

Thyroid hormone synthesis and anti-thyroid drugs: A bioinorganic chemistry approach

GOURIPRASANNA ROY and G MUGESH*

Department of Inorganic & Physical Chemistry, Indian Institute of Science, Bangalore 560 012
e-mail: muges@ipc.iisc.ernet.in

Abstract. Hydrogen peroxide, generated by thyroid oxidase enzymes, is a crucial substrate for the thyroid peroxidase (TPO)-catalysed biosynthesis of thyroid hormones, thyroxine (T4) and triiodothyronine (T3) in the thyroid gland. It is believed that the H_2O_2 generation is a limiting step in thyroid hormone synthesis. Therefore, the control of hydrogen peroxide concentration is one of the possible mechanisms for the inhibition of thyroid hormone biosynthesis. The inhibition of thyroid hormone synthesis is required for the treatment of hyperthyroidism and this can be achieved by one or more anti-thyroid drugs. The most widely used anti-thyroid drug methimazole (MMI) inhibits the production of thyroid hormones by irreversibly inactivating the enzyme TPO. Our studies show that the replacement of sulphur in MMI by selenium leads to a selone, which exists predominantly in its zwitterionic form. In contrast to the sulphur drug, the selenium analogue (MSeI) reversibly inhibits the peroxidase-catalysed oxidation and iodination reactions. Theoretical studies on MSeI reveal that the selenium atom in this compound carries a large negative charge. The carbon–selenium bond length in MSeI is found to be close to single-bond length. As the selenium atom exhibits a large nucleophilic character, the selenium analogue of MMI may scavenge the hydrogen peroxide present in the thyroid cells, which may lead to a reversible inhibition of thyroid hormone biosynthesis.

Keywords. Antioxidants; anti-thyroid drugs; iodination; selenium; thyroxine.

1. Introduction

Thyroxine (**T4**), the main secretory hormone of the thyroid gland, is produced on thyroglobulin by thyroid peroxidase (TPO)/hydrogen peroxide/iodide system. The synthesis of **T4** by TPO involves two independent steps: iodination of tyrosine and phenolic coupling of the resulting iodotyrosine residues (figure 1).¹ The prohormone **T4** is then converted to its biologically active form **T3** by iodothyronine deiodinase (ID-I), which is present in highest amounts in liver, kidney, thyroid and pituitary.² Although the deiodination reactions are essential for the function of the thyroid gland, the activation of thyroid stimulating hormone (TSH) receptor by auto-antibodies leads to an overproduction of thyroid hormones. As these antibodies are not under pituitary feedback control system, there is no negative influence on the thyroid activity and, therefore, the uncontrolled production of thyroid hormones leads to a condition called “hyperthyroidism” (figure 2). Under these conditions, the overproduction of **T4** and **T3** can be controlled by specific inhibitors, which either block

the thyroid hormone biosynthesis or reduce the conversion of **T4** to **T3**. A unique class of such inhibitors are the thiourea drugs, methimazole (**1**, MMI), 6-*n*-propyl-2-thiouracil (**3**, PTU), and 6-methyl-2-thiouracil (**5**, MTU) (figure 2).

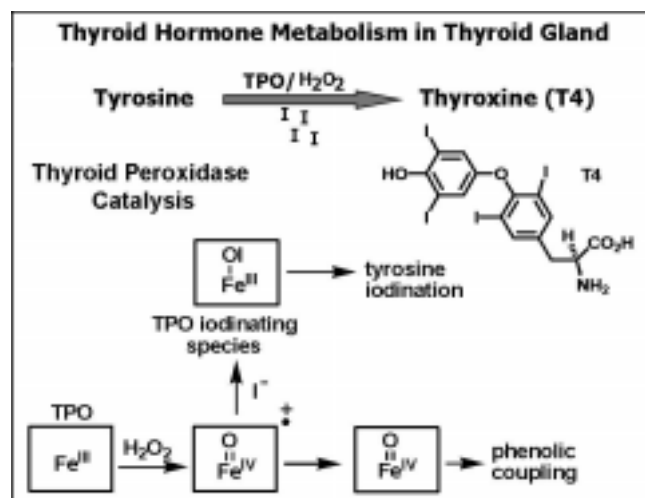


Figure 1. Synthesis of thyroid hormones by heme-containing thyroid peroxidase (TPO).

