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Article

Trichophycin A, a Cytotoxic Linear Polyketide Isolated from a *Trichodesmium thiebautii* Bloom

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Abstract: In an effort to isolate and characterize bioactive secondary metabolites from *Trichodesmium thiebautii* blooms, collected cyanobacteria biomass was subjected to bioassay-guided extraction and fractionation using the human colon cancer cell line HCT-116, resulting in the isolation and subsequent structure characterization of a linear polyketide trichophycin A (**1**). The planar structure of **1** was completed using 1D and 2D NMR spectroscopy and high-resolution electrospray ionization mass spectrometry (HRESIMS). Trichophycin A was moderately toxic against the murine neuroblastoma cell line Neuro-2A (EC₅₀: 6.5 μM) and HCT-116 cells (EC₅₀: 11.7 μM). Trichophycin A was significantly more cytotoxic than the previously isolated polyketides trichotoxin A and trichotoxin B. These cytotoxicity observations suggest that toxicity may be related to the polyol character of these polyketide compounds.

Keywords: *Trichodesmium thiebautii* blooms; polyketide; polyol; secondary metabolite

1. Introduction

Many biosynthetic systems in marine cyanobacteria allow for the incorporation of peptide and polyketide subunits resulting in remarkable structural diversity [1,2]. These resultant hybrid non-ribosomal peptide synthetase-polyketide synthase NRPS-PKS molecules have remained a frequently isolated secondary metabolite class from cyanobacteria [3] and they have shown diverse and therapeutically relevant biological activities [4–7]. Macrocyclic and linear polyketides have been isolated from cyanobacteria with less regularity than NRPS and NRPS-PKS molecules. Still, many of these polyketides have exhibited a diverse array of biological activities [8–10] and possess highly functionalized structural elements [11–13] demonstrating the importance of their continued isolation and biological evaluation.

Trichodesmium thiebautii, a marine filamentous cyanobacterium of the order *Oscillatoriales*, is globally significant both for its biogeochemical role in N₂ fixation [14] and the biological community associated with the “floating islands” formed by dense blooms that can cover many kilometers of surface waters [15]. *Trichodesmium* blooms occur over most of the subtropical and tropical oceans with the greatest concentration in the eastern tropical Pacific Ocean and the Arabian Sea [16]. Repeated blooms occur in the eastern and western Gulf of Mexico from February through August [17]. These blooms have been hypothesized to contribute the nitrogen required for the initiation of *Karenia brevis* blooms in the Gulf of Mexico [18]. Studies carried out using homogenized cells,

filtrates, aging cultures, and crude extracts of *Trichodesmium thiebautii* filaments have shown toxicity to copepods [19,20]. However, establishing uni-algal, actively growing cultures of *T. thiebautii* has been problematic and has limited the ability to characterize these toxic molecules [21]. Thus, members of the cyanobacterial genus *Trichodesmium* remain an under-sampled group with few metabolites described from the genus. The cyclic peptide trichamide was characterized from a cultured strain of *Trichodesmium erythraeum* [22]. The lipoamides, credneramides A and B, were isolated and characterized from a field-collected benthic cyanobacterium. This specimen showed a phylogenetic relationship to other *Trichodesmium* strains and may be a new species [23]. A chlorinated metabolite, trichotoxin, displaying some cytotoxicity was isolated from a field sample of *Trichodesmium thiebautii* [24]. Recently, our laboratory has revised the structure of trichotoxin, renaming it trichotoxin A, and have isolated and characterized an alkyne-containing analog trichotoxin B [25].

Herein, we report the isolation and structure characterization of trichophycin A (**1**), a linear triol polyketide. This molecule possesses, presumably, an aromatic biosynthetic starter unit, while the remainder of the molecule is recognizably derived from a PKS system, likely incorporating acetate extensions, the last of which we predict would undergo decarboxylation resulting in terminal olefin formation. Trichophycin A is structurally similar to the compounds trichotoxin A and B [25], which were also isolated from a *T. thiebautii* bloom. Trichophycin A (**1**) showed moderate cytotoxicity against Neuro-2A murine neuroblastoma cells and HCT-116 human colon cancer cells with EC₅₀ values of 6.5 ± 1.4 μM and 11.7 ± 0.6 μM, respectively.

