Thermodynamics of citrate complexation with Mn²⁺, Co²⁺, Ni²⁺ and Zn²⁺ ions

D. Wyrzykowski · L. Chmurzyński

Received: 3 August 2009/Accepted: 25 September 2009/Published online: 3 November 2009 © The Author(s) 2009. This article is published with open access at Springerlink.com

Abstract Isothermal titration calorimetry has been used to determine the stoichiometry, formation constants and thermodynamic parameters (ΔG° , ΔH , ΔS) for the formation of the citrate complexes with the Mn²⁺, Co²⁺, Ni²⁺ and Zn²⁺ ions. The measurements were run in Cacodylate, Pipes and Mes buffer solutions with a pH of 6, at 298.15 K. A constant ionic strength of 100 mM was maintained with NaClO₄. The influence of a metal ion on its interaction energy with the citrate ions and the stability of the resulting complexes have been discussed.

Keywords Citrate complexes · Thermodynamic parameters · Isothermal titration calorimetry

Introduction

The ions of the citric acid (2-hydroxy-1,2,3-propanetricarboxylic acid; H_4Cit) occur in small amounts in the majority of living organisms to act as bioactive ligands implicated in a number of biochemical processes [1–5]. Owing to their capacity to form thermodynamically stable complexes with a variety of metal ions, they found widespread use in food and pharmaceutical industries as well as in medicine. Over the past few years much attention has been paid to the synthesis of nanomaterials using metal citrates as precursors [6–9].

Thermodynamic stability of complexes is crucial for processes occurring in living organisms, as it determines, amongst others, biological and pharmacological activities

D. Wyrzykowski (🖂) · L. Chmurzyński

Faculty of Chemistry, University of Gdańsk, Sobieskiego 18, 80-952 Gdańsk, Poland e-mail: daro@chem.univ.gda.pl of complex compounds and plays an important role in the safety of their application. It is important to realize that the knowledge of thermodynamic parameters of reactions enables a better understanding of the processes involving complex compounds than that of simple equilibrium constants [10, 11]. Thermodynamic characteristics of a reaction also enable determination of the relationship between the structure of a ligand and chemical properties of new compounds, thus contributing to optimization of the conditions for their synthesis [12]. For this reason it seemed worthwhile to determine thermodynamic characteristics for the reactions of the citrate ions with some transition metal ions.

Experimental

Materials

All reagents: $C_6H_5O_7Na_3\cdot 2H_2O$ (sodium citrate dihydrate), Mn(NO₃)₂·6H₂O, Co(NO₃)₂·6H₂O, Ni(NO₃)₂·6H₂O, Zn(NO₃)₂·6H₂O, NaClO₄, Cacodylate (Cacodylic acid sodium salt trihydrate), Pipes (1,4-Piperazinediethanesulfonic acid) and Mes 2-(*N*-Morpholino)ethanesulfonic acid) were purchased from Aldrich Chemical Corp. These compounds were used without further purification.

Isothermal titration calorimetry (ITC)

All the ITC experiments were run at 298.15 K using an AutoITC isothermal titration calorimeter (MicroCal Inc., Northampton, USA) with a 1.4491-mL sample and a reference cell. The reference cell was filled with distilled water. The data, specifically the heat normalized per mole of injectant, were processed with Origin 7 from MicroCal.

An initial 2-uL injection sample was discarded from each data set to remove the effect of titrant diffusion across the syringe tip during the equilibration process. The experiment consisted of injection (29 injections, 2 µL for the first injection only) of a ca. 10-15-mM solution of appropriate salt into the reaction cell initially containing buffered solution of a ca. 1 mM sodium citrate (ionic strength $I = 100 \text{ mM NaClO}_4$). A background titration was performed using identical titrant with the buffer solution placed in the sample cell. The result was subtracted from each experimental titration to account for the heat of dilution. All the solutions were degassed before titrations were performed. Titrant was injected at 5-min intervals to ensure that the titration peak returned to the baseline prior to the next injection. Each injection lasted 20 s. To achieve a homogeneous mixing in the cell, the stirrer speed was kept constant at 300 rpm. Calibration of the AutoITC calorimeter was carried out using electrically generated heat pulses. The CaCl₂-EDTA titration was performed to check the apparatus and the results $(n, K, \Delta H)$ were compared with those obtained for the same samples (test kit) at MicroCal.