*For correspondence

Although these compounds are the most commonly employed drugs in the treatment of hyperthyroidism, the detailed mechanism of their action is still not clear. According to the initially proposed mechanism, these drugs may divert oxidized iodides away from thyroglobulin by forming stable electron donor–acceptor complexes with diiodine, which can effectively reduce thyroid hormone biosynthesis.³ It has also been proposed that these drugs may block the thyroid hormone synthesis by coordinating to the metal centre of thyroid peroxidase (TPO).⁴ After the discovery that ID-I is responsible for the activation of thyroxine, it has been reported that **PTU**, but not **MMI**, reacts with the selenenyl iodide intermediate (E-SeI) of ID-I to form a selenenyl sulphide as a dead end product, thereby blocking the conversion of **T4** to **T3** during the monodeiodination reaction.² The mechanism of anti-thyroid activity is further complicated by the fact that gold-containing drugs such as gold thioglucose (GTG) inhibit the deiodinase activity by reacting with the selenol group of the native enzyme.²

Recently, the selenium analogues **2** (**MSeI**), **4** (**PSeU**) and **6** (**MSeU**) attracted considerable attention.^{5,6} However, the data derived from the inhibition of TPO by selenium compounds show that these compounds may inhibit the TPO activity by a different mechanism. Therefore, further studies are required to understand the mechanism by which the selenium compounds exert their inhibitory action. Our initial attempts to isolate **2** were unsuccessful and the final stable compound in the synthesis was characterized to be the diselenide (**8**). In view of the current interest in anti-thyroid drugs and their mechanism, we

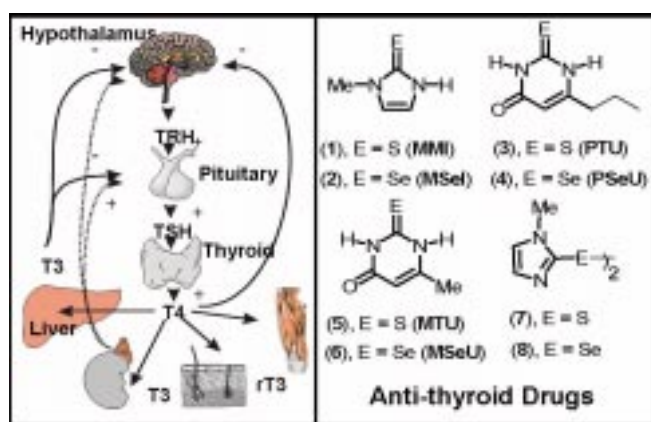


Figure 2. Pituitary feedback control system and anti-thyroid drugs.

extended our approach to the synthesis and biological activities of a number of sulphur and selenium derivatives bearing the methimazole pharmacophore.⁶ In this paper, we discuss our experimental evidences that replacement of the sulphur atom in methimazole by selenium leads to a completely different mechanism. In addition, we describe the effect of a range of sulphur and selenium compounds bearing the methimazole moiety on peroxidase-catalysed oxidation reactions. We also describe the effect of substituents attached to the imidazole moiety on the selenol-selone tautomerism by theoretical calculations.