2. Results

2.1. Structure Elucidation of **1**

Bioassay-guided fractionation of the *T. thiebautii* extract and chemical fractions against HCT-116 cells led to the isolation of **1** (Figure 1), an optically active pale yellow oil. HRESIMS analysis of **1** gave an [M + H]⁺ of *m/z* 479.3282, suggesting a molecular formula of C₂₉H₄₇ClO₃ and a requirement of 6 degrees of unsaturation. Examination and comparison of the ¹³C-NMR, HSQC and HMBC spectra of **1** showed the presence of 3 methyl groups, 11 methylenes, 13 methines, and 2 quaternary carbon atoms (Figures S2–S4).

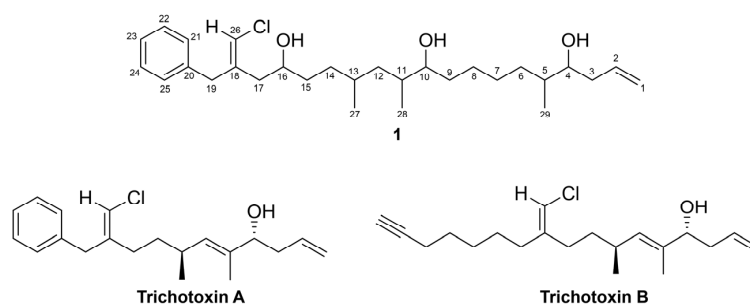


Figure 1. Structures of trichophycin A (**1**), trichotoxin A, and trichotoxin B.

The ¹³C-NMR spectra of **1** showed the characteristic signals of a phenyl group for positions C-20 to C-25 (δ_C 138.0, 129.0, 128.6, 126.7, 128.6, 129.0). A deshielded methylene (H₂-19, δ_H 3.46) showed HMBC correlations to the quaternary carbon of the aromatic ring (C-20, δ_C 138.0), and C-21 and C-25 on the aromatic ring (δ_C 129.0). Additionally, the H₂-19 methylene showed HMBC correlations to a moderately polarized olefin comprised of a quaternary carbon (C-18, δ_C 139.5) and a carbon (C-26, δ_C 116.2) bearing a singlet methine proton at δ_H 5.99. These chemical shifts were consistent with the presence of a vinyl chloride functionality, and considering the aromatic moiety, accounted for 5 of the 6 degrees of unsaturation required. NOE correlations between the H-26 (δ_H 5.99) and H-21 (δ_H 7.17) and H₂-19 (δ_H 3.46) supported a *Z* configuration of the vinyl chloride (Figure S7). The H₂-19 methylene showed HMBC correlation to C-17 (δ_C 38.2). The HSQC showed C-17 was correlated to

deshielded methylene protons (δ_{H} 2.38, 2.28). This second group of deshielded methylene protons (H₂-17) showed HMBC correlations to the vinyl chloride-containing olefin and to an oxygen-bearing carbon (C-16, δ_{C} 70.7). Examination of the COSY spectrum (see Figure S5) showed that the oxymethine (H-16, δ_{H} 3.79) was coupled to a methylene group (H₂-15 δ_{H} 1.52 and 1.44). These H₂-15 protons showed COSY correlations to the H₂-14 methylene (δ_{H} 1.35 and 1.22). The H₂-14 methylene protons showed HMBC correlations to C-13 (δ_{C} 29.8), C-12 (δ_{C} 40.8), and the C-27 methyl group (δ_{C} 20.4). The H-13 methine (δ_{H} 1.52) showed a COSY correlation to H₃-27 (δ_{H} 0.87). The H₂-12 methylene protons (δ_{H} 1.39, 1.00) showed HMBC correlations to a second tertiary carbon (C-11, δ_{C} 35.2) bearing an adjacent methyl group (C-28, δ_{C} 13.9) and an HMBC correlation to a second oxygen-bearing carbon (C-10, δ_{C} 74.5). Bidirectional HMBC correlations established four methylene groups between C-10 and C-5. The H-5 methine showed COSY correlations to H-6b (δ_{H} 1.19), H₃-29 (δ_{H} 0.90), and the H-4 oxymethine (δ_{H} 3.55). The oxymethine showed HMBC correlations to C-3 (δ_{C} 39.1) and C-2 (δ_{C} 135.6). A COSY correlation between H-2 (δ_{H} 5.83) and H₂-1 (δ_{H} 5.15, 5.12) established a monosubstituted terminal olefin functionality, satisfied the final degree of unsaturation, and demonstrated that **1** was a linear polyketide. Selected 2D correlations utilized in determining the planar structure of **1** are shown in Figure 2. Examination of the TOCSY spectrum supported COSY correlations (Figure S6).

