

Tunneling nanotubes spread fibrillar α -synuclein by intercellular trafficking of lysosomes

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Abstract

Synucleinopathies such as Parkinson's disease are characterized by the pathological deposition of misfolded α -synuclein aggregates into inclusions throughout the central and peripheral nervous system. Mounting evidence suggests that intercellular propagation of α -synuclein aggregates may contribute to the neuropathology; however, the mechanism by which spread occurs is not fully understood. By using quantitative fluorescence microscopy with co-cultured neurons, here we show that α -synuclein fibrils efficiently transfer from donor to acceptor cells through tunneling nanotubes (TNTs) inside lysosomal vesicles. Following transfer through TNTs, α -synuclein fibrils are able to seed soluble α -synuclein aggregation in the cytosol of acceptor cells. We propose that donor cells overloaded with α -synuclein aggregates in lysosomes dispose of this material by hijacking TNT-mediated intercellular trafficking. Our findings thus reveal a possible novel role of TNTs and lysosomes in the progression of synucleinopathies.

Keywords TNTs; α -synuclein; intercellular transfer; synucleinopathies

Subject Categories Membrane & Intracellular Transport; Neuroscience

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Introduction

Alpha-synuclein (α -synuclein) is a 140 amino acid protein widely expressed in the brain, localized principally in the cytosol and at presynaptic terminals in association with vesicles. Its exact function is still unclear, although it appears to be involved in neurotransmitter release (Bendor *et al.*, 2013; Burré *et al.*, 2013). Misfolded α -synuclein aggregates into intraneuronal inclusions called Lewy bodies (LB) and intragial inclusions, associated with neuronal and glial loss in specific regions of the brain. These inclusions represent the histopathological hallmark of synucleinopathies that include Parkinson's disease (PD), dementia with Lewy bodies (DLB) and

multiple system atrophy (MSA), which are characterized by a chronic and progressive decline in motor, cognitive, behavioural and autonomic functions (McCann *et al.*, 2014).

Considerable efforts have been made to understand the progression of synucleinopathies. Braak and colleagues were the first to show that α -synuclein neuropathology progresses throughout the brain in a specific and predictable manner, suggesting that the progression of the disease is associated with the propagation of a neurotropic infectious entity (Braak *et al.*, 2003a,b). Recent data indicate that this neurotropic agent might be aggregated α -synuclein. Pioneering studies supporting this hypothesis evidenced the presence of LB in grafted foetal mesencephalic dopaminergic neurons in the brain of PD subjects over 10 years after transplantation (Kordower *et al.*, 2008; Li *et al.*, 2008). Since then, intercellular α -synuclein transfer *in vitro* and *in vivo* has been reported (Desplats *et al.*, 2009; Hansen *et al.*, 2011; Kordower *et al.*, 2011; Angot *et al.*, 2012; Luk *et al.*, 2012a,b; Mougnot *et al.*, 2012; Masuda-Suzukake *et al.*, 2013, 2014; Bae *et al.*, 2014; Recasens *et al.*, 2014). Furthermore, it has been shown that fibrillar α -synuclein seeds the aggregation of the soluble form of the protein in a prion-like manner (Bousset *et al.*, 2013; Aulić *et al.*, 2014; Volpicelli-Daley *et al.*, 2014). Knowledge of the precise mechanism of α -synuclein propagation from one cell to another is therefore needed for better understanding the mechanism of disease progression. Although secretion was shown to be a possible mechanism of α -synuclein intercellular transfer (Desplats *et al.*, 2009; Lee *et al.*, 2010, 2013; Hansen *et al.*, 2011; Freundt *et al.*, 2012; Konno *et al.*, 2012; Brahic *et al.*, 2016), the predictable and specific patterns of spreading of α -synuclein inclusions in the brains of PD patients (Braak *et al.*, 2003b) together with *in vitro* and *in vivo* studies (Freundt *et al.*, 2012; Rey *et al.*, 2013; Holmqvist *et al.*, 2014; Peelaerts *et al.*, 2015) suggest a mechanism of spread following neuronal circuits.

Here, we develop a robust cell model to study *in vitro* the mechanism of cell-to-cell transfer of structurally and functionally characterized fluorescent human α -synuclein fibrils previously shown to recapitulate α -synuclein pathogenicity by being toxic and inducing seeding in neuronal cells and in animal models (Pieri *et al.*, 2012; Bousset *et al.*, 2013; Peelaerts *et al.*, 2015). We specifically

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investigated the role of tunneling nanotubes, F-actin containing membranous bridges that connect the cytoplasm of remote cells (Abounit & Zurzolo, 2012), allowing direct transfer of various cargoes (Austefjord *et al*, 2014) and thereby implicated in several physiological processes (Marzo *et al*, 2012). TNTs have been found in tissues of various origins (Chinnery *et al*, 2008; Lou *et al*, 2012; Seyed-Razavi *et al*, 2013; Ady *et al*, 2014), and recently TNT-like connections have been observed in human glioblastoma tumours implanted in mouse brains (Osswald *et al*, 2015). Of interest, data showing TNT-mediated HIV transfer from infected to uninfected T cells (Sowinski *et al*, 2008) and transfer of prions and other pathogens (Onfelt *et al*, 2004; Gousset & Zurzolo, 2009; Gousset *et al*, 2009) support the role of TNTs as a general conduit used by pathogens for spreading.

In the present study, we found that efficient transfer of α -synuclein fibrils between neuron-like cells and primary neurons in culture relied on tunneling nanotubes (TNTs). Following TNT-mediated transfer, α -synuclein fibrils were able to induce the aggregation of soluble, endogenous α -synuclein in acceptor cells. Most importantly, we show for the first time that fibrils normally directed to the lysosomal compartment for degradation both in neuron-like cells and in primary neurons can shuttle between cells in TNTs within lysosomal vesicles. Overall, our results support TNTs as efficient means for propagation of α -synuclein fibrils between neurons, and reveal a novel role played by lysosomes in this cell-to-cell transfer process.

Results

α -synuclein fibrils transfer efficiently between neuron-like cells

To investigate whether α -synuclein fibrils could transfer between neurons, we first assessed whether recombinant human α -synuclein fibrils of known dimension and molecular mass (on average 4,000 monomers for one fibril, Pieri *et al*, 2012) (Fig EV1A) were capable of entering catecholaminergic mouse (CAD) neuron-like cells. In time course experiments, we quantified the percentage of cells containing the fluorescent α -synuclein fibrils using flow cytometry and found a time-dependent internalization of fibrils with maximum fluorescence reached after 6 h (Figs 1A and EV1B). Time course measurements of lactate dehydrogenase (LDH) release after exposure to α -synuclein fibrils (up to 1 μ M) showed no change compared to control cells (Fig EV1C). This suggested that cells were viable, thus validating their use as donor cells in our transfer assay (see below and schematic in Fig EV2).

Although transfer of α -synuclein fibrils between neuronal cells has been shown, quantification (i.e. percentage of cells containing fibrils, number and size of α -synuclein foci) and intracellular localization of the transferred aggregates have not been thoroughly characterized. We set up a new co-culture assay where “donor” CAD cells (loaded overnight with α -synuclein fibrils) were co-cultured for 24 h at the ratio of 1:1 with naïve “acceptor” CAD cells expressing GFP. In order to remove any membrane-bound extracellular α -synuclein fibrils, donor cells were trypsin-washed before co-culture (Fig EV2A).

To automatically detect and quantify the number and size of fibrillar α -synuclein puncta in the donor and acceptor cell populations

separately, we used confocal microscopy and developed a specific script of the ICY software (see Appendix and Fig EV3). By this method, we found that 100% of acceptor cells contained fibrillar α -synuclein (Fig 1B and C). Furthermore, to determine the efficiency of transfer, we quantified the number of fluorescent puncta in the donor and acceptor cell populations. While donor cells contained about 70 puncta per cell, acceptor cells contained 38 puncta (Fig 1D) indicating that 35% of the puncta transferred from donor to acceptor cells. Together with the fact that 100% acceptor cells contained fibrillar α -synuclein puncta, this shows that in our model cell-to-cell transfer of fibrillar α -synuclein occurs and is very efficient.

We next analysed the size of α -synuclein puncta in donor and acceptor cells and found that the transferred puncta in acceptor cells correspond to smaller puncta (the median value is 0.15 μ m³) than the ones within the donor cells (the median value is 0.42 μ m³) (Fig 1E). This finding suggests size limitation of fibrillar α -synuclein puncta upon cell-to-cell transfer.

α -synuclein fibrils induce the aggregation of soluble α -synuclein after transfer in acceptor cells

We have previously demonstrated that α -synuclein fibrils identical to those used in this study efficiently seed the aggregation of soluble reporter α -synuclein (ChFP- α -synuclein) when exogenously added to neuron-like cells in culture or in rodent (Bousset *et al*, 2013; Peelaerts *et al*, 2015). However, a key question is whether these fibrils would seed cytoplasmic reporter α -synuclein, following their transfer from donor to acceptor cells. To address this question, we performed our co-culture experiment for 72 h using as acceptor CAD cells stably overexpressing soluble ChFP- α -synuclein (see schematic Fig EV2D). As expected for soluble α -synuclein, in control conditions, ChFP- α -synuclein cells cultured alone displayed a nucleocytoplasmic diffuse α -synuclein signal with a few ChFP- α -synuclein in discrete puncta, revealing low aggregation state of ChFP- α -synuclein (Fig 2A and B). In contrast, quantitative analysis revealed that after co-culture with donor cells loaded with α -synuclein fibrils, acceptor cells contained a significantly higher number of endogenous ChFP- α -synuclein in discrete puncta (4 and 15 α -synuclein-ChFP puncta per acceptor cell in control and co-culture conditions, respectively) (Fig 2A and B). In addition, 11% of the transferred Alexa-488 α -synuclein co-localized with ChFP- α -synuclein in discrete puncta (Fig 2A and C). These results indicate that α -synuclein fibrils are able to recruit soluble ChFP- α -synuclein, suggesting that they retain their seeding activity after being transferred from donor to acceptor cells.

α -synuclein fibril transfer is favoured by cell-to-cell contact

To further characterize the mechanism of transfer, we first determined whether transferred α -synuclein fibrils propagating from cell to cell co-localize with freshly taken up fibrils. Overnight co-cultures of donor (loaded with ATTO-550 α -synuclein fibrils) and acceptor cells were exposed for additional 12 h to α -synuclein fibrils labelled with Alexa-488. As expected, exogenously added Alexa-488 α -synuclein fibrils were taken up by both donor and acceptor cells with similar efficiency (Fig 3A). Alexa-488 fibrils co-localized with the ATTO-550 fibrils in donor cells indicating that fibrils taken up