Results and discussion

Thermodynamic parameters of interaction of the citrate ion with the Mn^{2+} , Co^{2+} , Ni^{2+} and Zn^{2+} ions, determined by the ITC technique in the Cacodylate, Pipes, and Mes buffer solutions witha pH of 6, at 298.15 K, are summarized in Table 1. The equilibrium constants, binding enthalpies and reaction stoichiometries were obtained from ITC experiments by fitting binding isotherms, using nonlinear leastsquares procedures, to a model that assumes a single set of identical binding sites. From the above experimental parameters, the free energy of binding (ΔG°) and entropy change (ΔS) could be determined from the standard thermodynamic relationship, $\Delta G^{\circ} = -RT \ln K_{obs} = \Delta_{obs}H - T\Delta S$.

The stoichiometry of the compounds indicates that at a pH of 6 almost equimolar metal/ligand complexes are formed. X-ray crystallographic results have shown that the citrate ions act as tridentate ligands [13–15]. Oxygen atoms of two carboxylic groups and an oxygen atom of the hydroxyl group participate in the metal binding (Fig. 1). A carboxyl group at the central carbon atom, C(3), is almost perpendicular to the carbon backbone, C(1)–C(2)–C(3)–C(4)–C(5), and is situated on one plane with the hydroxyl group. A third donor is the oxygen atom of the terminal carboxyl group, C(1) [16, 17]. A similar type of metal binding can also be expected in solutions.

In solution with a pH of 6, the citrate ions occur mostly as the H_2Cit^{2-} and $HCit^{3-}$ species, the equilibrium being displaced largely towards the former ones [18] $([H_2Cit^{2-}]:[HCit^{-}]$ equals approximately 4:1). In general, the stability constant (conditional stability constant) of considered reactions can be defined as

$$K = \frac{[\text{MeHCit}^-]}{[\text{Me}^{2+}]([\text{H}_2\text{Cit}^{2-}] + [\text{HCit}^{3-}])}$$

where Me^{2+} denotes Mn^{2+} , Co^{2+} , Ni^{2+} or Zn^{2+} ion.

Logarithmic stability constants of the examined metal citrates [MeHCit]⁻ are comparable with the values of the log $K(Me^{2+} + HCit^{3-} \rightleftharpoons MeHCit^{-})$: 3.54–3.67, 4.16–4.83, 4.99–5.11 and 4.25–4.9 for [MnHCit]⁻, [CoHCit]⁻, [NiHCit]⁻ and [ZnHCit]⁻, respectively, as found in the literature [19].

The fairly strong protonation of the carboxyl group not involved in the metal binding seems to impede the formation

Table 1 Thermodynamic parameters of metal binding to Na_3HCit in the buffer solutions with a pH of 6, at 298.15 K and at an ionic strength of I = 100 mM (NaClO₄)

