

Open access · Journal Article · DOI:10.12705/632.16

Three species for the price of one within the moss Homalothecium sericeum s.l. — Source link \square

Lars Hedenäs, Aurélie Désamoré, Benjamin Laenen, Beáta Papp ...+5 more authors

Institutions: University of Liège

Published on: 01 Apr 2014 - Taxon (International Association for Plant Taxonomy)

Related papers:

- Integrative taxonomy successfully resolves the pseudo-cryptic complex of the disjunct epiphytic moss Orthotrichum consimile s.l. (Orthotrichaceae)
- Unnoticed diversity within the disjunct moss Orthotrichum tenellum s.l. validated by morphological and molecular approaches
- jModelTest 2: more models, new heuristics and parallel computing.
- Integrative taxonomy of Lepidolejeunea (Jungermanniopsida: Porellales): Ocelli allow the recognition of two neglected species
- Integrative taxonomy resolves the cryptic and pseudo-cryptic Radula buccinifera complex (Porellales, Jungermanniopsida), including two reinstated and five new species



TAXON

Three species for the price of one within the moss Homalothecium sericeum s.l. --Manuscript Draft--

Manuscript Number:	TAXON-D-13-00151R1
Full Title:	Three species for the price of one within the moss Homalothecium sericeum s.l.
Short Title:	Species within Homalothecium sericeum s.l.
Article Type:	Research Article
Keywords:	Bryophytes; Cryptic species; Discriminant analysis; DNA-barcoding; Europe; Macaronesia; Mediterranean
Corresponding Author:	Lars Hedenäs Swedish Museum of Natural History Stockholm, SWEDEN
Corresponding Author Secondary Information:	
Corresponding Author's Institution:	Swedish Museum of Natural History
Corresponding Author's Secondary Institution:	
First Author:	Lars Hedenäs
First Author Secondary Information:	
Order of Authors:	Lars Hedenäs
	Aurélie Désamoré
	Benjamin Laenen
	Beata Papp
	Dietmar Quandt
	Juana Maria González-Mancebo
	Jairo Patiño
	Alain Vanderpoorten
	Michael Stech
Order of Authors Secondary Information:	
Abstract:	Phylogenetic analyses within the moss Homalothecium sericeum resolved two clades and four haplotypes lacking any molecular synapomorphy. Because they exhibit comparable levels of genetic divergence to those observed among sister species in the genus, significant morphological differences, and distinct geographic distributions, they are recognised as three distinct species. Discriminant analysis was employed to assign the types of 'forgotten' taxa previously recognized within H. sericeum s.l. to one of those three species based on their morphology. While a growing number of 'cryptic species' has been mentioned in the literature, the results suggest that thorough morpho-anatomical investigations are likely to reveal morphological discontinuities among such taxa and trigger their formal description at the appropriate taxonomic level. Homalothecium sericeum s.str., H. mandonii (Mitt.) Geh. and H. mediterraneum Hedenäs stat. et nom. nov. clearly differ in sporophytic traits but the identification of sterile specimens is challenged by the overlap in gametophytic characters. As a consequence, 8-37% of the specimens were mis-classified in discriminant analyses in an attempt to find the best combination of gametophytic traits to identify specimens that were assigned to one of the three species on the basis of their genotype. This points to the necessity of developing easy-to-use molecular identification tools in taxonomically challenging plant groups, such as bryophytes. Homalothecium mandonii is the second case of an endemic Macaronesian bryophyte species whose range encompasses the Cape Verde Islands, the Canary Islands, Madeira, and the Azores. Homalothecium

1	Three species for the price of one within the moss <i>Homalothecium sericeum</i> s.l.
2	
3	Lars Hedenäs ¹ , Aurélie Désamoré ² , Benjamin Laenen ² , Beata Papp ³ , Dietmar Quandt ⁴ ,
4	Juana Maria González-Mancebo ⁵ , Jairo Patiño ^{2,5} , Alain Vanderpoorten ^{2*} & Michael
5	Stech ^{6*}
6	
7	1 Swedish Museum of Natural History, Department of Cryptogamic Botany, Box 50007, SE-
8	104 05 Stockholm, Sweden.
9	2 Institute of Botany, University of Liège, B22 Sart Tilman, B-4000 Liège, Belgium
10	3 Botanical Department, Hungarian Natural History Museum, 1476 Budapest, Hungary.
11	4 Nees Institute for Biodiversity of Plants, University of Bonn, Meckenheimer Allee 170,
12	53115 Bonn, Germany.
13	5 Department of Botany, University of La Laguna, La Laguna, Tenerife (Canary Islands),
14	Spain.
15	6 Naturalis Biodiversity Center, Leiden University, PO Box 9517, 2300 RA Leiden, The
16	Netherlands.
17	* contributed equally to this paper
18	
19	Author for correspondence: Lars Hedenäs, lars.hedenas@nrm.se
20	
21	Abstract. Phylogenetic analyses within the moss Homalothecium sericeum resolved two
22	clades and four haplotypes lacking any molecular synapomorphy. Because they exhibit
23	comparable levels of genetic divergence to those observed among sister species in the genus,
24	significant morphological differences, and distinct geographic distributions, they are
25	recognised as three distinct species. Discriminant analysis was employed to assign the types

26 of 'forgotten' taxa previously recognized within *H. sericeum* s.l. to one of those three species based on their morphology. While a growing number of 'cryptic species' has been mentioned 27 28 in the literature, the results suggest that thorough morpho-anatomical investigations are likely 29 to reveal morphological discontinuities among such taxa and trigger their formal description 30 at the appropriate taxonomic level. Homalothecium sericeum s.str., H. mandonii (Mitt.) Geh. 31 and *H. mediterraneum* Hedenäs stat. et nom. nov. clearly differ in sporophytic traits but the 32 identification of sterile specimens is challenged by the overlap in gametophytic characters. 33 As a consequence, 8-37% of the specimens were mis-classified in discriminant analyses in an 34 attempt to find the best combination of gametophytic traits to identify specimens that were 35 assigned to one of the three species on the basis of their genotype. This points to the 36 necessity of developing easy-to-use molecular identification tools in taxonomically 37 challenging plant groups, such as bryophytes. *Homalothecium mandonii* is the second case of 38 an endemic Macaronesian bryophyte species whose range encompasses the Cape Verde 39 Islands, the Canary Islands, Madeira, and the Azores. Homalothecium mediterraneum is 40 circum-Mediterranean and was reported from one locality in Lanzarote in the Canary Islands, 41 while H. sericeum occurs across Europe, eastern North America, and western Asia. The 42 distributions of the segregate species within *H. sericeum* s.l. reinforce the notion that many 43 disjunctions typically observed in moss distribution ranges are due to taxonomic 44 shortcomings and call for the necessity of substantial taxonomic revisions of previously 45 broadly defined bryophyte species.

46

47 Key words: Bryophytes; Cryptic species; Discriminant analysis; DNA-barcoding; Europe;
48 Macaronesia; Mediterranean

49

50 Running head: Species within *Homalothecium sericeum* s.l.

51

52 **INTRODUCTION**

53 The acquisition of the different properties defining daughter species (when they become 54 phenotypically diagnosable, reciprocally monophyletic, reproductively incompatible, 55 ecologically distinct, etc.) is not simultaneous. Before the acquisition of any one of those 56 properties, everyone will agree that there is a single species, and after the acquisition of all, 57 everyone will agree that there are two (De Queiroz, 2007). With the increasing use of 58 molecular characters in systematics, a growing number of studies have revealed the existence 59 of lineages that are well-defined genetically but appear to be indistinguishable by normally 60 used morphological features, and are hence termed 'cryptic taxa' (see Bickford & al., 2007, 61 for review). Cryptic taxa have increasingly been mentioned in the recent literature (Bickford 62 & al., 2007), but are almost never formally described, typified, and named. This reflects the 63 reluctance of taxonomists to describe species only based on molecular characters. As Oliver 64 & Lee (2010) claimed, taxonomists need taxa that can be separated visually, because 65 'portable DNA barcoding probes are many years away, at best'. However, cryptic taxa do not 66 necessarily share a common ancestor (e.g., Goffinet & al., 2007), and in such cases 67 recognition of the genetically divergent but morphologically similar lineages is necessary 68 when species are defined based on the criterion of monophyly (Hutsemékers & al., 2012). By 69 showing that different entities exist, molecular analyses can help to detect species that would 70 otherwise have remained hidden and call for the careful search of morphological differences 71 among seemingly cryptic taxa, which is an essential step towards their effective recognition 72 (e.g., Szweykowski & al., 2005; Vanderpoorten & al., 2010; Sukkharak & al., 2011; Bell & 73 al., 2012; Medina & al., 2012).