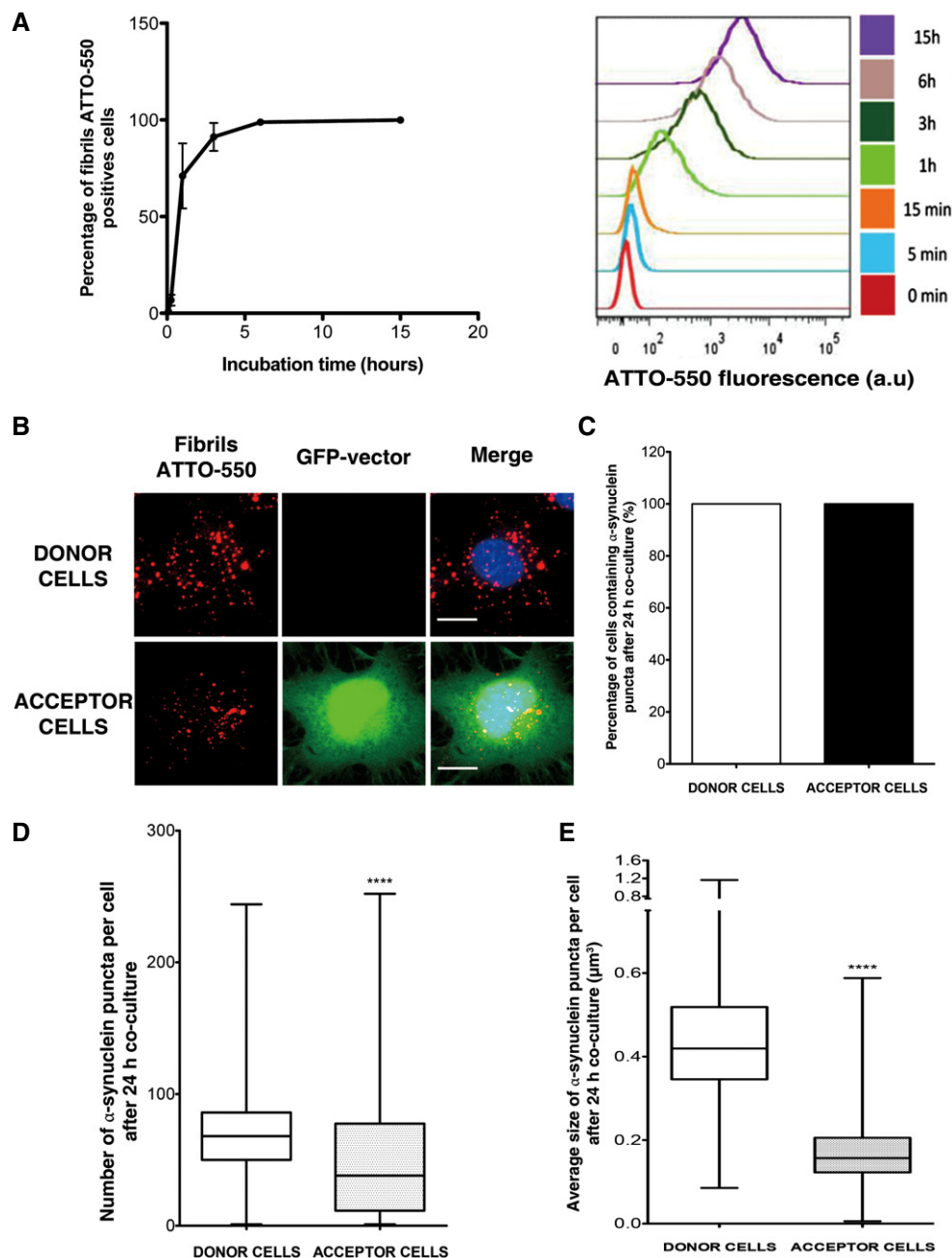


Figure 1. α -synuclein fibrils transfer efficiently between neuron-like CAD cells.

A Time course of rapid internalization of fluorescent α -synuclein fibrils by CAD cells (for characterization of α -synuclein assemblies, see Fig EV1A). Internalization was measured by recording ATTO-550-positive neuron-like cells by flow cytometry. Percentage of ATTO-550-positive cells was quantified (mean \pm s.e.m.) (left panel) and representative histograms of ATTO-550-positive cells are shown on the right panel (a.u., arbitrary units). $n = 3$ independent experiments. Similarly, α -synuclein fibril internalization was also confirmed by fluorescent microscopy (see Fig EV1B).

B Representative images of donor (upper panel) and acceptor cells (lower panel) after 24-h co-culture. Donor cells were loaded with α -synuclein fibrils prior to co-culture with GFP-transfected acceptor cells: in red, α -synuclein fibrils; in green, acceptor cells; and in blue, nuclei. Scale bars represent 10 μ m. $n = 3$ independent experiments. A larger field where donor and acceptor cells are shown is presented in Fig EV3A.

C Percentage of donor and acceptor cells containing α -synuclein fibrils after co-culture as in (B): all acceptor cells received α -synuclein fibrils.

D Quantification of the number of α -synuclein fibrils in donor and acceptor cells after co-culture as in (B). Donor cells contain around 70 α -synuclein fibril puncta (median), while acceptor cells contain 38 α -synuclein fibril puncta, respectively (**** $P < 0.0001$ by two-tailed Mann-Whitney U -test).

E Quantification of the average size of α -synuclein fibrillar foci in donor and acceptor cells after co-culture as in (B) (**** $P < 0.0001$ by two-tailed Mann-Whitney U -test). See also Fig EV3 for an example of α -synuclein fibrillar puncta detection in an acceptor cell. After detection, the number and the size of foci were determined using the ICY software.

Data information: In the box and whisker plots in (D and E), boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 100 cells scored for each independent experiment ($n = 3$).

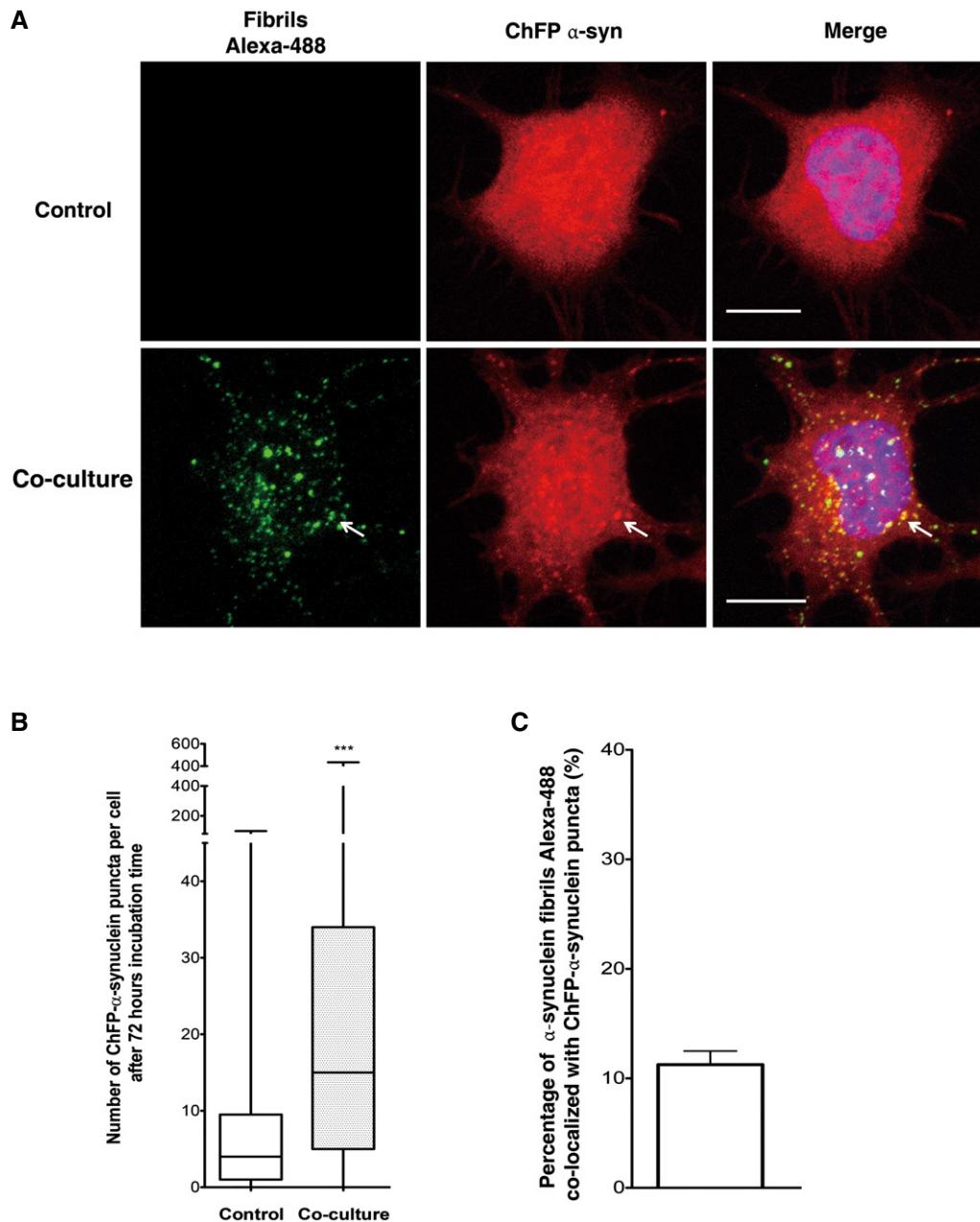


Figure 2. α -synuclein fibrils induce the aggregation of the soluble α -synuclein protein after transfer in acceptor cells.

- A** Representative confocal images showing ChFP- α -syn-transfected cells (in red) cultured alone (upper) or co-cultured with donor cells containing fluorescent Alexa-488 α -synuclein fibrils (in green) for 72 h (bottom). The arrow is pointing out representative α -synuclein fibrils co-localized with ChFP- α -syn puncta. Scale bars represent 10 μ m. Nuclei are stained with DAPI (blue). $n = 3$ independent experiments.
- B** Quantification of the number of ChFP- α -synuclein puncta in ChFP- α -syn acceptor cells showed a significant increase in ChFP- α -synuclein puncta number when cells were co-cultured with donor cells containing α -synuclein fibrils (** $P < 0.001$ by two-tailed Mann-Whitney U -test). In the box and whisker plot, boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 100 cells scored for each independent experiment ($n = 3$).
- C** Quantification of the percentage of co-localization of α -synuclein fibrils Alexa-488 and ChFP- α -syn puncta. Data are shown as mean \pm s.e.m. After co-culture, 11% of the transferred α -synuclein fibrils Alexa-488 co-localized with ChFP- α -syn puncta. $n = 3$ independent experiments.

initially (ATTO-550) and freshly taken up fibrils (Alexa-488) are directed to the same sub-cellular compartment (Fig 3B). Intriguingly, however, while in donor cells we observed high co-localization percentage between ATTO-550- and Alexa-488-labelled

fibrils (and reverse) ($66 \pm 0.66\%$), indicating that the two fibril batches were taken up following the same pathway (Fig 3B), in acceptor cells the taken up (Alexa-488) and transferred fibrils (ATTO-550) from the donor population co-localized much less ($16 \pm 0.9\%$)

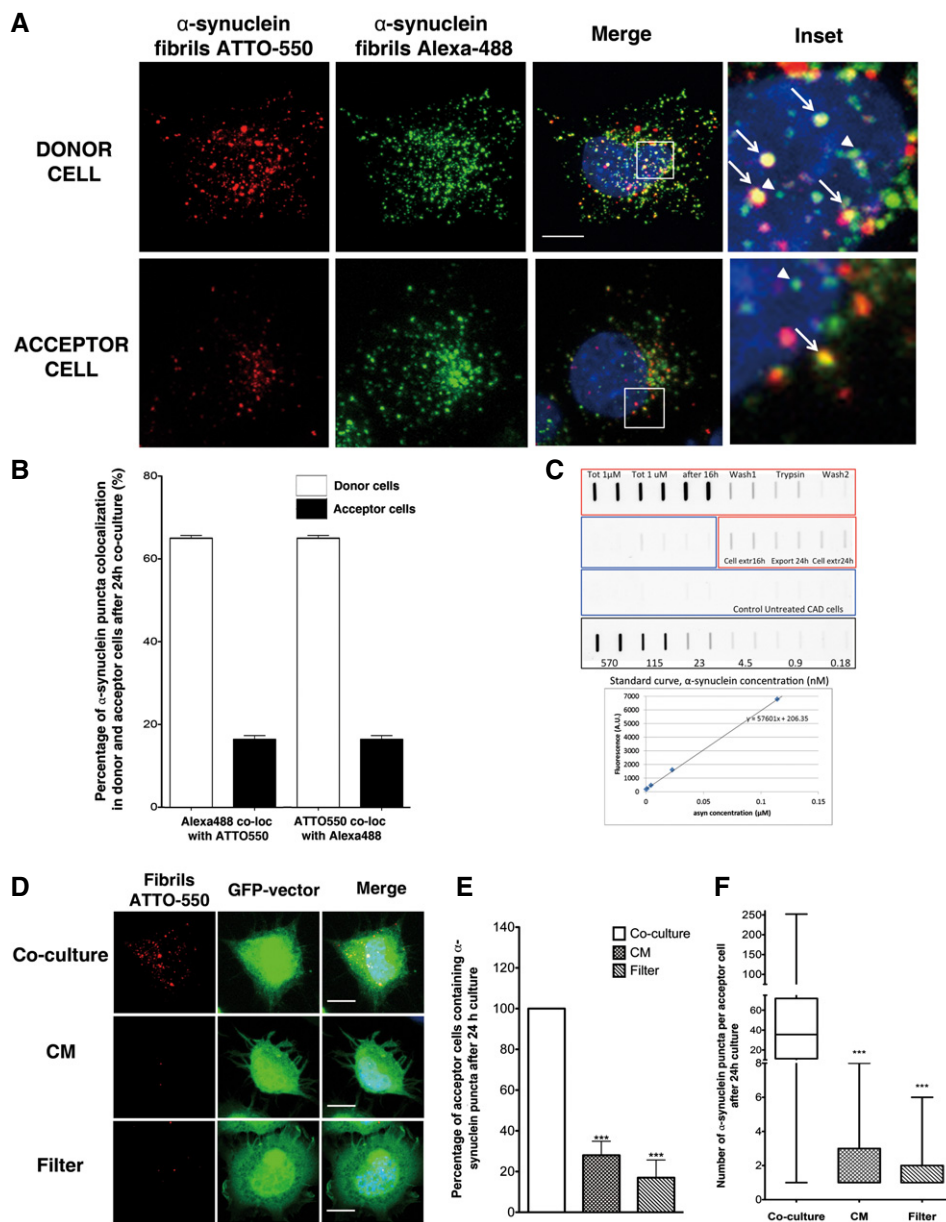


Figure 3. α -synuclein fibril transfer is favoured by cell-to-cell contact.

