

Two Panels of Plasma MicroRNAs as Non-Invasive Biomarkers for Prediction of Recurrence in Resectable NSCLC

Céline Sanfiorenzo^{1,2,3,9}, Marius I. Ilie^{1,3,4,5,9}, Amine Belaid¹, Fabrice Barlési⁶, Jérôme Mouroux^{1,3,7}, Charles-Hugo Marquette^{1,2,3}, Patrick Brest^{1,3†}, Paul Hofman^{1,3,4,5,*†}

1 Institute for Research on Cancer and Ageing in Nice IRCAN, INSERM U1081 – CNRS UMR 7284, Team 3, Nice, France, **2** University Hospital Center of Nice, Pasteur Hospital, Department of Pneumology, Nice, France, **3** University of Nice Sophia Antipolis, Faculty of Medicine, Team 3, Nice, France, **4** University Hospital Center of Nice, Pasteur Hospital, Laboratory of Clinical and Experimental Pathology, Nice, France, **5** University Hospital Center of Nice, Pasteur Hospital, Human Biobank, Nice, France, **6** Aix Marseille Université – Assistance Publique Hôpitaux de Marseille, Multidisciplinary Oncology and Therapeutic Innovations Department, Marseille, France, **7** University Hospital Center of Nice, Pasteur Hospital, Department of Thoracic Surgery, Nice, France

Abstract

The diagnosis of non-small cell lung carcinoma (NSCLC) at an early stage, as well as better prediction of outcome remains clinically challenging due to the lack of specific and robust non-invasive markers. The discovery of microRNAs (miRNAs), particularly those found in the bloodstream, has opened up new perspectives for tumor diagnosis and prognosis. The aim of our study was to determine whether expression profiles of specific miRNAs in plasma could accurately discriminate between NSCLC patients and controls, and whether they are able to predict the prognosis of resectable NSCLC patients. We therefore evaluated a series of seventeen NSCLC-related miRNAs by quantitative real-time (qRT)-PCR in plasma from 52 patients with I-IIIa stages NSCLC, 10 patients with chronic obstructive pulmonary disease (COPD) and 20-age, sex and smoking status-matched healthy individuals. We identified an eleven-plasma miRNA panel that could distinguish NSCLC patients from healthy subjects (AUC = 0.879). A six-plasma miRNA panel was able to discriminate between NSCLC patients and COPD patients (AUC = 0.944). Furthermore, we identified a three-miRNA plasma signature (high miR-155-5p, high miR-223-3p, and low miR-126-3p) that significantly associated with a higher risk for progression in adenocarcinoma patients. In addition, a three-miRNA plasma panel (high miR-20a-5p, low miR-152-3p, and low miR-199a-5p) significantly predicted survival of squamous cell carcinoma patients. In conclusion, we identified two plasma miRNA expression profiles that may be useful for predicting the outcome of patients with resectable NSCLC.

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* E-mail: hofman.p@chu-nice.fr

⁹ These authors contributed equally to this work.

[†] These authors also contributed equally to this work.

Introduction

Lung cancer, predominantly non-small cell lung cancer (NSCLC), is the leading cause of cancer-related deaths worldwide [1]. Despite subtle progress over recent years in terms of treatment strategies, the high mortality rate has not decreased significantly. NSCLC is often diagnosed at advanced stages with an overall 5-year survival less than 15% [2]. The poor prognosis of NSCLC patients is largely due to the lack of routine, validated, effective and low cost screening tools that allow detection of early-stage tumors. Developing such biomarkers is a public health imperative since diagnosis and treatment of early-stage NSCLC is associated with 60–80% survival at 5 years [1,3]. One of the major clinical determinants in NSCLC prognosis is tumor extension, roughly characterized by the pTNM stage. However, a large variability in disease outcome has been observed for a subset of patients with

similar clinical and pathological features, thus the current staging system may be insufficient to consistently predict the treatment outcome of NSCLC [4]. Therefore, prognostic assessment of the patients is essential to choose the best therapeutic strategy and may be improved by the integration of new robust prognostic biomarkers.

The impact on outcome of NSCLC of screening procedures such as chest X-rays, sputum cytology, spiral computed-tomography (CT), or a combination of these, has been evaluated in large-scale clinical trials. However, these analyses have not significantly affected overall survival (OS) and have demonstrated low sensitivity [5,6]. The search for non-invasive tumor biomarkers is rapidly expanding and investigation into circulating biomarkers is the subject of intense research. Several serum tumor markers such as the carcinoembryonic antigen or Cytokeratin-21-Fragment (CYFRA 21-1) may carry some prognostic and predictive

information in NSCLC, although their use is currently limited and the biochemical methodologies used to measure them are still labor-intensive [7,8].

One of the most exciting molecular markers in tumor diagnosis and prognosis are microRNAs (miRNAs) [9]. miRNAs are small RNA molecules (18 to 24 nt) that effect substantially the expression of multiple genes at a post-transcriptional level, via mRNA destabilization or translational repression [10]. Deregulation of miRNA expression is thought to be responsible of tumor initiation and progression [11]. MiRNAs are frequently deregulated in cancer and may act as oncogenes or tumor suppressors having regulatory functions on hundreds of downstream genes with different biologic functions [12,13]. Given the fundamental role of miRNAs in tumors and their global deregulation, miRNA profiles may provide a more accurate prediction of survival than the expression of a single-marker or expression profiles of protein-coding genes [13]. In addition, recent studies have demonstrated that specific expression profiles of circulating miRNAs could be promising blood-based non-invasive biomarkers useful for cancer detection and prognosis in different types of cancer, including NSCLC [14,15,16,17,18]. Human serum or plasma contains a large amount of intact and stable miRNAs, which can be detected with a simple assay such as quantitative real-time PCR (qRT-PCR) [19]. Therefore, the high stability of miRNAs allows for efficient identification in various clinical specimens including sputum, plasma, serum, and frozen and formalin-fixed paraffin embedded tissue samples [14,20,21,22].

