

Tumor-secreted miR-214 induces regulatory T cells: a major link between immune evasion and tumor growth

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An increased population of CD4⁺CD25^{high}Foxp3⁺ regulatory T cells (Tregs) in the tumor-associated microenvironment plays an important role in cancer immune evasion. However, the underlying mechanism remains unclear. Here we observed an increased secretion of miR-214 in various types of human cancers and mouse tumor models. Tumor-secreted miR-214 was sufficiently delivered into recipient T cells by microvesicles (MVs). In targeted mouse peripheral CD4⁺ T cells, tumor-derived miR-214 efficiently downregulated phosphatase and tensin homolog (PTEN) and promoted Treg expansion. The miR-214-induced Tregs secreted higher levels of IL-10 and promoted tumor growth in nude mice. Furthermore, *in vivo* studies indicated that Treg expansion mediated by cancer cell-secreted miR-214 resulted in enhanced immune suppression and tumor implantation/growth in mice. The MV delivery of anti-miR-214 antisense oligonucleotides (ASOs) into mice implanted with tumors blocked Treg expansion and tumor growth. Our study reveals a novel mechanism through which cancer cell actively manipulates immune response via promoting Treg expansion.

Keywords: secreted microRNA; regulatory T cell; PTEN; microvesicle; immune evasion; tumor

Cell Research (2014) 24:1164–1180. doi:10.1038/cr.2014.121; published online 16 September 2014

Introduction

Tumors develop multiple potent and overlapping mechanisms to mediate immune evasion [1, 2]. One such mechanism involves an increase in the population of CD4⁺CD25^{high}Foxp3⁺ regulatory T cells (Tregs), a subset

of CD4⁺ T cells that plays an important role in maintaining self-tolerance and modulating immune responses [3–5]. Tregs are increased during tumorigenesis [6, 7]. Tumor cells can also efficiently recruit Tregs to evade the host immune response [2]. Due to the importance of Tregs in tumor immune evasion, the depletion of Tregs is a potential therapy for cancer [8, 9]. Although multiple factors, such as IL-2 [10] and TGF-β [11, 12], are responsible for the expansion of Tregs, the precise mechanism by which tumor cells induce Tregs remains poorly understood.

The discovery of microRNAs (miRNAs), a class of noncoding RNAs with lengths of approximately 22 nucleotides, has revealed a new layer of gene regulation in

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Received 4 April 2014; revised 18 July 2014; accepted 5 August 2014; published online 16 September 2014

biological processes. Because miRNAs are abnormally expressed in tumors, an altered miRNA expression pattern is generally regarded as a major feature of tumorigenesis. Depending on the specific miRNAs and the targets in certain tumor types, miRNAs can function as either oncogenes or tumor suppressors. For example, miR-214, which is an important oncogenic miRNA, can promote tumorigenesis and induce cell survival and cisplatin resistance in human ovarian cancer by targeting the 3'-untranslated region (UTR) of phosphatase and tensin homolog (PTEN), which leads to the downregulation of the PTEN protein and upregulation of the Akt pathway [13]. The differential expression of miRNAs has also been reported in T cell maturation and activation and the modulation of immune responses [14]. For example, miR-214, miR-21, miR-31, miR-181a, and miR-155 have been reported to play pivotal roles in T cell function. Rouas *et al.* [15] also identified the miRNA signature of natural human Tregs and the regulatory roles of miR-31 and miR-21 in Foxp3 expression. The targeting of Foxp3 by miR-155 during the development of Tregs has also been reported by several groups [16, 17]. Despite these findings, the role of miRNAs in Treg function in tumorigenesis has not been fully elucidated.

Recently, we and others have demonstrated that cells can selectively package miRNAs into microvesicles (MVs), particularly exosomes, that are secreted under various pathophysiologic stimuli *in vitro* and *in vivo*. Through MV-cell interactions and, most likely, the endocytic pathway, MVs can efficiently deliver miRNAs into recipient cells, where they block the translation of their target genes and regulate recipient cell function [18]. Thus, secreted miRNAs can serve as a novel class of signaling molecules in mediating cell-cell communication. The identification of MV-mediated functional miRNA transport between non-adjacent cells significantly extends our understanding of the role of miRNAs. Because fast-growing tumor cells have been shown to highly express certain oncogenic miRNAs, such as miR-214 and miR-21 [19], and can generate and release high levels of miRNA-containing MVs [20], it is possible that tumor cells can actively manipulate the antitumor activities of immune cells by delivering tumor-specific miRNAs to target immune cells via an MV-based pathway.

In this study, we examined the modulation of CD4⁺C-D25^{high}Foxp3⁺ Tregs by tumor cell-secreted miRNAs to determine whether tumor cells can actively secrete particular miRNAs that induce suppression activities in immune cells. We identified miR-214 as a major tumor-specific miRNA secreted by various types of cancers and demonstrated for the first time that tumor cell-secreted miR-214 can promote Treg expansion by targeting PTEN

in CD4⁺ T cells, leading to host immune suppression and rapid tumor growth.

Results

miR-214 levels are elevated in tissues and plasma from cancer patients and tumor xenograft mice

To identify the secreted miRNAs that are able to regulate Tregs, the expression levels of oncogenic miRNAs, including miR-18a, miR-19a, miR-200a, miR-205, miR-21, miR-210, miR-214, and miR-92a [19], were assessed in tissues from patients with breast cancer, hepatocellular carcinoma, non-small-cell lung cancer, or pancreatic cancer. Although most of the oncogenic miRNAs had elevated expression levels in the 4 types of cancer, only miR-214 and miR-21 levels were increased in all 4 types of cancer (Figure 1A). The levels of the oncogenic miRNAs that were secreted into the plasma from the patients were then measured. Most of the miRNAs were increased in one or two types of cancer, but only miR-214 was significantly increased in all four types of cancer (fold change > 2 and $P < 0.05$) (Figure 1B). Further analysis revealed that the plasma levels of miR-214 in the tumor-bearing patients were markedly enriched in MVs (Figure 1C), by which miRNAs can be delivered into recipient cells. Secreted miR-214 levels were also investigated in mouse models. Mouse sarcoma S-180 cells and Lewis lung carcinoma (LLC) cells were used to establish a tumor xenograft mouse model. miR-214 expression levels were also increased in these two cell lines (Figure 1D). The elevation of circulating miR-214 and the enrichment of miR-214 in MVs was also observed in the two tumor xenograft mouse models (Figure 1E-1H). These results suggest that increased miR-214 secretion may occur in cancer cell biogenesis.

LLC cell-secreted miR-214 promotes Treg expansion

To determine whether secreted miR-214 was sufficiently delivered into the recipient Tregs, LLC-derived MVs containing a high level of miR-214 were incubated with primary CD4⁺ T cells in culture (Figure 2A). miR-214 levels were markedly increased in the recipient CD4⁺ T cells and peaked 24 h post MV treatment (12-fold induction) (Figure 2B), while no alterations in the levels of pre-miR-214 were observed (Figure 2C), suggesting that the elevation of miR-214 level in the CD4⁺ T cells was likely due to the MV delivery of exogenous miR-214. We also measured the levels of miR-199a, pre-miR-199a-2, and Dnm3os post MV treatment. miR-214 and miR-199a-2 are located within the same non-coding RNA, DN3os, and a change in endogenous miR-214 should be accompanied by a change in DN3os and