Here, we re-analyze previously published molecular data to revisit the significance of
morphological variation and taxonomy of the pleurocarpous moss *Homalothecium sericeum*(Hedw.) Schimp. Within *H. sericeum* s.l., numerous taxa were described during the second

77 half of the 19th and in the beginning of the 20th Century. These were later all synonymised 78 with *H. sericeum* (Hofmann, 1998), and this taxonomic position has been adopted in the most 79 recent check-lists of mosses of European and Macaronesian mosses (Hill & al., 2006; Ros & 80 al., 2013). Recent phylogenetic analyses (Désamoré & al., 2012) showed that accessions of 81 *H. sericeum* s.l. from its entire distribution range belong to three molecular groups, including 82 two sister clades hereafter referred to as *H. mandonii* and *H. sericeum* s.str., and four 83 haplotypes that did not share any synapomorphy and are hereafter referred to as H. 84 mediterraneum (Fig. 1).

85 In the present study, we compare the level of divergence of the molecular lineages 86 identified within H. sericeum s.l. with those observed among other species in the genus. We 87 then use the phylogenetic identity of a representative number of accessions to seek for 88 differences in morphological characters among lineages of the H. sericeum s.l. clade. 89 Because relevant type material is too old for sequencing, we compute a discriminant function 90 that optimizes morphological identification from the sample of molecularly analysed 91 specimens, and assign each type specimen to one of the molecular lineages. Finally, we make 92 the appropriate taxonomic changes based on the morphological and molecular data.

93

94 MATERIAL AND METHODS

95 Specimens representing each haplotype based on *rpl*16 and *atp*B-*rbc*L sequences from all

96 130 accessions of *H. sericeum* s.l. included in Désamoré & al. (2012) were combined with

97 those generated for multiple accessions of each species of the genus by Huttunen & al.

98 (2008). The dataset thus created included 68 accessions in total (Appendix 1).

99 Brachytheciastrum velutinum (Hedw.) Ignatov & Huttunen was employed as outgroup.

100 Indels were scored using simple index coding (Simmons & Ochoterena, 2000) as

101 implemented in the plugin SeqState (Müller, 2004) of PhyDE v0.995 (Müller & al., 2006).

102 The data matrix was submitted to a MP analysis using DNApars as implemented by Seaview 103 4.4.2 (Gouy & al., 2010) with 10 random starts, saving a maximum of 50000 most 104 parsimonious trees and using gaps as informative characters. Support for the branches was 105 assessed through a non-parametric bootstrap analysis with 100 replicates. 106 Fourty-three out of the 130 specimens of Homalothecium sericeum s.l. included by 107 Désamoré & al. (2012) were sampled to represent the morphological variation and 108 distribution range of *H. mediterraneum* (11 accessions), *H. mandonii* (12 accessions), and *H.* 109 sericeum s.str. (20 accessions) (see the Taxonomic treatment below for voucher information). 110 The types of a number of segregate taxa previously recognized within *H. sericeum* s.l., namely Camptothecium aureolum Kindb., Homalothecium sericeum var. meridionale 111 112 M.Fleisch. & Warnst., Hypnum mandonii Mitt., and Homalothecium barbelloides Dixon & 113 Cardot, were also examined. We did not score morphological characters of the type of Leskea 114 sericea Hedw. (Hedwig, 1801). In addition to the fact that Hedwig's type material should not 115 be sampled unless absolutely necessary, the type of L. sericea exhibits the long and narrow 116 leaves that are typical for *H. sericeum* s.str. Furthermore, Hedwig's European type material originates mainly from the non-Mediterranean regions where only the latter occurs. As a 117 118 dioicous species, *H. sericeum* s.l. is infrequently found with sporophytes and only three specimens included in Désamoré & al. (2012) indeed bore them. Therefore, a further 12 119 120 specimens with sporophytes were selected from herbarium material and assigned to one of 121 the three lineages on the basis of their gametophytic traits: six *H. mediterraneum*, five *H*. 122 mandonii, and one H. sericeum s.str. Sporophytes were only studied in one additional 123 specimen of the latter since their character states were already largely studied in previous 124 studies (Hedenäs, 2001, 2012). All morphologically studied specimens are cited under the 125 Taxonomic treatment.

126 Seven gametophytic traits were scored: leaf length (mm); leaf width (mm); leaf length to width ratio; median leaf lamina cell length (μ m); median leaf lamina cell width (μ m); median 127 128 leaf lamina cell length to leaf length ratio; and leaf margin denticulation (finely denticulate, 129 denticulate, strongly denticulate). Initial measurements were made in both stem and branch 130 leaves in three arbitrarily selected specimens of each lineage (*H. mediterraneum*: H69, H78, 131 H86; H. mandonii: H28, H29, H30; H. sericeum s.str.: H16, H19, H91; specimens cited under Taxonomic treatment). Three stem and branch leaves that had reached their final size 132 133 were measured, for median lamina cells the total size range was noted, and for all 134 measurements the mid-point (median) values were used in the comparisons (cf. Hedenäs, 135 1996). These characters showed parallel patterns of variation in the two kinds of leaves, and 136 since it was substantially easier to obtain undamaged branch leaves than stem leaves (results 137 not shown), it was decided to use only branch leaves in order to potentially find 138 distinguishing characters among the three lineages. Leaf lamina cell width did not distinguish 139 the lineages based on the initial three specimens per entity due to too great overlap (branch 140 leaf lamina cell width 4.4-6.9 µm in H. mediterraneum, 4.2-8.4 in H. mandonii, and 4.6-8.4 141 in *H. sericeum* s.str.). This feature was therefore not measured in the remaining material, as it 142 seemed unlikely that it would be a useful character for taxon identification. 143 Shapiro Wilk's W-test and Brown & Forsythe's test showed that the continuous 144 gametophytic variables branch leaf length, leaf width, leaf length to width ratio, median 145 lamina cell length, and median leaf lamina cell length to leaf length ratio did not significantly 146 depart from a normal distribution and did not exhibit significant differences in variance, 147 respectively. Morphological differences between H. sericeum, H. mandonii, and H. 148 *mediterrraneum* depending on the five variables were thus sought using parametric statistics, 149 namely Analysis of variance (ANOVA) and Fisher's Least Significant Difference (LSD) test as implemented by STATISTICA 8.0 (StatSoft, 2008). Linear discriminant analysis (LDA) 150

151 was then employed, using the same program, to identify which is the best combination of 152 morphological variables to identify *H. sericeum*, *H. mandonii*, and *H. mediterraneum*. 153 Variables were selected using backward selection with a probability to stay in the model of p = 0.01. The discriminant functions were employed to assign the types of taxa previously 154 155 recognized within *H. sericeum* s.l. to one of the three species based on their morphological 156 features. To determine what is the actual error rate when attempting at identifying specimens 157 from morphological characters only, a cross-validation procedure, during which each 158 specimen was successively removed from the matrix, was employed.

159

160 **RESULTS**

161 The datamatrix included 1273 characters, of which 94 (22 indels) were parsimony-

162 informative. The MP analysis of *rpl*16 and *atpB-rbcL* in *Homalothecium* resulted in 887

163 equally parsimonious trees of 121 steps, whose strict consensus is presented in Fig. 1. Within

164 *H. sericeum* s.l., a large polytomy comprising four haplotypes labelled as *H. mediterraneum*

as well as a clade holding the accessions of reciprocally monophyletic *H. mandonii* and *H.*

166 *sericeum* s. str. were recovered. Most branches did not receive bootstrap support >50%.

167 Branch leaf length (*H. mediterraneum*, mean \pm standard deviation: 1.74 ± 0.30 ; *H*.

168 *mandonii*: 1.60 ± 0.19 ; *H. sericeum* s.str.: 1.74 ± 0.28 ; p = 0.17--0.98, Fisher's LSD test) and

169 lamina cell width (see above) did not significantly differ among lineages, leaving four

170 continuous characters (leaf width, leaf length to width ratio, median lamina cell length, and

171 median lamina cell length to leaf length ratio; Table 1) as well as denticulation of leaf

172 margins for consideration as potentially discriminative gametophytic characters. Specimens

173 of *H. sericeum* s.str. exhibited significantly narrower leaves and a higher leaf length to width

174 ratio than those of the other clade and the grade according to Fisher's LSD test (Table 1). In

175 *H. mandonii*, the lamina cells were significantly longer and the lamina cell length to leaf

length ratio was significantly higher than in *H. mediterraneum* and *H. sericeum* s.str. (Table
1). All of the investigated characters were, however, overlapping among the three, with an
overlap of 9% in the lamina cell length to leaf length ratio to 61% in leaf width (Table 1). *Homalothecium sericeum* s.str. was further characterized by the strong denticulation or
sometimes weak dentation of the leaf margin in the alar region, with at least some of the teeth
distinctly bent outwards (Fig. 2).

182 Two variables, namely the leaf length to width ratio and the median lamina cell to leaf

183 length ratio, were selected in the LDA. On average, 79% of the specimens were assigned to

184 the correct taxa after cross-validation, which corresponds to a correct classification rate of

185 63%, 92%, and 80% in *H. mediterraneum*, *H. mandonii* and *H. sericeum* s.str., respectively.

186 The type specimens of *H. sericeum* var. *meridionale* and *Hypnum mandonii* were assigned to

187 *H. mediterraneum* and *H. mandonii*, respectively, while the types of *Camptothecium*

188 *aureolum* and *H. barbelloides* were assigned to *H. sericeum* s.str..

