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Transcriptome analysis of drough-tolerant CAM plants, Agave deserti and Agave tequilana

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ABSTRACT

Agaves are succulent monocotyledonous plants native to hot and arid environments of North America. Because of their adaptations to their environment, including crassulacean acid metabolism (CAM, a water-efficient form of photosynthesis) and existing technologies for ethanol production, agaves have gained attention both as potential lignocellulosic bioenergy feedstocks and models for exploring plant responses to abiotic stress. However, the lack of comprehensive Agave sequence datasets limits the scope of investigations into the molecular-genetic basis of Agave traits. Here, we present comprehensive, high quality de novo transcriptome assemblies of two Agave species, A. tequilana and A. deserti, from short-read RNA-seq data. Our analyses support completeness and accuracy of the de novo transcriptome assemblies, with each species having approximately 35,000 protein-coding genes. Comparison of agave proteomes to those of additional plant species identifies biological functions of gene families displaying sequence divergence in agave species. Additionally, we use RNA-seq data to gain insights into biological functions along the A. deserti juvenile leaf proximal-distal axis. Our work presents a foundation for further investigation of agave biology and their improvement for bioenergy development.

CAM PHOTOSYNTHESIS, ARID ENVIRONMENTS, AND BIOENERGY

Agave species are adapted to their native habitat in arid regions of Mexico and the United States. Agave thus holds promise as a biofuel feedstock [1,2], capable of growing on marginal lands where other proposed bioenergy The ability of agaves to withstand hot and arid conditions relies upon crassulacean acid metabolism (CAM)—a specialized form of photosynthesis allowing agaves to keep leaf stomata (pores) closed during the hot day, minimizing water loss through evapotranspiration.

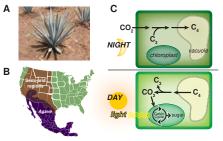


FIGURE 1: Agaves and CAM biology (A) Agave tequilana cultivated in Mexico.

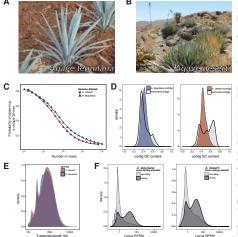
(B) Semi-arid regions of the United States (brown) are unsuitable for cultivation of other bioenergy plants, which require more temperate (green). Most Agave species regions adapted to semi-arid regions in Mexico and the extreme southwestern USA (purple).

(C) Crassulacean Acid Metabolism (CAM) CO (C) Crassulacean Acid Metabolism (GAM). CO₂ enters plant cells at night, joins with a 3-carbon molecule (C₂) and is stored in the vacuole as a 4-carbon molecule (C). During the day, C₄ molecules diffuse out of the vacuole, and CO₂ is relased and assimilated into sugar in the chloroplast.

	Inputs			Outputs		
Feedstock	Water (cm yr ⁻¹)	Drought tolerance	Nitrogen (kg ha ⁻¹ yr ⁻¹)	Dry biomass (Mg ha ^{.1} yr ^{.1})	Ethanol (liters yr ⁻¹)	
Corn grain	50-80	low	90-120	7–10	2900	
Corn stover	50 00	1011	50 120	3–6	900	
Miscanthus	75-120	low	0-15	15-40	4600-12,400	
Poplar coppice	70-105	moderate	0-50	5-11	1500-3400	
Agave spp.	3080	high	0-12	10-34	3000-10,500	

Comparison of inputs (water and nitrogen) and outputs (biomass and ethanol) of agaves and other biofuel feedstock species. Though agaves are harvested at several years of age, their annualized growth rate is on pa with *Miscanthus*. Table is modified from reference [2].

AGAVE TRANSCRIPTOME ASSEMBLIES FROM DEEP RNA-seq



To provide sequence resources for the Agave research community, we built de novo transcriptomes of Agave tequilana and Agave deserti from Illumina RNA-seq data. deep Sequences were assembled by Rnnotator [3], a *de novo* transcriptome assembly pipeline.

