

REVIEW PAPER

# Trace metal phytotoxicity in solution culture: a review

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## Abstract

Solution culture has been used extensively to determine the phytotoxic effects of trace metals. A review of the literature from 1975 to 2009 was carried out to evaluate the effects of As(V), Cd(II), Co(II), Cu(II), Hg(II), Mn(II), Ni(II), Pb(II), and Zn(II) on plants grown in solution. A total of 119 studies was selected using criteria that allowed a valid comparison of the results; reported toxic concentrations varied by five orders of magnitude. Across a range of plant species and experimental conditions, the phytotoxicity of the trace metals followed the trend (from most to least toxic):  $Pb \approx Hg > Cu > Cd \approx As > Co \approx Ni \approx Zn > Mn$ , with median toxic concentrations of ( $\mu M$ ): 0.30 Pb, 0.47 Hg, 2.0 Cu, 5.0 Cd, 9.0 As, 17 Co, 19 Ni, 25 Zn, and 46 Mn. For phytotoxicity studies in solution culture, we suggest (i) plants should be grown in a dilute solution which mimics the soil solution, or that, at a minimum, contains Ca and B, (ii) solution pH should be monitored and reported (as should the concentrations of the trace metal of interest), (iii) assessment should be made of the influence of pH on solution composition and ion speciation, and (iv) both the period of exposure to the trace metal and the plant variable measured should be appropriate. Observing these criteria will potentially lead to reliable data on the relationship between growth depression and the concentration of the toxic metal in solution.

**Key words:** Critical concentration, phytotoxicity, solution culture, trace metal.

## Introduction

Trace metals are natural components of the environment, but elevated and potentially toxic levels sometimes occur. Some soils contain trace metals at naturally elevated concentrations, such as with Ni in soils formed from ultramafic (serpentine) minerals (Anderson *et al.*, 1973; Batianoff and Singh, 2001). In acid soils, elevated levels of soluble metals (particularly Al or Mn) may occur, whilst high concentrations of other metals (such as Cu or Pb) may be present in sites contaminated by agriculture, mining, industry, or transport. The presence of excess trace metals represents a serious environmental and financial problem, with *c.* 30% of the world's land affected by acidity (Sumner and Noble, 2003) and hundreds of thousands of metal-contaminated sites worldwide which require remediation at an estimated cost of up to US\$35 billion (CEI, 2005). (Although strictly a metalloid, As will be grouped with the other trace metals for the purposes of this study.)

Many trace metals (such as Cu, Mn, and Zn) are essential for the growth of plants and animals, but are toxic at

elevated concentrations. There are numerous reviews in the literature which examine the influence of trace metals on plant growth and function (for example, see Clemens, 2006; Babula *et al.*, 2008). However, underlying all studies is the requirement to expose the plant to a toxic, but appropriate, concentration of the trace metal. For example, some solution culture studies have used up to 1000  $\mu M$  of the trace metal, despite published data demonstrating a complete cessation of growth at concentrations as low as 1  $\mu M$  (see later discussion). There is a need, therefore, to establish criteria for determining which data on trace metal phytotoxicity in solution culture are likely to be reliable and to summarize these high-quality data.

Whilst the phytotoxicity of trace metals has been studied for over a century (for example, see Jensen, 1907), there remains considerable variation within the literature as to the concentrations of trace metals necessary to induce toxic effects. An initial examination of the literature on trace metal toxicities in solution culture revealed that the

concentrations used to induce toxic effects varied by at least eight orders of magnitude, from 1 nM (Godbold, 1991) to high millimolar concentrations. Among factors likely to contribute to this variation are differences in: (i) the inherent toxicity of the various trace metals, (ii) tolerances among plant species, and (iii) the experimental techniques used in the various studies. Whilst it is these first two points (i.e. the toxicity of trace metals and the sensitivity of plants to them) which form the basis of many phytotoxicity studies, it appears that differences in 'true' toxic effects are often confounded by the experimental conditions employed. For example, Taylor and Foy (1985) reported that *c.* 30  $\mu\text{M}$  Cu is required to reduce growth of wheat (*Triticum aestivum* L.) by 50%, whereas Wheeler *et al.* (1993) found that only 0.5  $\mu\text{M}$  Cu was required for a 50% growth reduction in the same species. It is unlikely that such a large discrepancy could be due solely to genotypic effects.

The aim of the current study was to provide a comprehensive review of the literature to determine the range in concentrations over which nine trace metals [As(V), Cd(II), Co(II), Cu(II), Hg(II), Mn(II), Ni(II), Pb(II), and Zn(II)] have been reported to exert phytotoxic effects in solution culture. Although an important trace metal, Al was not included in the current study because its toxic effects result from soil acidification; neither were rare trace metals (such as Ga, Gd, and Sc) included. Also, Fe toxicity is confined to waterlogged soils, and whilst it may be of particular interest under paddy conditions, it was excluded from the present study. For arsenic, only arsenate, As(V), was considered; this species dominates under aerobic conditions. Given the wide range of concentrations which have been reported to be toxic, criteria were first established to minimize the influence of experimental conditions on apparent 'toxicity' of the nine trace metals. This review of the literature includes the results of only those studies meeting the criteria.

## Materials and methods

An extensive data set was collected from the literature for solution culture studies examining the phytotoxicity of As(V), Cd, Co, Cu, Hg, Mn, Ni, Pb, and Zn. Two databases (ISI Web of Science and Google Scholar) were searched from 1975 onwards, with the final date of searching being July 2009. Next, using the ISI Web of Science, all articles citing the retrieved references and all articles cited in the retrieved references were searched for further relevant publications. Two sets of criteria were used in this study; acceptance criteria and evaluation criteria. Acceptance criteria aimed to exclude investigations which were incompatible with the purpose of the current study (i.e. to determine the range in concentrations over which trace metals exert phytotoxic effects in solution culture). Once a study had been accepted, evaluation criteria were then applied to ascertain the suitability of the test procedures.

Ten acceptance criteria were used to determine eligibility for inclusion into the dataset, thus limiting data to those that were appropriate for the aims of the current study. Specifically, it was decided that the study must: (i) be conducted using solution culture (thus, excluding studies using agar, filter paper, sand, or soil), (ii) provide a direct measurement of plant growth (e.g. biomass or elongation of the root or shoot), (iii) examine the growth of intact plants (i.e. not excised portions), (iv) include

a control, which either contains no added metal or a basal (non-toxic) concentration in the case of essential trace metals, (v) utilize a minimum of four levels of the trace metal (inclusive of the control) with reported nominal or measured concentrations, (vi) report the duration of exposure and any non-exposure periods (e.g. during germination or early seedling growth), (vii) utilize metal concentrations sufficient to cause a significant decrease in growth, (viii) be the primary source of the data, (ix) utilize only a single stressor (or, if multiple stressors were examined, provide data for the stressors individually in addition to their combined effects), and (x) investigate the toxicity of the free, ionic metal. Regarding the last-named criterion, data from studies that examined the effects of chelation by organic complexes (such as EDTA) were excluded since chelation has marked effects on trace metal speciation (Parker and Norvell, 1999).

