

Under niche construction: an operational bridge between ecology,  
evolution, and ecosystem science

MATTHEWS, Blake, *et al.*

---

Reference

MATTHEWS, Blake, *et al.* Under niche construction: an operational bridge between ecology, evolution, and ecosystem science. *Ecological monographs*, 2014, vol. 84, no. 2, p. 245-263

DOI : 10.1890/13-0953.1

Available at:

<http://archive-ouverte.unige.ch/unige:32896>

Disclaimer: layout of this document may differ from the published version.



UNIVERSITÉ  
DE GENÈVE



# ECOLOGICAL SOCIETY OF AMERICA

*Ecology/Ecological Monographs/Ecological Applications*

## PREPRINT

This preprint is a PDF of a manuscript that has been accepted for publication in an ESA journal. It is the final version that was uploaded and approved by the author(s). While the paper has been through the usual rigorous peer review process of ESA journals, it has not been copy-edited, nor have the graphics and tables been modified for final publication. Also note that the paper may refer to online Appendices and/or Supplements that are not yet available. We have posted this preliminary version of the manuscript online in the interest of making the scientific findings available for distribution and citation as quickly as possible following acceptance. However, readers should be aware that the final, published version will look different from this version and may also have some differences in content.

The doi for this manuscript and the correct format for citing the paper are given at the top of the online (html) abstract.

Once the final published version of this paper is posted online, it will replace the preliminary version at the specified doi.

**Under niche construction: an operational bridge between  
ecology, evolution, and ecosystem science**

<sup>1</sup>Blake Matthews, <sup>2</sup>Luc De Meester, <sup>3</sup>Clive G. Jones, <sup>4</sup>Bas W. Ibelings, <sup>5</sup>Tjeerd  
J. Bouma, <sup>6</sup>Visa Nuutinen, <sup>7</sup>Johan van der Koppel, <sup>8</sup>John Odling-Smee

1. EAWAG, Aquatic Ecology department, Center for Ecology, Evolution and Biogeochemistry,  
Kastanienbaum, 6047, Switzerland, blake.matthews@eawag.ch

2. Laboratory of Ecology, Evolution and Conservation, University of Leuven, 3000 Leuven,  
Belgium, Luc.DeMeester@bio.kuleuven.be

3. Cary Institute of Ecosystem Studies, P.O. Box AB, Millbrook, NY 12545, USA,  
jonesc@caryinstitute.org

4. University of Geneva, Institut FA Forel, 10 Route de Suisse, Versoix, bastiaan.ibelings@unige.ch

5. Royal Netherlands Institute for Sea Research (NIOZ), Post Box 140, 4400 AC Yerseke,  
Netherlands, t.bouma@nioo.knaw.nl

6. MTT Agrifood Research Finland, FIN-31600 Jokioinen, Finland, visa.nuutinen@mtt.fi

7. Royal Netherlands Institute for Sea Research (NIOZ), Post Box 140, 4400 AC Yerseke,  
Netherlands, Johan.van.de.Koppel@nioz.nl

8. Mansfield College, University of Oxford, Oxford OX1 3TF, UK,  
john.odling-smee@mansfield.ox.ac.uk

**Article type:** Concepts and Synthesis

**Running title:** Under niche construction

**Statement of authorship:** All co-authors participated in the working group discussions, and contributed to subsequent discussions about the manuscript. BM wrote most of the first draft of the manuscript. LDM wrote several sections of the manuscript and made substantial contributions to the structure, concepts, and revisions of all manuscript drafts. BWI, VN, TB, and JvdK contributed substantially to revisions of the manuscript. JOS and CGJ made substantial contributions to the structure, concepts, and revisions of all manuscript drafts, and CGJ and TB organized, and CGJ chaired the working group.

**Corresponding author:** Blake Matthews:

blake.matthews@eawag.ch; Phone: +41 058 765 2120, Fax: +41 058 765 2168

## Abstract

1 All living organisms modify their biotic and abiotic environment. Niche construction  
 2 theory posits that organism-mediated modifications to the environment can change  
 3 selection pressures and influence the evolutionary trajectories of natural populations. While  
 4 there is broad support for this proposition in general, there is considerable uncertainty  
 5 about how niche construction is related to other similar concepts in ecology and evolution.  
 6 Comparative studies dealing with certain aspects of niche construction are increasingly  
 7 common, but there is a troubling lack of experimental tests of the core concepts of niche  
 8 construction theory. Here, we propose an operational framework to evaluate comparative  
 9 and experimental evidence of the evolutionary consequences of niche construction, and  
 10 suggest how such research can improve our understanding of ecological and evolutionary  
 11 dynamics in ecosystems. We advocate for a shift toward explicit experimental tests of how  
 12 organism-mediated environmental change can influence the selection pressures underlying  
 13 evolutionary responses, as well as targeted field-based comparative research to identify the  
 14 mode of evolution by niche construction and assess its importance in natural populations.

15 **Keywords:** niche construction, eco-evolutionary dynamics, ecosystem  
 16 engineering, alternative stable states, coevolution, diffuse coevolution, trophic  
 17 interactions, eco-evolutionary feedbacks, ecosystems

18 **Introduction**

19 The basic premise of niche construction theory is that organisms can act as potent agents  
 20 of natural selection by modifying biotic and abiotic environmental conditions (Lewontin,  
 21 1983; Odling-Smee et al., 2003, 2013). Previous research on niche construction has  
 22 extensively documented how living organisms, through their metabolism, activities, and  
 23 choices, can alter their surrounding environment and by doing so influence prevailing  
 24 selection pressures (Odling-Smee et al., 1996, 2003). Animals, for example, dig burrows,  
 25 build nests, aerate soils, construct webs, and forage for prey, while plants photosynthesize,  
 26 weather rocks, produce soil, and create shade (Odling-Smee et al., 2003). Such activities  
 27 can modify the selective environment of the organism doing the environmental modification  
 28 (Odling-Smee et al., 1996) or of an unrelated population (Odling-Smee et al., 2003, 2013).  
 29 Organism-mediated environmental modifications can also persist through time and affect  
 30 selection pressures experienced by future generations, a process referred to as ecological  
 31 inheritance (Odling-Smee et al., 2003). Ecological inheritance is a key element of niche  
 32 construction theory that is increasingly being integrated into evolutionary theory  
 33 (Bonduriansky and Day, 2009; Danchin et al., 2011; Bonduriansky, 2012).  
 34 When using the term niche construction (Odling-Smee et al., 2003, 2013), niche refers to  
 35 the sum of all natural selection pressures experienced by a population and construction  
 36 refers to the modification of selection pressures, either through physical modification of the

37 environment or through habitat choice (Odling-Smee et al., 2003). At the outset, niche  
 38 construction theory focused on how organisms can modify their own selective environments  
 39 (Odling-Smee et al., 1996), and so many classic examples of niche construction highlight  
 40 the importance of reciprocal interactions between organisms and their own selective  
 41 environment (Odling-Smee et al., 2003). Leaf cutter ants, for example, cultivate gardens of  
 42 fungus upon which they are obligately dependent (Mueller and Gerardo, 2002), and, in  
 43 some cases, this has culminated in a loss of genes associated with the acquisition of specific  
 44 nutrients (Ellers et al., 2012). Earthworms modify the structure of their soil environment  
 45 in a way that facilitates water uptake into their bodies, thereby partially solving a critical  
 46 physiological problem associated with living in terrestrial environments (Turner, 2002).  
 47 However, it is increasingly evident that organism-mediated environmental modifications  
 48 can have a wide range of direct and indirect evolutionary effects on multiple species in  
 49 natural communities (Odling-Smee et al., 2013; Walsh, 2013). Odling-Smee et al. (2003)  
 50 describe one type of indirect evolutionary effect as an environmentally mediated genotypic  
 51 association (EMGA), which is an association that develops between distinct genotypes in  
 52 the environment mediated by the effect of organisms on biotic or abiotic conditions. For  
 53 example, earthworms might influence the selective environment experienced by plants  
 54 growing in the same soils, potentially leading to covariance between the plant's fitness and  
 55 the worm's genes that underlie modifications to the soil environment (Odling-Smee et al.,  
 56 2003).

57 Clarifying the relationship between environment-modifying activities of organisms and  
 58 fitness variation has been controversial throughout the development of niche construction  
 59 theory (Dawkins, 2004; Laland and Sterelny, 2006). Dawkins (2004) argues that the  
 60 buildup of covariance between fitness and phenotype is much more likely to occur within a  
 61 gene pool, consistent with the idea of an extended phenotype (Dawkins, 1982), rather than  
 62 across gene pools (Dawkins, 2004). In the case of an extended phenotype, the phenotypic  
 63 trait that underlies the organism-mediated modifications of the environment must vary  
 64 within a population, have a genetic basis, and be the target of the altered selection regime  
 65 caused by the environmental modifications (Dawkins, 2004; Brodie III, 2005). For example,  
 66 genetically based variation among gall wasps in their ability to construct oak galls can  
 67 affect rates of parasitoid infection in the next generation of gall wasps, leading to a  
 68 covariance between gall forming traits and offspring fitness (Bailey et al., 2009). While not  
 69 disputing the importance of extended phenotypes, niche construction theory (Odling-Smee  
 70 et al., 2003) argues that the traits underlying specific environmental modifications neither  
 71 need to have a strong genetic basis (for example, they can be acquired characters) nor need  
 72 to be the same traits that develop strong associations with fitness. Hence, compared to  
 73 Dawkins (2004), Odling-Smee et al. (2003, 2013) consider a broader range of selective  
 74 agents that can potentially drive evolution, and suggest that covariance between fitness  
 75 and phenotype can frequently build up across species, resulting from organism-mediated  
 76 modifications to both biotic and abiotic environmental conditions. While empirical data



77 and theoretical work are increasingly supporting this view (Kerr et al., 1999; Odling-Smee  
 78 et al., 2003; Krakauer et al., 2009; Laland, 2010; Kylafis and Loreau, 2011), the ongoing  
 79 challenge is to determine how much of the variance in fitness of one organism can be  
 80 explained by organism-mediated environmental modifications compared to other agents of  
 81 selection.

