

# Journal of Colloid And Interface Science

## Triple-emission N, B co-doped carbon quantum dots from lignin: Highly fluorescent sensing platform for detection of Cr6+

--Manuscript Draft--

<b>Manuscript Number:</b>	
<b>Article Type:</b>	Full length article
<b>Section/Category:</b>	A. Colloidal Materials and Nanomaterials
<b>Keywords:</b>	Lignin; Carbon quantum dots; N, B co-Doped; Triple channel; Sensing Cr6+
<b>Corresponding Author:</b>	Dekui Shen Southeast University Nanjing, CHINA
<b>First Author:</b>	Lingli Zhu
<b>Order of Authors:</b>	Lingli Zhu Dekui Shen Kai Hong Luo
<b>Abstract:</b>	<p>Considering that Cr6+ with high toxicity poses huge threat to human health and ecological environment, the construction of a rapid and accurate fluorescence sensing platform is of great significance to detect the toxic substance. The N,B-CQDs from lignin waste are synthesized as fluorescent sensor for Cr6+ detection. The synthetic processes involve the acid hydrolysis step followed by the hydrothermal treatment. Lignin is firstly depolymerized by cleaving ether bonds under the action of acid, and N,B-CQDs are consequently formed by the aromatic re-fusion of lignin fragments in the carbonation process. The resultant N,B-CQDs show triple fluorescent emission of purple, blue and green color under the excitation of 300, 330, and 490 nm. The N,B-CQDs can be applied for the triple-channel detection of Cr6+, exhibiting highly sensitive and selective fluorescence quenching in the presence of Cr6+ with good linearity (<math>R^2 \leq 0.996</math>) and very low limit of detection as 0.054, 0.049 and 0.077 <math>\mu\text{M}</math> for Ex 300, 330 and 490 nm channel, respectively. The utilization of renewable lignin highlights the green, cheap and large-scale production of CQDs based fluorescent sensors and opens a new avenue for the rapid and accurate detection of Cr6+ through multichannel sensing platform.</p>
<b>Suggested Reviewers:</b>	Mingyue Ding Wuhan University dingmy@whu.edu.cn  Haiping Yang Huazhong University of Science and Technology yhping2002@163.com  Chunfei Wu Queen's University Belfast c.wu@hull.ac.uk  Sai Gu University of Surrey sai.gu@surrey.ac.uk  Dan Shan Nanjing University of Science and Technology Danshan@njust.edu.cn

## Cover letter

Dear Editor,

I, on behalf of all authors, would submit the manuscript entitled “Triple-emission N, B co-doped carbon quantum dots from lignin: Highly fluorescent sensing platform for detection of Cr<sup>6+</sup>” to the *Journal of Colloid and Interface Science*.

Cr<sup>6+</sup> with high toxicity poses huge threat to human health and ecological environment, the construction of a rapid and accurate fluorescence sensing platform is of great significance. CQDs based fluorescent sensors show promising potentials for the detection of Cr<sup>6+</sup> due to their nontoxicity, rapid response and adjustable fluorescence characteristics. Among them, biomass-derived CQDs with low cost, renewability and environmentally friendly are expected to realize green and mass production of fluorescent sensors. In this work, N, B-CQDs from lignin waste are synthesized as fluorescent sensor for Cr<sup>6+</sup> detection via a facile two-step method. The resultant N, B-CQDs exhibit triple fluorescent emission under different excitations and their formation mechanism is also excavated. N, B-CQDs are employed as fluorescent sensors for the triple-channel detection of Cr<sup>6+</sup> with superior sensitivity and selectivity. This work hopes to offer a new insight on in-depth understanding of formation mechanism for lignin-derived N, B-CQDs and open new avenue for the rapid and accurate detection of Cr<sup>6+</sup> through multichannel sensing platform.

I sincerely appreciate you for reviewing the manuscript and considering the publication of this review. Look forward to your positive feedback. Thanks a lot!

Prof. Dekui Shen

Key Laboratory of Thermal Conversion and Control, Ministry of China

School of Energy and Environment, Southeast University

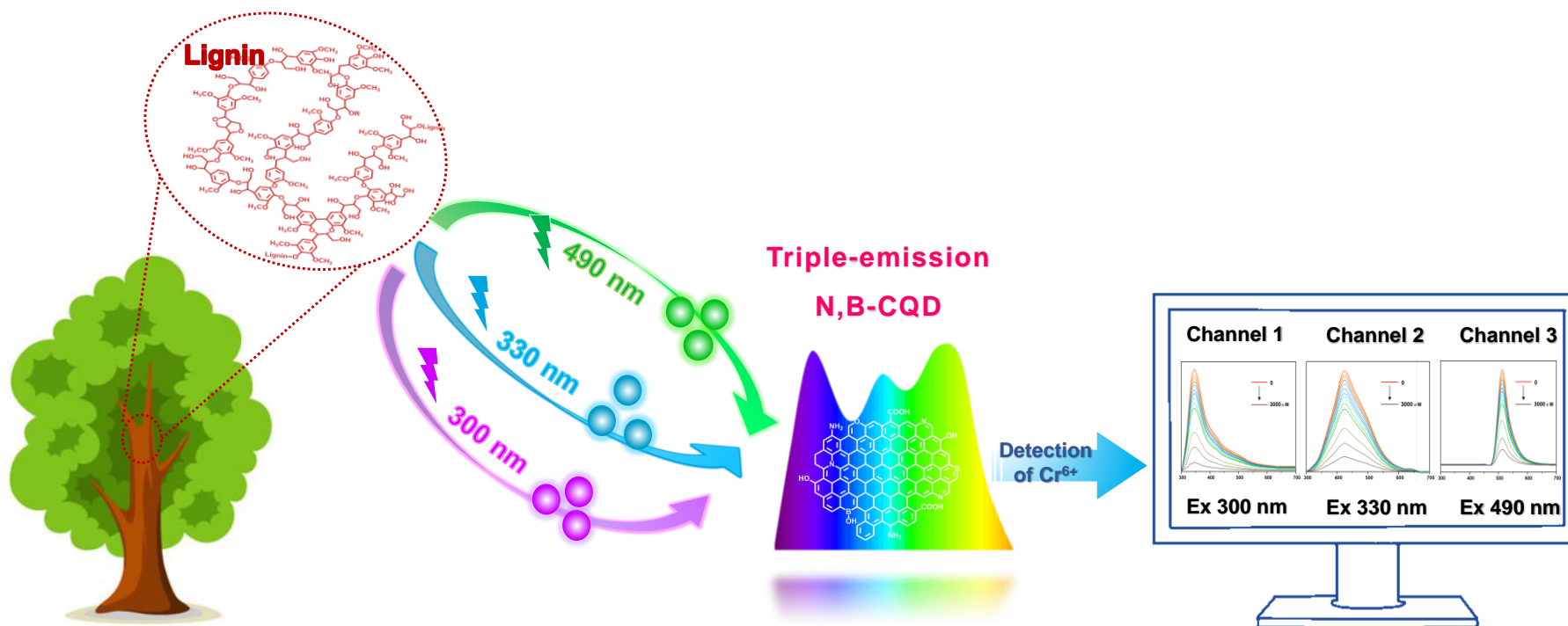
E-mail: 101011398@seu.edu.cn



Click here to access/download  
**2:Supplementary Material**  
Supporting Information.docx



# Graphic abstract



[Click here to view linked References](#)

1       **Triple-emission N, B co-doped carbon quantum dots from lignin:**  
2               **Highly fluorescent sensing platform for detection of Cr<sup>6+</sup>**

3                               Lingli Zhu<sup>a</sup>, Dekui Shen<sup>a,\*</sup> Kai Hong Luo<sup>b</sup>

4       <sup>a</sup> Key Laboratory of Energy Thermal Conversion and Control of Ministry of Education,  
5       School of Energy and Environment, Southeast University, Nanjing 210096, PR China.

6       <sup>b</sup> Department of Mechanical Engineering, University College London, London WC1E7JE,  
7       U.K.

8       E-mail address: [101011398@seu.edu.cn](mailto:101011398@seu.edu.cn) (D. Shen)

9       **ABSTRACT:** Considering that Cr<sup>6+</sup> with high toxicity poses huge threat to human health and  
10       ecological environment, the construction of a rapid and accurate fluorescence sensing  
11       platform is of great significance to detect the toxic substance. The N,B-CQDs from lignin  
12       waste are synthesized as fluorescent sensor for Cr<sup>6+</sup> detection. The synthetic processes involve  
13       the acid hydrolysis step followed by the hydrothermal treatment. Lignin is firstly  
14       depolymerized by cleaving ether bonds under the action of acid, and N,B-CQDs are  
15       consequently formed by the aromatic re-fusion of lignin fragments in the carbonation process.  
16       The resultant N,B-CQDs show triple fluorescent emission of purple, blue and green color  
17       under the excitation of 300, 330, and 490 nm. The N,B-CQDs can be applied for the  
18       triple-channel detection of Cr<sup>6+</sup>, exhibiting highly sensitive and selective fluorescence  
19       quenching in the presence of Cr<sup>6+</sup> with good linearity ( $R^2 \leq 0.996$ ) and very low limit of  
20       detection as 0.054, 0.049 and 0.077  $\mu\text{M}$  for Ex 300, 330 and 490 nm channel, respectively.  
21       The utilization of renewable lignin highlights the green, cheap and large-scale production of

22 CQDs based fluorescent sensors and opens a new avenue for the rapid and accurate detection  
23 of Cr<sup>6+</sup> through multichannel sensing platform.

24 **Keywords:** Lignin; Carbon quantum dots; N, B co-Doped; Triple channel; Sensing Cr<sup>6+</sup>

## 25 **1. Introduction**

26 Chromium is widely distributed in the ecosystem in the form of Cr<sup>3+</sup> and Cr<sup>6+</sup> [1]. Cr<sup>3+</sup> is  
27 considered as an indispensable trace element in the human body, while Cr<sup>6+</sup> is extremely  
28 highly toxic heavy metal contaminant, which is ca. 100 times more toxic than Cr<sup>3+</sup> [2].  
29 Massive Cr<sup>6+</sup> has been discharged into the biological environment from industrial effluents  
30 and waste residue produced by stainless steel manufacturing, ore smelting, dyeing, leather  
31 tanning, metal electroplating, etc. [3, 4]. Excessive Cr<sup>6+</sup> is carcinogenic and mutagenic to  
32 respiratory tract, internal organs and nerve tissue owing to its high oxidation potential and  
33 super ability to penetrate biomembranes, posing a great threat to the environment and human  
34 health [5-7]. The maximum allowable limit of Cr<sup>6+</sup> in drinking water is 1.92 μM as stipulated  
35 by the United States Environmental Protection Agency (USEPA), 0.962 μM by the World  
36 Health Organization (WHO), and in aquatic products is 2 mg/kg by the National Food Safety  
37 Standard of China (GB, 2762–2012) [8, 9]. Thus, the development of effective detection  
38 technology for Cr<sup>6+</sup> is a top priority to ensure the safety of food and drinking water sources.