The aim of our study was to: 1) select a large panel of miRNAs that have been reported to be highly deregulated in NSCLC, 2) determine whether the plasma expression profiles of these miRNAs were altered in NSCLC patients compared to healthy individuals and 3) evaluate whether the miRNA profile is able to predict the prognosis of resectable NSCLC. We evaluated a panel of seventeen miRNAs by qRT-PCR in the plasma of 52 patients with resectable NSCLC and 30 controls, 10 patients with chronic obstructive pulmonary disease (COPD) and 20-age, sex and smoking status-matched healthy individuals.

Materials and Methods

Study population

Sixty-two patients hospitalized from March 2008 to March 2010 at the Pasteur Hospital (Departments of Pulmonary Medicine, and Thoracic Surgery, CHU de Nice, France) were enrolled in this study. Among these patients, 52 patients had NSCLC and 10 had COPD. COPD patients did not have symptoms of lung cancer or other malignancies. The diagnosis of NSCLC patients was based on examination of all tumor specimens using the 7th pTNM classification and on the last histological classification of NSCLC [23]. In addition, twenty-age, -sex and -smoking status-matched healthy volunteers participated in this study. Written informed consent was obtained from participants after explaining the nature of the study, which was approved by the research ethics board of the Nice University hospital and was performed according to the guidelines of the Declaration of Helsinki. The main clinical and pathological data are summarized in Table 1. Enrollment of patients in our study was conditioned by stringent criteria such as obtained signed consent, availability of resected surgical specimens along with plasma samples, good quality RNA and minimum 18 months follow-up for surviving patients.

miRNA isolation

Peripheral blood (5 ml) was taken prior to surgery and kept in an EDTA-containing tube. The samples were centrifuged at 3000 rpm at 4°C for 10 minutes within 4 hours of collection. The plasma was collected and stored at -80°C until use. Total RNA containing small RNA was extracted from 100 µl of plasma using the miRNeasy Mini Kit (Qiagen GmbH, Hilden, Germany) according to the manufacturer's protocol. The concentration and purity of the RNA were determined with a NanoDrop 1000 (Thermo Fisher Scientific, Wilmington, DE).

Selection of control genes for quantification of plasma miRNAs

To select good candidates, we used some guidelines from Exiqon company (<http://www.exiqon.com/ls/Documents/Scientific/microRNA-serum-plasma-guidelines.pdf>) Endogenous controls such as U6, RNU19, miR-16-5p, miR-192-5p, and miR-103a-3p were analyzed in these samples to identify a small RNA expressed at a similar level in equal volume of sera from both healthy subjects and patients with cancer to serve as a normalization control. Only miR-16-5p, miR-192-5p, and miR-103a-3p were expressed at a high level in the samples of this study (median Ct<30; 100%, 100% and 85% detection, respectively) and not statistically different between the analyzed classes (*t*-test; *P*>0.05), and their levels were the least variable for the miRNAs in all samples (*SD*<0.9) (data not shown). Moreover, miR-103a-3p and miR-16-5p were used as markers of hemolysis [24].

Analysis of the miRNA expression level

Normalization of the results between patients was performed by subtracting the mean of miR-192-5p and miR-16-5p levels to all data (Δ CT) as previously described for other cancers [25,26,27]. Thus, the global mean of the relative expression of each miRNA was calculated and subtracted in order to have all miRNA centered on zero for further studies ($\Delta\Delta$ CT). For diagnosis-related analysis, the mean was based on the cohort of healthy controls Δ CT. For the prognosis-related study, the control Δ CT values were removed for the analysis.

Statistical Analysis

The statistical analyses were performed with SPSS 16.0 statistical software (SPSS Inc., Chicago, IL). Hierarchical clustering and pictures were generated using MeV (TM4 Microarray Software) [28]. The receiver-operator characteristic (ROC) curve and AUC analyses were used to determine the accuracy of each miRNA profile in a specimen with a given specificity rate and to determine the optimal cut-off point. We categorized each miRNA as high or low using the median value as the cut-off. The *chi*², Student or Mann-Whitney *U*-test tests were used to analyze the correlation between the miRNA expression levels and clinicopathological features of the patients. To assess the association of miRNA expression with disease-free survival (DFS), the Kaplan-Meier method and the log-rank test were used to compare survival times between groups. A Cox proportional hazards model was created to identify independent predictors of survival. Variables that were associated with survival with a *P*-value<0.20 in the univariate analysis were included in the multivariate Cox regression. All *P*-values shown were two sided, and a *P*-value≤0.05 was considered statistically significant.

Table 1. Clinicopathological characteristics of the 52 NSCLC patients, 10 COPD patients and 20 healthy individuals included in our study.

Variables	NSCLC patients n (%)	COPD patients n (%)	Healthy subjects n (%)
Overall	52 (100%)	10 (100%)	20 (100%)
Age (years)			
Mean ± SD	65.1 ± 11.1	68.9 ± 6.7	67.5 ± 5.3
Sex			
Male	39 (75%)	8 (80%)	14 (70%)
Female	13 (25%)	2 (20%)	6 (30%)
Smoking status			
Never smoked	8 (15%)	2 (20%)	5 (25%)
Former or current smokers	44 (85%)	8 (80%)	15 (75%)
Histological type			
Adenocarcinoma	27 (52%)	n/a	n/a
Squamous cell carcinoma	25 (48%)	n/a	n/a
pTNM stage			
IA	8 (19%)	n/a	n/a
IB	14 (33%)	n/a	n/a
IIA	5 (12%)	n/a	n/a
IIB	8 (19%)	n/a	n/a
IIIA	7 (17%)	n/a	n/a
Histologic grade			
Well	22 (42%)	n/a	n/a
Moderate	19 (37%)	n/a	n/a
Poor	11 (21%)	n/a	n/a
Adjuvant treatment	21 (40%)	n/a	n/a
Life status			
Alive	40 (77%)	10 (100%)	20 (100%)
Deceased; lung cancer	9 (17%)	n/a	n/a
Deceased; other cause	3 (6%)	n/a	n/a

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Results

Levels of expression of plasma miRNAs

Based on the literature, we selected seventeen miRNAs reported to be most frequently altered in primary NSCLC patients (Table S1). To determine whether aberrations in the specified miRNAs could be confirmed in independent plasma samples, we assessed expression of the candidate miRNAs in duplicate assays by qRT-PCR for 52 NSCLC plasma samples and 30 control samples (20 healthy subjects and 10 patients with COPD).