82 Since its inception, niche construction theory has captured the attention of a wide range of  
 83 evolutionary biologists, ecologists, and philosophers (Erwin, 2008; Lehmann, 2008;  
 84 Krakauer et al., 2009; Post and Palkovacs, 2009; Loreau, 2010; Kylafis and Loreau, 2011;  
 85 Van Dyken and Wade, 2012), but has also provoked considerable debate as to its novelty  
 86 (Brodie III, 2005), scope (Okasha, 2005; Kylafis and Loreau, 2008), and usefulness  
 87 (Dawkins, 2004). Niche construction has been defined with a deliberately broad scope  
 88 (Laland and Sterelny, 2006), and this has offered ecologists new insights about how  
 89 modifications to the environment by organisms might persist over time (e.g. ecological  
 90 inheritance), result from byproducts and acquired characters (Odling-Smee et al., 2003),  
 91 and interact with other environmental sources of selection so as to influence evolutionary  
 92 change in natural populations (Odling-Smee et al., 2013).

93 While generally received sympathetically, the broad scope of niche construction theory has  
 94 nonetheless led to some confusion and conflicts about how aspects of the theory are  
 95 positioned in relation to other closely related ideas in both ecology and evolution. For  
 96 example, the concept of reciprocal interactions between organisms and their selective

97 environments is both fundamental to niche construction theory and long-established in  
 98 some areas of standard evolutionary theory (Fisher, 1930; Roughgarden, 1976; Crespi,  
 99 2004; Frank, 2009), particularly in classic work on coevolution and diffuse coevolution  
 100 (Thompson, 2005; Haloin and Strauss, 2008). In ecology, there is also some uncertainty  
 101 about precisely what new insights niche construction theory can offer. On the one hand,  
 102 niche construction theory has already made important contributions to emerging syntheses  
 103 between ecological and evolutionary dynamics (Fussmann et al., 2007; Kokko and  
 104 López-Sepulcre, 2007; Post and Palkovacs, 2009; Schoener, 2011; Matthews et al., 2011b;  
 105 Kylafis and Loreau, 2011). In particular, niche construction research has documented a  
 106 broad range of organism-mediated environmental modifications that can influence selection  
 107 pressures (Odling-Smee et al., 2003). With the growing realization that ecological and  
 108 evolutionary timescales can be congruent (Hairston et al., 2005; Ellner et al., 2011), such  
 109 environmental modifications might turn out to be more important agents of selection and  
 110 drivers of evolutionary change than previously thought (Odling-Smee et al., 2013). On the  
 111 other hand, the precise relationship between niche construction theory and  
 112 eco-evolutionary dynamics is unclear, and there is confusion about how niche construction  
 113 is related to other ecological concepts in general, and to ecosystem engineering in particular  
 114 (Odling-Smee et al., 2003; Boogert et al., 2006; Pearce, 2011). Even though ecosystem  
 115 engineering theory clearly recognizes that the engineering effects of organisms can have  
 116 important evolutionary consequences (Jones et al., 1994), the strict definitions of ecosystem

117 engineering (Jones et al., 1994, 1997) and niche construction (Odling-Smee et al., 1996,  
 118 2003) refer to distinct concepts (see below).

119 In our view, niche construction theory has the potential to bridge many related concepts in  
 120 ecology, evolution, and ecosystem science. With the goal of integration in mind,  
 121 Odling-Smee et al. (2013) recently distinguished between two important "aspects" of the  
 122 process of niche construction. The first aspect is the environment-altering activities of  
 123 organisms, and the second is the subsequent modification of the selective environment  
 124 (Odling-Smee et al., 1996, 2003, 2013). Niche construction is only present if both aspects  
 125 occur, as not all environmental modifications will alter selection pressures. Similarly, not  
 126 all changes to selection pressures will cause an evolutionary response, meaning that niche  
 127 construction can occur without influencing evolution. In order to evaluate the importance  
 128 of evolution by organism-mediated environmental modification in natural populations, we  
 129 need to translate niche construction theory into empirical practice (Odling-Smee et al.,  
 130 2013). To do this, we propose the following criteria to test for the presence of niche  
 131 construction (Criteria 1 and 2) and determine when niche construction affects evolution  
 132 (Criterion 3).

133 **Criterion 1**

134 An organism (i.e. a candidate niche constructor) must significantly modify  
 135 environmental conditions.

136 **Criterion 2**

137 The organism-mediated environmental modifications must influence selection  
 138 pressures on a recipient of niche construction.

139 **Criterion 3**

140 There must be a detectable evolutionary response in a recipient of niche construction  
 141 that is caused by the environmental modification of the niche constructor.

142 Here, we refer to the environment in relation to both biotic and abiotic characteristics, and  
 143 the selective environment as the environmental context in which natural selection occurs.

144 The first two criteria define the term niche construction (Odling-Smee et al., 2013). The  
 145 organism changing the environmental conditions is only classified as a niche constructor if  
 146 criterion 2 is satisfied. The third criterion is a test of evolution by niche construction, or in  
 147 other words, evolution via selection that is mediated by organismal modification of the  
 148 environment. We consider an evolutionary response as a genetic change in a population  
 149 that alters the relationship between the phenotype distribution (including mean, variance,  
 150 and other moments of the distribution) and fitness variation. We distinguish between a  
 151 niche constructor and a recipient of niche construction, but explicitly recognize that both  
 152 can refer to the same organism. For example, in the case of an extended phenotype the  
 153 niche constructor and recipient of niche construction would be organisms within the same  
 154 gene pool, whereas in the case of an environmentally mediated genotypic association the

155 niche constructor and recipient could be different species.

156 Using these three criteria we can evaluate which sets of ecological and evolutionary

157 interactions describe evolution by niche construction, and which do not. We summarize

158 this approach graphically in Figure 1 where we consider a wide range of scenarios in which

159 organisms are connected with their biotic and abiotic environment via pathways of

160 evolutionary (dashed arrows) and non-evolutionary (solid arrows) effects. Evolutionary

161 effects are those cases where organisms cause an evolutionary response (e.g. Criterion 3),

162 while non-evolutionary effects include the effects organisms have on the abundance,

163 distribution, and behavior of interacting biota (e.g. collectively referred to as ecological

164 effects), as well as effects on the physical (e.g. engineering effects) and chemical state of

165 their environment (Criterion 1, Figure 2A). For a particular scenario in Figure 1 to satisfy

166 evolution by niche construction (i.e. the minimum condition for satisfying Criterion 3), the

167 pathway of effects must start (from the left) with a niche constructor, it must include at

168 least two sequential effects (i.e. connections in sequence along the pathway of effects), and

169 there must be an evolutionary effect beyond the first effect. This last condition follows

170 from our second criterion, which requires selection pressures to be mediated through some

171 form of environmental modification by an organism, including changes to either abiotic or

172 biotic conditions (Figure 2). Evolution by niche construction does not occur for scenarios

173 where the evolutionary response of an organism is caused solely by the direct selection

174 effects of another organism or by an environmental condition that is unmodified by another

175 organism. Such scenarios are examples of evolution, but not of evolution by niche  
 176 construction (Figure 1). Following our scheme, there are many simple modules of ecological  
 177 interactions that do not meet all three criteria (Figure 1: modules within the ecology box  
 178 but outside the evolution box). This highlights that there is considerable scope for  
 179 ecologists to use niche construction theory to help integrate evolution and ecosystem  
 180 ecology. To facilitate this, we clarify how niche construction (Criteria 1 and 2) and  
 181 evolution by niche construction (Criterion 3) are related to several key concepts, such as:  
 182 ecosystem engineering, (diffuse) co-evolution, and eco-evolutionary dynamics and feedbacks.  
 183 *Ecosystem engineering* - The distinction between ecosystem engineering and niche  
 184 construction is currently unclear in the literature (Boogert et al., 2006; Post and Palkovacs,  
 185 2009; Pearce, 2011; Odling-Smee et al., 2013). Ecosystem engineers are organisms that  
 186 modify their physical surroundings (e.g. light environment, physical habitat structure) so  
 187 as to modulate the availability of resources or energy fluxes in an ecosystem (Jones et al.,  
 188 1994, 1997). By comparison, niche constructors are organisms that alter selection pressures  
 189 of a recipient organism by modifying any aspect of the abiotic and biotic environment  
 190 (Figure 2). Evidence of ecosystem engineering would only satisfy our first criterion, and  
 191 would not provide evidence of niche construction. Nevertheless, ecosystem engineers are  
 192 excellent candidates for being niche constructors because their effects on the physical  
 193 environment can propagate to influence chemical fluxes and species interactions, and cause  
 194 ecosystem effects that are large, multidimensional, and persistent (Wright and Jones, 2006;

195 Hastings et al., 2007; Jones, 2012). Ecosystem engineering is hence a putative mechanism  
 196 of niche construction, and further work should focus on the how engineers might alter  
 197 selection pressures on themselves or on other species (Criterion 2).

198 *Coevolution and diffuse coevolution* - Based on our criteria and schematic (Figure 1), all  
 199 examples of pairwise coevolution and diffuse coevolution are examples of evolution by niche  
 200 construction. Pairwise coevolution is the situation where two interacting organisms are  
 201 both niche constructors and recipients of niche construction (Figure 2B) and they both  
 202 drive reciprocal evolutionary responses on one another. Diffuse coevolution is the case  
 203 where a niche constructor drives an evolutionary response of a recipient that is a different  
 204 species, and where this response is mediated through the niche constructors's ecological or  
 205 evolutionary effect on another species that interacts with the recipient (Haloin and Strauss,  
 206 2008). Hence, diffuse co-evolution is equivalent to evolution by niche construction where  
 207 the selective environment is modified by species interactions in the community. In sum,  
 208 compared to all forms of coevolution, evolution by niche construction considers a broader  
 209 range of potential agents of selection and effect pathways that underlie evolutionary  
 210 responses (Figure 1).