39 Currently, traditional detection techniques for Cr<sup>6+</sup> include atomic absorption  
40 spectrometry (AAS), atomic emission spectrometry (AES), inductively coupled plasma-mass  
41 spectrometry (ICP-MS), and electrochemical detection [10-12]. However, these methods have  
42 several drawbacks of large or costly equipment, time-consuming preprocessing, complex  
43 operation and tedious analysis, which limit the large-scale rapid detection of Cr<sup>6+</sup> [13]. In

44 comparison, the fluorescent detection method for  $\text{Cr}^{6+}$  can overcome these issues for its  
45 simple, fast, real-time and intuitive merits [14-16]. Among them, carbon quantum dots (CQDs)  
46 based fluorescent detection method for  $\text{Cr}^{6+}$  shows great promise due to its nontoxicity, good  
47 water solubility, rapid response as well as adjustable fluorescence characteristics [16]. Wang  
48 et al. [6] synthesized bright blue fluorescent N,S co-doped CQDs using citric acid  
49 monohydrate and thiosemicarbazide as raw materials via hydrothermal reaction. The N,  
50 S-CQDs based sensors not only exhibited sensitive fluorescence response for  $\text{Cr}^{6+}$  ions with  
51 the LOD of  $0.33 \mu\text{M}$ , but also for toluene in ethanol solution with the LOD of  $0.03 \mu\text{M}$ .  
52 Similarly, the N, S-CQDs based fluorescent sensors were obtained by Ji et al. [12] from the  
53 rapid stirring of L-cysteine and urea, which showed the LOD as low as  $0.02 \mu\text{M}$  for  $\text{Cr}^{6+}$   
54 detection along with  $0.28 \mu\text{M}$  for ascorbic acid detection. Zheng and co-workers [17]  
55 designed controllable functionalization of N-CQDs from carbon rods through electrochemical  
56 method followed by the solvothermal treatment with ammonia, N,N-Dimethylformamide or  
57 polyethylene glycol, respectively. The resultant three kinds N-CQDs were as fluorescent  
58 sensors for independent  $\text{Cr}^{6+}$ ,  $\text{Fe}^{3+}$  and  $\text{Cu}^{2+}$  ions detection. Nevertheless, the CQD-based  $\text{Cr}^{6+}$   
59 sensors are still largely underdeveloped with the bottleneck in selectivity and reproducibility.  
60 It might be attributed to their strong affinity for  $\text{Fe}^{3+}$  or other substances by naturally abundant  
61 phenolic hydroxyl attached on the surface of CQDs [18]. In addition, using toxic organic  
62 small molecules as raw materials for CQDs production is bound to add the risk to the  
63 environment and human health. Accordingly, there is an urgent need for the development of  
64 superior CQDs based fluorescent sensors with higher selectivity, greater practicability and  
65 more environmental friendliness.

66 In this work, the green synthesis of N,B co-doped CQDs from sustainable lignin via a  
67 mild two-step approach is proposed for the fluorescent detection of Cr<sup>6+</sup>. The  
68 physicochemical properties of N,B-CQDs are comprehensively characterized and the  
69 formation mechanism is also excavated. Utilizing its unique triple emission property, a highly  
70 fluorescent tri-channel sensing platform has established for rapid and accurate detection of  
71 Cr<sup>6+</sup> ions. The lignin derived CQDs using as fluorescent sensors offer a green, low-cost, and  
72 available strategy for the sensitive detection of Cr<sup>6+</sup> in a large scale.

## 73 **2. Experimental section**

### 74 *2.1 Reagents and materials*

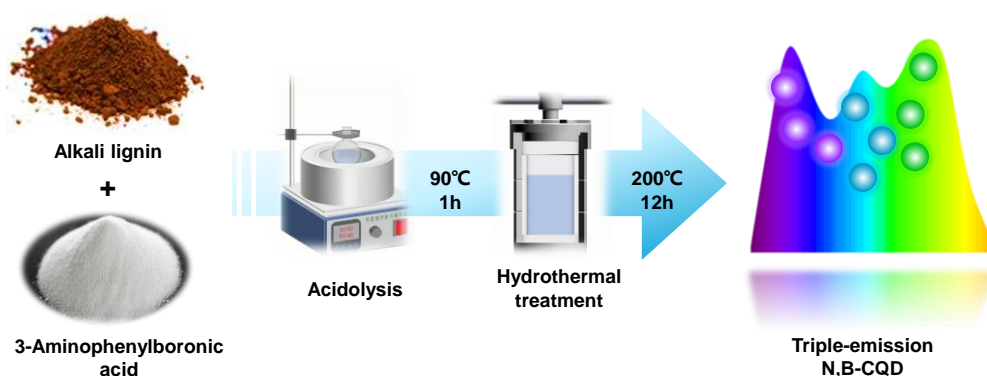
75 Alkali lignin with 4% sulfur was acquired from Sigma-Aldrich (Shanghai, China).  
76 3-Aminophenylboronic acid (≥98%) was purchased from Aladdin (Shanghai, China). Other  
77 chemicals, including NaCl<sub>2</sub>, NaF, Na<sub>2</sub>S, NaNO<sub>3</sub>, Na<sub>2</sub>SO<sub>3</sub>, Na<sub>2</sub>SO<sub>4</sub>, NaH<sub>2</sub>PO<sub>2</sub>, KCl,  
78 NiCl<sub>2</sub>·6H<sub>2</sub>O, MgCl<sub>2</sub>·6H<sub>2</sub>O, CuCl<sub>2</sub>·H<sub>2</sub>O, CaCl<sub>2</sub>, ZnCl<sub>2</sub>, FeCl<sub>3</sub>·6H<sub>2</sub>O, and K<sub>2</sub>CrO<sub>4</sub> were  
79 purchased from Macklin (Shanghai, China). Deionized (DI) water was used throughout this  
80 experiment. The dialysis bags with retained molecular weight of 1000 Da were provided by  
81 Viskase (USA). The microporous filter membranes of aqueous system with the diameter of  
82 1.0 and 0.10 μm were obtained by Jinteng (Tianjin, China). All chemicals were used without  
83 any further purification.

### 84 *2.2 Preparation of N,B-CQDs*

85 In a typical two-step synthetic route of N,B-CQDs, 0.3 g of 3-aminophenylboronic acid  
86 was added into 30 mL of DI water, and 0.3 g of AL was manually mixed under constant



87 stirring. The mixed solution was then heated at 90 °C for 1 h in water bath with constant  
88 stirring (350 rpm). The solution after the reaction was vacuum filtered by a PTFE  
89 microporous membrane (1.0 μm). The lignin nanoparticles (LNPs) can be obtained by directly  
90 freeze-drying the filtrate. Subsequently, the obtained filtrate was transferred into a  
91 Teflon-lined autoclave (50 mL) and kept at a certain temperature for 12 h. The resultant  
92 CQDs were cooled to room temperature and further filtered out by a microporous filter  
93 membrane (0.10 μm). The remaining filtrate was dialyzed in a dialysis bag against DI water  
94 for 2 days. The as-dialyzed CQDs aqueous solution was freeze-dried under -60 °C for 2 days  
95 and the powders were finally collected for characterization and property measurements. The  
96 effect of different temperature of 120, 150, 180, 200 and 220 °C on the lignin-derived CQDs  
97 has been investigated. The resultant N,B-CQDs are named as CQDs-120, CQDs-150,  
98 CQDs-180, CQDs-200, and CQDs-220, respectively.



100 **Scheme 1.** Illustration for the synthetic route of triple-emission N, B-CQDs.

### 101 2.3 Characterization

102 The detailed morphology was observed using a high-resolution transmission electron  
103 microscope (HR-TEM, Tecnai G2 F20, FEI, USA). The graphitized structure was recorded by  
104 a Micro-Raman spectrometer with InGaN laser excitation at 532 nm (DXR 2xi, ThermoFisher,

105 USA). The chemical composition was characterized by a Fourier transform infrared  
106 spectrometer (FT-IR, Nicolet Is5, ThermoFisher, USA) and an X-ray photoelectron  
107 spectrometer (XPS, K-Alpha, ThermoFisher, USA). The molecular weight was obtained by a  
108 matrix-assisted laser desorption/ ionization time of flight mass spectrometer (MALDI-TOF  
109 MS, ultrafleXtreme, Bruker, Germany). The  $^{13}\text{C}$ - $^1\text{H}$  chemical environment was measured by  
110 the two-dimensional heteronuclear singular quantum correlation nuclear magnetic resonance  
111 spectrometer (2D-HSQC NMR, Avance III 600 MHz, Bruker, Germany) with the dispersant  
112 of dimethyl sulfoxide- $d_6$  (DMSO) for AL and deuterium oxide ( $\text{D}_2\text{O}$ ) for LNP and CQD. The  
113 optical properties were measured by an ultraviolet–visible spectrophotometer (UV-vis,  
114 UV-5200, Yuanxi, China) and a fluorescence spectrophotometer (Cary Eclipse, Agilent, USA).  
115 The time-resolved PL spectra were acquired from a steady/transient state fluorescence  
116 spectrophotometer (TRPL, FLS1000, Edinburgh, UK) equipped with an integrating sphere to  
117 evaluate the absolute QY of CQDs. The obtained lifetime decay curves were fitted using a  
118 multi-exponential function  $R(t)$  (eq. (1)) and the average lifetime ( $\tau_{\text{avg}}$ ) was calculated  
119 according to eq. (2).

$$120 \quad R(t) = A_1 e^{(-t/\tau_1)} + A_2 e^{(-t/\tau_2)} \dots \dots + A_n e^{(-t/\tau_n)} \quad (1)$$

$$121 \quad \tau_{\text{avg}} = (A_1 \tau_1^2 + A_2 \tau_2^2 + \dots \dots A_n \tau_n^2) / (A_1 \tau_1 + A_2 \tau_2 \dots \dots + A_n \tau_n) \quad (2)$$

122 Where  $\tau_1, \tau_2 \dots \dots \tau_n$  represent fluorescence decay lifetimes and  $A_1, A_2 \dots \dots A_n$  represent  
123 pre-exponential factors of  $\tau_1, \tau_2 \dots \dots \tau_n$ , respectively.

