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Trophic relationships and pelagic-benthic coupling during summer in the Barents Sea Marginal Ice Zone, revealed by stable carbon and nitrogen isotope measurements

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ABSTRACT: Food web structure and pathways from primary production were studied in pelagic, sympagic (ice-associated) and benthic communities during summer in a seasonally ice covered region of the northern Barents Sea. Stable isotopes of carbon (δ^{13} C) and nitrogen (δ^{15} N) were used as tracers of organic material through marine food webs and trophic levels of organisms, respectively. Measurements of δ^{15} N show that sympacic amphipods occupy the lowest trophic levels (ca. 2), for primary consumers, followed by zooplankton (2.0 to 2.6), benthic suspension and deposit feeders (2.2 to 3.7), benthic carnivores (3.6 to 4.4) and fishes (3.3 to 4.4). The δ^{13} C values indicate that zooplankton mainly graze on suspended particulate organic material (POM). Sympagic amphipods derive most of their energy from ice POM, but some species had δ^{13} C values indicating that phytoplankton also contributes to their energy intake. δ^{13} C values of some components of the benthic community suggest that POM settling out of the water column is efficiently exploited by the benthic fauna. Elevated δ^{13} C values of the benthic fauna relative to zooplankton at some stations indicate that the degree of pelagic-benthic coupling at stations separated by only 90 km is determined by a combination of factors, including water-mass properties and the primary-production regime. These results may qualify findings of previous studies that have sampled from discrete locations or have pooled specimens collected from a broad area to make conclusions about food webs on a regional scale.

KEY WORDS: Stable isotopes $\cdot \, \delta^{13}C \cdot \delta \, ^{15}N \cdot$ Food web \cdot Arctic \cdot Barents Sea \cdot Marginal Ice Zone \cdot Pelagic–benthic coupling

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INTRODUCTION

Marginal Ice Zones (MIZs) are characterised by high annual primary production (Sakshaug & Skjoldal 1989, Niebauer et al. 1990), with significant seasonal and inter-annual variability (Falk-Petersen et al. 2000b, Engelsen et al. 2002). The seasonal retreat of the seaice edge is often associated with a pronounced phytoplankton bloom, governed by entrainment of phytoplankton and nutrients in a shallow, mixed surface layer, formed due to ice melt (Niebauer et al. 1990, Strass & Nöthig 1996, Engelsen et al. 2002). A latitudinal 'migration' of primary production, the so-called ice-edge effect, is a ubiquitous feature of seasonally ice-covered regions, and has a significant influence on food web structure and function in these regions (Sakshaug & Skjoldal 1989, Hansen et al. 2003). Dynamic ice conditions, however, may result in an ill-defined ice edge, creating a mosaic of ice-covered and open-water areas, where spatial and temporal patterns in the traditional MIZ bloom are decoupled (Falk-Petersen et al. 2000b, Engelsen et al. 2002).

There are 2 principal sources of primary production in the offshore Arctic MIZ; phytoplankton and ice algae (Sakshaug & Skjoldal 1989). Phytoplankton is responsible for the bulk of the annual production, but is confined to open waters or leads in the ice, and phytoplankton blooms tend to occur later in the year than ice algal production. Ice algae are mostly represented by diatoms growing in dense layers within or attached to the underside of the ice from February until May or June, when melting starts (Syvertsen 1991). Legendre et al. (1992) estimated that ice algal production in the Arctic averages 10 g C m^{-2} yr⁻¹. In the Barents Sea MIZ, ice algae may contribute 16 to 22% to the annual primary production, depending on the extent of ice-free areas (Hegseth 1998). Thus, in years with extended ice cover, the ice algae may comprise an important fraction of the primary production and vertical particle export in seasonally ice-covered regions (Michel et al. 2002). Ice algal-based detritus is an important food source for the sympagic, i.e. iceassociated, fauna (Bradstreet & Cross 1982, Poltermann 2001). The contribution of ice algae to the diet of other ecosystem components, i.e. zooplankton and benthos, however, remains to be quantified. Grazing by copepods on ice algae at the ice-water interface has been observed in the Canadian Arctic (Conover et al. 1986), which suggests that ice algae may be an important early season food source before phytoplankton become abundant.

The fate of primary production in Arctic shelf seas depends on the spatial and temporal variability in distribution and species composition of both the production regime and the pelagic grazers (Wassmann et al. 1996). From a food-web standpoint, phytoplankton- and ice algal-based organic material can either be retained in the upper water column or be exported to the benthos (e.g. Carroll & Carroll 2003). Primary trophic pathways, therefore, are determined in large by the match or mismatch between pelagic herbivores and the peak of primary production (Eilertsen et al. 1989, Falk-Petersen et al. 1999). The degree of heterotrophy in the plankton community has considerable implications for the retention versus vertical export of surface primary production (Olli et al. 2002). The highest vertical carbon fluxes are typically recorded during spring (Andreassen & Wassmann 1998), before the pelagic community has reached a state of development in which production and consumption are balanced (Olli et al. 2002, Verity et al. 2002, Sherr et al. 2003), resulting in a pulsed food supply to the benthos (Grebmeier et al. 1995, Wassmann et al. 1996). Benthic communities in Arctic shelf seas have been found to reflect the primary production regime of the overlying water column, in terms of biomass, abundance and production, which suggests a tight pelagic-benthic coupling in these regions (Grebmeier et al. 1988, Highsmith & Coyle 1990, Ambrose & Renaud 1995, Piepenburg et al. 1997).

Stable isotopes may permit the identification of energy sources assimilated by consumers, averaged over the turnover time for the species or tissue studied (Kling et al. 1992, Post 2002). In comparison, dietary analyses only provide a measure of recently ingested food, and are limited by the extent to which food items in stomachs can be identified. The stable isotope technique is particularly useful in food web studies in remote regions, such as the MIZ, where the possibility for long-term or high-frequency sampling is limited. When 2 isotopically distinct carbon sources are available to the food web, their relative importance can be estimated from the δ^{13} C value of consumer tissue relative to those of endpoints, corresponding to each source (Fry & Sherr 1984, Post 2002). Stable nitrogen isotopes (i.e. $\delta^{15}N$ values) can be used to estimate trophic levels of organisms in the community (Wada et al. 1987, Hobson & Welch 1992). Previous isotope studies have shown that in some regions of the Arctic, benthic communities may receive a substantial amount of ungrazed POM from the overlying water column (Hobson & Welch 1992, Hobson et al. 1995), whereas other regions are characterised by weaker pelagic-benthic coupling (McConnaughey & McRoy 1979, Dunton et al. 1989, Hobson et al. 2002). Some recent studies also suggest that ice algae may comprise a significant proportion of the diets of some benthic organisms (Hobson et al. 2002, McMahon et al. 2006, this volume).