2. Results and discussion

The tautomeric behaviour of **MMI** has been subjected to many investigations,⁷ which show that **MMI** exists almost exclusively as the thione tautomer (**1a**). Recent studies have shown that the thiourea-based drugs **PTU** and **MTU** also exist as thione tautomers.⁸ The stability of the thione tautomers may prevent these compounds from being oxidized to their corresponding disulphides, which may account for their high anti-thyroidal activity. Laurence *et al* have shown that the thione tautomer of **MMI** is responsible for its complexation with diiodine and the iodine complex of the thione tautomer **1a** is favoured by 13.2 kJ mol^{-1} compared to that of the thiol tautomer **1b**.^{9a} Therefore, the facile oxidation of **2** to the corresponding diselenide (**8**) requires the compound to be in its zwitterionic form (**2c**) and not in the true selone form (**2a**). Although compound **2** can exist in both selenol and selone forms in solution, the ⁷⁷Se NMR spectrum recorded immediately after the workup of the reaction showed a signal at -5 ppm , which can be ascribed to the zwitterionic (**2c**) tautomer (figure 3). In the presence of air, the zwitterion slowly oxidizes to the corresponding diselenide and this process continues until all the zwitterion is converted to diselenide **8**. In

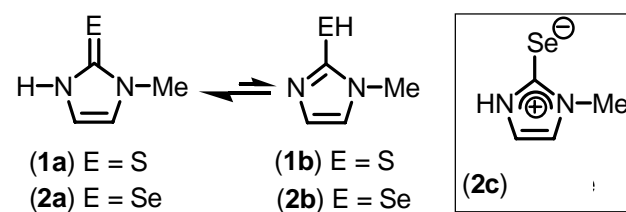


Figure 3. Proposed tautomeric structures of **MMI** and **MSeI**.

contrast, the sulphur analogue, which exists predominantly in its thione tautomer form (**1a**), was found to be very stable and could not be converted to the corresponding disulfide (**7**) even by using oxidizing agents such as O₂, H₂O₂ etc. It should be mentioned that MMI is readily oxidized by the TPO system to form the disulfide. Although the oxidation of this compound by iodine has been postulated to be a possible mechanism,¹⁰ the chemical way through which MMI is transformed into disulfide **6** *in vivo* is unknown.

The theoretical investigations on selones are highly limited to the compounds having simple substituents, mainly due to the requirement of large basis sets for the calculations.¹¹ The relatively larger size and greater polarizability of selenium as compared with sulphur have led to the assumption that the compounds with selone moiety are less stable than their sulphur analogues. Because the inhibition of TPO by antithyroid drugs depends upon the redox state of sulphur or selenium, we performed detailed quantum chemical calculations on **1** and **2** in gas phase. These studies show that the formation of the diselenide (**8**) from **2c** is energetically more favoured than the formation of the disulfide (**7**) from the corresponding thiol (**1b**). Interestingly, the conversion of thiol to thione is more favoured than the conversion of selenol to the corresponding selone. This can be rationalized by comparing the relative position of hydrogen on sulphur or selenium with respect to N1 in their most stable conformations. In the selenol (**2b**, figure 4), the H atom is located away from N1 leading to an increase in the energy barrier for the selenol–selone conversion. In contrast, the H atom is located in the close proximity of N1 in the thiol (**1b**, figure 2), which may favour the thiol–thione conversion. As the thione form is calculated to be much more stable than the corresponding thiol, it is quite unlikely that the thiol form contributes to the anti-thyroid activity of MMI. However, when the calculations were performed by including solvent effects, compound **2** was found to exist only in the zwitterionic form. Although the selone (**2a**) was also calculated to be more stable than the selenol (**2b**), the facile oxidation of the zwitterion to the corresponding diselenide disfavors the existence of selenol tautomer.

Although compound **2** readily oxidizes to diselenide **8**, the corresponding zwitterion (**2c**) can be conveniently obtained by reducing the diselenide by NaBH₄ or glutathione (GSH). The reaction of **8** with

