

PUBLISHED VERSION

Melanie J. Ceko, Sean O'Leary, Hugh H. Harris, Katja Hummitzsch, and Raymond J. Rodgers
Trace elements in ovaries: measurement and physiology
Biology of Reproduction, 2016; 94(4):86-1-86-14

© 2016 by the Society for the Study of Reproduction, Inc. This article is available under a Creative Commons License 4.0 (Attribution-Non-Commercial), as described at <http://creativecommons.org/licenses/by-nc/4.0>

Published version <http://dx.doi.org/10.1095/biolreprod.115.137240>

PERMISSIONS

<http://creativecommons.org/licenses/by-nc/4.0/>



Attribution-NonCommercial 4.0 International (CC BY-NC 4.0)

This is a human-readable summary of (and not a substitute for) the [license](#).

[Disclaimer](#)

You are free to:

Share — copy and redistribute the material in any medium or format

Adapt — remix, transform, and build upon the material

The licensor cannot revoke these freedoms as long as you follow the license terms.

Under the following terms:



Attribution — You must give **appropriate credit**, provide a link to the license, and **indicate if changes were made**. You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.



NonCommercial — You may not use the material for **commercial purposes**.

No additional restrictions — You may not apply legal terms or **technological measures** that legally restrict others from doing anything the license permits.

Notices:

You do not have to comply with the license for elements of the material in the public domain or where your use is permitted by an applicable **exception or limitation**.

No warranties are given. The license may not give you all of the permissions necessary for your intended use. For example, other rights such as **publicity, privacy, or moral rights** may limit how you use the material.

4 May, 2016

<http://hdl.handle.net/2440/98534>

Minireview

Trace Elements in Ovaries: Measurement and Physiology¹

Melanie J. Ceko,³ Sean O’Leary,⁴ Hugh H. Harris,³ Katja Hummitzsch,⁴ and Raymond J. Rodgers^{2,4}

³Department of Chemistry, The University of Adelaide, South Australia, Australia

⁴Discipline of Obstetrics and Gynaecology, School of Medicine, Robinson Research Institute, The University of Adelaide, South Australia, Australia

ABSTRACT

Traditionally, research in the field of trace element biology and human and animal health has largely depended on epidemiological methods to demonstrate involvement in biological processes. These studies were typically followed by trace element supplementation trials or attempts at identification of the biochemical pathways involved. With the discovery of biological molecules that contain the trace elements, such as matrix metalloproteinases containing zinc (Zn), cytochrome P450 enzymes containing iron (Fe), and selenoproteins containing selenium (Se), much of the current research focuses on these molecules, and, hence, only indirectly on trace elements themselves. This review focuses largely on two synchrotron-based x-ray techniques: X-ray absorption spectroscopy and x-ray fluorescence imaging that can be used to identify the in situ speciation and distribution of trace elements in tissues, using our recent studies of bovine ovaries, where the distribution of Fe, Se, Zn, and bromine were determined. It also discusses the value of other techniques, such as inductively coupled plasma mass spectrometry, used to garner information about the concentrations and elemental state of the trace elements. These applications to measure trace elemental distributions in bovine ovaries at high resolutions provide new insights into possible roles for trace elements in the ovary.

bromine, iron, ovary, selenium, synchrotron x-ray fluorescence, trace elements, zinc

INTRODUCTION

When living organisms migrated to the land from water during the Paleozoic period, they had to depend on the soil for their source of biological trace elements [1]. A total of 26 of the 90 naturally occurring elements are known to be essential for animal life, including copper (Cu), iron (Fe), selenium (Se),

zinc (Zn), cobalt, fluorine, iodine, manganese, molybdenum, nickel, silicon, and vanadium, with some disagreement over arsenic, chromium, and tin being essential trace elements [2–4]. Evidence for essentiality of a particular trace element is established when symptoms are induced by dietary deficiency, then subsequently reversed after a diet that is replete in this specific trace element is provided [5]. Essentiality is then generally acknowledged when it has been demonstrated by more than one independent investigator and in more than one animal species [6]. Evolutionary selection of these elements remains a mystery; the property that the suite of elements discussed in this review has in common is that they commonly occur and function in living organisms at low concentrations. Interest in the potential role of trace elements in human and animal physiology began over a century ago with the discovery that a number of compounds in living organisms contained metals not previously considered to be of biological significance [6]. When studying the relationship of trace elements in human health, it becomes increasingly evident that optimal levels of elements in every organ, tissue, and cell of the human body may be key to maintaining a healthy existence [7]. Trace elements are present in every biological process, from the production of hormones and energy, digestion, nerve transmission, and muscle contraction to the regulation of pH, metabolism, and cholesterol and blood sugar levels [7]. Stress and exposure to environmental pollution alter our requirements for these minerals, and scientists are becoming increasingly aware that trace element deficiencies can affect both physical and mental health.

In addition to identifying trace elements, elucidating the roles they might be playing at a biochemical level, and considering the optimal dietary intake, bioavailability, or amount of utilizable element relative to the amount present in the daily diet, are important considerations [8]. Trace element bioavailability is influenced by a complex matrix of interacting variables, including the chemical form of the element found in food, the nature of the food ingested, the composition of the total diet, and the health and nutritional well being of the individual [3, 9]. Dietary deficiencies of trace elements have been reported to alter various aspects of reproductive physiology [10]; however, to date, the majority of these studies have focused on their roles in male reproductive function.

Traditionally, research in the field of trace element biology and human health has largely depended on epidemiological methods to demonstrate involvement in biological processes. These studies were typically followed by trace element supplementation trials or attempts at identification of the biochemical pathways involved. With the discovery of

¹Funding was received from Australian Research Council grants DP0985807 and DP0984722 and the National Health and Medical Research Council of Australia.

²Correspondence: Ray Rodgers, School of Medicine, Robinson Research Institute, The University of Adelaide, SA 5005, Australia.
E-mail: ray.rodgers@adelaide.edu.au

Received: 20 November 2015.
First decision: 17 December 2015.
Accepted: 28 January 2016.

© 2016 by the Society for the Study of Reproduction, Inc. This article is available under a Creative Commons License 4.0 (Attribution-Non-Commercial), as described at <http://creativecommons.org/licenses/by-nc/4.0>

eISSN: 1529-7268 <http://www.biolreprod.org>
ISSN: 0006-3363

TABLE 1. Newer technical approaches to study trace elements in tissues.

Analytical platform	Mass of tissue	Tissue preparation	In situ quantification	Range of elements ^a	Accuracy ^b	Relative cost ^c
S-XRF	1 mg–1 g (wet weight)	No preconditioning of tissue	Yes, morphology intact. Single-cell level	Multielement, simultaneous	High	Low
XRF	1–7 g (dry weight)	Decomposition to powder or emulsion	No	Multielement (Mg to U), difficult with lighter elements	Medium	Medium
XAS	0.02–0.5 g (dry weight)	Unnecessary	Possible, although bulk analysis more common	Multielement (Mg to U), nonsimultaneous	Low, generally used to provide chemical rather than elemental (quantitative) information	Medium
ICP-MS	0.2 mg (dry weight)	Decomposition to liquid phase	No	Multielement, simultaneous	Medium to high	Low

^a Range of elements is generally considered to be from Mg to U for XAS and XRF, and lighter elements are possible with S-XRF and ICP-MS.

^b Accuracy: nearness of result to the true result, is dependent on sample preparation, homogeneity of tissue, and interfering matrix effects (see text).

^c Relative cost of analysis takes into account preparation of sample, start-up costs, etc. S-XRF and ICP-MS require expensive equipment that is normally shared between institutions, allowing for assay costs to be relatively low (shared) compared with platforms using less-expensive equipment, but operated within individual institutions.

biological molecules that contain the trace elements, such as matrix metalloproteinases containing Zn, superoxide dismutase containing Mn or Cu, cytochrome P450 enzymes containing Fe, or selenoproteins containing Se, much of the current research focuses on these proteins and, hence, only indirectly on trace elements themselves. While useful, these approaches overlook key roles of many trace elements in biological processes, including reproduction. In this review, we cover some of the newer techniques for studying trace elements and, in particular, their application to the study of the ovary.

NEWER TECHNICAL APPROACHES TO STUDY TRACE ELEMENTS

The objectives of newer approaches to study trace elements in cells and tissues are first to identify which trace elements are present, to quantify levels of trace elements, and to determine the location of the trace elements in tissues and cells. According to the findings of the study of these parameters, assumptions can be made with regard to tissue- and cell-specific function of trace elements. In a paper designed to evaluate laboratory methods for trace element determination, Bolann et al. [11] summarized that the analysis of trace elements in biological fluids and tissues serves multiple purposes. These include determining the concentration and distribution of essential trace elements in normal and disease conditions, detection and allocation of potentially toxic metals, diagnosis of trace element deficiency states, and trace element-related diseases [11]. According to their findings, the four most commonly used techniques for trace element analysis in human biological material are: 1) flame atomic absorption spectrometry, 2) graphite furnace atomic absorption spectrometry, 3) inductively coupled plasma atomic emission spectrometry, and 4) inductively coupled plasma mass spectrometry (ICP-MS), with ICP-MS being preferable for simultaneous screening of multiple elements [11].

In another study designed to ascertain the most effective means of analyzing the elemental composition of a geological sample (samples of sediment from the bottom of a lake), Phedorin et al. [12] compared synchrotron x-ray fluorescence (S-XRF), traditional XRF, ICP-MS, atomic absorption spectrometry, and instrumental neutron activation analysis, and concluded that S-XRF was superior from the point of view of its speed, ease of application, cost, nondestructive nature, and sensitivity [12]. These factors, coupled with the simultaneous determination of multiple elements of geochemical or biochemical interest, make synchrotron-based XRF a highly sought-after technique.

Weekley et al. [13] assert that XRF imaging and x-ray absorption spectroscopy (XAS) are ideal techniques for investigating the chemical speciation and distribution of elements heavier than silicon in biological systems, with minimal sample preparation required [13]. XRF imaging is now a well-established, nondestructive analytical method applicable across a large variety of fields, including materials science, environmental science, geology, life science, and archeological science [14]. The discussion below describes the newer approaches to studying trace elements, detailing the basic principle behind the approach, including the advantages and limitations of each platform. Table 1 compares the platforms with regard to sample preparation and salient features.

S-XRF Imaging

The principle of XRF involves using x-rays of sufficient energy to expel tightly held inner electrons of an element

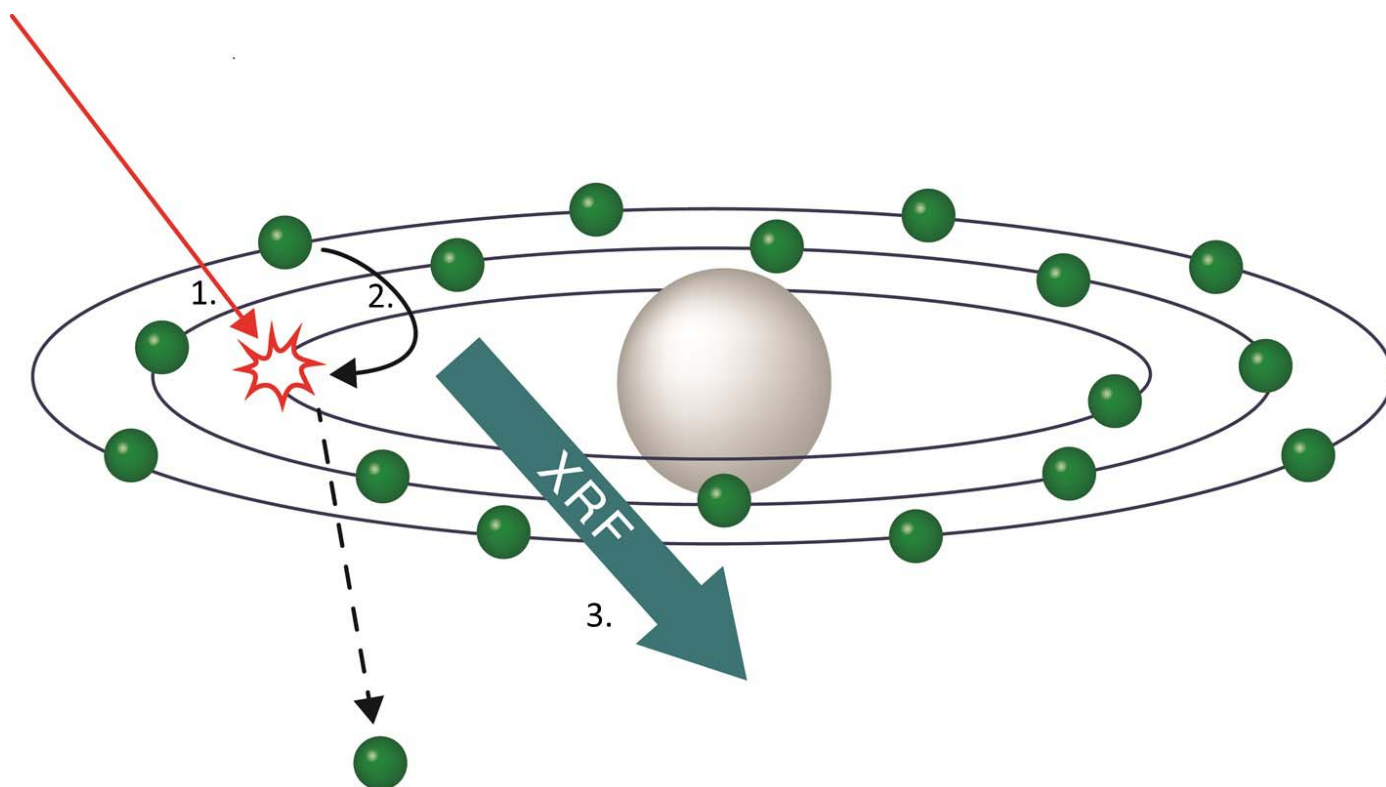


FIG. 1. Schematic diagram demonstrating the physical principles associated with S-XRF. 1) X-rays that strike an atom may knock electrons out of it, leaving behind an ionized version of the atom (i.e., one missing electron), and an unoccupied electron energy level. 2) An electron of a higher energy will then drop down to fill the lower-energy, unoccupied level, releasing an x-ray photon as a means to conserve energy. 3) The emission of the x-ray photon is the source of the fluorescence. The electron energy levels are very specific to each element, so that the energy of the S-XRF produced precisely describes the elements present in the biological sample.