Those studies meeting the above criteria were subjected to further quality assurance using the evaluation criteria. First, it was considered necessary that the plants had been grown in a complete nutrient solution, or at least in solution containing Ca (i.e. studies were excluded in which control plants were grown in deionized water). Second, the pH of the nutrient solution needed to be reported, given the importance of pH on solubility and ionic speciation. Third, to be included in the database, in those instances where modelling with PhreeqcI 2.15.0 (Parkhurst, 2009) indicated the solution to be supersaturated with respect to the metal of interest, it was necessary for the solution to have been sampled, filtered, and the soluble metal concentration measured. Fourth, in many studies, total growth of plants was reported for a period which included both an establishment phase and a phase involving exposure to the trace metal. Results of these studies were incorporated into the database only if: (i) a minimum of 50% of the time was spent in the metal-containing solution, or (ii) growth was assessed only for the period in the metal-containing solution (e.g. root elongation rate during metal-exposure versus the total root biomass).

The following parameters were recorded for each study entered into the database: (i) publication details, (ii) trace metal stressor, (iii) total number of treatments per stressor, (iv) concentrations (or activities) of a stressor determined as being toxic, (v) pH of nutrient solution, (vi) P concentration in solution, (vii) duration of exposure, (viii) plant species, (ix) plant growth variable measured, and (x) approximate growth reduction caused by the stressor at the toxic concentration.

In most studies, the concentration of the trace metal considered to be toxic was reported in the text of the article; alternatively, the values were determined from the figures or tables. Where an analysis of variance had been used, the lowest metal concentration causing a significant reduction in growth was selected. Values in the range of  $EC_{25}$ – $EC_{50}$  (i.e. 25–50% growth reduction) were selected from studies where the growth response had been modelled (e.g. by regression analysis). Where authors reported the toxicity of the stressor as the activity of the free ion (e.g. the activity of  $\text{Cu}^{2+}$ ), this was noted, but no discrimination was made between values reported as concentrations or activities. It was surprising, and rather disappointing, that the concentration of the trace metal of interest was measured in very few studies; rather, studies often simply reported the nominal (added) concentrations. Where losses of the metal have occurred, for example, by precipitation, relating growth reductions to the nominal concentration will lead to an underestimation of the actual toxicity of the trace metal (Lee *et al.*, 2005).

A total of 119 studies were entered into the database, including 28 for Cu, 22 for Cd, 17 for Mn, 13 for Ni, 13 for Zn, 11 for As, eight for Hg, four for Co, and three for Pb (see Supplementary Table S1 at *JXB* online). There was an overall total of 180 limiting metal concentrations; some studies including data on a number of plant species. The most commonly investigated species was wheat which was included in 19 studies. The median number of trace metal treatments was six (ranging from 4 to 58).

## Results and discussion

### Concentrations of trace metals found to be phytotoxic

The review of scientific literature over the past 34 years showed that trace metal phytotoxicity followed the general trend (from most toxic to least toxic):  $Pb \approx Hg > Cu > Cd \approx As > Co \approx Ni \approx Zn > Mn$  (Fig. 1). The median toxic concentration varied by about two orders of magnitude among the nine metals, being ( $\mu M$ ): 0.30 Pb, 0.47 Hg, 2.0 Cu, 5.0 Cd, 9.0 As, 17 Co, 19 Ni, 25 Zn, and 46 Mn (Fig. 1). This toxicity ranking is similar to that found in individual studies on the toxicity of a range of metals to a single species. For example, Wheeler *et al.* (1993) reported that wheat root mass was reduced by 50% in solutions containing ( $\mu M$ ) 0.5 Cu, 19 Zn, or 600 Mn (toxic values were also reported for Sc, La, Ga, Al, Fe, and B). Similarly, Taylor *et al.* (1991) reported that root mass of wheat was reduced by 5% in solutions containing ( $\mu M$ ) 0.02 Cd, 3.4 Cu, 11 Ni, 37 Mn, or 45 Zn (Al toxicity was also studied).

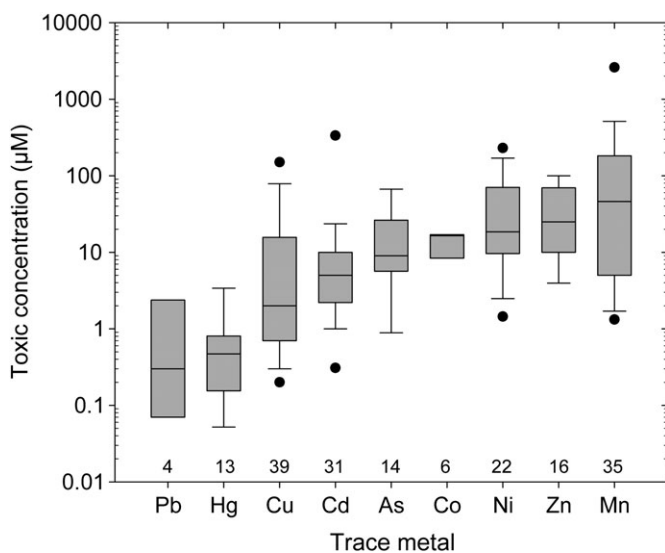
The median trace metal concentrations found to be toxic in the present review were all  $< 100 \mu M$  (c.  $1 \mu M$  for Hg and Pb to  $47 \mu M$  for Mn). These concentrations are comparable with those in soil solutions from metal-toxic soils, for example, c.  $1 \mu M$  Pb (Weng *et al.*, 2001; Degryse *et al.*, 2007),  $5 \mu M$  Cu (Aguirre-Gomez *et al.*, 2006; Luo *et al.*, 2006), or  $50 \mu M$  Ni (Anderson *et al.*, 1973; Proctor *et al.*, 1981). However, in the literature, numerous studies were found using concentrations of trace metals up to four orders of magnitude higher. For example, Zeid (2001) used up to  $50 \text{ mM}$  Co in a sand culture study on common bean

(*Phaseolus vulgaris* L.); Chi Yu *et al.* (2005) used  $10 \text{ mM}$  Cu in solution culture to investigate the influence of nitric oxide on Cu toxicity and  $NH_4^+$  accumulation in rice (*Oryza sativa* L.), and Sahi *et al.* (2007) used up to  $4.7 \text{ mM}$  Cu in solution culture when investigating rattlebush (*Sesbania drummondii* (Rydb.) Cory). Such studies are far removed from the metal-toxic field situation, and the results obtained are of little value in understanding metal phytotoxicity.

Whilst for each trace metal, the 25th and 75th percentile varied by about one order of magnitude (e.g. ranging from  $2.2 \mu M$  to  $10 \mu M$  for Cd), this variation in concentration required to induce toxic effects is not unexpected. Rather, there are several factors which may have contributed to this variability. These factors are related both to the plant species investigated and to the specific experimental conditions employed within each study.