211 *Eco-evolutionary dynamics* - The emerging field of eco-evolutionary dynamics has a very  
 212 broad focus that includes both the ecological and evolutionary responses of populations to  
 213 interactions between organisms and their environment (Fussmann et al., 2007; Urban et al.,  
 214 2008; Post and Palkovacs, 2009; Matthews et al., 2011b; Schoener, 2011). Eco-evolutionary



215 dynamics grew out the recognition that population dynamics and phenotypic evolution can  
 216 occur on similar timescales, leading to an important contemporary interplay between  
 217 evolutionary and ecological dynamics in natural populations (Thompson, 1998; Hairston  
 218 et al., 2005; Schoener, 2011).

219 Evolution by niche construction is closely related to eco-evolutionary dynamics but the two  
 220 concepts have slightly different emphases and are distinguishable in our schematic (Figure  
 221 1). Although the distinction is often likely to be subtle, it is useful to identify the  
 222 minimum conditions that constitute each process in order to perform more targeted  
 223 experimental tests of the specific mechanisms. Eco-evolutionary dynamic scenarios must  
 224 include at least two organisms and at least one evolutionary and one ecological effect (i.e. a  
 225 non-evolutionary effect terminating with a biotic recipient). Neither of these two conditions  
 226 are necessary for evolution by niche construction.

227 Following our scheme, there are simple cases of evolution by niche construction that do not  
 228 constitute eco-evolutionary dynamics, and vice versa (Figure 1). Unlike eco-evolutionary  
 229 dynamics, evolution by niche construction includes scenarios made up of entirely  
 230 evolutionary effects (Figure 1), including linked chains of evolutionary effects (e.g.  
 231 evolutionary cascades) and reciprocal evolutionary effects (e.g. coevolution). In addition,  
 232 evolution by niche construction includes simple scenarios where an evolutionary effect  
 233 follows from an organism's effect on abiotic environmental conditions. In relation to Figure  
 234 1, for example, worms (square) can modify (solid arrow) the soil environment (circle) and



235 affect the evolution (dashed arrow) of plants (square). Such chains of interactions where  
 236 abiotic modifications influence selection pressures are an important emphasis of niche  
 237 construction theory (Odling-Smee et al., 2013) but in their simplest form can fall outside  
 238 the domain of eco-evolutionary dynamics (Figure 1).  
 239 Eco-evolutionary dynamics scenarios can also occur without evolution by niche  
 240 construction. In relation to Figure 1, for example, a predator (square) may cause an  
 241 evolutionary response (dashed arrow) in the life history of a prey population (circle) that  
 242 subsequently changes prey consumption rates (solid arrow) on a resource (circle). This is  
 243 illustrated by recent work showing that alewives, a common planktivorous fish in  
 244 freshwater lakes of eastern North America, drive evolution in *Daphnia* in a way that alters  
 245 their grazing rates on phytoplankton (Walsh et al., 2012). This particular example does  
 246 not meet our second criterion for niche construction, because the selection pressure of  
 247 alewives on *Daphnia* is not mediated by an environmental modification caused by alewives.  
 248 In more complicated scenarios, evolution by niche construction and eco-evolutionary  
 249 dynamics will likely overlap, particularly when there are multiple interacting species and  
 250 complex networks of ecological and evolutionary effects. This may also be true for the  
 251 alewife system (as discussed below), where there is additional evidence for eco-evolutionary  
 252 feedbacks and niche construction (Palkovacs and Post, 2008; Post and Palkovacs, 2009).  
 253 *Eco-evolutionary feedbacks* - Eco-evolutionary feedbacks are a specific type of  
 254 eco-evolutionary dynamics that describe a reciprocal interaction between an ecological and

255 evolutionary process (Post and Palkovacs, 2009). To provide evidence of evolution by niche  
 256 construction an eco-evolutionary feedback must include an evolutionary response to  
 257 organism-mediated changes in the environment (Figure 1). Eco-evolutionary feedbacks do  
 258 not always satisfy the criteria for niche construction (Criterion 2) or for evolution by niche  
 259 construction (Criterion 3). In some situations, the sequence of the linked effects can be  
 260 important for identifying evolution by niche construction.

261 Eco-evolutionary feedbacks that begin with an ecological effect and subsequently cause an  
 262 evolutionary effect are clearly classified as evolution by niche construction. For example,  
 263 migratory and landlocked populations of the alewives can have contrasting effects on the  
 264 composition and size structure of their prey communities (Palkovacs and Post, 2008), and  
 265 this is thought to generate divergent selection and contribute to the phenotypic divergence  
 266 among allopatric populations of alewives (Post and Palkovacs, 2009). In this case,  
 267 evolution by niche construction has occurred if the ecological effects of alewives (i.e.  
 268 changes in prey species composition, or life history of a specific prey) drive phenotypic  
 269 evolution of the alewives themselves, or indirectly cause an evolutionary response of some  
 270 other organism in the system. Recently, Walsh et al. (2012) reviewed several studies that  
 271 piece together the network of ecological and evolutionary interactions between alewives,  
 272 zooplankton, and phytoplankton. Together these studies provide growing evidence for  
 273 eco-evolutionary feedbacks and niche construction in natural populations.

274 Eco-evolutionary feedbacks that begin with an evolutionary effect may or may not be

275 classified as evolution by niche construction. Consider an eco-evolutionary feedback in  
 276 which a predator is both causing the evolutionary effect on a prey species and is the  
 277 recipient of the ecological effect from the altered evolution of the prey. If the ecological  
 278 effect that feeds back on the predator subsequently modifies the predator's evolutionary  
 279 effect on the prey, then this would constitute evolution by niche construction (Criterion 3).  
 280 For example, in relation to Figure 1, evolution by niche construction would occur if the  
 281 predator (circle) directly alters the genotype distribution (dashed arrow) of prey (circle),  
 282 this has a feedback on the population dynamics of the predator (solid arrow), and this  
 283 subsequently changes the predator's effect on the genotype distribution (dashed arrow) of  
 284 prey. This is analogous to situations where predator-mediated selection pressures are  
 285 dependent on densities of predators and prey (i.e. density- and frequency-dependent  
 286 selection). Evolution by niche construction would not occur if the evolutionary responses of  
 287 the prey were independent of (or insensitive to) variation in predator density, because the  
 288 ecological effects of prey evolution on predator population dynamics would have no further  
 289 influence on prey evolution. In such a scenario, an eco-evolutionary feedback could occur in  
 290 the absence of evolution by niche construction. Again, we acknowledge this is subtle  
 291 distinction between eco-evolutionary feedbacks and evolution by niche construction, but  
 292 such considerations might help to decipher the mechanisms underlying coupled ecological  
 293 and evolutionary dynamics.

294 So far, we have used our criteria to clarify how key elements of niche construction theory

295 are positioned relative to other closely related concepts in ecology and evolution. In the  
 296 following sections, we (i) use our criteria to evaluate evidence of niche construction from a  
 297 wide range of studies and to identify new research directions, (ii) present new comparative  
 298 and experimental approaches for testing several elements of niche construction theory, and  
 299 (iii) describe a well-established model system in ecology that is useful for studying  
 300 evolution by niche construction in natural ecosystems.

301 **Bridging disciplinary gaps with niche construction**  
 302 **research**

303 We surveyed a wide selection of literature that was relevant to understanding the multiple  
 304 facets of niche construction theory and used our criteria to identify potential future  
 305 avenues of research (Table 1). Although our review is not exhaustive, it illustrates the  
 306 following three issues; (i) some of the potential mechanisms of niche construction (Figure  
 307 2) are well studied while others are not (Table 1: GAP I), (ii) several research areas in  
 308 ecology and evolution could be expanded to test for new examples of evolution by niche  
 309 construction by measuring evolutionary responses of organism-mediated environmental  
 310 modifications (Table 1: GAP II), (iii) many studies that explicitly discuss niche  
 311 construction are based on comparative evidence and would benefit from additional  
 312 experimental support (Table 1: see Criteria column).

313 (i) *Broaden the study of potential niche construction mechanisms: Table 1,*

314 *GAP I*

315 There are numerous mechanisms by which organisms can modify their environment, and  
 316 parsing these out (Figure 2A) can provide clues about the potential fitness effects on  
 317 recipient organisms (Figure 2B). To begin, it is useful to partition the environment into  
 318 components that are either modifiable or un-modifiable by a particular organism over a  
 319 relevant timescale necessary to assess a change in selection pressures or to track an  
 320 evolutionary response in a recipient organism. We then split each environmental partition  
 321 into the four categories shown in Figure 2A, which we discuss below, as a way to  
 322 summarize the wide range of potential mechanisms of niche construction that we have  
 323 identified from previous studies (Table 1).

324 *Abiotic effects: Physical* - The ubiquity of ecosystem engineers across a range of natural  
 325 systems testifies to the capacity for organisms to strongly modify their physical  
 326 environment (Figure 2A). Interestingly, such effects can also have strong cascading effects  
 327 on other biotic and abiotic factors (Jones et al., 1994; Hastings et al., 2007), but very little  
 328 is known about how ecosystem engineers mediate selection pressures and drive evolutionary  
 329 responses in natural populations.

330 *Abiotic effects : Chemical* - Organisms with strong nutrient homeostasis (Sturner and Elser,  
 331 2002) can affect their chemical environment through the acquisition and regeneration of

332 resources (Figure 2A). The evolution of consumer elemental ratios (e.g. C:N:P) is often  
 333 closely related to growth, such that variation in the growth rate among organisms can have  
 334 major impacts on biologically mediated flows of chemicals in the environment (Sterner and  
 335 Elser, 2002). Feedbacks between consumer growth rate and modifications to the chemical  
 336 environment have been addressed by theory (Mizuno and Kawata, 2009), but little is known  
 337 about how variation in organismal C:N:P ratios might affect selection pressures in nature.