Thirteen (76%) miRNAs including the overexpression of miR-20a-5p, miR-25-3p, miR-155-5p, miR-191-5p, miR-223-3p, miR-296-5p, and miR-320-3p along with the underexpression of let-7f-5p, miR-24-3p, miR-126-3p, miR-145-5p, miR-152-3p, miR-199a-5p, were consistently observed in all plasma samples ($\Delta\Delta CT < 32$) (Table 2). No expression was detected for miR-96-5p, miR-129-5p, miR-373-5p, and miR-516-5p, and these miRNAs were excluded from further analyses (Table 2).

MiRNA profiles of NSCLC patients and cancer-free controls

The hierarchical clustering based *t*-test along with the ROC curves were constructed to estimate the sensitivity and specificity of

the 13-plasma miRNA panel. The accuracy was 79.4% with a sensitivity of 79% and a specificity of 71% (data not shown).

However, the ANOVA test, along with the ROC curve estimation yielded a 12-miRNA signature with improved accuracy in discriminating between cancer-free controls and NSCLC patients (Figure 1). The expression of the twelve-plasma miRNA panel including miR-155-5p, miR-20a-5p, miR-25-3p, miR-296-5p, miR-191-5p, miR-126-3p, miR-223-3p, miR-152-3p, miR-145-5p, miR-199a-5p, miR-24-3p, and let-7f-5p allowed significant discrimination between controls and NSCLC patients with an accuracy of 82.1% (95% CI: 0.792–0.850; $P < 0.001$), demonstrating a sensitivity of 85% and a specificity of 75% (Figure 1). The twelve plasma miRNAs significantly discriminated between controls and stage I NSCLC patients (AUC = 0.806; Figure 1), stage II NSCLC cases (AUC = 0.849; Figure 1) and stage III NSCLC patients (AUC = 0.878; Figure 1).

Next, we carried out pairwise group comparisons to identify miRNAs that contribute significantly to the different separations, including NSCLC versus healthy controls, NSCLC versus COPD patients, and lung adenocarcinoma or squamous cell carcinoma patients versus either healthy individuals or COPD patients. There was a clear separation of the NSCLC patients from the healthy subjects based on an 11-plasma miRNA profile with an accuracy,

Table 2. Plasma expression levels of candidate miRNAs in all sample sets.

miRNA	CT Mean	SD	Deregulation in NSCLC cases
Let-7f-5p	30.76	2.99	Down-regulated
miR-20a-5p	23.08	2.39	Up-regulated
miR-24-3p	23.93	2.97	Down-regulated
miR-25-3p	24.29	1.80	Up-regulated
miR-126-3p	24.20	2.46	Down-regulated
miR-145-5p	31.18	3.01	Down-regulated
miR-152-3p	28.41	2.75	Down-regulated
miR-155-5p	30.50	1.80	Up-regulated
miR-191-5p	25.21	2.88	Up-regulated
miR-199a-5p	30.25	3.61	Down-regulated
miR-223-3p	22.54	3.21	Up-regulated
miR-296-5p	32.16	2.09	Up-regulated
miR-320-3p	22.62	2.18	Up-regulated
miR-96-5p	undetectable	undetectable	undetectable
miR-129-5p	undetectable	undetectable	undetectable
miR-373-5p	undetectable	undetectable	undetectable
miR-516-5p	undetectable	undetectable	undetectable

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sensitivity, and specificity of 87.9%, 81.1%, and 82.9%, respectively (Figure 1). In addition, the diagnostic sensitivity of the 11-plasma miRNA signature was higher for squamous cell carcinoma (91.3%) than for adenocarcinoma cases (85.7%) ($P < 0.05$; Figure S1).

Furthermore, a small subset of only six miRNAs still separated NSCLC from COPD with an accuracy, sensitivity, and specificity of 94.4%, 90.9%, and 83.3%, respectively (Figure 1). Interestingly, adenocarcinoma and squamous cell carcinoma cases shared five miRNAs when compared to COPD patients, including miR-20a-5p, miR-152-3p, miR-145-5p, miR-199a-5p, and miR-24-3p. However, miR-191-5p identified only adenocarcinoma patients versus COPD patients, and miR-25-3p, squamous cell carcinoma cases only (Figures S1 & S2). Finally, only three plasma miRNAs were differentially expressed when comparing adenocarcinoma and squamous cell carcinoma patients. Higher plasma levels of miR-20a-5p ($P = 0.034$) and miR-25-3p ($P = 0.013$) along with lower levels of miR-191-5p ($P = 0.008$) were observed in squamous cell carcinoma (Figure S2).

Correlation between plasma miRNAs and clinicopathological features of NSCLC

We then compared the plasma levels of miRNAs with patient clinicopathological parameters. Higher plasma levels of miR-20a-5p ($P = 0.012$) and miR-25-3p ($P = 0.04$) along with decreased levels of miR-191-5p ($P = 0.023$) were observed in squamous cell carcinoma (Table S2; Figure S2). No significant association was found between the levels of miRNAs and age, sex, history of smoking, tumor grade, and pathological stage ($P > 0.05$, Table S2).

Association of plasma miRNAs with DFS of NSCLC patients

We further investigated whether the expression of plasma miRNAs correlated with DFS in our group of NSCLC patients.