We used stable isotopes to study trophic relationships and the transfer of energy in pelagic, sympagic (ice-associated) and benthic communities, and in fish fauna in the north-western Barents Sea MIZ. The shallow, seasonally ice-covered Barents Sea borders the deep Arctic Ocean, which is ice-covered yearround. The hydrography of the Barents Sea is characterised by both Arctic and Atlantic water masses (Loeng 1991), resulting in a dynamic environmental setting and highly variable primary production regimes, which make this a complex region for the study of ecosystem structure and function (Falk-Petersen et al. 2000b, Engelsen et al. 2002). Consequently, we might expect trophic structures to vary spatially and temporally in response to ice cover, hydrographic parameters and production.

MATERIALS AND METHODS

Study area. The study area covers the north-western Barents Sea and the Arctic Ocean shelf-break region. Samples were collected from the RV 'Jan Mayen' (University of Tromsø) in July 2003 at stations located along a south-north transect between ca. 76 and 79°N, approximately along the 30°E meridian east of Spitsbergen (Table 1, Fig. 1). During the time of sampling, all stations had a 40 to 70% ice cover, which consisted of 80 to 90% first-year ice (Table 1). The ice extended relatively far to the south, but was in a melting stage. CTD profiles showed that the water column at Stns I and IV was dominated by Atlantic water (AtW) overlain by colder water with Arctic characteristics (ArW). Stn II was located in a region separated from the Atlantic water influence by topographic control and dominated by ArW with lower salinity than at the stations further south. At Stn III, the water column was mostly composed of ArW, but the bottom layer (140 m to bottom) was dominated by AtW,

likely a southward return-flow from the West Spitsbergen Current circulating around Spitsbergen to the north (A. Sundfjord, University of Bergen, pers. comm.).

Sampling and analyses. Particulate organic material (POM) from the ice and sympagic amphipods were sampled by SCUBA-diving below the ice. Ice POM and detritus were collected using 20 µm mesh nets mounted in electrical suction pumps, whereas a 500 µm mesh was used for animals. Depth-integrated water samples for suspended (water column) POM were collected with a Rosette sampler from 0, 5, 10 and 20 m depths. Samples of settling material were obtained from 24 h deployments of sediment traps, moored from the ice edge at 40, 90 and 200 m depth. The traps (KC Denmark Research Equipment) had duplicate transparent cylinders, with an internal diameter of 7.2 cm and a 6:1 aspect ratio, mounted in a gimballed frame, equipped with a vane. Because some sediment trap samples were too small for stable isotope analyses, data from the same depths is not available from every station.

Sediment was sampled with a multicorer (Bowers & Connely) containing 4 replicate tubes of 10 cm diameter. The sediment-surface nepheloid layer was sampled by siphoning the top millimetre of the sediment. Separate sediment samples were taken either from the multicorer or sub-sampled from box cores and sliced in 1 cm intervals to 5 cm depth. Samples of ice and suspended POM, settling material, and the nepheloid layer were filtered on pre-combusted (450°C, 4 h) Whatman GF/C glass fibre filters and stored frozen at -20° C.

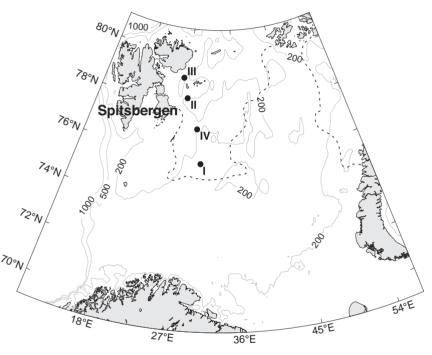


Fig. 1. Map of the studied region with bathymetry and main sampling locations indicated. Dashed line indicates approximate position of the ice edge (10 to 40% ice concentration) at the start of the cruise (10 July 2003)

Table 1. Stations visited, ice cover, proportions of first-year and multi-year and first-year ice (FYI/MYI), bottom depth, mixed layer depth (MLD), surface (mixed layer) and bottom (deepest 10 m) temperature and salinity (mean ± SD). Plankton, sympagic and benthic samples were collected at all main stations (I, II, III and IV). Stns IVa and IVb are bottom trawl (fish sampling) and horizontal MIK-trawl (zooplankton) stations. respectively

Stn	Position	Date visited	Ice coverage	FYI/MYI	Bottom	MLD	Tempera	ture (°C)	Salinit	y (psu)
	(latitude, longitude)	(2003)	(%)	(%)	depth (m)	(m)	Surface	Bottom	Surface	Bottom
I	75° 40.5' N, 30° 15.4' I	E 10 Jul	40-70	90/10	343	53	0.15 ± 1.41	0.06 ± 0.03	33.96 ± 0.81	34.92 ± 0
II	78° 14.8' N, 27° 1.3' E	14 Jul	40-70	90/10	319	35	-1.27 ± 0.57	0.26 ± 0.02	33.72 ± 0.68	34.85 ± 0
III	79° 2.1′ N, 25° 51.8′ E	E 16 Jul	50-70	80/20	213	43	-1.35 ± 0.38	1.87 ± 0.12	33.65 ± 0.62	34.74 ± 0.03
IV	77° 2.7′ N, 29° 18.9′ I	E 19 Jul	40-70	90/10	227	41	-1.52 ± 0.19	1.15 ± 0.03	33.91 ± 0.58	34.97 ± 0
IVa	77° 3.2′ N, 28° 19.1′ I	E 19 Jul	0	-	178 - 192	_	-	-	_	-
IVb	76° 58.4' N, 29° 1.7' E	19 Jul	0	_	239	_	_	_	_	_

Zooplankton was collected with vertical net tows from bottom to surface or bottom to 50 m with a WP2 net (mesh size 180 µm), and from bottom to surface with an MIK trawl ('Method Isaac Kidd'; mesh size 1000 µm, opening 3.14 m^2). The MIK trawl was also used for horizontal tows in open waters close to the main stations (Table 1). Zooplankton and sympagic amphipods were sorted into species and size groups, and kept live in filtered seawater in screened, falsebottom chambers at ca. 0°C for 24 to 48 h until evacuation of gut contents. Several individuals of the same species and size (length or developmental stage) were combined in order to obtain sufficiently large samples.