## 124 *2.4 Fluorescent detection of Cr(VI)*

125 The fluorescent detection of a series of interfering ions ( $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{F}^-$ ,  $\text{S}^{2-}$ ,  $\text{SO}_3^{2-}$ ,  $\text{SO}_4^{2-}$ ,  
126  $\text{H}_2\text{PO}_4^-$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{K}^+$ ,  $\text{Zn}^{2+}$ ,  $\text{Fe}^{3+}$ , and  $\text{Cr}^{6+}$ ) was performed at room

127 temperature. 3 mg of N,B-CQD powders were added into 1 L of ion solutions in a  
128 concentration of 600  $\mu\text{M}$  and rested for 30 min. Then their PL emission spectra were recorded  
129 at 300, 330 and 490 nm excitation, respectively. In addition, different concentrations of  $\text{Cr}^{6+}$   
130 including 0, 20, 40, 60, 80, 100, 200, 400, 600, 800, 1000, 1500, 2000 and 3000  $\mu\text{M}$  were  
131 added into N,B-CDs aqueous solution, respectively. The corresponding PL emission spectra  
132 of these mixtures were measured at 300, 330 and 490 nm excitation, respectively. The  
133 theoretical limit of detection (LOD) was calculated according to eq. (3):

$$134 \quad \text{LOD} = 3\sigma/K \quad (3)$$

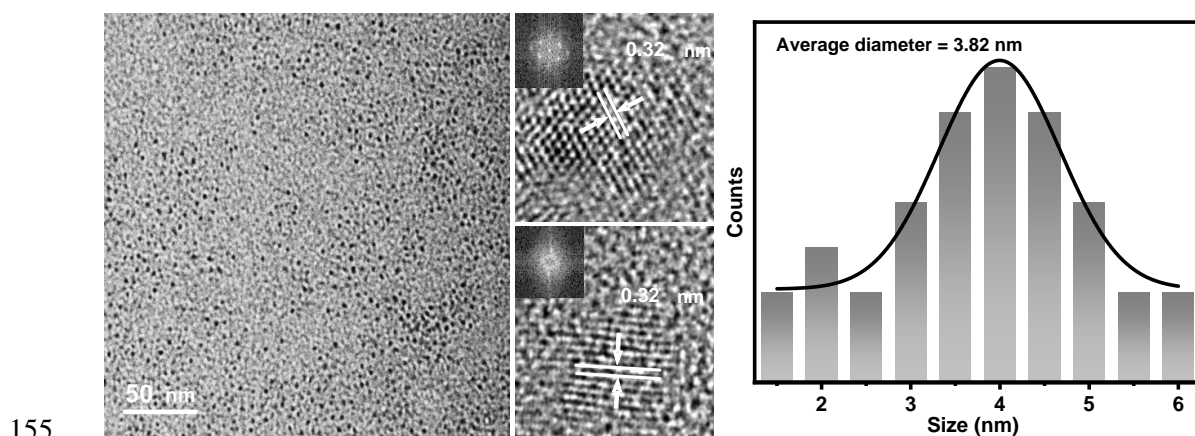
135 Where 3 refers to the signal-to-noise ratio;  $\sigma$  refers to the standard deviation of the blank  
136 signal;  $K$  refers to the slope of the linearly concentration-dependent response.

### 137 **3. Results and discussion**

#### 138 *3.1 Morphological structure of N, B-CQDs*

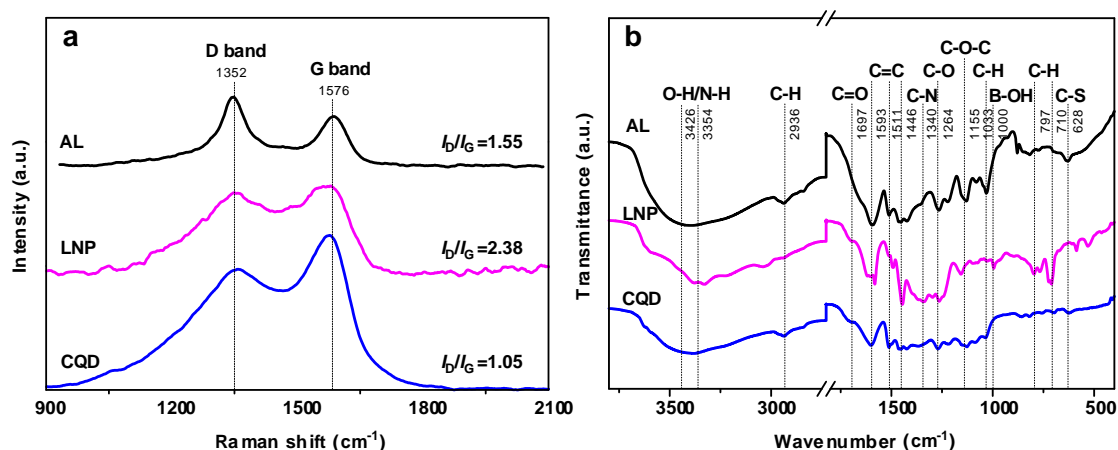
139 The lignin-derived N,B-CQDs are obtained through a mild two-step route involving acid  
140 hydrolysis followed by hydrothermal treatment. The hydrothermal temperatures ranging from  
141 120 to 220  $^{\circ}\text{C}$  are optimized based on the fluorescence QYs of N,B-CQDs at different  
142 excitation wavelengths. According to Fig.S1, N, B-CQDs synthesized at 200  $^{\circ}\text{C}$  shows the  
143 highest QYs under 300, 330 and 490 nm excitation, respectively, which is selected to be  
144 further characterized and investigated. The micromorphology of N, B-CQDs can be observed  
145 by TEM as shown in Fig.1, which are well-dispersed quasi-spherical nanoparticles with an  
146 average diameter of 3.82 nm. The HR-TEM images clearly exhibit one kind of lattice fringe  
147 with interplanar spacing of 0.34 nm, which represents the (002) crystal plane of graphite [19,

148 20]. The graphitization of AL, LNP and CQDs is recorded by Raman spectra (Fig. 2a). There  
 149 are two apparent peaks at  $1356\text{ cm}^{-1}$  (D band) and  $1572\text{ cm}^{-1}$  (G band), corresponding to the  
 150 vibrations of disordered  $\text{sp}^3$  carbon and graphitized  $\text{sp}^2$  carbon [21]. The relative intensities  
 151 ( $I_D/I_G$ ) of AL, LNP and CQDs, an indicator of the defect density in their structure [22, 23], are  
 152 1.55, 2.38, and 1.05, respectively. It indicates that more structural defects are introduced in  
 153 LNP after the acidolysis of AL while more graphitized structure in CQD after the  
 154 hydrothermal reaction.



155  
 156 **Fig. 1.** Morphology of N, B-CQDs: TEM, HRTEM images and size distribution diagram.

157



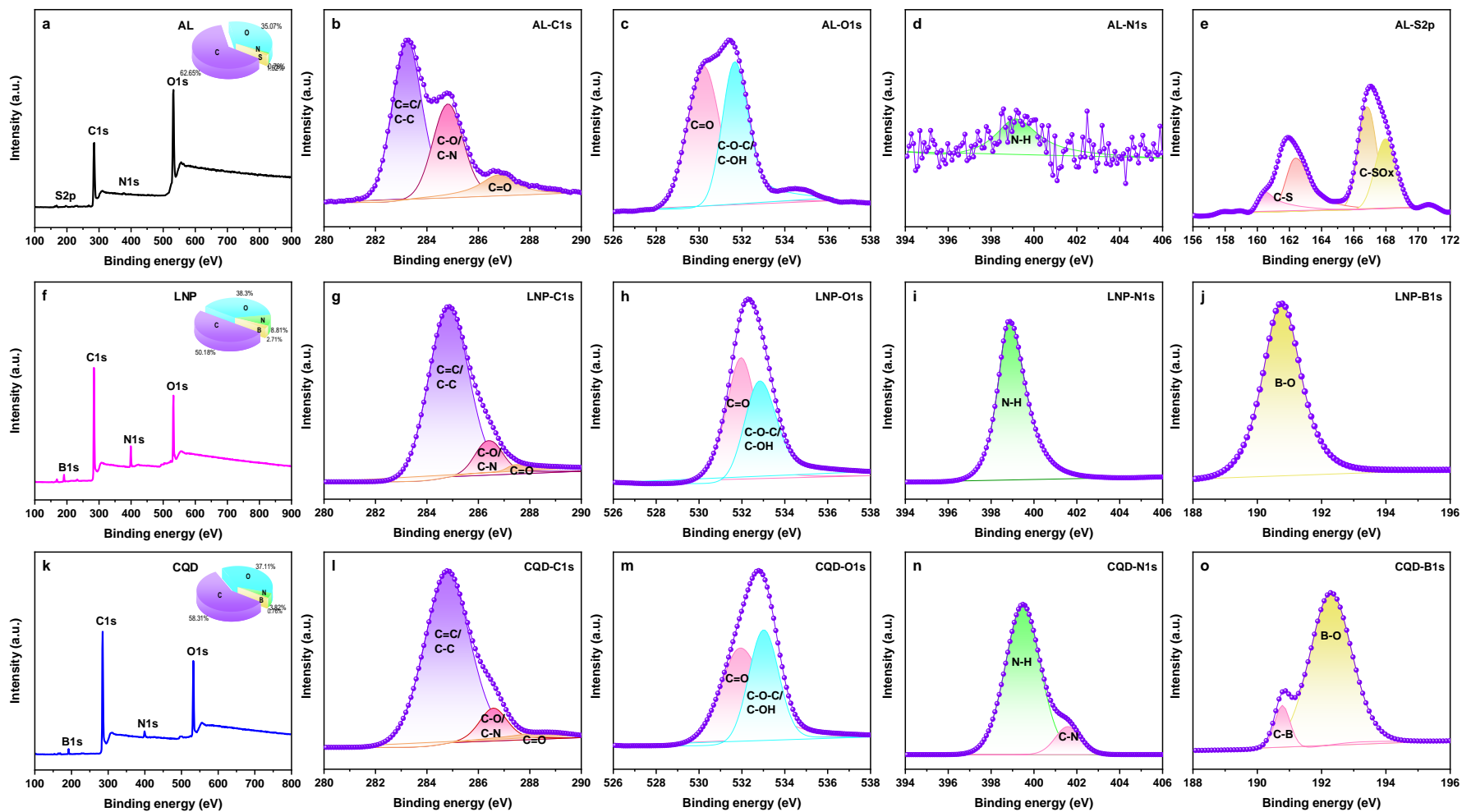
158

159 **Fig. 2.** Structure characterization: (a) Raman spectra, (b) FTIR spectra of AL, LNP and

