

Research Article

The Use of *Gigantochloa* Bamboo-Derived Biochar for the Removal of Methylene Blue from Aqueous Solution

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Received 8 January 2022; Revised 25 January 2022; Accepted 2 February 2022; Published 14 February 2022

Academic Editor: Selvaraju Narayanasamy

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In this study, locally grown bamboo (*Gigantochloa* spp.) was used as feedstock for pyrolysis production of biochar under various pyrolysis temperatures (400–800°C). The resultant biochars were tested for their performance in adsorptive removal of the methylene blue (MB) dye. The scope of the adsorption experiment includes the effects of adsorbent dosage, solution pH, initial adsorbate concentration, and contact time. The adsorption data confirmed that pyrolysis temperature has a significant effect on adsorptive performance, whereas biochar pyrolysed at 500°C (BC500) has the highest adsorptive performance with the maximum adsorption capacity (derived from the Langmuir model) being 86.6 mg g⁻¹. Basic characterisations (SEM, EDX, XRD, FTIR, and BET) were carried out for BC500 where FTIR and SEM confirmed the adsorption of MB onto the biochar, while the BET data showed the reduction of the BET surface area, total pore volume, and pore diameter after the adsorption process.

1. Introduction

The textile industry is an ever-growing market associated with the production of fabric. Despite its significance, this industrial sector contributes to the largest global polluters, particularly in releasing untreated effluents into the water bodies [1, 2]. These effluents consist of various organic compounds and toxic substances. When not adequately treated, they can not only degrade the environment but also degrade the health of humans and animals [2–5]. Dyes, which are widely used to impart colour onto products, are among the many effluents that are discharged from textile factories, which account for more than 50% of the world's dyestuff market [6]. Methylene blue (MB) is an example of a commonly used basic dye for dying wool, cotton, and silk [7]. Despite its many practical uses, MB is a known irritant that can cause eye burn and, if consumed, can cause nausea, headache, increased heart rate, and the inflammation of the gastrointestinal tracts [7–9].

There are many dye wastewater treatment methods such as chemical precipitation, adsorption, ion exchange, membrane technologies, electrodialysis, phytoremediation, and photocatalysis [10–15]. Adsorption is one of the popular wastewater remediation techniques due to its efficiency, simplicity, and low cost [16–20] and does not result in sludge production [21]. In addition, one of its major benefits is its eco-friendly features as its adsorbents can potentially be recovered and recycled [13, 22–24].

Activated carbon has undoubtedly been a conventional adsorbent in wastewater treatment universally [25, 26]. However, despite its prolific use, some drawbacks are its high cost and the dependency of complexing agents, which lead researchers to find a more suitable alternative [27]. Some low-cost and readily available adsorbents include natural materials such as biochar, zeolites, clay, or certain waste products from industrial operations such as fly ash and reused sanding wastes [14, 28, 29]. Biochar has been a popular choice among researchers due to its specific properties, including large specific surface area, porous structure, enriched surface functional groups, and mineral components, making it suitable as an adsorbent for wastewater remediation [30].

Bamboo is one of the many applicable biomasses with many benefits as feedstock for making biochar. They are found abundantly in natural habitats, especially in Asia and the tropical regions, due to their fast proliferative growth. Most bamboo species take only a few months to grow to full size and are among the fastest-growing plants [31]. Bamboo is also considered a renewable biomass resource. Bamboo is rich in lignocellulosic content, which plays a significant role in becoming an excellent raw material for biochar production [27, 32–34].

Regardless of the increasing published literature, more input is required to ensure an effluent clean-up as numerous factors affect this adsorption ability. From literature, it is known that altering pyrolysis conditions, particularly the pyrolysis temperature, can significantly impact the properties of the biochar. The biochar properties such as its elemental composition, physical structure, surface functional groups, surface area, pore size, and stability can influence the level of the adsorption capacity [35, 36]. In general, high pyrolysis temperature can increase the surface area and porosity of the adsorbent while keeping the minerals such as potassium, phosphorus, and calcium in the biochar. These minerals are known to enhance the adsorption capacity [37-39]. However, at high pyrolysis temperature, the functional groups such as carboxyl, hydroxyl, and amino groups on the surface of adsorbents may decrease, resulting in the reduced capacity of adsorption [37, 40]. According to Sun et al., the pyrolysis temperature and feedstock are responsible for the production rate, thermal stability, carbon content, and element composition of biochar [39]. Sizirici et al. showed that higher pyrolysis temperature resulted in biochar with lower adsorptive performance than those obtained at a lower temperature [37]. Another study shows the impact of pyrolysis temperature on lead adsorption and revealed that 550°C is the optimal temperature of making biochar for lead adsorption [41]. Primary cracking, secondary decomposition, and the formation of oxygen functional groups start around 400–500°C during pyrolysis. Hence, biochar samples produced within the temperature range of 500-700°C show better reactivity and adsorption behaviour than biochar samples produced below or above that temperature [42]. Those studies evidence that the adsorption mechanism and performance are highly affected by complex properties of biochar which are mainly based on the type of biomass and pyrolysis temperature [37-41].