189 Sporophytic characters, when available, further distinguished the three taxa. The seta was

rough throughout its length in *H. mandonii* and *H. sericeum* s.str., but completely rough,

191 smooth in the upper 1/4, or occasionally completely smooth, in *H. mediterraneum*. The outer

192 exostome ornamentation was clearly cross-striolate in *H. sericeum* s.str., but smooth or only

193 weakly cross-striolate in *H. mediterraneum* and *H. mandonii*. Finally, the exostome border in

194 the lower portion of the teeth was broad in *H. mediterraneum* but narrow in the two clades

195 (Fig. 3A, E).

Among the three plastid loci (*atp*B-*rbc*L, *rpl*16, *trn*G) investigated by Désamoré & al. (2012), *rpl*16 exhibited three substitutions and one indel (Table 2) within the *H. sericeum* complex, allowing for the unambiguous identification of any specimen that is recent enough

199 for DNA amplification. *Trn*G included one synapomorphic substitution for *H. sericeum s.l.*,

while in *atp*B-*rbc*L one synapomorphic substitution supports *H. sericeum s.str.* and *H. mandonii.*

202

203 **DISCUSSION**

204 The analyses confirmed the monophyly of accessions assigned to *H. mandonii* and *H.*

205 sericeum s. str. Levels of divergence between these two clades were similar to those observed

among other species of the genus. Although these relationships were supported by the strict

207 consensus resulting from the MP analysis of two cpDNA loci and are further fully consistent

with those resolved in other species-level phylogenies of the genus (Huttunen & al., 2008)

and with analyses of the *H. sericeum* s.l. clade (Désamoré & al., 2012), they lacked bootstrap

support in the present analyses. Although a complete set of ITS sequences was produced, we

211 refrained from combining them with the chloroplast data as accessions of morphologically

212 unambiguously identified species were resolved in completely unrelated clades with ITS.

213 Such a strong incongruence among partitions could result from the amplification of

214 paralogous ITS copies, as recently evidenced in mosses (Kosnar & al., 2012), and further

studies are therefore required to explore the utility of that locus in the genus.

216 Although variation in *rpl*16 and *atp*B-*rbc*L was sufficient to discriminate all 217 Homalothecium species recognized to date, the four haplotypes labelled as H. mediterraneum 218 formed a polytomy within *H. sericeum* s.l. One possibility to accommodate those haplotypes 219 taxonomically would be to include them within one of the closely related recognized species, 220 but this would be at odds with their levels of divergence that are of the same order as those 221 observed among sister species in the genus. Alternatively, each of those four haplotypes 222 could be recognized as an individual species. However, as they share the same geographic 223 origin and morphological identity (see below), we rather treat them here as conspecific,

although the lack of any molecular synapomorphy remains unexplained and is at odds with amonophyletic species concept.

226 The discriminant analysis assigned the types of *H. sericeum* var. meridionale and Hypnum mandonii to our groups of accessions labelled as H. mediterraneum and H. mandonii, 227 228 respectively, which are therefore formally recognized here as *H. mediterraneum* Hedenäs 229 stat. et nom. nov. (see Taxonomic treatment below) and H. mandonii (Mitt.) Hedenäs. The 230 new epithet *mediterraneum* was used instead of *meridionale*, since at the species level, the 231 latter is blocked by *H. meridionale* Ravaud, a synonym of *H. sericeum* var. robustum Boulay. 232 The recognition of *H. mediterraneum* and *H. mandonii* parallels previous 'resurrections' of ancient taxa (Rycroft & al., 2004) or de novo descriptions of moss species in the light of 233 234 molecular data (Hutsemékers & al., 2012; Medina & al., 2012). The type of Camptothecium 235 aureolum was unambiguously assigned to the *H. sericeum* s.str. clade, supporting the notion 236 that the former is a synonym of the latter (Hofmann, 1998). The isotype of *H. barbelloides* 237 has very narrow leaves, so that the specimen was assigned to *H. sericeum* by the discriminant 238 analysis, but its lamina cells and the shape and margin denticulation of the basal leaf are 239 similar to *H. mandonii*. The specimen appears to have grown as a pendent epiphyte, which 240 would explain the narrow leaves, and we therefore believe that *H*. *barbelloides* is most likely conspecific with *H. mandonii*. However, the very uncharacteristic leaf shape of this taxon 241 242 makes us refrain from making a formal reduction into synonymy. Unfortunately, only the old 243 type material is available, and the molecular identity could therefore not be determined. 244 Gametophytic traits significantly differ among *H. mandonii*, *H. mediterraneum* and *H.* 245 sericeum s.str., but exhibit substantial overlap (Table 1). In mosses, the gametophyte is the 246 dominant phase and gametophytic traits are largely employed for species identification. 247 During its lifetime the gametophyte is permanently exposed to environmental variation, and 248 is hence prone to plasticity and sometimes convergence (Vanderpoorten & al., 2002; Olsson

249 & al., 2011, 2012; but see Huttunen & al., 2012). Variation in gametophytic traits for 250 taxonomy and species identification might therefore be misleading (Zander & Vitt, 1979; 251 Olsson & al., 2011, 2012; Bell & Hyvönen, 2012; Câmara & Carvalho-Silva, 2013). In the H. sericeum complex and other moss genera where species differentiation mostly relies on 252 253 continuous characters (e.g., Leucobryum, Vanderpoorten & al., 2003; Rhynchostegium, 254 Hutsemékers & al., 2012), specimen identification is challenged by the overlap in characters 255 among species. In the *H. sericeum* complex, this overlap results in a misidentification rate of 256 8-37% when only gametophytic characters are used. Sporophytic traits substantially assist 257 species identification but, as in many of the about 60% of moss species that are dioecious (e.g., Wyatt, 1982; Hedenäs & Bisang, 2011), sporophytes are mostly lacking in the H. 258 259 sericeum complex. Although a combination of gametophytic and sporophytic characters 260 allows distinguishing the three species of the *H. sericeum* complex, as summarised in the 261 identification key given below, the present study points to the necessity of developing easy-262 to-use molecular identification tools. Such molecular identification tools (DNA barcoding 263 markers) will improve biodiversity assessments and ecological research in taxonomically 264 challenging bryophyte groups (e.g., Stech & al., 2013; Lang & Stech, in press). Of the three 265 plastid markers (atpB-rbcL, rpl16, trnG) used in the phylogeographic study of Désamoré & al. (2012), *rpl*16 was the only locus that displayed sufficient levels of variation to on its own 266 267 allow for an unambiguous differentiation between the three species (Table 2). Although 268 *atpB-rbcL* and *trnG* were also partly informative, *rpl*16 can best serve as an easy molecular 269 tool for identifying poorly developed and/or sterile *Homalothecium sericeum* s.l. specimens. 270 The locus was similarly shown to exhibit appropriate levels of inter-specific variation in 271 other genera, such as Plagiomnium (Wyatt & Odrzykoski, 2012), Forsstroemia (Olsson & 272 al., 2012), Leptodon (Sotiaux & al., 2009), and Cratoneuron (Hedenäs, 2011). Rpl16 is not 273 among the regions that have been explored as DNA barcoding marker in mosses so far (see

274 Stech & al., 2013; Lang & Stech, in press; and references therein), but can be considered a potential candidate for distinguishing closely related bryophyte species, although its 275 276 applicability to a wider range of mosses remains to be tested. Contrary to the original idea of species identification based on a single short, standardized DNA region, recent DNA 277 278 barcoding attempts of mosses indicate that different markers (or different combinations of 279 markers) may work best in different moss lineages, including standard markers such as trnL-F and ITS (but see above), but also newly considered regions such as *atpF-atpH* (Hassel & 280 281 al., 2013) or rps19-rpl12 (Lang & Stech, in press). The present results concerning rpl16 are 282 in line with these observations.

283 The three *Homalothecium* species exhibit distinct, albeit slightly overlapping geographic 284 ranges. Homalothecium mandonii is a strict Macaronesian endemic. It is distributed across 285 the four Macaronesian archipelagos, a pattern that is otherwise found in bryophytes only in 286 Exsertotheca intermedia (Brid.) S. Olsson, Enroth & D. Quandt, raising the question of why 287 apparently so vagile organisms failed to reach the North African and South-Western 288 European coasts. *Homalothecium mediterraneum* is a Mediterraneo-Macaronesian endemic 289 that is widespread across the Mediterranean but was found in one locality in Lanzarote in the 290 Canary Islands. Finally, *H. sericeum* is a temperate species distributed across central Europe 291 and the South of Scandinavia, with a few scattered localities in the Mediterranean and eastern 292 North America. The restricted distributions of the segregate species within *H. sericeum* s.l. 293 reinforce the notion that many disjunctions typically observed in moss distribution ranges are 294 due to taxonomic shortcomings (Hutsemékers & al., 2012; Medina & al., 2012) and call for 295 the necessity of substantial taxonomic revisions of previously broadly defined bryophyte 296 species.