A Representative images of donor cells (upper panel) and acceptor cells (bottom panel) of the co-culture system explained in Fig EV2E: in red, ATTO-550 α -synuclein fibrils; in green, α -synuclein fibrils Alexa-488; and in blue, nuclei. Scale bar represents 10 μ m ($n = 3$ independent experiments). In insets, arrows point to co-localized ATTO-550 and Alexa-488 α -synuclein fibril puncta, whereas arrowheads point to puncta of α -synuclein Alexa-488 fibrils that do not co-localize with fibrillar ATTO-550 α -synuclein.

B Quantification of the percentage of Alexa-488 α -synuclein fibrils co-localizing with ATTO-550 α -synuclein fibrils and the reverse co-localization in discrete puncta in donor and acceptor cells as in (A). Both measurements revealed high co-localization of the two fluorophores in donor cells (white bar), but less in acceptor cells. Data are shown as mean \pm s.e.m. of 3 independent experiments.

C The amount of fibrils taken up by CAD cells exposed to 1 μ M fibrils, remaining in the cells and exported into the medium after 24-h incubation was quantified by a filter trap assay. $n = 3$ independent measurements, filtered in duplicate. The standard fluorescence curve for increasing ATTO-550 α -synuclein fibril concentrations is given.

D Representative images of GFP-transfected acceptor cells that were either (i) co-cultured with donor cells (upper panel, Co-culture), (ii) cultured with the conditioned medium of donor cells (middle panel, CM) or (iii) physically separated from donor cells using a filter (bottom panel, Filter). Prior to culture, donor cells were loaded with ATTO-550 α -synuclein fibrils. In red: α -synuclein fibrils, in green: acceptor cells and in blue: nuclei. Scale bars represent 10 μ m. $n = 3$ independent experiments.

E Quantification of the percentage of acceptor cells containing α -synuclein fibrils from images such as those presented in (C). When acceptor cells were cultured with the conditioned medium from donor cells previously treated with α -synuclein fibrils (not diluted, concentrated or diluted) or co-cultured with a filter, the percentage of acceptor cells containing α -synuclein puncta was low. Data are shown as mean \pm s.e.m. of 3 independent experiments. $***P < 0.001$ by two-tailed Student's t -test.

F Quantification of the number of puncta of α -synuclein fibrils per acceptor cell from (D). While in co-culture conditions the number of α -synuclein puncta in acceptor cells is on average 35 (median), this number was on average 1 puncta per acceptor cells in both CM and filter conditions ($***P < 0.001$ by two-tailed Mann-Whitney U -test). In the box and whisker plot, boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 100 cells scored for each independent experiment ($n = 3$).

(Fig 3B). These data suggest that taken up and transferred fibrils in the acceptor cells may not undertake the same pathway.

We further observed that the internalization of exogenous α -synuclein fibrils is dependent on dynamin-1 as the expression of dominant-negative dynamin-1 (GFP-Dyn1 K44A) affected uptake (Fig EV4A and B). Interestingly, the overexpression of this mutant (GFP-Dyn1 K44A) did not affect α -synuclein fibril transfer in CAD cells (Fig EV4C–E). These data indicate that transfer of fibrils from donor to recipient cells in co-culture is dynamin independent in contrast to uptake of fibrils from the medium.

To directly assess the role of secretion in the transfer, we loaded CAD cells with ATTO-550 fibrils for 16 h and first assessed the amount of fibrils that remained within the cells and that were exported from the cells into the medium after 24 h. Our quantitative assessment indicates that the proportion of α -synuclein fibrils taken up by cells within 16 h is $2.7 \pm 0.5\%$ (25–30 nM internalized for 1 μ M in total). The proportion of fibrillar α -synuclein remaining in the cells and exported into the medium after 24-h incubation was 55 and $45 \pm 5\%$, respectively (Fig 3C). We then directly challenged the acceptor cell population with the 24-h conditioned medium (CM) of donor cells (Figs EV2B and 3D, panel CM). To faithfully mimic the condition of co-culture, the CM was not concentrated, diluted nor centrifuged. In this condition, the percentage of acceptor cells containing α -synuclein puncta decreased substantially ($28.3 \pm 6.9\%$ in CM condition compared to 100% in co-culture condition) (Fig 3E, CM). Noteworthy, the decrease of fibril transfer was accompanied with a drastic decrease in the number of α -synuclein puncta transferred per acceptor cell (i.e. less than two puncta per cell compared to 38 when direct contact was allowed) (Fig 3F, CM).

When co-culture experiments where the donor and acceptor cells were separated by filters were performed (Figs EV2C and 3D, Filter), the percentage of acceptor cells containing α -synuclein puncta and the number of puncta in acceptor cells were drastically reduced compared to the control condition (Fig 3E and F). It is worth noting that this percentage was even more reduced than for the CM condition ($17.4 \pm 8.7\%$ and $28.3 \pm 6.9\%$ in filter and CM conditions, respectively) (Fig 3E). In these conditions, a 0.4- μ m membrane pore size semipermeable filter was used to prevent contact between donor and acceptor cells, but to allow the passage of soluble factors between the two cell populations (Gousset *et al*, 2009; Pasquier *et al*, 2012; Costanzo *et al*, 2013). One possibility to explain this result is that filters are impairing the passage of large cell debris contained in CM, which in turn will be responsible of the transfer observed in CM condition.

Altogether, these data indicate that in our experimental condition intercellular transfer is favoured by cell-to-cell contacts, and exclude a role for dynamin.

α -synuclein fibrils induce TNT formation and are present inside TNTs

Because tunneling nanotubes (TNTs) allow direct communication between cells and were found to play a role in the spreading of prions (and prion-like proteins) (Gousset *et al*, 2009; Wang *et al*, 2011; Costanzo *et al*, 2013; Ding *et al*, 2015), we next assessed the involvement of these structures in α -synuclein fibril transfer. Noteworthy, cells exposed to α -synuclein fibrils formed 20% more TNTs

than control cells (Fig 4A and B), indicating that fibrils can induce TNT formation. Interestingly, following uptake of α -synuclein fibrils, we observed a significant and persistent increase in reactive oxygen species (ROS), which could contribute to the increase in the TNT number (Wang *et al*, 2011; Gousset *et al*, 2013) (Fig EV1D and E). Furthermore, as early as 16 h after uptake, α -synuclein fibrils were found in the lumen of TNTs between donor cells (Fig 4C). Most importantly, in co-culture α -synuclein fibrils were present inside TNTs that directly connect donor and acceptor cell populations (Fig 4D). These results together with the data indicating that cell-to-cell contact contributes to the efficient α -synuclein fibril transfer strongly support the implication of TNTs in this process.

α -synuclein fibrils use TNTs to efficiently spread between neuron-like cells

In order to directly assess the involvement of TNTs in α -synuclein fibril transfer, we repeated the co-culture experiment in conditions enhancing or reducing TNT formation. We have previously shown that myosin-10 increases both TNT number and TNT-mediated unidirectional transfer of intracellular vesicles (Gousset *et al*, 2013). Therefore, we analysed whether overexpression of myosin-10 in donor cells would affect the transfer of the fibrils. As control, we verified that upon overexpression of myosin-10 there was an increase in TNT formation and in the amount of DiI-labelled vesicle transfer (Fig 4E and F) and demonstrated that myosin-10 donor cells containing α -synuclein fibrils formed 50% more TNTs (Fig 4E). Noteworthy, after co-culturing donor cells overexpressing myosin-10 with acceptor cells for 24 h, a statistically relevant 1.3-fold increase in the transfer of α -synuclein fibrils was observed as compared to control conditions (Fig 4F). To further support the involvement of TNTs in α -synuclein fibril transfer, we impaired TNT formation by plating co-cultured cells in sparse conditions. Indeed, in order to establish TNTs, cells are required to be in relative close contact (sub-confluent cells); therefore, in sparse conditions, the percentage of TNT-connected cells was drastically reduced ($42.8 \pm 12.2\%$ versus 100% in sparse and sub-confluent conditions, respectively) (Fig 4G). In these conditions, we observed a decrease in the percentage of acceptor cells containing α -synuclein fibrils ($38.5 \pm 9.9\%$ versus 100% in sparse and sub-confluent conditions, respectively) (Fig 4H), which paralleled the decrease in TNT number. Altogether, these results clearly show that TNTs contribute to the intercellular transfer of α -synuclein fibrils.

α -synuclein fibrils are predominantly found in lysosomal vesicles in the cytosol of donor cells and inside TNTs

To determine whether α -synuclein fibrils transfer through TNTs “naked” or embedded in vesicles, we first characterized the subcellular localization of fibrils in donor cells. As expected from previous studies showing that fibrils taken up from the medium are preferentially directed to the lysosomal compartment for degradation (Sung *et al*, 2001; Lee *et al*, 2008; Liu *et al*, 2009; Hasegawa *et al*, 2011; Konno *et al*, 2012), we found α -synuclein puncta in the endolysosomal compartment (Fig 5A left panel). Quantifications of co-localization (see the Appendix) revealed that almost 90% of fibrils localized within endolysosomal vesicles with the majority of fibrils found in lysosomes ($16.7 \pm 6.31\%$ in EEA1-, $24.9 \pm 0.52\%$ in