Benthic invertebrates were collected from a triangular dredge (1 m per side) and from a Campelen 1800 bottom trawl (Table 1). Animals captured were sorted and kept in filtered seawater for evacuation of gut contents for 24 to 48 h. Small animals, such as pycnogonids and polychaetes, were analysed whole. Bivalves and gastropods were analysed based on their soft parts or only muscle tissue (larger animals). Sea stars were dissected, and the part of the stomach protruding to the arms (i.e. the pyloric caecum) was removed. Analyses were generally performed on 1 arm ind.⁻¹. Fish caught in the trawl were measured and dissected, and dorsal muscle samples were analysed for stable isotopes.

All samples were stored at -20°C until analysis. Prior to analyses, all samples were rinsed in distilled water in order to remove salt. Lipids were removed from all samples in order to reduce the variability of δ^{13} C values among species, seasons and developmental stages due to different body content of lipids, which are relatively depleted in ¹³C (Attwood & Peterson 1989, Hobson & Welch 1992). Carbonates were removed as well (Hobson et al. 1995). Samples were dried at 60° C, or freeze dried, and ground to a fine powder using mortar and pestle. Pre-treatment of the different types of samples differed slightly, as samples of the benthic fauna were analysed at the laboratory of K. Hobson (Saskatoon, Canada), whereas all other samples were analysed at the Institute for Energy Technology (Kjeller, Norway); these laboratories apply different methods for removal of carbonates and lipids.

Stable isotope ratios are expressed as a per mille (‰) enrichment relative to international standards according to the relationship:

$$\delta X = [(R_{\text{sample}} / R_{\text{standard}}) - 1] \times 1000$$

where X is ¹³C or ¹⁵N and R is the corresponding ratio ¹³C/¹²C or ¹⁵N/¹⁴N. Standards for δ^{13} C and δ^{15} N are PeeDee Belemnite and atmospheric nitrogen N₂ (air), respectively.

Parallel analyses of split samples revealed slight differences in the isotope enrichment values for some, although not all, species between the 2 laboratories (T. Tamelander unpubl. data). Even when statistically significant (maximum difference 1.1% for $\delta^{15}N$ and 1.4% for $\delta^{13}C$), the differences were small and would only have minor relevance for the food web model as a whole. Assuming a 1.4% offset for all samples analysed at the Canadian laboratory, corresponding to the maximum difference between the 2 laboratories, and recounting the $\delta^{13}C$ values accordingly would not affect our conclusions.

Trophic level values of consumer organisms were estimated assuming an average increase in δ^{15} N values of 3.4‰ between consecutive trophic levels (J. Søreide, Akvaplan-niva, Tromsø, pers. comm.). Previous food web studies have applied fractionation factors for δ^{15} N between 3 and 4‰ (Hobson et al. 1995, Post 2002, Bode et al. 2003, Iken et al. 2005). Trophic level values (TL) of consumer organisms were thus calculated according to the relationship:

$$TL = (\delta^{15}N_{Consumer} - \delta^{15}N_{PP}) / 3.4 + 1$$

where PP denotes primary producers (ice algae or phytoplankton, depending on the anticipated primary food source). The average $\delta^{15}N$ value of all samples of suspended POM (4.7‰), excluding 1 outlier, was used to calculate trophic levels of the pelagic, benthic and fish fauna, whereas the average ice algal $\delta^{15}N$ value (4.8‰) was used for trophic levels of sympagic amphipods.

Statistical analyses. Statistical tests were performed using Statistica 7. The isotope data was not normally distributed (Shapiro-Wilk's test, p < 0.05), and nonparametric statistical tests were therefore applied. Mann-Whitney *U*-tests were used for testing differences between 2 independent samples. A Kruskal-Wallis test, with a multiple comparisons post hoc test, was used for comparing several groups. For comparing the δ^{13} C and δ^{15} N signatures between the different animal communities sampled (pelagic, sympagic, benthic and fishes), only data from Stn IV (Stns IV, IVa and IVb) where all groups were sampled, was included in the analysis.

RESULTS

Stable isotope composition of particulate material and the biota

The lowest δ^{13} C values were found in suspended POM. Ice POM was significantly more enriched in ¹³C than suspended POM (Mann-Whitney *U*-test, p = 0.004, n = 4 and n = 12 for suspended and ice POM, respectively), but no difference was found for δ^{15} N (Mann-Whitney *U*-test, p = 0.8, n = 4 and n = 9 for

suspended and ice POM, respectively). Material from the sediment traps had δ^{13} C values that were 0.9 to 5.6% higher compared to the suspended POM (Fig. 2). Compared to settling POM, material from the sediment

nepheloid layer was depleted in $^{13}\mathrm{C}$ by 2.0 to 4.0% at Stns II, III and IV and enriched by 0.6% at Stn I. The sediment had consistently higher $\delta^{13}\mathrm{C}$ values than the nepheloid layer (Fig. 2).

Whereas ice fauna and zooplankton showed little variation in $\delta^{13}C$ values among species, benthic invertebrates revealed a wide range in values (Table 2, Fig. 3). At Stn IV, where samples of all organism groups were collected, zooplankton and sympagic amphipods had significantly lower $\delta^{13}C$ values than the benthos (Kruskal-Wallis test, p < 0.0001, Table 3). Fishes were significantly more enriched in ¹³C than zooplankton, but depleted by ca. 3‰ compared to benthos (Kruskal-Wallis test, p < 0.001, Table 3). Zooplankton had the same $\delta^{15}N$ values as sympagic amphipods and benthos, whereas fishes had significantly higher values than all other groups (Kruskal-Wallis test, p < 0.001, Table 3).

Zooplankton and ice fauna from all stations sampled were equally enriched in $^{13}\mathrm{C}$ (Mann-Whitney U-test, p = 0.5, n = 55 for ice fauna and n = 51 for zooplankton), with $\delta^{13}\mathrm{C}$ values around –21‰. Zooplankton had significantly higher (ca. 3‰) $\delta^{15}\mathrm{N}$ values than the ice fauna (Mann-Whitney U-test, p < 0.0001), however.

Within the group of sympagic amphipods, Apherusa glacialis and Onisimus glacialis had significantly higher $\delta^{13}C$ and lower $\delta^{15}N$ values, respectively, than Gammarus wilkitzkii and Onisimus nanseni (Kruskal-Wallis tests, p < 0.001, Tables 2 & 3). Among the zooplankton species, the pelagic amphipod Themisto libellula had significantly lower δ^{13} C values than *Calanus hyper*boreus and krill (Kruskal-Wallis test, p < 0.001, Table 3). Krill and T. libellula generally had higher $\delta^{15}N$ values than copepods (Table 2). A statistically significant difference was only found between Thysanoessa longicaudata and C. hyperboreus, however (Kruskal-Wallis test, p < 0.01, Table 3).