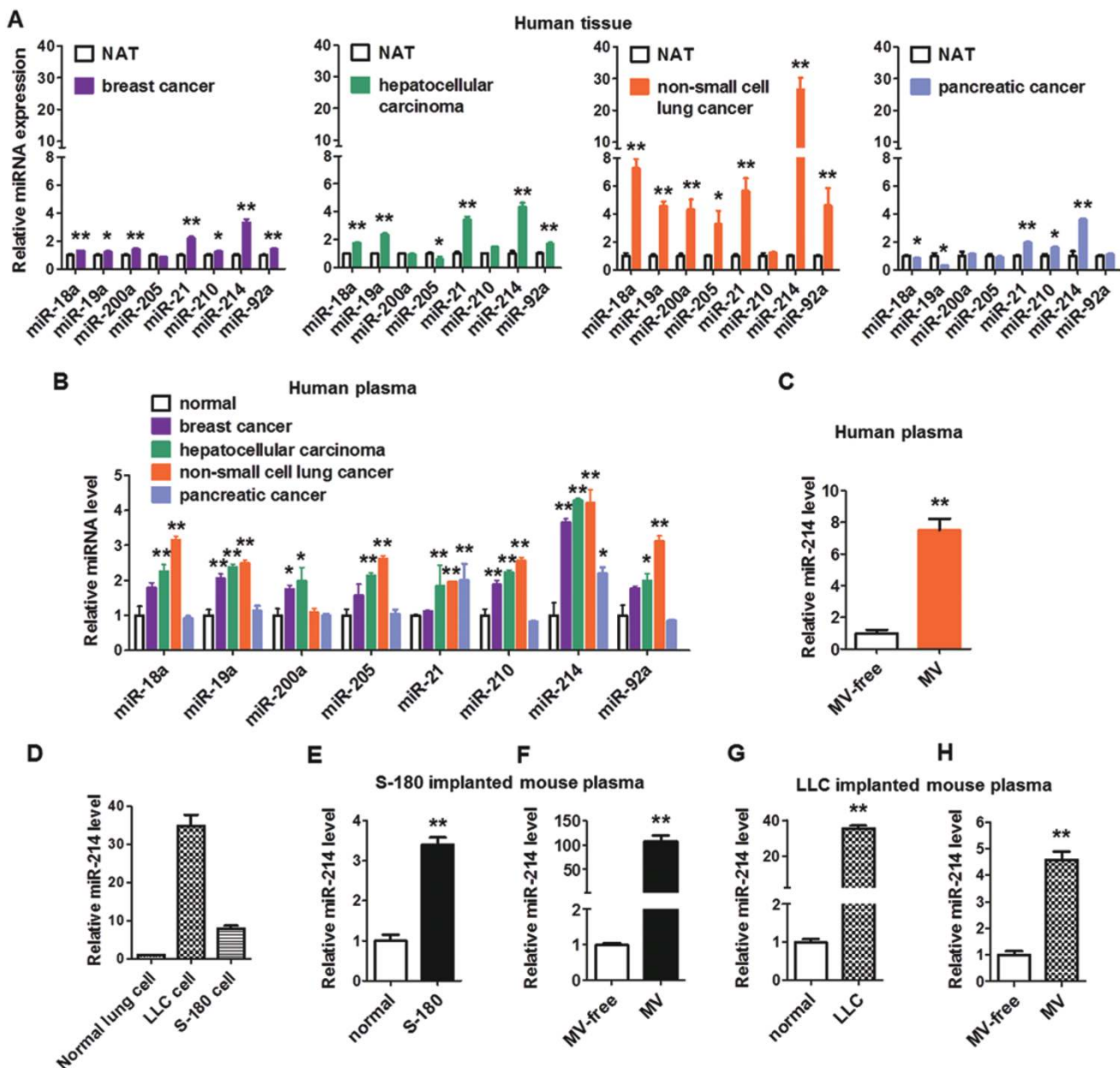


Figure 1 Increased miR-214 levels in cancer patients and mice implanted with tumors. **(A, B)** Elevated tumor-associated miRNAs in tissue and plasma samples from breast cancer, hepatocellular carcinoma, non-small-cell lung cancer, and pancreatic cancer patients. The miRNA expression levels were determined by qRT-PCR. The results are presented as the mean \pm SEM (tissue, $n = 4$; plasma, $n = 10$). NAT, normal adjacent tissues. **(C, F, H)** Comparison of the levels of miR-214 in the MV and MV-free fractions of plasma from the non-small-cell lung cancer patients and S-180- and LLC-implanted C57BL/6J mice. The expression levels of the miRNAs in the MV-free plasma were arbitrarily set to 1. **(D)** Comparison of the relative expression levels of miR-214 in normal lung cells, LLC cells, and S-180 cells. **(E, G)** Relative plasma miR-214 levels in C57BL/6J mice with or without the implantation of S-180 and LLC cells. The results are presented as the mean \pm SEM ($n = 10$). * $P < 0.05$; ** $P < 0.01$.

miR-199a-2 [21-23]. Consistent with previous studies [24, 25], we found that mature miR-199a and pre-miR-199a-2 gradually increased in cultured CD4⁺ T cells

(Figure 2D and 2E), suggesting that endogenous miR-214 levels increased in differentiated Tregs or activated T cells. However, the levels of mature miR-199a, pre-

miR-199a-2, and Dnm3os were the same in untreated CD4⁺ T cells and CD4⁺ T cells treated with MVs (Figure 2D-2F), suggesting that endogenous miR-214 level was not boosted by the incubation with MVs. Taken together, these results indicate that exogenous miR-214 was sufficiently delivered into the recipient T cells and that the increase in miR-214 levels in CD4⁺ T cells was not due

to the induction of endogenous miR-214 by MVs.

Next, the effect of LLC-derived MVs on recipient T cells was assessed. As shown in Figure 2G, LLC-derived MVs significantly increased the percentage of CD4⁺CD25^{high}Foxp3⁺ Tregs. To confirm that the expansion of CD4⁺CD25^{high}Foxp3⁺ Tregs was mainly due to exogenous miR-214 in MVs, miR-214-deficient LLC-derived

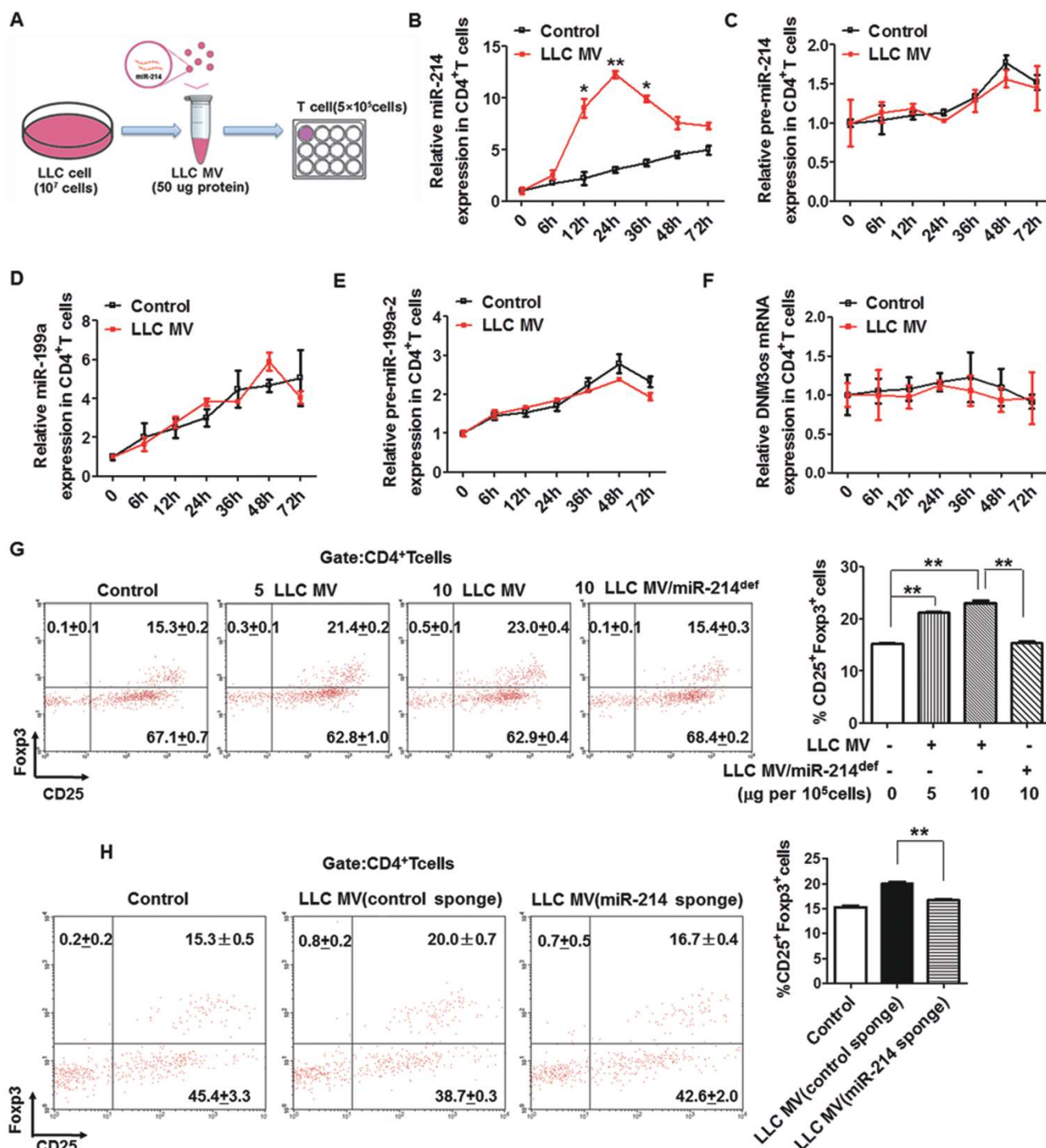


Figure 2 Effects of LLC MVs on miR-214 induction and Treg expansion in recipient CD4⁺ T cells. **(A)** Flow chart of the experimental design. The LLC MVs were isolated from LLC cells and were subjected to qRT-PCR to measure miR-214 level and the BCA method to quantify the total protein. According to our calculation, the amount of total protein in the LLC MVs isolated from 10⁷ LLC cells was 50 μg, and the level of miR-214 in these MVs was 3.125 fmol. Thus, miR-214 was present in the LLC MVs at a concentration of approximately 0.06 fmol/μg total protein. The LLC MVs were then incubated with 5 × 10⁵ primary CD4⁺ T cells. **(B, C)** The relative expression levels of miR-214 and pre-miR-214 in isolated CD4⁺ T cells incubated with or without the LLC MVs (10 μg protein/10⁵ CD4⁺ T cells) for 0, 6, 12, 24, 36, 48, and 72 h. The miRNA expression levels were determined by qRT-PCR and are presented as the mean ± SEM of three independent experiments. **(D-F)** The relative expression levels of mature miR-199a, pre-miR-199a-2, and Dnm3os in isolated CD4⁺ T cells incubated with or without LLC MVs (10 μg protein/10⁵ CD4⁺ T cells) for 0, 6, 12, 24, 36, 48, and 72 h. The expression levels were determined by qRT-PCR and are presented as the mean ± SEM of three independent experiments. **(G)** The induction of CD4⁺CD25^{high}Foxp3⁺ Tregs in isolated CD4⁺ T cells after treatment with different concentrations of LLC MVs or miR-214-deficient LLC MVs (LLC MV/miR-214^{def}) for 72 h. The left panels show a representative result of five experiments. The results from five independent experiments are plotted presented as the mean ± SEM (right panel). **(H)** The induction of CD4⁺CD25^{high}Foxp3⁺ Tregs in isolated CD4⁺ T cells after treatment with LLC MVs (control sponge) or miR-214-deficient LLC MVs (miR-214 sponge) for 72 h. The left panels show a representative result of five experiments. The results from five independent experiments are plotted and presented as the mean ± SEM (right panel). **P* < 0.05; ***P* < 0.01.