Often, the aim of phytotoxicity studies is to identify genotypic differences in sensitivity (e.g. to aid in the identification of mechanisms, or to select resistant or tolerant plants for revegetation of contaminated lands). Indeed, there is a large variation in the sensitivity of plant species to trace metals under the same experimental conditions, or even among populations within a species. In a study involving four Australian tree species, Reichman *et al.* (2004) reported that shoot mass was reduced by 10% at  $5.0 \mu M$  Mn for *Eucalyptus crebra* F. Muell. but only at  $330 \mu M$  Mn for *Eucalyptus camaldulensis* Dehnh. Similarly, Edwards and Asher (1982) found that across 13 crop and pasture species, the external Mn concentration needed to reduce plant dry mass by 10% varied from  $1.4 \mu M$  in two monocots (maize, *Zea mays* L., and wheat) to  $65 \mu M$  in a dicot (sunflower, *Helianthus annuus* L.). In a study with *Silene cucubalus* (Wib), de Vos *et al.* (1991) reported that the  $EC_{50}$  for root elongation was  $4.0 \mu M$  Cu in a sensitive population but was  $150 \mu M$  Cu in a tolerant population collected from a Cu-contaminated site.

Whilst comparisons in a specific experiment are possible, comparing metal toxicity between studies is often difficult because of differing experimental conditions which may markedly affect the concentration of metal found to be toxic. As part of the quality assessment in the current study, several evaluation criteria were developed to identify those studies where it is possible to compare results. It is proposed that these criteria should underpin all experiments on the phytotoxicity of trace metals.



**Fig. 1.** Concentrations of nine trace metals that reduced the growth of plants in solution culture, obtained from a review of the literature from 1975 to 2009 ( $n=180$ ). For each trace metal, the Tukey box plot represents the 25th and 75th percentiles (with the median contained therein). In cases where there are at least nine data points, the whiskers represent the largest and smallest values which are not outliers and, where shown, the dots represent the largest and smallest outliers. The numbers above each trace metal on the x-axis indicate the number of data points for that metal.

### Nutrient solution composition

The composition of the base nutrient solution has marked effects on the perceived toxicity of trace metals. Unfortunately, this does not seem to have been considered in many studies, resulting in toxicity data which are of limited value. Thus, besides ensuring appropriate trace metal concentrations, there is a need to pay particular attention to the overall composition of the nutrient solution.

Since plants can draw on their nutrient reserves for short periods of time, it is possible to conduct meaningful metal-toxicity experiments in simplified nutrient solutions which

do not contain all the essential elements. However, because Ca does not move towards the root tip, it must be present in the test solution to maintain structural and functional integrity. Indeed, it has been noted that root growth is reduced rapidly when placed in solutions lacking Ca (Burstrom, 1953; Kinraide, 1998; del Amor and Marcelis, 2003). Root tips of six tropical legumes were thick and blackened with  $<12 \mu\text{M}$  Ca in solution; indeed, symptoms were evident within 2 d at  $2 \mu\text{M}$  Ca (Bell *et al.*, 1989) and there was poor lateral root development at  $2 \mu\text{M}$  Ca also. Spehar and Galwey (1997) found that, in the absence of Ca, the primary root length of eight soybean [*Glycine max* (L.) Merr.] lines was only  $34 \pm 4$  mm after 7 d but ranged from 99 mm to 147 mm with  $500 \mu\text{M}$  Ca; at least  $100 \mu\text{M}$  Ca was needed to discriminate among lines varying in root growth. The absence of B in nutrient solutions also 'leads to morphological changes ... within hours or days' (Goldbach *et al.*, 2001). Therefore, at a minimum, the nutrient solution must contain Ca and B. However, examination of the literature revealed numerous studies where roots were grown in deionized water with no nutrients added. For example, Yildiz *et al.* (2009) conducted a study in which roots of onion (*Allium cepa* L.) were grown in deionized water for 4 d.

The composition of a nutrient solution should ideally mimic that of a soil solution (Table 1) (Parker and Norvell, 1999). This is especially important if the aim of the solution culture experiment is to study the effects of a toxic metal on plant growth in the field. However, for reasons of convenience, many well-known and commonly-used nutrient solutions, such as that of Hoagland

and Arnon (1950), employ high initial concentrations of nutrient salts. This allows a large total supply of nutrients in a conveniently small volume of solution, but the concentrations are typically 1–3 orders of magnitude higher than those commonly found in soil solutions (Table 1). This is particularly so for P, which is typically present in soil solution at a low concentration relative to those used in many nutrient solution culture studies. Soil solution P concentration is often  $<2 \mu\text{M}$  in unfertilized forest soils and in highly weathered soils (Gillman and Bell, 1978; Menzies and Bell, 1988) (Table 1). In agricultural soils, soil solution P is increased by fertilizer use, but the soil solution P concentration is still generally  $<10 \mu\text{M}$ . For example, 80% of 149 samples in the data compilation of Reisenauer (1966), and 80% of those in a study of 33 soils by Kovar and Barber (1988), fell below  $10 \mu\text{M}$  P (Table 1). It is only in soils which have recently received P fertilizer that soil solution P concentration of *c.*  $100 \mu\text{M}$  is evident (Wiklander and Andersson, 1974; Adams *et al.*, 1980; Wheeler and Edmeades, 1995). However, toxicities of trace metals such as Pb would not occur in these highly fertile (high-P) soils due to the precipitation of metal-phosphates. Indeed, P-fertilization is one method of reducing soluble Pb concentrations when remediating contaminated sites (Zhu *et al.*, 2004). Yet, in the solution culture studies reviewed, the median P concentration was  $100 \mu\text{M}$  (ranging from  $0 \mu\text{M}$  to  $4000 \mu\text{M}$ ) (Table 1).

In the studies reviewed, the median ionic strength was found to be  $4.7 \text{ mM}$  (ranging from  $0.29 \text{ mM}$  to  $46 \text{ mM}$ ), with soil solutions typically having an ionic strength of *c.*

**Table 1.** Comparison of the composition of Hoagland's No. 2 solution, a dilute nutrient solution, and soil solutions extracted from a Krasnozem (Oxisol) from Queensland, Australia and eight soils from New Zealand  
Ionic strength was calculated using Phreeqcl where sufficient data were available.

	Hoagland's No. 2 solution <sup>a</sup> ( $\mu\text{M}$ )	Dilute nutrient solution <sup>b</sup> ( $\mu\text{M}$ )	Soil solution <sup>c</sup> (unfertilized) ( $\mu\text{M}$ )	Soil solution <sup>d</sup> ( $0 \text{ kg P ha}^{-1} \text{ year}^{-1}$ ) ( $\mu\text{M}$ )	Soil solution <sup>d</sup> ( $80 \text{ kg P ha}^{-1} \text{ year}^{-1}$ ) ( $\mu\text{M}$ )
Ionic strength	26 000	2 700	4 900	–	–
$\text{NO}_3^- - \text{N}$	14 000	450	1 740	–	–
$\text{NH}_4^+ - \text{N}$	1 000	150	320	–	–
K	6 000	300	850	250	240
Ca	4 000	500	520	370	450
S	2 000	600	310	89	91
Mg	2 000	100	700	150	150
P	1 000	2.5	0.13	5	45
B	46	3	–	–	–
Fe	25	2.5	24	6.2	5.2
Cl	18	0	860	–	–
Mn	9	0.5	3.2	1.6	0.9
Zn	0.8	0.5	–	–	–
Cu	0.3	0.1	–	–	–
Na	0	0	250	490	510

<sup>a</sup> See Hoagland and Arnon (1950) or Parker and Norvell (1999).