Benthic amphipods had δ^{13} C values around -20%, similar to pelagic and sympagic amphipods (Table 2). Low δ^{13} C values were also observed in suspensionfeeding organisms, such as Bryozoa (-22.7%) and the

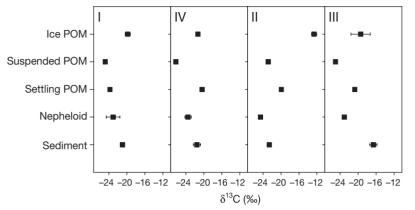


Fig. 2. δ^{13} C values (‰) of ice POM (particulate organic matter), suspended and settling POM, the nepheloid layer and sediment. Bars show standard deviation for ice POM, nepheloid layer and sediment. Station numbers are indicated in upper left panel corners, and are presented from south to north. Sediment trap depths at Stn I: 200 m, Stn IV: 90 m, Stn II: 40 m and Stn III: 90 m

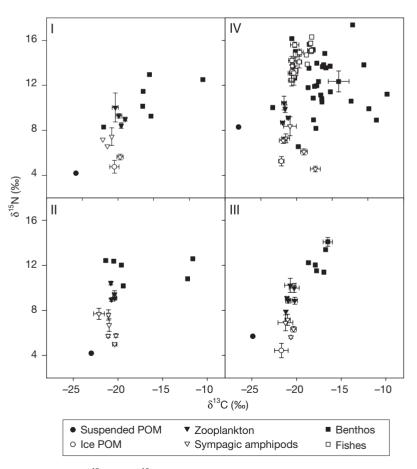


Fig. 3. Plots of δ^{13} C and δ^{15} N (‰) values (means ± SD) for organisms in the Barents Sea food web. Station numbers are indicated in upper left panel corners

Table 2. Stable carbon (8¹³C) and nitrogen (8¹⁵N) isotope values (‰, mean ± SD), number of samples (n), mean trophic level (TL), size (copepod size is given as developmental stage — CV: Copepodid Stage V; Fem: adult females) of sampled individuals and their trophic mode (H: herbivore; O: omnivore; C: carnivore; SF: suspension-feeder; DF: deposite a sit-feeder; SC: scavenger; P: pelagic fishes; D: demosal fishes; BP: benthic-pelagic species, based on the literature). Fish size is fork length (mm)

	Size (mm)	n 8	$\delta^{13}C$ Stn	.I 8 ¹⁵ N TL	4	→ Stn II 1 8 ¹³ C	Π δ ¹⁵ N	Ĕ	n $\delta^{13}C$	Stn IIIδ ¹⁵ Ν	IL	n $\delta^{13}C$	Stn IV δ^{15} N	
POM / algae Suspended POM Ice POM Ice POM		1 -2 3 -20.4	-24.7 -20.4 ± 0.5	4.2 4.8 ± 0.6		$\begin{array}{rrr} 1 & -23.0 \\ 3 & -12.6 \pm 0.5 \end{array}$	4.2		$\begin{array}{ccc} 1 & -24.9 \\ 3 & -21.7 \pm 0.8 \\ 3 & -17.3 \pm 0.4 \end{array}$	5.7 3 4.4 ± 0.6		$\begin{array}{rrr} 1 & -26.5 \\ 3 & -21.7 \pm 0.3 \end{array}$	8.3 3 5.3 ± 0.4	
Sympagic amphipods Apherusa glacialis (H ^{1,2}) Apherusa glacialis (H ^{1,2}) Commonone anticited and 30	4-6 7-12	3 -19.8	-19.8 ± 0.4	5.6 ± 0.3 1.2			5 ± 0.1 5.8 ± 0.2 5.7 ± 0.2		$\begin{array}{rrr} 3 & -20.6 \pm 0.1 \\ 3 & -20.3 \pm 0.3 \end{array}$	$\begin{bmatrix} 5.6 \pm 0.1 \\ 6.3 \pm 0.3 \end{bmatrix}$	$1.2 \\ 1.4$	3 −19.2 ± 0.4	$1 6.1 \pm 0.3$	3 1.4
Gammarus wuktizku (H) Gammarus wilkitzkii (O,C ^{1,3}) Gammarus wilkitzkii (O, ^{C^{1,3})} Onisimus glacialis (O ^{1,2})	••	$\begin{array}{ccc} 1 & -2 \\ 2 & -20.5 \\ 1 & -1 \end{array}$	-21.2 -20.7 ± 0.3 -18.2	$6.6 1.5 7.5 \pm 0.8 1.8 5.3 1.1 5.3 1.1 7.7 $		$\begin{array}{cccccccccccccccccccccccccccccccccccc$	5.7 ± 0.1 6.7 ± 0.6 7.6 ± 0.5	1.3 1.8 0	$\begin{array}{rrr} 3 & -21.2 \pm 0.8 \\ 3 & -21.0 \pm 0.3 \end{array}$	$\begin{array}{ccc} 6.9 \pm 0.7 \\ 7.2 \pm 0.5 \end{array}$	1.6 1.7	$\begin{array}{rrrr} 3 & -21.2 \pm 0.4 \\ 3 & -20.7 \pm 0.7 \\ 3 & -17.9 \pm 0.6 \\ \end{array}$	1 7.3 ± 0.4 7 8.4 ± 0.8 6 4.6 ± 0.3	1 1.7 3 2.0 3 0.9
Zooplankton Calanus glacialis (H ⁴)		3 -20.9	.4	1.3		1			$3 -20.9 \pm 0.3$	8.9 ± 0.2	2.2			
Calanus glactalis (H ⁺) Calanus hyperboreus (H ⁴) Calanus hyperboreus (H ⁴) Themisto libellula (O ²)	Fem CV Fem 4-10	3 -19.2	-19.2 ± 0.2	9.0 ± 0.2 2.3		$3 -20.4 \pm 0.2$ $3 -20.7 \pm 0.1$	9.1 ± 0.1 9.0 ± 0.2	2.3	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$			$3 -21.6 \pm 0.2$		
Themisto Ilbellula (O ⁻) Thysanoessa inermis (H ⁵) Thysanoessa loncicandata	26-35 25-30	3 -19.6	-19.6 ± 0.2	8.4 ± 0.3 2.1		$3 - 20.4 \pm 0.2$	9.4 ± 0.3	2.4	$3 -20.3 \pm 0.6$	10.0 ± 0.4	2.6	$3 -21.4 \pm 0.3$ $3 -21.3 \pm 0.1$	$5 10.4 \pm 0.0$ $1 9.9 \pm 0.3$	0 2.7 3 2.5
(C ⁵)	13 - 18	3 -19.9	-19.9 ± 0.4	9.3 ± 0.2 2.4		$3 - 20.8 \pm 0.1$	10.4 ± 0.2	2.7	$3 -20.7 \pm 0.6$	10.2 ± 0.6	2.6			
Benthic invertebrates Cnidaria Anemone, unid. (SF ^{6,7}) Mollusca												1 -18.6	13.5	3.6
Bivalvia Astarte spp. (SF, DF) Chlamys islandica (SF)					1	1 -21.4	12.4	3.3	1 -18.7	12.2	3.2	$\begin{array}{rrr} 1 & -18.1 \\ 1 & -17.2 \end{array}$	$10.9 \\ 10.5$	2.8 2.7
Gastropoda Gastropod, unid.												1 -16.7	13.6	3.6
Bryozoa Erect, lobed Bryozoa (SF ⁶) Erect, lobed Bryozoa (SF ⁶)												$\begin{array}{ccc} 1 & -11.1 \\ 1 & -22.7 \end{array}$	8.9 10.0	2.2 2.6
Crustacea Ampeliscidae (SF) Amphipod, unid. <i>Anonyx</i> sp. (DF, Sc ⁸) <i>Eupagurus pubescens</i>												$\begin{array}{ccc} 1 & -19.8 \\ 1 & -20.2 \\ 1 & -20.1 \\ 1 & -18.7 \end{array}$	6.6 12.7 15.0 11.8	1.5 3.3 4.0 3.1