297

298 **Taxonomic treatment**

299

300	Homalothecium sericeum (Hedw.) Schimp., Bryol. Eur. 5: 93. 456 (fasc. 46–47 Mon. 3. 1).
301	1851.
302	Basionym: Leskea sericea Hedw., Sp. Musc. Frond. 228. 1801.
303	Synonym: Camptothecium aureolum Kindb., Rev. Bryol. 22: 85. 1895. Lectotype:
304	Norwegen, Opdal, 22/2 80, C.Kaurin (Herb. N. C. Kindberg 19, Herb. H.Möller in S:
305	B17069) (Hofmann, 1998).
306	Plants medium-sized, sometimes small, irregularly or pinnately branched, branches and
307	upper shoot ± strongly curved upwards-inwards when dry. <i>Stem</i> with central strand, a cortex
308	(including epidermis) of 2-3(-4) layers of small and incrassate cells, without hyalodermis;
309	rhizoids inserted at or just below costa insertion, red-brown, slightly branched, smooth;
310	axillary hairs 1-2 per axil, strictly axillary, with 1-3 short, hyaline upper cells, 6.0-12.0 μ m
311	wide, basal cells 1-2, quadrate, hyaline or brownish; pseudoparaphyllia foliose; paraphyllia
312	absent. Stem leaves when moist erect to erecto-patent, when dry erect, straight or slightly
313	homomallous, from ovate-triangular triangular or narrowly triangular base gradually
314	narrowed to longly acuminate apex, not or slightly narrowed towards insertion, slightly
315	concave, plicate; costa single, ending 50-65% way up leaf, 29.5-80.0 µm wide near base,
316	cells on both ad- and abaxial sides linear and similar to adjoining lamina cells, smooth or
317	often ending in a spine on back, in transverse section near base plano-convex, 3-4-stratose,
318	cells homogeneous; margin plane or on one or both sides shortly recurved or reflexed,
319	without border, above finely denticulate or denticulate, sometimes partly entire, around upper
320	alar region mostly distinctly denticulate to dentate, denticles or teeth often spreading or
321	recurved; median leaf lamina cells 36.0-130.0 x 4.0-8.5 μ m, linear, with moderately to longly
322	tapering ends, slightly incrassate, scattered cells sometimes dorsally and distally prorate;
323	basal lamina cells slightly wider and much shorter than median cells, strongly incrassate,

324 more or less porose; *alar cells* triangular, transversely rectangular, guadrate or shortly 325 rectangular, in basal part rectangular and widened, irregular, incrassate, eporose, forming a 326 large and well differentiated, \pm isodiametric or approximately triangular group, extending from leaf margin 25-35% of distance to leaf middle at insertion, decurrent 50-80% way down 327 328 to leaf below. *Branch leaves* smaller and more shortly acuminate than stem leaves, widest 329 0-20% way up, costa ending in a spine, upper margin more strongly denticulate than in stem 330 leaves, median leaf lamina cells 19.0-119.5 x 4.5-8.5 µm, many lamina cells distally and 331 dorsally prorate; median values (three leaves) for leaf width 0.28-0.54 µm, length to width 332 ratio 3.94-5.88, mid-leaf lamina cell length 49.35-82.95 µm, lamina cell length (µm) to leaf 333 length (mm) ratio 29.70-45.64. Sexual condition dioicous, with normal-sized or dwarf male 334 plants. Perigonia lateral on stem, in dwarf males lateral or apical, paraphyses present, in 335 dwarf males absent. Perichaetia lateral on stem; inner perichaetial leaves straight and erect, 336 from ovate or triangular-ovate base suddenly or gradually narrowed to flexuose acumen, 337 longly acuminate, plicate; costa single, weak; margin in acumen denticulate or partly strongly 338 so; paraphyses 6-13 cells long, slightly incrassate. Calyptra cucullate, 3-5-stratose, smooth, 339 naked or with a few basal paraphyses. Seta 9-17 mm long, orange or red, rough throughout, when drv untwisted or dextrorse. Capsule longly cylindrical to longly elongate-ovoid, 340 341 straight, or slightly curved, not furrowed, not constricted at mouth when moist or dry, 342 orthotropous or slightly homotropous; exothecial cells 27.5-65.0 x 15.5-27.5 µm, quadrate to 343 elongate-rectangular, evenly incrassate, smooth, below mouth 2-4 rows of isodiametric or 344 transversely rectangular cells; stomata round-pored; annulus separating, of 2(-3) rows of 345 relatively small cells; operculum longly conical or short-rostrate, basal cells radial, slightly incrassate. *Exostome* reduced, teeth narrow, red or pale reddish, lower outside cross-striolate, 346 347 not furrowed, upper outside papillose or strongly so, margin entire, border in lower portion of 348 teeth narrow, gradually narrowed at transition zone, absent above, primary peristomial layer

349 reduced or strongly reduced. *Endostome* basal membrane low, with short and imperfect

350 processes, hyaline or brownish, papillose, cilia 1-2, short or rudimentary. Spores

 $14.5-23.0(-29.0) \mu m$, papillose, mature in winter half-year.

352 Known geographical distribution: *Homalothecium sericeum* occurs across extra-

353 Mediterranean Europe, eastern North America, western Asia eastwards to the Himalayas and

in Newfoundland (Hofmann, 1998). Earlier reports from China could not be substantiated

355 (Wang & Hu, 2008).

356 Specimens studied (except types; 'D' with number = specimens included in Désamoré &

357 al. (2012): Bulgaria. Strandzha Mts, Veleka reka at Kachul, 5 June 2009, B.Papp (D H82),

BP: 179829; Strandzha Mts., between Kachul and Malko Turnovo, 6 June 2009, B.Papp (D

H6), BP: 179866. Germany. Rheinland-Pfalz, Kr. Ahrweiler, Frahm 8604 (D H39), Herb. J.-

360 P.Frahm BONN. Greece. Chalkidiki, Galatista, 6 April 2006, B.Papp (D H77), BP: 174546.

361 Hungary. Veszprém County, Hálóeresztö, 19 June 2007, *B.Papp* (D H76), BP: 175051.

362 Norway. Finnmark, Söröysund, 24 July 2001, L.Hedenäs (D H25), S: B59862; Troms,

363 Lyngen, 18 July 2003, L.Hedenäs (D H26), S: B82906. Poland. Western Carpathians, Beskid

364 Maly Range, A.Stebel (Musci Macror. Merid. Pol. Exs. 1476) (D H23), S: B157625.

365 Slovakia. Mts Slovenský Raj, Vel'ký Sokol dolina Valley, 3 May 2008, B.Papp (D H10),

366 BP: 176082. Sweden. Åsele Lappmark, Dorotea, 29 June 2004, *L.Hedenäs* (D H57), S:

367 B96087; Medelpad, Torp, 3 October 2006, *L.Hedenäs* (D H59), S: B115534; Öland, E of

368 Skogsby, 28 April 2010, L.Hedenäs (D H19), S: B174820; Östergötland, Krokek, 23 October

369 2006, *L.Hedenäs* (D H56), S: B115519; Södermanland, Kålsö, 5 May 1991, *L.Hedenäs*, S:

370 B52012; Södermanland, Utö, 15 May 2010, *L.Hedenäs* (D H91), S: B175290. Switzerland.

371 Ct. Valais, Zermatt, 9 August 2007, L.Hedenäs (D H46), S: B121137; Ct. Ticino, Cerentino,

372 18 June 2008, L.Hedenäs (D H47), S: B138085. Russia. Krasnodar territory, Anapa district,

373 M.Ignatov & E.Ignatova (Mosses of Russia 24) (D H16), S: B113876; Caucasus, Malyi

Utrish, *Seregin & Lozhkina M-524* D H15), S: B110844. Turkmenistan. Aidere, 2 April
1989, *Levitskaya* (D H52), S: B110843. United Kingdom. England, Shropshire, 12 October
2008, *L.Hedenäs* (D H48), S: B144673.

Homalothecium mediterraneum Hedenäs stat. et nom. nov.