within a sample. This makes the electronic structure of the element unstable, and an electron from a higher orbit replaces the inner electron, at the same time releasing energy in the form of a fluorescent photon. The fluorescent photon has a characteristic energy equal to the difference in energy between the outer and inner orbital [15]. As the electron energy levels are specific for each element, the fluorescence produced by the x-ray precisely identifies the elements within the sample (see Fig. 1).

A synchrotron is a circular particle accelerator that uses both magnetic and electric fields to accelerate charged particles. During this process, bunches of electrons are accelerated to about the speed of light within an orbital storage ring, and give off synchrotron radiation, which can then be harnessed for the x-ray source for XRF (see Fig. 2). S-XRF imaging is a powerful technique that harnesses the finely focused x-ray beam that is several orders higher in intensity than radiation from traditional x-ray sources, and focuses this onto a small spot on a biological sample (e.g., an ovarian section). Digital images of microscopic tissue samples are built, pixel by pixel, by scanning the sample through the beam, and mathematical deconvolution analysis of the fluorescence spectrum reveals the elemental composition. Quantitative elemental distribution maps of the sample can be assembled [16] to reveal the spatial organization of trace elements within the sample with high spatial resolution and sensitivity (to ppm or better) for a large number of elements (Fig. 2) [17]. The S-XRF process is undertaken without destroying the tissue under study [18].

Applications of S-XRF Imaging to Biological Systems

To date, there is a plethora of studies, ranging from elemental tissue profiles and identifying toxic metals in human teeth [19–21], to determining levels of calcium and phosphorous in the bones in osteoporosis research [22, 23], and comparing chemical composition in liver biopsies from untreated cirrhosis patients with healthy controls [19, 24]. However, only a limited number of XRF imaging studies have been carried out involving male or female reproductive organs.

Quantitative analysis of Zn and Ca in human prostate cancer and normal tissues by S-XRF imaging was undertaken by Ide-Ektessabi et al. [25] in order to investigate differences in the distribution and concentrations of Zn in these tissues. Ortega et al. [26] mapped the distribution of chromium in the reproductive organs of male mice after exposure to CrCl_3 to elucidate the specific cellular regions where it had accumulated. Two more recent papers presented S-XRF imaging of the ovaries to investigate the bioaccumulation of dietary Zn in *Daphnia magna*, a common ecotoxicological model [27, 28]. In both cases, the researchers set out to investigate the hypothesis that Zn selectively accumulates in female reproductive tissues and adversely affects reproduction. S-XRF has also been used in recent years to analyze the trace elemental composition of human breast tissue, and has revealed statistically significant elevation of a multitude of elements in neoplastic tissue compared to normal samples [29–33].

In summary, the use of S-XRF in elucidating the distribution and quantification of both essential and toxic trace elements in tissues provides new insights into diseases and provides ever-increasing applications to biological systems

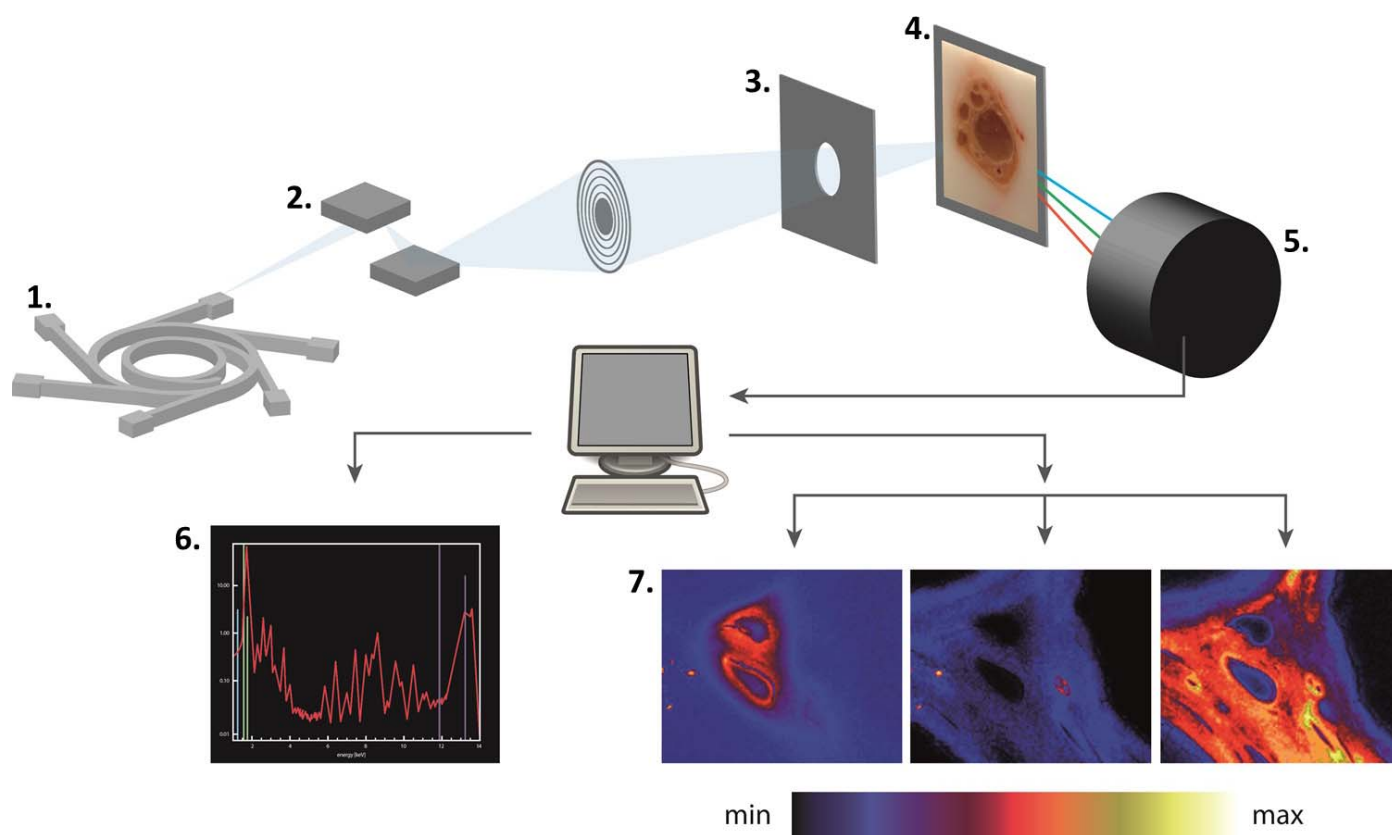


FIG. 2. Schematic representation of S-XRF. The sequence of steps includes: 1) X-rays produced by highly energetic electrons accelerated in the synchrotron storage ring; 2) x-ray optics are then used to energy filter; and 3) focus synchrotron sourced x-rays onto a small spot (several nanometers to micrometers in diameter) on the sample; 4) the sample is then raster scanned through the beam; 5) an energy-dispersive fluorescence detector is used to produce a full-fluorescence spectrum; and 6) peaks correspond to specific elements. By comparing these spectra to reference standards of known concentration, quantitative in situ maps of multiple elements (7) can be created simultaneously.

[34]. However, despite there being about 90 synchrotrons worldwide, only a handful have the capability of XRF imaging, and even fewer have the advanced capability to collect large amounts of data quickly, as was recently undertaken in the ovary [35, 36].

X-Ray Absorption Spectroscopy

The principle of XAS is similar to XRF, where synchrotron-sourced x-rays are used to ionize core electrons, creating an absorption edge with an energy that is characteristic to each element. XAS determines the local geometric and/or electronic structure of trace elements in matter [37]. More specifically, information can be gleaned on bond lengths, coordination numbers, local coordination geometry, and the oxidation state of atoms for a wide range of solid or liquid samples. Traditionally, the XAS spectrum is divided into two regions: the low-energy region, which covers photon energy up to about 50 eV above the absorption edge, often referred to as x-ray absorption near-edge structure (XANES); and the higher-energy region, from 50 to 1000 eV above the absorption edge, which is called the extended x-ray absorption fine structure (EXAFS) [38, 39]. Although the two regions have the same physical origin, this distinction is convenient for their interpretation [40]. XANES is highly sensitive to the spectral oxidation state and coordination chemistry of the absorbing atom, while the EXAFS is used to determine the distances, coordination number, and identity of the neighboring atoms. Thus, the information obtained using XAS is biologically

useful in ascertaining which biological molecules the elements are associated and, hence, their functions.

Inductively Coupled Plasma-Mass Spectrometry

The principle of ICP-MS involves a high-temperature inductively coupled plasma source, which converts the atoms of the elements within a sample into gaseous ions. These ions are then separated and detected by mass spectroscopy due to their mass-to-charge ratio. The multielemental capability of ICP-MS for the analysis of biological samples makes this an ideal supplementary technique for quantifying the trace elements identified by XRF imaging. ICP-MS is capable of detecting metals and several nonmetals at concentrations as low as one part per trillion [41].

ROLE OF TRACE ELEMENTS IN FEMALE REPRODUCTIVE FUNCTION

Over the last four decades, countless studies have been conducted on the role of dietary trace elements on reproductive function; however, a review of the literature quickly highlights how heavily the research has been weighted toward male compared to female reproductive function. Studies with a great deal of evidence on the impact of micronutrients, including trace elements, on female fertility are rare [42]. Although there are many human and animal studies focusing on the dietary intake of trace elements during pregnancy, and the resultant effects on pregnancy outcome, very few studies take into account the preconception period, or focus on general

TABLE 2. Trace elements and proposed effect on ovarian function.

Element	Bromine	Iron	Selenium	Zinc
Source	Soils, anthropogenic release	Food/supplements	Food/supplements	Food/supplements
Proposed mechanism/function	Halogen replacement BDBP & PBBs, collagen formation	Cell growth/function, LH & FSH secretion	↓ROS/oxidative stress	↓ROS/oxidative stress
Deficiency	↓ Tissue integrity[56]	Infertility [102], anovulation [193], hypoxia (?)	Unexplained infertility [194], POF (?)	↓ Oocyte maturation anovulation [195]
Toxicity	Transgenerational infertility [78]	Infertility [103, 104] iron-induced oxidative stress (?)	n/a	n/a

BDBP, brominated disinfection by-product; n/a: information not available; POF, premature ovarian failure; (?), putative mechanism.

reproductive health when it comes to ovarian function, including hormone synthesis and follicular development.

In the late 1990s, Xu et al. hypothesized that a disorder in the metabolism of trace elements may be one of the important factors causing unexplained infertility [43]. Recent studies, which are reviewed elsewhere [44], have reported that changes in the levels of the trace elements, Fe, Zn, and Ca, play important roles in female infertility, but there is still debate about the relationship between Fe and unexplained infertility [44]. A better understanding of the role of trace elements in the underlying mechanisms in infertility, as well as more rigorous studies clarifying the effectiveness of nutritional factors, are needed to improve diagnosis and treatment [45]. Despite the paucity of studies, it is clear that there is a strong relationship between nutritional status of trace elements prior to conception and successful conception and healthy progression of pregnancy [46].