In this paper, biochar prepared from locally grown *Gigantochloa* bamboo was employed to remove MB dye. The specific objectives were to optimise the pyrolysis temperature for the production of biochar to achieve the high

TABLE 1: Yield of biochar at different pyrolysis temperatures.

Sample	Biochar yield (%)
BC400	4.85
BC500	11.39
BC600	8.12
BC700	3.87
BC800	3.44

adsorption capacity. With the obtained temperature, adsorption conditions were optimised for efficient removal of dye, including adsorbent dosage, initial concentration, pH, and contact time. Further, the prepared biochar was characterised using Brunauer–Emmett–Teller (BET), Fourier transform infrared (FTIR), scanning electron microscope (SEM), and energy-dispersive X-ray spectroscopy (EDX). Lastly, the adsorption behaviour was modelled using various isotherm and kinetic models.

2. Methodology

2.1. Preparation of Adsorbent. The Gigantochloa bamboo stems were obtained locally from Kampong Kianggeh, Brunei-Muara District, Brunei Darussalam. The surface of the bamboo was washed and sun-dried, and the outer layer was peeled. They were then further dried in a drying oven at a temperature of $\sim 80^{\circ}$ C for a few days until there was no further noticeable decrease in weight.

The dried bamboo from the oven was directly transferred to the muffle furnace, where they undergo pyrolysis. The bamboo was divided into five groups, and each group was pyrolysed at different temperatures (400, 500, 600, 700, and 800°C). The heating rate was set at 10°C min⁻¹ and with 4 h of holding time upon reaching the desired temperatures. The flow rate of N₂ was at a constant 20 mL min⁻¹ throughout the pyrolysis process. After the furnace temperature reached room temperature, the biochar was taken out from the furnace. It was ground using pestle and mortar and passed through 150 μ m mesh sieves and stored in its respective airtight container. BC400, BC500, BC600, BC700, and BC800 were used to represent the biochar derived at different temperatures at 400, 500, 600, 700, and 800°C, respectively.

The yield of biochar was calculated according to

Biochar yield (%) =
$$\frac{\text{Mass of biochar}}{\text{Mass of bamboo}} \times 100.$$
 (1)

2.2. Chemical Reagents. The MB powder was purchased from Merck, Germany (dye purity 95%, reagent grade). The MB stock solution was prepared by dissolving the MB powder in distilled water, and all other concentrations were obtained via serial dilution. All other chemicals used in this study are of reagent grade and used as received without purification.

2.3. Batch Adsorption Experiments

2.3.1. Screening Test. After pyrolysis, 0.04 g from each biochar sample was mixed with 20 mL of MB dye. The solution was shaken for 3 h at a speed of 200 rpm with an orbital

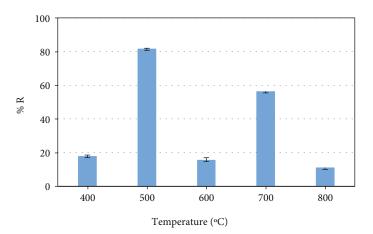


FIGURE 1: The performance of biochar produced by various pyrolysis temperatures on removing MB dye from solution (room temperature, unadjusted pH, $C_i = 100$ ppm, and contact time of 3 h).

shaker (MIULab GS-20). The purpose of this method was to find the most efficient adsorption process according to their pyrolysis temperature. With the optimum sample, it was then made to be the constant variable for the proceeding experiment.

2.3.2. Adsorbent Dosage, Initial Concentration of Solution, pH, and Contact Time. The general procedure for all adsorbent dosage, initial dye concentration, pH, and contact time studies involves shaking the known weight of adsorbent in 20 mL of known concentration of MB with an orbital shaker for 3 h and at agitating speed of 200 rpm at room temperature. The mixture was filtered, and the filtrate was measured using a UV-vis-NIR spectrometer (Agilent technologies, Cary 5000, US) at an absorbance (λ_{max}) of 664 nm. q_e and percentage removal were calculated using equations (2) and (3), respectively. The effects of adsorbent dosage (0.01-0.06 g), pH (2-10), initial dye concentration (20-400 mg L⁻¹), and contact time (5-180 min) were investigated. To increase accuracy of the data, each experiment was done twice, and the average was taken.

$$q_e = \frac{C_i - C_e}{m} \times V, \tag{2}$$

$$\%R = \frac{C_i - C_e}{C_i} \times 100,\tag{3}$$

where C_i (mg L⁻¹) is the initial concentration of the solution, m (g) is the mass of adsorbent used, and V (L) is the volume of dye used.