338 *Biotic effects : Consumer resource interactions* - Host-parasite and predator-prey  
 339 interactions are both archetypal consumer-resource interactions (Figure 2A) and provide  
 340 some of the best empirical examples of how organisms can modify their biotic environment  
 341 (Lafferty et al., 2008; Holt and Lawton, 1994). Predators, for example, can have strong  
 342 effects on community structure (Chase et al., 2009) and ecosystem functions (Schmitz,  
 343 2010) and can drive eco-evolutionary feedbacks (Post and Palkovacs, 2009; Becks et al.,  
 344 2012). The prevalence of trait-mediated indirect effects (Werner and Peacor, 2003; Walsh,  
 345 2013) suggests a rich set of ways that consumers can alter selection pressures through  
 346 modification of biotic interactions.

347 *Biotic effects : Non trophic direct interactions* - Non-trophic direct interactions between  
 348 species (Olf et al., 2009) can also drive changes to the biotic environment leading to  
 349 altered selective environments (Figure 2A). This category of potential mechanisms of niche  
 350 construction reflects the non-consumptive activities of organisms that might lead to  
 351 evolutionary changes, such as interference competition, cooperation, induced defence, and

352 behavioral modification. As one example, the relationship between a plant's fitness and its  
 353 tolerance to herbivory by deer (i.e. a selection gradient) is influenced by whether insect  
 354 herbivores are active in the system (Stinchcombe and Rausher, 2002).

355 Partitioning the mechanisms of organism-mediated environmental effects (e.g. Figure 2)  
 356 provides a structure for isolating the interactions underlying organismal effects on selective  
 357 environments (Criterion 2) and for detecting subsequent evolutionary responses (Criterion  
 358 3). In general, very little is known about how organism-mediated modifications to the  
 359 chemical and physical state of the environment can affect selection pressures (GAP I in  
 360 Table 1). Among the more evolutionarily oriented studies in our literature review, the  
 361 greater focus on the biotic effects (MacColl, 2011) over the abiotic effects (Jones et al.,  
 362 1994) of organisms is symptomatic of the limited cross-fertilization of ideas between  
 363 evolutionary biology and ecosystem ecology (Matthews et al., 2011*b*). For example, there is  
 364 considerable experimental work aimed at deciphering which species interactions underlie  
 365 the divergent selection regimes that drive ecological speciation (Schluter, 2000; Nosil,  
 366 2012), but there is much less research about how recent adaptive divergence between  
 367 closely related species can affect abiotic environmental conditions (Harmon et al., 2009),  
 368 and no experimental tests about whether such effects can influence selection pressures so as  
 369 to either promote or constrain further evolutionary divergence (Losos, 2010; Yoder et al.,  
 370 2010).

371 It is important to identify the modifiable components of the environment that might



372 underly selection pressures (Criterion 2) and drive evolutionary responses (Criterion 3),  
 373 because multiple interacting agents of selection can lead to complex relationships between  
 374 fitness and phenotype (Wade and Kalisz, 1990; MacColl, 2011). Organisms, for example,  
 375 might modify the environment in ways that either counteract or amplify other drivers of  
 376 environmental change (Odling-Smee et al., 2003, 2013), meaning that the various  
 377 mechanisms of niche construction (Figure 2B) may vary in their likelihood of driving  
 378 evolutionary responses in a particular environmental setting. Currently, we know little  
 379 about how selective agents interact across a range of environmental conditions (Wade and  
 380 Kalisz, 1990; MacColl, 2011), and this poses a major challenge for predicting the course of  
 381 adaptive evolution in natural populations (Barrett and Hoekstra, 2011). As part of an  
 382 intensive research effort integrated across disciplines, ecologists can use niche construction  
 383 theory to better understand the ecological causes of a broad range of evolutionary  
 384 dynamics.

385 *(ii) Measure evolutionary responses to organism-mediated environmental*  
 386 *effects: Table 1, GAP II*

387 Our literature review revealed that many of the more ecologically oriented studies rarely  
 388 investigate organism-mediated environmental effects together with evolutionary responses  
 389 (GAP II in Table 1). Recent research on the reciprocal interactions between ecological and



390 evolutionary dynamics is increasingly filling this gap (Hairston et al., 2005; Schoener, 2011;  
 391 Becks et al., 2012), but more studies are needed that examine how chemical modifications  
 392 of the environment by organisms affect the evolution of consumer resource demand  
 393 (Mizuno and Kawata, 2009; Matthews et al., 2011b), and how physical modification of the  
 394 environment by ecosystem engineers can modify selection gradients of the engineers  
 395 themselves or of other organisms (Wright et al., 2012). An interesting example of this gap,  
 396 and one we will return to later, is that while there is considerable research on  
 397 organism-mediated transitions between alternative stable states in ecosystems (Scheffer  
 398 et al., 2001), there is little research quantifying to what extent such states generate  
 399 contrasting selection pressures and lead to quantifiable differences in evolutionary  
 400 responses.

401 *(iii) Experimentally test more putative mechanisms of niche construction*

402 In our literature review, studies that explicitly discuss niche construction more often rely  
 403 on comparative (Beerling, 2005; Erwin and Tweedt, 2011) than experimental (Donohue  
 404 et al., 2005; Goddard, 2008) evidence to support their arguments (Table 1). For example,  
 405 the habitat modifying activities of bioturbating species, such as earthworms and bivalves,  
 406 are consistent with adaptive explanations (Turner, 2002; Odling-Smee et al., 2003), and the  
 407 adaptive radiations following the evolution of bioturbators strongly suggest a  
 408 macroevolutionary response driven by modifications to soils and sediments (Turner, 2002;

409 Meysman et al., 2006; Erwin and Tweedt, 2011). However, there is little experimental  
 410 evidence showing how bioturbation activities can affect selection pressures (Criterion 2) in  
 411 a way that would affect evolutionary responses (Criterion 3). There are, however,  
 412 experimental studies that measure changes in selection pressures caused by  
 413 organism-mediated modifications to the environment, illustrative of the type of research  
 414 needed to address the second criterion (Wright et al., 2012). In a study on ecosystem  
 415 engineers, Wright et al. (2012) showed that invasive seaweeds (*Caulerpa taxifolia*) modify  
 416 the physical and chemical characteristics of coastal marine sediments, and, in so doing,  
 417 alter selection gradients on native bivalves (*Anadara trapezia*). Specifically, the  
 418 relationships between several morphological traits (e.g. shell length, gill weight, and palp  
 419 weight) and relative performance (i.e. change in biomass over time) of *Anadara trapezia*  
 420 (the recipient of niche construction) differed in the presence and absence of *Caulerpa*  
 421 *taxifolia* (the niche constructor). While this study showed habitat-specific variation in  
 422 selection gradients, it did not document contrasting evolutionary responses and so does not  
 423 meet our third criterion. Nevertheless, similar experimental approaches could be expanded  
 424 upon to test for evolutionary responses of organisms to a broad range of environmental  
 425 modifications. In the following section, we expand on earlier ideas (Odling-Smee et al.,  
 426 2003) in order to develop new approaches to comparatively and experimentally test key  
 427 elements of niche construction theory.

428 **Designing comparative tests of niche construction**

429 **theory**

430 Many of the archetypical examples used to explain niche construction theory are largely  
 431 based on comparative evidence (Odling-Smee et al., 2003). Here, we summarize some  
 432 comparative approaches to identify niche construction and test for evolution by niche  
 433 construction.

434 *Do organism-mediated environmental modifications affect selective*  
 435 *environments?*

436 The environmental effects of organisms are often determined by their biomass and  
 437 dominance in an ecosystem (Vanni et al., 1997), by their functional role (Jones et al.,  
 438 1997), and by their phenotype (Schmitz, 2010). A comparative study that builds on such  
 439 ecological work, could gain support for the first criterion by finding contrasting  
 440 relationships between the un-modifiable and modifiable components of the environment in  
 441 the presence and absence of a putative niche constructor (Figure 3A). Further support  
 442 could come from relationships between the abundance of a niche constructor and  
 443 unexplained variation in the modifiable component of the environment (Figure 3B).

444 To test the second criteria, one could use well-established approaches to test how putative  
 445 selective agents (i.e. environmental modifications) shape the phenotypic distribution of a

446 population. Evidence of selection can be quantified by measuring selection differentials,  
 447 which are the mean trait differences between the entire population and the subset of  
 448 individuals that parent the next generation (Endler, 1986), and by quantifying selection  
 449 gradients, which are the slopes of the relationships between relative fitness and a  
 450 quantitative trait that is expressed in units of standard deviation (Hoekstra et al., 2001).  
 451 In a comparative study, one can either test for crossing reaction norms of the fitness of a  
 452 recipient organism in habitats with and without a niche constructor (Figure 3C), or test  
 453 whether variation in the environmental effects of a putative niche constructor covaries  
 454 positively or negatively with selection gradients of a recipient population (Figure 3D).  
 455 It is important to note that identifying such associations requires extensive data sets in  
 456 terms of the number of sampling sites or habitats. In addition, these studies would need to  
 457 rule out several alternative explanations for associations. These include non-random  
 458 habitat selection by the niche constructor based specifically on the environmental  
 459 conditions that it could otherwise modify, and habitat-specific variation in either the  
 460 carrying capacity of the niche constructor within its potential niche space or in the  
 461 selective environment favoring certain phenotypes. These alternate explanations can be  
 462 difficult to eliminate without experimental manipulations.

