Biol Ther.* 2012;12(5):623-632.
- [418] Gualberto A. Brentuximab Vedotin (SGN-35), an antibody-drug conjugate for the treatment of CD30-positive malignancies. *Expert Opin Investig Drugs.* 2012;21(2):205-216.
- [419] Minich SS. Brentuximab vedotin: a new age in the treatment of Hodgkin lymphoma and anaplastic large cell lymphoma. *Ann Pharmacother.* 2012;46(3):377-383.
- [420] Iagaru A, Mittra ES, Ganjoo K, Knox SJ, Goris ML. 131I-Tositumomab (Bexxar) vs. 90Y-Ibritumomab (Zevalin) therapy of low-grade refractory/relapsed non-Hodgkin lymphoma. *Mol Imaging Biol.* 2010;12(2):198-203.

- [421] Govindan SV, Goldenberg DM. New antibody conjugates in cancer therapy. *Scientific World Journal*. 2010;10:2070-2089.
- [422] Reichert JM. Marketed therapeutic antibodies compendium. *MAbs*. 2012;4(3):413-415.
- [423] Bagshawe KD. Antibody directed enzymes revive anti-cancer prodrugs concept. *Br J Cancer*. 1987;56(5):531-532.
- [424] Schellmann N, Deckert PM, Bachran D, Fuchs H, Bachran C. Targeted enzyme prodrug therapies. *Mini Rev Med Chem*. 2010;10(10):887-904.
- [425] Bagshawe KD. Antibody-directed enzyme prodrug therapy (ADEPT) for cancer. *Expert Rev Anticancer Ther*. 2006;6(10):1421-1431.
- [426] Bagshawe KD. Targeting: the ADEPT story so far. *Curr Drug Targets*. 2009;10(2):152-157.
- [427] Tietze LF, Schmuck K. Prodrugs for targeted tumor therapies: recent developments in ADEPT, GDEPT and PMT. *Curr Pharm Des*. 2011;17(32):3527-3547.
- [428] Balkwill F. Cancer and the chemokine network. *Nat Rev Cancer*. 2004;4(7):540-550.
- [429] Slettenaar VI, Wilson JL. The chemokine network: a target in cancer biology? *Adv Drug Deliv Rev*. 2006;58(8):962-974.
- [430] Kim-Schulze S, Taback B, Kaufman HL. Cytokine therapy for cancer. *Surg Oncol Clin N Am*. 2007;16(4):793-818.
- [431] Antony GK, Dudek AZ. Interleukin 2 in cancer therapy. *Curr Med Chem*. 2010;17(29):3297-3302.
- [432] Whitworth JM, Alvarez RD. Evaluating the role of IL-12 based therapies in ovarian cancer: a review of the literature. *Expert Opin Biol Ther*. 2011;11(6):751-762.
- [433] Yuzhalin AE, Kutikhin AG. Interleukin-12: Clinical usage and molecular markers of cancer susceptibility. *Growth Factors*. 2012;30(3):176-191.
- [434] Jakobisiak M, Golab J, Lasek W. Interleukin 15 as a promising candidate for tumor immunotherapy. *Cytokine Growth Factor Rev*. 2011;22(2):99-108.
- [435] Srivastava S, Salim N, Robertson MJ. Interleukin-18: biology and role in the immunotherapy of cancer. *Curr Med Chem*. 2010;17(29):3353-3357.
- [436] Hashmi MH, Van Veldhuizen PJ. Interleukin-21: updated review of Phase I and II clinical trials in metastatic renal cell carcinoma, metastatic melanoma and relapsed/refractory indolent non-Hodgkin's lymphoma. *Expert Opin Biol Ther*. 2010;10(5):807-817.
- [437] Yoshimoto T, Morishima N, Okumura M, Chiba Y, Xu M, *et al*. Interleukins and cancer immunotherapy. *Immunotherapy*. 2009;1(5):825-844.
- [438] Steel JC, Waldmann TA, Morris JC. Interleukin-15 biology and its therapeutic implications in cancer. *Trends Pharmacol Sci*. 2012;33(1):35-41.
- [439] Morishima N, Mizoguchi I, Okumura M, Chiba Y, Xu M, *et al*. A pivotal role for interleukin-27 in CD8<sup>+</sup> T cell functions and generation of cytotoxic T lymphocytes. *J Biomed Biotechnol*. 2010;2010:605483.