NaBH₄ followed by aqueous workup afforded the zwitterion as white solid, which was found to be stable under inert atmosphere and could be employed for *in vitro* biological assays without any noticeable oxidation. The treatment of diselenide **8** with 2 equiv. of GSH has also produced the zwitterion in nearly quantitative yield. Interestingly, the formation of a true selone (**2a**) was not observed in any of these processes, supporting the theoretical calculations that the selone exists as the zwitterion (**2c**) due to the nitrogen atoms present in the imidazole ring. The ¹H and ¹³C NMR data and the large upfield shift in the ⁷⁷Se NMR chemical shift for the zwitterion (–5 ppm), supports this assumption. In agreement with the theoretical data, the ⁷⁷Se NMR experiments show that the selone is more dissociated in water (–53 ppm) than in organic solvent (–5 ppm) as evidenced by a large upfield shift when changing the solvent from CDCl₃ to D₂O. The facile reduction of **8** by GSH suggests that this compound may exist in its zwitterionic form under *in vivo* conditions, because GSH is present in the thyroid gland in high concentrations. It has been shown that GSH is an important antioxidant in thyroid gland and the peroxide scavenging activity of MMI is considerably increased by GSH.¹²

The enzyme inhibition experiments were carried out with Fe-containing lactoperoxidase (LPO) since it is readily available in purified form. Furthermore, LPO has been shown to behave very similarly to TPO with respect to iodination of thyroglobulin, the natural substrate, and other iodide acceptors.¹³ Edelhock *et al* have reported the inactivation of LPO by thiourea-based drugs using LPO-N-acetyltyrosylamide assay.¹⁴ We have employed 2,2'-azobis(3-ethylbenzothiazoline-6-sulphonic acid) (ABTS) and H₂O₂ as substrates to determine the half-maximal inhibitory concentration (IC₅₀) of test compounds. The IC₅₀ values for the inhibition of LPO-catalysed oxidation of ABTS by **1–3** and **5** are summarized in table 2.

To obtain reliable IC₅₀ values for compound **2** and to make a direct comparison with the sulphur analogue, it is important to carry out the inhibition experiments with the completely reduced species. Therefore, we carried out the experiment with the reduced species (zwitterion, **2c**), which was obtained by reducing the diselenide (**8**) with NaBH₄ in an aqueous solution. As expected, MMI inhibited the LPO activity with an IC₅₀ value of 7.8 μM, which is much lower than that observed with PTU and MTU. The selenium analogue (**2c**) also inhibited LPO, and

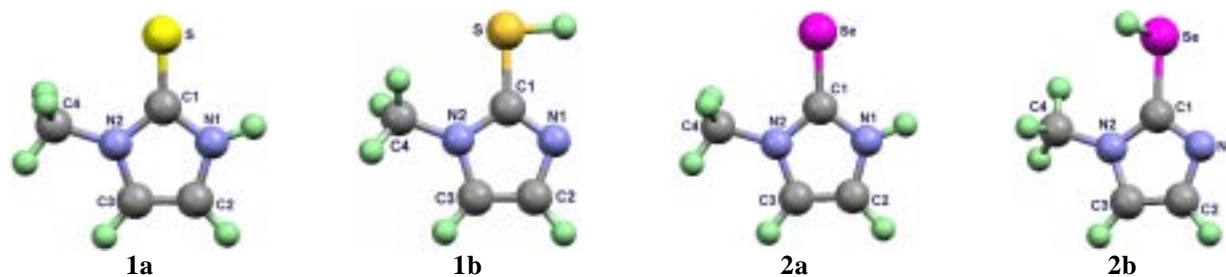


Figure 4. Optimized geometries of **1** and **2**. The conversion of thiol to thione is more favoured than the conversion of selenol to selone. The structures were optimised at the B3LYP level of theory using 6-311++G (*d, p*) basis set.

Table 1. The theoretical data for **2** and **8** obtained by DFT calculations at B3LYP/6-311++G (*d, p*) level along with the GIAO ^{77}Se NMR chemical shifts.

Compound	C–Se bond length (Å)	^{77}Se Chemical shift (ppm) ^b
2a	1.835	30
2b	1.917	–101
2c	1.835	30
8	1.902 ^a	386

^aCalculated using B3LYP/6-31G(d) level; ^bNMR values were calculated using B3LYP/6-311++G(2d,p) level and referenced to Me_2Se

Table 2. Inhibition of LPO activity by **1–3**, and **5**.