160 CQD.

161 The chemical compositions and functional groups of AL, LNP and CQD are identified  
162 by FT-IR and XPS. As depicted in Fig. 2b, the strong peaks at 3426 cm<sup>-1</sup> in FT-IR spectra are  
163 assigned to phenolic hydroxyl group [24], where the stretching vibration is significantly  
164 decreased from AL to CQD. The peaks located at 2936 and 1033 cm<sup>-1</sup> are corresponding to  
165 in-plane and out-of-plane bending of aromatic C-H [25]. The peaks at 1697, 1511/1593, 1264  
166 and 1155 cm<sup>-1</sup> are ascribed to C=O, aromatic C=C, C-O and C-O-C stretching vibration,  
167 respectively [26]. Notably, the ether vibration (C-O-C) from AL gradually disappears in LNP  
168 and CQD, while the new stretching peaks of N-H, C-N and B-OH groups are emerged at 3354,  
169 1340 and 1000 cm<sup>-1</sup> in LNP and CQD [18, 27]. These demonstrate that CQD inherits the  
170 aromatic skeletons from AL and derives the N, B co-doping from LNP by acid hydrolysis of  
171 AL. Meanwhile, plentiful polar functional groups (-OH, N-H, C=O, C-O, and B-OH) attached  
172 at the edge of the aromatic skeletons of CQD suggest its outstanding hydrophilicity [14, 21,  
173 28]. These results can be further confirmed by XPS analysis. The XPS survey spectra in  
174 Fig.3a, f and k show prominent peaks at 168.0, 190.1, 284.8, 399.1, and 532.1 eV, which  
175 assigned to S2p, B1s, C1s, N1s, and O1s, respectively [15, 18, 23, 29]. AL is mainly  
176 composed of C and O along with a small amount of N and S, while LNP and CQD have  
177 similar components of C, O, N and B. The high-resolution XPS spectra of C1s, O1s, N1s, and  
178 B1s are displayed in Fig. 4b-o. The C1s spectra can be fitted with three peaks at 284.8, 286.2,  
179 and 288.2 eV, corresponding to sp<sup>2</sup> carbon (C-C/C=C), sp<sup>3</sup> carbon (C-O/C-N), and carbonyl  
180 groups (C=O) [30]. There is a sharply decreased C-O peak from AL to CQD. The O1s spectra  
181 can be deconvoluted into two peaks at 531.8 and 533.2 eV for C=O and C-O-C/C-OH [31],  
182 where the latter is decreased from AL to LNP. This indicates the cleavage of ether bond by the

183 acidolysis of AL. The N1s spectra show two peaks at 399.7 and 401.8 eV for N-H and C-N  
184 groups [32]. The S2p spectra present two peaks at 162.2 and 167.8 eV corresponding to C-S  
185 and C-SO<sub>x</sub> groups [22]. The B1s spectra can be deconvoluted into two peaks at 190.6 and  
186 192.3 eV corresponding to B-C and B-O groups [32]. Remarkably, there are no significant  
187 signals of N1s and B1s of AL, which are consistent with the FT-IR results. The graphitic C-N  
188 and C-B units newly appear from LNP to CQD, inferring that doped N and B are inclined to  
189 combine at the edge sites of CQD along with a small amount of lattice doping. These findings  
190 can be evidently supported by the quantitative results of XPS analysis as listed in Table S1.  
191 The C content decreases first and then increases while O content is the opposite, implying the  
192 depolymerization of AL followed by the re-fusion of LNP into the aromatic skeletons of CQD  
193 during the synthetic process.



194

195

196

**Fig. 3.** Structure characterization: (a, f, k) survey and High-resolution (b, g, l) C1s, (c, h, m) O1s, (d, i, n) N1s, (e) S2p and (j, o) B1s XPS spectra of AL, LNP and CQD.

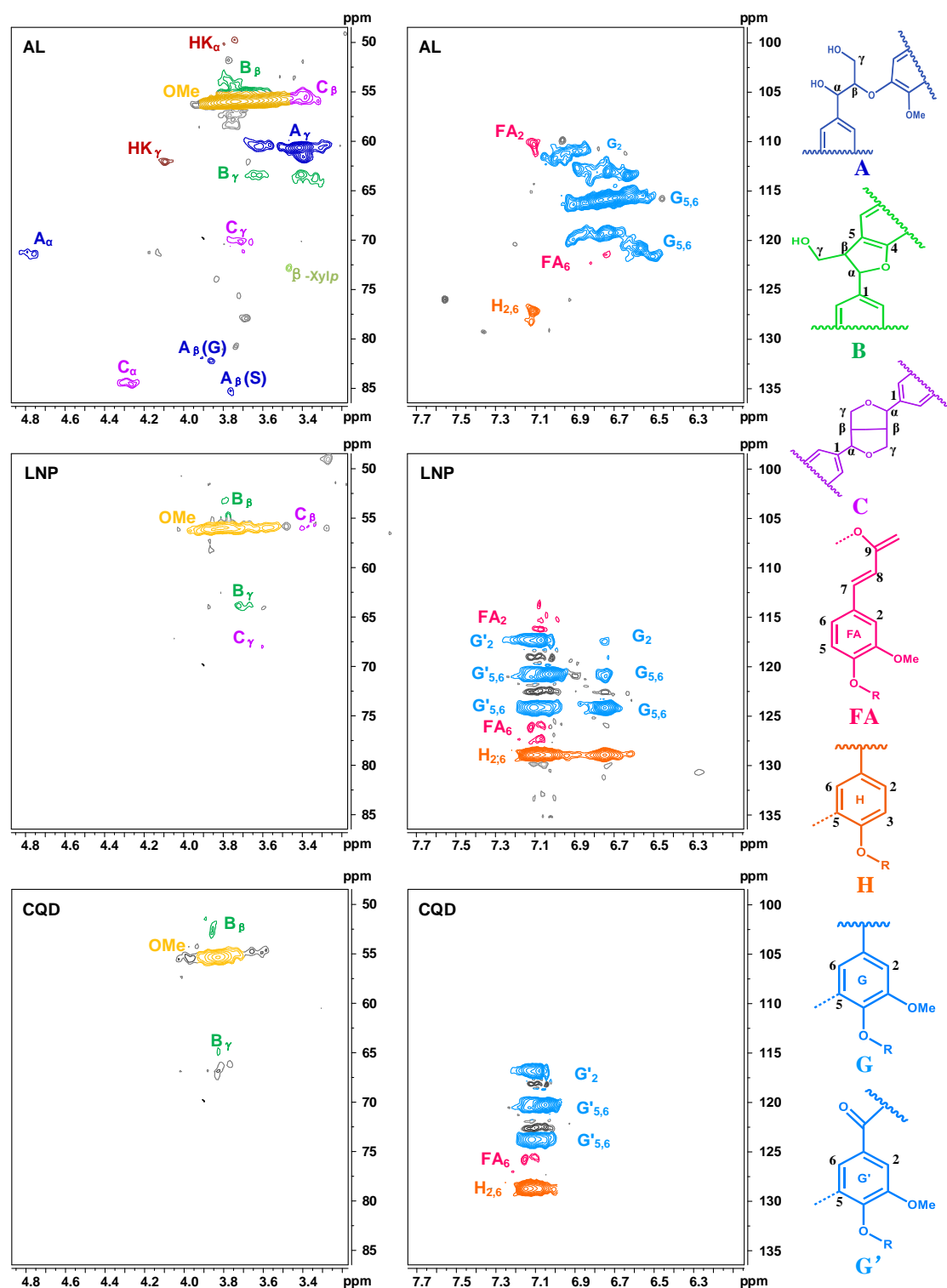
### 197 3.2 Formation mechanism of N, B-CQDs

198 The 2D- HSQC NMR spectra of AL, LNP and CQD are characterized for the in-depth  
199 structural insights on the transformation of lignin in the two-step process (Fig. 4). Lignin is an  
200 amorphous polymer with a three-dimensional network structure, which is composed of three  
201 types of phenylpropane units connected by ether bonds or C-C bonds [33, 34]. The HSQC  
202 data of AL show plentiful signals in the side-chain region ( $\delta_C/\delta_H$ : 50–90/3.2–5.0 ppm),  
203 assigned to the native structure of lignin including methoxyls (OCH<sub>3</sub>),  $\beta$ -aryl-ether linkages  
204 (A), phenylcoumarane structures (B) and resinol linkages (C) [35]. The signals of ferulate  
205 (FA), p-hydroxyphenyl (H), (G) guaiacyl units and (G') oxidized guaiacyl units are apparently  
206 exhibited in the aromatic region ( $\delta_C/\delta_H$ : 100–135/6.0–7.8 ppm) [36, 37]. In comparison, the  
207 side-chain signals of A, B and C linkages are sharply reduced in the HSQC data of LNP, and  
208 even almost vanish in the HSQC data of CQD, which are in consistent with the above Raman,  
209 FT-IR and XPS results. It qualitatively demonstrates that LNP is obtained by strongly  
210 de-etherified after the acid treatment of AL via the cleavage of A, B and C linkages.  
211 Meanwhile, the N and B dual-doping evidenced by FT-IR and XPS results may contribute to  
212 abundant active sites exposed on the surface of LNP for further transformation. The aromatic  
213 structures (FA, H, G and G') of AL are well inherited by LNP and CQD in the synthesis  
214 process.

215 According to the experimental evidences above, the formation mechanism of N,B-CQDs  
216 from AL via the two-step route is proposed as follows (Fig. 5): In the 1st step of acid  
217 hydrolysis, AL is de-ether decomposed into lignin fragments (LNP) through the catalytic  
218 cleavage of ether linkages, with simultaneous N, B co-doping via covalent conjugation [28].