2.3.3. Optimisation of Adsorption Conditions. Optimising the adsorption parameters is equally crucial as the pyrolysis temperature for biochar. These are several parameters (effects of adsorbent dosage, initial adsorbate concentration, solution pH, and contact time) that are essential in a typical adsorption study [43]. In general, increasing adsorbent dosage allows higher availability of adsorption sites for interaction with the adsorbates. However, when dosage exceeds a certain level, this leads to a plateau in adsorptive performance. This is due to the collision of adsorbent particles

TABLE 2: Effect of temperature on the specific surface area of biochar.

Sample	$S_{BET} \ (m^2 g^{-1})$	$V_{TOTAL} \ (\text{cm}^3 \text{g}^{-1})$	D_p (nm)
BC400	28.4270	0.032109	5.00934
BC500	4.1519	0.015165	14.61015
BC600	10.6617	0.020640	7.74367
BC700	8.3130	0.017765	8.54812
BC800	8.9699	0.019398	8.65043

and thus overlapping of the adsorption sites [44]. The initial adsorbate concentration typically results in increased adsorptive performance at higher adsorbate concentration due to the higher driving force caused by the concentration gradient according to Fick's law of diffusion. Similar to adsorbent dosage, the adsorptive performance becomes plateau at a particular level of adsorbate concentration due to the exhaustion of adsorption sites. The solution pH controls the electrostatic interaction between the functional groups on the adsorbent's surface and the adsorbate [45]. Lastly, the contact time is the duration the adsorbent is in contact with the adsorbate and is essential to confirm the adsorption process has reached equilibrium [46, 47].

2.3.4. Proper Disposal of Generated Wastes. All generated wastes were collected and are handled by a waste disposal company, specialised in the proper disposal of chemical wastes.

2.4. Biochar Characterisations. In this study, SEM, EDX, XRD, FTIR, and BET were used for characterising the biochar. The surface morphology and surface element analysis of BC500 before and after adsorption was characterised using a FE-SEM (JEOL JSM-7600F) equipped with an EDX spectrometer. An accelerating voltage of 10 kV with a working distance of 9–16 mm was maintained during the analysis. These analyses provide important visual insights as well as the information about the elemental composition of the adsorbent's surface. A Shimadzu X-ray diffractometer (XRD-7000) with Cu K α radiation source ($\lambda = 1.54060$ Å) at 2θ range between 20 and 80° and a sampling rate of 5° min⁻¹

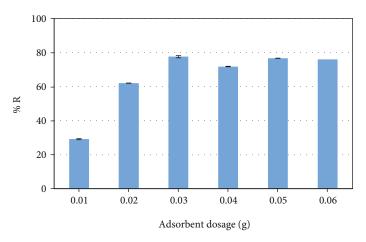


FIGURE 2: The effect of adsorbent dosage on removing MB dye from solution using BC500 (room temperature, unadjusted pH, $C_i = 100$ ppm, and contact time = 3 h).

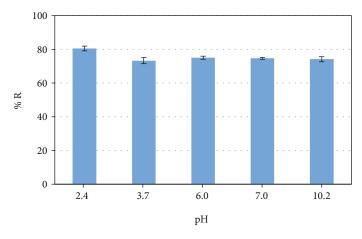


FIGURE 3: The effect of pH on removing MB dye from solution using BC500 (room temperature, $C_i = 100$ ppm, and contact time = 3 h).

was used to determine the XRD pattern of biochar before and after adsorption. The XRD data provides important information about the crystalline structure. A FTIR spectrometer was used to determine the functional groups of biochar involved before and after reacting with MB dye, and FTIR was carried out using an Agilent Cary 630 FTIR spectrometer with 400–4000 cm⁻¹ wavelength range at a resolution of 8 cm⁻¹, recording 32 scans per sample. The data derived from FTIR spectra is important for determining the functional group involved in the adsorption process. The biochar's BET surface area, total pore volume, and pore diameter were determined using the Micrometrics ASAP 2020 surface area and porosity analyser (USA), under nitrogen-adsorption desorption isotherm at 77 K.