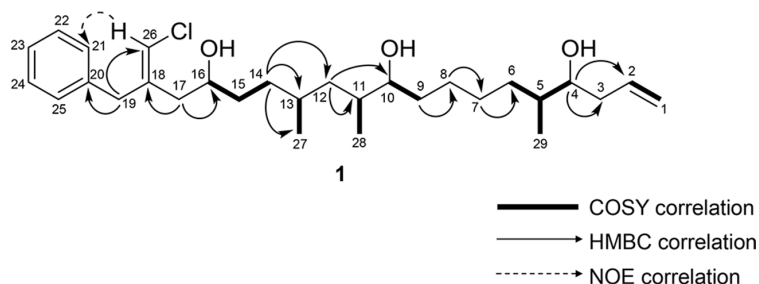


Figure 2. Key COSY, HMBC, and NOE correlations of **1**.

2.2. Cytotoxicity of Trichophycin A

Trichophycin A (**1**) showed moderate cytotoxicity against selected cells lines with EC_{50} values against Neuro-2A cells and HCT-116 cells of $6.5 \pm 1.4 \mu\text{M}$ and $11.7 \pm 0.6 \mu\text{M}$, respectively (Figure 3). However, there was no significant selectivity between the cell lines tested (*t*-test, $p > 0.05$). The EC_{50} values for trichotoxins A and B were greater than $50 \mu\text{M}$ against Neuro-2A cells and greater than $100 \mu\text{M}$ against HCT-116 cells (See Figure S8).

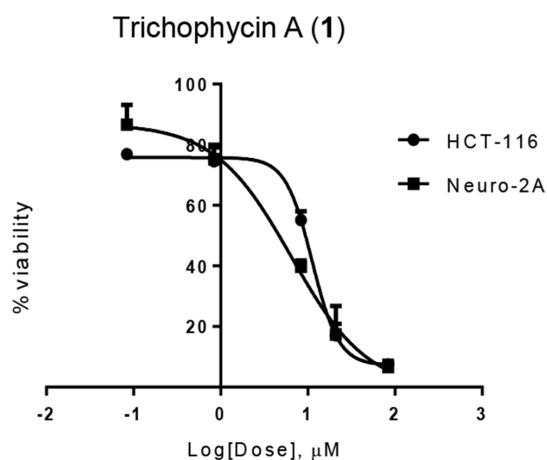


Figure 3. Dose-response curves of trichophycin A (**1**) tested against Neuro-2A murine neuroblastoma cells and HCT-116 human colon cancer cells.