377

378

379	Basionym: Homalothecium sericeum var. meridionale M.Fleisch. & Warnst., Bot.
380	Centralbl. 72: 395. 1897. Isolectotype: (Italy). Sicilia, Mola bei Taormina and
381	Kalkfelsen, 600 m., 6.4.1897, leg. Fleischer (Herb. H.Möller in S: B185199)
382	(Hofmann, 1998). [blocking name at species level: Homalothecium meridionale
383	Ravaud, Mém. Soc. Sc. Nat. Cherbourg 37: 303. 1910, nom. nud. in synon. ($\equiv H$.
384	sericeum var. robustum Boulay)]
385	Homalothecium sericeum var. tunetanum Besch. in Patouillard, Expl. Sci. Tunisie Bot. 8:
386	11. 1897 (lectotype in PC, Hofmann, 1998).
387	Plants medium-sized, sometimes small, irregularly or pinnately branched, branches and
388	upper shoot \pm strongly curved upwards-inwards when dry. <i>Stem</i> with central strand, a cortex
389	(including epidermis) of 2-4 layers of small and incrassate cells, without hyalodermis;
390	rhizoids inserted at or just below costa insertion, red-brown, hardly to moderately strongly
391	branched, smooth; axillary hairs 1-2 per axil, strictly axillary, with 1-4 short, hyaline upper
392	cells, 8.5-10.5 μ m wide, basal cells 1-2, transversely rectangular to shortly rectangular,
393	brownish; pseudoparaphyllia foliose; paraphyllia absent. Stem leaves when moist erect to
394	patent, when dry erect, straight or slightly homomallous, from cordate-triangular or rounded-
395	triangular base gradually narrowed to longly acuminate apex, markedly narrowed towards
396	insertion, slightly concave, plicate; <i>costa</i> single, ending 55-75% way up leaf, 38.0-73.5 μ m
397	wide near base, cells on both ad- and abaxial sides linear and similar to adjoining lamina
398	cells, smooth or often ending in a spine on back, in transverse section near base plano-

Fig. 3A-D

399 convex, 3-4-stratose, cells homogeneous; margin plane or on one or both sides entirely or 400 partly broadly recurved, without border, entire or finely denticulate, denticles around upper 401 alar region weak or absent, rarely bent slightly outwards; median leaf lamina cells 29.5-92.5 402 x 4.0-7.5 µm, linear, with shortly to moderately tapering ends, slightly incrassate, scattered 403 cells sometimes dorsally and distally prorate; *basal lamina cells* wider and much shorter than 404 median cells, incrassate, eporose; *alar cells* quadrate, transversely rectangular, rhomboidal, 405 especially towards insertion rectangular, in distal portion of group often irregular, slightly 406 incrassate, eporose, forming a large and well differentiated, \pm isodiametric or along margin 407 slightly elongate group, extending from leaf margin 25-35% of distance to leaf middle at insertion, decurrent 20-50% way down to leaf below. Branch leaves smaller and more ovate 408 409 than stem leaves, widest 15-30% way up, costa ending in a spine, upper margin more strongly denticulate than in stem leaves, median leaf lamina cells 25.0-115.5 x 4.5-7.0 µm, 410 411 many lamina cells distally and dorsally prorate; median values (three leaves) for leaf width 0.34-0.56 µm, length to width ratio 3.03-4.26, mid-leaf lamina cell length 46.20-81.90 µm, 412 413 lamina cell length (µm) to leaf length (mm) ratio 24.75-48.21. Sexual condition dioicous, 414 with normal-sized or dwarf male plants. Perigonia lateral on stem, in dwarf males lateral or 415 apical, paraphyses present, in dwarf males few. Perichaetia lateral on stem; inner perichaetial 416 leaves straight and erect, from ovate or ovate-oblong base suddenly or gradually narrowed to 417 flexuose acumen, acuminate, smooth or weakly plicate; costa single, weak; margin in acumen 418 denticulate or finely so, at shoulder strongly so or partly dentate; paraphyses 8-19 cells long, 419 slightly incrassate. Calvptra cucullate, 3-5-stratose, smooth, naked. Seta 8-15 mm long, red, 420 rough almost throughout or above weakly so or smooth, occasionally smooth almost 421 throughout, when dry dextrorse. *Capsule* cylindrical, shortly or gradually narrowed towards 422 mouth, straight, or mouth slightly oblique, not furrowed, often constricted at mouth when 423 moist but not more so when dry, orthotropous; exothecial cells 21.0-65.0 x 12.5-40.0 µm,

424 quadrate or shortly to longly rectangular, evenly incrassate or slightly collenchymatous with superficial thickenings, smooth, below mouth 1-4 rows of isodiametric, shortly transversely 425 426 rectangular, or rectangular cells; stomata round-pored, occasionally ovate-pored; annulus separating, of 1-3 rows of relatively small cells; operculum conical or rostrate, basal cells 427 428 radial, incrassate. *Exostome* reduced, teeth short and narrow, yellow-brown or pale yellow-429 brown, lower outside indistinctly cross-striolate to smooth, not furrowed, upper outside 430 strongly papillose, margin entire or irregular, border in lower portion of teeth broad, 431 gradually narrowed at transition zone, absent above, primary peristomial layer strongly 432 reduced. Endostome basal membrane low, with short and imperfect or vestigial processes, hyaline, vestigial processes sometimes brownish, papillose above, sometimes more finely so 433 434 below, cilia 1-3, vestigial. Spores 13.0-21.0 µm, finely or strongly papillose, mature in winter 435 half-year. 436 According to Hofmann (1998), H. sericeum var. meridionale (H. mediterraneum) and H. 437 sericeum var. tunetanum differ from H. sericeum s. str. only in their smooth seta. Because the 438 often occurring smooth or partly smooth seta is one feature that distinguishes H. 439 *mediterraneum* from the other two species recognized here (Table 2), *Homalothecium* 440 sericeum var. tunetanum is considered as a synonym of H. mediterraneum. Known geographical distribution: Homalothecium mediterraneum is circum-441 442 Mediterranean and is known from one locality in Lanzarote in the Canary Islands (Désamoré 443 & al., 2012). 444 Specimens studied (except types; 'D' with number = specimens included in Désamoré & 445 al. (2012): Cyprus. Trodos Mts, at Pano Platres village, 18 September 2001, B.Papp (D

446 H44), BP: 177881; Troodos Gebirge, Pano Platres, Frahm 200691 (D H31), Herb. J.-

447 P.Frahm BONN. Greece. Crete, Chania, Frahm K-158 (D H34), Herb. J.-P.Frahm BONN;

448 Crete, Irakleion region, 18 April 2001, B.Papp (D H86), BP: 170680; Crete, environs de

- 449 Heraklion, A.Désamoré, B.Laenen H108 (D H108), LG; Kephissos, Attica, 1873,
- 450 DeHeldreich 138, S B185184; Péloponnèse, Kalavryta, A.Désamoré, B.Laenen H110 (D
- 451 H110), LG; Péloponnèse, Sella, A.Désamoré, B.Laenen H115 (D H115), LG; Rhodos Island,
- 452 Mt. Profitis Ilias, 7 August 2006, *B.Papp* (D H78), BP: 174611. Italy. Sardinia merid.:
- 453 Iglesias, K.Holmen 70-34, S: B185180; Sardinia merid.: Iglesias, K.Holmen 71-53, S:
- 454 B185181. Montenegro. Lovcen Mts, Njegusi village to Kotor, 12-13 October 2003, B.Papp
- 455 (D H69), BP: 174306. Spain. Granada (Alpujarra de la Sierra). Sierra Nevada, May 2006,
- 456 R.M.Ros, S.Rams (D H50), S: B135972. Canary Islands. Lanzarote, Heleches,
- 457 A.Vanderpoorten HS41 (D HS41), LG. Tunisia. Aim Draham, C.J.Pirad 65, S: B185187.
- 458 Turkey. Prov. Aydin, north side of Samsundag, *E.Nyholm 335/71*, S: B99830; Prov. Mugla,
- 459 mainroad Marmaris-Mugla, *E.Nyholm* 475/71, S: B99831.
- 460
- 461 Homalothecium mandonii (Mitt.) Geh., Flora 69: 348. 1886. Fig. 3E-H
- 462 Basionym: *Hypnum mandonii* Mitt., Nat. Hist. Azores 311. 1870. Isolectotypes: 'Mandon:
- 463 Mousees de Madère no. 36. Homalothecium sericeum Sch. Rib. das Cales, Mars' (S:
- 464 B8824); 'Homalothecium sericeum, Madeira, Rib. d. Cales, Mandon 36, Mars.' (Herb.
- 465 H.Möller in S: B8823) (Hofmann, 1998).
- 466 *Homalothecium sericeum* var. *meridionale* Schimp. in Geh., Flora 69: 349. 1886, nom.
 467 *nud*. (Geheeb, 1886)
- 468 Plants medium-sized, sometimes small, irregularly pinnately branched, branches sometimes
- 469 turning to new stems, branching angle relatively narrow, branches straight or curved
- 470 upwards-inwards when dry. *Stem* with central strand, a cortex (including epidermis) of
- 471 1-2(-3) layers of small and incrassate cells, without hyalodermis; *rhizoids* inserted at or just
- 472 below costa insertion, red-brown, not or slightly branched, smooth; axillary hairs 1-3 per
- 473 axil, strictly axillary, with 1-2 short, hyaline upper cells, 10.0-12.5 μm wide, basal cells 1-2,