2.5. Adsorption Kinetic Modelling. The adsorption kinetics of adsorbate onto biochar was studied using the pseudo-first-order, pseudo-second-order, and Weber-Morris kinetic models.

This pseudo-first-order model describes the relationship between the sorption sites of the adsorbents and adsorbates [48]. The equation in its linear form is given by

$$\log (q_e - q_t) = \log q_e - \frac{t}{2.303} k_1, \tag{4}$$

where q_e and q_t are the amounts of adsorbate adsorbed at equilibrium and at time t (min), respectively, and k_1 is the rate constant of adsorption (min⁻¹). The linear plot of ln $(q_e - q_t)$ against time was used to determine the rate constant k_1 (mg g⁻¹).

The pseudo-second-order model focuses more on the chemisorption process and the dependency of the adsorption capacity on time [49]. This can be determined based on equation (5) expressed as

$$\frac{t}{q_t} = \frac{1}{q_e^2 k_2} + \frac{t}{q_e}.$$
 (5)

The linear plot of t/q_t against t was plotted to determine the pseudo-second-order rate constant, k_2 (g mg⁻¹ min⁻¹).

Weber-Morris is a suitable model when the internal surface or pore diffusion of pollutant inside the sorbent is the

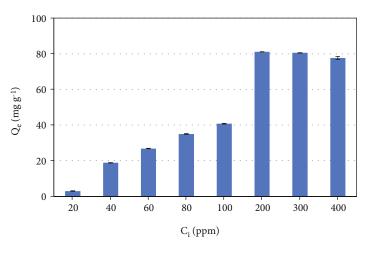


FIGURE 4: The effect of initial dye concentration on removing MB from solution using BC500 (room temperature, unadjusted pH, and contact time = 3 h).

rate-limiting step. Equation (6) shows the Weber-Morris model.

$$q_t = k_3 t^{1/2} + C. (6)$$

 $k_3 \pmod{\text{g}^{-1} \min^{-1/2}}$ is the intraparticle diffusion rate constant which can be determined by plotting linear graph of q_t against $t^{1/2}$.

2.6. Adsorption Isotherm. To illustrate the mechanism of distribution of the adsorption molecules between the liquid and solid phases when the adsorption process reaches the equilibrium state, several adsorption isotherm (Langmuir, Freundlich, Temkin, and Dubinin-Radushkevich (D-R)) models were used.

The Langmuir model describes the monolayer sorption on apparently localised adsorption sites [50]. The linearised form of the Langmuir isotherm is expressed as

$$\frac{C_e}{q_e} = \frac{1}{K_{\rm L}q_m} + \frac{C_e}{q_m},\tag{7}$$

where C_e (mgL⁻¹) is the concentration at equilibrium, q_e is the amount adsorbed, and q_m is the maximum amount of adsorbent to form a complete monolayer on the surface. K_L (mg⁻¹) is the Langmuir constant which indicates the extent of interaction between the adsorbate and surface.

The Freundlich model describes the nonideal and reversible adsorption processes that occur on heterogeneous surfaces and active sites of different energies based on multilayer adsorption and equilibrium [51]. The linear form of the Freundlich isotherm model is represented as follows:

$$\log q_e = \log K_{\rm F} + \frac{1}{n} \log C_e, \tag{8}$$

where K_F is the Freundlich isotherm constant and n is the adsorption intensity.

The D-R isotherm model assumes that the mechanism is related to the porosity of the adsorbent rather than surface

TABLE 3: Adsorption isotherm parameters for the adsorption of MB onto BC500.

Model	Parameter	Value
Langmuir	$q_m (\mathrm{mg g}^{-1})$	86.63
	$K_L \ (\mathrm{mg}^{-1})$	0.05
	R_L	0.048
	R^2	0.9795
Freundlich	$K_F (\text{mg g}^{-1} (\text{Lmg}^{-1})^{1/n})$	3.35
	n	3.44
	R^2	0.6117
Temkin	A_T	0.61
	b_T	0.53
	R^2	0.6066
D-R	$q_m \ (\mathrm{mg g}^{-1})$	-76.68
	$K_{dr} (\mathrm{mol}^2 \mathrm{kJ}^{-2})$	0.0130
	ε (kJ mol ⁻¹)	6.19
	$q_s \pmod{\mathrm{g}^{-1}}$	1.07
	R^2	0.7841

coverage with a monolayer or multilayer. One of its distinguishing features is that it is temperature-dependent and is mainly used to predict whether the adsorption process is physical or chemical [52]. The linearised equation is as follows:

$$\ln (q_e) = \ln (q_s) - K_{dr} \varepsilon^2, \tag{9}$$

where $q_e \pmod{\text{gg}^{-1}}$ represents the amount of adsorbate adsorbed at equilibrium. $K_{dr} \pmod{\text{kJ}^{-2}}$ is the mean free energy of adsorption, and ε refers to the Polanyi potential.