3. Discussion

Trichophycin A (**1**) represents a new addition to the diverse group of vinyl chloride-containing linear polyketides and polyketide-peptides isolated from cyanobacteria environmental collections. This group includes the jamaicamides [26], the credneramides [23], the kimbeamides [11], coibacin C and D [12], and pitiamides A and B [27] among others. We would predict that the biosynthesis of the vinyl chloride in trichophycin A would follow that of the jamaicamides in which an acetate-derived vinyl group is generated through the action of a hydroxymethylglutaryl-CoA (HMG-CoA) synthase cassette and subsequently halogenated [26]. Trichophycin A shows several structural similarities to trichotoxin A and B [25]. All possess vinyl chloride functionalities and terminal alkenes. Trichophycin A (**1**) features a longer polyketide chain than trichotoxin A with two additional secondary alcohol groups and one additional branched methyl. This increased polyol character may be a reason for the increased cytotoxicity of **1** ($EC_{50} = 6.5 \mu\text{M}$) compared to that originally reported for trichotoxin A ($LC_{50} = 106 \mu\text{M}$) against Neuro-2A cells [24] and this study ($EC_{50} > 50 \mu\text{M}$ against Neuro-2A cells). The number of free hydroxyl groups is critical to the antiproliferative effect of the macrocyclic polyketide amantelide A, as its peracetylated derivative was essentially inactive [9]. This effect was also observed with kalkipyron B and its acetylated derivative [8]. These modified products have decreased hydroxyl character. However, the acetyl groups add steric bulk which may interfere with receptor binding. Testing a comprehensive series of analogs with decreasing and increasing hydroxyl character will ultimately provide a more informative structure-activity relationship (SAR) study. Many of the previously mentioned vinyl chloride-containing compounds have shown neuromodulatory activity [11,23,26]. Trichophycin A (**1**) does show greater potency against Neuro-2A cells than HCT-116 cells. However, the difference in potency is not significant. Further evaluation of **1** against a diverse panel of eukaryotic cells will determine if trichophycin A is selective or if it is a broad-spectrum moderately toxic molecule. The amount of trichophycin A isolated (0.10% of dry weight of the cyanobacterial biomass) suggests this molecule may have an important ecological or physiological role for the organism. The complex methylene and methine envelope and overlapping signals from 1.20 to 1.70 ppm in the $^1\text{H-NMR}$ spectrum (see Figure S1) was not amenable to analysis following derivatization of the secondary alcohol groups in **1** with chiral shift reagents. Thus, absolute configuration analysis will necessitate total synthesis, X-ray diffraction analysis of a crystallized derivative or bioinformatic analysis of the ketoreductase (KR) domains in the trichophycin biosynthetic gene cluster. Ketoreductases generally control the configuration of β -hydroxy and secondary methyl groups in polyketide biosynthesis [28–30]. Work will continue to explore potential biological activities of **1** and search for structurally related analogs from *Trichodesmium* blooms in order to continue to evaluate the relationship between polyol character and cytotoxicity.

4. Materials and Methods

4.1. General Experimental Procedures

Optical rotations were measured using a Jasco P-2000 polarimeter (Jasco Inc., Easton, MD, USA). UV spectra were measured using a Beckman Coulter DU-800 spectrophotometer (Beckman Coulter Inc., Brea, CA, USA). NMR spectra were collected using a Bruker 800 MHz NMR instrument (Bruker, Rheinstetten, Germany) equipped with a cryoprobe. HRESIMS analysis was performed using a AB SCIEX TripleTOF 4600 mass spectrometer (SCIEX, Framingham, MA, USA) with Analyst TF software (version 1.7, SCIEX, Framingham, MA, USA). Semi-preparative HPLC was carried out using an Agilent 1100 series system equipped with a micro vacuum degasser, an autosampler, and a diode-array detector.

4.2. Biological Material

A localized bloom of *Trichodesmium* was collected from Padre Island, Corpus Christi, TX during 9–11 May 2014. Surface bloom material was collected in 5-gallon buckets from ca. 0.5-m water depth.

Approximately 300 g wet weight cell mass was concentrated from this material and frozen for further chemical analysis. In the laboratory, a subsample of the cell mass was examined microscopically and identified according to Komarek [31]. A preserved voucher sample of the biological material is kept in our laboratory with the identification number TTPI2014.

4.3. Extraction and Isolation of 1

Trichodesmium thiebautii filaments (14.4 g, dry weight), originally collected from a bloom near Padre Island, Corpus Christi, Texas in May of 2014, were extracted with five separate portions of 2:1 CH₂Cl₂-CH₃OH resulting in 3.95 g of crude extract. The crude residue was reconstituted in hexanes and fractionated over silica gel using vacuum liquid chromatography (VLC) using a stepped gradient of hexanes, EtOAc, and CH₃OH. The fractions eluting with 60% EtOAc in hexanes and 80% EtOAc in hexanes (Fractions E and F) were combined based on similarities in ¹H-NMR signals and similar potency in cytotoxicity assays. The combined material was fractionated over a 2 g Strata C18 SPE column eluting with 50% CH₃CN in H₂O, 100% CH₃CN, 100% CH₃OH, and 100% EtOAc. The fraction eluting with CH₃CN showed the most potent cytotoxicity and was subjected to reversed-phase HPLC using a YMC 5 μm ODS column (250 × 4.6 mm) with an elution solvent of 80% CH₃CN in H₂O with 0.1% formic acid added and trichophycin (**1**) (15 mg, rt: 9.25 min) was isolated.

Trichophycin A (1): pale yellow oil; α_D^{25} 4.3 (MeOH, *c* 0.20); UV (MeOH) λ_{\max} (log ϵ) 204 (4.0) nm; ¹H-NMR (800 MHz, CDCl₃); and ¹³C-NMR (200 MHz, CDCl₃), see Table 1; HRESIMS *m/z* 479.3282 [M + H]⁺ (calculated for C₂₉H₄₈ClO₃, 479.3292).