In order to gain a better understanding of the role of trace elements in the ovary, we recently undertook quantitative XRF imaging of bovine ovaries and conducted ICP-MS and XAS analyses [35, 36, 47, 48]. In particular, we obtained data on bromine (Br), Fe, Se, and Zn. These trace elements and their potential roles in the ovary form the basis of this review. However, some ovaries showed evidence of other trace elements, including Cu, cobalt, chromium, manganese, nickel, and titanium, but these were not in particularly high concentrations, nor were they found in a significant portion of samples (<10%), and so these elements were not examined further. In the following sections, we discuss what is known about Br, Fe, Se, and Zn in general and in reproduction, and, in particular, in the ovary. Table 2 summarizes the proposed effects of these trace elements on female fertility and ovarian function. Table 2 also includes proposed mechanisms associated with deficiency, and where possible toxicity of these trace elements is associated with ovarian function.

BROMINE

Br is one of the most abundant and ubiquitous trace elements in the biosphere, but, until recently, had not been conclusively shown to perform any essential function in plants, micro-organisms, or animals [2]. Up until the mid-1980s, only weak evidence existed to support the view that Br was essential, with one of the key findings being that bromide ($\text{Br}^-_{\text{[aq]}}$) could substitute for part of the chloride (Cl^-) requirement for chickens [49]. In 1990, one additional study reported that, when compared to goats fed a 20-mg Br/kg diet, goats fed a 0.8-mg Br/kg diet exhibited depressed growth and fertility, increased abortions, and reduced life expectancy [50]. In addition, microscopic examination of tissues and organs of

Br-deficient goats showed abnormalities in tissue sections of the thyroid, heart, lungs, pancreas, and ovaries [51]. In 1998, it was suggested that, although these additional studies gave more credibility to the concept of Br essentiality, the findings were still regarded as too limited [52]. The evidence provided little insight into a possible biochemical role for this element, beyond saying that it may serve as an electrolyte [52]. This led to the general assumption that the biological behavior of Br was similar to chlorine (Cl) in that the administration of it results in some displacement of body Cl^- , and vice versa [53]. This assumption has since been found to be invalid for the thyroid gland. In studies on the interaction of Br with I in the rat thyroid, under the conditions of enhanced Br intake, Pavelka et al. found that, contrary to other tissues, $\text{Br}^-_{\text{(aq)}}$ replaced iodide (I^-) as opposed to Cl^- [54, 55]. As previously mentioned, a recent study demonstrated that Br is required as a cofactor for peroxidase-catalyzed formation of sulfilimine crosslinks in *Drosophila* [56]. This process is involved in cross-linking the noncollagenous domains of collagens type IV, which are the collagen components of basal laminae found in many tissues. Br dietary deficiency led to physiologic dysfunction in *Drosophila*, while repletion of this trace element reversed the dysfunction, a key criterion for justifying essentiality [56].

Biological Activity of Brominated Species

Potential biological roles of brominated species have been under scrutiny for the past 13 yr, with several authors making important contributions to this area of knowledge. Hawkins et al. conducted an experiment to show that hypochlorous acid (HOCl) and hypobromous acid (HOBr) reacted with different selectivity with cellular targets, and that the resultant radical formation may result in cell lysis [57]. The reactivity of HOBr with biological molecules was not well characterized, but amino acids and proteins appear to be major targets, with the species reacting with fatty acid side chains and lipid-soluble antioxidants to a much greater extent than HOCl [58–60]. This species has also been reported to induce red blood cell lysis approximately 10 times more rapidly than HOCl [57]. In a later study to ascertain the rate constants for reactions of HOBr with protein components, most residues reacted 30- to 100-fold faster with this species than with the Cl^- -containing acid [58]. More recently, the contribution of HOBr to optimal and efficient microbial killing has been recognized as essential [61]. Marcinkiewicz et al. [62] investigated the role of another brominating oxidant, taurine bromamine, and found that this species exerted strong bactericidal effects on *Escherichia coli*. Maines et al. [63] investigated the cytotoxic effects of activated Br on human fetal osteoblasts in vitro and showed that sodium

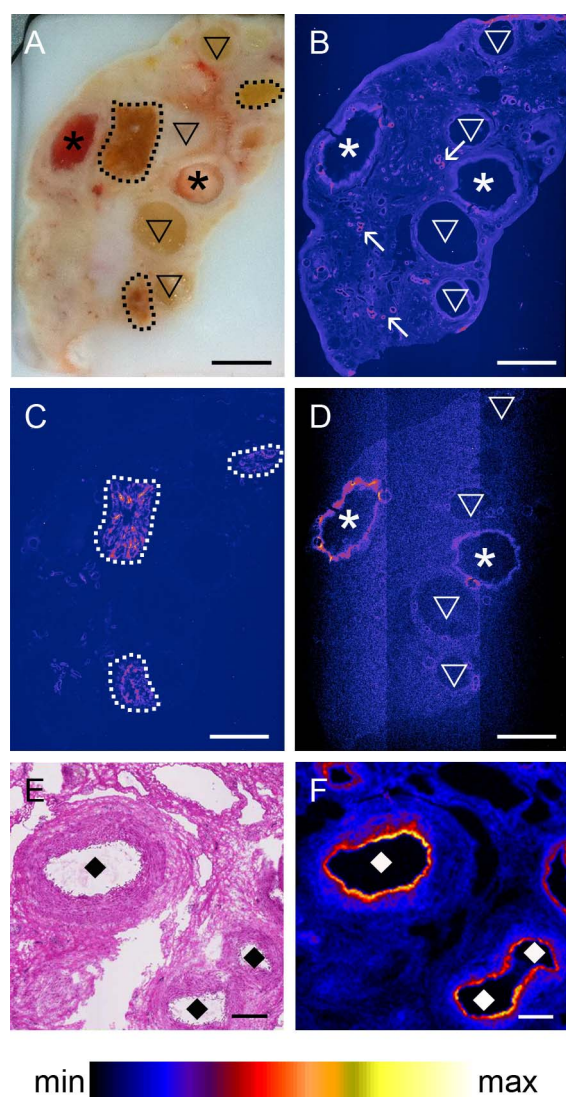


FIG. 3. Distribution of Zn, Fe, Se, and Br in bovine ovaries, as imaged by XRF. Comparison with the adjacent fresh frozen tissue (A) indicates that regions of high Zn (B, pink) intensity correspond to arterioles and capillaries; high Fe (C, pink) correspond to corpora lutea; and high Se (D, pink) is localized to the granulosa cell layers of the two largest healthy follicles. E) H&E-stained section of a bovine ovary containing arterioles alongside Br elemental distribution map (F) generated by XRF imaging. ▽, atretic follicles; *, healthy antral follicles; ---, corpus luteum or regressed follicles; †, capillaries; ◆, arterioles. Scale bars = 4 mm (A–D) and 200 μm (E and F). Reprinted from Ceko et al. *Metallomics* 2015; 7:71–82 and Ceko et al. *Metallomics*, 2015; 7:756–765 [48] and reproduced by permission from The Royal Society of Chemistry.

bromide was more cytotoxic than either sodium hypochlorite or activated sodium hypobromite. Thus, it appears that there is a range of chemical modifications induced by $\text{Br}^-_{(\text{aq})}$ -based species, many of which are still in the process of being fully elucidated.

Br Byproducts and Reproduction

In nature, Br is found mostly as Br^- , bound to metals in the form of inorganic salts [64]. The main natural source of $\text{Br}^-_{(\text{aq})}$ is in sea water, where the average concentration is several orders of magnitude larger than in freshwater systems [65]; however, it is also found naturally in soils [66] and a range of plant- and animal-based foods [65]. Major contribu-

tors to the anthropogenic release of Br into the environment include mining, industrial emissions, disinfectants, flame retardants, and the use of fertilizers and pesticides in agriculture [65]. These processes can significantly increase the concentration of Br in the environment. With such prevalent exposure to Br, coming from natural and anthropogenic sources, it is not surprising that numerous studies have been conducted on the impact of chronic Br exposure on mammalian systems. Throughout the 1980s, several authors confirmed that large doses of Br^- could reduce fertility, impact the rate of survival of offspring, and result in depressed production of thyroxine, because excess $\text{Br}^-_{(\text{aq})}$ competes with I^- in the synthesis of this hormone [67–69]. Flury et al. [65] suggested that chronic Br toxicity affects mainly the endocrine and reproductive systems of mammals, and Pavelka [64] further suggested that high $\text{Br}^-_{(\text{aq})}$ levels can influence iodine metabolism by either decreasing the iodine accumulation in the thyroid and skin and/or by increasing the excretion of iodine by the kidneys. More recent studies have focused less on $\text{Br}^-_{(\text{aq})}$ and more on the potential toxicology of brominated disinfection by-products. Several authors focusing on dibromoacetic acid have concluded that chronic exposure to even low levels of this chemical (1 mg/kg body weight/day) adversely affects reproductive function in male rabbits, and delays pubertal development and compromises sperm quality in rats [70–72].

In humans, most of the research investigating the effects of Br is centered around the possible toxic effects of brominated compounds and residues in food and water supplies [73, 74]. As a member of the halogen family, the propensity for substitution with other halogens (chlorine and iodine in particular) and the advent of newer technologies that identify and quantify the bioaccumulation of Br in tissue, more research is examining the toxic effects of Br exposure on pregnancy outcome. However, to date, there are inconsistent associations with spontaneous abortion, preterm birth, small-for-gestational-age babies, and birth defects [73]. In a study spanning 3 decades, Small et al. [75] investigated the effects of exposure of brominated flame retardants and polybrominated biphenyls (PBBs), a class of halogenated, brominated flame retardants, during pregnancy on the fertility and reproductive outcomes of offspring more than 20 yr after exposure. Of 194 women that were exposed to PBBs while in utero, there was a decrease in fertility and an increased risk of spontaneous abortions with increasing risk associated with greater exposure to PBBs [75]. The finding of an increase in spontaneous abortions suggests that the in utero exposure on reproductive outcomes is likely due to direct exposure to the fetal ovaries at the time when early follicular development is initiated. A hypothesis to account for the findings includes the effect that PBBs have on estrogen activity and progesterone production in in vitro models and animal studies [75]. Concurrent exposure to PBBs in pregnancy has led to spontaneous abortions in cattle [76, 77], but not in humans [75].

To our knowledge, our group was the first to identify and document the presence and nature of distribution of Br in the mammalian ovary [35]. Qualitative observations of the XANES region of the spectra coupled with principal component analyses of all XAS data led us to conclude that the predominant form of Br was aqueous Br^- . Using S-XRF, Br was identified in high concentrations in all sections from 45 bovine ovaries and was widely distributed throughout the ovarian tissue, but heavily concentrated in the walls of arterioles (Fig. 3F). The only known biological function of Br is for cross-linking of type IV collagen [56], which is predominantly a basal lamina component. Cross-linking of subunits would presumably increase the tensile strength of

basal laminas. The basal laminas of subendothelial cells of arterioles, venules, and capillaries and the basal laminas of smooth muscle cells of arterioles in bovine ovaries contain type IV collagens $\alpha 1$ and $\alpha 2$ [78]. Cross-linking of both type IV collagens $\alpha 1$ and $\alpha 2$ has been observed previously in bovine follicles [78]. That Br would be heavily concentrated in the walls of arterioles would suggest that its function is to facilitate substantial cross-linking of type IV collagens in the subendothelial basal laminas and that of the smooth muscles of arterial walls to withstand the high intra-arteriole blood pressures.

In our study [48], we observed tenfold higher concentrations of Br in bovine serum versus normal human serum concentrations, which raises some important questions regarding the dietary and environmental exposure to Br for the cattle. Due to previously determined toxic nature of Br, the origin and source of Br requires further investigation to determine whether it is from anthropogenic or natural sources.

