

Thymoquinone induces apoptosis through downregulation of c-FLIP and Bcl-2 in renal carcinoma Caki cells

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Abstract. Renal carcinoma is a common and frequently fatal carcinoma occurring worldwide and death rates due to this carcinoma are increasing with time. In the present study, we investigated the potential of thymoquinone a natural compound to induce apoptosis in renal carcinoma Caki cells. Thymoquinone efficiently enhanced the apoptotic population of Caki cells in a dose-dependent manner. Moreover, thymoquinone-mediated apoptosis caused downregulation of c-FLIP and Bcl-2, the master regulators of the anti-apoptotic mechanism. However, we did not find any changes in mRNA expression level of c-FLIP, therefore; this regulation of c-FLIP was a result of post-translation modification by thymoquinone. In contrast, expression of the Bcl-2 protein was observed at both transcriptional and translational level. However, we also observed that thymoquinone enhanced the level of intracellular reactive oxygen species (ROS) in Caki cells, which resulted in reduction of mitochondrial membrane potential (MMP) and cytochrome *c* release into cytoplasm. Our results postulate that thymoquinone induces apoptosis through downregulating c-FLIP and Bcl-2 which can be utilized as a chemotherapeutic agent to treat renal carcinoma.

Introduction

There are various kidney-related malignancies, yet, renal cell carcinoma is the most frequently occurring tumor among these. These tumor types are not responsive to chemotherapy or radiation therapy (1,2), thus, it is extremely hard to treat

them and this is the motivation why different groups of researchers are attempting to acquaint some novel chemotherapeutic agent that can treat this disease. Although, there have been many efforts to treat this carcinoma with molecular or gene therapy, for example, tyrosine kinase inhibitors, mammalian target of rapamycin (mTOR) and vascular endothelial growth factor (VEGF) inhibitors (3), the pharmacological agents that can play an important role in inhibition of these genes are the most important. However, the mechanism underlying the initiation to the progression of most cancers is similar to each other for example, there are various master regulators of anti-apoptotic mechanism such as Bcl-2 and c-FLIP that limit the potential of a drug to initiate apoptosis mechanism within the cell (4,5).

In light of the fact that renal carcinoma is one of the most deadly types of cancers, some candidates have been introduced to stop and/or limit the progression of the metastasis of this carcinoma, for example, sunitinib and pazopanib, drugs that are reported to be the inhibitors of tyrosine kinase (6,7). Indeed, these agents were found to be able to stop the growth of cancer cells but extensive research on these agents revealed that they are associated with drug resistance and they have various side-effects as well (8,9). Therefore, there is a need to introduce potent candidates that are originated from a natural source and are not destructive to the normal physiological condition of cells or tissue.

Thymoquinone is a natural polyphenolic compound found abundantly in black cumin (*Nigella sativa* L.) seeds and is grouped as monoterpenes. Thymoquinone has recently been reported for its various medicinal properties, for example, it is effective to treat gastroenteritis and various other organ and cell-associated inflammation (10,11). Moreover, few recent studies explored its efficacy to kill cancer cells via induction of apoptosis mechanism, and some researchers demonstrated that thymoquinone has potential to reduce the volume of breast, colon and gastric cancer using *in vivo* model (12,13), however, very limited studies have been carried out to uncover its mechanism of action in renal carcinoma cell lines.

In the present study, we have evaluated the efficacy of thymoquinone against Caki cells, a renal carcinoma cell line and uncovered its molecular mechanism of action against this cell line.

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Materials and methods

Cells and materials. Caki, A498 and ACHN cells were obtained from the American Type Culture Collection (ATCC; Manassas, VA, USA). The culture medium used throughout these experiments was Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS), 20 mM HEPES buffer and 100 $\mu\text{g}/\text{ml}$ gentamycin. The mouse kidney cells, TMCK-1, were a gift from Dr T.J. Lee (Yeungnam University, Korea). z-VAD-fmk was purchased from R&D Systems. Thymoquinone was purchased from Sigma Chemical Co. (St. Louis, MO, USA). Anti-Bcl-2 (sc-783), anti-Bcl-xL (sc-634), anti-Mcl-1 (sc-819) and anti-cIAP2 (sc-7944) were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Anti-XIAP (610762) was purchased from BD Biosciences (Bedford, MA, USA). Anti-PARP (#9542) antibody was obtained from Cell Signaling Technology (Beverly, MA, USA). Anti-actin (A5441) antibody was obtained from Sigma (St. Louis, MO, USA). Other reagents were purchased from Sigma Chemical Co.