MVs were prepared by knocking down miR-214 using antisense oligonucleotides (ASOs). The knockdown efficiency in the LLC cells is shown in Supplementary information, Figure S1A. Knockdown of miR-214 in the LLC cells resulted in the production of MVs containing low levels of miR-214 (Supplementary information, Figure 1B). miR-214-deficient MVs did not affect CD4⁺CD25^{high}Foxp3⁺ Treg expansion (Figure 2G). The effects of miR-214 loss-of-function in MVs were further tested using “miRNA sponge” technology to produce miR-214-deficient LLC-derived MVs (Supplementary information, Figure 2A) [26]. The miR-214 sponge efficiently blocked the activity of miR-214 (Supplementary information, Figure S2B). Importantly, the miR-214 sponge was not transported into MVs (Supplementary information, Figure S2C) and significantly decreased miR-214 levels in MVs (Supplementary information, Figure S2D). As shown in Figure 2H, miR-214-deficient MVs produced by the miR-214 sponge failed to affect Treg expansion. The iTRAQ quantitative proteomic technique and mRNA/miRNA microarray analysis of MVs with or without miR-214 were also performed to further confirm that the effect of miR-214-deficient MV on Tregs was due to the absence of miR-214 rather than non-specific changes in MVs. Most of the proteins in the miR-214-deficient MVs displayed little, if any, alterations (433 proteins were altered by 0.5-1-fold; 214 proteins were altered by 1-1.5-fold, and 7 proteins were altered by 1.5-2-fold) (Supplementary information, Figure S3A). Only 7 proteins displayed a greater than two-fold reduction (Supplementary information, Figure S3A), but to our knowledge, these proteins do not affect Tregs (Supplementary information, Table S1). Likewise, the transfection of anti-miR-214 did not substantially alter

mRNA or miRNA levels in the LLC MVs, with the exception of miR-214 (Supplementary information, Figure S3B and S3C). Based on the negligible changes in protein, mRNA, and miRNA levels in the miR-214-deficient MVs, it is unlikely that the recipient Tregs would be altered. Taken together, these results indicate that LLC-secreted miR-214 in MVs was delivered into the recipient CD4⁺ T cells sufficiently and promoted Treg expansion.

Decreased PTEN by LLC-secreted miR-214 results in Treg expansion

miR-214 targets PTEN and downregulates the PTEN-mediated signaling cascade [13, 25, 27]. PTEN is a negative modulator of CD4⁺CD25^{high}Foxp3⁺ Treg homeostasis *in vivo* and expansion *ex vivo* [28]. Therefore, we hypothesized that LLC-secreted miR-214 in MVs promotes Treg expansion by targeting PTEN in CD4⁺ T cells. To assess whether exogenous miR-214 in MVs was able to directly bind to the PTEN 3' UTR, luciferase reporter assays were performed (Supplementary information, Figure S4A and S4B). The activity of luciferase reporter containing the PTEN 3' UTR was significantly repressed upon treatment with LLC MV but not LLC MV/miR-214^{def}, whereas the mutant luciferase reporter activity was unaffected upon treatment with either LLC MV or LLC MV/miR-214^{def} (Supplementary information, Figure S4C). Next, the effects of LLC-secreted miR-214 on PTEN levels in the recipient CD4⁺ T cells were assessed. Treatment with LLC MVs reduced PTEN protein levels in the CD4⁺ T cells in a dose-dependent manner (Figure 3A), whereas the levels of PTEN mRNA in the CD4⁺ T cells were unchanged (Supplementary information, Figure S5A). Interestingly, although miR-214 levels were markedly increased in the recipient CD4⁺ T

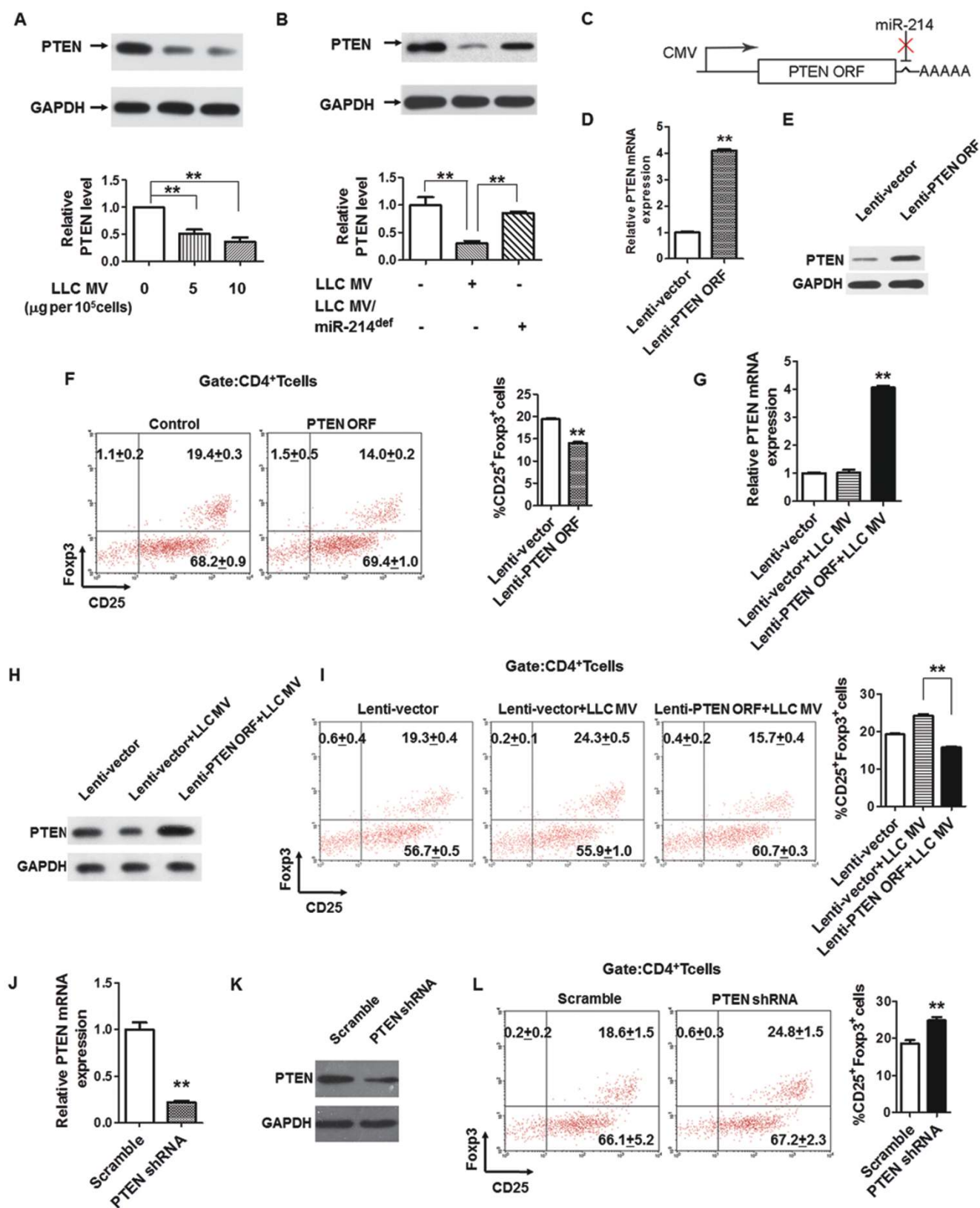


Figure 3 Effects of LLC-secreted miR-214 on PTEN suppression and Treg expansion in recipient CD4⁺ T cells. **(A, B)** Western blot analysis of PTEN protein levels in CD4⁺ T cells following incubation with different concentrations of LLC MVs (0, 5, and 10 μg protein/10⁵ cells) **(A)** and miR-214-deficient LLC MVs (10 μg protein/10⁵ cells) **(B)** for 72 h. **(C)** A diagram of the recombinant lentivirus carrying the miR-214-resistant version of PTEN (PTEN ORF). **(D, E)** qRT-PCR and western blot analysis of PTEN mRNA and protein levels in CD4⁺ T cells following infection with a control or PTEN ORF-overexpressing lentivirus. **(F)** Induction of CD4⁺CD25^{high}Foxp3⁺ Tregs in isolated CD4⁺ T cells after infection with a control or PTEN ORF-overexpressing lentivirus. The left panels show a representative result of five experiments. The results from five independent experiments are plotted and presented as the mean ± SEM (right panel). **(G, H)** qRT-PCR and western blot analysis of PTEN mRNA and protein levels in CD4⁺ T cells following treatment with the control lentivirus, control lentivirus plus LLC MVs, or PTEN ORF-overexpressing lentivirus plus LLC MVs. **(I)** Induction of CD4⁺CD25^{high}Foxp3⁺ Tregs in isolated CD4⁺ T cells after treatment with the control lentivirus, control lentivirus plus LLC MVs, or PTEN ORF-overexpressing lentivirus plus LLC MVs. The left panels show a representative result of five experiments. The results from five independent experiments are presented as the mean ± SEM (right panel). **(J, K)** qRT-PCR and western blot analysis of PTEN mRNA and protein levels in CD4⁺ T cells following infection with a lentiviral control or lentiviral shRNA targeting PTEN. **(L)** The induction of CD4⁺CD25^{high}Foxp3⁺ Tregs in isolated CD4⁺ T cells after infection with a lentiviral control or lentiviral shRNA targeting PTEN. The left panels show a representative result of five experiments. The results from five independent experiments are plotted and presented as the mean ± SEM (right panel). **P* < 0.05; ***P* < 0.01.

cells and peaked 24 h post MV treatment (Figure 2B), the PTEN protein was consistently downregulated from 24 to 72 h post treatment (Supplementary information, Figure S5B), whereas no alterations in PTEN mRNA level were observed (Supplementary information, Figure S5C). These results further demonstrate that miR-214 affects PTEN at the protein rather than the mRNA level. By contrast, neither LLC MV/miR-214^{def} (Figure 3B) nor “sponge”-produced miR-214-deficient MVs (Supplementary information, Figure S6A and S6B) affected PTEN protein levels. These data suggest that LLC-secreted miR-214 in MVs sufficiently reduces PTEN protein levels in recipient CD4⁺ T cells.