IRON

Fe has the longest and best-described history among all the trace elements; however, it was not until 1932 that the importance of Fe was finally confirmed with the evidence that inorganic Fe was needed for hemoglobin synthesis [79]. It is a key element in the metabolism of almost all living organisms, as it is involved in central cellular processes, including DNA synthesis, respiration, and oxygen transport. The biological functioning of Fe depends upon its ability to readily accept or donate electrons, interconverting between Fe^{3+} and Fe^{2+} forms, as it does in enzymes, such as cytochromes [80]. In the human body, Fe is an essential component of hundreds of proteins and enzymes [7], where it mainly exists in complex forms bound to heme protein (hemoglobin or myoglobin), as heme compounds, as heme enzymes, or as nonheme compounds (flavin-Fe enzymes, transferrin, and ferritin) [81]. In hemoglobin, Fe is in the Fe^{2+} form, whereas Fe transported on transferrin in the blood, or stored intracellularly in ferritin, is in the Fe^{3+} form [80]. An understanding is emerging on how these proteins, both individually and in unison, maintain cellular and whole-body homeostasis of this crucial trace element [82]. Fe hemostasis is a complex process, involving many proteins that respond not only to Fe load (intake and storage), but also to stimuli including inflammation, hypoxia, and anemia [83].

Although low Fe intake and its poor bioavailability are responsible for most anemia in industrialized countries, this accounts for only about half of the causes of anemia in developing countries [84]. In these regions, infectious and inflammatory diseases, such as malaria, blood loss caused by parasitic infections, and nutrient deficiencies, including those for vitamins A, B₂, B₉, and B₁₂, are also important causes [85]. The role of Fe deficiency in detrimentally influencing pregnancy outcomes is well established, and governments and researchers have focused a significant amount of attention on this aspect [86–91]. There has been little research, however, into the relationship between Fe deficiency and unexplained infertility [92].

Metabolism of Fe

The fraction of Fe absorbed relative to the amount ingested is typically quoted in the range of 5%–35%, depending on the combination of foods ingested and the oxidation state of Fe [81]. Fe absorption predominantly takes place in the duodenum and upper jejunum [93], and occurs by the enterocytes by divalent metal transporter 1, a member of the solute carrier group of membrane transport proteins. From here, it is

transferred across the duodenal mucosa into the blood, where it is transported by transferrin to the cells or the bone marrow for red blood cell production [94, 95]. Fe uptake, storage, and export by cells are tightly regulated by Fe regulatory proteins (IRPs) [96]. IRPs are cytosolic *trans* regulators that bind to specific RNA Fe-responsive elements that are present within the mRNA encoding transferrin receptors and ferritin, thus controlling the intracellular flux of Fe [97].

Dietary Fe occurs in both heme and nonheme forms [98]. Primary sources of heme Fe are hemoglobin and myoglobin from consumption of red meat, poultry, and fish, whereas nonheme Fe is obtained from cereals, pulses, legumes, fruits, and vegetables [99]. The difference in bioavailability of the two Fe forms (15%–35% and 2%–20% for heme and non-heme, respectively), in addition to the fact that dietary factors have little effect on absorption of heme Fe, make red meats an excellent nutrient source of this trace element [98]. Major inhibitors of Fe absorption are phytic acid, polyphenols, calcium, and peptides from partially digested proteins [98]. Enhancers include ascorbic acid and muscle tissue, which may reduce Fe^{3+} to Fe^{2+} and bind it in soluble complexes, which are available for absorption [98].

Fe and Reproduction

As previously mentioned, the effect on pregnancy of Fe deficiency in the mother and, more specifically, its negative impact on fetal growth and development, has been the focus of many studies [100]. Pregnancy increases the Fe requirement to nearly 6 mg/day by the second and third trimesters, due to the high growth rates of the placenta and the fetus, and the expansion of the maternal red blood cell mass [101]. Few studies have been published on the effect of Fe on female fertility; however, the preliminary consensus is that a greater intake of Fe, either due to increased dietary intake or supplementation, increases fertility levels in women, with the comment that the utilization by the ovary of this element is the likely reason for this observation [42]. In a large study involving 18 555 women, Chavarro et al. [102] evaluated whether Fe supplement use or greater intake of total heme and nonheme Fe is associated with lower risk of ovulatory infertility. Women who consumed Fe in supplements had a significantly lower risk of ovulatory infertility than women who did not use Fe. More specifically, total nonheme Fe intake, consumed as multivitamins and Fe supplements, was inversely associated with the risk of ovulatory infertility, whereas heme Fe intake was unrelated to ovulatory infertility in multivariable adjusted analyses [102]. A recently published study to ascertain the effects of severe Fe deficiency on fertility in female rats found that there was a significantly lower conception rate and a disruption of estrus in an Fe-deficient group relative to the controls [92]. These results suggest that the importance of Fe in female reproductive function begins well before pregnancy.

There are few studies investigating the effect of Fe overload (toxicity) and fertility, and even less with regard to female fertility. However, there is a link between Fe overload and female infertility [103, 104]. Excess Fe leads to reduced production of the hormones LH and FSH from the anterior pituitary, suggesting impaired oocyte maturation and low ovarian reserve [103]. In patients with beta thalassemia, multiple blood transfusions and increased gastrointestinal Fe absorption leads to Fe overload in the body [105] and infertility [103], likewise in patients with hemochromatosis, a genetic condition that leads to increased serum Fe levels and also sub-fertility in women [104].

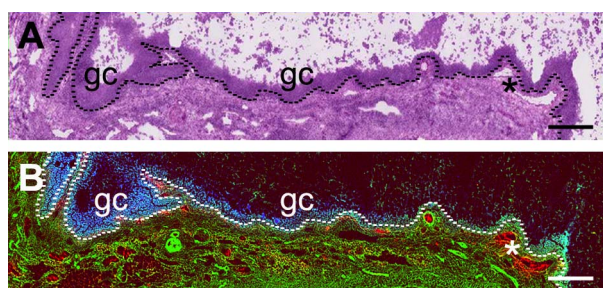


FIG. 4. Localization of Zn, Fe, and Se in a large bovine antral follicle. (a) H&E-stained serial section of a 15-mm-diameter healthy follicle. (b) The corresponding RGB image was generated from XRF elemental distribution maps and depicts the distribution of Zn (green), Fe (red), and Se (blue). *, vasculature; - - - -, the separation between granulosa layer and the thecal interna; gc, granulosa cells. Scale bar = 500 μm . Reprinted from Ceko et al. *Metallomics* 2015; 7:71–82 [47] and reproduced by permission from The Royal Society of Chemistry.

Using S-XRF, we were able to determine the precise location and levels of Fe in bovine ovarian sections [35, 36]. Bioaccumulation of Fe in the ovary is elevated in corpora lutea: low at early stages of development and high in mature corpora lutea. This would be consistent with Fe as a major component of cytochrome P450 cholesterol side-chain cleavage, which is elevated in mature bovine corpora lutea [106], and with the Fe in hemoglobin in the highly vascularized corpora lutea [76]. Fe in capillaries of follicles, likely due to hemoglobin, can be seen in Figure 4. Regressing corpora lutea and regressed follicles (Fig. 3C) are also high in Fe, which probably represents hemosiderin derived from the dead cells that contained the steroidogenic cytochrome P450 enzymes. Fe levels are also elevated in the walls of arterioles, and these levels are five times that of the levels within the lumen of the arterioles [36]. At least one other study has reported high Fe concentrations in blood vessels and in smooth muscle cells imaged by XRF [77]. They hypothesized that the elevated levels are owing to the presence of Fe in smooth muscle myoglobin. In addition to a clear higher density of Fe in the compartments discussed above, Fe is crucial for many basic processes in all ovarian cells, including respiration, DNA synthesis, antioxidant activities, and steroidogenesis, which is representative in the more dispersive distribution shown in Figure 4.

SELENIUM

The trace element Se was discovered in 1817 by the Swedish physician and chemist Berzelius; however, its essentiality in mammals was not discovered until the 1950s [107]. Conclusive evidence for the essentiality of Se in humans came with publication of the results of large-scale trials in China that showed the protective effect of Se supplementation on children and young adults suffering from Keshan disease, a cardiomyopathy endemic in regions with low soil Se levels [108]. Whole-body Se is about 15 mg, as estimated by direct tissue analysis and radioisotope techniques, with the tissue concentrations of this essential trace element being highest in the kidney and liver [109]. The most important biologically active compounds contain selenocysteine, where Se is substituted for sulfur in cysteine. The insertion of selenocysteine to form a selenoprotein is specified by the UGA codon in mRNA under specific conditions [110, 111].

In humans, the nutritional functions of Se are achieved by its incorporation into 25 selenoproteins (including the glutathione peroxidases [GPXs], thioredoxin reductases and deiodinases) that have selenocysteine at their active center [110].

These selenoproteins play an important role in many biological functions, such as antioxidant defense, formation of thyroid hormones, DNA synthesis, fertility, and reproduction [112]. By working synergistically with vitamin E, GPXs play an important role in the body, catalyzing the conversion of peroxides to nontoxic alcohols and thus protecting cells from membrane damage and oxidative stress [113, 114]. The concentrations of Se in whole blood and in plasma and/or serum are related to dietary intake, with 50%–60% of the total plasma Se being present as selenoprotein P [108, 109], about 30% being present as GPX3, and the remainder incorporated into albumin as selenomethionine [115]. Low serum Se levels have been linked to higher cancer risk based on evidence from observational studies conducted over a 40-yr period (see reviews [116–119]). Thus, adequate Se intake may have an anticancer effect, due to its contribution to antioxidant function or to immune system function, but the precise mechanism is uncertain. Weekley and Harris [117] highlight that epidemiological studies have reported an inverse relationship between Se status and the incidence of various diseases, but subsequent studies of Se supplementation and disease prevention have presented mixed results, with concerns about this trace element's toxicity [117].

Bioavailability and Absorption of Se

The Se content of food is dependent upon the Se content of the soil in which the plants are grown or the animals are raised. Se is found in highest amounts in organ meats, such as kidney and liver, while some seafood contains nearly as much [120]. That said, plant foods, such as grains, legumes, and cruciferous vegetables, are a major dietary sources of Se in most countries, where it enters the food chain predominantly as selenomethionine [109, 121]. Se, which is initially taken up from the soil and concentrated by plants, is absorbed in the small intestine and incorporated into proteins by complex mechanisms that remain to be fully elucidated [122]. Preintestinal absorption of Se is negligible, so the absorption operates mainly in the duodenum and caecum [112]. Similar to Fe, the mechanisms of intestinal absorption are different depending on the chemical form of the element. A recent study designed to investigate the bioavailability of four Se species using an in vitro model of the intestinal barrier determined that the efficiency of Se absorption decreased in the following order: selenomethionine, methylselenocysteine, selenate (SeO_4^{2-}), and selenite (SeO_3^{2-}) [123]. An earlier report presented by the U.S. Institute of Medicine, however, summarized that Se from the inorganic salts, SeO_4^{2-} and SeO_3^{2-} , is more rapidly incorporated into GPX and other selenoproteins than Se from organic sources containing selenomethionine. SeO_3^{2-} is absorbed by simple diffusion, and its subsequent reduction is a well-characterized metabolic pathway in animals [124]. The proportion of ingested Se excreted is dependent upon dietary intake, with high consumption resulting in high urinary and fecal excretion and vice versa [125, 126]. In cases of dietary Se deprivation, the synthesis of some selenoproteins (e.g., GPX4) is prioritized over that of others [111]. Conversely, GPX1 is one of the most highly sensitive selenoproteins to changes in Se status, with levels of mRNA and protein dramatically reduced under low Se conditions [127].

Se and Reproduction

Se has long been recognized in animal husbandry as being essential for successful reproduction [2]. Of particular importance to reproduction and pregnancy are the GPXs,

which play a crucial role in reducing hydrogen peroxide (H_2O_2) and lipid peroxides to harmless products, thereby reducing cellular damage by reactive oxygen species (ROS) [128]. The emerging role of Se in maintaining healthy reproductive function has been extensively studied in males, but data pertaining to Se status and fertility in females are sparse, with the majority of female-based studies having a tendency to focus on the role of this element in pregnancy [129–133].

Over 30 yr ago, Behne et al. conducted several studies in male rats and observed that, during insufficient Se intake, the supply of this trace element to the testes was prioritized over other tissues. This suite of studies led the authors to postulate the involvement of Se in the biosynthesis of testosterone [134, 135], and it is now commonly known that testicular tissue contains high concentrations of Se, predominantly as GPX4, and that this element is essential for the formation and normal development of spermatozoa [136, 137]. Some evidence suggests that increasing Se dietary intake increases antioxidant GPX activity, thereby increasing male fertility [138]. In a study of subfertile Norwegian men, the Se concentration of seminal plasma correlated positively with the concentration of spermatozoa [139]. Scott et al. [140] supplemented subfertile men with 100 μ g of Se per day for 3 mo and, in addition to their sperm motility significantly increasing, 11% of men receiving the active supplement fathered a child compared with none in the placebo group. It should be noted, however, that a similar study carried out over a similar length of time, but with double the daily dose of Se, showed no beneficial effect on sperm motility in subfertile Polish men [141]. Closer observation of the form of Se administered in each of these studies indicates that they differed, with the first being selenomethionine versus SeO_3^{2-} in the latter Polish study. This highlights that it is not necessarily the quantity of the trace element ingested that matters with regard to exerting a beneficial biological effect, but more so its bioavailability and subsequent absorption by the body.