Compound	IC_{50} (μM) ^a
MMI (1)	7.0 ± 1.1
MSeI (2)	16.4 ± 1.5
PTU (3)	45.0 ± 2.1
MTU (5)	47.8 ± 0.1

^aConcentration of the compound causing 50% inhibition. Each IC_{50} value was calculated from at least three independent experiments

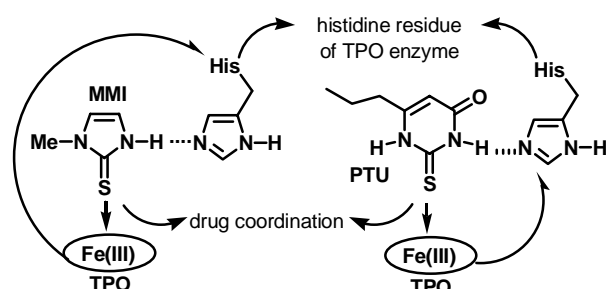


Figure 5. A hypothetical model for the coordination of thiourea drugs to the Fe-centre of TPO.

the IC_{50} value was found to be almost 4–5 times lower than that of PTU and MTU. The higher activity of MMI as compared with PTU and MTU is in

agreement with the previous studies on the inhibition of TPO. Since the activation of the iron centre in TPO must proceed through an interaction of Fe(III) with H_2O_2 , TPO inactivation may occur through a competitive coordination of the drug to iron, assisted by hydrogen bonding with a histidine residue of the TPO enzyme (figure 5). Under these conditions, MMI might compete more successfully than PTU with H_2O_2 , because the hydrogen-bond (hard) basicity pK_{HB} value of MMI (2.11) is much higher than that of PTU (~ 1.32). Similar to PTU, the methyl derivative **5** is also expected to be a weak inhibitor of TPO. On the other hand, compound **2**, which exists predominantly in its zwitterionic form, does not have the ability to coordinate to the iron centre and, therefore, this compound must inhibit the LPO activity by a different mechanism.

Taurog *et al* have shown that MMI and related derivatives irreversibly inhibit LPO and TPO, leading to a complete inactivation of the enzymes.¹⁵ Doerge and others have shown that mammalian peroxidases including LPO may activate the anti-thyroid drugs through S-oxygenation to produce the corresponding sulphoxides or sulphenic acids.¹⁶ They have also shown that the irreversible inactivation of LPO and TPO by MMI proceeds through the S-oxygenation of the thione moiety to form a reactive sulphenic acid, which binds covalently to the prosthetic heme and irreversibly blocks enzyme activity.¹⁷ Given the higher reactivity of selenium compounds as compared with the sulphur derivatives towards oxidation, it is possible that the facile oxidation of the selenium compounds may lead to an efficient inhibition of LPO activity. With this in mind, we treated all the compounds in this study with H_2O_2 before adding LPO and ABTS. The LPO activity was measured several times by increasing the time for the reaction of the test compounds with H_2O_2 . Remarkably, MSeI (**2**) inhibited the enzyme within few seconds

even at lower concentrations, which can be ascribed to the facile oxidation of the reactive selenium moiety in **2** (MSeI) by H_2O_2 . Because MMI also inhibits the enzyme very efficiently, we have carried out further experiments to prove that the mechanisms by which MMI and MSeI exert their inhibitory action are different. The initial rates (v_0) derived from various concentrations of H_2O_2 were plotted against the concentration of H_2O_2 . The LPO activity was completely inhibited by $40 \mu\text{M}$ MMI and the enzyme's activity could not be recovered by increasing the H_2O_2 concentration (figure 6, d). The LPO activity could not be recovered even at lower concentration of MMI ($10 \mu\text{M}$) and higher concentration of H_2O_2 ($230 \mu\text{M}$). This suggests that MMI does not act on H_2O_2 , but it acts on the enzyme itself leading to an irreversible inhibition as previously proposed. On the other hand, **2** also inhibited the LPO activity as efficient as MMI, but in this case, the enzyme's activity could be completely recovered by increasing H_2O_2 concentration (figure 6, b). The sigmoidal behaviour of the graph for this compound (figure 6, b) is probably due to the utilization of H_2O_2 for the oxidation of selenenic acid to other oxidized products at lower concentration of the peroxide.