219 Thereby, various active sites including oxygen-, nitrogen- and boron- functional groups can  
220 be generate on the LNP surface. 3-Aminophenylboronic acid plays a triple role of “top-down”  
221 scissor, N,B dopant as well as surfactant during the acidolysis process [14, 38]. In the 2nd  
222 step, the functionalized LNP serves as precursors for the preparation of CQD. The  
223 dehydration condensation of activated side chains on LNP result in the prompt  $\pi$ - $\pi$  stacking  
224 and the increased  $sp^2$  hybridization under the hydrothermal treatment [22, 39]. The C–N and  
225 C–B groups are simultaneously enhanced in the carbonization process, eventually resulting in  
226 the formation of CQD [40]. In summary, the formation of N,B co-doped CQDs involves the  
227 acid dissociation of AL first followed by the aromatic re-fusion of LNP.



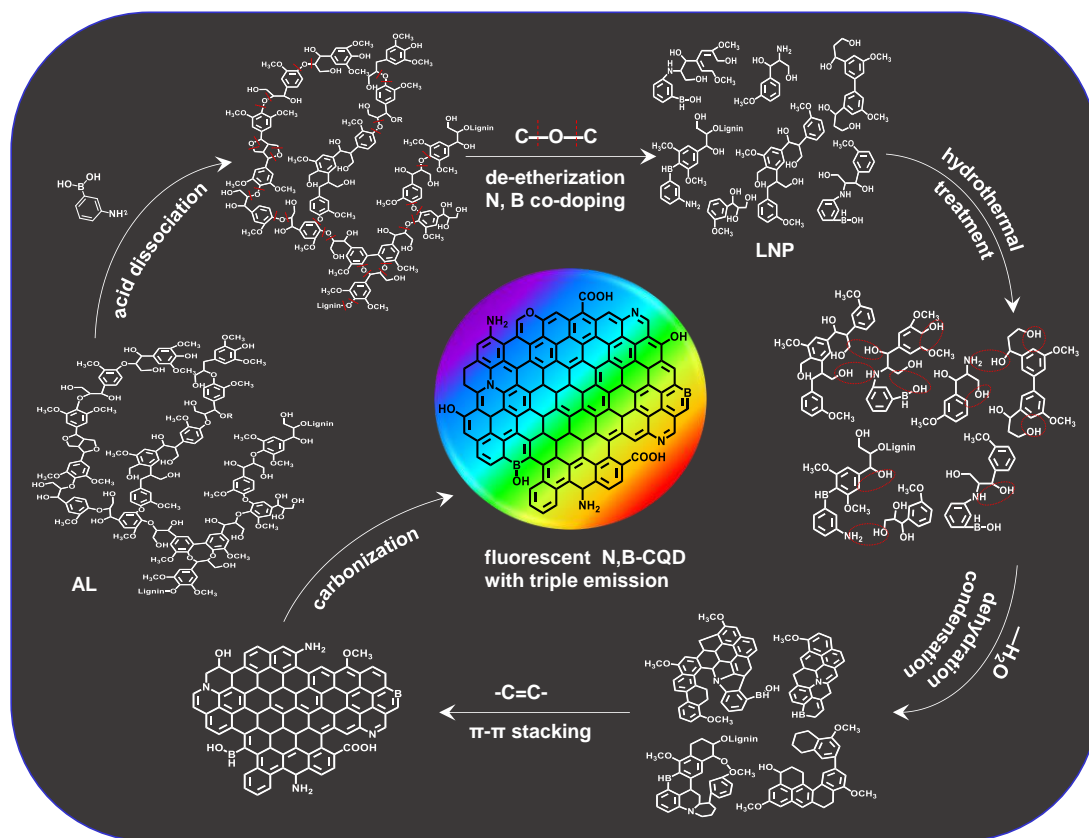
228

229 **Fig. 4.** Main substructures identified in the 2D- HSQC NMR spectra of AL, LNP and CQD:

230 (A)  $\beta$ -O-4' linkages; (B) phenylcoumaran with  $\alpha$ -O-4' and  $\beta$ -O-5' linkages; (C) resinol with

231  $\beta$ - $\beta'$ ,  $\alpha$ -O- $\gamma'$ , and  $\gamma$ -O- $\alpha'$  linkages; (FA) ferulate; (H) p-hydroxyphenyl; (G) guaiacyl units and

232 (G') oxidized guaiacyl unit bearing a carbonyl group at C $\alpha$ .



233

234 **Fig. 5.** Proposed formation mechanism of N,B-CQDs from AL via two-step route.

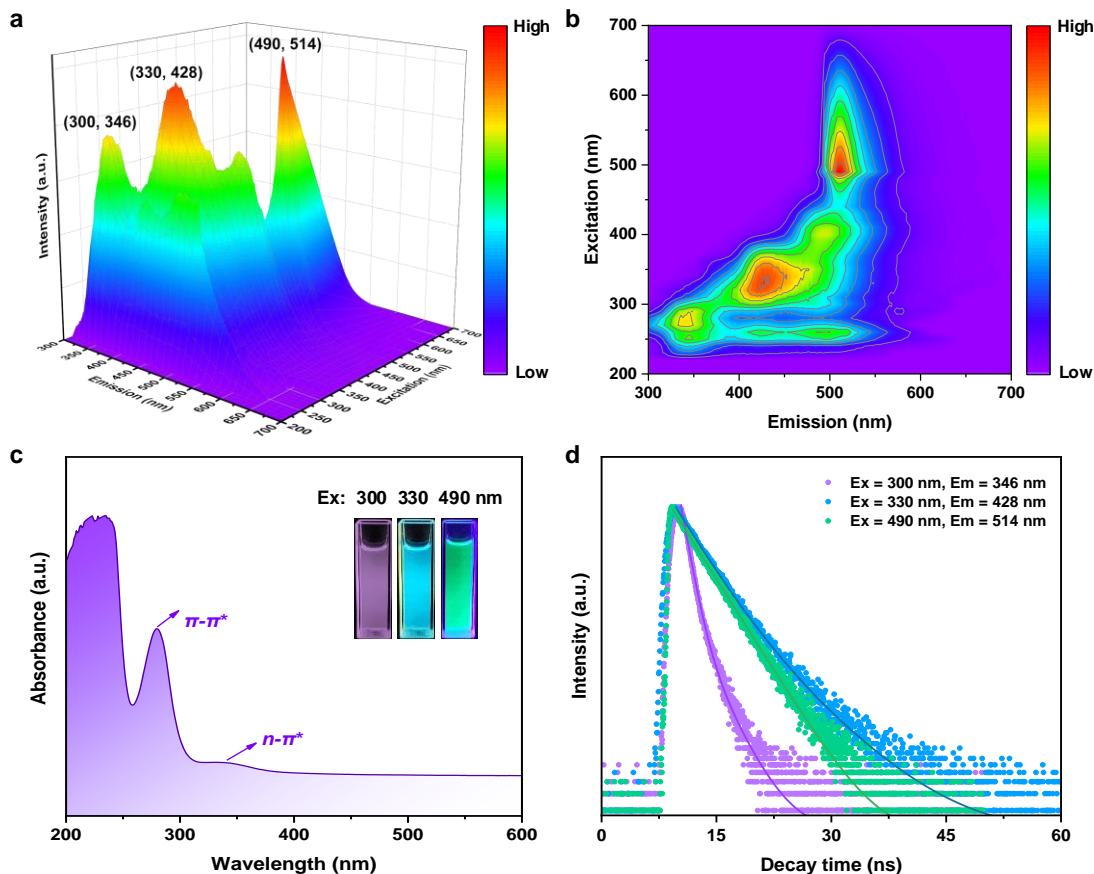
### 235 3.3 Optical properties of fluorescent N, B-CQDs

236 The optical properties of fluorescent N,B-CQDs are further investigated. The PL  
 237 emission spectra under different excitation wavelengths of 200-700 nm are displayed in Fig.  
 238 6a-b. There are three distinct PL centers located at (300, 346), (330, 428), and (490, 514) nm,  
 239 corresponding to the fluorescence emission of purple, blue and green color, respectively. This  
 240 indicates that the N,B-CQDs prepared from AL have unique triple-fluorescence emission,  
 241 which is rarely found in the previous reported biomass-derived CQDs [41, 42]. In detail,  
 242 N,B-CQDs exhibit almost excitation-independent property under the excitation of 230-300  
 243 nm and 440-490 (Fig. S2a, c). Oppositely, the emission peaks are gradually red-shifted from  
 244 350 to 496 nm when the excitation wavelengths increase from 310 to 430 nm with a 10 nm

245 increment (Fig. S2b). This pronounced excitation-dependent behavior is associated with  
246 diverse surface defects introduced by O, N or B functional groups [32, 38]. As for the UV-vis  
247 absorption spectra (Fig. 6c), N,B-CQDs show two strong adsorption peaks at 230, 281 nm  
248 and a weak one at 343 nm, with an optical adsorption edge at  $\sim$ 500 nm. The first two peaks  
249 below 300 nm are attributed to the  $\pi$ - $\pi^*$  transition of C=C in the carbon core, while the other  
250 are corresponding to the n- $\pi^*$  transition of C=O/C=N on the surface of CQDs [43, 44]. Their  
251 corresponding absorbance intensity depends on the conjugated degree in the carbon core and  
252 defects state on the surface, respectively.