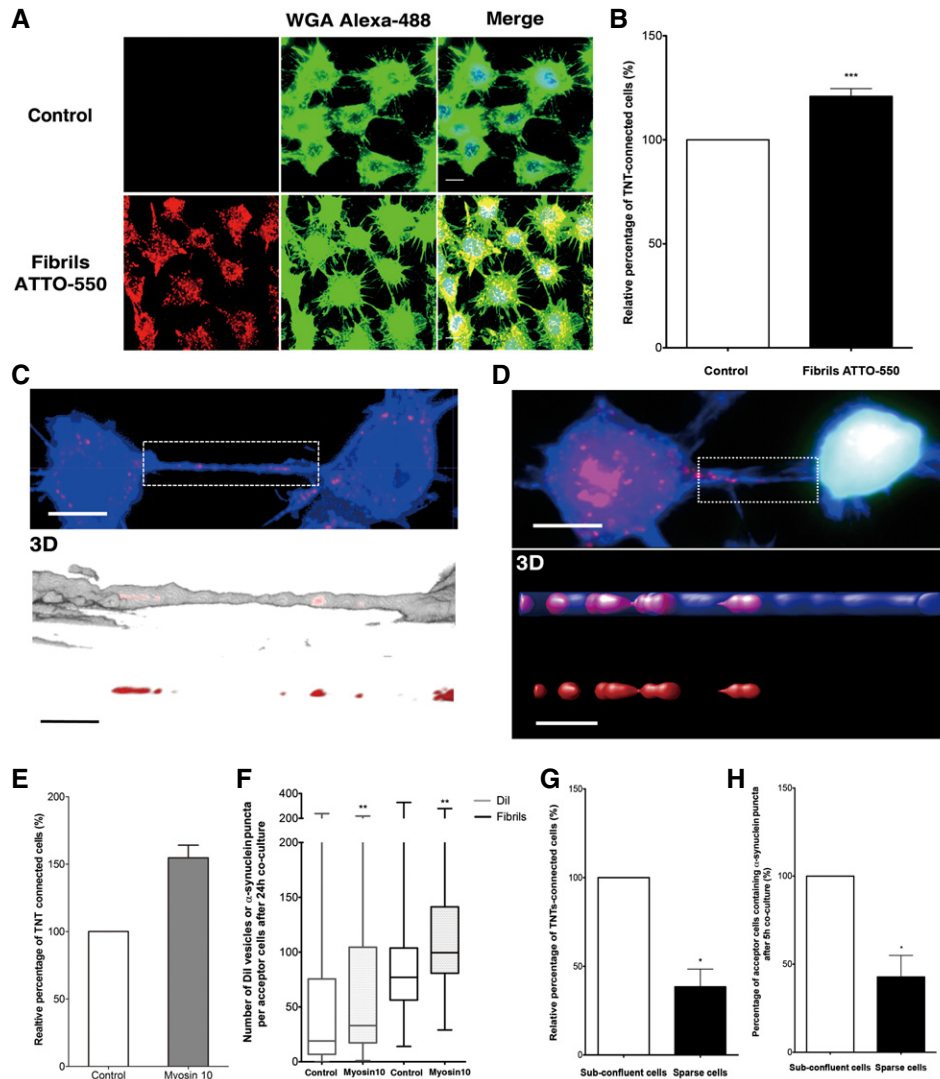


Figure 4. α -synuclein fibrils induce TNT formation and use TNTs to efficiently spread between neuron-like CAD cells.

A Representative images of control cells (upper panel) or cells treated with ATTO-550 α -synuclein fibrils (bottom panel) for 16 h: in red, α -synuclein fibrils; in green, WGA Alexa-488 (plasma membrane dye marker); in blue, DAPI. Scale bar represents 10 μ m. $n = 3$ independent experiments.

B Relative percentage of TNT-connected cells after α -synuclein fibril treatment as in (A) shows an increase in TNT number upon fibril treatment. Data are shown as mean \pm s.e.m. (***) $P < 0.001$ by paired, two-tailed Student's t -test.

C Confocal images showing one TNT connecting two cells loaded with ATTO-550 α -synuclein fibrils (in red) 16 h after co-culture and stained with WGA Alexa-488 (in blue). The image represents a Z-projection of the several middle stacks where the TNT is located (top panel). Insets show 3D reconstructions of the TNT using ICY software (middle panel, merge; bottom panel, red channel). Several α -synuclein fibrils (red) are present in the lumen of the TNT (WGA Alexa-488 in white). Scale bars represent 10 μ m (confocal images) and 4 μ m (insets).

D Confocal image showing a TNT connecting a donor cell containing α -synuclein fibrils and a GFP-transfected acceptor cell (top panel) strongly suggesting α -synuclein fibril transfer from donor to acceptor cell. Insets show 3D reconstructions of TNT containing α -synuclein fibrils in the lumen of the TNT: in red, α -synuclein fibrils; in green, GFP-transfected acceptor cells; in blue, WGA Alexa-633. Scale bars represent 10 μ m (confocal images) and 3 μ m (insets).

E Relative percentage of TNTs in cells containing α -synuclein fibrils untransfected (control) or transfected with myosin-10 (myosin-10). Overexpression of myosin-10 in the donor cell population led to an increase in the number of TNTs. Data are shown as mean \pm s.e.m.

F Quantification of the average number of ATTO-550 α -synuclein fibrils per acceptor cell after 24-h co-culture with donor cells containing α -synuclein fibrils untransfected (control) or transfected with myosin-10 (myosin-10). In the box and whisker plot, boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 100 cells scored for each independent experiment ($n = 3$). Note that myosin-10 overexpression increases both the transfer of vesicles labelled with the lipophilic dye Dil and the number of transferred α -synuclein puncta in acceptor cells; $n = 3$ independent experiments (***) $P < 0.01$ by two-tailed Mann-Whitney U -test.

G Quantification of the relative proportion of TNTs in co-culture of donor cells containing α -synuclein puncta and GFP acceptor cells under sub-confluent or sparse culture conditions. Data are shown as mean \pm s.e.m.; $n = 3$ independent experiments (* $P < 0.05$ by paired, two-tailed Student's t -test).

H Quantification of the percentage of acceptor cells containing α -synuclein puncta of the co-culture in (G). $n = 3$ independent experiments (* $P < 0.05$ by two-tailed Mann-Whitney U -test). Conditions impairing TNT formation (i.e. sparse conditions) induce a drastic decrease in TNT number and α -synuclein fibril transfer compared to conditions allowing TNT formation (sub-confluent conditions). Data are shown as mean \pm s.e.m.

Data information: In (B) and (E–H) at least 100 cells were counted for each independent experiment.

Vamp3- and 51.8 \pm 3.08% in Lamp1-positive vesicles) (Fig 5A right panel). Next, we investigated the subcellular localization of α -synuclein fibrils inside TNTs. Interestingly, the large bulk of fibrils in TNTs were found in Lamp1-positive vesicles and only rarely did they co-localize with markers of the early or recycling endosomes (Figs 5B and EV5A and B), indicating that fibrils travel inside TNT-connecting cells preferentially confined in lysosomal vesicles.

α -synuclein fibrils transferred in acceptor cell via TNTs are confined inside lysosomal vesicles derived from donor cells

To examine the destiny of α -synuclein fibrils after TNT-mediated transfer (i.e. once they reached the acceptor cells), we performed similar co-localization studies and found that transferred fibrils predominantly co-localized with Lamp1-positive vesicles (28.7 \pm 3.37%) in acceptor cells (Fig 5C). Only a few fibrils co-localized with EEA1- or Vamp3-positive vesicles (2.75 \pm 0.55% and 5.13 \pm 1.56% for EEA1 and Vamp3, respectively) (Fig 5C). These observations are consistent with the predominant lysosomal localization of α -synuclein fibrils in the TNT lumen and indicate that once transferred to acceptor cells, the majority of α -synuclein fibrils are confined inside lysosomal vesicles.

Finally, in order to demonstrate that lysosomal vesicles containing α -synuclein fibrils in acceptor cell came from donor cells (i.e. transfer), we pre-stained the lysosomes of donor cells with LysoTracker (i.e. lysosomotropic agent labelling lysosomal vesicles). After 24-h co-culture with acceptor cells, 100% of donor and acceptor cells contained LysoTracker-positive vesicles indicating transfer of lysosomes from donor to acceptor cells (Fig 5D). Importantly, almost 80% of LysoTracker-positive vesicles co-localized with α -synuclein fibrils in both donor and acceptor cells (76.2 \pm 1.2% and 79.5 \pm 0.9% for donor and acceptor cells, respectively) (Fig 5E). These results clearly indicate that α -synuclein fibrils in acceptor cells are inside lysosomes coming from donor cells. Consistent with this, transferred α -synuclein fibrils did not co-localize with GFP-Rab7-labelled lysosomal vesicles of the acceptor cells (Fig EV5C).

α -synuclein fibrils transfer efficiently between neurons

We next investigated α -synuclein fibril transfer between primary cortical neurons. Firstly, we determined by quantitative confocal microscopy the proportion of primary cortical neurons (6 div) that internalized fluorescent α -synuclein fibrils after 16 h of incubation with two different fibril concentrations (0.5 and 1 μ M). Internalization was very efficient and increased with the concentration of fibrils (around 75 and 100%, respectively) (Figs EV1F and 6A). Like in CAD cells, uptake of α -synuclein fibrils did not lead to an increase in LDH release over 72 h as compared to control neurons (Fig EV1G). Cortical neurons take up α -synuclein fibrils as early as 3 and up to 9 days *in vitro* (div) (Appendix Fig S1); however, we chose to perform the transfer experiments with 1-week-old neurons given that it gave us the best balance between good cell maturity and density.

To study neuron-to-neuron transfer of α -synuclein fibrils, “donor” neurons, exposed overnight to α -synuclein fibrils (1 μ M), were co-cultured with naïve freshly dissected “acceptor” CellTracker Green (CTG)-labelled neurons for 72 h (Fig EV2F). Appropriate controls were included in every experiment in order to assess health

of the young acceptor neurons (Appendix Fig S2). After 72-h co-culture, the number and size of puncta in both donor and acceptor cells were quantified by confocal microscopy using the same script of the ICY software employed for CAD cells (Appendix Fig S3). Note that because the acceptor neurons were pre-labelled in green with CTG, it was possible to easily discriminate them from the donor population (Appendix Figs S2 (lower panel) and S3). Under these experimental conditions, 94 \pm 1.6% of acceptor neurons contained at least one α -synuclein puncta (Fig 6B and C). More precisely, while donor cells contained a median of 26 puncta, acceptor cells had a median of 7 puncta (Fig 6D). Similar to CAD cells, the mean size of puncta in donor cells (the median value is 0.25 μ m³) was larger than in acceptor cells (the median value is 0.17 μ m³) (Fig 6E). Taken together, these results indicate that healthy neurons are able to take up and efficiently transfer α -synuclein fibrils to co-cultured neurons, in agreement with previous data (Freundt *et al*, 2012).

α -synuclein fibrils are located in lysosomal vesicles in both donor and acceptor neurons

In order to gain further insight into the mechanisms of α -synuclein transfer between neurons, we assessed the subcellular localization of α -synuclein puncta. Given our observations with neuron-like cells, we focused on the lysosomal pathway. After 72 h in co-culture, we found that the majority of α -synuclein puncta co-localized with Lamp1-positive vesicles in both donor (46 \pm 2.0%) and acceptor neurons (50 \pm 1.4%) (Fig 6B and F).

Cell–cell contacts facilitate α -synuclein fibril transfer between neurons

To determine whether physical contacts between neurons favour the transfer of α -synuclein fibrils as is the case for CAD cells, we grew donor neurons, containing α -synuclein fibrils, and CTG-labelled acceptor neurons on separated coverslips in the same Petri dish (Costanzo *et al*, 2013). Under our experimental conditions, the two populations of neurons share the same culture medium but establish no physical contacts (Fig 7A, No contact). Quantitative confocal microscopy revealed that transfer of fibrillar α -synuclein was significantly reduced under these conditions with only 3.1 \pm 1.8% of the acceptor cells containing α -synuclein puncta compared to co-culture conditions where cells could establish contacts (Fig 7B and C).

To assess the contribution of secretion of α -synuclein from donor cells to the propagation process, we added the 72-h conditioned medium from donor neurons (loaded overnight with fluorescent α -synuclein fibrils) to naïve acceptor neurons (Fig 7A, CM). As in the case of CAD cells (Fig 3C–E), the CM was not concentrated, diluted or centrifuged prior to its addition to acceptor cells. We found a significant decrease both in the percentage of acceptor cells containing α -synuclein (30 \pm 3%) and in the mean number of α -synuclein puncta per cell (0.9) (Fig 7B–D). We also analysed the presence of α -synuclein in the 72-h CM of donor cells. By Western blots, we detected a strong α -synuclein signal in the cell lysate of loaded neurons (overnight loading plus 72 h) compared to control cells. We also detected α -synuclein in the 72-h CM of loaded neurons, albeit this signal was quite low (Fig 7E). Altogether, these results suggest that cell-to-cell contacts contribute to neuron-to-neuron transfer of α -synuclein fibrils.