These observations strongly support the assumption that MSeI, in contrast to MMI, does not interfere with the enzyme directly, but it inhibits the LPO

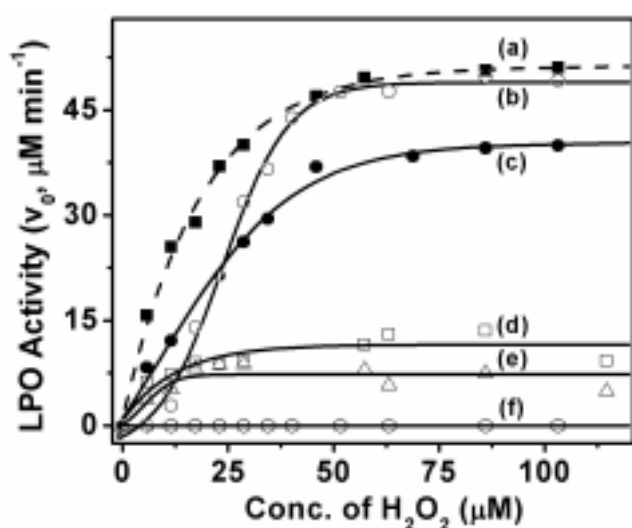


Figure 6. Plot of initial rates (v_0) for the LPO-catalysed oxidation of ABTS vs concentration of H_2O_2 . (a) Control activity, (b) $40 \mu\text{M}$ of **2**, (c) $40 \mu\text{M}$ of **8**, (d) $80 \mu\text{M}$ of PTU, (e) $80 \mu\text{M}$ of MTU, (f) $40 \mu\text{M}$ of MMI. Conditions: LPO: 6.5 nM ; H_2O_2 : $22.9 \mu\text{M}$.

activity by reducing the H_2O_2 , which is required for the oxidation of the iron centre in LPO (figure 7). When coupled with a suitable thiol such as GSH, compound **2** may constitute redox cycle involving a catalytic reduction of H_2O_2 (glutathione peroxidase (GPx) activity).¹⁸ In this way, compound **2** mimics the action of GPx, a selenoenzyme that protects the cellular components from oxidative damage by reducing H_2O_2 with the help of GSH. Recently, the GPx enzyme present in thyroid gland has been shown to inhibit the iodination reactions by degrading the intracellular H_2O_2 .¹⁹ In fact, the key compound **2** exhibited interesting GPx activity, leading to an assumption that some of the antithyroid drugs may act as antioxidants in addition to their inhibition behaviour. However, GSH does not appear to be a suitable thiol co-substrate for the catalytic antioxidant activity of **2**, because the reaction of **2** with hydrogen peroxide was found to be much faster than the reduction of the intermediate selenenic acid and/or other oxidized species by GSH. Similar to the zwitterion-mediated inhibition, the inhibition by diselenide **8** could also be reversed by increasing the H_2O_2 concentration (figure 6, c). Although compound **8** did not give any new ^{77}Se NMR signal with one equiv. H_2O_2 , addition of an excess amount of H_2O_2 to **8** produced a new signal at 1045 ppm, which cannot be ascribed to the selenenic acid because this signal appears to be different from that obtained from the reaction of **2** with H_2O_2 , which showed a signal at 1207 ppm.