To determine whether the effect of LLC-secreted miR-214 on Treg expansion is dependent on PTEN silencing, T cells were infected with a recombinant lentivirus carrying the miR-214-resistant form of PTEN (PTEN ORF) (Figure 3C). Efficient overexpression of the PTEN ORF in CD4⁺ T cells is shown in Figure 3D and 3E. The overexpression of PTEN potently suppressed Treg expansion (Figure 3F). Although the LLC MVs downregulated PTEN protein levels in CD4⁺ T cells, overexpression of PTEN ORF rescued the downregulation of PTEN protein induced by the miR-214-loaded MVs (Figure 3G and 3H). Consistent with these results, the T cells treated with the LLC MVs had a significantly higher proportion of Tregs compared with the control cells, but the cells treated with both the LLC MVs and PTEN ORF-overexpressing lentivirus exhibited significantly lower levels of Tregs (Figure 3I), suggesting that the miR-214-resistant PTEN compromised miR-214-induced Treg conversion. Moreover, a lentiviral shRNA was used to directly knock down PTEN expression in CD4⁺ T cells. Efficient knock-down of PTEN in CD4⁺ T cells is shown in Figure 3J and 3K. As expected, the inhibition of PTEN expression

potently promoted Treg expansion (Figure 3L).

Finally, to mimic the effects of miRNA secretion on recipient CD4⁺ T cell function *in vivo*, a transwell system was used in which secreted miR-214 crossed an insert to regulate PTEN expression and Treg expansion (Supplementary information, Figure S7A). CD4⁺ T cells co-cultured with the miR-214-deficient LLC cells had decreased levels of miR-214 and increased levels of PTEN protein compared with CD4⁺ T cells co-cultured with LLC cells (Supplementary information, Figure S7B-S7D). Consistent with these results, there was a significant decrease in the percentage of Tregs in the CD4⁺ T cells co-cultured with the miR-214-deficient LLC cells compared to the CD4⁺ T cells co-cultured with LLC cells (Supplementary information, Figure S7E). Taken together, LLC-secreted miR-214 targets PTEN in CD4⁺ T cells, leading to Treg expansion.

Exogenous miR-214 induces IL-10 secretion in CD4⁺ T cells and promotes xenograft tumor growth in nude mice

To monitor the functions of the induced Tregs, the levels of several cytokines [2, 5] associated with Tregs were measured in the culture supernatants of CD4⁺ T cells treated with or without the LLC MVs. Treatment with LLC MVs significantly increased the levels of interleukin-10 (IL-10) but not TGF-β or interleukin-12 (IL-12), and depletion of miR-214 from the LLC MVs significantly blocked IL-10 production (Figure 4A-4C). These results suggest that IL-10 might be involved in the suppression of the immune system by Tregs induced by tumor-secreted miR-214.

The functional activity of the Tregs induced by tumor-secreted miR-214 was further tested in nude mice. As depicted in Figure 4D, murine CD4⁺ T cells were pretreated with or without LLC MVs and then injected

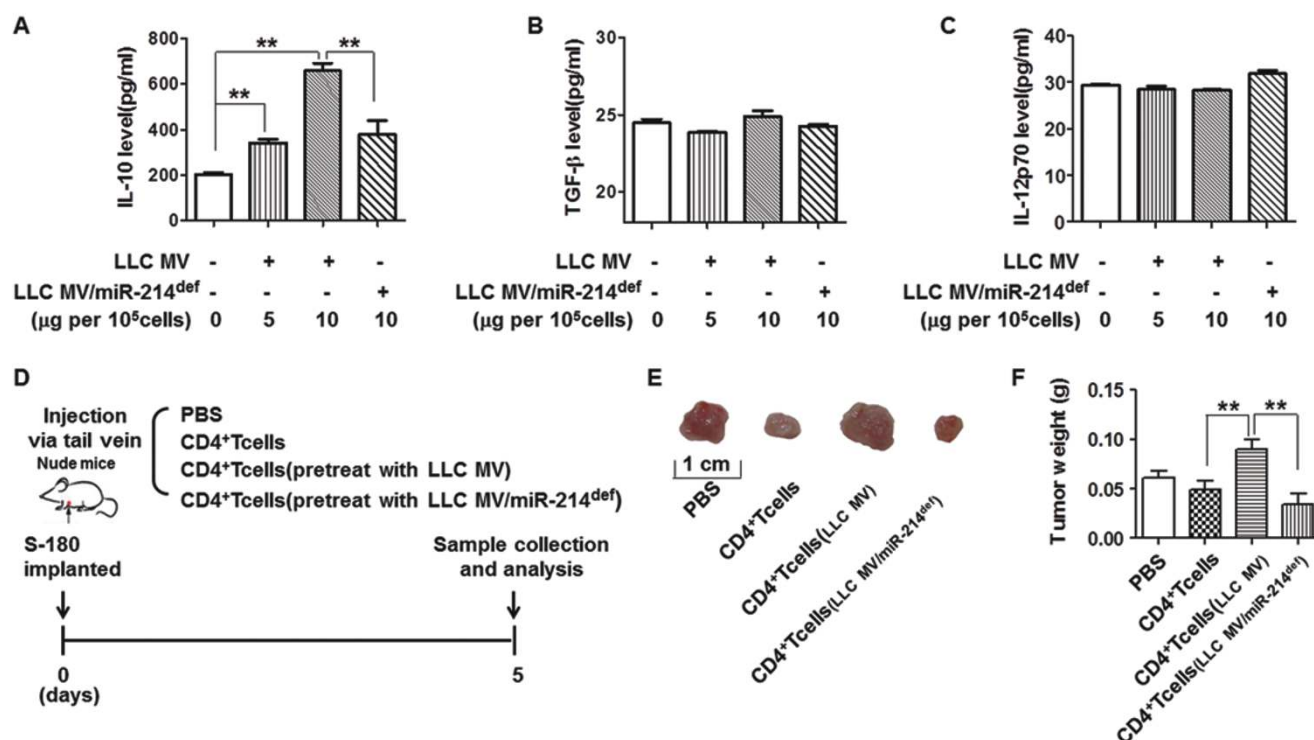


Figure 4 Effects of LLC-secreted miR-214 on cytokine secretion in CD4⁺ T cells and tumor growth in nude mice. **(A-C)** ELISA analysis of IL-10, TGF-β, and IL-12 levels in the supernatants of CD4⁺ T cells incubated with different concentrations of the LLC MVs or miR-214-deficient LLC MVs for 72 h. **(D)** Flow chart depicting the experimental design. Isolated murine CD4⁺ T cells, CD4⁺ T cells pretreated with LLC MVs (10 μg protein/10⁵ CD4⁺ T cells), CD4⁺ T cells pretreated with LLC MVs depleted of miR-214 (10 μg protein/10⁵ CD4⁺ T cells), or a saline control were administered to nude mice via tail vein injection (5 × 10⁶ cells), and sarcoma S-180 cells (1 × 10⁶ cells per mouse) were implanted. **(E)** Tumor size. **(F)** Tumor burden. The data are presented as the mean ± SEM of three independent experiments. In each experiment, each group consisted of 6-8 mice. *P < 0.05; **P < 0.01.

into nude mice via the tail vein. The nude mice were also implanted with S-180 cells. The Tregs induced by tumor-secreted miR-214 strongly promoted tumor growth in the nude mice (Figure 4E and 4F), suggesting that miR-214-induced Tregs can effectively suppress the immune responses of tumor-attacking cytotoxic cells, such as natural killer (NK) cells and macrophages.

Cancer cell-secreted miR-214 is sufficiently delivered into CD4⁺ T cells in vivo

Two approaches were performed to directly monitor the transport of miR-214 into peripheral CD4⁺ T cells via tumor-derived MVs. First, LLC MVs were fluorescently labeled and then intravenously injected into mice. A significant amount of fluorescent MVs were associated with the peripheral CD4⁺ T cells 1 h after injection (Figure 5A). In the second approach, the miR-214^{mut} (mimic of miR-214) was tracked. miR-214^{mut} could be distinguished from miR-214 by using specific probes and

primers in qRT-PCR assay. miR-214^{mut} was transfected into LLC cells, and the miR-214^{mut}-containing LLC MVs were collected and intravenously injected into C57BL/6J mice. The peripheral CD4⁺ T cells isolated from these mice contained a significant amount of miR-214^{mut} (Figure 5B and 5C), and miR-214^{mut} levels were markedly increased in the plasma and peaked 30 min post injection (Figure 5D). The miR-214^{mut}-transfected tumor cells were also directly injected into the mice. When the mice were directly implanted with 10⁶ miR-214^{mut}-transfected tumor cells, miR-214^{mut} was readily detected in the isolated CD4⁺ T cells (Figure 5E). Taken together, these results demonstrate that cancer cell-secreted miR-214 may be delivered into peripheral CD4⁺ T cells via MVs.