continued)	
Table 2 (

Sample Size (mm)	n – n		δ ¹³ C Stn I	$\delta^{15}N$	Ľ	- F	Stn II $\delta^{13}C$	I δ ¹⁵ Ν	TL	, E	Stn III $\delta^{13}C$	Π δ ¹⁵ N	TL	ц	δ ¹³ C	[V	L
Mesidotea sabini Pandalus borealis (O, C^9) Sclerocrangon sp. (O, C^9) Shrimp, unid. (1) Shrimp, unid. (2) Stegocephalus sp.	-		- 16.3	6.0	2.3		- 19.6	12.0	3.2		-17.7	11.5	3.0		-18.0 -17.0 -17.5 -17.1 -17.1	$11.9 \\ 13.8 \\ 12.4 \\ 13.7 \\ 15.2 \\ 15.2 \\$	3.1 3.7 3.3 3.6 4.1
Echinodermata Ophiacantha bidentata (SF, $O^{7,10}$) Ophiopholis aculeata (SF DF O, O^{10})	-		د 10	ر د د د	۲۲ ۲۲	1	-11.6	12.6	3.3					~ ~	-9.9 -11 q	11.2 9.9	2.9
(DF, O, C, C) Ophiopleura borealis (DF, O, C ¹⁰) Ophiura sarsi (DF, O, Sc ¹⁰)	-			2	5	1	-12.2	10.8	2.8	1	-16.7	13.4	3.6		-13.9	10.6	5.7
Crenodiscus crispatus (DF ¹⁰) Crossaster papposus (C ¹⁰) Pontaster tenuispinus (DF ¹⁰) Poraniomorpha tumida							-19.4	10.2	2.6	1	-16.9	11.4	3.0		-16.2 -12.5 -16.2 -13.7	11.4 13.8 13.7 17.4	3.0 3.7 3.6 4.7
Holothurian (DF ¹⁰) Crinoid (SF ¹⁰) Echiura, unid. (DF) Sipunculida (DF) <i>Strongylocentrotus</i> sp. (H ¹⁰)	-	-	-16.5	13.0	3.4					7	-17.9	12.0	3.2		-16.8 -17.2 -17.3 -17.3	14.8 10.8 11.1 8.2	4.0 2.8 2.9 2.0
Polychaeta Lumbrinereis fragilis (DF, C, Sc ¹¹) Maldane sarsi (DF ¹¹) Nephtys sp. (DF, C, Sc ¹¹) Pectinariidae			-17.2 -17.2	10.1 11.5	3.0	1	-20.5	12.4	3.3	1	-16.5 ± 0.5 14.1 ± 0.4	14.1 ± 0.4	1 3.8	- 7 - 7	-17.7 -15.3 ± 1.2 -18.1	12.0 12.3 ± 0.9 8.9	3.1 3.2 2.2
Pycnogonida Pycnogonida, unid. (C ¹²)														1	-20.5	16.2	4.4
Porifera Sponge 1, unid. (SF, O) Sponge 2, unid. (SF, O)															-18.6 -17.7	15.7 14.0	4.2 3.7
Fishes Gadus morhua (BP, O ¹³) 130–149 Mallotus villosus (P ¹³) 146–169 Himmorlossoides nlatessoides	149 169													6 5	-20.1 ± 0.2 -20.1 ± 0.3	14.8 ± 0.3 13.3 ± 0.4	3 4.0 4 3.5
(D ¹³) 86–98 Hippoglossoides platessoides	98													5	-18.8 ± 0.2	c.	1 3.7
(D ¹³) 178–265	265												۔ 2	-19.7 ± 0	$-19.7 \pm 0.414.9 \pm 0.3$	3 4.0	

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Sample	(mm) n $\delta^{13}C$	8 ¹⁵ N TL	n δ^{13} C δ^{15} N TL	L Nerg Derg u		
Reinhardtius hippoglossoides						
(BP^{13}) 2.2 2	92 - 94				$2 -20.6 \pm 0.4 12.5 \pm 0.5 3.3$	12.5 ± 0.5 3.3
<i>Clupea harengus</i> (P ¹³)	202				1 -20.3	12.8 3.4
Lumpenus lampraetaeformis						
(D^{13})	223-265				$5 -18.4 \pm 0.3$	$15 \pm 0.2 4.0$
Leptoclinus maculatus (D ¹³)	146 - 170				$5 -18.3 \pm 0.4 15.1 \pm 0.2$	$15.1 \pm 0.2 \ 4.1$
Liparis liparis (O ¹³)	112 - 144				$5 -20.6 \pm 0.4 13.1 \pm 0.7$	13.1 ± 0.7 3.5
$Lycodes$ sp. (D^{13})	100				1 -18.9	14.7 3.9
$Lycodes \ rossi \ (D^{13})$	207				1 -18.3	16.3 4.4
Eumicrotremus spinosus (D ¹³)	55 - 71				$3 -20.4 \pm 0.3$	13.8 ± 0.7 3.7
Leptagonus decagonus (BP ¹³)	154 - 180				$5 -20.2 \pm 0.5$	$15.6 \pm 0.2 \ 4.2$
Triglops murrayi (D ¹³)	101 - 134				$5 - 19.7 \pm 0.3$	14.1 ± 0.5 3.8
Triglops nybelini	105 - 111				$5 -20.5 \pm 0.1$	13.2 ± 0.3 3.5
Artediellus atlanticus (D ¹⁴)	90 - 109				$5 -18.4 \pm 0.2$	$15.7 \pm 0.2 4.2$
Boreogadus saida (P ¹³)	91 - 95				$5 -20.3 \pm 0.5$	13.1 ± 1.1 3.5
Boreogadus saida (P ¹³)	184 - 206				$5 -20.5 \pm 0.4$	14.2 ± 0.4 3.8