With regard to Se status and fertility in females, Paszkowski et al. [142] completed a study of 135 follicular fluid samples collected from patients during transvaginal oocyte retrieval, and found that patients with unexplained infertility had significantly decreased follicular Se concentrations compared with those with other known causes of infertility [142]. A case-controlled study from Turkey found lower Se concentrations in the serum and follicular fluid of women undergoing in vitro fertilization treatment compared with age-matched, nonpregnant control women [143], and a recent study, also from Turkey, found that plasma Se levels were significantly lower in women with polycystic ovary syndrome relative to the control group, indicating that this element may play a role in the pathogenesis of polycystic ovary syndrome related to hyperandrogenism [144]. Finally, women presenting with unexplained infertility or premature ovarian failure were found to have significantly increased serum levels of the ovarian autoantibody protein, Se-binding protein-1 [145]. Ultimately, Se plays a significant role in living organisms, and owing to its contribution to the formation of selenoproteins combined with its antioxidant action, a low Se status in the body induces a low resistance to free radical damage [112].

Our investigations into the bioaccumulation of Se in ovarian tissue using S-XRF demonstrated that Se was consistently localized to the granulosa cell layer of large, healthy follicles (Fig. 4) [36]. In addition, the increase in Se in these follicles was likely due to the increased expression of selenoprotein GPX1, which was significantly upregulated compared to small healthy or atretic follicles. Western blot analyses also

confirmed a strong association between GPX1 and large, healthy follicles, suggesting a role for this antioxidant in follicle dominance, protecting the dominant follicle from increasing levels of ROS [47]. Furthermore, an additional study using human cumulus cells derived from cumulus-oocyte complexes collected for both in vitro fertilization and intracytoplasmic sperm injection were assessed using RT-PCR for GPX1 expression before fertilization [47]. These cumulus cells retrieved from cumulus-oocyte complexes prior to embryo transfers that resulted in a pregnancy ($n = 12$) had significantly higher expression of GPX1 compared with those that did not result in a pregnancy ($n = 18$) [47]. Notwithstanding the fact that much further research is needed, these observations do suggest a strong role for selenoprotein GPX1 in determining follicle growth, maturation, and dominance in both the cow and in women, and lead to further specific dietary recommendations for Se and the role oxidative stress may play in infertility.

ZINC

Zn is an essential element in the nutrition of human beings, other animal species, and plants, and is required in every cell [7]. Zn plays an important role in growth and development, the immune response, neurological function, and reproduction [146, 147]. Zn is needed for DNA and RNA synthesis and is required at every step of the cell cycle, including proliferation, differentiation, and apoptosis [147]. Zn plays an important role in the structure of proteins and cell membranes, and is an integral component of over 300 metalloenzymes, including carbonic anhydrase, alcohol dehydrogenase, thymidine kinase, carboxypeptidase, glutamate dehydrogenase, lactate dehydrogenase, and alkaline phosphatase [5, 148]. Zn additionally participates in the synthesis and degradation of carbohydrates, lipids, proteins, and nucleic acids [3]. Furthermore, Zn finger proteins, formed when Zn atoms become tetrahedrally coordinated with histidine and cysteine [149], regulate the expression of many genes [150, 151], and Zn has been shown to have antiapoptotic and antioxidant properties [152–154]. Inadequate intake of this trace element in humans and other animal species results in immunodeficiency, increased numbers of infections, increased severity of infections, stunted growth, and delayed sexual maturation, as summarized by Qureshi et al. [7].

Bioavailability and Absorption of Zinc

Zn is found as organic complexes with protein in meats and as inorganic salts in plant foods [155, 156]; however, no absorption studies have been conducted in humans to determine whether these forms of Zn differ in their bioavailability [157]. Based on limited published data on the absorption of Zn from supplements ingested by humans, it appears that Zn gluconate, Zn citrate, and Zn sulfate are absorbed to a similar extent, and that Zn oxide is slightly less well absorbed [158, 159]. A recent study in rats, however, found all four of these species to be equally bioavailable [160]. Similar to the aforementioned trace elements (Cu, Fe, and Se), inadequate dietary intake of Zn can arise from low intake, poor bioavailability, or a combination of these dietary factors. Low dietary Zn intake is often associated with developing countries, owing to the prohibitive cost of Zn-rich foods (i.e., meat, poultry, and fish) or religious restrictions [161]. The total Zn content of one's diet is influenced, not only by the range of food items selected and their relative bioavailability, but also by the degree of refinement of any constituent cereals. In

addition, a diet rich in saturated or unsaturated fats tends to dilute the uptake of Zn [3].

Zn is absorbed in the small intestine by a carrier-mediated mechanism [162], with ongoing research over the last decade indicating that its transport across membranes is mediated by two subfamilies of mammalian Zn transporters, ZnT (Slc30) and Zip (Slc39) [163–166]. Following absorption through the enterocyte, dietary Zn is transported into the circulation and then to the tissues, where it is needed. Active transport of Zn into portal blood is mediated by metallothionein [167]. Zn is released from food as free ions during digestion and the liberated ions may then bind to endogenously secreted ligands before their transport into the enterocytes in the duodenum and jejunum [16, 168]. In contrast to Fe, the body regulates Zn homeostasis through gastrointestinal secretion and excretion of endogenous Zn, in addition to the absorption of exogenous Zn [169, 170]. Loss of Zn through the gastrointestinal tract accounts for approximately half of all Zn eliminated from the body, with pancreatic and biliary pathways being other modes of excretion. This is an important process in the regulation of Zn balance [171], and similar to Cu and Se, with higher Zn intake balanced by more secretion and vice versa [172].

As alluded to earlier, plant-based foods are the major source of dietary Zn in many low-income countries. Of the plant-based foods, cereals and legumes contain high levels of phytic acid and their magnesium, calcium, and potassium-associated salts, called phytates. While amino acids and peptides in chyme facilitate the absorption of Zn, the binding of Zn by phytate and dietary fibers forms insoluble complexes, rendering the Zn relatively unavailable [173, 174].

Zinc and Reproduction

Countless studies have focused on the role of Zn in male fertility, paying particular attention to its role in male sex hormone synthesis, as well as the resultant impact on sperm production and motility. Zn content is high in the adult testis, and the prostate has a higher concentration of Zn than any other organ of the body [1]. Zn deficiency first impairs angiotensin-converting enzyme activity, which leads to depletion of testosterone and inhibition of spermatogenesis. Defects in spermatozoa are frequently observed in Zn-deficient rats [175, 176]. In females, Zn also seems to be important in reproduction, but relatively few investigations have been performed [45]. Shaw et al. [177] found that Zn deficiency in female rabbits resulted in sexual disinterest by their male counterparts. Another study suggested that Zn deficiency in female rats led to abnormal estrous cycles [178], and the effects of Zn deficiency in two species of monkeys led the authors to conclude that normal reproduction was impaired in both the species through abnormal ovarian development [179]. Over 30 yr ago, Sato et al. [180] reported that even marginal Zn deficiency could affect oocyte maturation by doubling the number of degenerating oocytes and increasing chromosomal abnormalities in metaphase II oocytes. Despite the authors suggesting that preconception was a crucial time for women to ensure a sufficient intake of Zn, it would appear that attention in subsequent decades has shifted to Fe, folate, and, in more recent years, iodine supplementation [181–186]. In studies designed to ascertain the reproductive effects of feeding virgin female mice a Zn-deficient diet, Taneja and Kaur observed retardation of ovarian follicular growth with varying degrees of atresia, lack of preovulatory follicles, a reduced and shrunken corpus luteum, and a fragmented zona pellucida and vitelline membrane, indicating the cessation of oogenesis and ovulation [187, 188].

Kim et al. investigated the abundance of Zn throughout the ovarian cycle in mice using S-XRF [189]. Interestingly, during the period of meiotic maturation, between the LH surge and ovulation, there was a 50% increase in total Zn within the ovary [189]. Moreover, within the unfertilized egg, there is a polar distribution of Zn, which the authors suggest are Zn-enriched cortical granules that may play a role in the hardening of the zona pellucida required to block polyspermy at fertilization [189]. In a more recent study, Tian et al. [190] demonstrated that Zn deficiency in mice before fertilization led to reductions in placental and fetal development, highlighting the role of trace elements and the dependency that the embryo has on the integrity of the oocyte predecessor. Taken together, these studies demonstrate that Zn has a vital role in ovarian function, and cytoplasmic levels within the oocyte are crucial for early embryo and placental development [189, 190].

Similar to the alleged role of Se in reproduction, Zn is important for several antioxidant functions that provide protection of cells against oxidative and electrophilic stress caused by ROS. Many investigations have shown evidence for the role of ROS in the physiology and pathology of both male and female reproductive functions [191, 192], suggesting that the focus should shift to elucidating the biochemical roles these trace metals play in the process.

Our investigations using S-XRF into the accumulation of Zn in bovine ovarian tissue indicate that the highest Zn levels always corresponded to either the walls of arterioles or to capillaries (Figs. 3 and 4), and was associated with above-background Fe concentrations [36]. Although at lower levels than blood vessels, Zn was also elevated in the thecal and granulosa cell layers relative to the stromal tissue. Healthy follicles contain an average of three times more Zn than the regressed follicles. Using principal component analyses, a relationship was found to exist between levels of Br and Zn, showing that the healthy and early atretic follicles are different from atretic and regressed follicles, suggesting that the roles of these elements in healthy follicles may differ between growth and atresia [35].

FUTURE DIRECTIONS

The recent new capabilities to both localize and quantitate trace elements in tissues will allow us to identify where and when trace elements accumulate in reproductive tissues during their development and in disease states. Just as the recent studies in the ovary have demonstrated, this may lead to discoveries of new roles played by these elements in reproduction.

ACKNOWLEDGMENT

We thank the Australian Synchrotron, Victoria, Australia for their support and for the use of their facilities.

REFERENCES

1. Bedwal RS, Bahuguna A. Zinc, copper and selenium in reproduction. *Experientia* 1994; 50:626–640.
2. Underwood EJ (ed.). *Trace Elements in Human and Animal Nutrition*. New York: Academic Press, Inc.; 1977.
3. WHO. *Trace elements in human nutrition and health*. Geneva: World Health Organization 1996.
4. Bogden JD, Klevay LM. *Clinical Nutrition of the Essential Trace Elements and Minerals: The Guide for Health Professionals*. Totowa, NJ: Humana Press; 2000.
5. Burtis CA, Ashwood ER, Bruns DE. *Vitamins and Trace Elements*. St. Louis, MO: Elsevier Health Sciences; 2006.
6. Mertz W. The essential trace elements. *Science* 1981; 213:1332–1338.
7. Ali Qureshi GA, Memon SA, Memon AB, Ghouri RA, Memon JM,