253 As shown in Fig. 6d, the TRPL spectra of the triple-emission N,B-CQDs are monitored  
254 at  $\lambda_{\text{ex}}/\lambda_{\text{em}}$  of 300/346, 330/428 and 490/514 nm, respectively. The three decay curves can be  
255 bioexponentially fitted at the excitation of 300 and 330 nm while monoexponentially fitted at  
256 490 nm (the fitting parameters are summarized in Table S2). The  $\tau_{\text{avg}}$  of N,B-CQDs at 300,  
257 330 and 490 nm excitation are 1.26, 3.91 and 3.41 ns, respectively, which are consistent with  
258 the variation trend of QYs. The bioexponential lifetimes denote two PL centers including a  
259 fast component ( $\tau_1$ ) originated from the radiative recombination of intrinsic state ( $\pi$ - $\pi^*$ ) as  
260 well as a slow component ( $\tau_2$ ) ascribed to the non-radiative process of surface state (n- $\pi^*$ ) [29,  
261 45]. The percentages of  $\tau_1$  excited at 300 and 330 nm of the N,B-CQDs are 81.55% and  
262 75.93%, which demonstrate that their PL emissions are mainly attributed to the sp<sup>2</sup> conjugated  
263 structure in the carbon core. The percentage of  $\tau_2$  excited at 330 nm increases to 24.07%  
264 compared with that at 300 nm, corresponding to the enhanced effect of surface defects on PL  
265 emission. The PL emissions of N,B-CQDs excited at 490 nm present a monoexponential  
266 lifetime, implying a single PL center of molecular state [46, 47]. The difference concerning

267 PL centers at 300, 330 and 490 nm excitation further explains the triple-emission performance  
268 of N,B-CQDs. As well known, the O, N and B-related functional groups may serve as  
269 electron acceptor and surface-passivation agent to induce strong intramolecular charge  
270 transfer and improved emission efficiency, respectively. The O-related functional groups,  
271 especially C=O with strong electron-accepting property are considered to be chromophores.  
272 The N, B related groups are identified as auxochrome groups, the isolated n electrons of  
273 which and  $\pi$  electrons of the carbon ring are almost coplanar. The generated defect states lead  
274 to the additional energy levels (n- $\pi^*$  transitions) between  $\pi$  and  $\pi^*$  transition [48]. The  
275 additional n- $\pi^*$  transitions result in multiple radiation recombination paths back to the ground  
276 state, thereby exhibiting unique PL performances [49]. In general, the triple-emission  
277 behavior of N,B-CQDs depends on the comprehensive effect of carbon core and surface  
278 defect state.



279

280 **Fig. 6.** Optical properties of N,B-CQDs: (a) Three-dimensional (3D) fluorescence  
 281 spectra, (b) Detailed PL emission spectra under different excitation wavelengths in the range  
 282 from 200 to 700 nm, (c) UV-vis absorption spectra, inset: photographs of N,B-CQD aqueous  
 283 solution under different excitation lights (300, 330 and 490 nm), (d) TRPL decay curves at  
 284 emission of 346, 428, and 514 nm with the fitted curves.

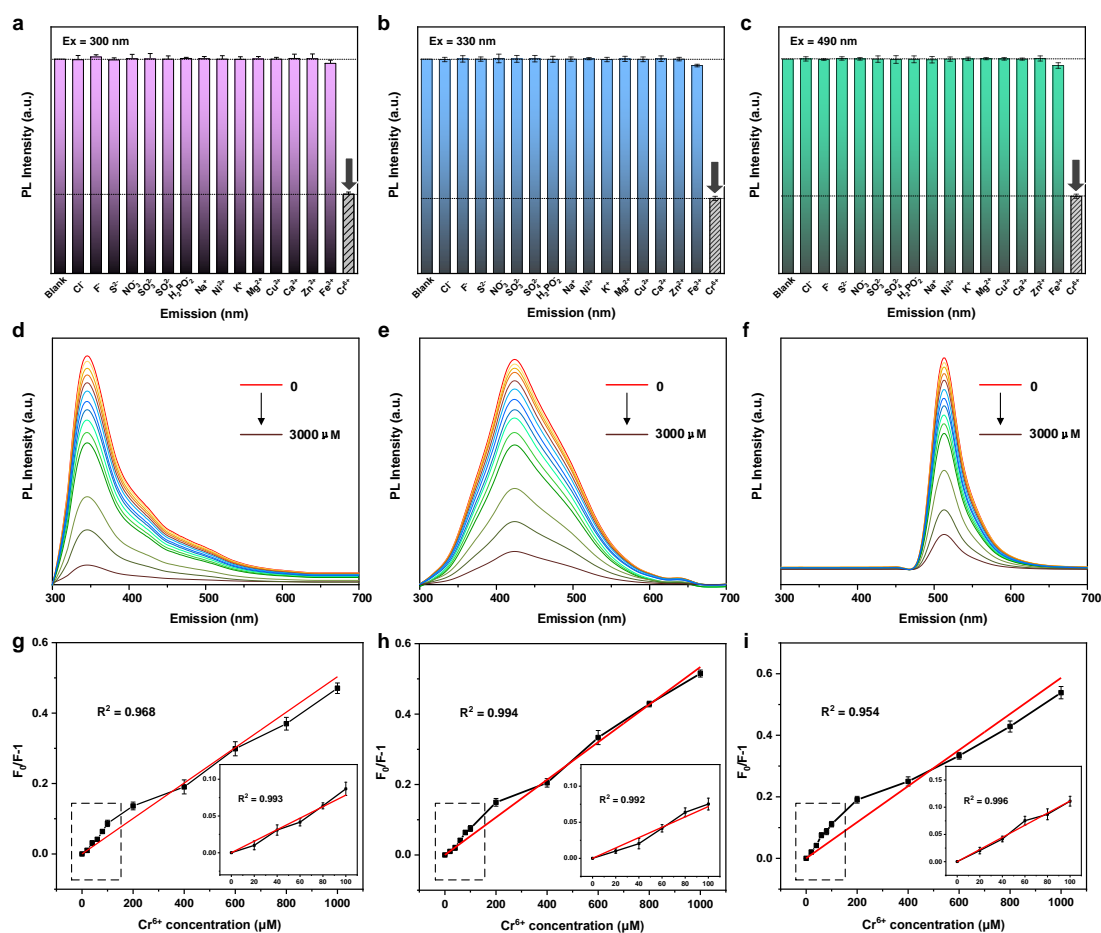
### 285 3.4 Tri-channel sensitive detection of $Cr^{6+}$

286 Based on the green synthesis, low toxicity, good water solubility and unique optical  
 287 properties, the selective and sensitive detection for  $Cr^{6+}$  is investigated to estimate  
 288 performance of N, B-CQDs-based fluorescent sensor. The selectivity of N,B-CQDs towards  
 289  $Cr^{6+}$  and other interfering ions is evaluated, including the anions of  $Cl^-$ ,  $F^-$ ,  $S^{2-}$ ,  $NO_3^-$ ,  $SO_3^{2-}$ ,  
 290  $SO_4^{2-}$ ,  $H_2PO_4^-$  and metal cations of  $Na^+$ ,  $K^+$ ,  $Ni^{2+}$ ,  $Mg^{2+}$ ,  $Cu^{2+}$ ,  $Ca^{2+}$ ,  $Zn^{2+}$ ,  $Fe^{3+}$ , and  $Cr^{6+}$ . As

291 presented in Fig.7a-c, the PL intensity of N,B-CQDs aqueous solution is maintained well in  
292 the presence of different interfering ions (600  $\mu\text{M}$ ). However, there is a significant  
293 fluorescence quenching in terms of  $\text{Cr}^{6+}$  under 300, 330 and 490 nm excitation, respectively.  
294 Comparing to other CQD-based sensors that also have a fluorescence quenching effect on  
295  $\text{Fe}^{3+}$ , this N,B-CQDs-based sensor exhibit the superiority in selectivity. The  $\text{Cr}^{6+}$  induced  
296 fluorescence quenching is considered to be originated from the dynamic quenching dominated  
297 by collisional deactivation [5, 15, 17, 18]. UV-Vis spectra of  $\text{Cr}^{6+}$ , N,B-CQDs and N,B-CQDs  
298 in the presence of  $\text{Cr}^{6+}$  are shown in Fig. S3. There are two strong absorption peaks around  
299 270 and 368 nm, which are overlapped with those of N,B-CQDs. It may suppress electron  
300 transitions and cause extinction effect on the fluorescence emission of N,B-CQDs [10]. In  
301 addition, N,B-CQDs are decorated with  $-\text{NH}_2$ ,  $-\text{COOH}$  and  $-\text{OH}$  surface groups as electron  
302 donors, and  $\text{Cr}^{6+}$  has stronger binding affinity than other interfering ions towards the  
303 electron-donating groups. It can induce considerable energy dissipation instead by rapider  
304 electron transfer and restrain recombination of excitons, thereby sharper fluorescence  
305 reduction of N,B-CQDs [14, 18].

306 The sensitivity of N,B-CQDs for  $\text{Cr}^{6+}$  detection is tested within a wide concentration  
307 range of  $\text{Cr}^{6+}$  from 0 to 1000  $\mu\text{M}$ . As displayed in Fig. d-f, the PL intensity gradually decrease  
308 with the increase of the  $\text{Cr}^{6+}$  concentration, implying the high sensitivity of the concentration  
309 dependent fluorescence quenching. It is worth noting that this phenomenon is simultaneously  
310 occurred under 300, 330 and 490 nm excitation, demonstrating the feasibility of N,B-CQDs  
311 as a tri-channel detection of  $\text{Cr}^{6+}$ . The linear correlation coefficients ( $R^2$ ) at concentration  
312 range of 0-1000  $\mu\text{M}$  are observed as 0.968, 0.994 and 0.954 for Ex 300, 330 and 490 nm

313 channel, respectively in Fig. g-i. Furthermore, the higher  $R^2$  are obtained as 0.993, 0.992 and  
 314 0.996 at a narrow concentration range of 0-100  $\mu\text{M}$ . Accordingly, the LOD can reach as low  
 315 as 0.054, 0.049 and 0.077  $\mu\text{M}$ , respectively, which are far lower than the maximum level of  
 316 drinking water (0.962  $\mu\text{M}$ ) permitted by the World Health Organization (WHO) [1]. The  
 317 lignin-derived N,B-CQDs are comparable to most previously reported CQDs for  $\text{Cr}^{6+}$   
 318 detection regarding the LOD and linear range (Table 1). These results demonstrate that  
 319 comparable accuracy is achieved for N,B-CQDs based fluorescent sensor, especially the Ex  
 320 330 nm channel. Therefore, N,B-CQDs have great potential in the tri-channel detection of  
 321  $\text{Cr}^{6+}$  in practical application.



322

323 **Fig. 7.** (a-c) The fluorescence responses of N,B-CQDs with the addition of various  
 324 interfering ions (600  $\mu\text{M}$ ); (d-f) the fluorescence spectra of N,B-CQDs in the presence of



325 different concentration of  $\text{Cr}^{6+}$  (0-1000  $\mu\text{M}$ ); (g-i) the linear fitting curves of  $(F_0-F)/F_0$  at the  
 326 different concentrations of  $\text{Cr}^{6+}$  in the range of 0-1000 mM and 0-100 (insert) excited under  
 327 300, 330 and 490 nm, respectively.

328 **Table 1.** Highly sensitive  $\text{Cr}^{6+}$  detection of N,B-CQDs compared with other reported  
 329 CQDs, based on the responsive fluorescence quenching.