The Temkin model considers the heat of adsorption as a function of temperature. All molecules in the layer would decrease in a linear manner instead of logarithmically with

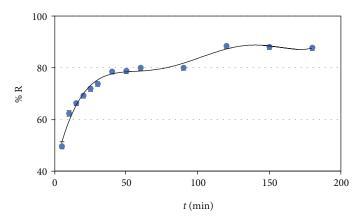


FIGURE 5: The effect of contact time on the adsorption of MB using BC500 (room temperature, unadjusted pH, and $C_i = 100$ ppm).

coverage at average concentrations [53]. The model is expressed as

$$q_e = \frac{RT}{b_{\rm T}} \ln A_{\rm T} + \left(\frac{RT}{b_{\rm T}}\right) \ln C_e,\tag{10}$$

where $A_{\rm T}$ is the Temkin isotherm equilibrium binding constant (Lg⁻¹), $b_{\rm T}$ is the Temkin isotherm constant, R is the universal gas constant (8.314 J mol⁻¹ K⁻¹), and T is the temperature at 298 K.

3. Results and Discussions

3.1. The Yield of Biochar. As seen from Table 1, the yield of biochar at pyrolysis temperature of 400°C is at 4.85%. As the pyrolysis temperature increases to 500°C, the biochar yield increases to 11.4%. Any further increase in temperature beyond 500°C resulted in a decrease in the biochar yield. This nonlinear behaviour is also reported by Açıkalın et al. in which biochar was produced using pistachio shell at pyrolysis temperature between 350 and 650°C, and the highest biochar yield was obtained at 500°C [54].

3.2. Effect of Pyrolysis Temperature in Dye Adsorption. The effect of pyrolysis temperature on the adsorption of MB using biochar can be seen in Figure 1. The pyrolysis temperature of 500°C resulted in the highest removal of MB dye from aqueous solution at 81.8% ($q_e = 28.1 \text{ mg g}^{-1}$), while the temperature of 800°C resulted in the lowest removal 11.1% ($q_e = 5.7 \text{ mg g}^{-1}$). This behaviour may be related to the average pore diameter (D_p) of BC. Naturally, a larger D_p will allow more dye molecules to move inside the pores with less chance of getting clogged when compared to a smaller D_p . As seen in Table 2, BC500 has the largest D_p at 14.6 nm, while BC400, BC600, BC700, and BC800 were much smaller at 5.00, 7.74, 8.54, and 8.65 nm, respectively. Since BC500 showed the utmost performance, therefore, complete adsorption experiment was only carried out for BC500.

3.3. *BET*. The pyrolysis of biomass resulted in the decomposition of lignin and the quick release of H_2 and CH_4 . These gases magnified the surface area and pore volume, particu-

 TABLE 4: Kinetic model parameters for the adsorption of MB onto BC500.

Model	Parameter	Value
Pseudo-first order	$k_1 \; (\min^{-1})$	0.0290
	R^2	0.9621
Pseudo-second order	$k_2 \ (\mathrm{mg g^{-1} min^{-1}})$	0.0037
	R^2	0.9986
Weber-Morris	$k_3 (\mathrm{mgg}^{-1}\mathrm{min}^{-0.5})$	1.4931
	R^2	0.9206

larly for the micropores. This is mostly attributed to the further degradation of lignin and the reaction of aromatic condensation, which in turn increases the release of volatile matter, creating more pores [42, 55, 56]. These pores can be characterised using the Brunauer–Emmett–Teller (BET) surface area analysis to obtain useful insights such as the adsorbent's surface area ($S_{\rm BET}$), total pore volume ($V_{\rm TOTAL}$), and $D_{\rm p}$.

The results of the BET surface area analysis are summarised in Table 2. BC400 has the highest S_{BET} and V_{TOTAL} at 28.4 m² g⁻¹ and 0.032 cm³ g⁻¹, respectively, while BC500 has the lowest S_{BET} at 4.15 m² g⁻¹ and 0.015 cm³ g⁻¹. However, the BC500 has the highest D_{p} at 14.6 nm, while the lowest was for BC400 at 5.0 nm.