Flow cytometric analysis. For flow cytometry, the cells were resuspended in 100 μl of phosphate-buffered saline (PBS), and 200 μl of 95% ethanol was added while the cells were being vortexed. The cells were then incubated at 4°C for 1 h, washed with PBS, resuspended in 250 μl of 1.12% sodium citrate buffer (pH 8.4) with 12.5 μg of RNase and incubated for an additional 30 min at 37°C. The cellular DNA was then stained by adding 250 μl of a propidium iodide solution (50 $\mu\text{g}/\text{ml}$) to the cells for 30 min at room temperature (14). The stained cells were analyzed by fluorescent-activated cell sorting on a FACScan flow cytometer to determine the relative DNA content, which was based on the red fluorescence intensity.

Western blot analysis. For the western blot experiments, the cells were washed with cold PBS and lysed on ice in modified RIPA buffer (50 mM Tris-HCl pH 7.4, 1% NP-40, 0.25% Na-deoxycholate, 150 mM NaCl, 1 mM Na_3VO_4 and 1 mM NaF) containing protease inhibitors (100 μM phenylmethylsulfonyl fluoride, 10 $\mu\text{g}/\text{ml}$ leupeptin, 10 $\mu\text{g}/\text{ml}$ pepstatin and 2 mM EDTA). The lysates were centrifuged at 10,000 \times g for 10 min at 4°C and the supernatant fractions were collected. The proteins were separated by SDS-PAGE electrophoresis and transferred to Immobilon-P membranes. The specific proteins were detected using an enhanced chemiluminescence (ECL) western blotting kit according to the manufacturer's instructions.

DNA fragmentation assay. DNA fragmentation was performed using the Cell Death Detection ELISAPLUS kit (Boehringer Mannheim, Indianapolis, IN, USA). Briefly, cells were centrifuged for 10 min at 200 \times g, the supernatant was removed, and pellet was lysed for 30 min. After centrifuging the plate again at 200 \times g for 10 min, and the supernatant that contained the cytoplasmic histone-associated DNA fragments was collected and incubated with an immobilized anti-histone antibody. The reaction products were incubated with a peroxidase substrate for 5 min and measured by spectrophotometry at 405 and 490 nm (reference wavelength) with a microplate reader. The signals in the wells containing the substrate alone were subtracted as background.

Asp-Glu-Val-Asp-ase (DEVDase) activity assay. To evaluate DEVDase activity, cell lysates were prepared after their respective treatments with thymoquinone. Assays were performed in 96-well microtiter plates by incubating 20 μg of cell lysates in 100 μl of reaction buffer (1% NP-40, 20 mM Tris-HCl, pH 7.5, 137 mM NaCl, 10% glycerol) containing a caspase substrate [Asp-Glu-Val-Asp-chromophore-p-nitroanilide (DVAD-pNA)] at 5 mM. Lysates were incubated at 37°C for 2 h. Thereafter, the absorbance at 405 nm was measured with a spectrophotometer.

RNA isolation, reverse transcription polymerase chain reaction (RT-PCR). Total cellular RNA was extracted from cells using TRIzol reagent (Life Technologies, Gaithersburg, MD, USA). Complementary DNA was synthesized from 2 μg of total RNA using M-MLV reverse transcriptase (Promega, Madison, WI, USA). The cDNA for c-FLIP, Bcl-2 and actin were amplified by a PCR using specific primers: c-FLIP (forward) 5'-CGG ACT ATA GAG TGC TGA TGG-3' and (reverse) 5'-GAT TAT CAG GCA GAT TCC TAG-3'; Bcl-2 (forward) 5'-GTC CTC AGC CCT CGC TCT-3' and (reverse) 5'-CAC CTA ATT GGG CTC CAT CT-3'; actin (forward) 5'-GGC ATC GTC ACC AAC TGG GAC-3' and (reverse) 5'-CGA TTT CCC GCT CGG CCG TGG-3'. PCR products were analyzed by agarose gel electrophoresis and visualized using ethidium bromide staining.

Transfection and promoter activity assay. Transient transfection was performed in 6-well plates. One day before the transfection, Caki cells were plated at ~60-80% confluence. The Bcl-2/-3254 promoter- or NF- κ B-luciferase plasmid was transfected into the cells using Lipofectamine™ 2000 (Invitrogen, Carlsbad, CA, USA). To assess the promoter-driven expression of the luciferase gene, the cells were collected and disrupted by sonication in lysis buffer (25 mM Tris-phosphate pH 7.8, 2 mM EDTA, 1% Triton X-100 and 10% glycerol), and aliquots of the supernatants were used to analyze the luciferase activity according to the manufacturer's instructions (Promega).

Measurement of reactive oxygen species (ROS). Intracellular accumulation of ROS was determined using the fluorescent probes 2',7'-dichlorodihydrofluorescein diacetate (H2DCFDA). H2DCFDA is commonly used to measure ROS generation. Caki cells were pretreated with NAC for 30 min, and then added with thymoquinone. Cells were stained with the fluorescent dye H2DCFDA for an additional 10 min. Then, cells were observed using a fluorescence microscope (Axiovert 200M; Carl Zeiss, Oberkochen, Germany).