- Parvez SH. The emerging role of iron, zinc, copper, magnesium and selenium and oxidative stress in health and diseases. *Biogenic Amines* 2005; 19:147–169.
8. Mertz W. Review of the scientific basis for establishing the essentiality of trace elements. *Biol Trace Elem Res* 1998; 66:185–191.
 9. Dendougui F, Schwedt G. Use of an ion selective electrode to determine the complexing of copper in food extracts dependent upon the pH. *Eur Food Res Technol* 2002; 215:76–82.
 10. Hackbart KS, Ferreira RM, Dietsche AA, Socha MT, Shaver RD, Wiltbank MC, Fricke PM. Effect of dietary organic zinc, manganese, copper, and cobalt supplementation on milk production, follicular growth, embryo quality, and tissue mineral concentrations in dairy cows. *J Anim Sci* 2010; 88:3856–3870.
 11. Bolann BJ, Rahil-Khazen R, Henriksen H, Isrenn R, Ulvik RJ. Evaluation of methods for trace-element determination with emphasis on their usability in the clinical routine laboratory. *Scand J Clin Lab Invest* 2007; 67:353–366.
 12. Pheodorin MA, Bobrov VA, Chebykin EP, Goldberg EL, Melgunov MS, Filippova SV, Zolotarev KV. Comparison of synchrotron radiation X-ray fluorescence with conventional techniques for the analysis of sedimentary samples. *Geostandards Newslett* 2000; 24:205–216.
 13. Weekley CM, Aitken JB, Vogt S, Finney LA, Paterson DJ, de Jonge MD, Howard DL, Musgrave IF, Harris HH. Uptake, distribution, and speciation of selenoamino acids by human cancer cells: X-ray absorption and fluorescence methods. *Biochemistry* 2011; 50:1641–1650.
 14. Janssens K, De Nolf W, Van Der Snickt G, Vincze L, Vekemans B, Terzano R, Brenker FE. Recent trends in quantitative aspects of microscopic X-ray fluorescence analysis. *Trends Analyt Chem* 2010; 29:464–478.
 15. Ortega R, Deves G, Carmona A. Bio-metals imaging and speciation in cells using proton and synchrotron radiation X-ray microspectroscopy. *J R Soc Interface* 2009; 6(suppl):S649–S658.
 16. Tubek S. Selected zinc metabolism parameters in premenopausal and postmenopausal women with moderate and severe primary arterial hypertension. *Biol Trace Elem Res* 2007; 116:249–256.
 17. Eichert D, Gregoratti L, Kaulich B, Marcello A, Melpignano P, Quaroni L, Kiskinova M. Imaging with spectroscopic micro-analysis using synchrotron radiation. *Anal Bioanal Chem* 2007; 389:1121–1132.
 18. James SA, Myers DE, de Jonge MD, Vogt S, Ryan CG, Sexton BA, Hoobin P, Paterson D, Howard DL, Mayo SC, Altissimo M, Moorhead GF, et al. Quantitative comparison of preparation methodologies for x-ray fluorescence microscopy of brain tissue. *Anal Bioanal Chem* 2011; 401:853–864.
 19. Carvalho ML, Casaca C, Marques JP, Pinheiro T, Cunha AS. Human teeth elemental profiles measured by synchrotron x-ray fluorescence: dietary habits and environmental influence. *Xray Spectrom* 2001; 30:190–193.
 20. Harris HH, Vogt S, Eastgate H, Lay PA. A link between copper and dental caries in human teeth identified by X-ray fluorescence elemental mapping. *J Biol Inorg Chem* 2008; 13:303–306.
 21. Harris HH, Vogt S, Eastgate H, Legnini DG, Hornberger B, Cai Z, Lai B, Lay PA. Migration of mercury from dental amalgam through human teeth. *J Synchrotron Radiat* 2008; 15:123–128.
 22. Zhang YX, Cheng F, Li DY, Wang YS, Zhang GL, Xu HJ, Liao WS, Tang TT, Huang YY, He W. Synchrotron radiation XRF microprobe investigation of elemental distribution in femoral head slice with osteoporosis. *Chin Sci Bull* 2001; 46:1138–1141.
 23. Senda J, Hashiguchi N, Tanaka Y, Komiyama S, Shimahara M, Kono K, Watanabe T, Dote T, Usuda K. Determination of bone calcium and phosphorus in osteoporosis model rats by X-ray fluorescent analysis. *Trace Elem Electrolyt* 2001; 18:55–58.
 24. Le Naour F, Sandt C, Peng CY, Trcera N, Chiappini F, Flank AM, Guettier C, Dumas P. In situ chemical composition analysis of cirrhosis by combining synchrotron fourier transform infrared and synchrotron X-ray fluorescence microspectroscopies on the same tissue section. *Anal Chem* 2012; 84:10260–10266.
 25. Ide-Ektessabi A, Fujisawa S, Sugimura K, Kitamura Y, Gotoh A. Quantitative analysis of zinc in prostate cancer tissues using synchrotron radiation microbeams. *Xray Spectrom* 2002; 31:7–11.
 26. Ortega R, Deves G, Bonnin-Mosbah M, Salome M, Susini J, Anderson LM, Kasprzak KS. Chromium mapping in male mice reproductive glands exposed to CrCl₃ using proton and X-ray synchrotron radiation microbeams. *Nucl Instrum Meth B* 2001; 181:485–488.
 27. De Samber B, Silversmit G, De Schampelaere K, Evens R, Schoonjans T, Vekemans B, Janssen C, Masschaele B, Van Hoorebeke L, Szaloki I, Vanhaecke F, Rickers K, et al. Element-to-tissue correlation in biological samples determined by three-dimensional X-ray imaging methods. *J Anal At Spectrom* 2010; 25:544–553.
 28. Evens R, De Schampelaere KA, De Samber B, Silversmit G, Schoonjans T, Vekemans B, Balcaen L, Vanhaecke F, Szaloki I, Rickers K, Falkenberg G, Vincze L, et al. Waterborne versus dietary zinc accumulation and toxicity in *Daphnia magna*: a synchrotron radiation based X-ray fluorescence imaging approach. *Environ Sci Technol* 2012; 46:1178–1184.
 29. Piacenti da Silva M, Mara da Silva D, Ribeiro-Silva A, Poletti ME. Correlations of trace elements in breast human tissues: evaluation of spatial distribution using mu-XRF. *AIP Conf Proc* 2012; 1437:45–49.
 30. Al-Ebraheem A, Goettlicher J, Geraki K, Ralph S, Farquharson MJ. The determination of zinc, copper and iron oxidation state in invasive ductal carcinoma of breast tissue and normal surrounding tissue using XANES. *Xray Spectrom* 2010; 39:332–337.
 31. Al-Ebraheem A, Geraki K, Leek R, Harris AL, Farquharson MJ. The use of bio-metal concentrations correlated with clinical prognostic factors to assess human breast tissues. *Xray Spectrom* 2013; 42:330–336.
 32. Farquharson MJ, Geraki K, Falkenberg G, Leek R, Harris A. The localisation and micro-mapping of copper and other trace elements in breast tumours using a synchrotron micro-XRF system. *Appl Radiat Isot* 2007; 65:183–188.
 33. Geraki K, Farquharson MJ, Bradley DA, Gundogdu O, Falkenberg G. The localisation of biologically important metals in soft and calcified tissues using a synchrotron X-ray fluorescence technique. *Xray Spectrom* 2008; 37:12–20.
 34. Pushie MJ, Pickering IJ, Korbas M, Hackett MJ, George GN. Elemental and chemically specific X-ray fluorescence imaging of biological systems. *Chem Rev* 2014; 114:8499–8541.
 35. Ceko MJ, Hummitzsch K, Hatzirodos N, Rodgers RJ, Harris HH. Quantitative elemental analysis of bovine ovarian follicles using X-ray fluorescence imaging. *Metallomics* 2015; 7:828–836.
 36. Ceko MJ, Hummitzsch K, Bonner WM, Aitken JB, Spiers KM, Rodgers RJ, Harris HH. Localization of the trace elements iron, zinc and selenium in relation to anatomical structures in bovine ovaries by X-ray fluorescence imaging. *Microsc Microanal* 2015; 21:695–705.
 37. Henderson GS, de Groot FMF, Moulton BJA. X-ray Absorption Near-Edge Structure (XANES) Spectroscopy In: Henderson GS, Neuville DR, Downs RT (eds.), *Spectroscopic Methods in Mineralogy and Materials Sciences*, vol. 78. Chantilly: Mineralogical Soc Amer; 2014; 75–138.
 38. Limpjumnong S, Rujirawat S, Boonchun A, Smith MF, Cherdhirunkorn B. Identification of Mn site in Pb(Zr,Ti)O₃ by synchrotron X-ray absorption near-edge structure: theory and experiment. *Appl Phys Lett* 2007; 90:103–113.
 39. Koningsberger D, Prins R. X-ray absorption: principles, applications, techniques of EXAFS, SEXAFS, and XANES. Hoboken, NJ: Wiley-Interscience; 1988:3–4.
 40. Newville M. *Fundamentals of XAFS*. Chicago: University of Chicago; 2004.
 41. Heitland P, Köster HD. Biomonitoring of 30 trace elements in urine of children and adults by ICP-MS. *Clin Chim Acta* 2006; 365:310–318.
 42. Buhling KJ, Grajecki D. The effect of micronutrient supplements on female fertility. *Curr Opin Obstet Gynecol* 2013; 25:173–180.
 43. Xu X, Cao Z, Chen X. Study of the interaction between serum trace elements and reproductive hormone in menstrual cycle. *J Xi'an Med Univ* 1997; 18:455–458.
 44. Pathak P, Kapil U. Role of trace elements zinc, copper and magnesium during pregnancy and its outcome. *Indian J Pediatr* 2004; 71:1003–1005.
 45. Ebisch IMW, Thomas CMG, Peters WHM, Braat DDM, Steegers-Theunissen RPM. The importance of folate, zinc and antioxidants in the pathogenesis and prevention of subfertility. *Hum Reprod Update* 2007; 13:163–174.
 46. Cetin I, Berti C, Calabrese S. Role of micronutrients in the periconceptional period. *Hum Reprod Update* 2010; 16:80–95.
 47. Ceko MJ, Hummitzsch K, Hatzirodos N, Bonner WM, Aitken JB, Russell DL, Lane M, Rodgers RJ, Harris HH. X-Ray fluorescence imaging and other analyses identify selenium and GPX1 as important in female reproductive function. *Metallomics* 2015; 7:71–82.
 48. Ceko MJ, Hummitzsch K, Hatzirodos N, Bonner W, James SA, Kirby JK, Rodgers RJ, Harris HH. Distribution and speciation of bromine in mammalian tissue and fluids by X-ray fluorescence imaging and X-ray absorption spectroscopy. *Metallomics* 2015; 7:756–765.
 49. Leach RM Jr, Nesheim MC. Studies on chloride deficiency in chicks. *J Nutr* 1963; 81:193–199.
 50. Anke M, Regius A, Groppe B, Arnold W. Essentiality of the trace element bromine. *Acta Agron Hung* 1990; 39:297–304.
 51. Zhavoronkov AA, Kakturskii LV, Anke M, Groppe B, Mikhaleva LM.