463 *Does the modified environment by an organism cause an evolutionary*  
 464 *response in a recipient?*

465 To test for evolution by niche construction (Criterion 3), one must determine whether an  
 466 organism-mediated environmental modification acts as an agent of selection and causes an  
 467 evolutionary response in a recipient species. One potential comparative approach would be  
 468 to quantify how the rate of evolutionary change of a recipient differs in environments that  
 469 are either modified or unmodified by a niche constructor. Rates of evolution can be  
 470 quantified in Haldane units, which measure the change in a mean trait value per generation  
 471 relative to its standard deviation (Hendry and Kinnison, 1999). However, such an approach  
 472 would also include any phenotypic changes caused by plasticity, and would not satisfy our  
 473 third criterion that requires a genetic component of evolutionary change. This could be  
 474 addressed by performing common garden experiments with organisms from the recipient  
 475 population that have been exposed to the modified and unmodified environments.

476 In a recent review, Hansen et al. (2012) propose clear criteria for quantifying adaptive  
 477 genetic responses to specific environmental changes, and these can be adopted to test  
 478 criterion 3. In summary, the approach is to (i) demonstrate that suitable genetic variation  
 479 exists that could respond to a specific environmental modification, (ii) test for a genetic  
 480 change over time consistent with selection, and (iii) confirm that the environmental  
 481 modification caused the observed genetic change within the defined population (Hansen

482 et al., 2012). Indeed, it is not easy to unambiguously show that the environmental  
 483 modifying activities of organisms affect their own evolutionary trajectory, or that of  
 484 another recipient population. The most direct way is to test for relationships between allele  
 485 frequencies or genotypic trait values of a recipient species and the extent of environmental  
 486 modification caused by the niche constructor (Figure 4C panels i and ii). We are not aware  
 487 of any studies that have attempted this in the framework of niche construction theory.

## 488 **Designing experimental tests of niche construction**

489 It is not a trivial task to determine whether or not organism-mediated environmental  
 490 modifications can alter selection pressures and subsequently drive an evolutionary response,  
 491 and it is likely best addressed by experimental tests (MacColl, 2011; Barrett and Hoekstra,  
 492 2011). In general, it is much easier to measure the strength and form of natural selection  
 493 (Hoekstra et al., 2001; Siepielski et al., 2009) than to determine the underlying causes (i.e.  
 494 agents) (Wade and Kalisz, 1990; MacColl, 2011) and eventual outcomes (Barrett and  
 495 Hoekstra, 2011). It is even difficult to identify the agents of selection in nature for  
 496 well-described polymorphic traits with a known genetic basis (MacColl, 2011). The  
 497 challenge partly stems from interactions among multiple selective agents that can lead to  
 498 complex fitness landscapes where selection differentials are a function of multiple axes of  
 499 modifiable or un-modifiable environmental conditions (Wade and Kalisz, 1990; MacColl,

500 2011). The most convincing experimental studies are those that manipulate putative  
 501 agents of selection and measure the consequences for the strength of selection (MacColl,  
 502 2011; Barrett and Hoekstra, 2011). Even more persuasive, and decidedly rare, are  
 503 experiments that simultaneously manipulate both the agent of selection and the target of  
 504 selection (Lankau and Strauss, 2007). As a first approach, one could carry out targeted  
 505 experiments in which the biomass or dynamics of a potential niche constructor is  
 506 manipulated and evolutionary responses are monitored in a recipient population  
 507 (Odling-Smee et al., 2003). To illustrate this idea, we propose the following series of  
 508 questions as a guide for future experimental tests of evolution by niche construction.

509 *Question 1: What is the effect-size distribution of organism-mediated*  
 510 *environmental effects?*

511 It is useful to quantify the distribution of organisms' environmental effects (Criterion 1)  
 512 because niche construction is more likely to occur if such effects are not too weak, too  
 513 diffuse, or too transient to cause a detectable change in selection. Common gardening  
 514 experiments (Matthews et al., 2011b) that are conducted in outdoor experimental  
 515 ecosystems that are either self-contained (Harmon et al., 2009; Matthews et al., 2011a) or  
 516 located in situ (Palkovacs and Post, 2009) are particularly useful for quantifying the  
 517 distribution of organisms' environmental effects. Such experiments are designed to



518 investigate how an organism modifies its environment, either relative to the absence of the  
 519 organism or relative to how another organism modifies the same environment. By  
 520 analyzing time-series of multiple environmental metrics in replicate ecosystems (that start  
 521 with identical conditions), one can disentangle the environmental effects of a putative niche  
 522 constructor from external forcing by temperature, rainfall, or incident radiation (Matthews  
 523 et al., 2011*a*). This is possible for experiments in which the organism causing the  
 524 environmental modification is either present or absent, and in designs where the biomass of  
 525 the organism is kept constant but its phenotype or genotype varies among treatments  
 526 (Harmon et al., 2009; Matthews et al., 2011*a*). A more elaborate experimental design  
 527 would be to manipulate the niche-constructing activities of an organism, while still keeping  
 528 the organisms in the system. This might be possible by routinely removing structures  
 529 created by the organism, or by homogenizing some aspect of the environment that the  
 530 organism modifies and that is thought to affect selection pressures. Such experiments  
 531 would require a detailed knowledge about both the traits underlying the environmental  
 532 modification, and about how variation in fitness of the recipient organism is aligned with  
 533 the modifiable environmental conditions.

534 Quantifying how organisms differentially affect their environment (e.g. Figure 4A) might  
 535 help predict how they shape selection pressures and drive evolutionary responses  
 536 (Odling-Smee et al., 2003, 2013). One possibility is that organisms may differentially  
 537 modify multiple axes of environmental variation so as to increase the dimensionality of



538 selection regimes and strengthen divergent selection (Nosil et al., 2009). Another  
 539 possibility is that organisms narrow the range of environmental conditions experienced by  
 540 the organism and impose stabilizing selection, which could happen by habitat choice  
 541 (Donohue et al., 2005) or by physical manipulation of the environment that buffers the  
 542 evolutionary response of populations to external environmental drivers (Turner, 2002;  
 543 Laland and Brown, 2006).

544 *Question 2: How persistent through time are organism-mediated*  
 545 *environmental effects?*

546 The environmental effects of organisms range from trivial modifications that dissipate  
 547 quickly, to long lasting habitat modifications that persist beyond the lifetime of the  
 548 organism (Odling-Smee et al., 2003; Hastings et al., 2007; Jones, 2012). Persistence time  
 549 can be measured in a simple common gardening experiment by extending the design  
 550 proposed in Figure 4A to include a phase in which the niche constructor is removed (Figure  
 551 4B). Upon removal of the niche constructor, persistence time is the interval over which one  
 552 can statistically differentiate the modified and unmodified ecosystems (Figure 4B). This  
 553 metric is analogous to quantifying the rate of ecosystem recovery to a pulsed stressor (i.e. a  
 554 putative niche constructor), which is often measured in experimental tests of ecosystem  
 555 resilience (Cottingham and Carpenter, 1994). Persistence is closely related to the concept

556 of ecological inheritance (Odling-Smee et al., 2003), which posits that organisms not only  
 557 transmit genes to subsequent generations, but also leave a legacy of environmental  
 558 modification that can affect selective pressures beyond their own lifetime. Ecosystem  
 559 engineers, for example, can affect environments over a very broad range of spatial and  
 560 temporal scales (Hastings et al., 2007), allowing ample opportunity for evolutionary effects  
 561 to occur. We are unaware of any experimental tests of how the ecosystem engineering  
 562 activities of organisms can alter selection pressures and drive evolutionary responses.

563 *Question 3: Do modifiable components of the environment affect selection*  
 564 *pressures and evolutionary responses?*

565 Both selection experiments and experimental evolution trials are useful to test how  
 566 organism-mediated environmental modifications might influence the environmental sources  
 567 of selection and drive evolutionary responses (MacColl, 2011; Barrett and Hoekstra, 2011).  
 568 Selection experiments can test whether heritable phenotypic changes within a population  
 569 are caused by a particular environmental modification, and are well suited for testing  
 570 criterion 2. Experimental evolution trials performed over one or more generations can test  
 571 for evolutionary responses to selection, and are well suited for testing criterion 3. Designing  
 572 robust experiments to test criteria 2 and 3 is not trivial, because it requires that the  
 573 ecosystem modification caused by the niche constructor is the reason for a particular

574 evolutionary response.

575 One robust experimental approach for testing criterion 2 and 3, is to do a common  
 576 gardening experiment, with treatments that manipulate either the abundance or modifying  
 577 activities of a niche constructor, followed by either a selection experiment (Criterion 2) or  
 578 an experimental evolution trial (Criterion 3) within the same set of experimental  
 579 ecosystems. The common gardening experiment would reveal the effect size distribution of  
 580 organism-mediated environmental modifications (Figure 4A). The selection phase of the  
 581 experiment would specifically test for niche construction, and reveal whether selection  
 582 pressures on a recipient organism differed among treatments in the common gardening  
 583 phase (Figure 4C). Alternatively (or additionally), an experimental evolution trial could be  
 584 performed in the modified environment in order to assess if evolutionary responses in a  
 585 recipient population differed among treatments, providing evidence that niche construction  
 586 led to alternative evolutionary outcomes (Figure 4C). In practice, working with relatively  
 587 isolated and controlled ecosystems (e.g. mesocosms) affords the opportunity to monitor  
 588 evolutionary changes in recipient populations over time.