Determination for the mitochondrial membrane potential (MMP) by Rhodamine 123 and DiOC₆. Rhodamine 123 and DiOC₆ (Molecular Probes, Carlsbad, CA, USA) uptake by mitochondria is directly proportional to its membrane potential. Caki cells 4 h after treatment were incubated with Rhodamine 123 (5 μM) and DiOC₆ for 30 min in the dark at 37°C (15). The cells were harvested and suspended in PBS. The MMP was subsequently analyzed using a flow cytometer (Becton-Dickinson, Franklin Lakes, NJ, USA).

Analysis of cytochrome c release. Caki cells (1.2×10^6 cells/ml) were harvested, washed once with ice-cold PBS and gently

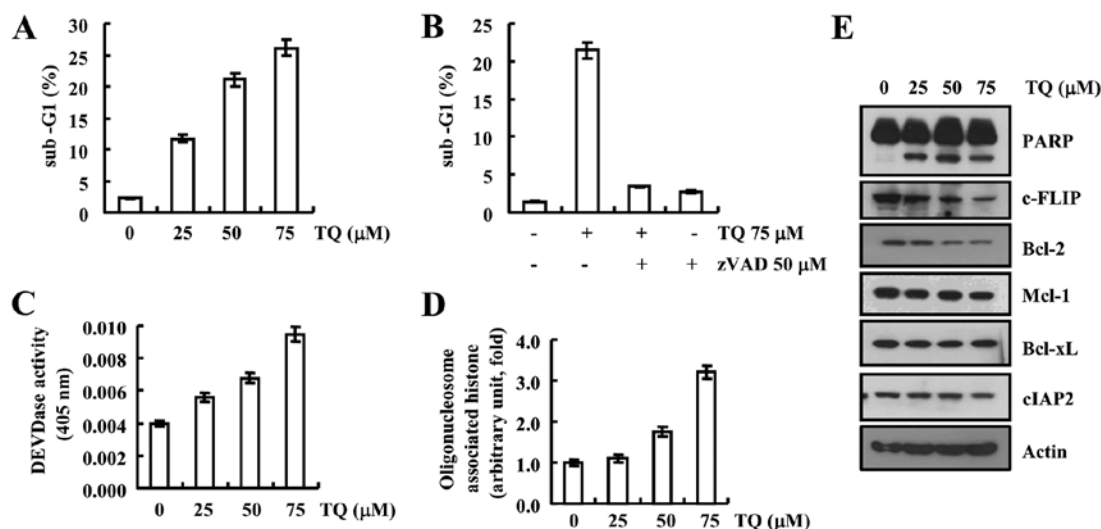


Figure 1. Thymoquinone induces apoptosis in Caki cells. (A) Cells were treated with the indicated concentrations of thymoquinone for 24 h. Apoptosis was analyzed as a sub-G1 fraction by FACS. (B) Caki cells were pretreated with 25 μ M z-VAD-fmk for 30 min, and then 75 μ M thymoquinone was added for 24 h. Apoptosis was analyzed as a sub-G1 fraction by FACS. (C) Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. Caspase activities were determined with colorimetric assays using caspase-3 DEVDase assay kits. (D) DNA fragmentation in Caki cells was determined by the DNA fragmentation detection kit. (E) Cells were treated with the indicated concentrations of thymoquinone for 24 h. Equal amounts of cell lysates (60 μ g) were subjected to electrophoresis and analyzed by western blotting for PARP, c-FLIP, Bcl-2, Mcl-1, Bcl-xL, cIAP2 and actin as a control for protein loading.

lysed for 2 min in 80 μ l ice-cold lysis buffer [250 mM sucrose, 1 mM EDTA, 20 mM Tris-HCl (pH 7.2), 1 mM DTT, 10 mM KCl, 1.5 mM MgCl₂, 5 μ g/ml pepstatin A, 10 μ g/ml leupeptin and 2 μ g/ml aprotinin]. Lysates were centrifuged at 12,000 \times g at 4°C for 10 min to obtain the supernatants (cytosolic extracts free of mitochondria) and the pellets (fraction that contains mitochondria). The resulting cytosolic fractions were used for western blot analysis with an anti-cytochrome *c* antibody.

Statistical analysis. The data were analyzed using a one-way ANOVA followed by post hoc comparisons (Student-Newman-Keuls) using the Statistical Package for Social Sciences version 22.0 (SPSS, Inc., Chicago, IL, USA).

