## Two New Aromatic Compounds from *Hericium erinaceum* (Bull.: Fr.) PERS.<sup>1)</sup>

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Two new aromatic compounds, erinacerins A (1) and B (2), were isolated from the fruiting bodies of *Hericium erinaceum* (BULL: FR.) PERS. (Hericiaceae) together with a known compound, hericenone A (3). The structures of the new compounds were elucidated on the basis of their spectral data. It was found that 1 occurred as a racemate.

Key words Hericium erinaceum; Hericiaceae; aromatic compound; mushroom

The fruiting bodies of *Hericium erinaceum* (BULL: FR.) PERS. (Yamabushitake in Japanese, Hericiaceae) are known as an edible mushroom. The constituents of H. erinaceum have been previously investigated and shown to contain aromatic compounds,<sup>2-5)</sup> fatty acids,<sup>6)</sup> sterols<sup>7)</sup> and polysaccharides.<sup>8)</sup> Some aromatic compounds, including hericenone A (3), were found to have significant cytotoxicity against HeLa cells<sup>2)</sup> and stimulating activity of the synthesis of the nerve growth factor.<sup>3)</sup> In a continuation of our investigation of chemical constituents from mushrooms, we describe here the isolation and structural elucidation of two new aromatic compounds, erinacerins A (1) and B (2), together with a known compound (3) from the fruiting bodies of *H. eri*naceum. Compound 3 was identified as hericenone A by comparison of its spectroscopic data with those previously described in the literature.<sup>2,4)</sup>

Compound 1, called erinacerin A, was obtained as a colorless oil. The molecular formula was determined to be  $C_{27}H_{31}NO_4$  by high-resolution (HR)-electron ionization (EI)-MS. The IR spectrum showed the presence of a  $\gamma$ -lactam (1705 cm<sup>-1</sup>), an  $\alpha,\beta$ -unsaturated ketone (1680 cm<sup>-1</sup>) and phenyl groups (1614 cm<sup>-1</sup>). The <sup>1</sup>H-NMR spectrum (*vide* Experimental) showed signals due to a tertiary methyl group  $[\delta_H 1.41 (3H, CH_3-9')]$ , two olefinic methyl groups  $[\delta_H 1.85 (3H, CH_3-8'), 2.14 (3H, CH_3-10')]$ , six methylenes  $[\delta_H 1.90 (1H, H-2'a), 2.02 (1H, H-2'b), 2.64 (1H, H-4'a), 2.68 (1H, H-1'a), 2.69 (1H, H-1'b), 2.76 (1H, H-4'b), 2.98 (2H, H_2-2''), 3.83 (1H, H-1''a), 3.84 (1H, H-1''b), 4.11 (1H, H-3a), 4.17 (1H, H-3b)], a methoxyl group <math>[\delta_H 3.87 (3H)]$ , a trisubsti-