- Pathology of congenital bromine deficit (experimental observation) [article in Russian]. *Arkh Patol* 1996; 58:62–67.
52. Nielsen FH. Ultratrace elements in nutrition: current knowledge and speculation. *J Trace Elem Exp Med* 1998; 11:251–274.
 53. Hellerstein S, Kaiser C, Darrow DD, Darrow DC. Distribution of bromide and chloride in the body. *J Clin Invest* 1960; 39:282–287.
 54. Pavelka S, Babický A, Vobecký M, Lener J. Effect of high dose of bromide on iodine metabolism in the rat. In: *Industrial Toxicology '99 Bratislava, Slovakia: Slovak Technical University; 1999:224–228.*
 55. Vobecky M, Babicky A, Pavelka S, Lener J. Determination of bromine and iodine in the rat thyroid by short-term INAA. *J Trace Microprobe T* 2000; 18:467–473.
 56. McCall AS, Cummings CF, Bhawe G, Vanacore R, Page-McCaw A, Hudson BG. Bromine is an essential trace element for assembly of collagen IV scaffolds in tissue development and architecture. *Cell* 2014; 157:1380–1392.
 57. Hawkins CL, Brown BE, Davies MJ. Hypochlorite- and hypobromite-mediated radical formation and its role in cell lysis. *Arch Biochem Biophys* 2001; 395:137–145.
 58. Pattison DI, Davies MJ. Kinetic analysis of the reactions of hypobromous acid with protein components: Implications for cellular damage and use of 3-bromotyrosine as a marker of oxidative stress. *Biochemistry* 2004; 43:4799–4809.
 59. Hawkins CL, Davies MJ. The role of reactive N-bromo species and radical intermediates in hypobromous acid-induced protein oxidation. *Free Radic Biol Med* 2005; 39:900–912.
 60. Skaff O, Pattison DI, Davies MJ. Kinetics of hypobromous acid-mediated oxidation of lipid components and antioxidants. *Chem Res Toxicol* 2007; 20:1980–1988.
 61. Justino GC, Rodrigues M, Florencio MH, Mira L. Structure and antioxidant activity of brominated flavonols and flavanones. *J Mass Spectrom* 2009; 44:1459–1468.
 62. Marcinkiewicz J, Mak M, Bobek M, Biedron R, Bialecka A, Koprowski M, Kontny E, Maslinski W. Is there a role of taurine bromamine in inflammation? Interactive effects with nitrite and hydrogen peroxide. *Inflamm Res* 2005; 54:42–49.
 63. Maines J, Khurana NR, Roman K, Knaup D, Ahmad M. Cytotoxic effects of activated bromine on human fetal osteoblasts in vitro. *J Endoc* 2006; 32:886–889.
 64. Pavelka S. Metabolism of bromide and its interference with the metabolism of iodine. *Physiol Res* 2004; 53:S81–S90.
 65. Flury M, Papritz A. Bromide in the natural environment—occurrence and toxicity. *J Environ Qual* 1993; 22:747–758.
 66. Kabata A, Pendias H. *Trace Elements in Soils and Plants*. New York: CRC Press; 2001.
 67. Owens LB, Vankeuren RW, Edwards WM. Groundwater quality changes resulting from a surface bromide application to a pasture. *J Environ Qual* 1985; 14:543–548.
 68. Vanleeuwen FXR, Dentonkelaar EM, Vanlogten MJ. Toxicity of sodium bromide in rats: effects on endocrine system and reproduction. *Food Chem Toxicol* 1983; 21:383–389.
 69. Loeber JG, Franken MAM, Vanleeuwen FXR. Effect of sodium bromide on endocrine parameters in the rat as studied by immunocytochemistry and radioimmunoassay. *Food Chem Toxicol* 1983; 21:391–404.
 70. Veeramachaneni DNR, Palmer JS, Klinefelter GR. Chronic exposure to low levels of dibromoacetic acid, a water disinfection by-product, adversely affects reproductive function in male rabbits. *J Androl* 2007; 28:565–577.
 71. Linder RE, Klinefelter GR, Strader LF, Narotsky MG, Suarez JD, Roberts NL, Perreault SD. Dibromoacetic acid affects reproductive competence and sperm quality in the male rat. *Fundam Appl Toxicol* 1995; 28:9–17.
 72. Klinefelter GR, Strader LF, Suarez JD, Roberts NL, Goldman JM, Murr AS. Continuous exposure to dibromoacetic acid delays pubertal development and compromises sperm quality in the rat. *Toxicol Sci* 2004; 81:419–429.
 73. Grellier J, Bennett J, Patellarou E, Smith RB, Toledano MB, Rushton L, Briggs DJ, Nieuwenhuijsen MJ. Exposure to disinfection by-products, fetal growth, and prematurity: a systematic review and meta-analysis. *Epidemiology* 2010; 21:300–313.
 74. Nieuwenhuijsen M, Dadvand P, Grellier J, Martinez D, Vrijheid M. Environmental risk factors of pregnancy outcomes: a summary of recent meta-analyses of epidemiological studies. *Environ Health* 2013; 12:6.
 75. Small CM, Murray D, Terrell MI, Marcus M. Reproductive outcomes among women exposed to a brominated flame retardant in utero. *Arch Environ Occup Health* 2011; 66:201–208.
 76. O'Shea JD, Rodgers RJ, D'Occhio MJ. Cellular composition of the cyclic corpus luteum of the cow. *J Reprod Fertil* 1989; 85:483–487.
 77. Gajda M, Banaś K, Banaś A, Jawień J, Mateuszuk Ł, Chlopicki S, Kwiatek WM, Cichoński T, Falkenberg G. Distribution of selected elements in atherosclerotic plaques of apoE/LDLR-double knockout mice assessed by synchrotron radiation-induced micro-XRF spectrometry. *Xray Spectrom* 2011; 37:495–502.
 78. Rodgers HF, Irvine CM, van Wezel IL, Lavranos TC, Luck MR, Sado Y, Ninomiya Y, Rodgers RJ. Distribution of the alpha1 to alpha6 chains of type IV collagen in bovine follicles. *Biol Reprod* 1998; 59:1334–1341.
 79. Yip R, Dallman P (eds.). *Iron*. Washington, DC: ILSI Press; 1996.
 80. Linder MC. Mobilization of stored iron in mammals: a review. *Nutrients* 2013; 5:4022–4050.
 81. McDowell LR. *Minerals in Animal and Human Nutrition*. Amsterdam: Elsevier Science BV; 2003.
 82. Polla AS, Polla LL, Polla BS. Iron as the malignant spirit in successful ageing. *Ageing Res Rev* 2003; 2:25–37.
 83. Jomova K, Valko M. Advances in metal-induced oxidative stress and human disease. *Toxicology* 2011; 283:65–87.
 84. WHO. Guidelines on food fortification with micronutrients In: Allen LH, de Benoist B, Dary O, Hurrell R (eds.), *Guidelines on Food Fortification with Micronutrients*. Geneva: World Health Organization, Department of Nutrition for Health and Development; 2006.
 85. Brabin BJ, Premji Z, Verhoeff F. An analysis of anemia and child mortality. *J Nutr* 2001; 131:636S–645S.
 86. Brabin L, Brabin BJ, Gies S. Influence of iron status on risk of maternal or neonatal infection and on neonatal mortality with an emphasis on developing countries. *Nutr Rev* 2013; 71:528–540.
 87. Grant S, Ford P, Keck G. Iron deficiency anemia: an important yet underrecognized and unmet need in pregnancy. *J Womens Health* 2014; 23:7–7.
 88. Ma AG, Schouten EG, Sun YY, Yang F, Han XX, Zhang FZ, Jiang DC, Kok FJ. Supplementation of iron alone and combined with vitamins improves haematological status, erythrocyte membrane fluidity and oxidative stress in anaemic pregnant women. *Br J Nutr* 2010; 104:1655–1661.
 89. Sangare L, van Eijk AM, ter Kuile FO, Walson J, Stergachis A. The Association between Malaria and Iron Status or Supplementation in Pregnancy: A Systematic Review and Meta-Analysis. *Plos One* 2014; 9:e87743.
 90. Souza de Camargo RM, Pereira RA, Yokoo EM, Schirmer J. Factors associated with iron deficiency in pregnant women seen at a public prenatal care service. *Rev Nutr* 2013; 26:455–464.
 91. Yogender P, Sujatha R, Rangaswamy R, Sreekantha, Avinash SS. The study of iron related parameters in iron deficiency anaemia in pregnancy. *Res J Pharm Biol Chem Sci* 2014; 5:980–992.
 92. Li YQ, Cao XX, Bai B, Zhang JN, Wang MQ, Zhang YH. Severe iron deficiency is associated with a reduced conception rate in female rats. *Gynecol Obst Invest* 2014; 77:19–23.
 93. Muir A, Hopfer U. Regional specificity of iron uptake by small intestinal brush-border membranes from normal and iron-deficient mice. *Am J Physiol* 1985; 248:G376–G379.
 94. Frazer DM, Anderson GJ. Iron imports. I. Intestinal iron absorption and its regulation. *Am J Physiol Gastrointest Liver Physiol* 2005; 289:G631–G635.
 95. Nadadur S, Srirama K, Mudipalli A. Iron transport & homeostasis mechanisms: their role in health & disease. *Indian J Med Res* 2008; 128:533–544.
 96. Eisenstein RS, Blemings KP. Iron regulatory proteins, iron responsive elements and iron homeostasis. *J Nutr* 1998; 128:2295–2298.
 97. Anderson CP, Shen M, Eisenstein RS, Leibold EA. Mammalian iron metabolism and its control by iron regulatory proteins. *Biochim Biophys Acta* 2012; 1823:1468–1483.
 98. Hurrell R, Egli I. Iron bioavailability and dietary reference values. *Am J Clin Nutr* 2010; 91:1461S–1467S.
 99. FAO, WHO. *Human Vitamin and Mineral Requirements In: Food-Based Approaches to Meeting Vitamin and Mineral Needs*. Rome: Food and Agricultural Organization of the United Nations World Health Organization; 2001:7–25.
 100. Beard JL. Iron-deficiency—assessment during pregnancy and its importance in pregnant adolescents. *Am J Clin Nutr* 1994; 59:S502–S510.
 101. Bothwell TH, Charlton R, Cook J, Finch CA. *Iron metabolism in man*. In: *Iron Metabolism in Man*. Hoboken, NJ: Wiley-Blackwell; 1979; 7–87.
 102. Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Iron intake and risk of ovulatory infertility. *Obstet Gynecol* 2006; 108:1145–1152.