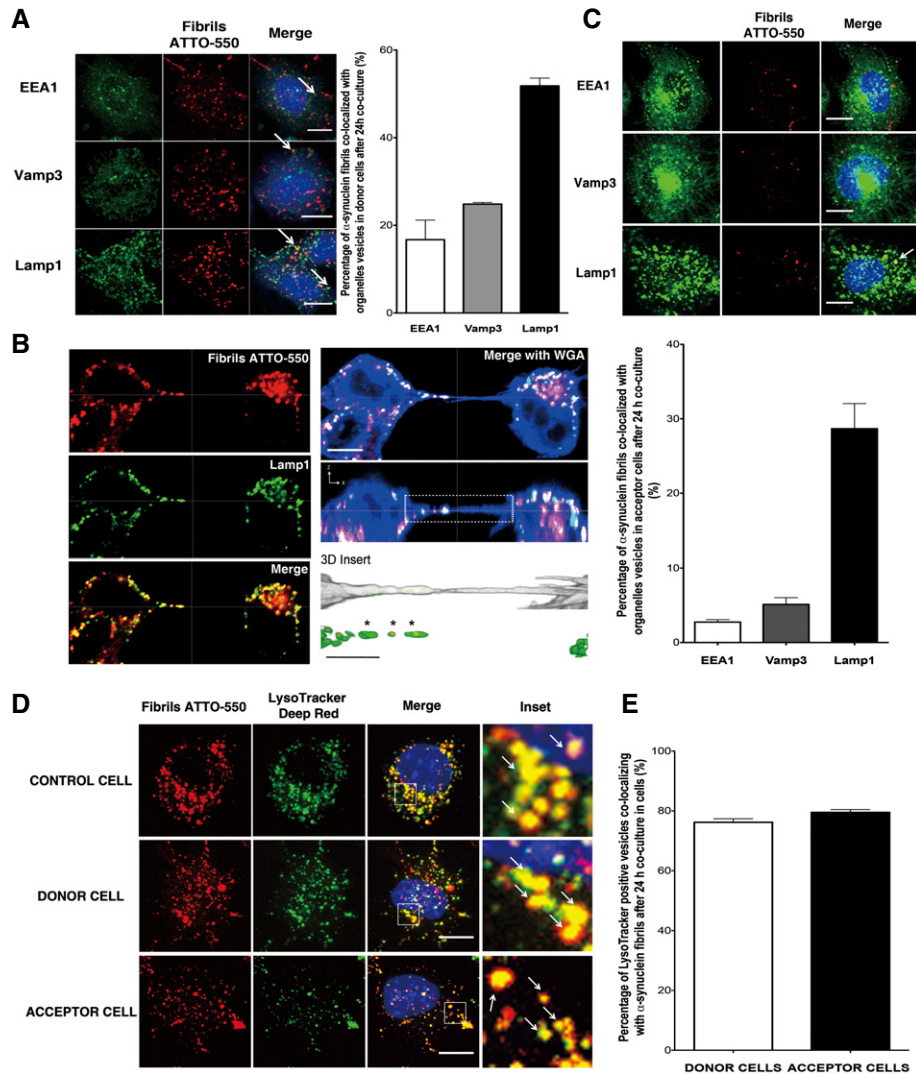


Figure 5. α -synuclein fibrils are predominantly found in lysosomal vesicles in the cytosol of donor cells and inside TNTs after transfer.

- A** Subcellular localization of α -synuclein fibrils in donor cells after 24-h co-culture. The left panels show representative images of co-localization of α -synuclein puncta with early endosomes (EEA1, top panel), recycling endosomes (Vamp3, middle panel) and lysosomal vesicles (Lamp1, bottom panel) (see arrows for co-localized spots): in red, α -synuclein fibrils; in green, EEA1, Vamp3 or Lamp1; and in blue, nuclei. Scale bars represent 10 μ m. On the right, the bar graph represents the percentage of α -synuclein puncta co-localized with endolysosomal vesicles. Data are shown as mean \pm s.e.m. from three independent experiments.
- B** Confocal images showing a TNT connecting two cells loaded with ATTO-550 α -synuclein fibrils immunostained with Lamp1 (in green). Labelling of TNTs is performed using a plasma membrane dye (WGA Alexa-633, in blue). The image represents a Z-projection of the several middle stacks where the TNT is located. The merged image with WGA Alexa-633 (blue), the orthogonal view and insets showing 3D reconstructions of the TNT (with WGA in white) are on the right. Several α -synuclein fibrillar foci (in red) are present in the lumen of the TNT and embedded in Lamp1-positive vesicles (in green). Of note, only tubular structures physically connecting remote cells and not touching the substratum were identified as TNTs to distinguish them from filopodia. Scale bars represent 10 μ m (confocal images) and 3 μ m (insets). The merged image of α -synuclein fibrils (red) and Lamp1 (green) channels and the respective single channels are shown on the left panel.
- C** The top panels show representative images of GFP-transfected acceptor cells after 24-h co-culture with donor cells containing ATTO-550 α -synuclein fibrils. Immunofluorescence was performed with endolysosomal organelle markers (top panel, EEA1; middle panel, Vamp3; and bottom panel, Lamp1) and revealed that α -synuclein fibrils co-localized with lysosomal vesicles (see arrow): in red, α -synuclein fibrils; in green, organelle markers; and in blue, nuclei. Scale bars represent 10 μ m. The bar graph below shows the quantification of the percentage of co-localization of α -synuclein fibrils with organelle markers, revealing that α -synuclein puncta co-localized mostly with lysosomal vesicles. Data are shown as mean \pm s.e.m. from three independent experiments.
- D** Representative confocal pictures of control cells loaded with ATTO-550 α -synuclein fibrils and LysoTracker Deep Red (top panel), and of donor cells previously loaded with ATTO-550 α -synuclein fibrils and LysoTracker Deep Red (middle panel), that were co-cultured for 24 h with GFP-transfected acceptor cells (bottom panel, GFP vector not shown). In red, α -synuclein fibrils; in green, LysoTracker Deep Red; and in blue, nuclei. Scale bars represent 10 μ m, and arrows in inset pinpoint fibrils co-localizing with LysoTracker-positive vesicles.
- E** The percentage of co-localization of LysoTracker-positive vesicles with α -synuclein fibrils revealed that 80% of lysosomal vesicles that transferred from donor to acceptor cells contained α -synuclein fibrils, thus demonstrating direct transfer of α -synuclein fibrils from donor to acceptor cells in majority inside lysosomal vesicles derived from donor cells. Data are shown as mean \pm s.e.m. from three independent experiments.

Data information: In (A, C and E) at least 100 cells were scored for each independent experiment.

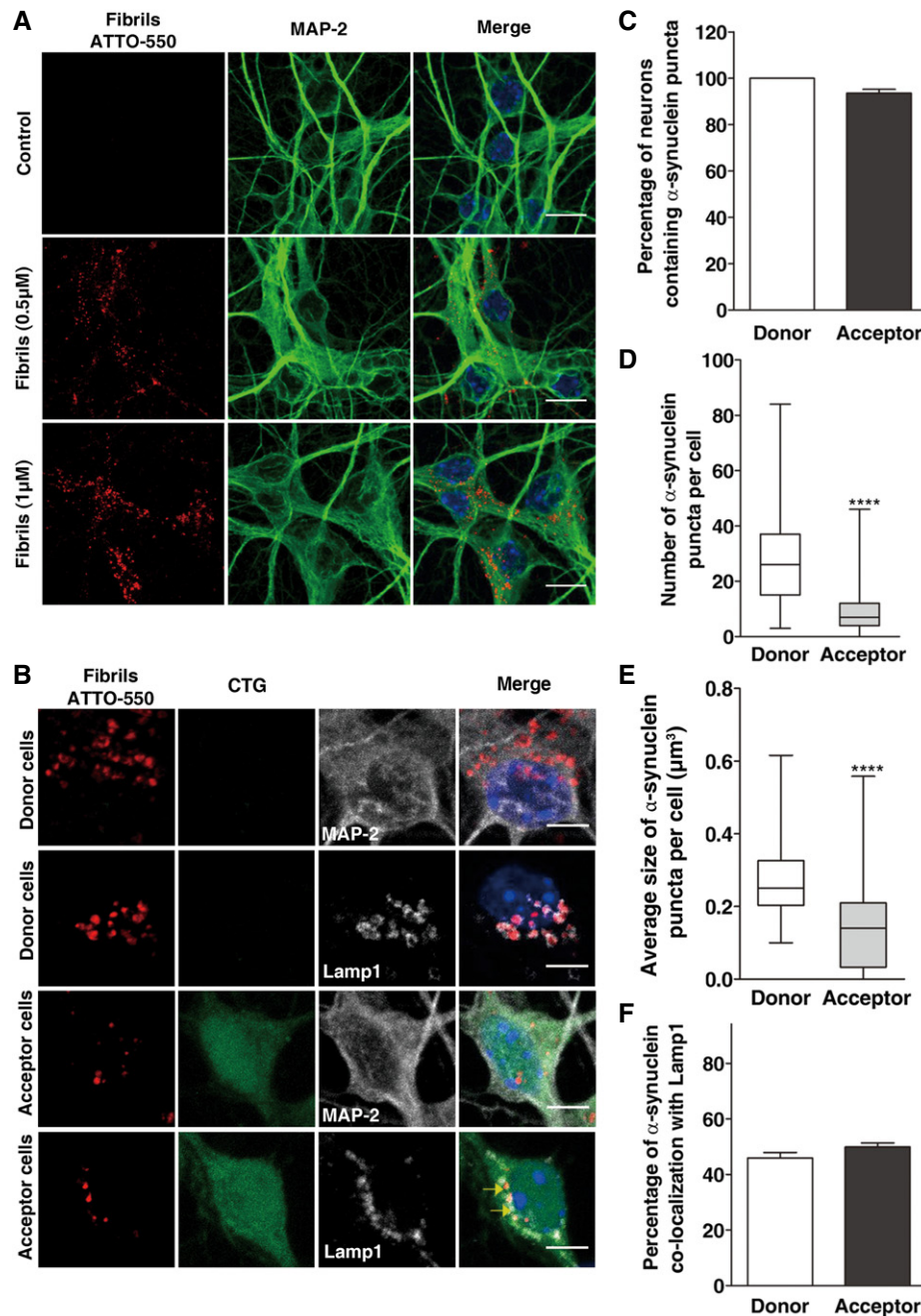


Figure 6. α -synuclein fibrils are found in lysosomes and are efficiently transferred between primary neurons.

- A Representative images of control (upper panel) and α -synuclein-loaded (middle, 0.5 μ M; bottom, 1 μ M) neurons after 16 h. In red, α -synuclein fibrils; in green, MAP-2; and in blue, nuclei. Scale bars represent 10 μ m.
- B Representative images of donor (upper panels) and acceptor neurons (bottom panels) after 72 h of co-culture. Donor neurons were loaded with α -synuclein fibrils prior to co-culture with CTG-labelled acceptor neurons and then cells were labelled either with MAP-2 (first and third panels) or Lamp1 (second and fourth panels). In red, α -synuclein fibrils; in green, CTG; in white, MAP-2/Lamp1; and in blue, nuclei. Scale bars represent 5 μ m. Yellow arrows point to co-localized ATTO-550 α -synuclein puncta and Lamp1.
- C The graph bar shows the percentage of donor (white bar) and acceptor (black bar) neurons containing α -synuclein puncta. Data are shown as mean \pm s.e.m.
- D The box plot depicts the number of puncta in donor (white) and acceptor (grey) neurons.
- E Box plot showing the distribution of the average size of α -synuclein puncta in donor (white) and acceptor (grey) neurons.
- F The graph bar represents the percentage of α -synuclein puncta co-localized with lysosomes in donor (white) and acceptor (black) neurons. Data are shown as mean \pm s.e.m.

Data information: Graphs in (C–F) correspond to data from at least 100 cells scored for each of three independent experiments analysed after 72 h of co-culture.

**** $p < 0.0001$ compared to donor by two-tailed Mann–Whitney U -test. In the box and whisker plots of (D and E), boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value.