<sup>b</sup> Taken from Wheeler *et al.* (1993).

<sup>c</sup> Surface soil of a highly weathered Krasnozem (Oxisol) from Queensland, Australia (Menzies and Bell, 1988).

<sup>d</sup> Average values of soil solutions collected from eight surface soils (0–50 mm) from New Zealand receiving P fertilizer at either  $0$  or  $80 \text{ kg P ha}^{-1} \text{ year}^{-1}$  for 4 years (Wheeler and Edmeades, 1995).

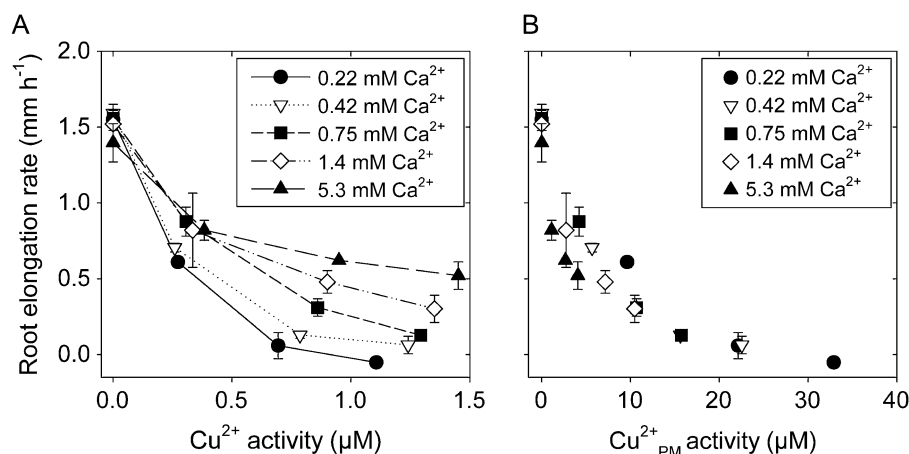
0.5–10 mM (Edmeades *et al.*, 1985; Menzies and Bell, 1988; Bruce *et al.*, 1989; Agbenin, 2003). High ionic strength solutions often affect trace metal toxicity as the concentration of other nutrients has an influence on the toxicity of the metal. For example, Lock *et al.* (2007b) reported that the activity of  $\text{Ni}^{2+}$  required to reduce root length of barley (*Hordeum vulgare* L.) by 50% increased 20-fold (from 5.05  $\mu\text{M}$  to 105  $\mu\text{M}$ ) as the solution Mg concentration increased from 0.05 mM to 3.9 mM. Similarly, a study using the technique of Kopittke *et al.* (2008b) on short-term root growth in cowpea [*Vigna unguiculata* (L.) Walp. cv. Caloona] showed that an increase in the activity of  $\text{Ca}^{2+}$  increased the  $\text{EC}_{50}$  of  $\text{Cu}^{2+}$  activity from *c.* 0.24  $\mu\text{M}$  to 0.59  $\mu\text{M}$  (Fig. 2). This cation amelioration of cation toxicity probably does not result from changes in metal-speciation, but is attributable to changes in cation activity both in the bulk solution (Taylor *et al.*, 1998) and, perhaps more importantly, at the root-cell plasma membrane surface (Kinraide, 2006). For example, the data in Fig. 2 show the influence of cation composition, in this case Ca concentration, on the toxicity of  $\text{Cu}^{2+}$  [as determined from the activity of  $\text{Cu}^{2+}$  either in the bulk solution (Fig. 2A) or at the root-cell plasma membrane surface (Fig. 2B)]. However, in this review, no relationship was found between the concentration of metal which is toxic and solution ionic strength (data not presented). It is likely that the toxic values decrease in high ionic strength solutions, but we consider that the data from the reviewed studies is confounded by other variables (e.g. differences in sensitivity among plant genotypes). The effects of specific ions should also be considered. For example, phosphate inhibits arsenate uptake due to a competitive interaction (Asher and Reay, 1979; Tamaki and Frankenberger, 1992); hence, the phytotoxicity of As is likely to be underestimated where high P concentrations are used. It is interesting that Zn toxicity was alleviated in wheat and radish (*Raphanus*

*sativus* L.) by as little as 1–5  $\mu\text{M}$  Mg, concentrations too low to affect Zn activity in the bulk solution or at the plasma membrane (Pedler *et al.*, 2004). While the ameliorative mechanism in this instance remains unknown, it appears distinct from that of Ca (illustrated in Fig. 2).

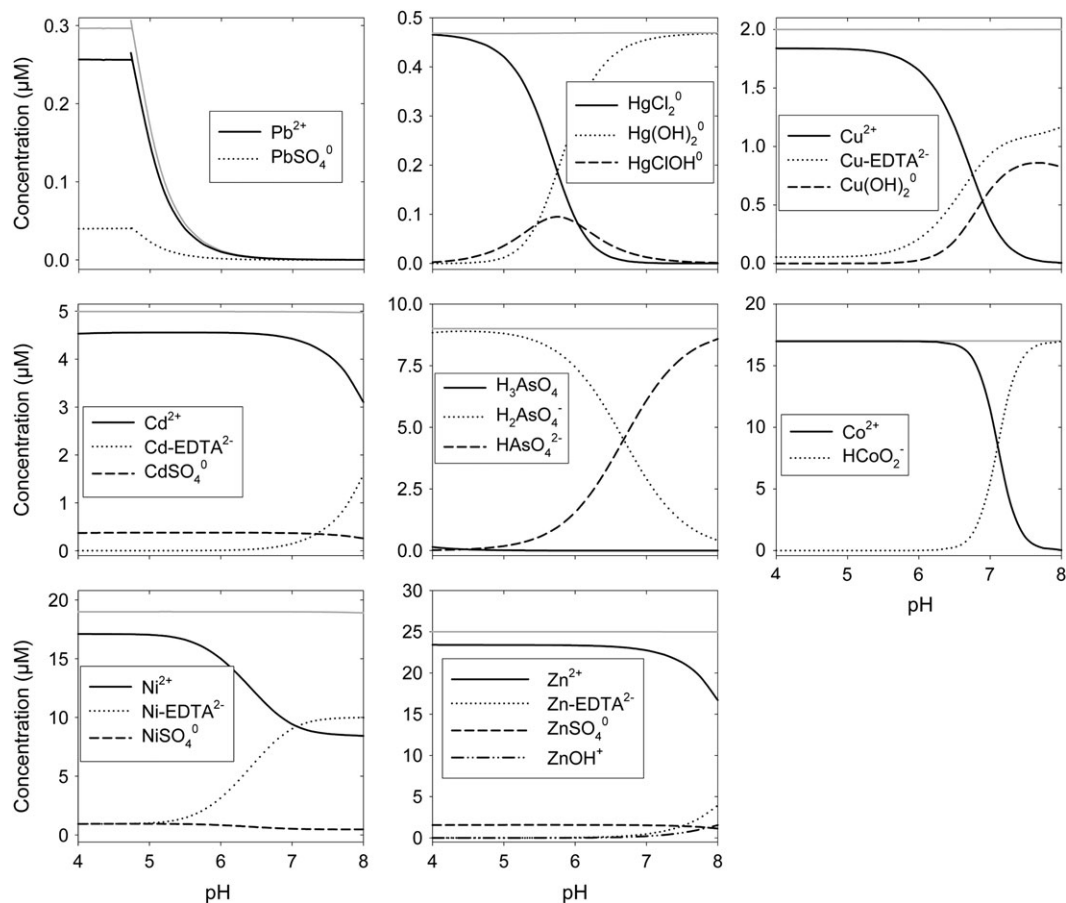
#### Solution pH and trace metal speciation