Cancer cell-secreted miR-214 increases the number of Tregs and promotes tumor growth in vivo

We next determined the effects of secreted miR-214 on Tregs and tumor growth *in vivo*. Mice were given

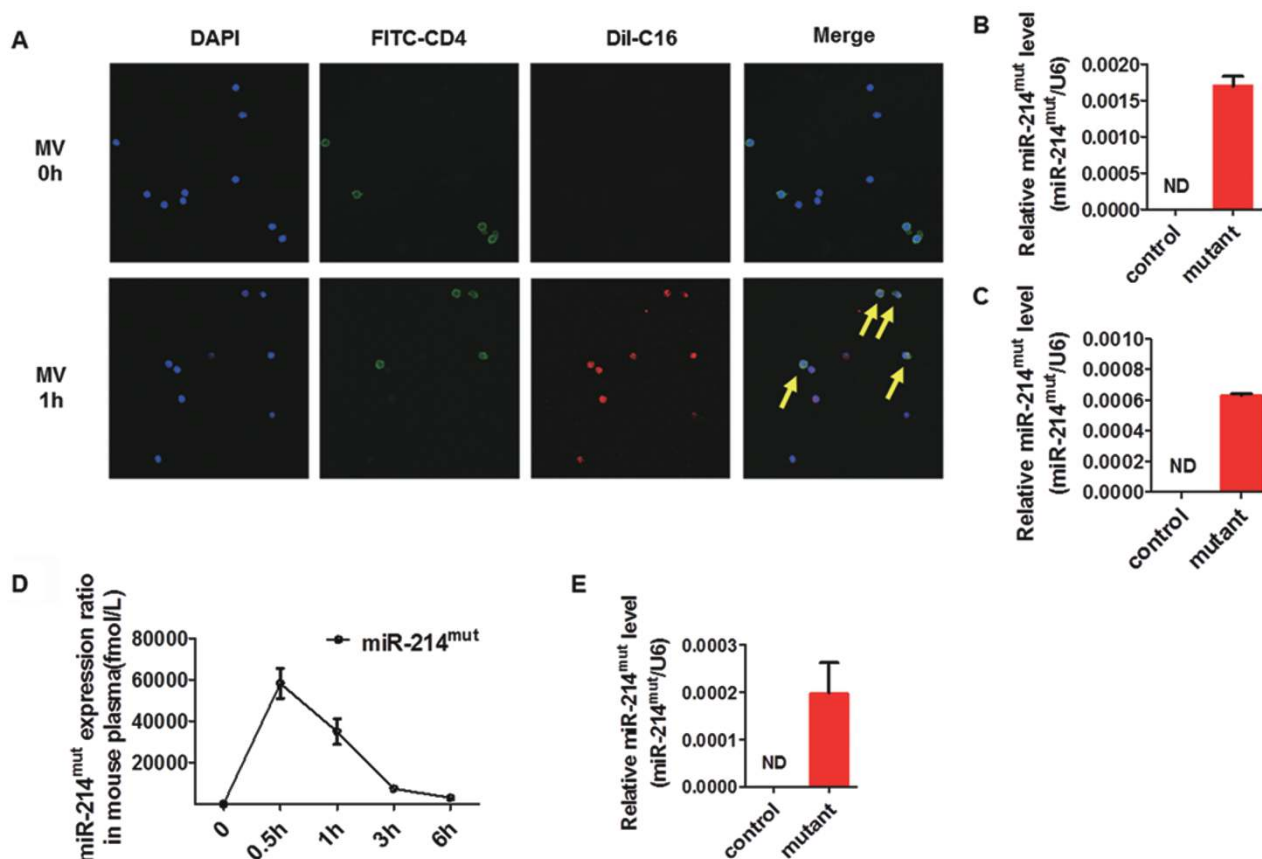


Figure 5 LLC-derived MV sufficiently delivers exogenous miR-214 into the recipient CD4⁺ T cells *in vivo*. **(A)** Confocal microscopy image of the internalization of fluorescently labeled MVs in peripheral CD4⁺ T cells. A total of 100 μg of the DiIC18(3)-labeled LLC MVs was injected into each C57BL/6J mouse via the tail vein. After 1 h, the peripheral lymphocytes were incubated with FITC-conjugated anti-CD4⁺ antibody and DAPI, and the cells were washed, fixed, and observed by confocal microscopy. **(B, C)** Either 600 or 0.6 pmol miR-214^{mut} (ACAGCAGGAACAGUCAGCCAAU) was transfected into 10⁷ LLC cells, and the miR-214^{mut}-containing LLC MVs were collected and injected into C57BL/6J mice via the tail vein. After 1 h, the peripheral CD4⁺ T cells were isolated and assayed for miR-214^{mut} using TaqMan probes specifically designed for miR-214^{mut}. **(D)** A total of 600 pmol of miR-214^{mut} was transfected into 10⁷ LLC cells, and the miR-214^{mut}-containing LLC MVs were injected into the C57BL/6J mice. The miR-214^{mut} levels in the plasma of the C57BL/6J mice were assayed at 0, 0.5, 1, 3, and 6 h post injection. **(E)** A total of 0.6 pmol miR-214^{mut} was transfected into 10⁷ LLC cells, and the miR-214^{mut}-transfected tumor cells were implanted into the C57BL/6J mice (10⁶ per mouse). After 5 days, the peripheral CD4⁺ T cells were isolated and assayed for miR-214^{mut}. The results are presented as the mean ± SEM of three independent experiments. **P* < 0.05; ***P* < 0.01.

seven injections of LLC MVs over a span of 2 weeks (Supplementary information, Figure S8A). Consistent with findings in cancer patients and tumor-implanted mice (Figure 1), miR-214 levels in the plasma (Supplementary information, Figure S8B) and peripheral CD4⁺ T cells (Supplementary information, Figure S8C) of LLC MV-injected mice were significantly higher compared with those in saline-injected mice. However, the levels of pre-miR-214 (Supplementary information, Figure S8D), as well as miR-199a, pre-miR-199a-2, and Dnm3os (Supplementary information, Figure S8E-S8G), in the peripheral CD4⁺ T cells did not change. These results con-

firmed the transfer of exogenous miR-214 into the CD4⁺ T cells. Following LLC MV treatment, the percentage of CD4⁺CD25^{high}Foxp3⁺ Tregs in the peripheral and splenic CD4⁺ T cells increased from 3.2% and 7.8% to 5.5% and 12.7%, respectively (Supplementary information, Figure S8H and S8I). C57BL/6J mice were also directly implanted with S-180 sarcoma cells (Supplementary information, Figure S9A). As expected, the percentage of CD4⁺CD25^{high}Foxp3⁺ Tregs in the total CD4⁺ T cell population isolated from the peripheral blood and spleen was significantly higher in the tumor-implanted mice than the control mice (Supplementary information, Figure S9B).

miR-214 levels were also increased in peripheral CD4⁺ T cells (Supplementary information, Figure S9C), whereas there was no change in pre-miR-214 levels (Supplementary information, Figure S9D), suggesting that the elevated miR-214 levels in the CD4⁺ T cells were likely due to cancer cell secretion. Consistent with the results in tumor-implanted mice, the percentage of CD4⁺CD25^{high}Foxp3⁺ Tregs was higher in peripheral blood from lung cancer patients than that from normal subjects (Supplementary information, Figure S9E). Likewise, the level of miR-214 was significantly higher in plasma from lung cancer patients than that from normal subjects (Supplementary information, Figure S9F). We next examined the effect of LLC MVs on the growth of xenograft tumors. As depicted in Supplementary information, Figure S10A, C57BL/6J mice were pre-treated with or without LLC MVs and implanted with mouse sarcoma S-180 cells. When the mice were implanted with 10⁶ S-180 cells, the plasma miR-214 levels were more than 20-fold higher in the LLC MV-pretreated mice compared with their control counterparts (Supplementary information, Figure S10B). Consistent with previous findings, the expansion of the Tregs compromised the immune response to the tumors, and the growth of the implanted tumors was significantly more rapid in the LLC MV-pretreated mice (Supplementary information, Figure S10C, left panel; Figure S10D, upper panel). Interestingly, when the mice were implanted with 10⁵ S-180 cells, there was no tumor growth in the control mouse group, whereas successful tumor implantation and rapid tumor growth occurred in the LLC MV-pretreated mice (Supplementary information, Figure S10C, right panel; Figure S10D, lower panel). The tumor weight in the LLC MV-pretreated mice was also significantly higher than that in the control group (Supplementary information, Figure S10E).

Next, we confirmed the effects of tumor-secreted miR-214 on Treg expansion and tumor growth *in vivo* by comparing tumor-derived MVs with MVs released from 293T cells [18]. Both the 293T cells and 293T MVs contained low levels of miR-214 (Supplementary information, Figure S11A and S11B). The overexpression of miR-214 in 293T cells (Supplementary information, Figure S11C) resulted in the production of MVs containing high levels of miR-214 (Supplementary information, Figure S11D). Transfection of pre-miR-214 in 293T cells did not substantially affect the protein, mRNA, and miRNA levels in the 293T MVs, with the exception of miR-214 (Supplementary information, Figure S11E-S11G). Various MVs from 293T and LLC cells were then used to pretreat mice, as depicted in Figure 6A. The LLC MV-induced downregulation of PTEN expression in mouse peripheral CD4⁺ T cells was reversed after the depletion

of miR-214 from the LLC MVs (Figure 6B). In addition, although the control 293T MVs did not affect CD4⁺ T cell PTEN expression, the “modified” 293T MVs, which contained high levels of miR-214, resulted in the downregulation of PTEN expression in CD4⁺ T cells (Figure 6B). Compared with the control 293T MVs, which did not affect the expansion of CD4⁺CD25^{high}Foxp3⁺ Tregs, LLC MVs and “modified” 293T MVs with high miR-214 levels significantly increased the percentage of Tregs in peripheral and splenic CD4⁺ T cells (Figure 6C and 6D). This induction of Tregs by LLC MVs, however, was largely inhibited by the removal of miR-214 from the LLC MVs (Figure 6C and 6D). Consistent with this result, the growth of the implanted sarcoma in the LLC MV-treated mice was significantly faster than that in the saline- or normal 293T MV-treated control mice (Figure 6E and 6F). The enhancement of tumor growth following LLC MV treatment was inhibited by the depletion of miR-214 from the LLC MVs (Figure 6E and 6F). As expected, the “modified” 293T MVs with high miR-214 levels were able to promote tumor growth (Figure 6E and 6F).