clam Astarte sp. (–21.4 to –18.1‰). Deposit-feeding omnivorous decapods and polychaetes generally had higher $\delta^{13}C$ values (–20.5 to –15.3 ‰). The highest $\delta^{13}C$ values were observed in the brittle stars *Ophiura sarsi* and *Ophiacantha bidentata* (–12.2 and –9.9‰, respectively, Table 2).

Of the fishes sampled at IVa, 2 groups with different δ^{13} C values could be identified (Table 2). Demersal fishes were significantly more enriched than pelagic species (Mann-Whitney *U*-test, p < 0.0001, n = 37 for pelagic and n = 33 for demersal fishes).

Spatial variations in $\delta^{13}C$

The $\delta^{13}C$ values of suspended POM were elevated at Stn II (-23.0%) compared to Stns I, III and IV (-26.5 to -24.7 ‰, Table 2). The δ^{13} C values of the settling material generally followed those of the suspended POM (Fig. 2). Zooplankton generally had δ^{13} C values reflecting those of the suspended POM, although with a narrower range (-21.6 to -19.2%) (Fig. 3). The δ^{13} C values of the benthic fauna, however, showed an opposite trend compared to the suspended POM and the zooplankton. Several species (Astarte sp., Pandalus borealis, Lumbrinereis fragilis, Ctenodiscus crispatus) had lower δ^{13} C values at Stn II than at the other stations. At Stn II, the δ^{13} C values of these species ranged from -21.4 to -19.4%, which is approximately the same as for pelagic grazers (Fig. 3). At Stns I, III and IV, the same species had $\delta^{13} C$ values between –18.7 and -16.2‰, more than 3‰ higher than for pelagic grazers (Table 2).

DISCUSSION

Our results demonstrate a variable degree of pelagic-benthic coupling over a relatively small spatial scale. At 1 location, fresh phytodetritus reached the seabed and was efficiently exploited by the benthos, as indicated by the similarity in δ^{13} C values for the suspension- and surface-depositfeeding benthos and pelagic grazers. At other locations, higher enrichment of ¹³C in the benthos relative to the zooplankton and sympagic fauna suggests weaker pelagic-benthic coupling. Distribution of water masses and the associated primary production regime seem to be important factors governing pelagic-benthic coupling in the northern Barents Sea. The spatial heterogeneity of the system is a feature that needs to be considered in food web studies in environments characterised by complex and dynamic environmental settings, such as marginal ice zones.

Table 3. Summary of results from Kruskal-Wallis multiple comparisons tests. Animal groups or species are arranged from left to right in order of increasing δ^{13} C (less negative) or δ^{15} N values. Statistically homogeneous groups (p > 0.05) are marked with lines (Zp: zooplankton; Sa: sympagic amphipods; F: fishes; B: benthos; On: *Onisimus nanseni*; Gw: *Gammarus wilkitzkii*; Ag: *Apherusa glacialis*; Og: *Onisimus glacialis*; TI: *Themisto libellula*; Cg: *Calanus glacialis*; T lo: *Thysanoessa longicaudata*; Ch: *Calanus hyperboreus*; Ti: *Thysanoessa inermis*)

Comparison	Isotope	Stations included	Ν	p-value		Gro	up comparis	ons ———	
Animal groups	$\delta^{13}C$	IV	134	< 0.0001	Zp	Sa	F	В	
	$\delta^{15}N$	IV	134	< 0.0001	Sa	Zp	В	F	
Sympagic fauna	$\delta^{13}C$	I, II, III, IV	54	< 0.0001	On	Gw	Ag	Og	
	$\delta^{15}N$	I, II, III, IV	53	< 0.0001	Og	Ag	Gw	On	
Zooplankton	$\delta^{13}C$	I, II, III, IV	51	0.0003	<u>T1</u>	Cg	T lo	Ch	Ti
	$\delta^{15}N$	I, II, III, IV	51	0.005	Ch	Tl	Cg	Ti	T lo

Sources of organic C to the food web

Suspended POM, derived mainly from phytoplankton, was found to have significantly lower $\delta^{13}C$ values than ice POM (-24.8 versus -20.3%, on average), although there was considerable variability in values for both groups. Samples collected in the following summer (2004) at stations further north at the shelf break and in the Arctic Ocean revealed the same pattern, although the δ^{13} C values of both suspended POM and ice POM were depleted by ca. 2% compared to the 2003 data (T. Tamelander unpubl. data). Similar enrichment patterns have been observed in other regions of the Arctic (Hobson et al. 1995, Cooper et al. 2002) as well as the Antarctic (Gibson et al. 1999). The wide range in δ^{13} C values in both groups probably reflects short-term variations in growth conditions, CO₂ limitation and species-specific differences in isotope fractionation (Rau et al. 1992, Kopczynska et al. 1995, Fry 1996, Ostrom et al. 1997). Samples of ice POM and suspended POM were most likely dominated by autotrophic algal cells, but may have included a variable fraction of heterotrophic microorganisms and detritus, which could have increased the stable isotope values of these samples (Kopczynska et al. 1995).

Material collected in the sediment traps between 40 and 200 m depth was derived mainly from the suspended POM. The δ^{13} C values of the sediment were mostly within the range of δ^{13} C values of ice POM and suspended POM, which shows that autochthonous phytoplankton and ice algae both contribute to the organic material buried in the sediment of the Barents Sea MIZ (Fig. 2). The relative proportion of ice algae to the vertical particle flux may, however, be higher earlier in the season. A significant input of either macroalgal detritus or terrestrial plant material would likely have affected the δ^{13} C values of the sediment, as these were respectively enriched and depleted in ¹³C relative to phytoplankton (Fry & Sherr 1984, Fredriksen 2003). Allochthonous sources of organic material therefore seem to be of minor importance in this region. The sediment nepheloid layer was generally depleted in ¹³C compared to the settling material. One possible mechanism for this would be bacterial degradation of material reaching the seabed. Bacteria are generally enriched in ¹³C compared to their substrate (Macko & Estep 1984, Coffin et al. 1989), resulting in a depletion of ¹³C in the remnant material (Lehmann et al. 2002). In the Northeast Water Polynya, Ritzrau & Thomsen (1997) found that bacterial activity was significantly higher in the benthic boundary layer than in the water column, and that bacteria in the benthic boundary layer responded rapidly to the input of fresh phytodetritus. Organic material may therefore be modified in the nepheloid layer before it is incorporated in the sediment (Ritzrau & Thomsen 1997, Lovvorn et al. 2005). In the Barents Sea, Thomsen & Graf (1994) estimated that bacteria may comprise 15% of the POC in the benthic boundary layer.