474 transversely rectangular to shortly rectangular, brown; *pseudoparaphyllia* foliose; 475 paraphyllia absent. Stem leaves when moist erect to patent, when dry erect, straight or 476 slightly homomallous, triangular or ovate-triangular, from shortly above insertion gradually 477 narrowed to acuminate apex, distinctly constricted at insertion, concave, strongly plicate; 478 costa single, ending 50-75% way up leaf, 31.5-63.0 µm wide near base, cells on both ad- and 479 abaxial sides linear and similar to adjoining lamina cells, smooth, in transverse section near 480 base plano-convex, 4(-5)-stratose, cells homogeneous; *margin* plane or shortly to longly 481 weakly reflexed, without border, finely denticulate throughout, a few denticles around upper 482 alar region sometimes stronger, rarely bent slightly outwards; median leaf lamina cells 483 46.0-178.5 x 4.0-8.0 µm, linear, with moderately to longly tapering ends, slightly incrassate 484 or incrassate, smooth or slightly distally and dorsally prorate; basal lamina cells wider and 485 much shorter than median cells, strongly incrassate, porose; *alar cells* in upper portion of 486 group transversely rectangular, quadrate, or rectangular, often rounded, below rectangular to 487 longly rectangular, incrassate, slightly porose, forming a rounded, shortly oblong, or 488 triangular group, extending from margin 35-40% of distance to leaf middle at insertion, 489 decurrent or shortly so. Branch leaves smaller than stem leaves, widest 15-25% way up, costa 490 occasionally ending in small, obtuse spine, upper margin denticulate, median leaf lamina 491 cells 33.5-147.0 x 4.0-8.5 µm, occasional lamina cells distally and dorsally prorate; median 492 values (three leaves) for leaf width 0.37-0.61 um, length to width ratio 2.96-3.60, mid-leaf 493 lamina cell length 65.10-103.95 µm, lamina cell length (µm) to leaf length (mm) ratio 494 44.93-59.64. Sexual condition dioicous, with normal-sized or dwarf male plants. Perigonia 495 lateral on stem, in dwarf males lateral or apical, paraphyses present, in dwarf males few. 496 Perichaetia lateral on stem and branch bases; inner perichaetial leaves straight and erect, 497 narrowly ovate or triangular ovate, above shortly narrowed to acumen, apex narrowly 498 acuminate, smooth or plicate; costa single, weak, indistinct; margin in acumen entire or

499 weakly denticulate, at shoulder denticulate, strongly so, or with single teeth, not or weakly 500 bordered; paraphyses 6-19 cells long, incrassate. Calyptra cucullate, 3-5-stratose, smooth or 501 sometimes with one low ridge, naked. Seta 11-22 mm long, red, rough throughout, when dry 502 dextrorse. *Capsule* ovoid to cylindrical, not furrowed, not or when dry sometimes weakly 503 constricted below mouth, orthotropous or almost so; exothecial cells 21.0-50.5 x 12.5-31.5 504 um, quadrate or rectangular, slightly evenly incrassate or longitudinal walls incrassate, not collenchymatous, smooth, below mouth 1-4 rows of small, rectangular, transversely 505 506 rectangular, or quadrate cells; stomata round-pored; annulus separating, of 1-3 rows of 507 relatively small cells; operculum shortly rostrate, basal cells radial, incrassate. Exostome reduced or strongly so, teeth narrow and sometimes short, light orange-brown, lower outside 508 509 weakly cross-striolate to smooth, not furrowed, upper outside papillose to almost smooth, 510 margin entire, border absent, or present, narrow, and gradually narrowed upwards, primary 511 peristomial layer strongly reduced. *Endostome* basal membrane low, with short and imperfect 512 or vestigial processes, yellowish or brownish, papillose or finely so, cilia 0-2(-3), absent or 513 vestigial. Spores 14.5-22.0 µm, rather strongly papillose, mature in winter half-year. 514 In the original description of Hypnum mandonii Mitt. (Mitten, 1870), this taxon was said 515 to have more closely imbricate leaves than *H. sericeum*, and acute rather than more narrowly 516 pointed leaves. The latter is probably to some degree reflected in the narrower leaves that 517 widen from closer to the leaf insertion in *H. sericeum* than in *H. mandonii*. 518 Known geographical distribution: Homalothecium mandonii is a Macaronesian endemic 519 distributed across Cape Verde, all of the Canary Islands, Madeira, and the Azorean islands 520 São Miguel and Santa Maria 521 (http://www.azoresbioportal.angra.uac.pt/listagens.php?sstr=3&lang=en; accessed 15 August

522 2013).

- 523 Specimens studied (except types; 'D' with number = specimens included in Désamoré &
- 524 al. (2012): Canary Islands. El Hierro, Arbol Santo, I.Hildebrandt 06-13 (D H30), L; El
- 525 Hierro, El Brezal, I.Hildebrandt 06-17 (D H29), L; Hierro, Riscos de Sabinosa, March 1906,
- 526 C.J.Pitard, S: B185186; Fuerteventura, A.Vanderpoorten HS43 (D HS43), LG; La Palma, los
- 527 Tilos, A.Vanderpoorten 1577 (D HS25), LG; Lanzarote, Peñas del Cache, A.Vanderpoorten
- 528 HS39 (D HS39), LG; Lanzarote, Pico de la Zarza, A.Vanderpoorten HS32 (D HS32), LG;
- 529 Tenerife, Anaga-Gebirge, Abts 6145 (D H33), Herb. J.-P.Frahm BONN. Cape Verde. Santo
- 530 Antão, Ribeira da Torre, A.Lindlar 193 (D H32), Herb. J.-P.Frahm BONN. Madeira. Lapa
- do Passo, Mandon 37, S: B8799; Pico Arieiro to Pico Ruivo, Stech 04-387 (D H28), L;
- 532 Rabaçal, Cascata de Risco, Frahm M-138 (D H67), Herb. J.-P.Frahm BONN; Ribeira Brava,
- 533 Pico Ferreiro, 8 May 2007, L.Hedenäs (D H27), S: B119087; São Jorge, Ribeira Grande,
- 534 S.Fontinha, L.Hedenäs MA91-199 (D H88), S: B8835; Ribeiro Frio, 3 January 1880,
- 535 *R.Fritze*, S: B8831; Ribeiro Frio, 8 April 1864, *L.Kny*, S: B8833.
- 536
- 537 Taxon with uncertain position
- 538 Homalothecium barbelloides Dixon & Cardot, J. Bot. 49: 6. 2. 1911.
- 539 Isotype: (Canary Islands). Teno Mts., Teneriffe, Evergreen woods, Dec. 1909, leg. Dr.
- 540 Salter; No. 52. Ex herb. H.N.Dixon. Orig. spec. (Herbarium G.Roth. 1842-1915, Herb.
- 541 H.Möller in S; reg. no. B107660) (Hofmann, 1998).
- 542

547

- 543 *Key to the European and Macaronesian species of* Homalothecium sericeum *s.l.*
- 1. Branch leaves 3.9-5.9 times as long as wide, mostly widest 0-20% way up leaf; margin
- 545 denticulation at alar region well developed, with at least some teeth distinctly bent
- 546 outwards (Fig. 2). Exostome distinctly cross-striolate on lower outside.
 - *H. sericeum* s.str.

548	1. Branch leaves 3.0-4.3 times as long as wide, mostly widest 15-30% way up leaf; margin
549	denticulation at alar region weak or absent, teeth rarely and only slightly bent outwards
550	(Fig. 3C, G). Exostome smooth or weakly cross-striolate on lower outside. 2
551	2. Ratio between branch leaf lamina median cell length (μ m)/median leaf length (mm)
552	24.7-48.2. Seta frequently partly or entirely smooth; exostome border broad (Fig. 3A).
553	H. mediterraneum
554	2. Ratio between branch leaf lamina median cell length (μ m)/median leaf length (mm)
555	44.9-59.6. Seta rough throughout; exostome border narrow (Fig. 3E).
556	H. mandonii
557	
558	ACKNOWLEDGEMENTS
559	We thank Ina Hildebrandt for providing material included in the study. Comments by two
560	reviewers significantly improved the manuscript. This research was funded through an
561	Integrating Research Grant (IRG) of the European Distributed Institute of Taxonomy (EDIT).
562	
563	LITERATURE CITED
564	Bell, N. & Hyvönen, J. 2012. Gametophytic simplicity in Laurasian and Gondwanan
565	Polytrichopsida — the phylogeny and taxonomy of the Oligotrichum morphology. J.
566	Bryol. 34: 160172.
567	Bell, D., Long, D.G., Forrest, A.D., Hollingsworth, M.L., Blom, H.H. & Hollingsworth,
568	P.M. 2012. DNA barcoding of European Herbertus (Marchantiopsida, Herbertaceae)
569	and the discovery and description of a new species. Molec. Ecol. Res. 12: 3647.
570	Bickford, D., Lohman, D.J., Sodhi, N.S., Ng, P.K.L., Meier, R., Winker, K., Ingram,
571	K.K. & Das, I. 2007. Cryptic species as a window on diversity and conservation.
572	Trends Ecol. Evol. 22: 148155.