The pH of the nutrient solution is an extremely important property in regulating the solubility, speciation, and toxicity of trace metals; hence, the results of a study are of limited value without knowledge of solution pH. Perhaps rather surprisingly, about one-third of the studies did not list the pH used, including recently published studies (Israr *et al.*, 2006; Sahi *et al.*, 2007; Krantev *et al.*, 2008). Given that metal toxicity is most commonly encountered on acidic soils, studies should typically be conducted at low pH. Indeed, the median pH of studies included in the database was 5.5 (ranging from pH 4.0 to pH 7.5).

Firstly, solution pH has a major influence on the solubility of many trace metals, this being well known for Al. This is particularly important for Pb (Fig. 3) among the trace metals examined in the current review. Lead phosphates are highly insoluble (Kopittke *et al.*, 2008a), and large amounts of Pb would have precipitated in the study of Malone *et al.* (1974) who added up to 4.8 mM Pb to Hoagland's solution (1000  $\mu\text{M}$  P) even at pH 3.5–4.0 when investigating Pb toxicity in maize. Similarly, investigating the toxicity of Pb to *Beta vulgaris* L., Larbi *et al.* (2002) noted the 'immediate formation of a white precipitate cloud' following the addition of up to 2 mM Pb to a nutrient solution at pH 5.5. The importance of pH can also be seen in the study of Wong and Bradshaw (1982) (which, as of July 2009, has been cited 100 times). The concentrations of Al, Fe, Mn, or Pb reported to reduce root growth of ryegrass (*Lolium perenne* L.) by 50% in 3 mM  $\text{Ca}(\text{NO}_3)_2$



**Fig. 2.** Effects of the activities of  $\text{Cu}^{2+}$  [in either the bulk solution (A) or at the plasma membrane surface (B)] and  $\text{Ca}^{2+}$  and on the average root elongation rate (0–24 h) of 3-d-old cowpea seedlings grown in a solution containing 5  $\mu\text{M}$   $\text{H}_3\text{BO}_3$  at pH 5.3. All bulk solution  $\text{Ca}^{2+}$  and  $\text{Cu}^{2+}$  activities were calculated using Phreeqcl from measured concentrations (see Kopittke *et al.*, 2008b, for more details). The  $\text{Cu}^{2+}$  activity at the plasma membrane surface was calculated as described by Kinraide (2006). Vertical bars represent the standard deviations of the arithmetic mean of two replications (where not visible, the vertical bars are smaller than the symbol). The Ca was supplied as  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$  and the Cu as  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ .



**Fig. 3.** Speciation of nine trace metals in a dilute nutrient solution containing a total of ( $\mu\text{M}$ ): 0.30 Pb, 0.47 Hg, 2.0 Cu, 5.0 Cd, 9.0 As, 17 Co, 19 Ni, 25 Zn, and 46 Mn (the median toxic concentrations listed in Fig. 1). The solid grey line represents the total soluble concentration (for Pb, the soluble concentration decreased markedly with increasing pH due to precipitation as  $\text{Pb}_5(\text{PO}_4)_3\text{Cl}$ ). The solutions were modelled using PhreeqcI 2.15.0 (Parkhurst, 2009), with the Minteq database (other than for Co), using a dilute nutrient solution containing ( $\mu\text{M}$ ): 680  $\text{NO}_3^-$  - N, 120  $\text{NH}_4^+$  - N, 650 Ca, 502 S, 302 K, 140 Cl, 50 Mg, 10 Fe (as EDTA), 3 B, 2 P, 2 Mn, 1 Zn, 0.2 Cu, and 0.02 Mo (Kopittke *et al.*, 2008a) and in equilibrium with atmospheric  $\text{O}_2$ . The Minteq database contained no constants for Co, so the 'Iln' database (prepared by Jim Johnson, Lawrence Livermore National Laboratory) supplied with PhreeqcI 2.15.0 was used. Only the soluble species with the highest concentrations are presented. Solutions were not modelled for Mn as the relationship between measured and predicted concentrations is often poor (Norvell, 1988).

adjusted to pH 7.0 were considerably higher than those predicted to have remained in solution. Indeed, of the 30.8  $\mu\text{M}$  Al added to reduce growth by 50%, it is predicted using PhreeqcI that  $<1$   $\mu\text{M}$  remained in solution. Similarly,  $<1$   $\mu\text{M}$  of the 256  $\mu\text{M}$  Fe (assuming  $\text{Fe}^{2+}$  was oxidized to  $\text{Fe}^{3+}$  and no chelators were used) and 2.5  $\mu\text{M}$  of the 8.2  $\mu\text{M}$  Pb is predicted to have remained in solution. Although much of the Mn was also likely to have precipitated, Mn solutions were not modelled as the relationship between measured and predicted concentrations is often poor (Norvell, 1988). It is possible, therefore, that the solutions in the studies of Malone *et al.* (1974), Wong and Bradshaw (1982), and Larbi *et al.* (2002) may not have reached equilibrium. This further emphasizes the need to measure the soluble trace metal concentrations in filtered solutions, thereby establishing with greater certainty the concentrations or activities that are toxic to plants.

Comparatively few studies have considered trace metal speciation when examining their phytotoxicity. For the nine trace metals included in this study (and within the pH range commonly employed), consideration of speciation is particularly important for Hg, since it is unlikely that the free  $\text{Hg}^{2+}$  will be the dominant ion. Rather, solutions will tend to be dominated by  $\text{HgCl}_2^0$  or  $\text{Hg}(\text{OH})_2^0$  (Fig. 3). The influence of Fe-chelators (such as EDTA) on solution speciation should also be considered, particularly in solutions at  $\geq c.$  pH 5.5 (Fig. 3).