Table 1. NMR data for trichophycin A (**1**) (800 MHz, CDCl₃ for ¹H-NMR).

Pos	δ_C , Type	δ_H (J in Hz)	HMBC	COSY	NOESY
1a	117.9, CH ₂	5.15, m	2, 3	2	
1b		5.12, m	2, 3	2	
2	135.6, CH	5.83, m	3, 4	1a, 1b, 3a, 3b	
3a	39.1, CH ₂	2.26, m	1, 2, 4, 5	2, 3b, 4	29
3b		2.16, m	1, 2, 4, 5	2, 3a, 4	6b, 29
4	73.9, CH	3.55, dt (8.9, 4.0)	2, 3, 5, 6, 29	3a, 3b, 5	6b, 7b, 29
5	37.8, CH	1.53, ovlp ^a	4, 29	4, 6b, 29	3a, 3b
6a	33.0, CH ₂	1.48, m	4, 5, 7, 8, 29	6b, 7a, 7b	3b
6b		1.19, m	4, 5, 7, 8, 29	5, 6a	3b, 4
7a	27.4, CH ₂	1.38, ovlp	5, 6, 8		
7b		1.32, ovlp	6, 8		4
8a	26.7, CH ₂	1.44, ovlp	7, 9		
8b		1.32, ovlp	6, 7, 10		
9	34.6, CH ₂	1.44, ovlp	10		10
10	74.5, CH	3.50, m	8, 9, 11, 12, 28	9, 11	12b, 13, 28
11	35.2, CH	1.59, m	9, 10, 12, 13, 28	10, 12a, 12b, 28	
12a	40.8, CH ₂	1.39, ovlp	10, 11, 13, 14, 27, 28	12b	
12b		1.00, m	10, 11, 13, 14, 27, 28	11, 12a	10
13	29.8, CH	1.52, ovlp	12, 14, 27	12b, 14b, 27	10, 16
14a	32.5, CH ₂	1.35, ovlp	12, 13, 15, 16, 27	14b, 15b	16, 17b
14b		1.22, m	12, 13, 15, 16, 27	14a, 15b	16, 17b
15a	34.8, CH ₂	1.52, ovlp	13, 14, 16, 17	14a, 14b, 16	17a, 17b
15b		1.44, ovlp	13, 14, 16, 17	14b, 16	17a, 17b
16	70.7, CH	3.79, m	14, 15, 17, 18	15a, 15b, 17a, 17b	4a, 13, 14b, 27
17a	38.2, CH ₂	2.38, dd (13.7, 8.8)	15, 16, 18, 19, 26	16, 17b	15a, 15b, 19
17b		2.28, dd (13.7, 4.2)	15, 16, 18, 19, 26	16, 17a	15a, 15b, 19
18	139.5, qC				
19	41.9, CH ₂	3.46, d (5.5)	17, 18, 20, 21, 25, 26		17a, 17b, 26
20	138.0, qC				
21	129.0, CH	7.17, d (7.6)	19, 22, 23	22	19, 26
22	128.6, CH	7.30, t (7.6)	20	21	
23	126.7, CH	7.23, t (7.6)	22, 24	22, 24	

Table 1. Cont.

Pos	δ_C , Type	δ_H (J in Hz)	HMBC	COSY	NOESY
24	128.6, CH	7.30, t (7.6)	20	25	
25	129.0, CH	7.17, d (7.6)	19, 23, 24	24	19
26	116.2, CH	5.99, s	16, 17, 18, 19		19, 21
27	20.4, CH ₃	0.87, d (6.6)	12, 13, 14	13	10, 11, 14b, 16
28	13.9, CH ₃	0.84, d (6.8)	10, 11, 12	11	9, 10
29	13.9, CH ₃	0.90, d (6.8)	4, 5, 6	5	3a, 3b, 4, 6b

^a overlapping signals.