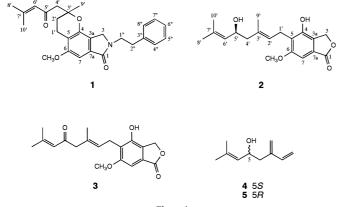


Chart 1

tuted double bond [ $\delta_{\rm H}$  6.05 (1H, H-6')] and six aromatic protons [ $\delta_{\rm H}$  6.90 (1H, H-7), 7.20–7.32 (5H, H-4"–H-8")]. The <sup>13</sup>C-NMR spectrum (vide Experimental), obtained with the aid of a distortionless enhancement by polarization transfer (DEPT) spectrum, showed characteristic signals appearing to be due to a carbonyl [ $\delta_{\rm C}$  198.1 (C-5')], a carbonyl of  $\gamma$ -lactam  $[\delta_{\rm C} 168.9 \text{ (C-1)}]$ ,<sup>9)</sup> two methylene carbons linked to  $\gamma$ lactam nitrogen [ $\delta_{\rm C}$  44.4 (C-1"), 48.0 (C-3)]<sup>9)</sup> and an oxygenated quaternary carbon [ $\delta_{\rm C}$  75.9 (C-3')]. There were thirteen degrees of unsaturation in the molecule according to the molecular formula. Two benzene rings, a  $\gamma$ -lactam and an  $\alpha,\beta$ -unsaturated ketone accounted for twelve of those. One degree of unsaturation remained, implying that there was one more ring in this structure. The <sup>1</sup>H–<sup>1</sup>H shift correlation spectroscopy (<sup>1</sup>H–<sup>1</sup>H COSY) spectrum of **1** implied connectivities for H<sub>2</sub>-1'-H<sub>2</sub>-2' and H<sub>2</sub>-1"-H<sub>2</sub>-2". Interpretation of the <sup>1</sup>H-detected heteronuclear multiple-bond coherence (HMBC) spectrum revealed correlations from H<sub>2</sub>-3 to C-1, C-3a and C-7a; H-7 to C-1 and C-6; H<sub>2</sub>-1' to C-4, C-5 and C-6; H-4' and H-6' to C-5'; CH<sub>3</sub>-8' and CH<sub>3</sub>-10' to C-6'; CH<sub>3</sub>-9' to C-2', C-3' and C-4'; H<sub>2</sub>-1" to C-3; H<sub>2</sub>-2" to C-4" and C-8"; and CH<sub>3</sub>O-6 to C-6. As a result, C-3' carbon is linked to an oxygen atom at C-4 to establish a dihydropyran ring. Therefore, the structure of 1 was deduced to be as shown in Fig. 1. Despite the presence of an asymmetric carbon at C-3', the specific rotation of 1 was almost zero, suggesting its racemic nature. This was proved by chiral reverse-phase HPLC, in which 1 was separated into two peaks in a ratio of 1:1 [peak A, (-)-1:  $[\alpha]_D^{25}$  -16.8° (MeOH); peak B, (+)-1:  $[\alpha]_D^{26}$  $+16.1^{\circ}$  (MeOH)], both of which gave <sup>1</sup>H-NMR spectra identical to those of the starting material. The circular dichroism (CD) spectra of (+)-1 and (-)-1 exhibited mirror images [(+)-1: 261.5 nm ( $\Delta \varepsilon$  +4.6), 224.0 nm ( $\Delta \varepsilon$  -5.2), (-)-1: 260.5 nm ( $\Delta \varepsilon$  -5.4), 223.1 nm ( $\Delta \varepsilon$  +5.2)], confirming their enantiomeric relationship or, in other words, the racemic na-

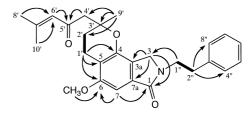


Fig. 1. <sup>1</sup>H–<sup>1</sup>H COSY (Bold Line) and HMBC (Full-Line Arrows) Correlations for **1** 

ture of our initial preparation.

Compound 2, called erinacerin B, was obtained as an amorphous powder,  $[\alpha]_D$  +12.7°. The molecular formula was determined to be  $C_{19}H_{24}O_5$  by HR-EI-MS [m/z 314  $(M^+-H_2O)$ ] and <sup>13</sup>C-NMR data. The IR spectrum showed the presence of hydroxyl groups  $(3383 \text{ cm}^{-1})$ , a phthalide  $(1763 \text{ cm}^{-1})$  and a benzene ring  $(1603 \text{ cm}^{-1})$ . The <sup>1</sup>H- and <sup>13</sup>C-NMR spectra of **2** were similar to those of **3**, except that the C-5' carbonyl group in 3 was replaced by a hydroxyl group  $[\delta_{\rm H} 4.54 \ ({\rm H}{-}5'), \delta_{\rm C} 66.0 \ ({\rm C}{-}5')]^{10}$  in **2**. The position of this hydroxyl group was confirmed by the <sup>1</sup>H–<sup>1</sup>H COSY spectrum, in which H-5' showed connectivity to  $H_2$ -4' and H-6'. The absolute configuration of the hydroxyl group at C-5' was determined as S by comparing the specific rotation values of **2** [[ $\alpha$ ]<sub>D</sub> +12.7° (MeOH)], (S)-(+)-ipsdienol [4,  $[\alpha]_{\rm D}$  +15.7° (MeOH)]<sup>11</sup> and (R)-(-)-ipsdienol [5,  $[\alpha]_{\rm D}$  $-15.3^{\circ}$  (MeOH)].<sup>11)</sup> On the basis of the above data, the structure of 2 was represented as shown in the formula.