589 A potentially more practical approach for testing criteria 2 and 3, is to experimentally  
 590 manipulate environmental factors that are known to be modifiable by a putative niche  
 591 constructor (e.g. emulate the physical conditions affected by an engineer) and perform  
 592 selection experiments and experimental evolution trials under these manipulated  
 593 conditions. For example, there is experimental evidence that Trinidadian guppies adapted

594 to different predation regimes (Reznick and Endler, 1982) can alter the flux of nutrients in  
 595 streams and have different effects on algal growth (Palkovacs and Post, 2009; Bassar et al.,  
 596 2010, 2012). Odling-Smee et al. (2013) hypothesized that changes in algal biomass might  
 597 alter the distribution of dietary algal pigments that influence the coloration of male  
 598 guppies, which can subsequently affect either sexual selection or predator mediated  
 599 selection pressures. In order to test specific effect pathways in this system, one could mimic  
 600 the contrasting environmental effect of locally adapted guppies by manipulating the level of  
 601 nutrients in the system. Such an approach is eminently more feasible than common  
 602 gardening experiments in which variation in the density, genotype or phenotype of the  
 603 putative niche constructor (e.g. guppies) is used to modify the environment directly (see  
 604 Figure 4A). One drawback, however, is that the experimentally modified environments may  
 605 lack realism and not reflect the subtleties of the modifying activities of the niche  
 606 constructor. Indeed, the foraging activities of organisms can shape community composition  
 607 and ecosystem properties in ways that might not be reproducible by direct manipulation  
 608 (Vanni et al., 1997; Schmitz, 2010). In some cases, ecosystem engineers can shape the  
 609 geometry of their physical environment in complex ways that might be impossible to  
 610 recreate by experimental manipulation (Jones, 2012), but in other cases the effects of  
 611 engineers on physical habitat structure can be mimicked in an experimental setting  
 612 (Crooks and Khim, 1999; Lill and Marquis, 2003). Regardless, using artificially modified  
 613 environments to mimic the effect of a selective agent might lead to associations between

614 fitness and phenotype that do not reflect a realistic set of environmental conditions. For  
 615 this same reason, many laboratory manipulations of selection pressures have led to  
 616 misleading conclusions about the associations between genotype, phenotype, and fitness  
 617 (Barrett and Hoekstra, 2011).

618 Another complication with testing the third criterion is the need to demonstrate an  
 619 evolutionary response in a recipient population using a natural range of phenotypic and  
 620 genetic variation. Ideally, one should work with the standing genetic variation that is  
 621 present in a population of a recipient. This stringent condition is justified by the  
 622 prevalence of genotype x environment interactions and genotype x genotype x environment  
 623 interactions (Barrett and Hoekstra, 2011). Careful consideration of the genetic background  
 624 of the recipient is a critical step in both selection experiments and experimental evolution  
 625 trials, because putative adaptive alleles in one genetic background can produce different  
 626 fitness effects in another genetic background and fundamentally change selection  
 627 coefficients. Furthermore, epistatic interactions between genes can differ among  
 628 populations, and the environmental conditions in which they evolve might influence the  
 629 relationship between phenotype and fitness in a novel environment (Barrett and Hoekstra,  
 630 2011).

631 Overall, testing the wide range of potential niche construction mechanisms is best achieved  
 632 by an integrative research effort that combines comparative and experimental approaches.  
 633 In the following section we outline a model system for testing niche construction theory

634 that has been extensively studied by aquatic ecologists but has received comparably little  
 635 attention from evolutionary biologists.

636 **A case study: Alternative stable states in shallow lake**  
 637 **ecosystems**

638 The presence of alternate stable states is the main explanation for sudden and dramatic  
 639 shifts observed in terrestrial, marine and inland water ecosystems (Scheffer et al., 2001).

640 Considerable research has been directed towards understanding the mechanistic basis of  
 641 tipping points between states (Scheffer and Carpenter, 2003; Carpenter et al., 2011). Shifts  
 642 in ecosystem state often occur because a specific group of organisms that has a stabilizing  
 643 effect on environmental conditions (e.g. trees on microclimate in a forest) is overwhelmed  
 644 by some environmental stressor (e.g. drought, exploitation), leading to dramatic changes in  
 645 both community composition and environmental conditions. Here, we use one of the  
 646 best-studied regime shifts, namely that between the turbid and clear-water state in shallow  
 647 lakes (Scheffer and Carpenter, 2003), to illustrate the existing evidence for niche  
 648 construction and exciting avenues for future research.

649 Organisms in shallow lakes modify the abiotic and biotic environment in multiple ways  
 650 that can influence the transition between stable states (i.e. clear and turbid states).

651 Macrophytes, for example (Figure 5), act as ecosystem engineers in shallow lakes because

652 their roots stabilize sediments and reduce phosphorus recycling to phytoplankton, and  
 653 their canopies reduce turbulent mixing and attenuate light availability (Byers et al., 2006).  
 654 There is both comparative (Scheffer et al., 2001) and experimental (Declerck et al., 2007)  
 655 evidence for the strong impact of macrophytes on the relationship between phosphorus  
 656 concentration and phytoplankton biomass (paralleling Figure 3A), and, as a result, in the  
 657 maintenance of the clear water state. Macrophytes can also affect the transition between  
 658 different states in shallow lakes by modifying trophic interactions (Byers et al., 2006). In  
 659 shallow lakes and ponds, omnivorous fish can dramatically modify the environment by  
 660 uprooting macrophytes, re-suspending sediments, and increasing turbidity levels (Drenner  
 661 et al., 1998). More generally, there is overwhelming comparative (Jeppesen et al., 1997)  
 662 and compelling experimental evidence (Carpenter and Kitchell, 1993; Vanni et al., 1997)  
 663 that fish in aquatic systems have important impacts on prey community composition and  
 664 size structure, particularly through their effects on large bodied cladocerans that play a  
 665 pivotal role in the cascading trophic interactions that influence phytoplankton abundance  
 666 (Carpenter and Kitchell, 1993). Mesocosm studies have demonstrated that the presence of  
 667 the large-bodied cladoceran *Daphnia magna* can have a significant impact on the  
 668 community composition of phyto- and bacterioplankton (Verreydt et al., 2012), and that  
 669 the genotype of *Daphnia magna* can impact the community composition of zooplankton  
 670 (De Meester et al., 2007).  
 671 Despite a plethora of evidence for the effects of organisms on the environmental conditions



672 of shallow lakes, so far no studies have specifically tested for evolution by niche  
 673 construction. Macrophytes are good candidates for being niche constructors because of  
 674 their strong impacts on the abiotic and biotic conditions of shallow-lake ecosystems and  
 675 their central role in mediating the transition between alternative stable states. In  
 676 particular, there are many potential ways that macrophytes might influence evolutionary  
 677 responses through their modification of abiotic conditions (Figure 5). For example,  
 678 macrophytes might cause evolutionary responses in phytoplankton by directly  
 679 manipulating the light and nutrient environment of shallow lakes (Collins and Bell, 2004;  
 680 Stomp et al., 2004), or, alternatively, by providing habitat for zooplankton communities  
 681 that graze on phytoplankton. Differences in grazing pressure might explain the genetic  
 682 differentiation in the size and number of cells in colonies of the planktonic alga  
 683 *Desmodesmus armatus* isolated from a neighbouring turbid and clear-water system  
 684 (Vanormelingen et al., 2009). Macrophytes can also provide refugia for prey species that  
 685 are vulnerable to visually foraging predators. In recent work combining paleolimnology  
 686 with quantitative genetics, where organisms can be resurrected from resting stages  
 687 preserved in lake sediments, there is evidence for the adaptation of *Daphnia* phototactic  
 688 behaviour in response to changes in fish predation pressure (Cousyn et al., 2001). Similarly,  
 689 the genetic adaptation in pigmentation of *Asellus aquaticus*, a common freshwater  
 690 crustacean of shallow lakes, might be explained by a combined response to  
 691 macrophyte-mediated changes to habitat structure, light environments, and predation



692 pressure (Hargeby et al., 2005). While Figure 5 is not exhaustive, it highlights the  
 693 potential for one prominent group of organisms (i.e. macrophytes) to cause evolutionary  
 694 effects by altering abiotic conditions of shallow-lake ecosystems.

695 There are well-established experimental designs to test for the presence of alternative  
 696 stable states (Schröder et al., 2005), and combining these with selection experiments and  
 697 experimental evolution trials (e.g. Figure 4) would help uncover the specific mechanisms  
 698 underlying how species interactions and organism-mediated changes to shallow-lake  
 699 ecosystems can influence selection pressures and drive evolutionary responses. In  
 700 particular, such research might offer new insights into which modifications to the abiotic  
 701 environment might persist and influence selection pressures through time (i.e. ecological  
 702 inheritance), and where such effects might lead to evolutionary responses (i.e. evolution by  
 703 niche construction). In aquatic mesocosms, one could use a common gardening experiment  
 704 to establish alternate stable states in replicated experimental mesocosms by directly  
 705 manipulating macrophytes, nutrients, and fish. To test criterion 2, one could measure  
 706 selection gradients of a target organism inhabiting both ecosystem states. To test criterion  
 707 3, one could track changes in the phenotype and genotype of short-lived organisms (e.g.  
 708 phytoplankton) through time in both clear and turbid states. To perform experiments at a  
 709 larger scale, one could also capitalize on whole-lake manipulations where fish are removed  
 710 in order to create opportunities for the establishment of macrophytes. In such cases, one  
 711 could monitor evolutionary responses of organisms at different trophic levels over time. In

712 sun, shallow lakes offer a model system with many uncharted dimensions to explore in the  
 713 context of niche construction. A fundamental question to address is how evolution by niche  
 714 construction might stabilize or destabilize equilibrium states in systems that show regime  
 715 shifts.