To determine whether the observed pro-tumor effects of the tumor-derived MVs were truly Treg-dependent, inducible Treg-deficient mice (diphtheria toxin-treated Foxp3^{DTR} mice) [29, 30] were treated with tumor-derived MVs (Supplementary information, Figure S12A). Our results indicated that the levels of miR-214 in peripheral CD4⁺ T cells of Foxp3^{DTR} mice and wild-type Foxp3⁺ littermates were significantly increased after LLC MV injection, whereas the levels of miR-214 were unchanged in mice injected with LLC MV/miR-214^{def} (Supplementary information, Figure S12B). Wild-type Foxp3⁺ mice treated with LLC MVs had a higher percentage of Tregs in peripheral CD4⁺ T cells compared to wild-type Foxp3⁺ mice treated with LLC MV/miR-214^{def} (Supplementary information, Figure S12C). However, neither the LLC MVs nor LLC MV/miR-214^{def} affected Treg levels in the Foxp3^{DTR} mice (Supplementary information, Figure S12C). The growth rate of the implanted sarcoma in the LLC MV-treated Foxp3⁺ mice was significantly faster than that in the LLC MV/miR-214^{def}-treated Foxp3⁺ mice (Supplementary information, Figure S12D and S12E). In contrast, neither LLC MVs nor LLC MV/miR-214^{def} promoted tumor growth in Foxp3^{DTR} mice (Supplementary information, Figure S12F and S12G). Importantly, Treg elimination in the adult Foxp3^{DTR} mice resulted in a more rapid development of terminal autoimmune disease compared with the Foxp3⁺ mice. In addition, several Foxp3^{DTR} mice became moribund. However, the MV treatment did not affect the death rate in the Foxp3^{DTR} mice (Supplementary information, Figure S12H).

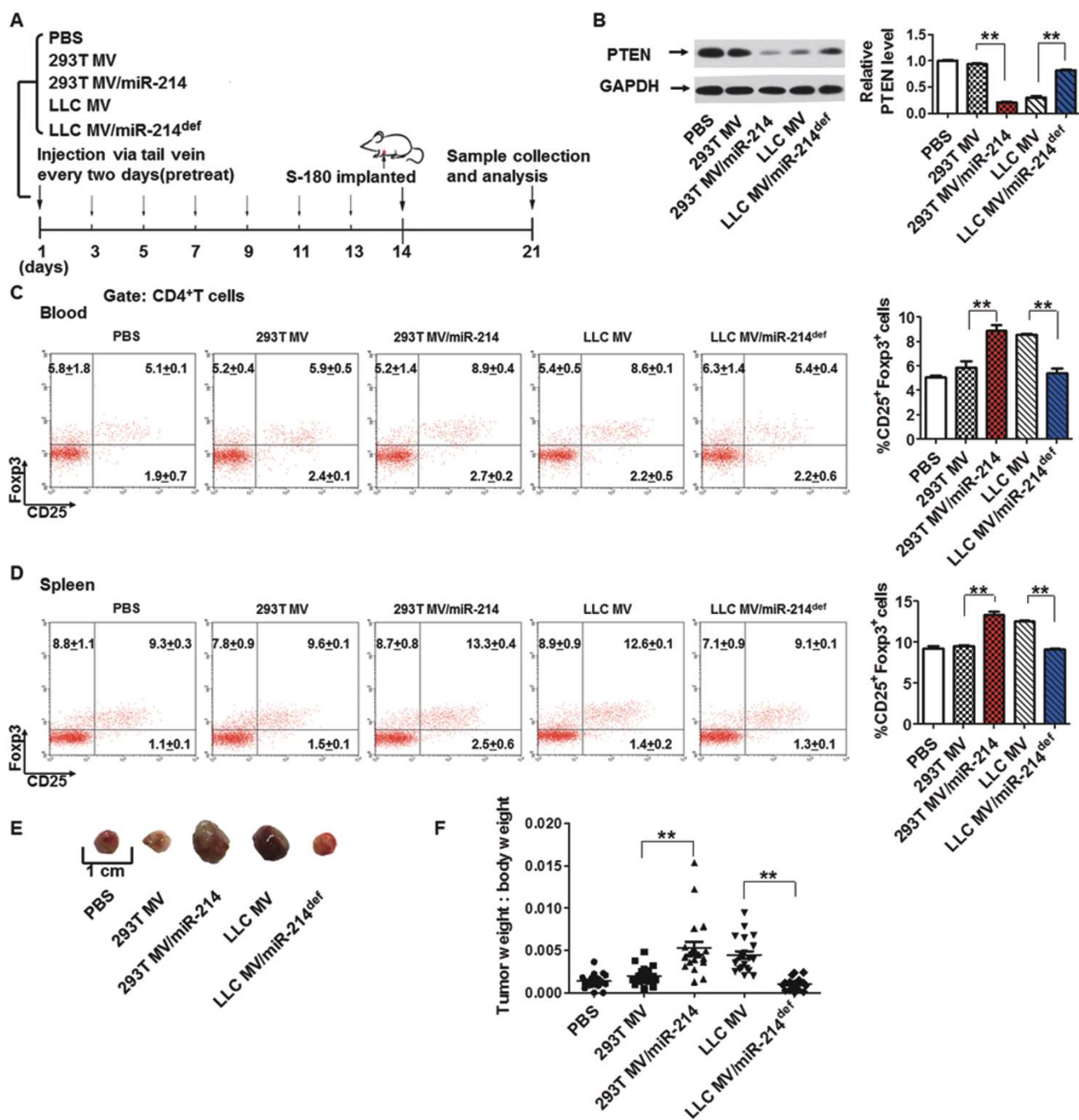
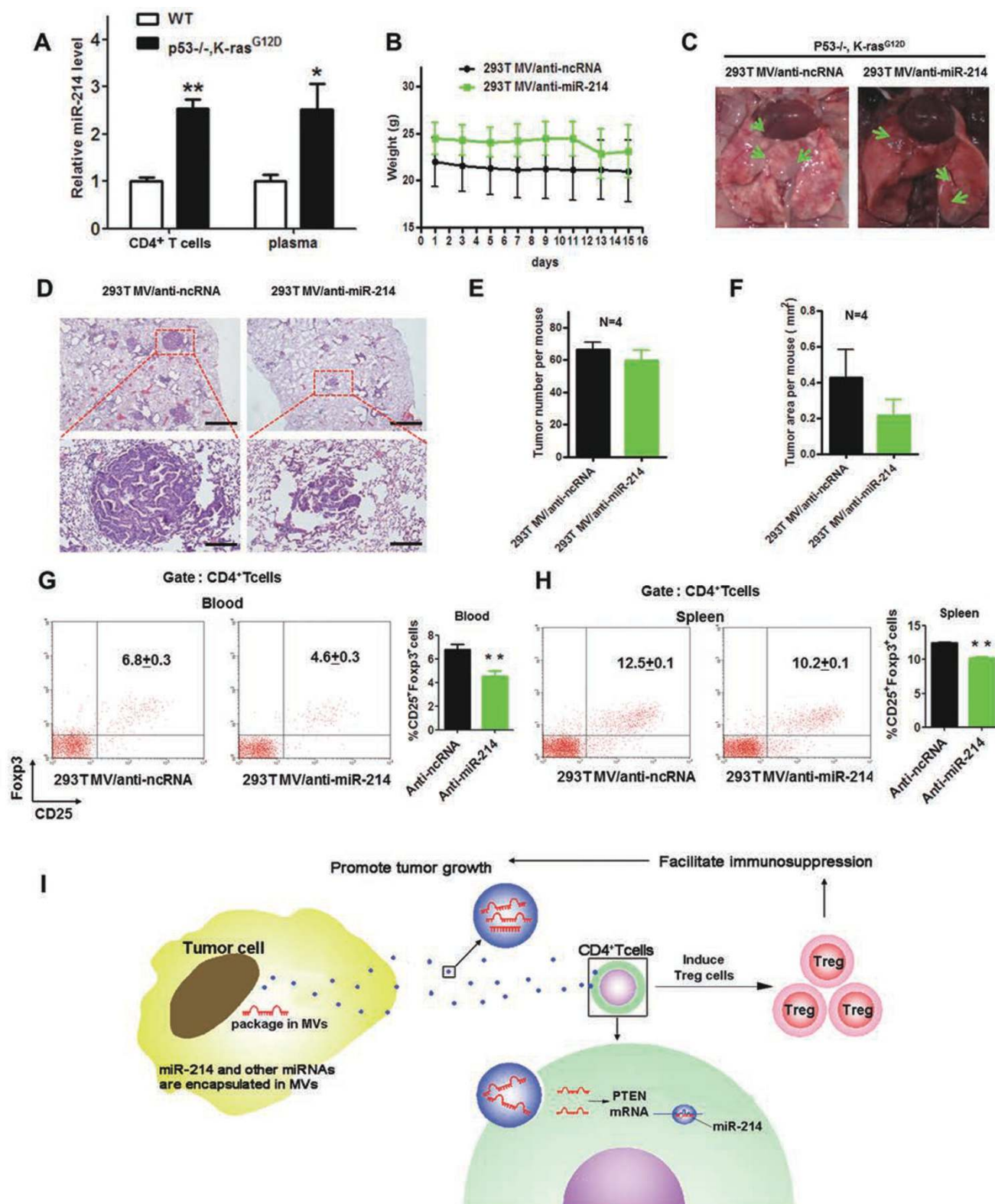


Figure 6 Effects of tumor-secreted miR-214 on CD4⁺ T cell PTEN expression, Treg induction, and tumor growth in C57BL/6J mice. **(A)** Flow chart of the experimental design. C57BL/6J mice were inoculated via the tail vein with saline (control) or MVs from normal 293T cells, 293T cells overexpressing miR-214, normal LLC cells, or LLC cells depleted of miR-214. The MVs were administered once every 2 days for 2 weeks (MV with 20 μg of total protein in 100 μl of PBS per mouse). The mice were then implanted with an S-180 sarcoma cells (10⁶ cells per mouse) and sacrificed after 7 days. **(B)** Western blot analysis of PTEN protein levels in CD4⁺ T cells from the mouse spleen. **(C, D)** Flow cytometric analysis of the population of Tregs in mouse whole blood **(C)** and spleen **(D)**. **(E, F)** Tumor growth in mice treated with various types of MVs. The data are presented as the mean ± SEM of three independent experiments. A total of six to eight mice were used in each group for each experiment. **P* < 0.05; ***P* < 0.01.