The mean DFS in our study population was 46 months (95% CI, 39.4 to 52.9). In the univariate analysis, the clinical factor that significantly associated with DFS was the pTNM stage ($P < 0.0001$). The unadjusted survival analysis showed that high plasma levels of miR-155-5p ($P = 0.068$) and miR-20a-5p ($P = 0.018$) along with a low level of miR-152-3p ($P = 0.049$) were associated with poor DFS of NSCLC patients (Figure S3). The remaining miRNAs such as miR-223-3p ($P = 0.348$), miR-191-5p ($P = 0.671$), miR-320-3p ($P = 0.322$), miR-126-3p ($P = 0.131$), miR-145-5p ($P = 0.705$), miR-199a-5p ($P = 0.612$), miR-24-3p ($P = 0.364$), miR-25-3p ($P = 0.816$), miR-296-5p ($P = 0.853$), and let-7f-5p ($P = 0.964$) were not associated with survival (Figure S4).

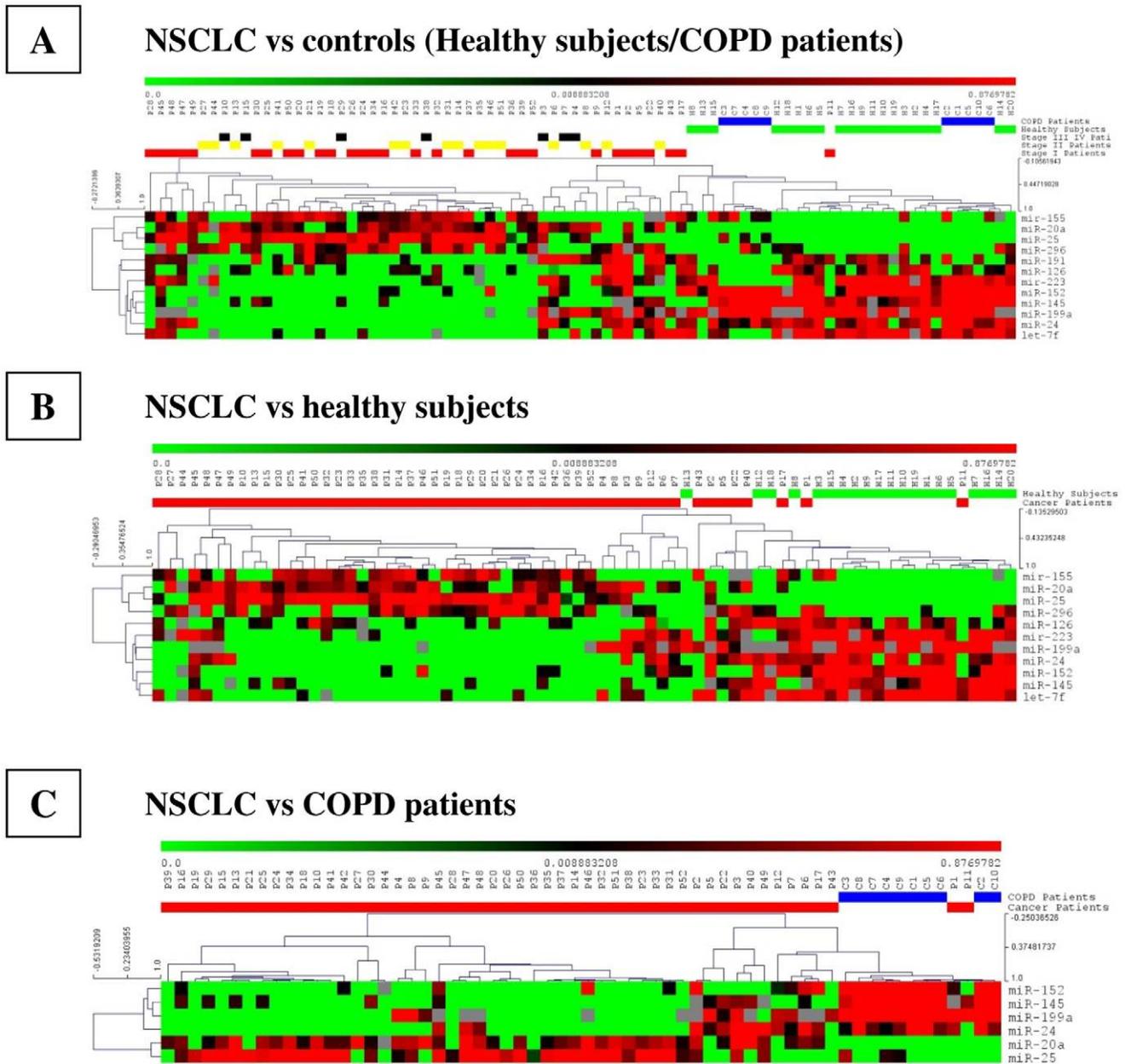
Due to the biological differences in the miRNA expression the survival analyses were conducted separately for adenocarcinoma and squamous cell carcinoma patients. Interestingly, the high plasma levels of miR-155-5p ($P = 0.008$), and miR-223-3p ($P = 0.038$) with low plasma level of miR-126-3p ($P = 0.008$; Figure 2) were significantly associated with poor DFS in lung adenocarcinoma patients. In addition, low plasma levels of miR-152-3p ($P = 0.035$) and miR-199a-5p ($P = 0.05$) along with a high plasma level of miR-20a-5p ($P = 0.001$; Figure 2) significantly correlated to decreased DFS in lung squamous cell carcinoma patients. Age, gender, history of smoking, histological subtype, and tumor grade were not associated with DFS. In a multivariate analysis, the independent factors for improved DFS were stage I ($P < 0.001$), low plasma levels of miR-155-5p ($P = 0.030$) and miR-20a-5p ($P = 0.048$) along with high plasma levels of miR-152-3p ($P = 0.029$) and miR-199a-5p ($P = 0.038$) (Table 3).