- [440] Bracarda S, Eggermont AM, Samuelsson J. Redefining the role of interferon in the treatment of malignant diseases. *Eur J Cancer*. 2010;46(2):284-297.
- [441] Breitenstein S, Dimitroulis D, Petrowsky H, Puhon MA, Mullhaupt B, *et al*. Systematic review and meta-analysis of interferon after curative treatment of hepatocellular carcinoma in patients with viral hepatitis. *Br J Surg*. 2009;96(9):975-981.
- [442] Spadaro F, Lapenta C, Donati S, Abalsamo L, Barnaba V, *et al*. IFN-alpha enhances cross-presentation in human dendritic cells by modulating antigen survival, endocytic routing, and processing. *Blood*. 2012;119(6):1407-1417.

- [443] Santini SM, Lapenta C, Santodonato L, D'Agostino G, Belardelli F, *et al.* IFN-alpha in the generation of dendritic cells for cancer immunotherapy. *Handb Exp Pharmacol.* 2009;188:295-317.
- [444] Bracci L, Proietti E, Belardelli F. IFN-alpha and novel strategies of combination therapy for cancer. *Ann N Y Acad Sci.* 2007;1112:256-268.
- [445] Kane A, Yang I. Interferon-gamma in brain tumor immunotherapy. *Neurosurg Clin N Am.* 2010;21(1):77-86.
- [446] Lasfar A, Abushahba W, Balan M, Cohen-Solal KA. Interferon lambda: a new sword in cancer immunotherapy. *Clin Dev Immunol.* 2011;2011:349575.
- [447] Tagawa M, Kawamura K, Li Q, Tada Y, Hiroshima K, *et al.* A possible anticancer agent, type III interferon, activates cell death pathways and produces antitumor effects. *Clin Dev Immunol.* 2011;2011:479013.
- [448] Merhavi-Shoham E, Haga-Friedman A, Cohen CJ. Genetically modulating T-cell function to target cancer. *Semin Cancer Biol.* 2011.
- [449] Westwood JA, Berry LJ, Wang LX, Duong CP, Pegram HJ, *et al.* Enhancing adoptive immunotherapy of cancer. *Expert Opin Biol Ther.* 2010;10(4):531-545.
- [450] Morgan RA, Dudley ME, Wunderlich JR, Hughes MS, Yang JC, *et al.* Cancer regression in patients after transfer of genetically engineered lymphocytes. *Science.* 2006;314(5796):126-129.
- [451] Wang Z, Rao DD, Senzer N, Nemunaitis J. RNA interference and cancer therapy. *Pharm Res.* 2011;28(12):2983-2995.
- [452] Aharinejad S, Sioud M, Lucas T, Abraham D. Targeting stromal-cancer cell interactions with siRNAs. *Methods Mol Biol.* 2009;487:243-266.
- [453] Aharinejad S, Sioud M, Lucas T, Abraham D. Target validation using RNA interference in solid tumors. *Methods Mol Biol.* 2007;361:227-238.
- [454] Bora RS, Gupta D, Mukkur TK, Saini KS. RNA interference therapeutics for cancer: challenges and opportunities (review). *Mol Med Report.* 2012;6(1):9-15.
- [455] Huang L, Liu Y. *In vivo* delivery of RNAi with lipid-based nanoparticles. *Annu Rev Biomed Eng.* 2011;13:507-530.
- [456] Tan SJ, Kiatwuthinon P, Roh YH, Kahn JS, Luo D. Engineering nanocarriers for siRNA delivery. *Small.* 2011;7(7):841-856.
- [457] Burnett JC, Rossi JJ. RNA-based therapeutics: current progress and future prospects. *Chem Biol.* 2012;19(1):60-71.
- [458] Petrocca F, Lieberman J. Promise and challenge of RNA interference-based therapy for cancer. *J Clin Oncol.* 2011;29(6):747-754.
- [459] Tong AW, Zhang YA, Nemunaitis J. Small interfering RNA for experimental cancer therapy. *Curr Opin Mol Ther.* 2005;7(2):114-124.
- [460] Phadke AP, Jay CM, Wang Z, Chen S, Liu S, *et al.* *In vivo* safety and antitumor efficacy of bifunctional small hairpin RNAs specific for the human Stathmin 1 oncoprotein. *DNA Cell Biol.* 2011;30(9):715-726.