	Metal ion	K/M ⁻¹	$\Delta G^{\rm o}/{\rm kJ}~{\rm mol}^{-1}$	$\Delta H/kJ \text{ mol}^{-1}$	$T\Delta S/kJ \text{ mol}^{-1}$	Stoichiometry
Cacodylate	Mn ²⁺	$(0.247 \pm 0.004) \times 10^4$	-19.31	12.74 (±0.16)	32.05	0.985 (±0.01)
Pipes	Mn^{2+}	$(0.24 \pm 0.002) \times 10^4$	-19.25	10.41 (±0.06)	29.66	0.931 (±0.004)
Mes	Mn^{2+}	$(0.305 \pm 0.002) \times 10^4$	-19.93	8.90 (±0.03)	28.83	0.954 (±0.002)
Cacodylate	Co ²⁺	$(3.57 \pm 0.04) \times 10^4$	-25.99	9.53 (±0.01)	35.52	0.950 (±0.001)
Pipes	Co ²⁺	$(2.89 \pm 0.10) \times 10^4$	-25.24	6.13 (±0.03)	31.55	0.936 (±0.003)
Mes	Co ²⁺	$(4.03 \pm 0.05) \times 10^4$	-26.28	4.64 (±0.01)	30.92	0.968 (±0.001)
Cacodylate	Ni ²⁺	$(10.9 \pm 0.05) \times 10^4$	-28.71	7.06 (±0.03)	35.77	0.969 (±0.003)
Pipes	Ni ²⁺	$(10.3 \pm 0.04) \times 10^4$	-28.60	3.45 (±0.02)	32.05	0.955 (±0.003)
Mes	Ni ²⁺	$(14.5 \pm 0.07) \times 10^4$	-29.49	2.18 (±0.01)	31.67	0.979 (±0.003)
Cacodylate	Zn^{2+}	$(3.53 \pm 0.12) \times 10^4$	-25.97	6.08 (±0.03)	32.05	0.981 (±0.004)
Pipes	Zn^{2+}	$(3.47 \pm 0.13) \times 10^4$	-25.88	3.78 (±0.03)	29.66	1.01 (±0.005)
Mes	Zn^{2+}	$(4.0 \pm 0.2) \times 10^4$	-26.25	2.45 (±0.02)	28.70	0.972 (±0.005)

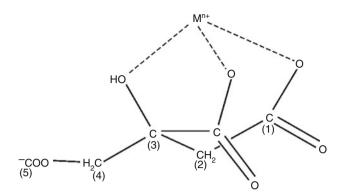


Fig. 1 A diagram indicating coordination of the metal ions to the citrate anion

of binuclear species of the type $[Co_2(C_6H_5O_7)_2(H_2O)_4]^{2-}$ [20] and $[Ni_2(C_6H_5O_7)_2(H_2O)_4]^{2-}$ [21].

The overall energetic effect of the complexation consists mainly of the dehydration of the metal cation and the ligand, $\Delta_{dehyd}H > 0$, on the one hand, and the formation of new ion–ligand bonds, $\Delta_{bind}H < 0$, on the other hand.

In the set of the compounds studied, the energy released by metal ion–citrate interaction is overcompensated by endothermic dehydration of the ion ($\Delta_{dehyd}H > \Delta_{bind}H$). Positive values of the measured reaction enthalpy, ΔH_{obs} , show the thermodynamic stability of the complexes to be strongly dependent on the entropy change and to increase in the order Mn²⁺ < Co²⁺ = Zn²⁺ < Ni²⁺. An additional factor affecting experimental $\Delta_{obs}H$ values is the identity of buffer solution. That is, the enthalpy change decreases with an increase in buffer ionization energy. The energy is the sum of all energetic effects accompanying the reaction, i.e. the enthalpy due to the heat of complex formation, $\Delta_{bind}H$, which is independent of the identity of buffer, and the energy due to proton transfer from the ligand to the buffer [22–24]:

$$\Delta_{\rm obs}H = \Delta_{\rm bind}H + (\Delta n)\Delta_{\rm ion}H_{\rm buf}$$

where $\Delta_{ion}H_{buf}$ is the enthalpy of buffer ionization, and Δn is the number of protons exchanged during binding.