Discussion

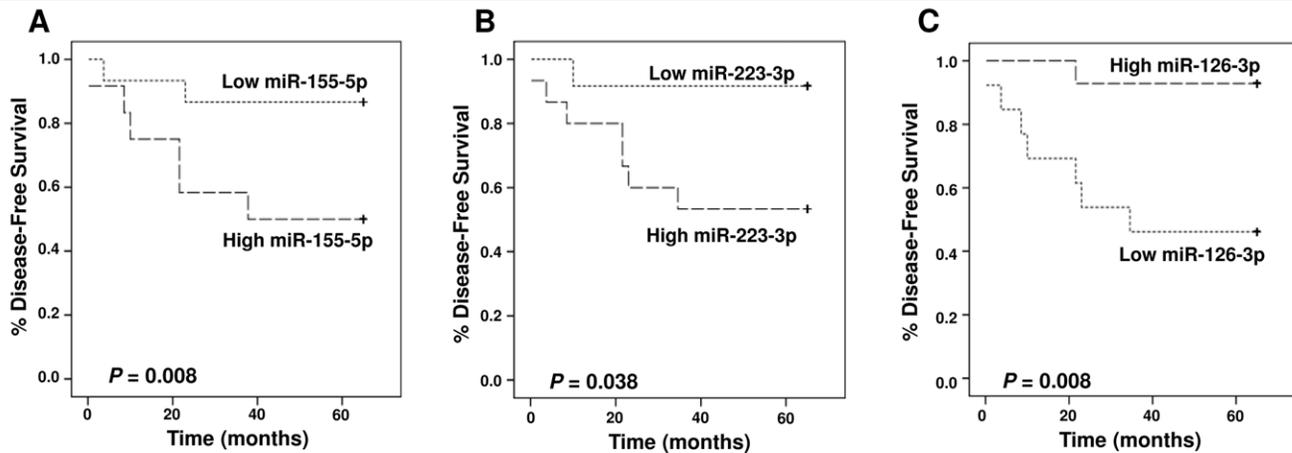
Despite recent advances in diagnosis and treatment strategies, the prognosis of NSCLC across all stages remains unchanged and early detection and prediction of outcome is critical in improving survival. However, it can be sometimes difficult to obtain tissue for diagnosis, in particular in patients with metastatic lung cancer.

Profiling of miRNA expression in lung tumor tissues discriminated cancer patients from cancer-free individuals, and specific miRNAs correlated with disease diagnosis and clinical outcome [29]. Therefore, developing minimally invasive methods by integrating the recent advances in the field of miRNAs for early diagnosis and prognosis of NSCLC is of great interest. Accumulating reports suggest that unique patterns of circulating miRNAs may act as novel biomarkers for early detection of lung cancer and for prediction of outcome [4,19,30]. Endogenous circulating miRNAs are stable and resistant to RNases [14,21]. Because of the simplicity and reproducibility of getting a blood sample, the levels of easily testable miRNAs in plasma seem suited to surveillance of NSCLC outcome [4]. However, it seems that the expression of a single miRNA may not be a reliable biomarker for cancer diagnosis and prognosis [31,32]. Simultaneous assessment of a panel of tumor-specific circulating miRNAs may improve the sensitivity and specificity for diagnosis of lung cancer and may better predict development of the cancer. Therefore, the investigation of a plasma miRNA signature in NSCLC patients using a qRT-PCR assay, as shown in this study, may be of great clinical interest as a routine procedure.

In our study, an 11-plasma miRNAs signature significantly discriminated healthy individuals from NSCLC patients. The accuracy, sensitivity, and specificity for NSCLC detection by the 11-plasma miRNA panel are 87.9%, 85% and 82.9%, respectively, which are higher to those of blood-based single biomarker, such as CYFRA 21-1 (AUC \approx 0.84, sensitivity \approx 50%, specificity \approx 95%), tissue polypeptide specific antigen (AUC \approx 0.74, sensitivity \approx 34%,



LUNG ADENOCARCINOMA



LUNG SQUAMOUS CELL CARCINOMA

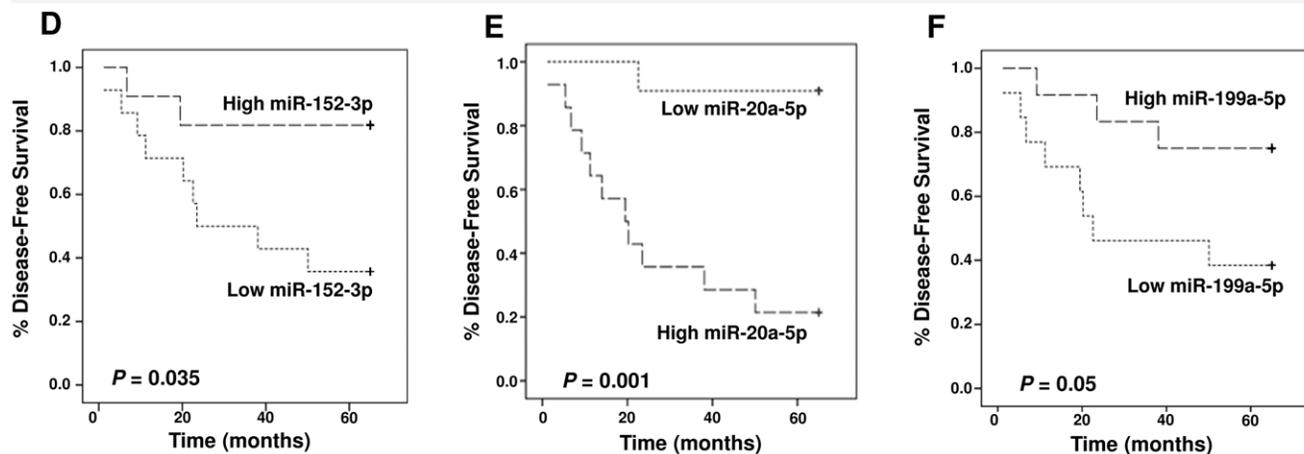


Figure 2. Kaplan-Meier DFS curves for lung adenocarcinoma patients (upper panels) and lung squamous cell carcinoma patients (lower panels) stratified according to plasma levels of miR-155-5p (A), miR-223-3p (B), miR-126-3p (C), miR-152-3p (D), miR-20a-5p (E), and miR-199a-5p (F). The *P*-values were calculated using the log-rank test between patients with high- and low-fold changes.
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in COPD patients, which has been explored as a method for early diagnosis of questionable lung densities [39,41].