Blocking of cancer cell-secreted miR-214 downmodulates Treg induction and reduces tumor growth

Because tumor-secreted miR-214 promotes the expansion

of Tregs and the growth of implanted cancer cells, we determined whether the downregulation of circulating miR-214 levels could decrease the numbers of Tregs and



suppress tumor growth. We utilized 293T cells because they contain low levels of miR-214. We generated 293T MVs containing anti-miR-214 ASOs by transfecting 293T cells with anti-miR-214 ASOs. Transfection of anti-miR-214 ASOs into the 293T cells did not substantially affect the protein, mRNA, or miRNA levels in the 293T MVs (Supplementary information, Figure S13A-S13C). The mice were implanted with S-180 cells and then treated with anti-miR-214 ASO-containing 293T MVs as depicted in Supplementary information, Figure S14A. As expected, treatment with the anti-miR-214 ASOs significantly decreased sarcoma-induced miR-214 levels in CD4⁺ T cells (Supplementary information, Figure S13D). Consequently, treatment with the anti-miR-214 ASOs successfully rescued the S-180 sarcoma-induced reduction of PTEN levels (Supplementary information, Figure S14B) and thus suppressed the expansion of CD4⁺CD25^{high}Foxp3⁺ Tregs (Supplementary information, Figure S14C and S14D). Importantly, compared with the control mice treated with saline or 293T MVs containing a control oligonucleotide (anti-ncRNA), the mice treated with anti-miR-214 ASO-containing 293T MVs exhibited a significantly decreased growth rate and tumor size (Supplementary information, Figure S14E-S14G).

Next, we tested the effect of miR-214 in a *de novo* mouse model that spontaneously develops lung tumors [31-34]. This *de novo* lung cancer mouse model is sufficient to drive lung adenoma formation with the continuous activation of *K-ras*^{G12D} and loss of *p53* function *in vivo* by Adeno-Cre treatment. First, we measured the expression levels of miR-214 in CD4⁺ T cells and plasma from *p53*^{-/-},*K-ras*^{G12D} mice. The results indicate that the miR-214 levels in the CD4⁺ T cells and plasma from the *p53*^{-/-},*K-ras*^{G12D} mice were significantly higher than those in wild-type mice (Figure 7A). We next used the *p53*^{-/-},*K-ras*^{G12D} mouse model to test the effects of MV delivery

of anti-miR-214 ASOs. Anti-miR-214 ASO treatment prevented weight loss in the *p53*^{-/-},*K-ras*^{G12D} mice (Figure 7B). Compared with the *p53*^{-/-},*K-ras*^{G12D} mice treated with 293T MVs containing anti-ncRNA, the *p53*^{-/-},*K-ras*^{G12D} mice treated with the anti-miR-214 ASO-containing 293T MVs displayed decreased growth of spontaneous lung tumors (Figure 7C-7F). Anti-miR-214 ASOs also successfully suppressed the expansion of CD4⁺CD25^{high}Foxp3⁺ Tregs in the blood and spleen of *p53*^{-/-},*K-ras*^{G12D} mice (Figure 7G and 7H). These results indicate that MV delivery of anti-miR-214 ASOs is an efficient method to abolish tumor-induced Treg expansion and inhibit the growth of tumors.

Discussion

Investigations of the biological functions of cell-secreted miRNAs are in their infancy. While secreted miRNAs in MVs have been shown to function as novel components of the intercellular communication system, the role of secreted miRNAs in tumor progression has only recently been investigated. In the present study, we report a new signaling pathway between tumor and immune cells. As a result of this type of intercellular communication, tumor cell-secreted miRNAs play a fundamental role in tumorigenesis by suppressing the immune response in host cells, which in turn accelerates tumor growth. Interestingly, some recent studies have shown that tumor-secreted miRNAs can participate in tumor spread or promote muscle loss during tumorigenesis [35, 36]. These studies, combined with us, reveal the importance of tumor-secreted miRNAs as a novel regulator in cancer progression.

A significant alteration of miRNA expression profiles is a major feature of tumorigenesis. As an oncogenic miRNA, miR-214 is upregulated in various types of

Figure 7 The inhibition of spontaneous lung tumor progression in *p53*^{-/-},*K-ras*^{G12D} mice by 293T MVs containing anti-miR-214 ASOs. **(A)** qRT-PCR analysis of the miR-214 levels in the CD4⁺ T cells and plasma of *p53*^{-/-},*K-ras*^{G12D} mice compared with wild-type mice. **(B)** The *p53*^{-/-},*K-ras*^{G12D} mice were treated with Adeno-Cre at week 8. After 5 weeks of Adeno-Cre treatment, the mice were then treated with 293T MVs derived from 293T cells transfected with anti-ncRNA or anti-miR-214 ASOs (MV with 20 μg of total protein in 100 μl of PBS per mouse) once every 2 days for 2 weeks. The mice were then sacrificed and analyzed. A total of four mice were used in each group for the anti-ncRNA or anti-miR-214 treatment experiment. The body weights of the *p53*^{-/-},*K-ras*^{G12D} mice treated with anti-ncRNA or anti-miR-214 for 2 weeks were recorded. **(C)** Lung tumor formation in *p53*^{-/-},*K-ras*^{G12D} mice treated with 293T MVs derived from 293T cells transfected with anti-ncRNA or anti-miR-214 for 2 weeks. **(D)** Pathological photos of the lung tumors from the *p53*^{-/-},*K-ras*^{G12D} mice that were treated with 293T MVs derived from 293T cells transfected with anti-ncRNA or anti-miR-214 for 2 weeks. Scale bars: 500 μm (top panels) and 100 μm (bottom panels). **(E, F)** The tumor number and area were evaluated for each mouse. **(G, H)** Flow cytometric analysis of the numbers of Tregs in the whole blood and spleen of *p53*^{-/-},*K-ras*^{G12D} mice treated with 293T MVs derived from 293T cells transfected with anti-ncRNA or anti-miR-214 for 2 weeks. **P* < 0.05, ***P* < 0.01. **(I)** Working model for the role of tumor-secreted miR-214 in the conversion of CD4⁺ T cells into immune-suppressive Tregs and promotion of tumor immune escape.

tumors [37, 38] and induces cell survival and cisplatin resistance in human cancers by targeting the 3' UTR of PTEN, which leads to the activation of the Akt pathway. In the present study, we confirmed the elevated expression of miR-214 in different human cancer tissues (Figure 1A). Increased levels of plasma miR-214 were also detected in cancer patients (Figure 1B) and tumor-implanted mice (Figure 1E and 1G), suggesting that circulating miR-214 may be secreted by tumor cells. Furthermore, we observed that the majority of the plasma miR-214 was stored in MVs (Figure 1C, 1F and 1H), by which miRNAs can be delivered into other cells and tissues. Because miR-214 is a major oncogenic miRNA and its levels are generally elevated in the plasma of various cancer patients, the signaling events induced by tumor-secreted miR-214 may serve as a common mechanism in various cancer cells to create a tumor-promoting microenvironment.

The following results suggest a direct delivery of tumor-secreted miR-214 into CD4⁺ T cells through an MV-mediated pathway. (a) Treatment with the LLC MVs increased the levels of miR-214 but not pre-miR-214 in the recipient CD4⁺ T cells (Figure 2 and Supplementary information, Figures S8 and S9). By contrast, treatment with MVs that contained deficient miR-214 did not upregulate miR-214 in CD4⁺ T cells (Supplementary information, Figure S7B). (b) The intravenous injection of fluorescently labeled tumor-derived MVs into mice resulted in the association of the fluorescent MVs with peripheral CD4⁺ T cells (Figure 5A). (c) Similarly, the administration of tumor MVs containing a mutant miR-214 significantly increased the amount of mutant miR-214 in the peripheral CD4⁺ T cells of mice (Figure 5B-5E). Because most of the tumor cells expressed high levels of miR-214, the secretion of miR-214 by the tumor cells and the delivery of secreted miR-214 from the tumor cells to CD4⁺ T cells via the MVs may represent a common mechanism for cancers.