Finally, a decrease in solution pH decreases the adsorption of metals onto and absorption into plant roots (Rengel, 2002). This was reflected, for example, in the study of Lock *et al.* (2007a) in which the root growth  $EC_{50}$  in barley for  $\text{Cu}^{2+}$  activity was 0.083  $\mu\text{M}$  at pH 7.7, but this increased to 0.44  $\mu\text{M}$  at pH 4.5. Weng *et al.* (2003) also reported that the  $EC_{50}$  for  $\text{Ni}^{2+}$  activity increased from 1.7  $\mu\text{M}$  to 23  $\mu\text{M}$  with

a decrease from pH 7.0 to pH 4.0. Similarly, a short-term study, similar to that of Kopittke *et al.* (2008b), showed that poor root elongation rate ( $0.2 \text{ mm h}^{-1}$ ) was evident at pH 4.0 irrespective of Cu concentration (data not shown). At higher pH, however, the  $EC_{50}$  for  $\text{Cu}^{2+}$  toxicity in cowpea increased from  $0.52 \text{ }\mu\text{M}$  to  $0.87 \text{ }\mu\text{M}$   $\text{Cu}^{2+}$  as the pH decreased from 5.3 to 4.6 (Fig. 4A). As with the effect of Ca (Fig. 2B), this effect of pH can potentially be explained due to a change in the activity of the toxicant (in this case,  $\text{Cu}^{2+}$ ) at the plasma membrane surface (Fig. 4B). Interestingly, the data from the effects of Ca (Fig. 2B) and pH (Fig. 4B) on Cu toxicity can be combined to produce a single relationship between root elongation rate and changes in  $\text{Cu}^{2+}$  activity at the plasma membrane surface.

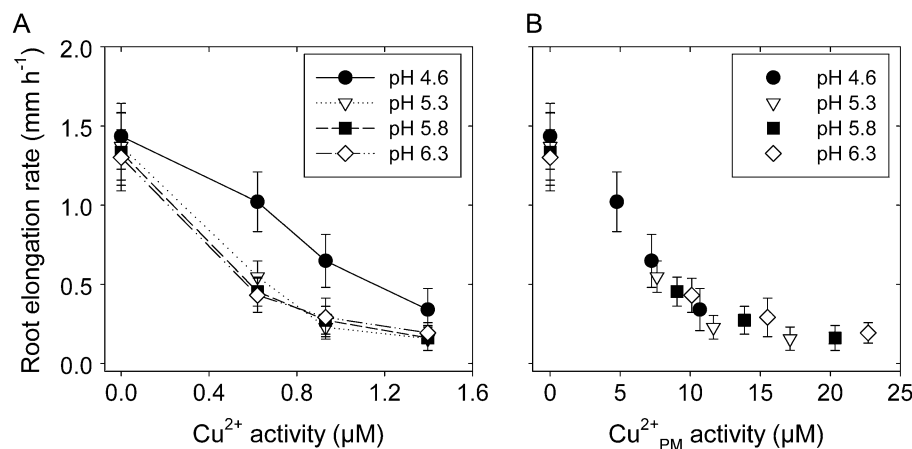
#### Time of exposure to metals

The length of time that roots are exposed to trace metals is important in determining their toxicity; the median duration of metal-exposure for studies incorporated into the database was 14 d (ranging from 2 d to 90 d). Many trace metals exert toxic effects within minutes or hours (Rengel, 1996; Blamey *et al.*, 2004; Kopittke *et al.*, 2008b, 2009), and the relative magnitude of their influence on plant growth increases with time of exposure. For example, Charpentier *et al.* (1987) reported that the  $EC_{50}$  for duckweed (*Lemna polyrrhiza* L.) exposed to Cd decreased from  $1.5 \text{ }\mu\text{M}$  after 4 d exposure to  $0.8 \text{ }\mu\text{M}$  after 14 d exposure. The length of exposure is particularly important in studies where plants are initially grown in a toxicant-free environment before transfer to metal-containing solutions and growth is measured as a 'bulk' variable. For example, root elongation rate during the metal-exposure period would be a more sensitive indicator of toxicity than the total mass of roots (i.e. including those produced during the non-exposure period).

This does not seem to have been considered in the study of Mourato *et al.* (2009) who grew yellow lupin (*Lupinus luteus* L.) for 49 d in a toxicant-free environment before exposing them to excess Cu for 15 d. These authors reported that a Cu concentration of  $\leq 50 \text{ }\mu\text{M}$  did not affect the total biomass of the plant. This most likely occurred not because  $50 \text{ }\mu\text{M}$  Cu is not toxic (see Fig. 1), but because most of the biomass had been produced in the toxicant-free environment with insufficient time allowed for differences to develop between treatments.

## Conclusions

A review of the literature over the past 34 years showed that the concentration required to reduce plant growth followed the general trend (from most to least toxic):  $\text{Pb} \approx \text{Hg} > \text{Cu} > \text{Cd} \approx \text{As} > \text{Co} \approx \text{Ni} \approx \text{Zn} > \text{Mn}$ . The median toxic concentration was ( $\mu\text{M}$ ): 0.30 Pb, 0.47 Hg, 2.0 Cu, 5.0 Cd, 9.0 As, 17 Co, 19 Ni, 25 Zn, and 46 Mn, but the 25th and 75th percentile causing toxicity varied by about one order of magnitude for each trace metal. We conclude that this was due to differences among plant species and in experimental conditions, the latter sometimes making it difficult to apply the results to field situations. To improve the utility of the data, it is suggested that studies investigating trace metal phytotoxicity in solution culture: (i) use a solution containing, as a minimum, Ca and B, but ideally one which mimics the composition of the soil solution, particularly for P, (ii) carefully monitor and report solution pH, (iii) filter the solution and measure the actual concentration of metal in solution and assess the influence of pH on solution composition and speciation, and (iv) use an appropriate period of exposure and ensure that the plant variable measured is a sensitive indicator of toxicity.



**Fig. 4.** Effects of solution pH and the activity of  $\text{Cu}^{2+}$  (in either the bulk solution (A) or at the plasma membrane surface (B)) on root elongation rate (0–26 h) of 3-d-old cowpea seedlings grown in solution containing  $1000 \text{ }\mu\text{M}$  Ca and  $5 \text{ }\mu\text{M}$   $\text{H}_3\text{BO}_3$ . All bulk solution  $\text{Cu}^{2+}$  activities were calculated using Phreeqcl from measured concentrations (see Kopittke *et al.*, 2008b, for more details). The  $\text{Cu}^{2+}$  activity at the plasma membrane surface was calculated as described by Kinraide (2006). Vertical bars represent the standard deviations of the arithmetic mean of two replications. The Ca was supplied as  $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$  and the Cu as  $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ .

## Supplementary data

Supplementary data are available at *JXB* online.

**Supplementary Table S1.** Data set collected from the literature (1975–2009) for solution culture studies examining the phytotoxicity of As, Cd, Co, Cu, Hg, Mn, Ni, Pb, and Zn.