Results

Thymoquinone efficiently induces apoptosis in Caki cells. We first determined the cytotoxic effect of thymoquinone against Caki cells and found that after treatment of thymoquinone (25, 50 and 75 μ M) for 24 h apoptotic population (sub-G1) was increased in a dose-dependent manner (Fig. 1A). Furthermore, we used caspase inhibitor zVAD to examine the association of caspases in apoptosis induced by thymoquinone, and we found that after inhibition of caspase apoptotic population was decreased even at a higher concentration of thymoquinone which was further validated by DEVDase activity suggesting that thymoquinone induced caspase-dependent apoptosis (Fig. 1B and C). Moreover, the level of cytoplasmic histone was observed to be increased that is an outcome of DNA fragmentation thus thymoquinone-induced DNA fragmentation which resulted in increased cytoplasmic histone (Fig. 1D). We further determined the expression pattern of apoptotic and anti-apoptotic proteins to understand the molecular mechanism underlying the thymoquinone toxicity, as depicted in Fig. 1E, thymoquinone enhanced the cleavage of PARP protein in a

dose-dependent manner. Expression of anti-apoptotic proteins such as c-FLIP and Bcl-2 was downregulated, but expression of Mcl-1, Bcl-xL and cIAP2 did not change in western blot examination (Fig. 1E).

Thymoquinone downregulates c-FLIP expression to induce apoptosis in Caki cells. We found that thymoquinone downregulated the expression of c-FLIP in our previous experiment, next we decided to validate our data. We then examined the expression pattern of c-FLIP at the transcriptional level, results of RT-PCR analysis showed that there was no any change in the expression of c-FLIP at the transcriptional level (Fig. 2A). Therefore, we investigated whether thymoquinone modulates the protein stability of c-FLIP in Caki cells. Cells were treated with cycloheximide (CHX), an inhibitor of *de novo* protein synthesis, in the presence or absence of thymoquinone. CHX gradually decreased c-FLIP, but co-treatment with CHX and thymoquinone reduced more c-FLIP protein expression (Fig. 2B). To investigate the importance of downregulation of c-FLIP expression on thymoquinone-induced apoptosis, c-FLIP protein was overexpressed in Caki cells. Overexpression of c-FLIP markedly inhibited thymoquinone-induced increase of sub-G1 cell population (Fig. 2C) and PARP cleavage (Fig. 2D). These results indicate that downregulation of c-FLIP may be involved in thymoquinone-mediated apoptosis.

Expression of Bcl-2 is downregulated after treatment with thymoquinone through hindering the NF- κ B cascade. We evaluated the effect of thymoquinone on the expression pattern of Bcl-2, another major regulator of the anti-apoptotic mechanism. Results are depicted in Fig. 3, where we first determined the expression of Bcl-2 at the transcriptional level and found that Bcl-2 mRNA expression was reduced in a dose-dependent manner (Fig. 3A). Furthermore, we validated

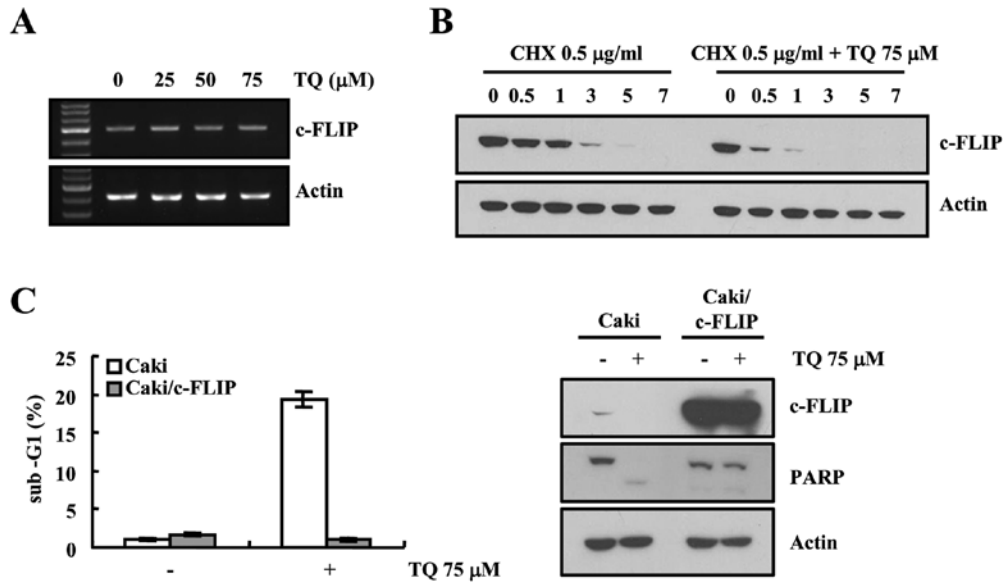


Figure 2. Thymoquinone downregulates c-FLIP protein at the post-transcriptional level in Caki cells. (A) Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. c-FLIP and actin mRNA expression was determined using RT-PCR. (B) Caki cells were treated with cyclohexamide (CHX) 20 $\mu\text{g}/\text{ml}$ alone or CHX plus 75 μM thymoquinone for indicated time periods. Equal amounts of cell lysates (60 μg) were subjected to electrophoresis and analyzed by western blotting for c-FLIP and actin as a control for protein loading. (C) Vector (Caki/Vec) and c-FLIP overexpressed cells (Caki/cFLIP) were treated with 75 μM thymoquinone for 24 h. Apoptosis was analyzed as a sub-G1 fraction by FACS. Equal amounts of cell lysates (60 μg) were subjected to electrophoresis and analyzed by western blotting for c-FLIP and actin as a control for protein loading. The data represent three independent experiments.