CD4⁺CD25^{high}Foxp3⁺ Tregs are a subset of CD4⁺ T cells that play an important role in maintaining self-tolerance and modulating immune responses in both normal and disease states [3-5, 39]. Because PTEN is a direct target of miR-214 [13, 25] and the targeted depletion of PTEN can regulate peripheral homeostasis of CD4⁺CD25^{high}Foxp3⁺ Tregs *in vivo* and their expansion *ex vivo* [28], tumor-secreted miR-214 may play a role in Treg expansion. In agreement with previous studies, our *in vitro* assay indicated that the expansion of Tregs could be induced by molecules that were present in the tumor-derived MVs, which likely included miR-214. Subsequently, we clearly demonstrated that the LLC MVs that contained high levels of miR-214 decreased the

protein levels of PTEN in CD4⁺ T cells and increased the percentage of CD4⁺CD25^{high}Foxp3⁺ Tregs in a dose-dependent manner (Figure 3A). The effects of the LLC MVs on the levels of PTEN and CD4⁺CD25^{high}Foxp3⁺ Tregs were reversed by depleting miR-214 in the LLC MVs (Figure 3B and Supplementary information, Figure S6B). These results further indicate that PTEN in CD4⁺ T cells was specifically targeted by the cancer cell-secreted miR-214. Our results indicate that secreted miR-214 targets PTEN and may serve as a critical "switch" molecule in Treg expansion, similar to previous reports that PTEN is a negative regulator of Treg expansion [28]. The activation of CD4⁺CD25^{high}Foxp3⁺ Tregs has a profound inhibitory effect on T cell-mediated immunity, particularly on CD4⁺ and CD8⁺ T cell function [6, 40, 41]. Several mechanisms, including the secretion of IL-10 and TGF- β and/or direct cell-cell contact, have been proposed to explain the suppression of effector immunocytes by Tregs [5]. Because the treatment of CD4⁺ T cells with the LLC MVs resulted in an increase in IL-10 but not IL-12 or TGF- β (Figure 4A-4C), our results suggest the involvement of IL-10 in the suppression of the immune system by tumor-induced Tregs.

Tumor cells utilize two major mechanisms to evade the immune response: (a) modification of the tumor cells themselves to decrease their sensitivity to attacks by functional immune cells [1], and (b) direct modulation of the host immune cells to secrete cytokines such as TGF- β and IL-10 [2]. The expansion of CD4⁺CD25^{high}Foxp3⁺ Tregs is one of the major mechanisms used by cancer cells to suppress immune cell function and facilitate tumor immune evasion [42, 43]. In tumor patients, levels of CD4⁺CD25^{high}Foxp3⁺ Tregs are generally increased and associated with compromised immune responses [6]. Although multiple factors, such as IL-2 [10] and TGF- β [11, 12], have been identified as being responsible for Treg expansion, the precise mechanisms that are responsible for the induction of tumor-associated Tregs remain poorly understood. Here, we present evidence suggesting that cancer cell-secreted miR-214 could expand the CD4⁺CD25^{high}Foxp3⁺ Treg population by decreasing the levels of PTEN in CD4⁺ T cells, which may promote tumor growth (Figure 6 and Supplementary information, Figure S10). The secretion of miRNAs by cancer cells to target immune cells and alter the immune system represents a novel pathway for tumor immune evasion. On the other hand, it is also possible that tumor-secreted miR-214 enters into other cells (e.g., effector T cells, macrophages, NK cells, and tumor cells), in which miR-214 may have different effect. Because peripheral T cells and tumor-secreted miR-214 are both in circulation, peripheral T cells may have more ample access to miR-214

compared to other cells in different organs/tissues. By directly tracking exogenous miR-214 both *in vitro* and *in vivo*, our data clearly show that tumor cell-secreted miR-214 can sufficiently enter into CD4⁺ T cells (Figure 5) and modulate PTEN expression and target cell function (Figures 3 and 6). Combining with our result that isolated CD4⁺ T cells can be directly converted into Tregs by miR-214-containing MVs (Figures 2 and 6), we conclude that downregulation of PTEN in CD4⁺ T cells and the expansion of Tregs are likely direct effects of miR-214 delivered by tumor MVs. Thus, tumor-secreted miR-214 inducing Tregs is one of major pathways to promote immune evasion and tumor growth.

Because Tregs are essential in tumorigenesis, the modulation of Tregs is an important area of research for the effective treatment of tumors. To evoke effective antitumor immunity, CD4⁺CD25^{high}Foxp3⁺ Tregs have been depleted with anti-CD25 antibodies [44] or the recombinant IL-2 diphtheria toxin conjugate DAB₃₈₉IL-2 [45]. A recent study also employed the adenovirus-mediated intratumoral expression of immunostimulatory proteins in combination with systemic CD4⁺CD25^{high}Foxp3⁺ Treg inactivation to induce tumor-destructive immune responses in mouse models [46]. Although the results were promising, human Treg-based therapies have previously been difficult to implement in the clinic, and relatively few clinical trials have been initiated. Because our results suggest that the expansion of the CD4⁺CD25^{high}Foxp3⁺ Treg population by tumor-secreted miR-214 likely serves as a common mechanism for various cancer cells to create a tolerant immune environment, inhibiting the transport of tumor-secreted miR-214 to immune cells may be a novel strategy to reverse tumor-induced immune tolerance. By employing cell-derived MVs to deliver anti-miR-214 ASOs into peripheral CD4⁺ T cells, we successfully reversed the downregulation of PTEN in CD4⁺ T cells and blocked the expansion of the CD4⁺CD25^{high}Foxp3⁺ Treg population (Figure 7 and Supplementary information, Figure S14). As expected, the MV-delivered anti-miR-214 ASOs effectively decreased the growth of the implanted and spontaneous tumors (Figure 7 and Supplementary information, Figure S14). In fact, MVs are natural nanocarriers derived from endogenous cells and would therefore be better tolerated by the immune system. Moreover, MVs protect small RNAs against RNase digestion and contain proteins that are required for small RNA function, such as Argonaute 2 (AGO2) [18, 47]. Thus, MVs have the intrinsic ability to traverse biological barriers and naturally transport functional small RNAs (including miRNA and siRNA) between cells. MV-mediated small RNA delivery may fill the void left by current delivery methods for systemic gene therapy.

In summary, our study provides the first evidence that tumor cells can actively modulate immune cell function through the secretion of MVs containing tumor-specific miR-214. As depicted in Figure 7I, tumor cells secrete miR-214 as well as other tumor-specific miRNAs through MVs that are then delivered into the peripheral CD4⁺ T cells. After the MVs enter the recipient CD4⁺ T cells, miR-214 decreases PTEN protein expression and facilitates Treg expansion, which, in turn, results in host immune suppression and tumor growth. Thus, tumor-secreted miRNA targeting of immune cells and the immune system represents an active pathway for tumor immune evasion. As an extension of this idea, we demonstrated that the inhibition of Treg-mediated immune suppression induced by tumor-secreted miR-214 via MV delivery of functional anti-miR-214 ASOs into CD4⁺ T cells represents a novel and effective therapeutic approach for cancer treatment.

Materials and Methods

Cell culture

LLC and 293T cells were purchased from the China Cell Culture Centre (Shanghai, China). The cells were cultured in high-glucose (4.5 g/L) DMEM (Gibco, Invitrogen, China) supplemented with 10% fetal bovine serum (FBS) (Gibco), penicillin, and streptomycin in a 5% CO₂ water-saturated atmosphere. The S-180 mouse sarcoma cell line was obtained from the China Cell Culture Centre, grown and maintained in RPMI-1640 medium (Gibco) supplemented with 10% FBS.

Mice

The 6-8-week-old male C57BL/6J mice were purchased from the Model Animal Research Centre of Nanjing University (Nanjing, China) and maintained under specific pathogen-free conditions at Nanjing University. The 7-week-old male nude mice were provided by the Experimental Animal Centre of Nanjing General Hospital (Nanjing, China). The Foxp3^{DTR} mice (where DTR is diphtheria toxin receptor) were generously provided by Prof Alexander Rudensky (Memorial Sloan-Kettering Cancer Center, New York) and maintained under specific pathogen-free conditions at Nanjing University.

To generate the heterotypic tumor model, S-180 mouse sarcoma cells were subcutaneously injected into the left armpits of the animals. After the tumors were separated from the animals, the length, width, and height of the tumors was measured with digital calipers. The tumor weights were determined, and the ellipsoid volume was calculated using the following formula: Volume = $\pi/6 \times$ (length) \times (width) \times (height). All animal care and handling procedures were performed in accordance with the National Institutes of Health's Guide for the Care and Use of Laboratory Animals and were approved by the Institutional Review Board of Nanjing University.

The *LSL-K-ras^{G12D}* and *p53 L/L* mice were originally generously provided by T Jacks (Massachusetts Institute of Technology) [31-34]. All mice were housed in a specific pathogen-free environ-

ment at the Shanghai Institute of Biochemistry and Cell Biology and treated in strict accordance with protocols approved by the Institutional Animal Care and Use Committee of the Shanghai Institute of Biochemistry and Cell Biology, Chinese Academy of Sciences.

Statistical analysis

All of the western blot images and semi-quantitative qRT-PCR results are representative of at least three independent experiments. The real-time PCR and cell migration assays were performed in triplicate, and each experiment was repeated several times. The data are presented as the means \pm SEM of at least three independent experiments. The differences were considered statistically significant if P was < 0.05 by Student's t -test.

For all the other materials and methods, see Supplementary information, Data S1 for details.

Acknowledgments

We thank Prof Alexander Rudensky (Memorial Sloan-Kettering Cancer Center, New York) for diphtheria toxin-treated Foxp3^{DTR} mice. We thank Drs Zheng-Gang Liu (Cancer Research Institute, NIH) and Dan Wu (Yale University) for their comments on the manuscript. This work was supported by grants from the National Basic Research Program of China (973 Program; 2014CB542300), the National Natural Science Foundation of China (81101330, 31271378 and 81250044), the Natural Science Foundation of Jiangsu Province (BK2011013 and BK2012014) and the Research Special Fund for Public Welfare Industry of Health (201302018). This work was also supported by the program for New Century Excellent Talents in University from the Ministry of Education of China (NCET-12-0261).

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