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## References

- Adams F, Burmester C, Hue NV, Long FL.** 1980. A comparison of column-displacement and centrifuge methods for obtaining soil solutions. *Soil Science Society of America Journal* **44**, 733–735.
- Agbenin JO.** 2003. Soil saturation extract composition and sulfate solubility in a tropical semiarid soil. *Soil Science Society of America Journal* **67**, 1133–1139.
- Aguirre-Gomez A, McBride MB, Norvell WA.** 2006. A voltammetric method for determining free metal activities in aqueous solutions. 2. Anodic stripping voltammetry of Cd, Cu, Pb, and Zn in synthetic and soil solutions. *International Journal of Environment and Pollution* **26**, 68–89.
- Anderson AJ, Meyer DR, Mayer FK.** 1973. Heavy metal toxicities: levels of nickel, cobalt, and chromium in soil and plants associated with visual symptoms and variation in growth of an oat crop. *Australian Journal of Agricultural Research* **24**, 557–571.
- Asher CJ, Reay PF.** 1979. Arsenic uptake by barley seedlings. *Australian Journal of Plant Physiology* **6**, 459–466.
- Babula P, Adam V, Opatrilova R, Zehnalek J, Havel L, Kizek R.** 2008. Uncommon heavy metals, metalloids and their plant toxicity: a review. *Environmental Chemistry Letters* **6**, 189–213.
- Batianoff GN, Singh S.** 2001. Central Queensland serpentine landforms, plant ecology and endemism. *South African Journal of Science* **97**, 495–500.
- Bell RW, Edwards DG, Asher CJ.** 1989. External calcium requirements for growth and nodulation of six tropical food legumes grown in flowing solution culture. *Australian Journal of Agricultural Research* **40**, 85–96.
- Blamey FPC, Nishizawa NK, Yoshimura E.** 2004. Timing, magnitude, and location of initial soluble aluminium injuries to mungbean roots. *Soil Science and Plant Nutrition* **50**, 67–76.
- Bruce RC, Warrell LA, Bell LC, Edwards DG.** 1989. Chemical attributes of some Queensland acid soils. I. Solid and solution phase compositions. *Australian Journal of Soil Research* **27**, 333–351.
- Burström H.** 1953. Physiology of root growth. *Annual Review of Plant Physiology* **4**, 237–252.
- CEI.** 2005. *Soil remediation technologies: assessment, clean-up, decommissioning, rehabilitation.* Canadian Environmental Industries (Energy and Environmental Industries Branch), available at: <http://www.ic.gc.ca/eic/site/ea-ae.nsf/eng/ea02201.html>.
- Charpentier S, Garnier J, Flaugnatti R.** 1987. Toxicity and bioaccumulation of cadmium in experimental cultures of duckweed, *Lemna polyrrhiza* L. *Bulletin of Environmental Contamination and Toxicology* **38**, 1055–1061.
- Chi Yu C, Tung Hung K, Huei Kao C.** 2005. Nitric oxide reduces Cu toxicity and Cu-induced  $\text{NH}_4^+$  accumulation in rice leaves. *Journal of Plant Physiology* **162**, 1319–1330.
- Clemens S.** 2006. Toxic metal accumulation, responses to exposure and mechanisms of tolerance in plants. *Biochimie* **88**, 1707–1719.
- de Vos CHR, Schat H, Dewaal MAM, Vooijs R, Ernst WHO.** 1991. Increased resistance to copper-induced damage of the root cell plasmalemma in copper tolerant *Silene cucubalus*. *Physiologia Plantarum* **82**, 523–528.
- Degryse F, Waegeneers N, Smolders E.** 2007. Labile lead in polluted soils measured by stable isotope dilution. *European Journal of Soil Science* **58**, 1–7.
- del Amor FM, Marcelis LFM.** 2003. Regulation of nutrient uptake, water uptake and growth under calcium starvation and recovery. *Journal of Horticultural Science and Biotechnology* **78**, 343–349.
- Edmeades DC, Wheeler DM, Clinton OE.** 1985. The chemical composition and ionic strength of soil solutions from New Zealand topsoils. *Australian Journal of Soil Research* **23**, 151–165.
- Edwards DG, Asher CJ.** 1982. Tolerance of crop and pasture species to manganese toxicity. In: Scaife A, ed. *Proceedings of the ninth international plant nutrition colloquium.* Warwick University, England: Commonwealth Agricultural Bureaux, 145–150.
- Gillman GP, Bell LC.** 1978. Soil solution studies on weathered soils from tropical North Queensland. *Australian Journal of Soil Research* **16**, 67–77.
- Godbold DL.** 1991. Mercury-induced root damage in spruce seedlings. *Water, Air, and Soil Pollution* **56**, 823–831.
- Goldbach HE, Yu Q, Wingender R, Schulz M, Wimmer M, Findekle P, Baluska F.** 2001. Rapid response reactions of roots to boron deprivation. *Journal of Plant Nutrition and Soil Science-Zeitschrift Fur Pflanzenernahrung Und Bodenkunde* **164**, 173–181.
- Hoagland DR, Arnon DI.** 1950. *The water-culture method for growing plants without soil.* Berkeley, California: University of California.
- Israr M, Sahi S, Datta R, Sarkar D.** 2006. Bioaccumulation and physiological effects of mercury in *Sesbania drummondii*. *Chemosphere* **65**, 591–598.
- Jensen GH.** 1907. Toxic limits and stimulation effects of some salts and poisons on wheat. *Botanical Gazette* **43**, 11–44.
- Kinraide TB.** 1998. Three mechanisms for the calcium alleviation of mineral toxicities. *Plant Physiology* **118**, 513–520.
- Kinraide TB.** 2006. Plasma membrane surface potential ( $\psi_{PM}$ ) as a determinant of ion bioavailability: a critical analysis of new and



published toxicological studies and a simplified method for the computation of plant  $\psi_{PM}$ . *Environmental Toxicology and Chemistry* **25**, 3188–3198.

**Kopittke PM, Asher CJ, Menzies NW.** 2008a. Prediction of Pb speciation in commonly used nutrient solutions. *Environmental Pollution* **153**, 548–554.

**Kopittke PM, Blamey FPC, Menzies NW.** 2008b. Toxicities of soluble Al, Cu, and La include ruptures to rhizodermal and root cortical cells of cowpea. *Plant and Soil* **303**, 217–227.

**Kopittke PM, McKenna BA, Blamey FPC, Wehr JB, Menzies NW.** 2009. Metal-induced cell rupture in elongating roots is associated with metal ion binding strengths. *Plant and Soil* **322**, 303–315.

**Kovar JL, Barber SA.** 1988. Phosphorus supply characteristics of 33 soils as influenced by 7 rates of phosphorus addition. *Soil Science Society of America Journal* **52**, 160–165.

**Krantev A, Yordanova R, Janda T, Szalai G, Popova L.** 2008. Treatment with salicylic acid decreases the effect of cadmium on photosynthesis in maize plants. *Journal of Plant Physiology* **165**, 920–931.