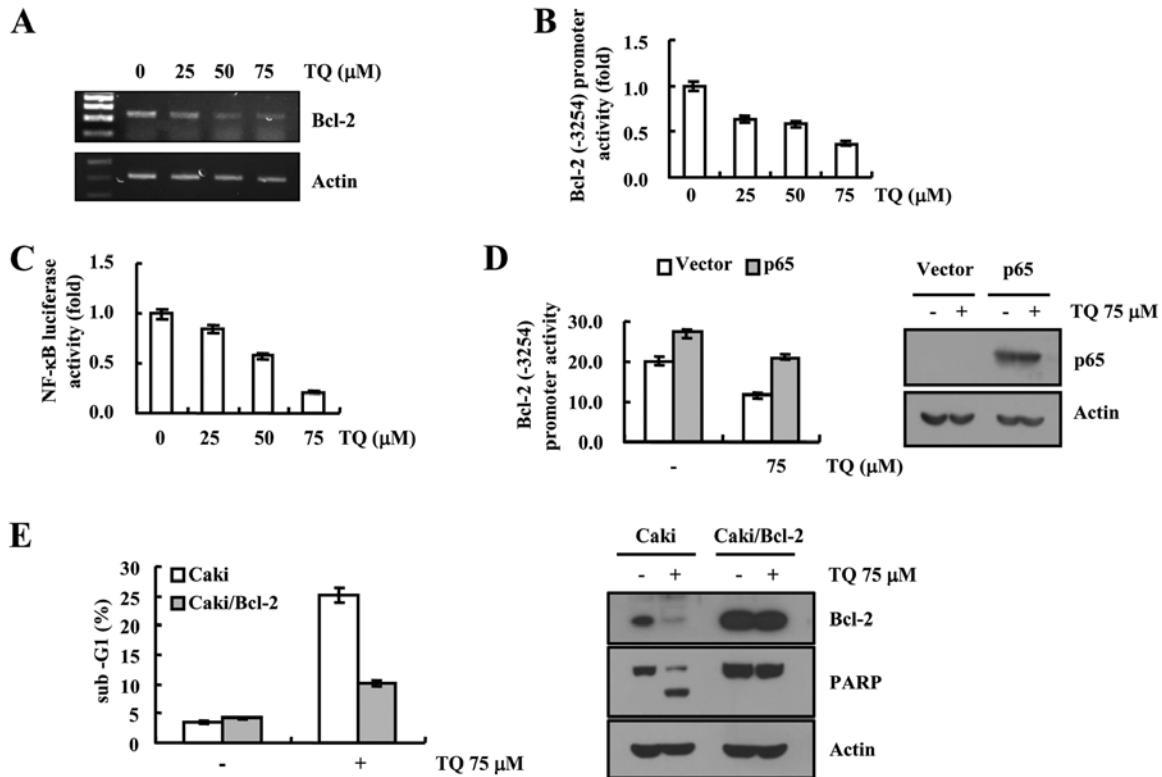


Figure 3. Thymoquinone downregulates Bcl-2 expression at the transcriptional level in Caki cells. (A) Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. Bcl-2 and actin mRNA expression were determined using RT-PCR. (B) Caki cells were transiently transfected with a plasmid harboring the luciferase gene under the control of the Bcl-2/-3254 promoter. After transfection, the Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. After treatment, the cells were lysed, and the luciferase activity was analyzed. (C) Caki cells were transiently transfected with NF- κ B-luciferase construct. After transfection, the Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. After treatment, the cells were lysed and the luciferase activity was analyzed. (D) Caki cells were transiently co-transfected with NF- κ B subunit (p65) and Bcl-2-luciferase construct. After transfection, the Caki cells were treated with 75 μM thymoquinone for 24 h. After treatment, the cells were lysed, and the luciferase activity was analyzed. Equal amounts of cell lysates (60 μg) were subjected to electrophoresis and analyzed by western blotting for p65 and actin as a control for protein loading. (E) Vector (Caki/Vec) and Bcl-2 overexpressed cells (Caki/Bcl-2) were treated with 75 μM thymoquinone for 24 h. Apoptosis was analyzed as a sub-G1 fraction by FACS. Equal amounts of cell lysates (60 μg) were subjected to electrophoresis and analyzed by western blotting for PARP, Bcl-2 and actin as a control for protein loading. The values in (B, C, D and E) represent the mean \pm SD from three independent samples.