- [461] Gomes-da-Silva LC, Fonseca NA, Moura V, Pedrosa de Lima MC, Simoes S, *et al.* Lipid-Based Nanoparticles for siRNA Delivery in Cancer Therapy: Paradigms and Challenges. *Acc Chem Res.* 2012.
- [462] Chono S, Li SD, Conwell CC, Huang L. An efficient and low immunostimulatory nanoparticle formulation for systemic siRNA delivery to the tumor. *J Control Release.* 2008;131(1):64-69.

- [463] Cukierman E, Bassi DE. The mesenchymal tumor microenvironment: A drug-resistant niche. *Cell Adh Migr*. 2012;6(3):285-296.
- [464] Correia AL, Bissell MJ. The tumor microenvironment is a dominant force in multidrug resistance. *Drug Resist Updat*. 2012;15(1-2):39-49.
- [465] Houthuijzen JM, Daenen LG, Roodhart JM, Voest EE. The role of mesenchymal stem cells in anti-cancer drug resistance and tumour progression. *Br J Cancer*. 2012;106(12):1901-1906.
- [466] Gilbert LA, Hemann MT. Chemotherapeutic resistance: surviving stressful situations. *Cancer Res*. 2011;71(15):5062-5066.
- [467] Frame FM, Maitland NJ. Cancer stem cells, models of study and implications of therapy resistance mechanisms. *Adv Exp Med Biol*. 2011;720:105-118.
- [468] Anton K, Glod J. Targeting the tumor stroma in cancer therapy. *Curr Pharm Biotechnol*. 2009;10(2):185-191.
- [469] Galmarini CM, Galmarini FC. Multidrug resistance in cancer therapy: role of the microenvironment. *Curr Opin Investig Drugs*. 2003;4(12):1416-1421.
- [470] Malik B, Nie D. Cancer stem cells and resistance to chemo and radio therapy. *Front Biosci (Elite Ed)*. 2012;4:2142-2149.
- [471] McCubrey JA, Steelman LS, Abrams SL, Misaghian N, Chappell WH, *et al*. Targeting the cancer initiating cell: the ultimate target for cancer therapy. *Curr Pharm Des*. 2012;18(13):1784-1795.
- [472] Wilting RH, Dannenberg JH. Epigenetic mechanisms in tumorigenesis, tumor cell heterogeneity and drug resistance. *Drug Resist Updat*. 2012;15(1-2):21-38.
- [473] Balch C, Nephew KP. Epigenetic targeting therapies to overcome chemotherapy resistance. *Adv Exp Med Biol*. 2013;754:285-311.
- [474] Balch C, Fang F, Matei DE, Huang TH, Nephew KP. Minireview: epigenetic changes in ovarian cancer. *Endocrinology*. 2009;150(9):4003-4011.
- [475] Baguley BC. Multiple drug resistance mechanisms in cancer. *Mol Biotechnol*. 2010;46(3):308-316.
- [476] Sharom FJ. The P-glycoprotein multidrug transporter. *Essays Biochem*. 2011;50(1):161-178.
- [477] Fukuda Y, Schuetz JD. ABC transporters and their role in nucleoside and nucleotide drug resistance. *Biochem Pharmacol*. 2012;83(8):1073-1083.
- [478] Wojtkowiak JW, Verduzco D, Schramm KJ, Gillies RJ. Drug resistance and cellular adaptation to tumor acidic pH microenvironment. *Mol Pharm*. 2011;8(6):2032-2038.
- [479] Bailey KM, Wojtkowiak JW, Hashim AI, Gillies RJ. Targeting the metabolic microenvironment of tumors. *Adv Pharmacol*. 2012;65:63-107.
- [480] Solyanik GI. Multifactorial nature of tumor drug resistance. *Exp Oncol*. 2010;32(3):181-185.
- [481] Tannock IF. Tumor physiology and drug resistance. *Cancer Metast Rev*. 2001;20(1-2):123-132.
- [482] Shree T, Olson OC, Elie BT, Kester JC, Garfall AL, *et al*. Macrophages and cathepsin proteases blunt chemotherapeutic response in breast cancer. *Genes Dev*. 2011;25(23):2465-2479.
- [483] Jinushi M, Chiba S, Yoshiyama H, Masutomi K, Kinoshita I, *et al*. Tumor-associated macrophages regulate tumorigenicity and anticancer drug responses of cancer stem/initiating cells. *Proc Natl Acad Sci U S A*. 2011;108(30):12425-12430.