FIGURE 2: A. tequilana, A. deserti, and their respective transcriptomes

(A) Cultivated A. tequilana in Jalisco, Mexico

(B) A. deserti (foreground) in natural habitat, Riverside County, California, USA. (C) Plot of the fraction of unique 25-mers over

ndicated read depth (log2 scale).

(D) Density plot of GC content of agave transcript contigs vs. contigs from contamination and commensal organisms. (E) Density plots of A. deserti and A. tequilana transcript lengths. Note \log_{10} scale. Peaks at 150 and 250 nt represent single reads or paired-end reads, respectively, that were not

mbled into larger contigs (F) Density plot of locus RPKM values for coding (dark shading) and non-coding (light shading) loci.

OVERVIEW OF AGAVE TRANSCRIPTOME ASSEMBLIES

Species	Total Sequencing	No. of loci	No. transcript contigs	N50 length	Sum assembled length	No. protein-coding loci
A. tequilana	293.5 Gbp	139,525	204,530	1387 bp	204.9 Mbp	34,870
A. deserti	184.7 Gbp	88,718	128,869	1323 bp	125.0 Mbp	35,086



and accurate

Rnnotator assemblies

Analysis of assembled contigs suggest the Agave de novo

assemblies are comprehensive

FIGURE 3: Comparison of the de novo Agave transcriptome assemblies

(A) Comparisons of the A. tequilana de novo Rnnotator assembly to error corrected Pacific Biosciences subreads,

82 GenBank A. tequilana sequences

and an additional A. tequilana dataset from McKain et al. 2012. [4]

(B) Comparisons between the A. tequilana and A. deserti de novo

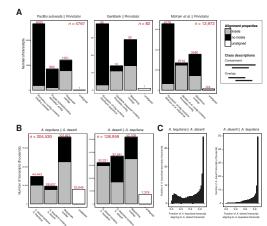
(C) Histograms of the fraction of aligned sequence lengths between A. deserti and A. tequilana.

Symbol || separates query sequence

dataset from subject sequence dataset. Total number of sequences (n) is noted in

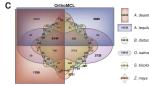
chart, total number sequences in alignment classes are noted above bar.

COMPARISON OF AGAVE DE NOVO ASSEMBLIES



PROTEOMIC ANALYSES SUPPORT COMPREHENSIVE AGAVE TRANSCRIPTOME ASSEMBLIES





Proteome comparisons between Agave species and additional monocot species suggest the majority of Agave proteins are conserved across taxa. We can also identify protein families specific to agaves

each bar

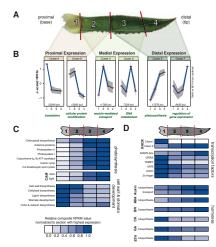
FIGURE 4: Proteomic comparison of agaves to other plant species

(A) Venn diagram of BLASTP-based one-to-one reciprocal best hit proteins shared between A. deserti and A. tequilana.

(B) Venn diagram of OrthoMCL-defined protein families shared between agaves

(C) Edwards-Venn diagram of OrthoMCL-defined plant orthologous-group protein families (Plant OCs) shared between agave and 4 additional monocotyledonous plant species. Shape and color used for each species is at the right with the total number of Plant OGs within each species.

PROFILING OF THE A. DESERTI LEAF HIGHLIGHTS REGIONS CRITICAL TO DEVELOPMENT AND PHOTOSYNTHESIS



Agaves spend the majority of their lives as compact rosettes, thus leaves are important organs in which to study Agave developmental and bioenergetic processes.

FIGURE 5: Transcriptomic analysis of the *A. deserti* leaf proximal-distal axis. f proxin

(A) One of the A. deserti leaves used for analysis, indicating proximal-distal (PD) sections 1–4.

(B) Six major K-means clusters of gene expression along the PD axis. Clusters are manually grouped by highest expression in proximal, medial, or distal tissues Blue lines connect mean z-scaled RPKM values, shaded areas represent the 25th and 75th percentiles, red lines indicate standard error at each mean. Green text beneath each cluster denotes the description of the most significantly enriched GO term in each cluster.

(C, D) Heatmaps of composite gene expression for indicated biological processes along the leaf PD axis.

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