For a given pH value, the relationship between $\Delta_{obs}H$ and $\Delta_{ion}H_{buf}$ is a straight line whose slope corresponds to the number of protons interchanged during the reaction. The ionization energies of the buffers used in this study are -2.97, 11.21 and 14.81 kJ mol⁻¹ for Cacodylate, Pipes and Mes, respectively [25]. The number of protons interchanged during citrate complex formation, determined in this way at a pH of 6, are $0.2(\pm 0.12)$, $0.27(\pm 0.03)$, $0.27(\pm 0.02)$ and $0.19(\pm 0.03)$ for Mn²⁺, Co²⁺, Ni²⁺ and Zn²⁺, respectively (Fig. 2). The $\Delta_{obs}H$ versus $\Delta_{ion}H_{buf}$ relationship is a decreasing function ($\Delta n < 0$), this indicating that in the complexation process the proton is transferred from the ligand onto a buffer component [26].

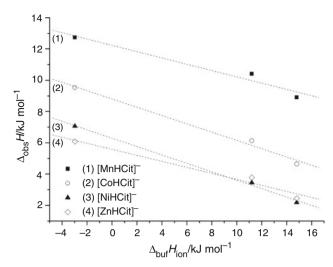


Fig. 2 Plot of $\Delta_{obs}H$ against $\Delta_{ion}H_{buf}$ for the metal–ligand interaction in 10 mM Cacodylate, Pipes and Mes, at a pH of 6, at 298.15 K: (1) Mn²⁺/HCit³⁻; (2) Co²⁺/HCit³⁻; (3) Ni²⁺/HCit³⁻; (4) Zn²⁺/HCit³⁻

Arithmetic means of the enthalpy changes for interaction of the ions with the citrate ligand ($\Delta_{bind}H$), accounting for the number of protons exchanged during complex formation and the ionization enthalpy of the Cacodylate, Pipes and Mes components, are 9.2, 8.83, 6.32 and 5.56 kJ mol⁻¹ for Mn²⁺/HCit³⁻, Co²⁺/HCit³⁻, Ni²⁺/ HCit³⁻ and Zn²⁺/HCit³⁻, respectively. The determined thermodynamic characteristics of the complexes show that the entropy term, $T\Delta S$, has a greater impact on stability of the resulting species than does the enthalpy term ΔH associated with the energy of the donor–acceptor bonds.

Conclusions

Interaction of the metal ions with the citrate ligand in solution of a pH of 6, at 298.15 K, is an endothermic process resulting in the formation of 1:1 complexes. The enthalpy change of the reaction, $\Delta_{bind}H$, accounting for the energy contribution due to binding proton by a buffer component, has been found to depend on the metal ion identity and to decrease in the order $Mn^{2+} > Co^{2+} > Ni^{2+} > Zn^{2+}$. Thermodynamic stability of the complexes is determined by the entropy term that overcompensates the positive value of the enthalpy change. These findings may be useful for optimization of synthetic procedures of the complexes. They would also enable to predict the influence of the presence of metal ions on biological activity of citrate ions.