- [484] Martinez-Outschoorn UE, Goldberg A, Lin Z, Ko YH, Flomenberg N, *et al.* Anti-estrogen resistance in breast cancer is induced by the tumor microenvironment and can be overcome by inhibiting mitochondrial function in epithelial cancer cells. *Cancer Biol Ther.* 2011;12(10):924-938.
- [485] Eyler CE, Rich JN. Survival of the fittest: cancer stem cells in therapeutic resistance and angiogenesis. *J Clin Oncol.* 2008;26(17):2839-2845.
- [486] Nakasone ES, Askautrud HA, Kees T, Park JH, Plaks V, *et al.* Imaging tumor-stroma interactions during chemotherapy reveals contributions of the microenvironment to resistance. *Cancer Cell.* 2012;21(4):488-503.
- [487] Bergers G, Hanahan D. Modes of resistance to anti-angiogenic therapy. *Nat Rev Cancer.* 2008;8(8):592-603.
- [488] Eikesdal HP, Kalluri R. Drug resistance associated with antiangiogenesis therapy. *Semin Cancer Biol.* 2009;19(5):310-317.
- [489] Paez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, *et al.* Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell.* 2009;15(3):220-231.
- [490] Bridges EM, Harris AL. The angiogenic process as a therapeutic target in cancer. *Biochem Pharmacol.* 2011;81(10):1183-1191.
- [491] Ebos JM, Kerbel RS. Antiangiogenic therapy: impact on invasion, disease progression, and metastasis. *Nat Rev Clin Oncol.* 2011;8(4):210-221.
- [492] Rapisarda A, Melillo G. Role of the hypoxic tumor microenvironment in the resistance to anti-angiogenic therapies. *Drug Resist Updat.* 2009;12(3):74-80.
- [493] Rapisarda A, Melillo G. Overcoming disappointing results with antiangiogenic therapy by targeting hypoxia. *Nat Rev Clin Oncol.* 2012;9(7):378-390.
- [494] Li XH, Li C, Xiao ZQ. Proteomics for identifying mechanisms and biomarkers of drug resistance in cancer. *J Proteomics.* 2011;74(12):2642-2649.
- [495] Li SL, Ye F, Cai WJ, Hu HD, Hu P, *et al.* Quantitative proteome analysis of multidrug resistance in human ovarian cancer cell line. *J Cell Biochem.* 2010;109(4):625-633.
- [496] Alvarez-Calderon F, Gregory MA, Degregori J. Using functional genomics to overcome therapeutic resistance in hematological malignancies. *Immunol Res.* 2012. [Epub ahead of print]
- [497] Rodrigues AS, Dinis J, Gromicho M, Martins C, Laires A, *et al.* Genomics and cancer drug resistance. *Curr Pharm Biotechnol.* 2012;13(5):651-673.
- [498] Curiel TJ. Immunotherapy: a useful strategy to help combat multidrug resistance. *Drug Resist Updat.* 2012;15(1-2):106-113.
- [499] Vanneman M, Dranoff G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat Rev Cancer.* 2012;12(4):237-251.
- [500] Palakurthi S, Yellepeddi VK, Vangara KK. Recent trends in cancer drug resistance reversal strategies using nanoparticles. *Expert Opin Drug Deliv.* 2012;9(3):287-301.
- [501] Shapira A, Livney YD, Broxterman HJ, Assaraf YG. Nanomedicine for targeted cancer therapy: towards the overcoming of drug resistance. *Drug Resist Updat.* 2011;14(3):150-163.
- [502] Jabr-Milane LS, van Vlerken LE, Yadav S, Amiji MM. Multi-functional nanocarriers to overcome tumor drug resistance. *Cancer Treat Rev.* 2008;34(7):592-602.
- [503] Zamboni WC, Torchilin V, Patri AK, Hrkach J, Stern S, *et al.* Best practices in cancer nanotechnology: perspective from NCI nanotechnology alliance. *Clin Cancer Res.* 2012;18(12):3229-3241.

- [504] Jabir NR, Tabrez S, Ashraf GM, Shakil S, Damanhoury GA, *et al.* Nanotechnology-based approaches in anticancer research. *Int J Nanomedicine*. 2012;7:4391-4408.
- [505] Hu CM, Zhang L. Nanoparticle-based combination therapy toward overcoming drug resistance in cancer. *Biochem Pharmacol*. 2012;83(8):1104-1111.