Trophic relationships in the northern Barents Sea

Pelagic grazers receive most of their energy from suspended POM. Trophic level values of the copepods *Calanus glacialis* and *C. hyperboreus* between 2.2 and 2.6, however, suggest that their diet includes a fraction of the suspended POM with elevated δ^{15} N values. The δ^{15} N value of phytoplankton and suspended POM is influenced by a variety of factors, including nitrogen limitation (Pennock et al. 1996), inter-specific differences in isotope fractionation (Needoba et al. 2003), form of nitrogen assimilated (i.e. nitrate vs. ammonium, e.g. Ostrom et al. 1997) and grazing by protists (Hoch et al. 1996). Verity et al. (2002) estimated that microzooplankton was capable of grazing 64 to 97 % of the daily chlorophyll production in the surface layer in the Barents Sea during summer. It is possible that microzooplankton, at least during a part of the productive season, provides an important link between primary producers and mesozooplankton in the Arctic pelagic marine ecosystem (Sato et al. 2002). A contribution of heterotrophic cells to the diet of zooplankton could also account for the 2.3 to 5.2‰ steps in δ^{13} C values between suspended POM and pelagic grazers (Table 2, Kopczynska et al. 1995).

Based on their respective stable isotopic compositions, the sympagic amphipods *Onisimus glacialis* and *Apherusa glacialis* are primarily herbivorous, deriving a proportionately larger fraction of their energy directly from ice POM. In contrast, *Gammarus wilkitzkii* and *Onisimus nanseni* forage on pelagic prey in addition to grazing on POM. This interpretation agrees with dietary and lipid-biomarker studies (Scott et al. 1999, Poltermann 2001, T. Tamelander unpubl. data). Krill and pelagic amphipods had slightly higher trophic level values than the copepods. Consistently low δ^{13} C values in the zooplankton, however, suggest that all species to some degree feed directly on suspended POM.

Suspension-feeding bivalves and bryozoans at 1 station (Stn II) had δ^{13} C values comparable to pelagic copepods, indicating that fresh phytodetritus reaching the seabed was immediately exploited by the benthic community at this location (Fig. 3). Previous isotope studies in the Arctic have shown this to be the case at least in the Chukchi Sea (Dunton et al. 1989) and in the Northeast Water Polynya (Hobson et al. 1995). Suspension feeders had $\delta^{15}N$ values corresponding to TL 2.2 to 3.3. Deposit-feeding isopods, decapods, polychaetes and echinoderms occupied TL 2.3 to 3.6. Scavengers (the polychaete Nephtys sp., brittle stars Ophiura spp., the amphipod Anonyx sp.) were at TL 2.7 to 4.0, whereas predators such as sea spiders (Pycnogonida) and the sea star Crossaster papposus occupied TL 3.7 to 4.4 (Table 2). No information was found on the feeding behaviour of the sea star Poraniomorpha tumida, but the high δ^{15} N value, corresponding to TL 4.7, suggests that it is mainly a carnivore or scavenger.

The highest δ^{13} C values were observed in brittle stars. A wide range of δ^{13} C values have been reported in this group elsewhere in the Arctic. *Ophiura sarsi* and *Ophiacantha bidentata* collected in the Northeast Water Polynya had δ^{13} C values of -18.0 and -22.0%, respectively (Hobson et al. 1995). The δ^{13} C value of *O. sarsi* collected in the north-central Bering Sea (-19.0‰; Lovvorn et al. 2005) was in the same range,

whereas values from the North Water Polynya and Chukchi Sea, -11.3‰ (Hobson et al. 2002) and -11.1‰ (W. G. Ambrose unpubl. data), respectively, were similar to the level in the northern Barents Sea. Unidentified deep-sea ophiuroids in the Canada Basin (NW Canada) had δ^{13} C values between -23.5 and -20.8‰ (Iken et al. 2005). These regional variations may reflect a highly flexible omnivorous feeding strategy, and an ability to switch feeding strategy depending on available resources (Jangoux & Lawrence 1982, Graeve et al. 1997). Brittle stars occupied trophic levels 2.6 to 3.6, the same as polychaetes, crustaceans and bivalves, but their higher δ^{13} C values suggest that brittle stars may utilise a proportionally different array of food sources. Feeding on isotopically heavy ice POM reaching the seabed early in the season could increase their $\delta^{13}C$ values. However, when accounting for a trophic enrichment of ca. 1‰ between diet and consumer (Michener & Schell 1994), -14 to -10‰ still seem high, since ice POM generally had δ^{13} C values between –17 and –21‰. Input of allochthonous material with higher δ^{13} C values, such as kelp (Fredriksen 2003), was not likely an important food source, as the δ^{13} C values of the nepheloid layer indicate that phytoplankton and ice algae were the most important sources of organic material reaching the seabed. Different biochemical fractionation pathways in different organism groups, or a faster turnover of body carbon than nitrogen when shifting from an herbivorous to a carnivorous diet, could perhaps contribute to the observed pattern of comparably low δ^{15} N and high δ^{13} C values in this group.

The wider range in δ^{13} C and δ^{15} N values of benthic organisms compared to zooplankton and ice fauna is consistent with the higher number of species, feeding strategies and habitat types that are represented in the benthic samples (Fig. 3). The variation in δ^{13} C values of the benthos among stations probably reflects more persistent differences in energy input and utilisation in the benthic community rather than seasonal variations in δ^{13} C values of the POM reaching the seabed.