716 **Back to nature: some further challenges**

717 While experimental approaches allow for strong tests of certain aspects of niche  
 718 construction theory, it remains a monumental challenge to identify the importance of  
 719 evolution by niche construction in nature. One can make progress by building on existing  
 720 studies of keystone species, ecosystem engineers, and the newly emerging model systems in  
 721 eco-evolutionary dynamics, but there are numerous complexities to consider. At the level  
 722 of niche constructors, environmental modifications that influence selection might be driven  
 723 by the combined action of multiple species, rather than any single species. Here, we only  
 724 consider niche construction to operate if the environmental modification leading to altered  
 725 selection pressures is attributable to particular organisms. At the level of the recipients of  
 726 niche construction, it is possible that there are no species with the evolutionary potential  
 727 to respond to organism-modified environmental conditions (Vincent and Brown, 2005) or  
 728 that an ecological response will preempt any evolutionary responses (Urban et al., 2008).  
 729 This latter point highlights an important conceptual link between niche construction and

730 evolving metacommunities (Urban et al., 2008). Following an organism-mediated change to  
 731 the environment, a community may change its average trait values (e.g. body size) by a  
 732 combination of changes in the relative abundance of species (cf. species sorting) and  
 733 evolutionary changes in the species that make up the community (Urban et al., 2008).  
 734 Evolution by niche construction is only present if the latter outcome occurs.

735 We believe that the testing of niche construction theory is still in its infancy and that the  
 736 approaches we advocate will lead to greater integration among related disciplines (Figure  
 737 1). There are ample examples in which species modify the environment, and where  
 738 environmental change alters selection pressures and induces evolutionary responses in focal  
 739 species, but only a handful of studies that show all of these aspects in the same system,  
 740 and even fewer that test a mechanistic link between evolutionary responses and the  
 741 environmental modifying activities of organisms. There are numerous descriptive cases of  
 742 niche construction and some intriguing experimental tests (e.g. Table 1), but there are  
 743 many plausible mechanisms of environmental modification (Figure 2) and numerous  
 744 organisms that could act as putative niche constructors and recipients of niche construction  
 745 (Table 1). Intriguingly, there are also numerous well-developed model systems that provide  
 746 exciting avenues for both evolutionary biologists and ecologists to explore niche  
 747 construction dynamics. Indeed, we are well poised to elucidate the network of interactions  
 748 between niche constructors and their environment, and to assess the importance of niche  
 749 construction in explaining ecological and evolutionary changes in nature.

750 **Acknowledgments**

751 This paper is a product of the Working Group "Ecosystem Engineering and Evolution"  
 752 convened at the Centre for Estuarine and Marine Ecology, Netherlands Institute of Ecology  
 753 (NIOO), Yerseke, The Netherlands. Workshops were organized by Tjeerd Bouma, Clive  
 754 Jones, and Peter Herman and financially supported by KNAW grant ISK/6445/VPP and  
 755 the NIOO. We thank Joke van Houte, Anneke van der Endt, Elly van Hulsteijn, Christine  
 756 van der Jagt, Laura Wisse and Jan Megens for logistical support of the workshops. We also  
 757 thank Laurent Lehmann, Monique de Jager, Peter Herman, and Carlos Melian for useful  
 758 discussions and comments on the manuscript. We particularly thank Kevin Laland for  
 759 helping us revise the manuscript. CGJ thanks The Royal Netherlands Academy for Arts  
 760 and Sciences and The Cary Institute of Ecosystem Studies for financial support.  
 761 Contribution to the program of the Cary Institute of Ecosystem Studies. BM was  
 762 supported by SNF grant number 31003A-125006, and LDM by KU Leuven Research Fund  
 763 Excellence Center PF/2010/007.

764 **References**

- 765 Bailey, R., K. Schönrogge, J. Cook, and G. Melika. 2009. Host Niches and Defensive  
 766 Extended Phenotypes Structure Parasitoid Wasp Communities. *PLoS Biology* **7**:e1000179.
- 767 Barrett, R. D. H., and H. E. Hoekstra. 2011. Molecular spandrels: tests of adaptation at  
 768 the genetic level. *Nature Reviews Genetics* **12**:767–780.
- 769 Bassar, R. D., R. Ferriere, A. López-Sepulcre, M. C. Marshall, J. Travis, C. M. Pringle, and  
 770 D. N. Reznick. 2012. Direct and Indirect Ecosystem Effects of Evolutionary Adaptation  
 771 in the Trinidadian Guppy (*Poecilia reticulata*). *The American Naturalist* **180**:167–185.
- 772 Bassar, R. D., M. C. Marshall, A. Lopez-Sepulcre, E. Zandonà, S. K. Auer, J. Travis, C. M.  
 773 Pringle, A. S. Flecker, S. A. Thomas, D. F. Fraser, and D. N. Reznick. 2010. Local  
 774 adaptation in Trinidadian guppies alters ecosystem processes. *Proceedings of the National*  
 775 *Academy of Sciences of the United States of America* **107**:3616–3621.
- 776 Becks, L., S. P. Ellner, L. E. Jones, and J. N. G. Hairston. 2012. The functional genomics of  
 777 an eco-evolutionary feedback loop: linking gene expression, trait evolution, and community  
 778 dynamics. *Ecol. Lett.* **15**:492–501.
- 779 Beerling, D. 2005. Leaf Evolution: Gases, Genes and Geochemistry. *Annals of Botany* **96**.
- 780 Benkman, C. W., T. L. Parchman, and E. T. Mezquida. 2010. Patterns of coevolution in  
 781 the adaptive radiation of crossbills. *Ann Ny Acad Sci* **1206**:1–16.

- 782 Bonduriansky, R. 2012. Rethinking heredity, again. *Trends in Ecology and Evolution*  
 783 **27**:330–336.
- 784 Bonduriansky, R., and T. Day. 2009. Nongenetic Inheritance and Its Evolutionary  
 785 Implications. *Annual Review of Ecology and Systematics* **40**:103–125.
- 786 Boogert, N., D. Paterson, and K. Laland. 2006. The Implications of Niche Construction and  
 787 Ecosystem Engineering for Conservation Biology. *Bioscience* **56**:570.
- 788 Bouma, T. J., M. B. De Vries, and P. M. J. Herman. 2010. Comparing ecosystem engineering  
 789 efficiency of two plant species with contrasting growth strategies. *Ecology* **91**:2696–2704.
- 790 Brodie III, E. 2005. Caution: Niche Construction Ahead. *Evolution* **59**:249–251.
- 791 Byers, J. E., K. Cuddington, C. G. Jones, T. S. Talley, A. Hastings, J. G. Lambrinos, J. A.  
 792 Crooks, and W. G. Wilson. 2006. Using ecosystem engineers to restore ecological systems.  
 793 *Trends in Ecology and Evolution* **21**:493–500.
- 794 Carpenter, S., and J. Kitchell. 1993. *The Trophic Cascade in Lakes*. Cambridge University  
 795 Press.
- 796 Carpenter, S. R., J. J. Cole, M. L. Pace, R. Batt, W. A. Brock, T. Cline, J. Coloso, J. R.  
 797 Hodgson, J. F. Kitchell, D. A. Seekell, L. Smith, and B. Weidel. 2011. Early Warnings of  
 798 Regime Shifts: A Whole-Ecosystem Experiment. *Science* **332**:1079–1082.

- 799 Chase, J. M., E. G. Biro, W. A. Ryberg, and K. G. Smith. 2009. Predators temper the  
 800 relative importance of stochastic processes in the assembly of prey metacommunities. *Ecol.*  
 801 *Lett.* **12**:1210–1218.
- 802 Collins, S., and G. Bell. 2004. Phenotypic consequences of 1,000 generations of selection at  
 803 elevated CO<sub>2</sub> in a green alga. *Nature* **431**:566–569.
- 804 Corenblit, D., J. Steiger, A. Gurnell, and R. J. Naiman. 2009. Plants intertwine fluvial  
 805 landform dynamics with ecological succession and natural selection: a niche construction  
 806 perspective for riparian systems. *Global Ecology and Biogeography* **18**:507–520.
- 807 Cottingham, K., and S. Carpenter. 1994. Predictive Indexes of Ecosystem Resilience in  
 808 Models of North Temperate Lakes. *Ecology* **75**:2127–2138.
- 809 Cousyn, C., L. De Meester, J. K. Colbourne, L. Brendonck, D. Verschuren, and F. Volckaert.  
 810 2001. Rapid, local adaptation of zooplankton behavior to changes in predation pressure in  
 811 the absence of neutral genetic changes. *Proceedings of the National Academy of Sciences*  
 812 *of the United States of America* **98**:6256–6260.
- 813 Crespi, B. 2004. Vicious circles: positive feedback in major evolutionary and ecological  
 814 transitions. *Trends in Ecology and Evolution* **19**:627–633.
- 815 Crooks, J. A., and H. S. Khim. 1999. Architectural vs. biological effects of a habitat-altering,



- 816 exotic mussel, *Musculista senhousia*. *Journal of Experimental Marine Biology and Ecology*  
 817 **240**:53–75.
- 818 Danchin, É., A. Charmantier, F. A. Champagne, A. Mesoudi, B. Pujol, and S. Blanchet.  
 819 2011. Beyond DNA: integrating inclusive inheritance into an extended theory of evolution.  
 820 *Nature Reviews Genetics* **12**:475–486.
- 821 Dawkins, R. 1982. *Dawkins: Organisms, groups and memes: Replicators...* - Google Scholar.  
 822 *The extended phenotype* Oxford University Press .
- 823 Dawkins, R. 2004. Extended phenotype—but not too extended. A reply to Laland, Turner  
 824 and Jablonka. *Biology and Philosophy* **19**:377–396.
- 825 De Meester, L., G. Louette, C. Duvivier, C. Van Damme, and E. Michels. 2007. Genetic  
 826 composition of resident populations influences establishment success of immigrant species.  
 827 *Oecologia* **153**:431–440.
- 828 Declerck, S., M. Vanderstukken, A. Pals, K. Muylaert, and L. De Meester. 2007. Plankton  
 829 biodiversity along a gradient of productivity and its mediation by macrophytes. *Ecology*  
 830 **88**:2199–2210.
- 831 Dickman, E. M., J. M. Newell, M. J. González, and M. J. Vanni. 2008. Light, nutrients, and  
 832 food-chain length constrain planktonic energy transfer efficiency across multiple trophic  
 833 levels. *Proceedings of the National Academy of Sciences* **105**:18408–18412.