4.4. Cytotoxicity Assays

HCT-116 cells were added to 96-well plates in 100 μ L of McCoy's 5A media at a density of 2000 cells/well. Neuro-2A cells were added to assay plates in 100 μ L of Eagle's Minimum Essential Media (EMEM) supplemented with 10% fetal bovine serum at a density of 5000 cells/well. Cells were incubated overnight (37 $^{\circ}$ C, 5% CO₂) and examined microscopically to confirm confluence and adherence. Impure fractions were dissolved in CH₃OH (1% *v/v*) and tested at concentrations of 40 and 4 μ g/mL. Purified trichophycin A was dissolved in DMSO (1% *v/v*) and added to the cells in the range of 100, 10, 1, 0.1, and 0.01 μ M in order to construct a dose response curve. Three technical replicates were prepared for each concentration and each assay was performed in triplicate. Paclitaxel was used as a positive control (EC₅₀ = 3.1 nM against HCT-116 cells; EC₅₀ = 15.8 nM against Neuro-2A cells) and DMSO (1% *v/v*) was used as a negative control. Trichotoxin A and B were tested in the same manner. However, the concentrations tested were between 1 and 160 μ M. Trichotoxin A and B assays were run in duplicate. Plates were incubated for 72 h after which 15 μ L of MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium) dye were added each assay well. The dye was allowed to incubate with the cells for 4 h after which media was aspirated and the remaining formazan crystals were solubilized in 100 μ L of DMSO. Absorbance at 570 nm was measured using a Molecular Devices SpectraMax plate reader and EC₅₀ curves were generated and statistical procedures were performed using Graphpad Prism software.

Supplementary Materials: The following are available online at www.mdpi.com/1660-3397/15/1/10/s1, Figures S1–S7: 1H-NMR, 13C-NMR, HSQC, HMBC, COSY, TOCSY, NOESY data for compound 1. Figure S8: Cytotoxicity data of trichotoxin A and B against HCT-116 cells.

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Author Contributions: M.J.B., P.G.Z. and P.D.R.M. conceived the project. M.J.B. and P.G.W. performed the experiments. M.J.B. designed the experiments and wrote the paper; M.J.B., P.D.R.M., P.G.W. and H.H. analyzed the data; P.V.Z. contributed reagents/materials/analysis tools.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Kehr, J.C.; Picchi, D.G.; Dittmann, E. Natural product biosynthesis in cyanobacteria: A treasure trove of unique enzymes. *Beilstein J. Org. Chem.* **2011**, *7*, 1622–1635. [[CrossRef](#)] [[PubMed](#)]
- Calteau, A.; Fewer, A.; Latifi, A.; Coursin, T.; Laurent, T.; Jokela, J.; Kerfeld, C.A.; Sivonen, K.; Piel, J.; Gugger, M. Phylum-wide comparative genomics unravel the diversity of secondary metabolism in Cyanobacteria. *BMC Genom.* **2014**, *15*, 997. [[CrossRef](#)] [[PubMed](#)]
- Tan, L.T. Pharmaceutical agents from filamentous marine cyanobacteria. *Drug Discov. Today* **2013**, *18*, 863–871. [[CrossRef](#)] [[PubMed](#)]
- Pereira, A.; Kale, A.J.; Fenley, A.T.; Byrum, T.; Debonsi, H.M.; Gilson, M.K.; Valeriote, F.A.; Moore, B.S.; Gerwick, W.H. The carmaphyccins: New proteasome inhibitors exhibiting an α,β -epoxyketone warhead from a marine cyanobacterium. *ChemBioChem* **2012**, *13*, 810–817. [[CrossRef](#)] [[PubMed](#)]