Acknowledgements This research was supported by the Polish State Committee for Scientific Research under grant DS/8230-4-0088-9.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- 1. Martin RB. Citrate binding of Al^{3+} and Fe^{3+} . J Inorg Biochem. 1989;28:181–7.
- Beinert H, Kennedy MC. Engineering of protein bound ironsulfur clusters. A tool for the study of protein and cluster chemistry and mechanism of iron-sulfur enzymes. Eur J Biochem. 1989;186:5–15.
- Brynhildsen L, Allard B. Influence of metal complexation on the metabolism of citrate by *Klebsiella oxytoca*. Biometals. 1994;7:163–9.
- Krom BP, Warner JB, Konings WN, Lolkema JS. Complementary metal ion specificity of the metal-citrate transporters CitM and CitH of *Bacillus subtilis*. J Bacteriol. 2000;182:6374–81.
- Lippard SJ. Principles of bioinorganic chemistry. Mill Valley, CA: University Science Books; 1994. p. 352.
- Bi J, Wu L, Li Z, Wang X, Fu X. A citrate complex process to prepare nanocrystalline PbBi₂Nb₂O₉ at a low temperature. Mater Lett. 2008;62:155–8.
- 7. Mesquita A, Bernardi MIB, Maia LJQ, Mastelaro VR. Synthesis and characterization of $Pb_{1-x}La_xTiO_3$ nanocrystalline powders. J Therm Anal Calorim. 2007;87:747–51.
- Delmon B. Preparation of heterogeneous catalysts. J Therm Anal Calorim. 2007;90:49–65.
- 9. Fuentes RO, Baker RT. Synthesis of nanocrystalline CeO_2 – ZrO_2 solid solutions by a citrate complexation route: a thermochemical and structural study. J Phys Chem C. 2009;113:914–24.
- Velazquez-Campoy A, Luque I, Freire E. The application of thermodynamic methods in drug design. Thermochim Acta. 2001;380:217–27.
- Freire E. Isothermal titration calorimetry: controlling binding forces in lead optimization. Drug Discov Today Technol. 2004;1:295–9.
- Holdgate GA, Ward WHJ. Measurements of binding thermodynamics in drug discovery. Drug Discov Today. 2005;10:1543–50.
- Deng Y-F, Zhou Z-H, Wan H-L, Ng SW. Δ-Aqua-S-citrato(2-) manganese(II). Acta Crystallogr. 2003;E59:m310–2.

- Zhou Z-H, Deng Y-F, Wan H-L. Structural diversities of cobalt(II) coordination polymers with citric acid. Cryst Growth Des. 2005;5:1109–17.
- Zhang G, Yang G, Ma JS. Versatile framework solids constructed from divalent transition metals and citric acid: syntheses, crystal structures, and thermal behaviors. Cryst Growth Des. 2006; 6:375–81.
- 16. Glusker JP. Citrate conformation and chelation: enzymatic implications. Acc Chem Res. 1980;13:345–52.
- Carrell HL. Metal chelation versus internal hydrogen bonding of the α-hydroxy carboxylate group. J Am Chem Soc. 1987;109: 8067–71.
- Al-Khaldi MH, Nasr-El-Din HA, Mehta S, Al-Aamri AD. Reaction of citric acid with calcite. Chem Eng Sci. 2007;62: 5880–96.
- Sillen LG, Martel AE. Stability constants of metal-ion complexes. Spec. Publ. 17. London, Great Britain: The Chemical Society; 1966.
- 20. Kotsakis N, Raptopoulou CP, Tangoulis V, Terzis A, Giapintzakis J, Jakusch T, et al. Correlations of synthetic, spectroscopic, structural, and speciation studies in the biologically relevant cobalt(II)-citrate system: the tale of the first aqueous dinuclear cobalt(II)-citrate complex. Inorg Chem. 2003;42:22–31.
- Baker EN, Baker HM, Anderson BF, Reeves RD. Chelation of nickel(II) by citrate. The crystal structure of a nickel–citrate complex, K₂[Ni(C₆H₅O₇)(H₂O)₂]₂·4H₂O. Inorg Chim Acta. 1983;78:281–5.
- Baker BM, Murphy KP. Evaluation of linked protonation effects in protein binding reactions using Isothermal Titration Calorimetry. Biophys J. 1996;71:2049–55.
- Fukada H, Takahashi K. Enthalpy and heat capacity changes for the proton dissociation of various buffer components in 0.1 M potassium chloride. Proteins. 1998;33:159–66.
- 24. Haq I, O'Brien R, Lagunavicius A, Siksnys V, Ladbury JE. Specific DNA recognition by the type II restriction endonuclease *MunI*: the effect of pH. Biochemistry. 2001;40:14960–7.
- Goldberg RN, Kishore N, Lennen RM. Thermodynamic quantities for the ionization reactions of buffers. J Phys Chem Ref Data. 2002;31:231–70.
- Gomez J, Freire E. Thermodynamic mapping of the inhibitor site if the aspartic protease endothiapepsin. J Mol Biol. 1995;252: 337–50.