The S_{BET} , V_{TOTAL} , and D_{p} are observed not to be in a linear relationship with the increasing temperature. This is partially due to varying proportions of micropore ($D_{\text{p}} < 2 \text{ nm}$) and mesopore ($D_{\text{p}} 2-50 \text{ nm}$) that form at a different range of temperature, which is observed by Fu et al. in their study of rice straw biochar [57].

3.4. Effect of Adsorbent Dosage. The effect of the adsorbent dosage on the MB removal was investigated using various masses of BC500 (0.01 g to 0.06 g) at a fixed volume (20 mL) of the adsorbate. It can be observed in Figure 2 that the removal of dye increased as the mass of adsorbents increased from 0.01 g to 0.03 g. This is due to more adsorption sites being available at higher adsorbent dosages. However, beyond the adsorbent dosage of 0.03 g, there is no

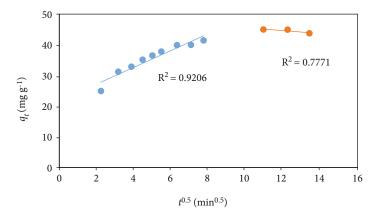


FIGURE 6: Weber-Morris plot.

further increase in the removal of MB, which may be due to the overcrowding of adsorbent particles in such a way that overlapped the adsorption sites [58]. As the adsorbent dosage that performed the best was 0.03 g with a percentage removal of 77.6% ($q_e = 51.0 \text{ mg g}^{-1}$), therefore, the rest of the adsorption experiment was carried out at this adsorbent dosage.

3.5. Effect of pH. As seen from Figure 3, the change in pH has no large impact on the pH. The removal of MB $(C_i = 100 \text{ ppm}, \text{ unadjusted pH at 7.0})$ was at 74.3% $(q_e = 39.2 \text{ mg g}^{-1})$. As pH increased to 10, the removal of MB was at 74.1% $(q_e = 40.3 \text{ mg g}^{-1})$, while for acidic pH (2.4), the removal was at 80.6% $(q_e = 41.0 \text{ mg g}^{-1})$. This behaviour is commonly observed with the adsorption of MB dye, which was also reported in removing MB using biomass adsorbent derived from Azolla pinnata [59].

3.6. Effect of Initial Concentration and Isotherm Modelling. The effect of the initial MB concentration is shown in Figure 4. There was a gradual increase in q_e as the C_i increased from 20 ppm to 200 ppm. The reason for this observation is the availability of adsorption sites on the adsorbent's surface. Beyond 200 ppm, no further increase in q_e is due to the saturation of the adsorption sites and no further possible uptake of MB molecules at this adsorption condition.

To understand more about the adsorption system, the adsorption data were fitted into various adsorption isotherm models. The parameters of the isotherm models are summarised in Table 3, where it can be observed that the correlation coefficient of determination (R^2) is highest for the Langmuir at 0.9795, followed by Dubinin-Radushkevich ($R^2 = 0.7841$), Freundlich ($R^2 = 0.6117$), and Temkin ($R^2 = 0.6066$) models. The highest value of R^2 makes the Langmuir model most suitable to describe the adsorption process and indicate the possibility of forming a single layer of MB on the surface of BC500. The R_L value assumes the nature and the feasibility of the adsorption process, and since the value was determined to be 0.048, the adsorption process is considered favourable.

The q_m , calculated from the Langmuir isotherm model, is one of the most commonly used metrics for comparing the performance of various adsorbents in the removal of a particular adsorbate. The q_m of the BC500 was calculated to be at 86.3 mg g⁻¹ and is compared to adsorbents reported in the literature such as *Azolla pinnata* (80.6 mg g⁻¹) [59], *Lathyrus sativus* husk (98.3 mg g⁻¹) [60], *Casuarina* leaf (110.8 mg g⁻¹) [9], eucalyptus leaves (194.3 mg g⁻¹) [61], jackfruit leaf (271 mg g⁻¹) [62], and zinc chloride activated carbon (462.9 mg g⁻¹) [63].

3.7. Effect of Contact Time and Kinetic Modelling. The effect of contact time on removing MB ($C_i = 100$ ppm, unadjusted pH, and room temperature) is shown in Figure 5, where there was a rapid increase in adsorption of MB from time 0 to 40 min. The rapid rate of adsorption was mainly due to the availability of sites for dye adsorption [44]. Beyond the contact time of 40 min, the adsorption rate slows down as adsorption sites are gradually taken up until there is no further increase in adsorption beyond 120 min. Thus, the contact time of 120 min is deemed sufficient for this adsorption system to achieve equilibrium.