- 573 Câmara, P. & Carvalho-Silva, M. 2013. The genus Schroeterella Herzog
- 574 (Sematophyllaceae) revisited. *Bryologist* 116: 146--148.
- 575 De Queiroz, K. 2007. Species concepts and species delimitation. *Syst. Biol.* 56: 879--886.
- 576 Désamoré, A., Laenen, B., Stech, M., Papp, B., Hedenäs, L., Mateo, R.G. &
- 577 **Vanderpoorten, A.** 2012. How do temperate bryophytes face the challenge of a
- 578 changing environment? Lessons from the past and predictions for the future. *Global*
- 579 *Change Biol.* 18: 2915--2924.
- 580 Geheeb, A. 1886. Bryologische Fragmente. *Flora* 69: 339–353.
- 581 Goffinet, B., Buck, W.R. & Wall, M.A. 2007. Orthotrichum freyanum (Orthotrichaceae), a
 582 new epiphytic moss from Chile 1. Nova Hedwigia Beih. 131: 1--11.
- 583 Gouy, M., Guindon, S. & Gascuel, O. 2010. SeaView Version 4: A multiplatform graphical
 584 user interface for sequence alignment and phylogenetic tree building. *Mol. Biol. Evol.*585 27: 221-224.
- 586 Hassel, K., Segreto, R. & Ekrem, T. 2013. Restricted variation in plant barcoding markers
- 587 limits identification in closely related bryophyte species. *Mol. Ecol. Resour.* 13: 1047588 -1057.
- Hedenäs, L. 1996. On the interdependence of some leaf characters within the *Drepanocladus aduncus-polycarpus* complex. J. Bryol. 19: 311--324.
- Hedenäs, L. 2001. Environmental factors potentially affecting character states in
 pleurocarpous mosses. *Bryologist* 104: 72--91.
- Hedenäs, L. 2011. Relationships among *Cratoneuron curvicaule*, *C. filicinum* var. *filicinum*,
 and *C. filicinum* var. *atrovirens* (Bryophyta: Amblystegiaceae). *J. Bryol.* 33: 99--104.
- 595 **Hedenäs**, L. 2012. Morphological and anatomical features associated with epiphytism
- among the pleurocarpous mosses one basis for further research on adaptations and
- 597 their evolution. J. Bryol. 34: 79--100.

598	Hedenäs, L. & Bisang, I. 2011. The overlooked dwarf males in mosses - Unique among
599	green land plants. Perspect. Plant Ecol. Evol. Syst. 13: 121135
600	Hedwig J. 1801. Species muscorum frondosorum. Lipsiae: Joannis Ambrosii Barthii.
601	Hill, M.O., Bell, N., Bruggeman-Nannenga, M.A., Brugués, M., Cano, M.J., Enroth, J.,
602	Flatberg, K.I., Frahm, JP., Gallego, M.T., Garilleti, R., Guerra, J., Hedenäs, L.,
603	Holyoak, D.T., Hyvönen, J., Ignatov, M.S., Lara, F., Mazimpaka, V., Muñoz, J.
604	& Söderström, L. 2006. An annotated checklist of the mosses of Europe and
605	Macaronesia. J. Bryol. 28: 198267.
606	Hofmann, H. 1998. A monograph of the genus Homalothecium (Brachytheciaceae, Musci).
607	Lindbergia 23: 119159.
608	Hutsemékers, V., Vieira, C.C., Ros, R.M., Huttunen, S. & Vanderpoorten, A. 2012.
609	Morphology informed by phylogeny reveals unexpected patterns of species
610	differentiation in the aquatic moss Rhynchostegium riparioides s.l. Molec.
611	Phylogenet. Evol. 62: 748755.
612	Huttunen, S., Hedenäs, L., Ignatov, M.S., Devos, N. & Vanderpoorten, A. 2008. Origin
613	and evolution of the northern hemisphere disjunction in the moss genus
614	Homalothecium (Brachytheciaceae). Amer. J. Bot. 95: 720730.
615	Huttunen, S., Olsson, S., Buchbender, V., Enroth, J., Hedenäs, L. & Quandt, D. 2012.
616	Phylogeny-based comparative methods question the adaptive nature of sporophytic
617	specializations in mosses. PloS One 7: e48268.
618	Košnar, J., Herbstová, M., Kolář, F., Koutecký, P. & Kučera, J. 2012. A case study of
619	intragenomic ITS variation in bryophytes: Assessment of gene flow and role of
620	polyploidy in the origin of European taxa of the Tortula muralis (Musci: Pottiaceae)
621	complex. <i>Taxon</i> 61: 709-720.

	622	Lang, A. S. & Stech	, M. (ir	n press) What's in a nam	ne? Disentangling	g the Dicranun
--	-----	---------------------	----------	---------	-------------------	-------------------	----------------

623 scoparium species complex (Dicranaceae, Bryophyta). Syst. Bot.

624 Medina, R., Lara, F., Goffinet, B., Garilleti, R. & Mazimpaka, V. 2012. Integrative

- taxonomy successfully resolves the pseudo-cryptic complex of the disjunct epiphytic
 moss *Orthotrichum consimile* (Orthotrichaceae). *Taxon* 61: 1180--1198.
- 627 Mitten, W. 1870. Musci. Pp. 288--316 in: Godman, F.D.C. (ed.), Natural history of the
- 628 *Azores, or western islands*. London: John van Voorst.
- Müller, K. 2004. SeqState primer design and sequence statistics for phylogenetic DNA
 data sets. *Applied Bioinformatics* 4: 65--69.
- 631 Müller, K., Müller, J., Neinhuis, C. & Quandt, D. 2006. PhyDE Phylogenetic Data

Editor, v0.995. Program distributed by the authors http://www.phyde.de

- 633 Oliver, P.M. & Lee, M.S.Y. 2010. The botanical and zoological codes impede biodiversity
 634 research by discouraging publication of unnamed new species. *Taxon* 59: 1201--1205.
- 635 Olsson, S., Enroth, J., Buchbender, V., Hedenäs, L., Huttunen, S. & Quandt, D. 2011.
- 636 *Neckera* and *Thamnobryum* (Neckeraceae, Bryopsida): paraphyletic assemblages.
- 637 *Taxon* 60: 36--50.
- 638 Olsson, S., Enroth, J., Huttunen, S. & Quandt, D. 2012. Forsstroemia Lindb.
- 639 (Neckeraceae) revisited. J. Bryol. 34: 114--122.
- 640 Ros, R.M., Mazimpaka, V. Abou-Salama, U., Aleffi, M., Blockeel, T.L., Brugués, S.M.,
- 641 Cros, R.M., Dia, M.G., Dirkse, G.M., Draper, I., El-Saadawi, W., Erda, I.,
- 642 Ganeva, A., Gabriel, R., Gonzalez-Mancebo, J.M., Granger, C., Herrnstadt, I.,
- 643 Hugonnot, V., Khalil, K., Kürschner, H., Losada-Lima, A., Luis, L., Mifsud, S.,
- 644 Privitera, M., Puglisi, M., Sabovljevic, M., Sérgio, C., Shabbara, H.M., Sim-
- 645 Sim, M., Sotiaux, A., Tacchi, R., Vanderpoorten, A., Werner, O. 2013. Mosses of
- the Mediterranean, an annotated checklist. *Cryptog.*, *Bryol.* 34: 99--283.

647	Rycroft, D. S., Groth, H. & Heinrichs, H. 2004. Reinstatement of Plagiochila maderensis
648	(Jungermanniopsida: Plagiochilaceae) based on chemical evidence and nrDNA ITS
649	sequences. J. Bryol. 26: 3745.
650	Simmons, M.P. & Ochoterena, H. 2000. Gaps as characters in sequence-based phylogenetic
651	analysis. Syst. Biol. 49: 369-381.
652	Sotiaux, A., Enroth, J., Olsson, S., Quandt, D. & Vanderpoorten, A. 2009. When
653	morphology and molecules tell us different stories: a case-in-point with Leptodon
654	corsicus, a new and unique endemic moss species from Corsica. J. Bryol. 31: 186
655	196.
656	StatSoft, I. 2008. STATISTICA (data analysis software system), version 8.0.
657	(http://www.statsoft.com).
658	Stech, M., Veldman, S., Larraín, J., Muñoz, J., Quandt, D., Hassel, K. & Kruijer, H.
659	2013. Molecular species delimitation in the Racomitrium canescens complex
660	(Grimmiaceae) and implications for DNA barcoding of species complexes in mosses.
661	<i>PloS One</i> 8: e53134.
662	Sukkharak, P., Gradstein, S.R. & Stech, M. 2011. Phylogeny, taxon circumscriptions and
663	character evolution in the core Ptychanthoideae (Lejeuneaceae, Marchantiophyta).
664	<i>Taxon</i> 60: 16071622.
665	Szweykowski, J., Buczkowska, K. & Odrzykoski, I.J. 2005. Conocephalum salebrosum
666	(Marchantiopsida, Conocephalaceae) - a new Holarctic liverwort species. Plant Syst.
667	<i>Evol</i> . 253: 133158.
668	Vanderpoorten, A., Boles, S. & Shaw, A.J. 2003. Patterns of molecular and morphological
669	variation in Leucobryum albidum, L. glaucum, and L. juniperoideum. Syst. Bot. 28:
670	651656.