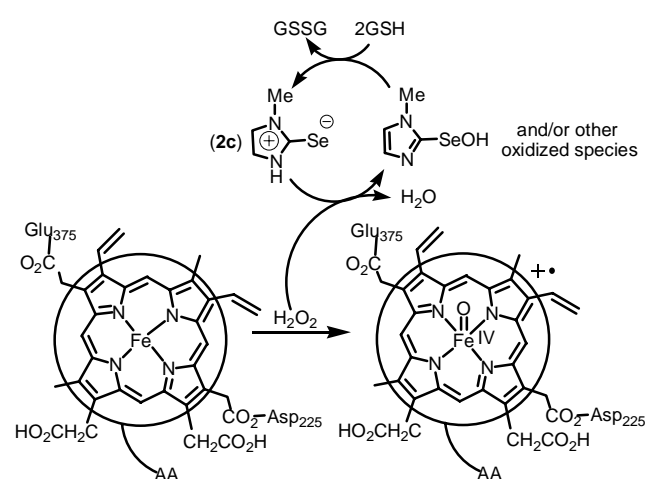


Figure 7. A hypothetical model representing the inhibition of LPO by **2**. The porphyrin core inside the circle represents the active centre of LPO. AA: amino acid residues.

As expected, the plot of initial rates (v_o) vs concentration of MSeI shows that the rate of the reaction decreases with increasing concentration of MSeI. In all these cases, the LPO activity could be recovered by increasing the hydrogen peroxide concentration. These experimental observations support the conclusions made by Taurog *et al* that MSeI, unlike MMI, cannot act as an irreversible inhibitor of TPO. These observations also support the *in vivo* experiments, which showed that MMI is at least 50 times more potent than MSeI as an inhibitor of organic iodine formation in the thyroid. Crucially, the treatment of **2** with the selenolate specific reagent, iodoacetic acid, abolished the inhibitory potency of **2**, confirming that the oxidation of the selenium centre by H_2O_2 is responsible for the inhibition. In contrast, the sulphur analogue MMI was found to be less sensitive to the iodoacetic acid treatment and this also confirms that the thiol form of MMI is not only less predominant in solution, but also less reactive as compared with the thione form.

3. Conclusion

In summary, our experimental and theoretical studies show that the selenium analogue of methimazole (MSeI) exists predominantly in its zwitterionic form whereas the sulphur compound exists in its thione form and the oxidation of the zwitterion to the corresponding diselenide is energetically more favoured than the conversion of thione/thiol to the corresponding disulfide. Although MSeI readily oxidizes to produce the diselenide, the oxidized form can be easily reduced by reducing agents such as $NaBH_4$ or glutathione (GSH). The ^{77}Se NMR studies show that the selenone form of MSeI dissociates in solution to form a more reactive selenolate, which could be trapped by selenolate specific reagents such as iodoacetic acid. In its reduced form, MSeI effectively and reversibly inhibits the iron-containing lactoperoxidase (LPO). In contrast to methimazole, MSeI does not interfere with the enzyme directly, but it inhibits LPO by reducing the H_2O_2 that is required for the oxidation of the iron centre in LPO. In the presence of GSH, MSeI constitutes a redox cycle involving a catalytic reduction of H_2O_2 and thereby mimics the glutathione peroxidase (GPx) activity *in vitro*. These studies reveal that the degradation of the intracellular H_2O_2 by the selenium analogues of anti-thyroid drugs may be beneficial to the thyroid gland as these compounds may act as antioxidants

and protect thyroid cells from oxidative damage. Because the drugs with an action essentially on H_2O_2 can reversibly inhibit the thyroid peroxidase, such drugs with a more controlled action could be of great importance in the treatment of hyperthyroidism.

Acknowledgments

This study was supported by the Department of Science and Technology (DST), and Council of Scientific and Industrial Research (CSIR), New Delhi. GM acknowledges the DST for the Ramanna Fellowship and GR thanks the CSIR for a research fellowship.