our data with Bcl-2 promoter assay and found that thymoquinone reduced the Bcl-2 promoter activity in a dose-dependent manner (Fig. 3B). Moreover, we determined the effect of thymoquinone on NF- κ B expression as it is associated with expression of Bcl-2 protein. NF- κ B luciferase activity was reduced after treatment of thymoquinone. In addition, overexpression of p65, a subunit of NF- κ B heightened the Bcl-2 promoter activity in case of thymoquinone treated cells, which was further confirmed by immunoblot analysis (Fig. 3D). We examined the efficacy of thymoquinone to induce apoptosis in Bcl-2 overexpressed Caki cells and observed that overexpression of Bcl-2 reversed the apoptotic potential of thymoquinone as evidenced by flow cytometric analysis (Fig. 3E). Moreover, thymoquinone-induced cleavage of PARP protein was blocked in Bcl-2 overexpressed cells (Fig. 3E), suggesting the downregulation of Bcl-2 plays a critical role in thymoquinone-induced apoptosis.

Thymoquinone-induced ROS generation resulting in loss of MMP in Caki cells. The induction of ROS production plays an important role in apoptosis. Therefore, we carried out H2DCFDA staining to determine the level of intracellular ROS. Thymoquinone (75 μ M) potentially induced the generation of intracellular ROS within 10 min, and then slightly reduced after 30 min of treatment (Fig. 4A and B). Furthermore, we determined the effect of thymoquinone-induced ROS on cell death with or without using the ROS scavenger *N*-acetylcysteine (NAC). Thymoquinone-induced ROS generation caused apoptosis in Caki cells which was reduced after using the ROS scavenger (Fig. 4C). We observed increased intensity of MitoSOX Red dye which detects mitochondrial ROS production (Fig. 4D). Furthermore, we determined the MMP, as the exaggerated production of ROS leads to the mitochondrial damage. Rhodamine 123 and DiOC₆ fluorometry data revealed the loss of MMP in time-dependent manner at 75 μ M thymoquinone treatment (Fig. 4E and F). Treatment with thymoquinone caused cytochrome *c* release into cytoplasm (Fig. 4G). These results suggest that thymoquinone reduces the MMP levels and induces cytochrome *c* release.

Thymoquinone exhibits cytotoxic effect against various renal carcinoma cells, but not in normal cells. Although, our results demonstrated that thymoquinone potentially induced apoptosis in Caki cells, we further examined the efficacy of thymoquinone against other renal carcinoma cell line for example ACHN and A498 and found that thymoquinone has efficacy to induce apoptosis in all the renal carcinoma cells used in the present study (Fig. 5A and B). However, there was the insignificant cytotoxic effect of thymoquinone on normal TMCK-1 cells neither on both morphology and cell population analysis, even at a higher concentration of 75 μ M which induced the apoptosis in Caki cells (Fig. 5C and D) suggesting the safe use of thymoquinone.

Discussion

There is an intricate balance between pro- and anti-apoptotic proteins in normal cells; however, in cancer cells there is a dysregulation of this balance and that may be due to overex-

pression of anti-apoptotic proteins which demise the process of apoptosis (16), for instance, overexpression of Bcl-2, an anti-apoptotic protein, protects the apoptosis of prostate cancer cells (17). Consequently, in the present study we evaluated for example the apoptotic PARP as well as the anti-apoptotic (c-FLIP, Bcl-2, Mcl-1, Bcl-xL and cIAP2) markers through immunoblot analysis and observed that thymoquinone treatment induced the cleavage of PARP and downregulation of c-FLIP and Bcl-2 proteins.

c-FLIP is a master regulator of anti-apoptotic mechanism and is found to be abundantly expressed in a variety of cancer cells (18). Overexpression of c-FLIP interferes with caspase-8 protein and is able to block the cleavage of caspase-8 which ultimately terminates the apoptotic machinery (4). Thymoquinone downregulated the expression of c-FLIP at translation level but surprisingly there was no effect of thymoquinone on transcriptional regulation of c-FLIP, however, it is well documented in literature that ROS contribute to post-translation modification of c-FLIP, through upregulation of proteasomal activity (19,20). Notably, we found that thymoquinone potentially induced production of intracellular ROS in Caki cells, thus, we hypothesize that this property of thymoquinone may be responsible for the downregulation of c-FLIP.

Another key regulator protein of anti-apoptotic machinery is Bcl-2 (21-23), and in the present study, we found that thymoquinone potentially suppressed the expression of this protein at both transcriptional and translation level resulting in the initiation of the apoptotic cascade. However, recently numerous studies have explored the association of NF- κ B with Bcl-2, for example, expression of the NF- κ B targets BCL-2 expression on the transcriptional level (24,25). We, therefore, examined the possible role of thymoquinone in the regulation of NF- κ B activity and notably, a highly decreased NF- κ B activity was observed in luciferase reporter assay in a concentration-dependent manner suggesting NF- κ B inhibitory efficacy of thymoquinone. In contrast, when we overexpressed p65, a subunit of NF- κ B it caused increased promoter activity of Bcl-2 even at a higher concentration of thymoquinone which clearly indicates that downregulation of Bcl-2 by thymoquinone was associated with NF- κ B activity. Heckman *et al* (25) reported similar results in lymphoma cells where overexpression of NF- κ B was responsible for the Bcl-2 overexpression, taken together, we can conclude that hindrance in NF- κ B activity may be an effective tool to target the Bcl-2 expression which was accomplished by thymoquinone in our case.