5. Gutiérrez, M.; Pereira, A.R.; Deboni, H.M.; Ligresti, A.; Di Marzo, V.; Gerwick, W.H. Cannabinomimetic lipid from a marine cyanobacterium. *J. Nat. Prod.* **2011**, *74*, 2313–2317. [[CrossRef](#)] [[PubMed](#)]
6. Kwan, J.C.; Eksioğlu, E.A.; Liu, C.; Paul, V.J.; Luesch, H. Grassystatins A–C from marine cyanobacteria, potent cathepsin E inhibitors that reduce antigen presentation. *J. Med. Chem.* **2009**, *52*, 5732–5747. [[CrossRef](#)] [[PubMed](#)]
7. Luesch, H.; Yoshida, W.Y.; Moore, R.E.; Paul, V.J.; Corbett, T.H. Total structure determination of apratoxin A, a potent novel cytotoxin from the marine cyanobacterium *Lyngbya majuscula*. *J. Am. Chem. Soc.* **2001**, *123*, 5418–5423. [[CrossRef](#)] [[PubMed](#)]
8. Bertin, M.J.; Demirkiran, O.; Navarro, G.; Moss, N.A.; Lee, J.; Goldgof, G.M.; Vigil, E.; Winzeler, E.A.; Valeriote, F.A.; Gerwick, W.H. Kalkipyronone B, a marine cyanobacterial γ -pyrone possessing cytotoxic and anti-fungal activities. *Phytochemistry* **2016**, *122*, 113–118. [[CrossRef](#)] [[PubMed](#)]
9. Salvador-Reyes, L.A.; Sneed, J.; Paul, V.J.; Luesch, H. Amantelides A and B, polyhydroxylated macrolides with differential broad-spectrum cytotoxicity from a Guamanian marine cyanobacterium. *J. Nat. Prod.* **2015**, *78*, 1957–1962. [[CrossRef](#)] [[PubMed](#)]
10. Navarro, G.; Cummings, S.; Lee, J.; Moss, N.; Glukhov, E.; Valeriote, F.A.; Gerwick, L.; Gerwick, W.H. Isolation of polycavernoside D from a marine cyanobacterium. *Environ. Sci. Technol. Lett.* **2015**, *2*, 166–170. [[CrossRef](#)]
11. Nunnery, J.K.; Engene, N.; Byrum, T.; Cao, Z.; Jabba, S.V.; Pereira, A.R.; Matainaho, T.; Murray, T.F.; Gerwick, W.H. Biosynthetically intriguing chlorinated lipophilic metabolites from geographically distant tropical marine cyanobacteria. *J. Org. Chem.* **2012**, *77*, 4198–4208. [[CrossRef](#)] [[PubMed](#)]
12. Balunas, M.J.; Grosso, M.F.; Villa, F.A.; Engene, N.; McPhail, K.L.; Tidgewell, K.; Pineda, L.M.; Gerwick, L.; Spadafora, C.; Kyle, D.E.; et al. Coibacins A–D, antileishmanial marine cyanobacterial polyketides with intriguing biosynthetic origins. *Org. Lett.* **2012**, *14*, 5543–5554. [[CrossRef](#)] [[PubMed](#)]
13. Williamson, R.T.; Boulanger, A.; Vulpanovici, A.; Roberts, M.A.; Gerwick, W.H. Structure and absolute stereochemistry of phormidolide, a new toxic metabolite from the marine cyanobacterium *Phormidium* sp. *J. Org. Chem.* **2002**, *67*, 7927–7936. [[CrossRef](#)] [[PubMed](#)]
14. Karl, D.; Michaels, A.; Bergman, B.; Capone, D.; Carpenter, E.; Letelier, R.; Lipschultz, F.; Paerl, H.; Sigman, D.; Stal, L. Dinitrogen fixation in the world's oceans. *Biogeochemistry* **2002**, *57/58*, 47–98. [[CrossRef](#)]
15. Bergman, B.; Sandh, G.; Lin, S.; Larsson, J.; Carpenter, E.J. *Trichodesmium*—A widespread marine cyanobacterium with unusual nitrogen fixation properties. *FEMS Microbiol. Rev.* **2013**, *37*, 286–302. [[CrossRef](#)] [[PubMed](#)]
16. Westberry, T.K.; Siegel, D.A. Spatial and temporal distribution of *Trichodesmium* blooms in the world's oceans. *Glob. Biogeochem. Cycles* **2006**, *20*, GB4016. [[CrossRef](#)]
17. Mulholland, M.R.; Bernhardt, P.W.; Heil, C.A.; Bronk, D.A.; O'Neil, J.M. Nitrogen fixation and release of fixed nitrogen by *Trichodesmium* spp. in the Gulf of Mexico. *Limnol. Oceanogr.* **2006**, *51*, 1762–1776. [[CrossRef](#)]
18. Walsh, J.J.; Weisberg, R.H.; Lenes, J.M.; Chen, F.R.; Dieterle, D.A.; Zheng, L.; Carder, K.L.; Vargo, G.A.; Havens, J.A.; Peebles, E.; et al. Isotopic evidence for dead fish maintenance of Florida red tides, with implications for coastal fisheries over both source regions of the West Florida shelf and within downstream waters of the South Atlantic Bight. *Prog. Oceanogr.* **2009**, *80*, 51–73. [[CrossRef](#)]
19. Hawser, S.P.; O'Neil, J.M.; Roman, M.R.; Codd, G.A. Toxicity of blooms of the cyanobacterium *Trichodesmium* to zooplankton. *J. Appl. Phycol.* **1992**, *4*, 79–86. [[CrossRef](#)]
20. Guo, C.; Tester, P.A. Toxic effect of the bloom-forming *Trichodesmium* sp. (Cyanophyta) to the copepod *Acartia tonsa*. *Nat. Toxins* **1994**, *2*, 222–227. [[CrossRef](#)] [[PubMed](#)]
21. Ohki, K.; Fujita, Y. Laboratory culture of the pelagic blue-green alga *Trichodesmium thiebautii*: Conditions for unialgal culture. *Mar. Ecol. Prog. Ser.* **1982**, *7*, 185–190. [[CrossRef](#)]
22. Sudek, S.; Haygood, M.G.; Youssef, D.T.A.; Schmidt, E.W. Structure of trichamide, a cyclic peptide from the bloom-forming cyanobacterium *Trichodesmium erythraeum*, predicted from the genome sequence. *Appl. Environ. Microbiol.* **2006**, *72*, 4382–4387. [[CrossRef](#)] [[PubMed](#)]
23. Malloy, K.L.; Suyama, T.L.; Engene, N.; Deboni, H.; Cao, Z.; Matainaho, T.; Spadafora, C.; Murray, T.F.; Gerwick, W.H. Credneramides A and B: Neuromodulatory phenethylamine and isopentylamine derivatives of a vinyl chloride-containing fatty acid from cf. *Trichodesmium* sp. nov. *J. Nat. Prod.* **2012**, *75*, 60–66. [[CrossRef](#)] [[PubMed](#)]