The adsorption data were fitted with various kinetic models to study the kinetics of the adsorption processes, and the kinetic parameters are summarised in Table 4. By comparing the R^2 values between the models, the pseudo-second order best described the experimental kinetic data ($R^2 = 0.9986$), implying that the chemisorption may dominate the adsorption process.

To gain insight into the diffusion process, kinetic data were fitted into the Weber-Morris model and the plot is shown in Figure 6. The Weber-Morris plot did not cross the origin (*Y*-intercept \neq 0), which indicates that the intraparticle diffusion is not the rate-limiting step, and the "bottleneck" of the adsorption may be due to other processes.

3.8. Characterisations of the Adsorbents

3.8.1. SEM and EDX. The SEM images are shown in Figure 7. The surface morphology of the BC500 is rough and distorted in appearance with some irregular microstructures. However, after adsorption with MB, the surface is smoother with less visible microstructures, and this may indicate the deposition of MB onto the adsorbent's surface.

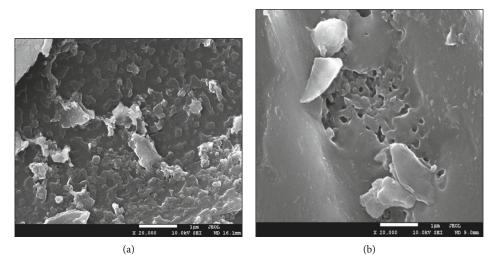


FIGURE 7: SEM images of BC500 (a) before and (b) after MB adsorption.

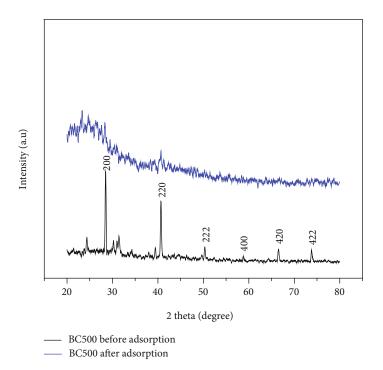


FIGURE 8: The XRD spectra of BC500 before and after MB adsorption.

The elements present on BC500 were identified using EDX. The EDX revealed the presence of C, O, S, Cl, and K with the normalised weight percentage at 82.9, 11.3, 0.33, 0.29, and 4.65%, respectively. However, after the MB adsorption, only the elements C, O, S, and K were detected at 88.0, 9.42, 0.23, and 2.36%, respectively. Also, notice that K and Cl percentages have reduced after adsorption; this could be due to the mobility and solubility of the K and Cl ions easily washed into the aqueous phase during the adsorption process. A similar finding is also reported in the adsorption of methyl violet 2B dye using biomass *Azolla pinnata* as the adsorbent [64].

3.8.2. XRD. XRD is a valuable analytical technique for the identification of crystallinity in a material. The XRD spectra of the BC500 before and after the adsorption of MB are shown in Figure 8. For BC500 before the adsorption, diffraction peaks at 28.4, 40.6, 50.2, 58.7, 66.4, and 73.7° are assigned (200), (220), (222), (400), (420), and (422) crystal planes of sylvite (KCl), respectively, and are consistent with findings by Yuan et al. and He et al. [65, 66]. Those intense sharp peaks suggest that KCl content is high in BC500 before the adsorption. There was no intense peak observed in the XRD spectra for BC500 after the MB adsorption; however, there is a broad diffraction peak at 2θ value

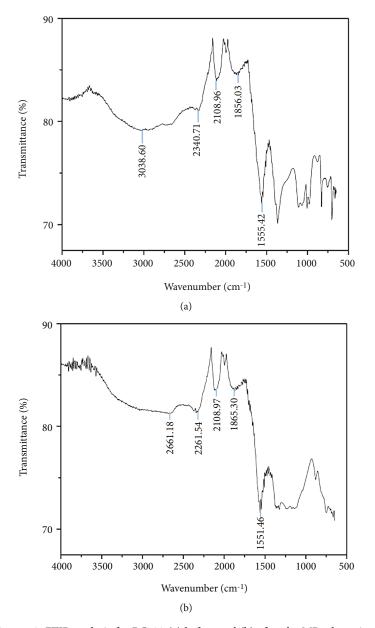


FIGURE 9: FTIR analysis for BC500 (a) before and (b) after the MB adsorption.

22°, which is due to the amorphous carbon of the adsorbent after adsorption. This might be that KCl is dissolved and washed away into the solution during the adsorption process, as discussed in the previous section.

3.8.3. FTIR. The purpose of FTIR analysis is to identify the functional groups that interact with the MB molecules. The molecular structure was similar to the biochar produced; hence, for brevity, only the FTIR plot for BC500 before and after treatment is shown in Figure 9.