Fluorescent sensors	Performances		References
	Limit of detection ( $\mu\text{M}$ )	Linear range ( $\mu\text{M}$ )	
Cysm-CQDs	1.570	1-10	[50]
CQDs	0.160	2-80	[51]
N,B-CQDs	0.41	0-100	[18]
af-CQDs	0.242	0.8-80	[17]
N-CQDs	0.260	0-12	[10]
S,N-CDs/PVA film	0.093	0.1–50	[12]
Co-CQDs	1.17	5 -125	[52]
N,B-CQDs	0.054	0-100	This work
N,B-CQDs	0.049	0-100	This work
N,B-CQDs	0.077	0-100	This work

## 330 4. Conclusions

331 The fluorescent N,B co-doped CQDs from renewable lignin waste are fabricated by a  
 332 green two-step strategy to develop a versatile sensitive platform for detection of  $\text{Cr}^{6+}$  ions.  
 333 The formation mechanism of N,B co-doped CQDs involves the de-etherization and  
 334 depolymerization of AL in the acidolysis step, followed by the aromatic re-fusion of LNP in  
 335 the hydrothermal treatment. The resultant N,B-CQDs exhibit triple fluorescent emission under  
 336 the excitation of 300, 330, and 90 nm, which are considered to be induced by the synergistic  
 337 effect of carbon core state and surface defect state. The triple-emission N,B-CQDs are utilized  
 338 for the highly selective and sensitive detection of  $\text{Cr}^{6+}$  with the LOD as low as 0.144, 0.082

339 and 0.094  $\mu\text{M}$  in Ex 300, 330 and 490 nm channel, respectively. The sensing mechanism may  
340 be attributed to the decorated electron-donating amino, carboxyl, hydroxyl groups on the  
341 CQD surface, which combine  $\text{Cr}^{6+}$  to cause the dynamic fluorescence quenching. The  
342 fluorescence sensing platform based on N,B-CQDs is constructed, which provides new  
343 insights on the rapid, visualized and accurate detection of  $\text{Cr}^{6+}$  through triple channel in  
344 aqueous media.

### 345 **CRedit authorship contribution statement**

346 Lingli Zhu: Investigation, Methodology, Data curation, Writing-original draft, Validation.

347 Dekui Shen: Conceptualization, Investigation, Writing-original draft, Funding acquisition.

348 Kai Hong Luo: Writing-review, Supervision.

### 349 **Declaration of Competing Interest**

350 The authors declare that they have no known competing financial interests or personal  
351 relationships that could have appeared to influence the work reported in this paper.

### 352 **Acknowledgements**

353 The authors gratefully acknowledge the financial support from National Natural Science  
354 Foundation of China (grant numbers. 51676047 and 51861145102) and the Key Research &  
355 Development Program of Jiangsu Province (grant number: BE2020114). The authors also  
356 acknowledge the funding support from the Scientific Research Foundation of Graduate  
357 School of Southeast University, China (YBPY2109) and Postgraduate Research & Practice  
358 Innovation Program of Jiangsu Province from the Education Department of Jiangsu  
359 (KYCX21\_0094).

## 360 **Appendix A. Supplementary data**

361 Supplementary material related to this article can be found, in the online version, at

## 362 **References**

- 363 [1] Y. Mao, S. Gao, L. Yao, L. Wang, H. Qu, Y. Wu, Y. Chen, L. Zheng, Single-atom  
364 nanozyme enabled fast and highly sensitive colorimetric detection of Cr(VI), *J Hazard*  
365 *Mater*, 408 (2021) 124898.
- 366 [2] S. Li, T. Wei, G. Ren, F. Chai, H. Wu, F. Qu, Gold nanoparticles based colorimetric probe  
367 for Cr(III) and Cr(VI) detection, *Colloids and Surfaces A: Physicochemical and*  
368 *Engineering Aspects*, 535 (2017) 215-224.
- 369 [3] S. Shahim, R. Sukesan, I. Sarangadharan, Y.L. Wang, Multiplexed Ultra-Sensitive  
370 Detection of Cr(III) and Cr(VI) Ion by FET Sensor Array in a Liquid Medium, *Sensors*  
371 (Basel), 19 (2019).
- 372 [4] O. Dvoynenko, S.L. Lo, Y.J. Chen, G.W. Chen, H.M. Tsai, Y.L. Wang, J.K. Wang,  
373 Speciation Analysis of Cr(VI) and Cr(III) in Water with Surface-Enhanced Raman  
374 Spectroscopy, *ACS Omega*, 6 (2021) 2052-2059.
- 375 [5] Y. Wang, X. Hu, W. Li, X. Huang, Z. Li, W. Zhang, X. Zhang, X. Zou, J. Shi, Preparation  
376 of boron nitrogen co-doped carbon quantum dots for rapid detection of Cr(VI),  
377 *Spectrochimica Acta Part a-Molecular and Biomolecular Spectroscopy*, 243 (2020).
- 378 [6] J. Wang, Z. Wu, S. Chen, R. Yuan, L. Dong, A novel multifunctional fluorescent sensor  
379 based on N/S co-doped carbon dots for detecting Cr (VI) and toluene, *Microchemical*  
380 *Journal*, 151 (2019).
- 381 [7] Y. Yang, X. Chen, Y. Wang, M. Wu, Y. Ma, X. Yang, A Novel Fluorescent Test Papers  
382 Based on Carbon Dots for Selective and Sensitive Detection of Cr (VI), *Front Chem*, 8  
383 (2020) 595628.
- 384 [8] A. Tall, F. Antônio Cunha, B. Kaboré, C. d'Angeles do E. S. Barbosa, U. Rocha, T.O. Sales,  
385 M.O. Fonseca Goulart, I. Tapsoba, J. Carinhanha Caldas Santos, Green emitting N,  
386 P-doped carbon dots as efficient fluorescent nanoprobe for determination of Cr(VI) in  
387 water and soil samples, *Microchemical Journal*, 166 (2021).
- 388 [9] X. Luo, P. Bai, X. Wang, G. Zhao, J. Feng, H. Ren, Preparation of nitrogen-doped carbon  
389 quantum dots and its application as a fluorescent probe for Cr(vi) ion detection, *New*  
390 *Journal of Chemistry*, 43 (2019) 5488-5494.
- 391 [10] S. Liu, J. Cui, J. Huang, B. Tian, F. Jia, Z. Wang, Facile one-pot synthesis of highly  
392 fluorescent nitrogen-doped carbon dots by mild hydrothermal method and their  
393 applications in detection of Cr(VI) ions, *Spectrochim Acta A Mol Biomol Spectrosc*, 206  
394 (2019) 65-71.
- 395 [11] B. Li, H. Ma, B. Zhang, J. Qian, T. Cao, H. Feng, W. Li, Y. Dong, W. Qin, Dually  
396 emitting carbon dots as fluorescent probes for ratiometric fluorescent sensing of pH  
397 values, mercury(II), chloride and Cr(VI) via different mechanisms, *Mikrochim Acta*, 186  
398 (2019) 341.
- 399 [12] Y. Ji, X. Zou, W. Wang, T. Wang, S. Zhang, Z. Gong, Co-Doped S, N-Carbon dots and its

- 400 fluorescent film sensors for rapid detection of Cr (VI) and Ascorbic acid, *Microchemical*  
401 *Journal*, 167 (2021).
- 402 [13] A.K. Singh, V.K. Singh, M. Singh, P. Singh, S.R. Khadim, U. Singh, B. Koch, S.H.  
403 Hasan, R.K. Asthana, One pot hydrothermal synthesis of fluorescent NP-carbon dots  
404 derived from *Dunaliella salina* biomass and its application in on-off sensing of Hg (II),  
405 Cr (VI) and live cell imaging, *Journal of Photochemistry and Photobiology a-Chemistry*,  
406 376 (2019) 63-72.
- 407 [14] R. Wang, L. Jiao, X. Zhou, Z. Guo, H. Bian, H. Dai, Highly fluorescent graphene  
408 quantum dots from biorefinery waste for tri-channel sensitive detection of Fe<sup>3+</sup> ions,  
409 *Journal of hazardous materials*, 412 (2021) 125096-125096.
- 410 [15] J. Shen, S. Shang, X. Chen, D. Wang, Y. Cai, Highly fluorescent N, S-co-doped carbon  
411 dots and their potential applications as antioxidants and sensitive probes for Cr (VI)  
412 detection, *Sensors and Actuators B: Chemical*, 248 (2017) 92-100.
- 413 [16] S. Chaudhary, S. Kumar, B. Kaur, S.K. Mehta, Potential prospects for carbon dots as a  
414 fluorescence sensing probe for metal ions, *Rsc Advances*, 6 (2016) 90526-90536.
- 415 [17] X. Zheng, S. Ren, L. Wang, Q. Gai, Q. Dong, W. Liu, Controllable functionalization of  
416 carbon dots as fluorescent sensors for independent Cr(VI), Fe(III) and Cu(II) ions  
417 detection, *Journal of Photochemistry and Photobiology A: Chemistry*, 417 (2021).
- 418 [18] M. Jia, L. Peng, M. Yang, H. Wei, M. Zhang, Y. Wang, Carbon dots with dual emission:  
419 A versatile sensing platform for rapid assay of Cr (VI), *Carbon*, 182 (2021) 42-50.
- 420 [19] F. Li, Y. Li, X. Yang, X. Han, Y. Jiao, T. Wei, D. Yang, H. Xu, G. Nie, Highly Fluorescent  
421 Chiral N-S-Doped Carbon Dots from Cysteine: Affecting Cellular Energy Metabolism,  
422 *Angewandte Chemie-International Edition*, 57 (2018) 2377-2382.
- 423 [20] H. Zhang, S. Kang, G. Wang, Y. Zhang, H. Zhaou, Fluorescence Determination of Nitrite  
424 in Water Using Prawn-Shell Derived Nitrogen-Doped Carbon Nanodots as Fluorophores,  
425 *Acs Sensors*, 1 (2016) 875-881.
- 426 [21] B. Zhang, Y. Liu, M. Ren, W. Li, X. Zhang, R. Vajtai, P.M. Ajayan, J.M. Tour, L. Wang,  
427 Sustainable Synthesis of Bright Green Fluorescent Nitrogen-Doped Carbon Quantum  
428 Dots from Alkali Lignin, *Chemsuschem*, 12 (2019) 4202-4210.
- 429 [22] L. Zhu, D. Shen, Q. Liu, C. Wu, S. Gu, Sustainable synthesis of bright green fluorescent  
430 carbon quantum dots from lignin for highly sensitive detection of Fe<sup>3+</sup> ions, *Applied*  
431 *Surface Science*, 565 (2021).
- 432 [23] Y. Liu, H. Yang, C. Ma, S. Luo, M. Xu, Z. Wu, W. Li, S. Liu, Luminescent Transparent  
433 Wood Based on Lignin-Derived Carbon Dots as a Building Material for Dual-Channel,  
434 Real-Time, and Visual Detection of Formaldehyde Gas, *ACS Appl Mater Interfaces*, 12  
435 (2020) 36628-36638.
- 436 [24] Y. Zhao, C. Ou, J. Yu, Y. Zhang, H. Song, Y. Zhai, Z. Tang, S. Lu, Facile Synthesis of  
437 Water-Stable Multicolor Carbonized Polymer Dots from a Single Unconjugated Glucose  
438 for Engineering White Light-Emitting Diodes with a High Color Rendering Index, *ACS*  
439 *Appl Mater Interfaces*, (2021).
- 440 [25] H. Zhao, X. Yu, C.-F. Li, W. Yu, A. Wang, Z.-Y. Hu, S. Larter, Y. Li, M. Golam Kibria, J.  
441 Hu, Carbon quantum dots modified TiO<sub>2</sub> composites for hydrogen production and  
442 selective glucose photoreforming, *Journal of Energy Chemistry*, 64 (2022) 201-208.
- 443 [26] J. Zhang, Y. Lin, S. Wu, X. Hou, C. Zheng, P. Wu, J. Liu, Self-photo-oxidation for