Vanderpoorten, A., Hedenäs, L., Cox, C.J. & Shaw, A.J. 2002. Phylogeny and
morphological evolution of the Amblystegiaceae (Bryophyta, Musci). Mol.
Phylogenet. Evol. 23: 121.
Vanderpoorten, A., Schäfer-Verwimp, A., Heinrichs, J., Devos, N. & Long, D.G. 2010.
The taxonomy of the leafy liverwort genus Leptoscyphus (Lophocoleaceae) revisited.
<i>Taxon</i> 59: 176186.
Wang, Yf. & Hu, Rl. 2008. Brachytheciaceae. Pp. 71167 in: Hu, Rl., You, Yf.,
Crosby, M. R. & He, S. (ed.), Moss flora of China. English version. Volume 7.
Amblystegiaceae-Plagiotheciaceae. Beijing & St. Louis: Science Press & Missouri
Botanical Garden.
Wyatt, R. 1982. Population ecology of bryophytes. J. Hattori Bot. Lab. 52: 179198.
Wyatt, R. & Odrzykoski, I.J. 2012. Plagiomnium floridanum sp. nov. (Mniaceae), a new
moss from the southeastern United States. The Bryologist 115: 527535.
Zander, R. H. & Vitt, D.H.1979. Gametophytic distinctions of Zygodon (Orthotrichaceae),
Anoectangium and Leptodontium (Pottiaceae) and the status of Anoectangium
rubrigemmium of Hawaii. Canad. J. Bot. 57: 292296.

Table 1 Branch leaf and sporophyte characters that differentiate the three *Homalothecium sericeum* s.l. species. 1--4: Average and standard deviation of the median (mid-point) values of measured characters in *Homalothecium mediterraneum* (n = 11), *H. mandonii* (n = 12), and *H. sericeum* s. str. (n = 20) and median values of the measures from type specimens of *Camptothecium aureolum*, *Homalothecium sericeum* var. *meridionale*, *Hypnum mandonii* and *Homalothecium barbelloides*. Minimum and maximum values for each variable are shown in parentheses. Measurements highlighted in bold in one species indicate a significant difference (p < 0.05) from those observed in the two other species according to Fisher's LSD test. 5--9. Other branch leaf and sporophyte characters that differentiate the three *Homalothecium* species.

	1. Leaf	2. Leaf	3. Median	4. Median	5.Leaf	6. Alar leaf	7. Seta	8. Lower	9.
	width	length to	lamina cell	lamina cell	widest at	margin	ornamentation	exostome	Exostome
	(mm)	width ratio	length (μm)	length (μm)		denticulation		outside	border
				to leaf					
				length (mm)					
				ratio					
H. mediterraneum	(0.34)	(3.03)	(46.20)	(24.75)	15-30%	Weak or	Rough, smooth	Smooth or	Broad
	0.48±0.02	3.60±0.11	61.09±3.44	35.92±2.32	above	absent, rarely	in upper ¼, or	weakly	(Fig. 3A)
	(0.56)	(4.26)	(81.90)	(48.21)	leaf base	bent slightly	occasionally	cross-	

						outwards	completely	striolate	
							smooth		
H. mandonii	(0.37)	(2.96)	(65.10)	(44.93)	15-25%	Weak,	Rough	Smooth or	Narrow
	0.49±0.01	3.29±0.06	88.26±3.64	55.05±1.27	above	sometimes a	throughout	weakly	(Fig. 3E
	(0.61)	(3.60)	(103.95)	(59.64)	leaf base	few stronger		cross-	
						teeth, rarely		striolate	
						bent			
						outwards			
H. sericeum	(0.28)	(3.94)	(49.35)	(29.70)	0-20%	Mostly	Rough	Cross-	Narrow
	0.38±0.01	4.61±0.10	65.31±2.12	37.92±1.02	above	strong,	throughout	striolate	
	(0.54)	(5.88)	(82.95)	(45.64)	leaf base	occasionally			
						dentate, often			
						some teeth			
						bent			
						outwards			
C. aureolum	0.22	4.62	52.50	51.55	-	-	-	-	-

H. sericeum var.	0.50	3.23	71.40	43.87	-	-	-	-	-
meridionale									
H. mandonii	0.48	3.06	95.55	64.54	-	-	-	-	-
H. barbelloides	0.20	5.63	79.80	69.09	-	-	-	-	-

Table 2. Species-specific polymorphisms in the *rpl*16 gene among *Homalotheciummandonii*, *H. sericeum*, and *H. mediterraneum*.

Position	594	703	619	832
H. mandonii	С	Poly-A (8 repeats)	Т	А
H. sericeum	Т	Poly-A (9 repeats)	С	G
H. mediterraneum	Т	Poly-A (9 repeats)	Т	А

Appendix 1. Voucher information and Genbank accession numbers of the specimens of *Homalothecium* included in the present phylogenetic analysis. *Hap* and *SH* numbers are used in Fig. 1, and refer to haplotypes included in Désamoré & al. (2012) and specimens in Huttunen & al. (2008), respectively. The GenBank accession numbers for *SH* specimens correspond with *rpl*16 and *atp*B-*rbc*L.

Homalothecium aeneum (Mitt.) E.Lawton - SH308: EF531042, EF530983. SH347: EF531043, EF530985. SH348: EF531045, EF530984. SH349: EF531044, EF530986. Homalothecium aureum (Spruce) H.Rob. - SH142: EF531081, EF530969. SH302: EF531082, EF530970. SH303: EF531083, EF530967. SH306: EF531080, EF530968. Homalothecium californicum Hedenäs, Huttunen, Shevock & D.H.Norris - SH345: EF531037, EF531016. Homalothecium lutescens(Hedw.) H.Rob. - SH20: EF531053, EF530971. SH304: EF531052, EF530974. SH318: EF531051, EF530972. SH321: EF531050, EF530973. SH380: EF531054, EF530975. Homalothecium lutescens var. fallax (H.Philib.) Hedenäs & L.Söderstr. - SH305: EF531056, EF530977. SH392: EF531057, EF531003. SH397: EF531055, EF530976. Homalothecium fulgescens (Müll.Hal.) Lawt. - SH140: EF531038, EF530978. SH320: EF531040, EF530980. SH363: EF531039, EF530979. SH365: EF531041, EF530982. Homalothecium mandonii (Mitt.) Geh. - Hap11, Hap12, Hap13: Désamoré & al. (2012). SH322: EF531063, EF531009. SH357: EF531064, EF531010. SH358: EF531065, EF531011. Homalothecium mediterraneum Hedenäs - Hap14, Hap15, Hap16, Hap21: Désamoré & al. (2012). Homalothecium megaptilum (Sullivant) Schofield - SH344: EF531036, EF531014. SH346: EF531035, EF531015. Homalothecium nevadense (Lesg.) Renauld & Cardot - SH307: EF531046, EF530987. SH309: EF531048, EF530989. SH355: EF531047, EF530988. SH356: EF531049, EF530990. Homalothecium nuttallii (Wilson) A.Jaeger - SH68: EF531077, EF530993. SH362: EF531078, EF530991.

SH391: EF531079, EF530992. *Homalothecium philippeanum* (Spruce) Schimp. - *SH121*:
EF531069, EF530994. *SH310*: EF531074, EF531000. *SH315*: EF531071, EF530996. *SH316*:
EF531073, EF530999. *SH317*: EF531070, EF530995. *SH323*: EF531072, EF530997. *SH389*:
EF531068, EF531002. *SH390*: EF531076, EF530998. *SH395*: EF531075, EF531001. *Homalothecium sericeum* (Hedw.) Schimp. s.str. - *Hap1*, *Hap2*, *Hap3*, *Hap4*, *Hap5*, *Hap6*, *Hap7*, *Hap9*, *Hap10*, *Hap17*, *Hap18*, *Hap20*: Désamoré & al. (2012). *SH35*: EF531061,
EF531007. *SH319*: EF531066, EF531012. *SH324*: EF531067, EF531013. *SH359*: EF531060,
EF531006. *SH360*: EF531062, EF531008. *SH393*: EF531059, EF531005. *SH394*: EF531058,
EF531004. **OUTGROUP:** *Brachytheciastrum velutinum* (Hedw.) Huttunen& Ignatov – *SH78*: EF531033, EF530965.

Figure captions:

Figure 1. Strict consensus of 887 equally parsimonious trees resulting from the MP analysis of *rpl*16 and *atp*B-*rbc*L in the moss genus *Homalothecium*. Thick branches indicate Bootstrap support above 50.

Figure 2. Variation in leaf margin denticulation in the alar region of branch leaves in *Homalothecium sericeum* s.str. (Sweden. Södermanland, Utö, 15 May 2010, *L.Hedenäs*, S: B175290). Scale: 50 μm.

Figure 3. A-D: *Homalothecium mediterraneum* (Greece. Crete, Chania, *Frahm K-158*, Herb. J.-P.Frahm BONN); E-H: *H. mandonii* (Canary Islands. El Hierro, Riscos de Sabinosa, March 1906, *C.J.Pitard*, S: B185186). A, E: lower exostome seen from the outside – note the exostome borders, visible as semi-translucent marginal portions of each tooth; B, F: branch leaves; C, G: leaf margin in alar region; D, H: median leaf lamina cells. Scales: a: 50 μm (A, E); b: 1 mm (B, F); c: 50 μm (C, D, G, H).