24. Schock, T.B.; Huncik, K.; Beauchesne, K.R.; Villareal, T.A.; Moeller, P.D.R. Identification of trichotoxin, a novel chlorinated compound associated with the bloom forming cyanobacterium, *Trichodesmium thiebautii*. *Environ. Sci. Technol.* **2011**, *45*, 7503–7509. [[CrossRef](#)] [[PubMed](#)]
25. Bertin, M.J.; Zimba, P.V.; He, H.; Moeller, P.D.R. Structure revision of trichotoxin, a chlorinated polyketide isolated from a *Trichodesmium thiebautii* bloom. *Tetrahedron Lett.* **2016**, in press. [[CrossRef](#)]
26. Edwards, D.J.; Marquez, B.L.; Nogle, L.M.; McPhail, K.; Goeger, D.E.; Roberts, M.A.; Gerwick, W.H. Structure and biosynthesis of the jamaicamides, new mixed polyketide-peptide neurotoxins from the marine cyanobacterium *Lyngbya majuscula*. *Chem. Biol.* **2004**, *11*, 817–833. [[CrossRef](#)] [[PubMed](#)]
27. Cai, W.; Matthews, J.H.; Paul, V.J.; Luesch, H. Pitiamides A and B, multifunctional fatty acid amides from marine cyanobacteria. *Planta Med.* **2016**, *82*, 897–902. [[CrossRef](#)] [[PubMed](#)]
28. Keatinge-Clay, A.T.; Stroud, R.M. The structure of a ketoreductase determines the organization of the β -carbon processing enzymes of modular polyketide synthases. *Structure* **2006**, *14*, 737–748. [[CrossRef](#)] [[PubMed](#)]
29. Bonnett, S.A.; Whicher, J.R.; Papireddy, K.; Florova, G.; Smith, J.L.; Reynolds, K.A. Structural and stereochemical analysis of a modular polyketide synthase ketoreductase domain required for the generation of *cis*-alkene. *Chem. Biol.* **2013**, *20*, 772–783. [[CrossRef](#)] [[PubMed](#)]
30. Zheng, J.; Keatinge-Clay, A.T. The status of type I polyketide synthase ketoreductases. *Med. Chem. Commun.* **2013**, *4*, 34–40. [[CrossRef](#)]
31. Komárek, J.; Anagnostidis, K. *Cyanoprokarota 19 Part 2: Oscillatoriales*; Elsevier: München, Germany, 2005; pp. 1–759.



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