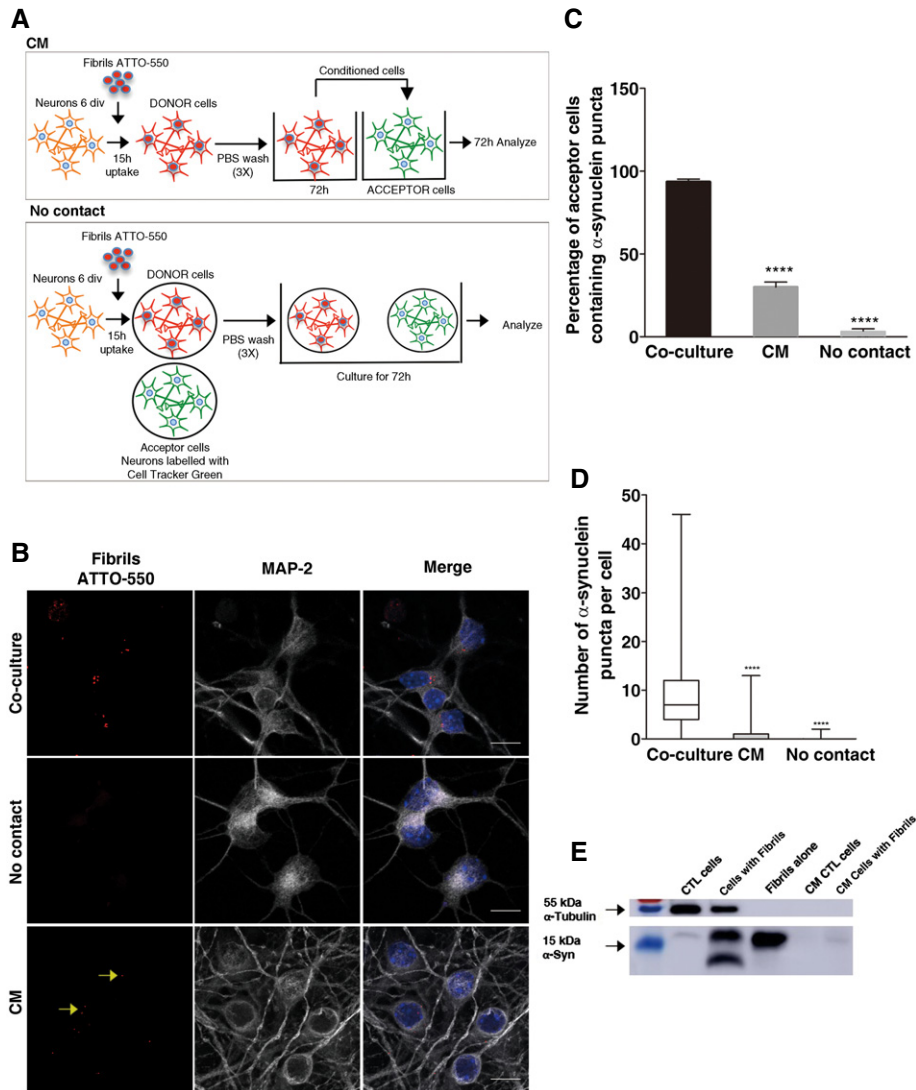


Figure 7. Cell-to-cell contact facilitates α -synuclein transfer between primary neurons.

A Experimental set-up used to assay transfer through conditioned medium (upper scheme) and cell-to-cell contacts (lower scheme) in neurons. For the CM experiments, donor neurons are loaded with α -synuclein fibrils for 16 h, washed 3 times with PBS, and then fresh medium is added and left to condition for 72 h. CM is then collected and transferred to acceptor neurons (10 div) for 72 h. In the cell-to-cell contact impairment (No contact) experiment, the donor (already loaded) and newly dissected acceptor (already labelled) neurons are plated on different coverslips placed in the same Petri dish. In both experiments, the percentage of cells containing α -synuclein ATTO-550 puncta, and the number and mean size of α -synuclein puncta per cell are quantified using the ICY software after 72 h.

B Representative images of acceptor neurons that were either (i) co-cultured with donor neurons (upper panel, Co-culture), (ii) physically separated from donor cells (middle panel, No contact) or (iii) cultured with the conditioned medium of donor neurons (bottom panel, CM). In red, α -synuclein fibrils; in white, acceptor neurons; and in blue, nuclei. Scale bars represent 10 μ m. Yellow arrows point to ATTO-550 α -synuclein puncta detected in the CM condition.

C The bar graph shows the percentage of acceptor cells containing α -synuclein puncta from images such as those presented in (B). **** $P < 0.001$ compared to the co-culture condition by two-tailed Student's *t*-test. Data are shown as mean \pm s.e.m. from three independent experiments (where at least 80 cells were counted per experiment).

D The box plot shows the number of α -synuclein puncta in acceptor neurons in co-culture (white), in CM (grey) and in the No contact condition (light grey). Boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 80 cells scored for each independent experiment. **** $P < 0.001$ compared to the co-culture condition by two-tailed Mann-Whitney *U*-test ($n = 3$ independent experiments).

E Western blot analysis of α -synuclein indicates the presence of some α -synuclein in the CM of neurons loaded with the fibrils after 72 h compared to control medium and to lysate of cells directly exposed to the fibrils. α -tubulin was used as a loading control.

α -synuclein fibrils efficiently transfer between neurons and CAD cells

Because of the lack of a specific TNT marker, identification of TNTs in primary neurons is challenging. To determine whether

TNTs are involved in the transfer of α -synuclein from neurons to recipient cells, we co-cultured acceptor H2B-GFP-transfected CAD cells with mature donor neurons, pre-exposed overnight to fluorescent α -synuclein fibrils. The proportion of the recipient cells containing α -synuclein puncta was 100% after 24 h of co-culture

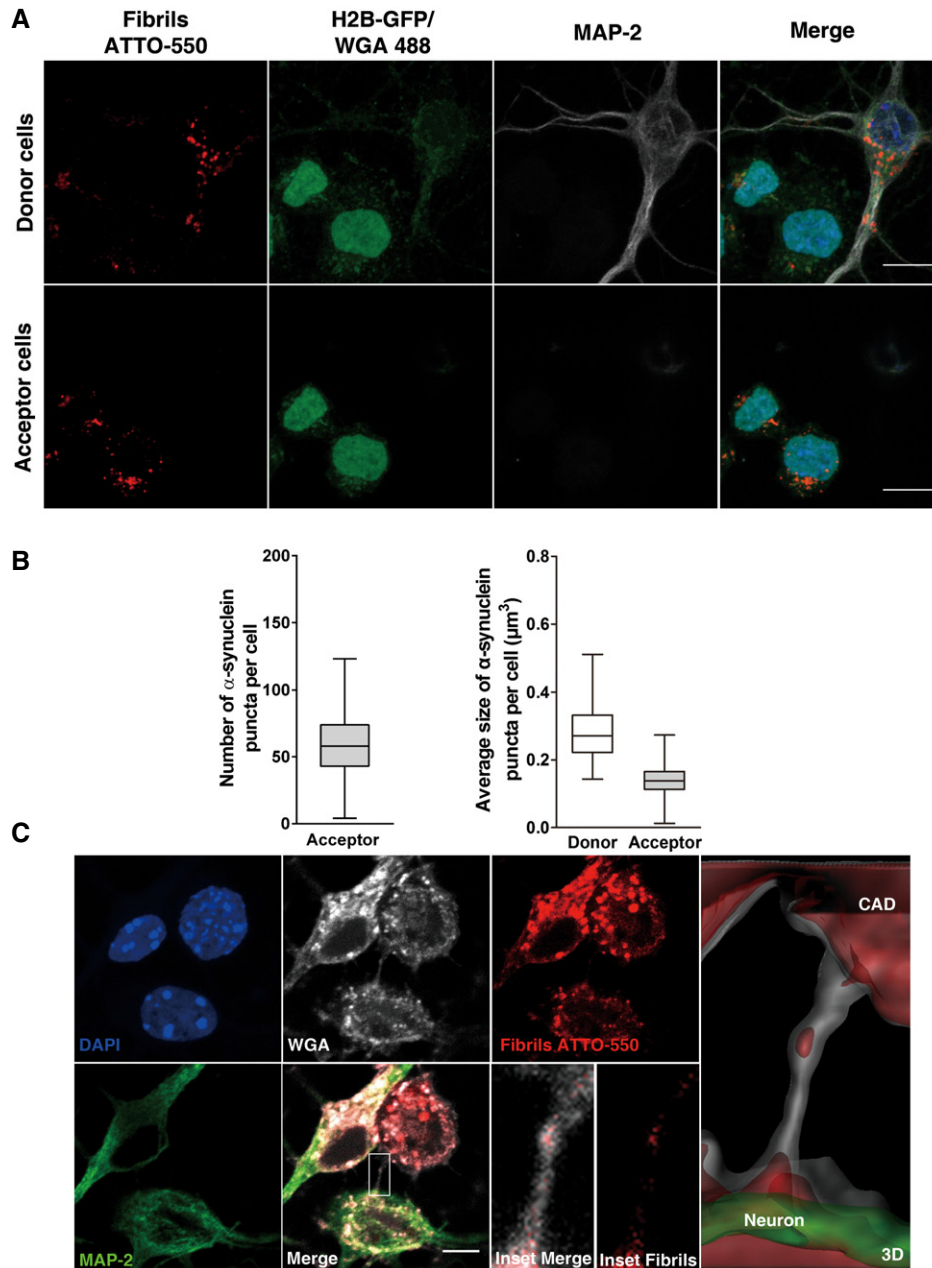


Figure 8. Transfer of α -synuclein fibrils between neurons and neuron-like CAD cells.

- A** Representative images of donor (upper panel) and acceptor cells (bottom panel) after 24 h of co-culture. Donor neurons were loaded with α -synuclein fibrils prior to co-culture with H2B-GFP acceptor CAD cells. In red, α -synuclein puncta; in green, acceptor CAD cells; in white, donor neurons; and in blue, nuclei. The images of donor cells are representative Z-stack projections of the lower slices where these cells are located, and the images corresponding to acceptor CAD cells are Z-stack projections of the upper slices, covering the whole cell body. Scale bars represent 10 μm .
- B** Box plots showing the number (left side) and average size (right side) of α -synuclein puncta in acceptor CAD cells after 24 h of co-culture with donor neurons. ($n=3$ independent experiments). In the box and whisker plots, boxes extend from the first to the third quartile, the line inside the boxes shows the median and the whiskers represent the min/max value of at least 80 cells scored for each independent experiment.
- C** Representative images showing a TNT detected between CAD cells and neurons after 24 h of co-culture. In green, MAP-2; in white, WGA; in red, α -synuclein puncta; and in blue, nuclei. The merged image in the bottom panel shows in detail the TNT connecting the cells. The insets and the 3D reconstruction of the neuron (green) and the CAD cell (white) show the presence of α -synuclein puncta inside the TNT. Scale bar represents 5 μm .

(Fig 8A). Of interest, the number (the median value is 58) and average size (the median value is 0.14 μm^3) of α -synuclein foci were very similar to the number and size found in the acceptor

cells upon cell-to-cell-mediated transfer of fibrillar α -synuclein between CAD cells (compare Fig 8B to Fig 1D and E). Most importantly, under these conditions we could identify TNTs

extending between CAD cells and the soma and neurites of primary cortical neurons and in some cases TNTs containing α -synuclein puncta (Fig 8C). We conclude from these observations that TNT-mediated cell-to-cell contacts contribute to the transfer of fibrillar α -synuclein.

Discussion

While the transfer of α -synuclein between co-cultured cells *in vitro* has been convincingly demonstrated, the nature of the transferred α -synuclein and the mechanism of transfer have not been defined (Desplats *et al*, 2009; Danzer *et al*, 2011; Hansen *et al*, 2011; Freundt *et al*, 2012; Bae *et al*, 2014). Characterizing both these issues is critical for designing strategies aimed at interfering with the propagation component of the pathology. To address this, we assessed the intercellular transfer of well-characterized recombinant human fluorescent α -synuclein fibrils of defined structure and molecular mass proven to induce Lewy body-like inclusion in rat (Peelaerts *et al*, 2015), in co-culture of CAD cells and primary neurons. To our knowledge, our study is the first evidencing high levels of transfer in neuronal-like cells and in primary neurons (100% of acceptor cells compared to on average 2–10% for equal or longer duration of co-culture) (Desplats *et al*, 2009; Hansen *et al*, 2011; Bae *et al*, 2014). This may be attributed to the nature of α -synuclein fibrils used in our study as opposed to polymorphic preparations and overexpressed α -synuclein (which does not appear to form disease-associated fibrillar aggregates) used in previous studies (Desplats *et al*, 2009; Hansen *et al*, 2011; Bae *et al*, 2014). In addition using confocal microscopy combined with a dedicated plugin for quantitative image analysis, we could document both the number and size of transferred α -synuclein fibrils, which has been proven very challenging using manual methods (Desplats *et al*, 2009; Danzer *et al*, 2011; Hansen *et al*, 2011; Kondo *et al*, 2011; Freundt *et al*, 2012; Konno *et al*, 2012; Lee *et al*, 2013; Bae *et al*, 2014).