Although comparison of smokers and never smokers did not demonstrate significant results in our study, possibly because of the small number of never smokers, it was interesting to note that three of the miRNA of the six-plasma miRNA panel, including miR-152-3p, miR-199a-5p and miR-20a-5p, differentially expressed between COPD patients and squamous cell carcinoma patients were able to significantly predict cancer relapse, as previously reported [16,35,36,37]. Our findings emphasize the potential role of these miRNAs as plasma biomarkers playing an important role in lung tumorigenesis and squamous cell carcinoma progression. Finally, we identified a three-plasma miRNA signature including high plasma levels of miR-155-5p and miR-223-3p with low plasma level of miR-126-3p associated with a higher risk for lung adenocarcinoma progression.

Several recent studies have proposed sets of plasma miRNAs as potential markers to monitor the development of lung cancer and its prognosis, in particular for NSCLC [4,14,17,19,30]. However the results of different studies are quite variable and often identified different miRNAs signatures. The reason for this variability is

complex, but likely arises from the differences in patients' ethnicities, sample subtypes (plasma vs. serum vs. whole blood), sample collection methods, technology platforms (microarray or qRT-PCR), as well as the bioinformatic analyses, as previously suggested [42]. We therefore carefully designed our study in order to ensure the identification of reliable miRNAs as plasma biomarkers and employed established methods that reveal valuable clinical information. We only analyzed Caucasian patients to reduce the effect of ethnicity. NSCLC patients were matched according to histology in order to minimize pathological subtype effects and to amplify the molecular homogeneity of tumor specimens. The patients were strictly selected from individuals who had not been previously treated with neoadjuvant treatment to avoid therapy bias. In addition, we demonstrated that our markers could be conveniently measured by qRT-PCR in plasma.

Interestingly, the two panels of plasma miRNAs comprising our lung squamous cell carcinoma and adenocarcinoma progression classifiers have been reported individually to have a prognostic impact in other studies.

Table 3. Multivariate analysis of prognostic factors identified in our study with DFS as the end point in patients with NSCLC.

Prognostic factor	HR ¹	95% CI ²	P-value ³
pTNM stage			
I vs. II+III	0.095	0.030–0.303	<0.001
miR-155-5p			
Low vs. high	0.060	0.005–0.767	0.030
miR-223-3p			
Low vs. high	1.449	0.581–3.614	0.426
miR-20a-5p			
Low vs. high	2.881	1.009–8.227	0.048
miR-152-3p			
High vs. low	0.333	0.125–0.892	0.029
miR-126-3p			
High vs. low	0.497	0.191–1.295	0.153
miR-199a-5p			
High vs. low	0.204	0.045–0.918	0.038

¹HR; hazard ratio. ²CI; confidence interval. ³P-value<0.05 statistically significant. doi:10.1371/journal.pone.0054596.t003

Several studies have recently validated the prognostic utility of the high plasma levels of miR-20a-5p in lung squamous cell carcinoma patients [16,35,36,37]. Furthermore, in our study, miR-199a-5p was a consistent element of the plasma miRNA signature for NSCLC diagnosis in comparison to both healthy individuals and COPD patients. This finding was in agreement with previous results obtained in a tobacco-specific carcinogen-induced lung cancer model that suggested a key role of miR-199a-5p as a part of an early-response miRNAs signature associated with pulmonary tumorigenesis [43]. Chen *et al.* reported a profiling study on serum miRNA expression among 400 NSCLC cases and 220 controls. MiR-152-3p and miR-223-3p were among the 10-serum miRNAs differentially expressed between cancer patients and control subjects in their cohort [44]. In addition, the serum expression level of miR-223 was associated with cancer-specific mortality in stage IA/B NSCLC patients [17]. Moreover, by negatively regulating tumor-suppressor genes, miR-155-5p promotes malignant transformation and cancer progression in many types of cancer, including NSCLC [45,46]. In our study, the up-regulation of miR-155-5p was found to be an independent negative prognostic factor in lung adenocarcinoma patients, as previously reported [34,47]. Shen *et al.* reported a four-miRNA panel in plasma, including miR-126 that distinguished NSCLC from healthy controls with a sensitivity and specificity of 73% and 96%, respectively [30]. One study reported the role as prognostic biomarker for miR-126-3p in NSCLC patients [48]. Finally, the fact that the six plasma miRNAs identified with prognostic utility in our study are common to other studies of NSCLC patients validates our findings.

In conclusion, we developed a robust methodology to study miRNAs in the plasma of NSCLC patients by using a clinically applicable qRT-PCR method. We identified two-plasma miRNA

signatures-histology related that are highly predictive of cancer progression. Three-plasma miRNA panel, including miR-152-3p, miR-199a-5p and miR-20a-5p, was associated with a higher risk for squamous cell carcinoma recurrence, and three-plasma miRNA signature including miR-155-5p, miR-223-3p and miR-126-3p was associated with a higher risk of lung adenocarcinoma progression. However, further studies are needed to fully validate these signatures, so as to investigate the mechanism by which miRNAs enter the bloodstream, to further elucidate the biological significance of miRNAs in the circulation and to evaluate therapeutic response.