Demersal fishes such as Atlantic hookear sculpin Artediellus atlanticus, eelpouts Lycodes spp. and long rough dab Hippoglossoides platessoides generally had higher δ^{13} C values than herring Clupea harengus and capelin Mallotus villosus, which primarily feed on zooplankton, or Atlantic cod Gadus morhua and polar cod Boreogadus saida, which may both forage on zooplankton and selectively on benthic invertebrates (G. morhua) or ice fauna (B. saida). Demersal fishes also tended to occupy slightly higher trophic levels than pelagic species, although the δ^{15} N values did not differ significantly between these groups. Fishes in the respective groups were of similar size, and the stable isotope composition therefore presumably reflects

feeding habits rather than size effects. Fishes in our study had trophic levels in the upper range of values reported from the same or other regions of the Arctic. We calculated mean TLs of 4.0 and 3.8 for juvenile G. morhua and adult B. saida, respectively. For individuals of the same size range as in our study, Hop et al. (2002) estimated a TL of 3.2 for G. morhua in the Barents Sea near Bear Island. In other Arctic studies, B. saida's TL has ranged from around 3.2 to 3.7 (Hobson & Welch 1992, Hobson et al. 1995, 2002). The demersal fishes Triglops nybelini and Lycodes rossi were 0.2 and 0.7 TL higher, respectively, than in the Northeast Water Polynya food web (Hobson et al. 1995). Since most values stem from the summer months (Hobson & Welch 1992, Hobson et al. 1995, Hop et al. 2002), these variations are probably diet related, reflecting slightly different trophic positions of fishes across the Arctic. The use of a smaller fractionation factor for the Barents Sea food chain in this study (3.4%) compared to the others (3.8‰) would also slightly inflate TL values.

Spatial variability of the pelagic-benthic coupling

The higher $\delta^{13}C$ values of the suspended POM and settling material at Stn II compared to the other stations likely reflected different successional stages of the phytoplankton bloom (Kopczynska et al. 1995, Ostrom et al. 1997). Stn II was characterised by nutrient depletion in the upper 10 m and a high chlorophyll biomass below 20 m, dominated by diatoms, indicating bloom conditions. Stn III exhibited a similar pattern, but low primary production rates suggested that the bloom was declining. At Stn I, the bloom was likely in a late stage, indicated by nutrient depletion in the surface layer and a deep chlorophyll maximum, with a lower abundance of diatoms, whereas at Stn IV, the bloom was in an earlier stage, since nutrients still remained in the surface layer (H. Hodal, S. Kristiansen, M. Reigstad, Norwegian College of Fishery Science, University of Tromsø, and T. Ratkova, Shirshov Institute of Oceanology, Moscow, pers. comms.). The δ^{13} C value of phytoplankton has been shown to increase progressively with decreasing concentrations of CO₂ during a bloom (Rau et al. 1992, Ostrom et al. 1997). In the Southern Ocean, Kopczynska et al. (1995) found that pennate diatoms and heterotrophic dinoflagellates were associated with increased $\delta^{13}C$ values of the suspended POM, whereas centric diatoms and flagellates, such as Phaeocystis sp., and autotrophic dinoflagellates caused depleted $\delta^{13}C$ values. The elevated $\delta^{13}C$ value of suspended POM at Stn II (-23.0%) compared with the other stations (-24.7 to -26.5%) was, therefore, likely caused by the dense diatom bloom at this station.

The similarity in δ^{13} C values for the suspension-feeding benthos and zooplankton at Stn II suggests that these groups derive their energy from a common food source (Fig. 3), and, hence, there is strong pelagic-benthic coupling in this region (Hobson et al. 1995). The bloom conditions observed in the surface layer at Stn II and the δ^{13} C values of the benthic community (Table 2) suggest diatom-based primary production to be the most likely source. At other stations the differences in δ^{13} C values between the pelagic and benthic food webs were more pronounced, perhaps indicating a weaker coupling between surface primary production and benthic animal communities. It is uncertain if these among-station differences represent a persistent spatial heterogeneity of pelagic-benthic coupling in this region, or if they reflect the stage of the phytoplankton bloom and timing of sedimentation at the individual stations.

The elevated δ^{13} C values of the benthic fauna at Stns I, III and IV are not easily explained (Fig. 3). Trophic fractionation of stable carbon isotopes increases the δ^{13} C of a consumer by 0.5 to 1% relative to the diet (Michener & Schell 1994), but the benthic organisms were enriched by >7% relative to suspended POM. The following factors could potentially have contributed to the elevated δ^{13} C values in benthic organisms compared to the zooplankton in the AtW region.

The benthic communities in regions dominated by AtW may receive a larger proportion of their energy from degraded material than communities in the ArW region (McConnaughey & McRoy 1979, Hobson et al. 1995). Reworking of organic material by the benthic community and feeding on detritus with associated microfauna could increase the overall δ^{13} C value of the benthic food web (Macko & Estep 1984, Coffin et al. 1989, Lovvorn et al. 2005).

It is possible that ambient temperature may affect stable isotope fractionations between diet and consumer tissues. Power et al. (2003) showed that freshwater zooplankton raised at 26.5°C were enriched in δ^{13} C by 3.1% relative to their diet, whereas animals raised at 12.5°C were enriched by only 1.7%. The effect on δ^{15} N values was opposite. Applying these results to our data should be performed with caution, however, as they stem from experiments with different organisms and environments and were performed at higher temperatures than in our study. Bottom water temperatures only ranged from 0.06 to 1.9 among stations and it is unlikely that this would have caused a higher δ^{13} C fractionation at the warmer stations.

The higher δ^{13} C values of the suspension- and surface-deposit-feeding benthos compared to zooplankton indicate that isotopically light POM exported from the surface during non-diatom bloom conditions does not contribute much to the benthic energy intake. The benthic fauna therefore seems to be dependent on the episodic primary production events giving rise to pulses of vertical export of ungrazed material, conditions that are commonly associated with diatom blooms (Wassmann et al. 1996, Tamelander & Heiskanen 2004). Early season sedimentation of ice algae, mostly represented by diatoms, could provide an additional input of energy to the benthic community.

Our results show that tight linkages between surface primary production and benthic animal communities in the northern Barents Sea are mainly governed by diatom blooms during which rapid sedimentation of ungrazed POM occurs. The distribution of different water masses and dynamic bloom conditions seems to give rise to spatial heterogeneities in the pelagicbenthic coupling in this region, as revealed by the different $\delta^{13}C$ values of benthic organisms relative to zooplankton at stations dominated by AtW and ArW, respectively. Previous isotope studies in the Arctic have assessed the average food web structure of special types of environments such as polynyas (Hobson et al. 1995, 2002) or examined geographical trends in isotope signatures over wide geographical areas (Dunton et al. 1989, Saupe et al. 1989, France et al. 1998). Our findings show that 'average' food web models based on isotope signatures gathered from a large geographical area may easily oversee important aspects of the structure and function of the food web. In a complex environment such as the MIZ, spatial variability may likewise induce errors in extrapolations from singlesite observations to a regional scale.

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