- 444 extending visible light absorption of carbon dots and oxidase-like activity, *Carbon*, 182  
445 (2021) 537-544.
- 446 [27] J. Wang, Q. Li, J. Zheng, Y. Yang, X. Liu, B. Xu, N. B-Codoping Induces  
447 High-Efficiency Solid-State Fluorescence and Dual Emission of Yellow/Orange Carbon  
448 Dots, *ACS Sustainable Chemistry & Engineering*, 9 (2021) 2224-2236.
- 449 [28] R. Wang, G. Xia, W. Zhong, L. Chen, L. Chen, Y. Wang, Y. Min, K. Li, Direct  
450 transformation of lignin into fluorescence-switchable graphene quantum dots and their  
451 application in ultrasensitive profiling of a physiological oxidant, *Green Chemistry*, 21  
452 (2019) 3343-3352.
- 453 [29] Q. Su, X. Yang, Promoting Room Temperature Phosphorescence through Electron  
454 Transfer from Carbon Dots to Promethazine, *ACS Appl Mater Interfaces*, 13 (2021)  
455 41238-41248.
- 456 [30] J. Li, H. Zhao, X. Zhao, X. Gong, Red and yellow emissive carbon dots integrated  
457 tandem luminescent solar concentrators with significantly improved efficiency,  
458 *Nanoscale*, 13 (2021) 9561-9569.
- 459 [31] W. Chang, B. Ning, Q. Xu, H. Jiang, Y. Hu, C. Li, Strongly coupled N-doped graphene  
460 quantum dots/Ni(Fe)OxHy electrocatalysts with accelerated reaction kinetics for water  
461 oxidation, *Chemical Engineering Journal*, (2021).
- 462 [32] L. Zhu, D. Shen, Q. Wang, K.H. Luo, Green Synthesis of Tunable Fluorescent Carbon  
463 Quantum Dots from Lignin and Their Application in Anti-Counterfeit Printing, *ACS  
464 Applied Materials & Interfaces*, (2021).
- 465 [33] C. Zhu, X. Dou, W. Li, X. Liu, Q. Li, J. Ma, Q. Liu, L. Ma, Efficient depolymerization of  
466 Kraft lignin to liquid fuels over an amorphous titanium-zirconium mixed oxide  
467 supported partially reduced nickel-cobalt catalyst, *Bioresour Technol*, 284 (2019)  
468 293-301.
- 469 [34] J. Zhu, F. Chen, Z. Zhang, M. Li, Q. Yang, Y. Yang, Z. Bao, Q. Ren, M-Gallate (M = Ni,  
470 Co) Metal–Organic Framework-Derived Ni/C and Bimetallic Ni–Co/C Catalysts for  
471 Lignin Conversion into Monophenols, *ACS Sustainable Chemistry & Engineering*, 7  
472 (2019) 12955-12963.
- 473 [35] H. Luo, M.M. Abu-Omar, Lignin extraction and catalytic upgrading from genetically  
474 modified poplar, *Green Chemistry*, 20 (2018) 745-753.
- 475 [36] W. Zhao, L.-P. Xiao, G. Song, R.-C. Sun, L. He, S. Singh, B.A. Simmons, G. Cheng,  
476 From lignin subunits to aggregates: insights into lignin solubilization, *Green Chemistry*,  
477 19 (2017) 3272-3281.
- 478 [37] M. Wang, J. Lu, X. Zhang, L. Li, H. Li, N. Luo, F. Wang, Two-Step, Catalytic C–C Bond  
479 Oxidative Cleavage Process Converts Lignin Models and Extracts to Aromatic Acids,  
480 *ACS Catalysis*, 6 (2016) 6086-6090.
- 481 [38] Z. Ding, F. Li, J. Wen, X. Wang, R. Sun, Gram-scale synthesis of single-crystalline  
482 graphene quantum dots derived from lignin biomass, *Green Chemistry*, 20 (2018)  
483 1383-1390.
- 484 [39] M. Si, J. Zhang, Y. He, Z. Yang, X. Yan, M. Liu, S. Zhuo, S. Wang, X. Min, C. Gao, L.  
485 Chai, Y. Shi, Synchronous and rapid preparation of lignin nanoparticles and carbon  
486 quantum dots from natural lignocellulose, *Green Chemistry*, 20 (2018) 3414-3419.
- 487 [40] S. Jing, Y. Zhao, R.-C. Sun, L. Zhong, X. Peng, Facile and High-Yield Synthesis of

488 Carbon Quantum Dots from Biomass-Derived Carbons at Mild Condition, *ACS*  
489 *Sustainable Chemistry & Engineering*, 7 (2019) 7833-7843.

490 [41] A. Abbas, L.T. Mariana, A.N. Phan, Biomass-waste derived graphene quantum dots and  
491 their applications, *Carbon*, 140 (2018) 77-99.

492 [42] L. Zhu, D. Shen, C. Wu, S. Gu, State-of-the-Art on the Preparation, Modification, and  
493 Application of Biomass-Derived Carbon Quantum Dots, *Industrial & Engineering*  
494 *Chemistry Research*, 59 (2020) 22017-22039.

495 [43] Y. Zheng, K. Arkin, J. Hao, S. Zhang, W. Guan, L. Wang, Y. Guo, Q. Shang, Multicolor  
496 Carbon Dots Prepared by Single- Factor Control of Graphitization and Surface  
497 Oxidation for High- Quality White Light- Emitting Diodes, *Advanced Optical Materials*,  
498 (2021).

499 [44] B. Wang, H. Song, Z. Tang, B. Yang, S. Lu, Ethanol-derived white emissive carbon dots:  
500 the formation process investigation and multi-color/white LEDs preparation, *Nano*  
501 *Research*, (2021).

502 [45] H. Li, S. Han, B. Lyu, T. Hong, S. Zhi, L. Xu, F. Xue, L. Sai, J. Yang, X. Wang, B. He,  
503 Tunable light emission from carbon dots by controlling surface defects, *Chinese*  
504 *Chemical Letters*, (2021).

505 [46] T. Zhang, J. Zhu, Y. Zhai, H. Wang, X. Bai, B. Dong, H. Wang, H. Song, A novel  
506 mechanism for red emission carbon dots: hydrogen bond dominated molecular states  
507 emission, *Nanoscale*, 9 (2017) 13042-13051.

508 [47] M. Park, H.S. Kim, H. Yoon, J. Kim, S. Lee, S. Yoo, S. Jeon, Controllable Singlet-Triplet  
509 Energy Splitting of Graphene Quantum Dots through Oxidation: From Phosphorescence  
510 to TADF, *Adv Mater*, 32 (2020) e2000936.

511 [48] X. Li, S.P. Lau, L. Tang, R. Ji, P. Yang, Sulphur doping: a facile approach to tune the  
512 electronic structure and optical properties of graphene quantum dots, *Nanoscale*, 6 (2014)  
513 5323-5328.

514 [49] Y. Chen, H. Lian, Y. Wei, X. He, Y. Chen, B. Wang, Q. Zeng, J. Lin,  
515 Concentration-induced multi-colored emissions in carbon dots: origination from triple  
516 fluorescent centers, *Nanoscale*, 10 (2018) 6734-6743.

517 [50] R. V. V. Gujar, H. Pathan, S. Islam, M. Tawre, K. Pardesi, M.K. Santra, D. Ottoor,  
518 Bioimaging Applications of Carbon dots (C. dots) and its Cystamine Functionalization  
519 for the Sensitive Detection of Cr(VI) in Aqueous Samples, *J Fluoresc*, 29 (2019)  
520 1381-1392.

521 [51] Y. Liu, Z. Chen, W. Li, C. Ma, P. Wu, X. Wu, S. Li, S. Liu, A nanocomposite probe  
522 consisting of carbon quantum dots and phosphotungstic acid for fluorometric  
523 determination of chromate(VI) with improved selectivity, *Mikrochim Acta*, 185 (2018)  
524 470.

525 [52] H.-Y. Zhang, Y. Wang, S. Xiao, H. Wang, J.-H. Wang, L. Feng, Rapid detection of Cr(VI)  
526 ions based on cobalt(II)-doped carbon dots, *Biosensors & Bioelectronics*, 87 (2017)  
527 46-52.

## Highlights

- Facile two-step route is proposed for synthesizing triple-emission fluorescent N, B-CQDs derived from lignin waste.
- The formation mechanism of N,B-CQDs from lignin-derived via two-step route is deeply excavated.
- The highly fluorescent tri-channel sensing platform based on N,B-CQDs sensor has been established.

**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Dekui Shen reports financial support was provided by National Natural Science Foundation of China. Dekui Shen reports financial support was provided by Key Research & Development Program of Jiangsu Province. Dekui Shen reports financial support was provided by Postgraduate Research & Practice Innovation Program of Jiangsu Province from the Education Department of Jiangsu.