Supporting Information

Figure S1 Heat-map clustering analysis of the deregulated miRNAs expression levels stratified according to NSCLC histology subtypes and either COPD patients or healthy individuals. Average linkage and 1-Pearson correlation as distance metric were used for the clustering. Abbreviations: ADC, adenocarcinoma; COPD, chronic obstructive pulmonary disease; SCC, squamous cell carcinoma. (TIF)

Figure S2 The expression levels of 13-plasma miRNAs included in our study and detected by qRT-PCR. Paired Student's *t*-test was performed to ascertain statistical significance between the expression levels across groups. Abbreviations: ADC, adenocarcinoma; COPD, chronic obstructive pulmonary disease; SCC, squamous cell carcinoma. (TIF)

Figure S3 Kaplan-Meier DFS curves for NSCLC patients, independently of histology, stratified according to plasma levels of miR-155-5p (A), miR-20a-5p (B), and miR-152-3p (C). The *P*-values were calculated using the log-rank test between patients with high- and low-fold changes. (TIF)

Figure S4 Kaplan-Meier DFS curves for NSCLC patients, independently of histology, stratified according to plasma levels of miR-223-3p (A), miR-191-5p (B), miR-320-3p (C), miR-126-3p (D), miR-145-5p (E), miR-199a-5p (F), miR-24-3p (G), miR-25-3p (H), miR-296-5p (I), and let-7f-5p (J). The *P*-values were calculated using the log-rank test between patients with high- and low-fold changes. (TIF)

Table S1 The panel of miRNAs included in the study. (DOC)

Table S2 Correlation of plasma miRNAs expression with clinicopathological parameters of NSCLC patients. (DOC)

Author Contributions

Conceived and designed the experiments: CS MI PB PH. Performed the experiments: CS AB MI. Analyzed the data: CS MI PB. Contributed reagents/materials/analysis tools: CHM FB JM PH. Wrote the paper: MI PB PH.

References

- Jemal A, Bray F, Center MM, Ferlay J, Ward E, et al. (2011) Global cancer statistics. *CA Cancer J Clin* 61: 69–90.
- Blanchon F, Grivaux M, Asselain B, Lebas FX, Orlando JP, et al. (2006) 4-year mortality in patients with non-small-cell lung cancer: development and validation of a prognostic index. *Lancet Oncol* 7: 829–836.
- Henschke CI, Yankelevitz DF, Libby DM, Pasmantier MW, Smith JP, et al. (2006) Survival of patients with stage I lung cancer detected on CT screening. *N Engl J Med* 355: 1763–1771.