We further evaluated the effect of thymoquinone on MMP, as we found that thymoquinone potentially downregulated Bcl-2 expression which in turn is associated with the loss of MMP leading to mitochondrial death, a potent and common anticancer target for many pharmacological agents (26-28). In addition, a well-known inducer of mitochondrial stress is generation of exaggerated ROS inside the cell which eventually kill the mitochondria (29-31). Interestingly, both the phenomenon associated with mitochondrial stress (Bcl-2 downregulation and ROS generation) were found to be followed by thymoquinone in the present study, thus, we examined the mitochondrial membrane potential (MMP) in the presence of thymoquinone and as expected, the loss of MMP

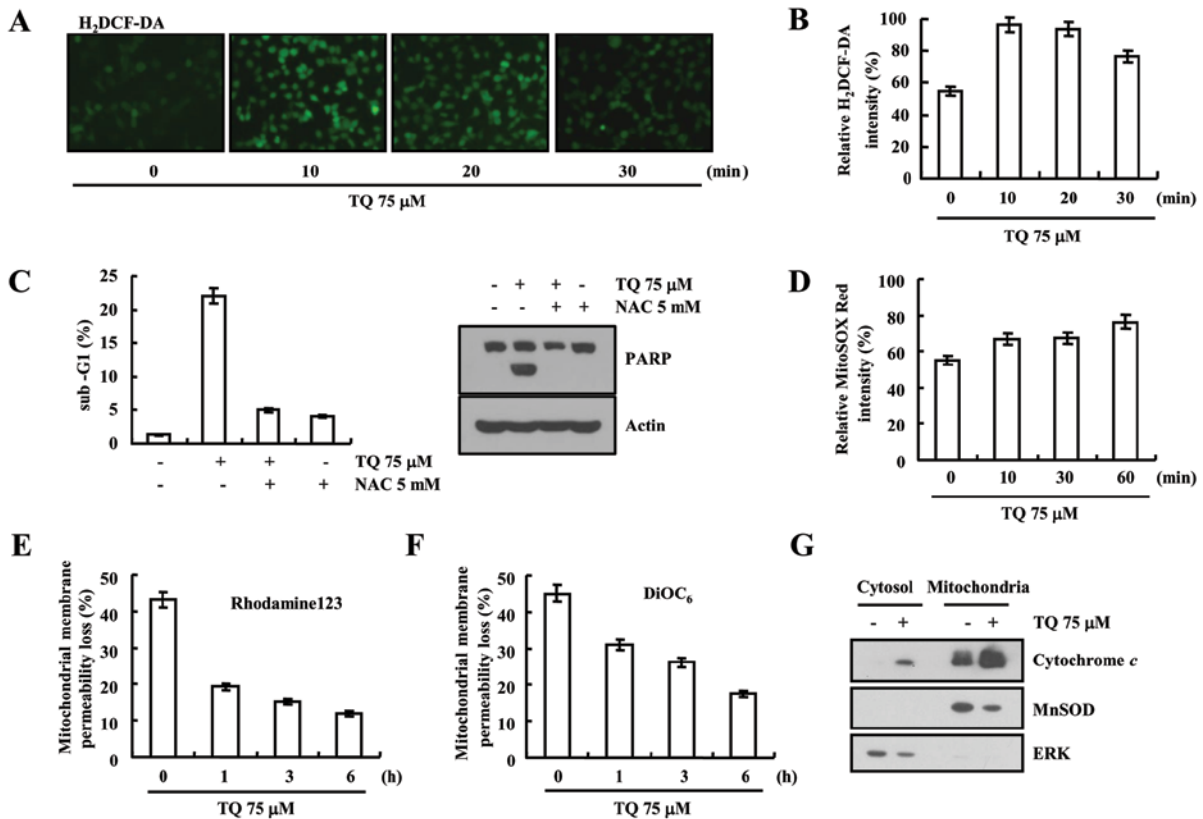


Figure 4. Thymoquinone induces ROS and release of cytochrome *c* into the cytoplasm. (A and B) Caki cells were stimulated with 75 μM thymoquinone for the indicated time points and loaded with a H2DCF-DA fluorescent dye. H2DCF-DA fluorescence intensity was detected by (A) fluorescence microscopy and (B) FACS. (C) Caki cells were pretreated with NAC (5 mM) for 30 min, and then stimulated with 75 μM thymoquinone for 24 h. Apoptosis was analyzed as the sub-G1 population by FACS analysis. Equal amounts of cell lysates (60 μg) were subjected to electrophoresis and analyzed by western blotting for PARP and actin as a control for protein loading. The data represent three independent experiments. (D) Caki cells were stimulated with 75 μM thymoquinone for the indicated time points and loaded with a MitoSOX Red dye. Fluorescence intensity was detected by FACS. (E and F) Caki cells were treated with 75 μM thymoquinone for 4 h. The mitochondrial membrane potential was measured as described under 'Materials and methods'. The data represent three independent experiments. (G) Caki cells were treated with 75 μM thymoquinone for 4 h. Cytosol fraction and mitochondria fraction (Mito). Extracts were prepared as described under 'Materials and methods'. Equal amounts of cell lysates (40 μg) were subjected to electrophoresis