## Experimental

**General Procedures** Optical rotations were determined using a JASCO DIP-360 digital polarimeter. CD spectra were measured on a JASCO J-720 spectropolarimeter. IR spectra were recorded with a Perkin-Elmer Spectrum One FT-IR spectrophotometer and UV spectra with a Beckman DU-64 spectrophotometer. <sup>1</sup>H- and <sup>13</sup>C-NMR spectra were recorded on JEOL JNM-LA 600 (600 and 150 MHz, respectively) and JEOL JNM-LA 400 (400 and 100 MHz, respectively) spectrometers. Chemical shifts are given on a  $\delta$  (ppm) scale, with tetramethylsilane as internal standard. HR-EI-MS were recorded out on a Kieselgel 60 (230—400 mesh, Merck). HPLC was carried out on a Tosoh HPLC system (pump, CCPD; detector, RI-8010).

**Fungal Material** The fresh fruiting bodies of *Hericium erinaceum* (from Sendai, Miyagi Prefecture, Japan) were purchased in a food market.

**Extraction and Isolation** The fresh fruiting bodies of *H. erinaceum* (1.3 kg) were extracted three times with Et<sub>2</sub>O at room temperature for 2 weeks. The Et<sub>2</sub>O extract (6.8 g) was chromatographed on a silica gel column using *n*-hexane–EtOAc (7:3–1:7), EtOAc, and MeOH to afford 32 fractions. Fraction 14 was purified by preparative HPLC [column, TSKgel ODS-120T (7.8 mm i.d.×30 cm, Tosoh), column temperature, 40 °C; mobile phase, MeOH–H<sub>2</sub>O (7:1); flow rate, 1.0 ml/min] to give 1 (3.6 mg) and 3 (5.0 mg). The separation of 1 into its enantiomers [(–)-1 (1.2 mg,  $t_R$  19.2 min)] was achieved by HPLC using a chiral column chromatograph [column, Chiralpak AD-RH (4.6 mm i.d.×15 cm, Daicel Chemical Industries, Ltd.); mobile phase, CH<sub>3</sub>CN–H<sub>2</sub>O (7:3); flow rate, 0.5 ml/min]. Fraction 16 was purified by preparative HPLC [column, TSKgel ODS-120T (7.8 mm i.d.×30 cm, Tosoh), column temperature, 40 °C; mobile phase, MeOH–H<sub>2</sub>O (7:1); flow rate, 1.0 ml/min] to give 2 (1.6 mg).

Erinacerin A (1): Colorless oil.  $[\alpha]_D^{21} \pm 0^\circ$  (*c*=0.4, MeOH). IR *v*<sub>max</sub> (CHCl<sub>3</sub>) cm<sup>-1</sup>: 1705, 1680, 1614. UV  $\lambda_{max}$  (MeOH) nm (log  $\varepsilon$ ): 215 (4.6), 244 (4.2), 292 (3.4). HR-EI-MS *m/z*: 433.2237 (M<sup>+</sup>, Calcd for C<sub>27</sub>H<sub>31</sub>NO<sub>4</sub>: 433.2253). <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.41 (3H, s, CH<sub>3</sub>-9'), 1.85 (3H, d, *J*=1.2 Hz, CH<sub>3</sub>-8'), 1.90 (1H, ddd, *J*=13.9, 7.1, 6.8 Hz, H-2'a), 2.02 (1H,