4. Hu Z, Chen X, Zhao Y, Tian T, Jin G, et al. (2010) Serum microRNA signatures identified in a genome-wide serum microRNA expression profiling predict survival of non-small-cell lung cancer. *J Clin Oncol* 28: 1721–1726.
5. Chiriac LR, Flieder DB (2010) High-resolution computed tomography screening for lung cancer: unexpected findings and new controversies regarding adenocarcinogenesis. *Arch Pathol Lab Med* 134: 41–48.
6. Jiang F, Todd NW, Li R, Zhang H, Fang H, et al. (2010) A panel of sputum-based genomic marker for early detection of lung cancer. *Cancer Prev Res (Phila)* 3: 1571–1578.
7. Duffy MJ (2007) Role of tumor markers in patients with solid cancers: A critical review. *Eur J Intern Med* 18: 175–184.
8. Margolis ML, Hyzy JB, Schenken LL, Schepart BS (1994) Serum tumor markers in non-small cell lung cancer. A comparative analysis. *Cancer* 73: 605–609.
9. Wang QZ, Xu W, Habib N, Xu R (2009) Potential uses of microRNA in lung cancer diagnosis, prognosis, and therapy. *Curr Cancer Drug Targets* 9: 572–594.
10. Dannemann M, Pruffer K, Lizano E, Nickel B, Burbano HA, et al. (2012) Transcription Factors Are Targeted by Differentially Expressed miRNAs in Primates. *Genome Biol Evol* 4: 552–564.
11. McDermott AM, Henehgan HM, Miller N, Kerin MJ (2011) The therapeutic potential of microRNAs: disease modulators and drug targets. *Pharm Res* 28: 3016–3029.
12. Calin GA, Croce CM (2006) MicroRNA signatures in human cancers. *Nat Rev Cancer* 6: 857–866.
13. Nana-Sinkam SP, Croce CM (2011) Non-coding RNAs in cancer initiation and progression and as novel biomarkers. *Mol Oncol* 5: 483–491.
14. Chen X, Ba Y, Ma L, Cai X, Yin Y, et al. (2008) Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* 18: 997–1006.
15. Zhao H, Shen J, Medico L, Wang D, Ambrosone CB, et al. (2010) A pilot study of circulating miRNAs as potential biomarkers of early stage breast cancer. *PLoS One* 5: e13735.
16. Landi MT, Zhao Y, Rotunno M, Koshiol J, Liu H, et al. (2010) MicroRNA expression differentiates histology and predicts survival of lung cancer. *Clin Cancer Res* 16: 430–441.
17. Heegaard NH, Schetter AJ, Welsh JA, Yoneda M, Bowman ED, et al. (2012) Circulating micro-RNA expression profiles in early stage nonsmall cell lung cancer. *Int J Cancer* 130: 1378–1386.
18. Zheng D, Haddadin S, Wang Y, Gu LQ, Perry MC, et al. (2011) Plasma microRNAs as novel biomarkers for early detection of lung cancer. *Int J Clin Exp Pathol* 4: 575–586.
19. Boeri M, Verri C, Conte D, Roz L, Modena P, et al. (2011) MicroRNA signatures in tissues and plasma predict development and prognosis of computed tomography detected lung cancer. *Proc Natl Acad Sci U S A* 108: 3713–3718.
20. Xie Y, Todd NW, Liu Z, Zhan M, Fang H, et al. (2010) Altered miRNA expression in sputum for diagnosis of non-small cell lung cancer. *Lung Cancer* 67: 170–176.
21. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, et al. (2008) Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 105: 10513–10518.
22. Xi Y, Nakajima G, Gavin E, Morris CG, Kudo K, et al. (2007) Systematic analysis of microRNA expression of RNA extracted from fresh frozen and formalin-fixed paraffin-embedded samples. *RNA* 13: 1668–1674.
23. Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger KR, et al. (2011) International association for the study of lung cancer/american thoracic society/european respiratory society international multidisciplinary classification of lung adenocarcinoma. *J Thorac Oncol* 6: 244–285.
24. Kirschner MB, Kao SC, Edelman JJ, Armstrong NJ, Valley MP, et al. (2011) Haemolysis during sample preparation alters microRNA content of plasma. *PLoS One* 6: e24145.
25. Appaiah HN, Goswami CP, Mina LA, Badve S, Sledge GW Jr., et al. (2011) Persistent upregulation of U6: SNORD44 small RNA ratio in the serum of breast cancer patients. *Breast Cancer Res* 13: R86.
26. Lawrie CH, Gal S, Dunlop HM, Pushkaran B, Liggins AP, et al. (2008) Detection of elevated levels of tumour-associated microRNAs in serum of patients with diffuse large B-cell lymphoma. *Br J Haematol* 141: 672–675.
27. Vasilescu C, Rossi S, Shimizu M, Tudor S, Veronese A, et al. (2009) MicroRNA fingerprints identify miR-150 as a plasma prognostic marker in patients with sepsis. *PLoS One* 4: e7405.
28. Saeed AI, Bhagabati NK, Braisted JC, Liang W, Sharov V, et al. (2006) TM4 microarray software suite. *Methods Enzymol* 411: 134–193.
29. Yu SL, Chen HY, Chang GC, Chen CY, Chen HW, et al. (2008) MicroRNA signature predicts survival and relapse in lung cancer. *Cancer Cell* 13: 48–57.
30. Shen J, Todd NW, Zhang H, Yu L, Lingxiao X, et al. (2011) Plasma microRNAs as potential biomarkers for non-small-cell lung cancer. *Lab Invest* 91: 579–587.
31. Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, et al. (2005) MicroRNA expression profiles classify human cancers. *Nature* 435: 834–838.
32. Jay C, Nemunaitis J, Chen P, Fulgham P, Tong AW (2007) miRNA profiling for diagnosis and prognosis of human cancer. *DNA Cell Biol* 26: 293–300.
33. Nisman B, Lafair J, Heching N, Lyass O, Baras M, et al. (1998) Evaluation of tissue polypeptide specific antigen, CYFRA 21-1, and carcinoembryonic antigen in nonsmall cell lung carcinoma: does the combined use of cytokeratin markers give any additional information? *Cancer* 82: 1850–1859.
34. Yanaihara N, Caplen N, Bowman E, Seike M, Kumamoto K, et al. (2006) Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* 9: 189–198.
35. Lu Y, Govindan R, Wang L, Liu PY, Goodgame B, et al. (2012) MicroRNA profiling and prediction of recurrence/relapse-free survival in stage I lung cancer. *Carcinogenesis* 33: 1046–1054.
36. Raponi M, Dossey L, Jatke T, Wu X, Chen G, et al. (2009) MicroRNA classifiers for predicting prognosis of squamous cell lung cancer. *Cancer Res* 69: 5776–5783.
37. Huang W, Hu J, Yang DW, Fan XT, Jin Y, et al. (2012) Two MicroRNA Panels to Discriminate Three Subtypes of Lung Carcinoma in Bronchial Brushing Specimens. *Am J Respir Crit Care Med*.
38. McDaniels-Silvers AL, Nimri CF, Stoner GD, Lubet RA, You M (2002) Differential gene expression in human lung adenocarcinomas and squamous cell carcinomas. *Clin Cancer Res* 8: 1127–1138.
39. Leidinger P, Keller A, Borries A, Huwer H, Rohling M, et al. (2011) Specific peripheral miRNA profiles for distinguishing lung cancer from COPD. *Lung Cancer* 74: 41–47.
40. Celli BR (2012) Chronic obstructive pulmonary disease and lung cancer: common pathogenesis, shared clinical challenges. *Proc Am Thorac Soc* 9: 74–79.
41. Akbas F, Coskunpinar E, Aynaci E, Oltulu YM, Yildiz P (2012) Analysis of serum micro-RNAs as potential biomarker in chronic obstructive pulmonary disease. *Exp Lung Res* 38: 286–294.
42. Tan X, Qin W, Zhang L, Hang J, Li B, et al. (2011) A 5-microRNA signature for lung squamous cell carcinoma diagnosis and hsa-miR-31 for prognosis. *Clin Cancer Res* 17: 6802–6811.
43. Kalscheuer S, Zhang X, Zeng Y, Upadhyaya P (2008) Differential expression of microRNAs in early-stage neoplastic transformation in the lungs of F344 rats chronically treated with the tobacco carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone. *Carcinogenesis* 29: 2394–2399.
44. Chen X, Hu Z, Wang W, Ba Y, Ma L, et al. (2012) Identification of ten serum microRNAs from a genome-wide serum microRNA expression profile as novel noninvasive biomarkers for nonsmall cell lung cancer diagnosis. *Int J Cancer* 130: 1620–1628.
45. Jiang S, Zhang HW, Lu MH, He XH, Li Y, et al. (2010) MicroRNA-155 functions as an OncomiR in breast cancer by targeting the suppressor of cytokine signaling 1 gene. *Cancer Res* 70: 3119–3127.
46. Donnem T, Eklo K, Berg T, Sorbye SW, Lonvik K, et al. (2011) Prognostic impact of MiR-155 in non-small cell lung cancer evaluated by in situ hybridization. *J Transl Med* 9: 6.
47. Takamizawa J, Konishi H, Yanagisawa K, Tomida S, Osada H, et al. (2004) Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res* 64: 3753–3756.
48. Donnem T, Lonvik K, Eklo K, Berg T, Sorbye SW, et al. (2011) Independent and tissue-specific prognostic impact of miR-126 in nonsmall cell lung cancer: coexpression with vascular endothelial growth factor-A predicts poor survival. *Cancer* 117: 3193–3200.