We demonstrate that cell-to-cell contacts contribute to efficient transfer of α -synuclein fibrils in CAD cells and primary neurons. This is in agreement with a recent elegant *in vivo* study showing that in mice the intercellular transfer of α -synuclein from the olfactory bulb to other brain regions was likely not due to passive diffusion through cerebrospinal fluid or brain parenchyma, but rather relied on cell-to-cell contacts (Rey *et al*, 2013). Impairing cell-to-cell contact in our model systems (both in CAD cells and in primary cortical neurons) reduced the transfer of α -synuclein fibrils. Transfer through conditioned media was not as efficient as that through donor cells physically contacting recipient cells. Previous studies where the conditioned medium was concentrated (200X) suggested secretion as a major path for α -synuclein transfer (Desplats *et al*, 2009; Hansen *et al*, 2011; Bae *et al*, 2014). While secretion in the medium contributes to propagation of α -synuclein fibrils, physical contacts between cells significantly increased transfer. This means that either physical contacts between cells or tight proximity is essential for efficient transfer of α -synuclein fibrils. It is also worth noting that studies *in vivo* have demonstrated lower levels of the secreted α -synuclein protein in the cerebrospinal fluid in PD subjects compared to control subjects (Tokuda *et al*, 2006).

Uptake of preformed α -synuclein fibrils has been shown to be partially dependent on dynamin (Hansen *et al*, 2011; Konno *et al*, 2012; Reyes *et al*, 2014). To assess the contribution of dynamin-dependent processes on fibrillar α -synuclein transfer, we overexpressed the dominant-negative form of dynamin-1 in the acceptor CAD cells. We confirmed that uptake of exogenous fibrils is partially dynamin-1 dependent. However, the overexpression of dominant-negative forms of dynamin-1 did not affect the cell-to-cell transfer of α -synuclein fibrils. Previous studies employed donor cells overexpressing α -synuclein, which does not appear to form aggregates and was reported to be secreted. Significantly, we found that in acceptor cells taken up α -synuclein fibrils co-localized little with fibrils transferred from donor cells, which suggest that fibrils may use different pathways to be internalized or to transfer between neuron-like cells in our co-culture conditions.

By analysing three dimensionally reconstructed confocal images, we observed α -synuclein fibrils in the lumen of TNT-connecting donor and acceptor CAD cells. Importantly, the transfer of fibrils was greatly increased when TNT formation was enhanced by overexpression of the TNT inducer myosin-10 (Gousset *et al*, 2013), while it decreased when TNT formation was impaired (i.e. in sparse conditions or using physical barriers). This suggests that TNTs contribute to intercellular transfer of α -synuclein fibrils. Interestingly, we found that α -synuclein fibrils enhance TNT formation. The mechanism for this increase needs to be further studied; however, we report a sustained increase in ROS, which could account for this observation. We have previously reported that oxidative stress (hydrogen peroxide) induces TNT formation in CAD cells (Gousset *et al*, 2013). Thus, α -synuclein fibrils may induce TNT formation through ROS and hijack this process to spread efficiently. In support of this hypothesis, Wang and colleagues have shown that rat astrocytes, subjected to oxidative stress, form TNTs to selectively connect with naïve unstressed cells (Wang *et al*, 2011). We propose that this should favour the oriented propagation of α -synuclein aggregates from stressed, aggregate-containing cells, to healthy counterparts, and could explain (in part) the topographical and stereotypical spreading of α -synuclein inclusions described in the Braak staging (Braak *et al*, 2003b). Consistent with this view, myosin-10 both enhances the unidirectional transfer of vesicles through TNTs (Gousset *et al*, 2013) and increases the number of fibrils transferred in acceptor cells. These observations suggest that intercellular transfer of α -synuclein fibrils through TNTs is mainly unidirectional (from donor stressed cells to unstressed acceptor cells).

How α -synuclein fibrils are transported in TNTs and to what compartment are they directed to after they reach the acceptor cells are important questions. Endogenous α -synuclein aggregates are targeted to lysosomes both *in vivo* and in cell models (Cuervo *et al*, 2004; Spencer *et al*, 2009; Mak *et al*, 2010), suggesting an important role of the autophagic and lysosomal pathways in the degradation of misfolded proteinaceous aggregates in neurodegenerative diseases (Zhang *et al*, 2009; Nixon, 2013). We found the majority of preformed α -synuclein fibrils inside lysosomal vesicles in donor cells (Sung *et al*, 2001; Lee *et al*, 2008; Liu *et al*, 2009; Konno *et al*, 2012). We further discovered that fibrils are transported to acceptor cells in the lumen of TNTs within lysosomal vesicles. The finding that α -synuclein fibrils traffic inside lysosomes in TNTs is reminiscent of other studies reporting a TNT-dependent mechanism of lysosome transfer in both healthy and pathological conditions (Rustom

et al, 2004; Gousset et al, 2009; Yasuda et al, 2011; Naphade et al, 2015). Of specific interest, in the context of a lysosomal disorder, Naphade and colleagues have shown that damaged lysosomes are transferred from diseased fibroblasts towards healthy macrophages (Naphade et al, 2015). Interestingly, α -synuclein aggregation has been shown to cause lysosomal dysfunction *in vivo* and *in vitro* (Chu et al, 2009; Dehay et al, 2010; Alvarez-Erviti et al, 2011). This appears to be very relevant for PD pathology, which is associated with lysosomal dysfunctions (Bourdenx et al, 2014). In our cell

models (CAD cells and primary neurons), α -synuclein fibrils accumulate in lysosomal vesicles. We further observed an increase in the lysosome size similar to what was reported in α -synuclein-induced lysosomal dysfunction (Tanik et al, 2013). Based on these findings, we formulate the hypothesis that donor cells overloaded with damaged lysosomes containing α -synuclein fibrils dispose of this material by transferring it in a “targeted way” to healthy cells by hijacking the trafficking of TNT-mediated lysosomal vesicles (see model, Fig 9). This hypothesis is fully supported by and explains

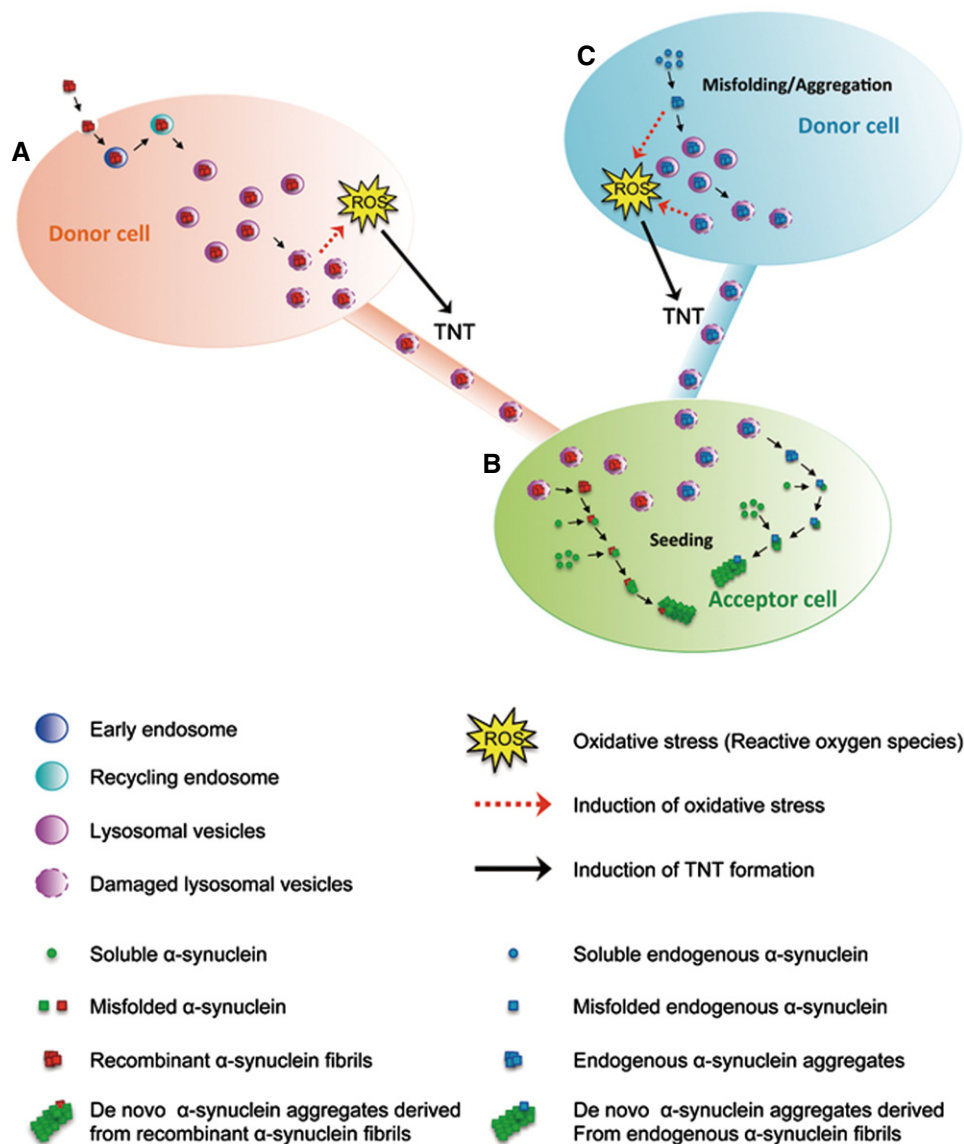


Figure 9. Model for TNT-mediated intercellular transfer of fibrillar α -synuclein.

A–C Once internalized in donor cells (A), recombinant human α -synuclein fibrils are mainly targeted to lysosomal vesicles (and found to a lesser extent in early and recycling endosomes). Lysosomal vesicles containing α -synuclein fibrils transfer from donor to acceptor cells (B) inside TNTs. Once in the acceptor cells, α -synuclein fibrils induce the formation of *de novo* α -synuclein aggregates (seeding). We propose that (A) α -synuclein fibrils accumulated inside lysosomes would impair their function leading to cellular stress (e.g. oxidative stress). In turn, this would induce TNT formation allowing spread to (B) healthy cells in order to lower the burden of α -synuclein fibrils inside lysosomes. α -synuclein fibrils would then escape lysosomes possibly by inducing LMP. Once free in the cytoplasm, they would recruit soluble α -synuclein and induce *de novo* formation of α -synuclein aggregates. As a consequence, α -synuclein fibrils would propagate from stressed to unstressed cells through TNTs contributing to the intercellular spreading of the pathology. Because endogenous α -synuclein when aggregated is targeted to lysosomes *in vivo*, we propose a similar mechanism occurring for endogenously formed aggregates (C).

the evidence in the literature showing that lysosomal impairment increases the transfer of α -synuclein between cells (Bae *et al*, 2014). TNTs may also be involved in the oriented transfer of aggregated α -synuclein from neurons to astrocytes. The latter have been indeed shown to take up and concentrate very efficiently fibrillar α -synuclein (Reyes *et al*, 2014).