and analyzed by western blotting for cytochrome *c* and ERK as a control for protein loading (right panel). To show there is no mitochondrial contamination in the cytosol fraction, we carried out western blot analysis using antibody against MnSOD that was expressed in mitochondria. Mitochondrial fraction derived from non-treated cells was used as a positive control.

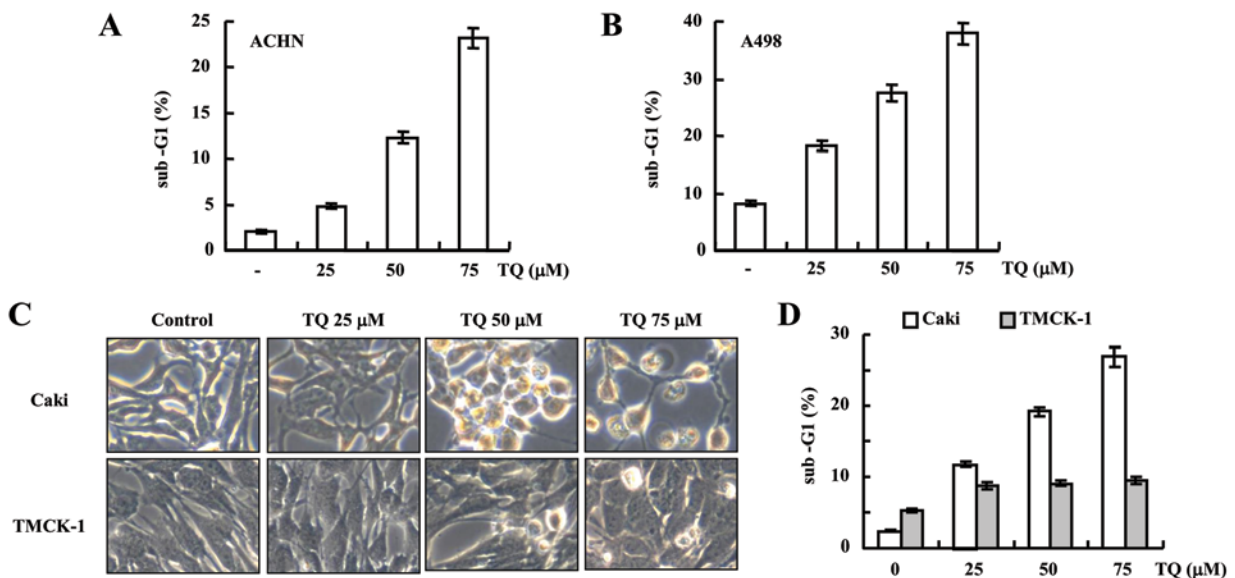


Figure 5. Thymoquinone induces apoptosis in other renal carcinoma cells, but not in normal cells. Renal carcinoma (A) ACHN and (B) A498 cells were treated with the indicated concentrations of thymoquinone for 24 h. Apoptosis was analyzed as a sub-G1 fraction by FACS. (C) TMCK-1 and Caki cells were treated with the indicated concentrations of thymoquinone for 24 h. The cell morphologies were determined by interference light microscopy. (D) Apoptosis was analyzed as the sub-G1 fraction by FACS analysis. Values in (A, B and D) are expressed as mean \pm SD of three independent experiments.

was the outcome of the experiment with an intense release of cytochrome *c* into cytosol, a defining feature of mitochondrial damage (31). However, undoubtedly, a potent candidate for anticancer therapy should be one which possess toxic effect against cancer cells but not to normal cells and we in the case of thymoquinone found similar results showing insignificant toxic effect on normal cells (TMCK-1 cells).

On account of the results obtained in the present study, we conclude that thymoquinone has potential to induce apoptosis in renal cell carcinoma with a major mechanism of c-FLIP and Bcl-2 downregulation. In addition, it is able to induce mitochondrial dysfunctioning through the generation of intracellular ROS in Caki cells which eventually leads to cell death. Moreover, it originates from natural plants and its insignificant toxicity to the normal cells warrant its safe use as a drug against renal cell carcinoma. However, further experiments on an animal model is needed to evaluate its efficacy and precise mechanism *in vivo*.

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