**Larbi A, Morales F, Abadia A, Gogorcena Y, Lucena JJ, Abadia J.** 2002. Effects of Cd and Pb in sugar beet plants grown in nutrient solution: induced Fe deficiency and growth inhibition. *Functional Plant Biology* **29**, 1453–1464.

**Lee D-Y, Fortin C, Campbell PGC.** 2005. Contrasting effects of chloride on the toxicity of silver to two green algae, *Pseudokirchneriella subcapitata* and *Chlamydomonas reinhardtii*. *Aquatic Toxicology* **75**, 127–135.

**Lock K, Criel P, De Schamphelaere KAC, Van Eeckhout H, Janssen CR.** 2007a. Influence of calcium, magnesium, sodium, potassium and pH on copper toxicity to barley (*Hordeum vulgare*). *Ecotoxicology and Environmental Safety* **68**, 299–304.

**Lock K, Van Eeckhout H, De Schamphelaere KAC, Criel P, Janssen CR.** 2007b. Development of a biotic ligand model (BLM) predicting nickel toxicity to barley (*Hordeum vulgare*). *Chemosphere* **66**, 1346–1352.

**Luo XS, Zhou DM, Wang YJ.** 2006. Free cupric ions in contaminated agricultural soils around a copper mine in eastern Nanjing City, China. *Journal of Environmental Sciences (China)* **18**, 927–931.

**Malone C, Koeppe DE, Miller RJ.** 1974. Localization of lead accumulated by corn plants. *Plant Physiology* **53**, 388–394.

**Menzies NW, Bell LC.** 1988. Evaluation of the influence of sample preparation and extraction technique on soil solution composition. *Australian Journal of Soil Research* **26**, 451–464.

**Mourato M, Martins L, Campos-Andrada M.** 2009. Physiological responses of *Lupinus luteus* to different copper concentrations. *Biologia Plantarum* **53**, 105–111.

**Norvell WA.** 1988. Inorganic reactions of manganese in soils. In: Graham RD, Hannam RJ, Uren NC, eds. *Manganese in soils and plants*. Dordrecht, The Netherlands: Kluwer Academic Publishers, 37–58.

**Parker DR, Norvell WA.** 1999. Advances in solution culture methods for plant mineral nutrition research. *Advances in Agronomy* **65**, 151–213.

**Parkhurst D.** 2009. *Phreeqc v2.15.00*. United States Geological Survey. <http://water.usgs.gov/software/> (Accessed March 2009).

**Pedler JF, Kinraide TB, Parker DR.** 2004. Zinc rhizotoxicity in wheat and radish is alleviated by micromolar levels of magnesium and potassium in solution culture. *Plant and Soil* **259**, 191–199.

**Proctor J, Johnston WR, Cottam DA, Wilson AB.** 1981. Field-capacity water extracts from serpentine soils. *Nature* **294**, 245–246.

**Reichman SM, Menzies NW, Asher CJ, Mulligan D.** 2004. Seedling responses of four Australian tree species to toxic concentrations of manganese in solution culture. *Plant and Soil* **258**, 341–350.

**Reisenauer HM.** 1966. Mineral nutrients in soil solution. In: Altman PL, Dittmer DS, eds. *Environmental biology*. Bethesda, MD: Federation of American Societies for Experimental Biology, 507–508.

**Rengel Z.** 1996. Tansley review No. 89. Uptake of aluminium by plant cells. *New Phytologist* **134**, 389–406.

**Rengel Z.** 2002. *Handbook of plant growth: pH as the master variable*. New York: Marcel Dekker.

**Sahi SV, Israr M, Srivastava AK, Gardea-Torresdey JL, Parsons JG.** 2007. Accumulation, speciation and cellular localization of copper in *Sesbania drummondii*. *Chemosphere* **67**, 2257–2266.

**Spehar CR, Galwey NW.** 1997. Screening soya beans [*Glycine max* (L.) Merrill] for calcium efficiency by root growth in low-Ca nutrient solution. *Euphytica* **94**, 113–117.

**Sumner ME, Noble AD.** 2003. Soil acidification: the world story. In: Rengel Z, ed. *Handbook of soil acidity*. New York, USA: Marcel Dekker, 1–28.

**Tamaki S, Frankenberger WT.** 1992. Environmental biochemistry of arsenic. *Reviews of Environmental Contamination and Toxicology* **124**, 79–110.

**Taylor GJ, Blamey FPC, Edwards DG.** 1998. Antagonistic and synergistic interactions between aluminum and manganese on growth of *Vigna unguiculata* at low ionic strength. *Physiologia Plantarum* **104**, 183–194.

**Taylor GJ, Foy CD.** 1985. Differential uptake and toxicity of ionic and chelated copper in *Triticum aestivum*. *Canadian Journal of Botany - Revue Canadienne De Botanique* **63**, 1271–1275.

**Taylor GJ, Stadt KJ, Dale MRT.** 1991. Modeling the phytotoxicity of aluminum, cadmium, copper, manganese, nickel, and zinc using the Weibull frequency-distribution. *Canadian Journal of Botany* **69**, 359–367.

**Weng L, Temminghoff EJM, Van Riemsdijk WH.** 2001. Determination of the free ion concentration of trace metals in soil solution using a soil column Donnan membrane technique. *European Journal of Soil Science* **52**, 629–637.

**Weng LP, Lexmond TM, Wolthoorn A, Temminghoff EJM, Van Riemsdijk WH.** 2003. Phytotoxicity and bioavailability of nickel: chemical speciation and bioaccumulation. *Environmental Toxicology and Chemistry* **22**, 2180–2187.

**Wheeler DM, Edmeades DC.** 1995. Effect of depth and lime or phosphorus-fertilizer applications on the soil solution chemistry of some New Zealand pastoral soils. *Australian Journal of Soil Research* **33**, 461–476.

**Wheeler DM, Power IL, Edmeades DC.** 1993. Effect of various metal ions on growth of two wheat lines known to differ in aluminium tolerance. *Plant and Soil* **155/156**, 489–492.

**Wiklander L, Andersson A.** 1974. The composition of the soil solution as influenced by fertilization and nutrient uptake. *Geoderma* **11**, 157–166.

**Wong MH, Bradshaw AD.** 1982. A comparison of the toxicity of heavy metals, using root elongation of rye grass, *Lolium perenne*. *New Phytologist* **91**, 255–261.

**Yildiz M, Cigerci IH, Konuk M, Fatih Fidan A, Terzi H.** 2009. Determination of genotoxic effects of copper sulphate and cobalt chloride in *Allium cepa* root cells by chromosome aberration and comet assays. *Chemosphere* **75**, 934–938.

**Zeid IM.** 2001. Responses of *Phaseolus vulgaris* to chromium and cobalt treatments. *Biologia Plantarum* **44**, 111–115.

**Zhu YG, Chen SB, Yang JC.** 2004. Effects of soil amendments on lead uptake by two vegetable crops from a lead-contaminated soil from Anhui, China. *Environment International* **30**, 351–356.