In order to propagate the pathology, α -synuclein fibrils have to amplify after transfer by seeding the aggregation of their soluble counterpart. While several reports have demonstrated a seeding activity of the transferred α -synuclein (with 2–5% of inclusion formation upon co-culture of neuronal cells), none has so far identified the species of α -synuclein responsible for seeding after transfer (Desplats *et al*, 2009; Lee *et al*, 2010; Hansen *et al*, 2011; Angot *et al*, 2012; Bae *et al*, 2014). As the majority of α -synuclein fibrils are confined within lysosomes, how α -synuclein fibrils seed the aggregation of their cytosolic counterpart becomes intriguing. α -synuclein assemblies have been previously shown to induce the rupture of lysosomes *in vitro* (Freeman *et al*, 2013) and lysosomal membrane permeation (LMP) has been reported in a mouse model of PD (Dehay *et al*, 2010). The α -synuclein fibrils used in this study were shown to permeabilize membranes (Pieri *et al*, 2012). Based on these evidences, we propose that seeding may be the consequence of lysosomal rupture and release of α -synuclein fibrils into the cytosol (see model, Fig 9). Further studies are required to determine whether this occurs or how α -synuclein fibrils escape the lysosomal vesicles during or after TNT-mediated transfer.

We demonstrated here that TNTs contribute significantly to intercellular α -synuclein fibril transfer and evidenced a specific role of lysosomes in this process. This represents a major breakthrough in understanding the mechanisms underlying the progression of synucleinopathies. Furthermore, our work has revealed a novel role of lysosomes for intercellular spreading of α -synuclein aggregates and calls for a re-evaluation of lysosome function in neurodegenerative diseases. Similar to α -synuclein fibrils, huntingtin (Costanzo *et al*, 2013), PrP^{Sc} (Gousset & Zurzolo, 2009; Gousset *et al*, 2009; Zhu *et al*, 2015) and TDP-43 (Ding *et al*, 2015) aggregates were shown to induce TNT formation and to propagate from cell to cell through TNTs. Our compelling findings together with previous reports allow us to propose a general role for TNTs in the propagation of prion-like proteins in neurodegenerative diseases (Gousset *et al*, 2009; Wang *et al*, 2011; Costanzo *et al*, 2013; Ding *et al*, 2015). The discovery of specific TNT markers is paramount to investigate the involvement of TNT-mediated transfer *in vivo*. Such studies could open new avenues for the discovery of therapeutic targets and the development of innovative drugs to prevent the progression of neurodegenerative diseases.

Materials and Methods

Cell lines, reagents, plasmids and transfection

Mouse neuron-like CAD cells were a gift from Hubert Laude (Institut National de la Recherche Agronomique, Jouy-en-Josas, France), and they were cultured in Opti-MEM (Invitrogen) and 10% foetal bovine serum (FBS). Transient transfections were performed with Lipofectamine 2000[®] (Invitrogen) in accordance with the manufacturer's

instructions. The plasmids used were GFP vector (EGFP-C1), GFP-dynamin-1 K44A that was a gift from Mark A. McNiven (Mayo Clinic, Rochester, USA), GFP-myosin-10 that was a gift from Richard E. Cheney (University of North Carolina, Chapel Hill, USA) and ChFP- α -synuclein as previously described (Bousset *et al*, 2013). Transfected cells were selected and maintained in the same culture medium as untransfected CAD cells containing 500 μ g/ml of G 418 disulfate salt (Sigma Aldrich) during at least 3 weeks, before fluorescent cell sorting according to their ChFP expression levels, using FACS Aria III (BD Biosciences), with an excitation laser at 561 nm and speed of sorting ranging from 3,000 to 4,000 events per second.

Preparation of α -synuclein fibrils

The expression and purification of human wild-type α -synuclein was performed as previously described (Chee *et al*, 2005) and detailed in the Appendix.

Flow cytometry measurement of time course of internalization

CAD cells were plated in 24-well plates at a density that would allow sub-confluency to be reached after 10 h. Then, cells were treated with 1 μ M of ATTO-550 fluorescent-tagged human recombinant α -synuclein fibrils. Fibrils were sonicated prior to internalization for 5 min at 80% amplitude with a pulse cycle of 5 s on and 2 s off. Internalization was followed for time points of 5, 15 min, 1, 3, 6 and 16 h. To remove plasma membrane-bound assemblies, cells were washed three times with 0.1% trypsin (Gibco) and were detached by pipetting and then passed through sterile 40- μ m nylon cell strainers (BD Falcon[™]) in order to obtain single-cell suspensions. Cells were fixed using 4% PFA. The percentage of ATTO-550-positive cells at each time point was scored using BD LSRFortessa Cell Analyzer (BD Biosciences).

Co-culture system in neuron-like CAD cells

Acceptor cells were plated in T25 flasks and transiently transfected the day after with GFP vector or GFP-dynamin-1 K44A. The same day of transfection, donor cells were plated in 35-mm dishes at a density that would allow sub-confluency to be reached after 10 h. The donor cells were then treated with sonicated α -synuclein fibrils as described above for 16 h. The day after, donor cells were detached and mixed with acceptor cells at the ratio of 1:1 and plated on μ -Dish^{35 mm, high} Ibidi (Biovalley, France) at a confluency that allows TNT formation (sub-confluent cells) or impairs TNT formation (sparse cells). After 24-h incubation, cells were briefly washed with PBS, fixed with PFA 4%, stained with DAPI (1/10,000) and mounted with Aqua-Poly/Mount (Polysciences, Inc.). Images were acquired on an LSM 700 confocal microscope (Zeiss) with a 40 \times objective by taking stacks covering the whole cellular volume. The number of acceptor cells containing α -synuclein fibrils (referred as percentage of transfer) and the number and the volume (referred as size) of fibrils per acceptor cells were assessed using a script developed by Fabrice de Chaumont for use in the ICY software. This script allows the automated detection and quantification of number and size of fibrils within donor and acceptor cells (described in image analysis).

Filter and CM controls

In order to co-culture donor and acceptor cells in conditions that allow their physical separation, GFP-transfected acceptor cells were plated on poly-D-lysine-coated coverslips at the bottom of 24-well plates. An equal number of donor cells containing sonicated α -synuclein fibrils were plated in 6.5-mm transwell filters with 0.4- μ m pore polyester membrane insert (Sigma Aldrich) which were then placed within the 24-well plates. The cells therefore shared only the culture medium but were unable to physically contact each other. After 24-h co-culture, the filter was removed and the acceptor cells on the coverslip were fixed, stained and imaged as described above. For the conditioned medium control, donor cells containing sonicated α -synuclein fibrils and GFP-transfected acceptor cells were plated separately on μ -Dish^{35 mm, high} Ibidi for 24 h. The next day, the medium of acceptor cells was removed and replaced by the complete untransformed 24-h conditioned medium of donor cells (i.e. not diluted, concentrated or centrifuged). After 24 h, cells were fixed, stained and imaged and pictures were analysed as described above.

LysoTracker-positive vesicle transfer

To investigate whether α -synuclein transfer from donor to acceptor cells inside lysosomal vesicles derived exclusively from donor cells, donor cells containing sonicated α -synuclein fibrils and previously trypsin-washed were incubated with LysoTracker Deep Red (Life Technologies) for 30 min at 1:100 dilution. After extensive washes with PBS, donor cells were cultured with GFP-transfected acceptor cells at 1:1 ratio. After 24-h co-culture, cells were fixed, stained and imaged as described above. After detection of α -synuclein fibrils and LysoTracker-positive vesicles in both the donor and acceptor cells, the percentage of co-localization between α -synuclein fibrils and LysoTracker-positive vesicles was quantified in both cell populations using a co-localization script developed by Fabrice de Chaumont (described in the Appendix).

Exogenous α -synuclein fibril internalization in co-cultured cells

To get insight into the mechanism of α -synuclein internalization and transfer, donor cells containing sonicated α -synuclein fibrils ATTO-550 were co-cultured with untransfected acceptor cells as described above with the exception that after 12 h of co-culture plating, 0.3 μ M of sonicated α -synuclein fibrils Alexa-488 was added in the co-culture for an additional 12 h. After a total of 24-h co-culture, cells were trypsin-washed, fixed, stained and imaged. After detection of ATTO-550 and Alexa-488 α -synuclein fibrils in the donor and acceptor cells, the percentage of co-localization of α -synuclein fibrils Alexa-488 with ATTO-550 α -synuclein fibrils was quantified in both cell populations using a co-localization script developed by Fabrice de Chaumont (described in the Appendix).

Co-culture with CAD cells overexpressing soluble α -synuclein

Donor cells were loaded with 1 μ M of sonicated Alexa-488 α -synuclein fibrils for 16 h as described above and then co-cultured sub-confluently with acceptor ChFP- α -synuclein cells for 72 h. A simultaneous control was performed, wherein ChFP- α -synuclein cells were cultured separately for 72 h to determine whether spontaneous aggregation of the ChFP- α -synuclein occurred in this time frame. Cells were then fixed, stained and imaged as described above. Automated detection and quantification of the number of

ChFP- α -synuclein puncta was assessed by using the ICY script. After detection of transferred Alexa-488 α -synuclein fibrils in the acceptor cells, the percentage of co-localization of transferred α -synuclein fibrils and ChFP- α -synuclein puncta was quantified using the co-localization script previously mentioned.

TNTs imaging

To image TNTs within the donor cell population, cells were treated with 1 μ M of ATTO-550 sonicated α -synuclein fibrils for 16 h. Then, cells were several times gently washed with PBS. To preserve TNTs, cells were first fixed with fixative solution 1 for 20 min (2% PFA, 0.05% glutaraldehyde and 0.2 M HEPES in PBS) and then with fixative solution 2 for another 20 min (4% PFA and 0.2 M HEPES in PBS). Cells were then washed with PBS and incubated for 20 min with WGA Alexa-488 (Life Technologies) and mounted. Cells were imaged using LSM700 confocal microscope and serial Z-stacks of 0.25 nm were acquired to image the whole cell volume. Three-dimensional (3D) reconstruction was performed using the ICY software.

To image TNTs between donor and acceptor cells, cells were co-cultured and then fixed as described above. In addition, cells were also stained with WGA-350 (Life Technologies) for 30 min. Epifluorescence microscope was used to image the whole cell volume using Z-stacks of 0.25 nm distance (Zeiss Axiovert 200M controlled by Axiovision software). Deconvolution and 3D reconstruction were obtained by using the Huygens Professional software and Huygens Essential software, respectively.

TNT quantification experiments

To determine whether internalized α -synuclein fibrils affected TNT number, CAD cells were plated in culture inserts in μ -Dish^{35 mm, high} Ibidi (Biovalley, France). After 10 h, sub-confluent cells were treated with 1 μ M of ATTO-550-labelled sonicated α -synuclein fibrils. After 16 h, cells were fixed, stained with WGA Alexa-488 and mounted as described above. After imaging using fluorescence microscope as described above, the TNT structures connecting remote cells and not touching the substratum were manually counted for both α -synuclein fibril-loaded cells and the unloaded control cells. TNT quantification was also performed in cells untransfected and GFP-myosin-10-transfected cells. Both cell populations were separately loaded with α -synuclein fibrils for 16 h and then plated in μ -Dish^{35 mm, high} Ibidi (Biovalley, France) for an additional 16 h. Cells were fixed, stained with WGA Alexa-488 and mounted. TNT quantification was performed as described above.

After 24-h co-culture (see co-culture system described above), cells were fixed, stained with WGA Alexa-633 and mounted as described above. Images were similarly acquired and the percentage of TNT-connecting donor and acceptor cells was quantified in sub-confluent and sparse conditions.

Statistical analysis

The statistical relevance of the bar graphs was obtained by calculating the *P*-value using the paired two-tailed Student's *t*-test. The bar graphs showed in the figures are presented as mean \pm s.e.m. The statistical relevance of the box-and-whisker plots was calculated

with the Mann–Whitney *U*-test. All column graphs, plots and statistical analyses were done using GraphPad Prism version 5 software.

Expanded View for this article is available online.

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Author contributions

CZ conceived the project and discussed the planning and execution of the experiments. SA planned and performed the majority of the experiments. FL prepared and performed all experiments in primary neurons. SZ established stable CAD cell lines and performed some experiments and quantifications. LP performed trap assay. LB generated α -synuclein fibrils, established stable CAD cell lines and performed quantitative assessments for trap assay. FC devised the script for spot detection and co-localization of the ICY software. JCOM and RM discussed the project with CZ and directed the research of FC and LB. SA, RM and CZ wrote the paper. All authors contributed to revising the manuscript.

Conflict of interest

The authors declare that they have no conflict of interest.

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