103. Singer S, Vichinsky E, Gildengorin G, van Disseldorp J, Rosen M, Cedars M. Reproductive capacity in iron overloaded women with thalassemia major. *Blood* 2011; 118:2878–2881.
104. Tweed M, Roland J. Haemochromatosis as an endocrine cause of subfertility. *BMJ* 1998; 316:915–916.
105. Mishra A, Tiwari A. Iron overload in beta thalassaemia major and intermedia patients. *Maedica (Buchar)* 2013; 8:328–332.
106. Rodgers RJ, Waterman MR, Simpson ER. Cytochromes P-450_{scc}, P-450(17)alpha, adrenodoxin, and reduced nicotinamide adenine dinucleotide phosphate-cytochrome P-450 reductase in bovine follicles and corpora lutea: changes in specific contents during the ovarian cycle. *Endocrinology* 1986; 118:1366–1374.
107. Schwarz K, Foltz CM. Selenium as an integral part of factor 3 against dietary necrotic liver degeneration. *J Am Chem Soc* 1957; 79: 3292–3293.
108. O'Dell BL, Sunde RA. *Handbook of Nutritionally Essential Mineral Elements*. New York, Basel, Hong Kong: CRC Press; 1997:493–557.
109. Tietz NW, Burtis CA, Ashwood ER. *Tietz Textbook of Clinical Chemistry and Molecular Diagnostics*. St. Louis, Mo: Elsevier Saunders; 2006.
110. Kryukov GV, Castellano S, Novoselov SV, Lobanov AV, Zehtab O, Guigo R, Gladyshev VN. Characterization of mammalian selenoproteomes. *Science* 2003; 300:1439–1443.
111. Reeves MA, Hoffmann PR. The human selenoproteome: recent insights into functions and regulation. *Cell Mol Life Sci* 2009; 66:2457–2478.
112. Mehdi Y, Hornick JL, Istasse L, Dufrasne I. Selenium in the environment, metabolism and involvement in body functions. *Molecules* 2013; 18:3292–3311.
113. Frausto da Silva JJR, Williams RJP. *The Biological Chemistry of the Elements: The Inorganic Chemistry of Life*. New York, NY: Oxford University Press; 1991.
114. Gutmann F, Johnson C, Keyzer H, Molnar J. *Charge Transfer Complexes in Biological Systems*. Boca Raton, FL: CRC Press; 1997.
115. Harrison I, Littlejohn D, Fell GS. Distribution of selenium in human blood plasma and serum. *Analyst* 1996; 121:189–194.
116. Whanger PD. Selenium and its relationship to cancer: an update. *Br J Nutr* 2004; 91:11–28.
117. Weekley CM, Harris HH. Which form is that? The importance of selenium speciation and metabolism in the prevention and treatment of disease. *Chem Soc Rev* 2013; 42:8870–8894.
118. Rayman MP. Selenium and human health. *Lancet* 2012; 379:1256–1268.
119. *Nutrient Reference Values for Australia and New Zealand: Including Recommended Dietary Intakes*. National Health and Medical Research Council: Commonwealth of Australia; 2005.
120. Rayman MP. Food-chain selenium and human health: emphasis on intake. *Br J Nutr* 2008; 100:254–268.
121. Combs GF. Selenium in global food systems. *Br J Nutr* 2001; 85: 517–547.
122. Reilly C. *Selenium in Food and Health*, 2nd ed. New York, NY: Springer; 2006.
123. Thiry C, Ruttens A, Pussemier L, Schneider YJ. An in vitro investigation of species-dependent intestinal transport of selenium and the impact of this process on selenium bioavailability. *Br J Nutr* 2013; 109:2126–2134.
124. Hsieh HS, Ganther HE. Biosynthesis of dimethyl selenide from sodium selenite in rat liver and kidney cell-free systems. *Biochim Biophys Acta* 1977; 497:205–217.
125. Thomson CD, Robinson MF. Urinary and fecal excretions and absorption of a large supplement of selenium - superiority of selenate over selenite. *Am J Clin Nutr* 1986; 44:659–663.
126. Oster O, Prellwitz W. The renal excretion of selenium. *Biol Trace Elem Res* 1990; 24:119–146.
127. Sunde RA, Raines AM, Barnes KM, Evenson JK. Selenium status highly regulates selenoprotein mRNA levels for only a subset of the selenoproteins in the selenoproteome. *Biosci Rep* 2009; 29:329–338.
128. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Science* 1973; 179:588–590.
129. Al-Kunani AS, Knight R, Haswell SJ, Thompson JW, Lindow SW. The selenium status of women with a history of recurrent miscarriage. *BJOG* 2001; 108:1094–1097.
130. Guvenc M, Guvenc H, Karatas F, Aygun AD, Bektas S. Low levels of selenium in miscarriage. *J Trace Elem Exp Med* 2002; 15:97–101.
131. Rumiris D, Purwosunu Y, Wibowo N, Farina A, Sekizawa A. Lower rate of preeclampsia after antioxidant supplementation in pregnant women with low antioxidant status. *Hypertens Pregnancy* 2006; 25:241–253.
132. Watson M, van Leer L, Vanderlelie JJ, Perkins AV. Selenium supplementation protects trophoblast cells from oxidative stress. *Placenta* 2012; 33:1012–1019.
133. Tsuzuki S, Morimoto N, Hosokawa S, Matsushita T. Associations of maternal and neonatal serum trace element concentrations with neonatal birth weight. *Plos One* 2013; 8:e75627.
134. Behne D, Hofer T, Vonberswordtwallrabe R, Elger W. Selenium in the testis of the rat: studies on its regulation and its importance for the organism. *J Nutr* 1982; 112:1682–1687.
135. Behne D, Duk M, Elger W. Selenium content and glutathione-peroxidase activity in the testis of the maturing rat. *J Nutr* 1986; 116:1442–1447.
136. Behne D, Weiler H, Kyriakopoulos A. Effects of selenium deficiency on testicular morphology and function in rats. *J Reprod Fertil* 1996; 106: 291–297.
137. Flohe L. Selenium in mammalian spermiogenesis. *Biol Chem* 2007; 388: 987–995.
138. Irvine DS. Glutathione as a treatment for male infertility. *Rev Reprod* 1996; 1:6–12.
139. Oldereid NB, Thomassen Y, Purvis K. Selenium in human male reproductive organs. *Hum Reprod* 1998; 13:2172–2176.
140. Scott R, Macpherson A, Yates RWS, Hussain B, Dixon J. The effect of oral selenium supplementation on human sperm motility. *Br J Urol* 1998; 82:76–80.
141. Iwanier K, Zachara BA. Selenium supplementation enhances the element concentration in blood and seminal fluid but does not change the spermatozoal quality characteristics in subfertile men. *J Androl* 1995; 16: 441–447.
142. Paszkowski T, Traub AI, Robinson SY, McMaster D. Selenium dependent glutathione peroxidase activity in human follicular fluid. *Clin Chim Acta* 1995; 236:173–180.
143. Ozkaya MO, Naziroglu M, Barak C, Berkkanoglu M. Effects of multivitamin/mineral supplementation on trace element levels in serum and follicular fluid of women undergoing in vitro fertilization (IVF). *Biol Trace Elem Res* 2011; 139:1–9.
144. Coskun A, Arikan T, Kilinc M, Arikan DC, Ekerbicer HC. Plasma selenium levels in Turkish women with polycystic ovary syndrome. *Eur J Obstet Gynecol Reprod Biol* 2013; 168:183–186.
145. Edassery SL, Shatavi SV, Kunkel JP, Hauer C, Brucker C, Penumatsa K, Yu Y, Dias JA, Luborsky JL. Autoantigens in ovarian autoimmunity associated with unexplained infertility and premature ovarian failure. *Fertil Steril* 2010; 94:2636–2641.
146. Butzow JJ, Eichhorn GL. Different susceptibility of DNA and RNA to cleavage by metal ions. *Nature* 1975; 254:358–359.
147. Maret W, Sandstead HH. Zinc requirements and the risks and benefits of zinc supplementation. *J Trace Elem Med Biol* 2006; 20:3–18.
148. Oberleas D, Harland BF. Nutritional agents which affect metabolic zinc status. *Prog Clin Biol Res* 1977; 14:11–27.
149. Berg JM, Shi YG. The galvanization of biology: a growing appreciation for the roles of zinc. *Science* 1996; 271:1081–1085.
150. Favier AE. The role of zinc in reproduction: hormonal mechanisms. *Biol Trace Elem Res* 1992; 32:363–382.
151. Freedman LP. Anatomy of the steroid receptor zinc finger region. *Endocr Rev* 1992; 13:129–145.
152. Zhang XL, Zhao Y, Chu QQ, Wang ZY, Li HJ, Chi ZH. Zinc modulates high glucose-induced apoptosis by suppressing oxidative stress in renal tubular epithelial cells. *Biol Trace Elem Res* 2014; 158:259–267.
153. Chimienti F, Jourdan E, Favier A, Seve M. Zinc resistance impairs sensitivity to oxidative stress in HeLa cells: protection through metallothionein expression. *Free Radic Biol Med* 2001; 31:1179–1190.
154. Zago MP, Oteiza PI. The antioxidant properties of zinc: Interactions with iron and antioxidants. *Free Radic Biol Med* 2001; 31:266–274.
155. Gibson RS. The role of diet- and host-related factors in nutrient bioavailability and thus in nutrient-based dietary requirement estimates. *Food Nutr Bull* 2007; 28:S77–S100.
156. Solomons NW, Jacob RA, Pineda O, Viteri F. Studies on the bioavailability of zinc in man. 2. Absorption of zinc from organic and inorganic sources. *J Lab Clin Med* 1979; 94:335–343.
157. Lim KHC, Riddell LJ, Nowson CA, Booth AO, Szymlek-Gay EA. Iron and zinc nutrition in the economically-developed world: a review. *Nutrients* 2013; 5:3184–3211.
158. Siepmann M, Spank S, Kluge A, Schappach A, Kirch W. The pharmacokinetics of zinc from zinc gluconate: a comparison with zinc oxide in healthy men. *Int J Clin Pharmacol Ther* 2005; 43:562–565.
159. Wolfe SA, Gibson RS, Gadowsky SL, Oconnor DL. Zinc status of a group of pregnant adolescents at 36 weeks of gestation living in southern Ontario. *J Am Coll Nutr* 1994; 13:154–164.
160. Bertinato J, Sherrard L, Plouffe LJ. EDTA disodium zinc has superior bioavailability compared to common inorganic or chelated zinc

- compounds in rats fed a high phytic acid diet. *J Trace Elem Med Biol* 2012; 26:227–233.
161. Gibson RS. Zinc deficiency and human health: etiology, health consequences, and future solutions. *Plant Soil* 2012; 361:291–299.
 162. Cousins RJ. Absorption, transport, and hepatic metabolism of copper and zinc: special reference to metallothionein and ceruloplasmin. *Physiol Rev* 1985; 65:238–309.
 163. Lichten LA, Cousins RJ. Mammalian zinc transporters: nutritional and physiologic regulation. *Annu Rev Nutr* 2009; 29:153–176.
 164. Eide DJ. The SLC39 family of metal ion transporters. *Pflugers Arch* 2004; 447:796–800.
 165. Palmiter RD, Huang LP. Efflux and compartmentalization of zinc by members of the SLC30 family of solute carriers. *Pflugers Arch* 2004; 447:744–751.
 166. Lye JC, Richards CD, Dechen K, Paterson D, de Jonge MD, Howard DL, Warr CG, Burke R. Systematic functional characterization of putative zinc transport genes and identification of zinc toxicosis phenotypes in *Drosophila melanogaster*. *J Exp Biol* 2012; 215:3254–3265.
 167. Lonnerdal B. Dietary factors influencing zinc absorption. *J Nutr* 2000; 130:1378S–1383S.
 168. Report of joint FAO. WHO Expert Consultation on Human Vitamin and Mineral Requirements. Vitamin and Mineral Requirements in Human Nutrition. Geneva: World Health Organization and Food and Agriculture Organization of the United Nations; 2004:246–278.
 169. Dietary Reference Intakes for Vitamin A, Vitamin K, Arsenic, Boron, Chromium, Copper, Iodine, Iron, Manganese, Molybdenum, Nickel, Silicon, Vanadium, and Zinc: A Report of the Panel on Micronutrients. Food and Nutrition Board, Institute of Medicine. Washington DC: National Academies Press; 2001.
 170. Sandstrom B. Bioavailability of zinc. *Eur J Clin Nutr* 1997; 51:S17–S19.
 171. Roohani N, Hurrell R, Kelishadi R, Schulin R. Zinc and its importance for human health: an integrative review. *J Res Med Sci* 2013; 18: 144–157.
 172. Sian L, Xiang MY, Miller LV, Tong L, Krebs NF, Hambidge KM. Zinc absorption and intestinal losses of endogenous zinc in young Chinese women with marginal zinc intakes. *Am J Clin Nutr* 1996; 63:348–353.
 173. Iqbal TH, Lewis KO, Cooper BT. Phytase activity in the human and rat small intestine. *Gut* 1994; 35:1233–1236.
 174. Sandstead HH. Nutritional Aspects of Zinc Consumption. Clifton, VA: IOS Press; 2011.
 175. Dinsdale D, Williams RB. Ultrastructural changes in the sperm tail of zinc-deficient rats. *J Comp Pathol* 1980; 90:559–566.
 176. Wallace E, Calvin HI, Salgo MP, Dennis JE, Ploetz K. Normal levels of zinc and sulfhydryls in morphologically abnormal populations of spermatozoa from moderately zinc-deficient rats. *Gamete Res* 1984; 9: 375–386.
 177. Shaw NA, Dickey HC, Brugman HH, Blamberg DL, Witter JF. Zinc deficiency in female rabbits. *Lab Anim* 1974; 8:1–7.
 178. Swenerton H, Hurley LS. Zinc deficiency in rhesus and bonnet monkeys including effects on reproduction. *J Nutr* 1980; 110:575–583.
 179. Hurley LS, Keen CL. Fetal and neonatal development in relation to maternal trace element nutrition manganese zinc and copper. In: Berger H (ed.), *Vitamins and Minerals in Pregnancy and Lactation*, vol. 16. New York, NY: Vevy/Raven Press Ltd.; 1988:215–230.
 180. Sato F, Watanabe T, Endo A. Cytogenetic effects of zinc deficiency on oogenesis and spermatogenesis. *Teratology* 1982; 26:A13–A14.
 181. Branum AM, Bailey R, Singer BJ. Dietary supplement use and folate status during pregnancy in the United States. *J Nutr* 2013; 143:486–492.
 182. Glinoe D. Pregnancy and iodine. *Thyroid* 2001; 11:471–481.
 183. Patey-Pirra S, Keriell-Gascou M, Borson-Chazot F. Benefits and risks of iodine supplementation during pregnancy: a review of observational and experimental studies in mild-to-moderate iodine deficiency areas. *Rev Epidemiol Sante Publique* 2014; 62:65–74.
 184. Paudel P, Wing K, Silpakar SK. Awareness of periconceptional folic acid supplementation among Nepalese women of childbearing age: a cross-sectional study. *Prev Med* 2012; 55:511–513.
 185. Ramakrishnan U, Grant F, Goldenberg T, Zongrone A, Martorell R. Effect of women's nutrition before and during early pregnancy on maternal and infant outcomes: a systematic review. *Paediatr Perinat Epidemiol* 2012; 26:285–301.
 186. Zhou SJ, Anderson AJ, Gibson RA, Makrides M. Effect of iodine supplementation in pregnancy on child development and other clinical outcomes: a systematic review of randomized controlled trials. *Am J Clin Nutr* 2013; 98:1241–1254.
 187. Taneja SK, Kaur R. Effect of dietary Zn deficiency on the distribution of lipids in the ovary of mature mouse. *Indian J Exp Biol* 1988; 26: 271–273.
 188. Taneja SK, Kaur R. Pathology of ovary, uterus, vagina and gonadotrophs of female mice fed on Zn-deficient diet. *Indian J Exp Biol* 1990; 28: 1058–1065.
 189. Kim AM, Vogt S, O'Halloran TV, Woodruff TK. Zinc availability regulates exit from meiosis in maturing mammalian oocytes. *Nat Chem Biol* 2010; 6:674–681.
 190. Tian X, Anthony K, Neuberger T, Diaz FJ. Preconception zinc deficiency disrupts postimplantation fetal and placental development in mice. *Biol Reprod* 2014; 90:83.
 191. Riley JCM, Behrman HR. Oxygen radicals and reactive oxygen species in reproduction. *Proc Soc Exp Biol Med* 1991; 198:781–791.
 192. Delamirande E, Gagnon C. Reactive oxygen species (ROS) and reproduction. In: Armstrong D (ed.), *Free Radicals in Diagnostic Medicine: a Systems Approach to Laboratory Technology, Clinical Correlations, and Antioxidant Therapy*, vol. 366. New York, NY: Springer US; 1994:185–197.
 193. Chavarro J, Rich-Edwards J, Rosner B, Willett W. Diet and lifestyle in the prevention of ovulatory disorder infertility. *Obstet Gynecol* 2007; 110:1050–1058.
 194. Pieczynska J, Grajeta H. The role of selenium in human conception and pregnancy. *J Trace Elem Med Biol* 2015; 29:31–38.
 195. Tian X, Diaz FJ. Zinc depletion causes multiple defects in ovarian function during the periovulatory period in mice. *Endocrinology* 2012; 153:873–886.