ddd, J=13.9, 6.6, 6.6 Hz, H-2'b), 2.14 (3H, d, J=1.2 Hz, CH<sub>3</sub>-10'), 2.64 (1H, d, J=14.1 Hz, H-4'a), 2.68 (1H, dd, J=7.1, 6.6 Hz, H-1'a), 2.69 (1H, dd, J=6.8, 6.6 Hz, H-1'b), 2.76 (1H, d, J=14.1 Hz, H-4'b), 2.98 (2H, t, J=7.6 Hz,  $H_2$ -2"), 3.83 (1H, t, J=7.6 Hz, H-1"a), 3.84 (1H, t, J=7.6 Hz, H-1"b), 3.87 (3H, s, CH<sub>3</sub>O-6), 4.11 (1H, d, J=16.8 Hz, H-3a), 4.17 (1H, d, J=16.8 Hz, H-3b), 6.05 (1H, br s, H-6'), 6.90 (1H, s, H-7), 7.20—7.32 (5H, m, H-4"—H-8"). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ : 17.4 (C-1'), 20.8 (C-10'), 24.8 (C-9'), 27.8 (C-8'), 30.6 (C-2'), 34.9 (C-2"), 44.4 (C-1"), 48.0 (C-3), 52.3 (C-4'), 55.9 (CH<sub>3</sub>O-6), 75.9 (C-3'), 96.2 (C-7), 113.5 (C-5), 121.9 (C-3a), 125.0 (C-6'), 126.5 (C-6"), 128.6, 128.7 (C-4", C-5", C-7", C-8"), 132.2 (C-7a), 138.8 (C-3"), 148.6 (C-4), 155.6 (C-7'), 158.5 (C-6), 168.9 (C-1), 198.1 (C-5').

(+)-1: Colorless oil.  $[\alpha]_D^{2\delta}$  +16.1° (*c*=0.1, MeOH). CD  $\lambda_{ext}$  (*c*=2.15×10<sup>-5</sup> M, MeOH) (nm) ( $\Delta \epsilon$ ): 261.5 (+4.6), 224.0 (-5.2).

(-)-1: Colorless oil.  $[\alpha]_D^{25} - 16.8^{\circ}$  (*c*=0.1, MeOH). CD  $\lambda_{ext}$  (*c*=2.06× 10<sup>-5</sup> M, MeOH) (nm) ( $\Delta \epsilon$ ): 260.5 (-5.4), 223.1 (+5.2).

Erinacerin B (2): Amorphous powder.  $[\alpha]_{2}^{24}$  +12.7° (*c*=0.2, MeOH). IR  $V_{max}$  (CHCl<sub>3</sub>) cm<sup>-1</sup>: 3383, 1763, 1603. UV  $\lambda_{max}$  (MeOH) nm (log  $\varepsilon$ ): 212 (4.5), 259 (4.0), 296 (3.5). HR-EI-MS *m/z*: 314.1526 (M<sup>+</sup>-H<sub>2</sub>O, Calcd for C<sub>19</sub>H<sub>22</sub>O<sub>4</sub>: 314.1518). <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$ : 1.71 (3H, d, *J*= 1.1 Hz, CH<sub>3</sub>-10'), 1.72 (3H, d, *J*=1.5 Hz, CH<sub>3</sub>-8'), 1.87 (3H, s, CH<sub>3</sub>-9'), 2.21 (1H, dd, *J*=13.6, 4.8 Hz, H-4'a), 2.26 (1H, dd, *J*=13.6, 8.8 Hz, H-4'b), 3.51 (1H, dd, *J*=16.5, 7.7 Hz, H-1'a), 3.58 (1H, dd, *J*=16.5, 6.2 Hz, H-1'b), 3.87 (3H, s, CH<sub>3</sub>O-6), 4.54 (1H, dt, *J*=4.8, 8.8 Hz, H-5'), 5.16 (1H, brd, *J*= 8.8 Hz, H-6'), 5.20 (2H, s, H<sub>2</sub>-3), 5.37 (1H, m, H-2'), 6.97 (1H, s, H-7). <sup>13</sup>C-NMR (150 MHz, CDCl<sub>3</sub>)  $\delta$ : 16.4 (C-9'), 18.2 (C-10'), 23.3 (C-1'), 25.8 (C-8'), 47.7 (C-4'), 56.2 (CH<sub>3</sub>O-6), 66.0 (C-5'), 68.0 (C-3), 98.5 (C-7), 121.1 (C-5), 123.6 (C-2'), 125.1 (C-3a), 127.4 (C-6'), 127.8 (C-7a), 136.1 (C-7'), 137.1 (C-3'), 150.4 (C-4), 159.2 (C-6), 171.7 (C-1).

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## **References and Notes**

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