The FTIR spectrum of BC500 without MB treatment showed a broad peak at 3038 cm^{-1} , which is assigned to the O-H stretching, indicating that some moisture was absorbed from the air. The stretching at 2340 cm^{-1} is attributed to the O-H bond stretch of a carboxylic acid group [67]. The peak at 1856 cm^{-1} was related to the stretching vibration

of C=O of the carbonyl group. In contrast, the 1555 cm⁻¹ was associated with the stretching vibration of C=C and may be due to the benzene ring skeleton in lignin mainly from aromatic [54]. For the MB-treated BC500, there is a shift of peaks such as 2261 cm⁻¹ (O-H stretch of the carboxylic group), 1865 cm⁻¹ (C=O stretch of the carbonyl group), and 1551 cm⁻¹ (aromatic), indicating that these functional groups may have interacted with the MB dyes.

The BET analysis was carried out for BC500 before and after the MB adsorption. The results revealed that after MB adsorption, the BET surface area decreased from 4.1519 to $3.2413 \text{ m}^2 \text{ g}^{-1}$, while the total pore volume was from 0.015165 to 0.008817 cm³ g⁻¹, and lastly, the pore diameter was from 14.61015 to 9.7603 nm, respectively. The molecular dimension of MB is approximately $0.7 \times 1.7 \text{ nm}$ [68], which indicates that the pore diameter and pore volume of

BC500 are relatively higher than those of the MB molecules. Therefore, the MB molecules easily enter the pores of BC500, filling them and leading to the decreasing surface area, pore volume, and pore diameter of BC500 after the adsorption process.

3.9. Adsorption Mechanism. The mechanism of dye adsorption is well established, and the three dominant forces involved in dye interaction are electrostatic interaction, hydrogen bonding, and hydrophobic-to-hydrophobic interaction [69, 70]. Several active functional groups such as O-H stretch of the carboxylic group and C=O stretch of the carbonyl group (as described in the previous section) are capable of interacting with MB by hydrogen bonding or electrostatic attraction, while the aromatic groups on the adsorbent's surface may interact by hydrophobic-tohydrophobic interaction. Similar findings were reported in removing MB using *Casuarina* leaf [9] and removing malachite green dye using *Azolla pinnata* [71].

4. Conclusion

In summary, this study investigated the effect of pyrolysis temperature on a species of Gigantochloa bamboo to derive biochar for the removal of MB from the aqueous solution. It was revealed that pyrolysis temperature of 500°C resulted in the highest biochar yield at 11.39% and also possessed the highest adsorptive performance when compared to biochar produced at other temperatures. The SEM image revealed the rough surface of biochar which becomes smoother after MB adsorption, while EDX data reveal the composition of C, O, S, Cl, and K elements in BC500. The XRD data reconfirms the presence of KCl by having crystal peaks at 28.4, 40.6, 50.2, 58.7, 66.4, and 73.7°, while FTIR confirms the BC500 interaction with MB by having carboxylic, carbonyl, and aromatic functional groups. S_{BET} data revealed a decrease in the BET surface area and pore volume after adsorption, further confirming the adsorption of MB. The optimised adsorbent dosage for removal of MB is at 0.03 g and aqueous pH of 2, and the equilibrium was achieved within 120 min of contact time. The kinetic data is best represented by the pseudo-secondorder model, indicating that the chemisorption dominates during the adsorption process. On the other hand, the Langmuir model best describes the adsorption process hinting at the formation of a monolayer of the adsorbate on the adsorbent's surface. Based on the results, it can be concluded that the pyrolysis temperature significantly impacts the physicochemical characteristics of the biochar and thus influences the adsorptive performance of MB.

Data Availability

All the data used to support the findings of this study are included within the article.

Additional Points

Research Highlight. Biochar was produced using the Gigantochloa bamboo as feedstock for pyrolysis. Several pyrolysis temperatures (400 to 800°C) were investigated. The adsorption conditions for removal of methylene blue (MB) were optimised. Pyrolysis temperature of 500°C produced biochar with the highest adsorptive performance in removal of MB from aqueous solution.

Conflicts of Interest

All authors declare no conflict of interest.

Acknowledgments

We, the authors, would like to thank the Government of Brunei Darussalam and Universiti Brunei Darussalam for their continuous support. The work described in this paper was supported by Universiti Brunei Darussalam research grants UBD/RSCH/1.9/FICBF/2021/010 and UBD/RSCH/ 1.9/FICBF/2021/008 and Ministry of Science and Technology of Taiwan (MOST 109-2112-M-019-001).

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