References

- (a) Taurog A 1951 *Thyroid hormone synthesis*. In *Werner's the Thyroid* (eds) L E Braverman and R D Utiger, pp 51–97; (b) Taurog A, Dorris M L and Dörge D R 1996 *Arch. Biochem. Biophys.* **330** 24
- (a) Berry M J, Banu L and Larsen P R 1991 *Nature (London)* **349** 438; (b) Larsen P R and Berry M J 1995 *Annu. Rev. Nutr.* **15** 323; (c) St Germain D L and Galton V A 1997 *Thyroid* **7** 655; (d) Köhrle J 1999 *Biochimie* **81** 527; (e) Bianco A C, Salvatore D, Gereben B, Berry M J and Larsen P R 2002 *Endocr. Rev.* **23** 38; (f) Köhrle J 2002 *Methods Enzymol.* **347** 125
- (a) Buxeraud J, Absil A C, Claude J, Raby C, Catanzano G and Beck C 1985 *Eur. J. Med. Chem.* **20** 43; (b) Raby C, Lagorce J F, Jambut-Absil A C, Buxeraud J and Catanzano G 1990 *Endocrinology* **126** 1683
- Bassosi R, Nicolai N and Rossi C 1978 *Biophys. Chem.* **8** 61
- (a) Visser T J, Kaptein E and Aboul-Enein H Y 1992 *Biochem. Biophys. Res. Commun.* **189** 1362; (b) Taurog A, Dorris M L, Guziec L J and Guziec F S Jr 1994 *Biochem. Pharmacol.* **48** 1447; (c) Guziec L J and Guziec F S Jr 1994 *J. Org. Chem.* **59** 4691; (d) Taurog A, Dorris M L, Hu W-X and Guziec F S Jr 1995 *Biochem. Pharmacol.* **49** 701
- (a) Roy G, Nethaji M and Mugesh G 2004 *J. Am. Chem. Soc.* **126** 2712; (b) Roy G and Mugesh G 2005 *J. Am. Chem. Soc.* **127** 15207; (c) Roy G, Nethaji M and Mugesh G 2006 *Org. Biomol. Chem.* **4** 2883; (d) Roy G, Das D and Mugesh G 2007 *Inorg. Chem. Acta* **360** 000, doi: 10.1016/j.ica.2006.07.052
- Balestrero R S, Forkey D M and Russell J G 1986 *Magn. Reson. Chem.* **24** 651
- Antoniadis C D, Corban G J, Hadjikakou S K, Hadjiliadis N, Kubicki M, Warner S and Butler I S 2003 *Eur. J. Inorg. Chem.* **8** 1635
- Laurence C, El Ghomari M J, Le Questel J-Y, Berthelot M and Mokhlisse R 1998 *J. Chem. Soc., Perkin Trans. 2* 1545

10. Aragoni M C, Arca M, Demartin F, Devillanova F A, Garau A, Isaia F, Lippolis V and Verani G 2002 *J. Am. Chem. Soc.* **124** 4538
11. (a) Ha T-K and Puebla C 1994 *Chem. Phys.* **181** 47; (b) Jemmis E D, Giju K T and Leszczynski J 1997 *J. Phys. Chem.* **A101** 7389
12. Kim H, Lee T-H, Hwang Y S, Bang M A, Kim K H, Suh J M, Chung H K, Yu D-Y, Lee K-K, Kwon O-Y, Ro H K and Shong M 2001 *Mol. Pharmacol.* **60** 972
13. Taurog A, Dorris M L and Lamas L 1974 *Endocrinology* **94** 1286
14. Edelhoch H, Irace G, Johnson M L, Michot J L and Nunez J 1979 *J. Biol. Chem.* **254** 11822
15. (a) Taurog A, Dorris M L and Guziec F S Jr 1989 *Endocrinology* **124** 30, and references therein
16. (a) Doerge D R 1986 *Arch. Biochem. Biophys.* **244** 678; (b) Kobayashi S, Nakano M, Goto T, Kimura T and Schaap A P 1986 *Biochem. Biophys. Res. Commun.* **135** 166
17. Doerge D R, Cooray N M and Brewster M E 1991 *Biochemistry* **30** 8960
18. Sarma B K and Mughesh G 2005 *J. Am. Chem. Soc.* **127** 11477, and references therein
19. (a) Björkman U, Ekholm R 1995 *Mol. Cell. Endocrinol.* **111** 99; (b) Ekholm R and Björkman U 1997 *Endocrinology* **138** 2871