- 834 Donohue, K., C. R. Polisetty, and N. J. Wender. 2005. Genetic Basis and Consequences  
 835 of Niche Construction: Plasticity Induced Genetic Constraints on the Evolution of Seed  
 836 Dispersal in *Arabidopsis thaliana*. *The American Naturalist* **165**:537–550.
- 837 Drenner, R. W., K. L. Gallo, and R. M. Baca. 1998. Synergistic effects of nutrient loading and  
 838 omnivorous fish on phytoplankton biomass. *Canadian Journal of Fisheries and Aquatic  
 839 Sciences* **55**:2087–2096.
- 840 Ellers, J., E. T. Kiers, C. Currie, and B. McDonald. 2012. Ecological interactions drive  
 841 evolutionary loss of traits. *Ecol. Lett.* **15**:1071–1082.
- 842 Ellner, S. P., M. A. Geber, and J. N. G. Hairston. 2011. Does rapid evolution matter?  
 843 Measuring the rate of contemporary evolution and its impacts on ecological dynamics.  
 844 *Ecol. Lett.* **14**:603–614.
- 845 Endler, J. 1986. *Natural Selection in the Wild*. Princeton University Press, New Jersey.
- 846 Erwin, D., and S. Tweedt. 2011. Ecological drivers of the Ediacaran-Cambrian diversification  
 847 of Metazoa. *Evolutionary Ecology* **26**:417–433.
- 848 Erwin, D. H. 2008. Macroevolution of ecosystem engineering, niche construction and  
 849 diversity. *Trends in Ecology and Evolution* **23**:304–310.
- 850 Fisher, R. A. 1930. *The genetical theory of natural selection*. Clarendon Press.

- 851 Flack, J. C., M. Girvan, F. B. M. de Waal, and D. C. Krakauer. 2006. Policing stabilizes  
 852 construction of social niches in primates. *Nature* **439**:426–429.
- 853 Forbes, A., T. Powell, L. Stelinski, and J. Smith. 2009. Sequential Sympatric Speciation  
 854 Across Trophic Levels. *Science* **323**:776–779.
- 855 Frank, S. A. 2009. Natural selection maximizes Fisher information. *Journal of Evolutionary*  
 856 *Biology* **22**:231–244.
- 857 Fussmann, G. F., M. Loreau, and P. A. Abrams. 2007. Eco-evolutionary dynamics of  
 858 communities and ecosystems. *Functional Ecology* **21**:465–477.
- 859 Goddard, M. R. 2008. Quantifying the complexities of *Saccharomyces cerevisiae*'s ecosystem  
 860 engineering via fermentation. *Ecology* **89**:2077–2082.
- 861 Goodman, B. A., and P. T. J. Johnson. 2011. Disease and the Extended Phenotype: Parasites  
 862 Control Host Performance and Survival through Induced Changes in Body Plan. *PLoS*  
 863 *ONE* **6**:e20193.
- 864 Grant, P., and B. Grant. 2006. Evolution of character displacement in Darwin's finches.  
 865 *Science* **313**:224–226.
- 866 Hairston, J. N. G., S. P. Ellner, M. A. Geber, T. Yoshida, and J. A. Fox. 2005. Rapid  
 867 evolution and the convergence of ecological and evolutionary time. *Ecol. Lett.* **8**:1114–1127.

- 868 Haloin, J. R., and S. Y. Strauss. 2008. Interplay between Ecological Communities and  
 869 Evolution. *Annals of the New York Academy of Sciences* **1133**:87–125.
- 870 Hansen, M. M., I. Olivieri, D. M. Waller, E. E. Nielsen, and GeM Working Group. 2012.  
 871 Monitoring adaptive genetic responses to environmental change. *Molecular Ecology*  
 872 **21**:1311–1329.
- 873 Hargeby, A., J. Stoltz, and J. Johansson. 2005. Locally differentiated cryptic pigmentation  
 874 in the freshwater isopod *Asellus aquaticus*. *Journal of Evolutionary Biology* **18**:713–721.
- 875 Harmon, L. J., B. Matthews, S. Des Roches, J. M. Chase, J. B. Shurin, and D. Schluter.  
 876 2009. Evolutionary diversification in stickleback affects ecosystem functioning. *Nature*  
 877 **458**:1167–1170.
- 878 Hastings, A., J. E. Byers, J. A. Crooks, K. Cuddington, C. G. Jones, J. G. Lambrinos, T. S.  
 879 Talley, and W. G. Wilson. 2007. Ecosystem engineering in space and time. *Ecol. Lett.*  
 880 **10**:153–164.
- 881 Hendrix, P. F. 2006. Biological invasions belowground—earthworms as invasive species.  
 882 Springer Netherlands, Dordrecht.
- 883 Hendry, A., and M. Kinnison. 1999. Perspective: The pace of modern life: Measuring rates  
 884 of contemporary microevolution. *Evolution* **53**:1637–1653.
- 885 Hoekstra, H. E., J. M. Hoekstra, D. Berrigan, S. N. Vignieri, A. Hoang, C. E. Hill,

- 886 P. Beerli, and J. G. Kingsolver. 2001. Strength and tempo of directional selection in the  
 887 wild. *Proceedings of the National Academy of Sciences of the United States of America*  
 888 **98**:9157–9160.
- 889 Holt, R., and J. H. Lawton. 1994. The ecological consequences of shared natural enemies.  
 890 *Annual Review of Ecology and Systematics* **25**:495–520.
- 891 Hoover, J. P., and S. K. Robinson. 2007. Retaliatory mafia behavior by a parasitic cowbird  
 892 favors host acceptance of parasitic eggs. *Proceedings of the National Academy of Sciences*  
 893 of the United States of America **104**:4479–4483.
- 894 Hoover, K., M. Grove, M. Gardner, D. P. Hughes, J. McNeil, and J. Slavicek. 2011. A gene  
 895 for an extended phenotype. *Science* **333**:1401.
- 896 Jeppesen, E., J. P. Jensen, M. Sondergaard, T. Lauridsen, L. Pedersen, and L. Jensen. 1997.  
 897 Top-down control in freshwater lakes: the role of nutrient state, submerged macrophytes  
 898 and water depth. *Hydrobiologia* **342**:151–164.
- 899 Jones, C. G. 2012. Ecosystem engineers and geomorphological signatures in landscapes.  
 900 *Geomorphology* **157-158**:75–87.
- 901 Jones, C. G., J. H. Lawton, and M. Shachak. 1994. Organisms as ecosystem engineers. *Oikos*  
 902 **69**:373–386.

- 903 Jones, C. G., J. H. Lawton, and M. Shachak. 1997. Positive and negative effects of organisms  
 904 as physical ecosystem engineers. *Ecology* **78**:1946–1957.
- 905 Kerr, B., D. W. Schwilk, and A. Bergman. 1999. Rekindling an old flame: a haploid model  
 906 for the evolution and impact of flammability in resprouting plants. *Evolutionary Ecology*  
 907 **1**:807–833.
- 908 Kokko, H., and A. López-Sepulcre. 2007. The ecogenetic link between demography and  
 909 evolution: can we bridge the gap between theory and data? *Ecol. Lett.* **10**:773–782.
- 910 Krakauer, D. C., K. M. Page, and D. H. Erwin. 2009. Diversity, Dilemmas, and Monopolies  
 911 of Niche Construction. *The American Naturalist* **173**:26–40.
- 912 Kylafis, G., and M. Loreau. 2008. Ecological and evolutionary consequences of niche  
 913 construction for its agent. *Ecol. Lett.* **11**:1072–1081.
- 914 Kylafis, G., and M. Loreau. 2011. Niche construction in the light of niche theory. *Ecol. Lett.*  
 915 **14**:82–90.
- 916 Lafferty, K. D., S. Allesina, M. Arim, C. J. Briggs, G. De Leo, A. P. Dobson, J. A. Dunne,  
 917 P. T. J. Johnson, A. M. Kuris, D. J. Marcogliese, N. D. Martinez, J. Memmott, P. A.  
 918 Marquet, J. P. McLaughlin, E. A. Mordecai, M. Pascual, R. Poulin, and D. W. Thieltges.  
 919 2008. Parasites in food webs: the ultimate missing links. *Ecol. Lett.* **11**:533–546.

920 Laland, K. 2010. Niche construction, co-evolution and biodiversity. *Ecological economics*  
 921 **69**.

922 Laland, K. N., and G. R. Brown. 2006. Niche construction, human behavior, and the  
 923 adaptive-lag hypothesis. *Evol. Anthropol.* **15**:95–104.

924 Laland, K. N., and K. Sterelny. 2006. Seven reasons (not) to neglect niche construction.  
 925 *Evolution* **60**:1751–1762.

926 Lankau, R. A., and S. Y. Strauss. 2007. Mutual Feedbacks Maintain Both Genetic and  
 927 Species Diversity in a Plant Community. *Science* **317**:1561–1563.

928 Lehmann, L. 2008. The adaptive dynamics of niche constructing traits in spatially subdivided  
 929 populations: evolving posthumous extended phenotypes. *Evolution* **62**:549–566.

930 Lewontin, R. C., 1983. Gene, organism, and environment. Pages 273–285 *in* D. S. Bendall,  
 931 editor. *Evolution from Molecules to Men*. Cambridge University Press.

932 Lill, J. T., and R. J. Marquis. 2003. Ecosystem Engineering by Caterpillars Increases Insect  
 933 Herbivore Diversity on White Oak. *Ecology* **84**:682–690.

934 Loreau, M. 2010. *From Populations to Ecosystems: Theoretical Foundations for a New*  
 935 *Ecological Synthesis*. Monographs in Population Biology, Princeton University Press.

936 Losos, J. B. 2010. Adaptive Radiation, Ecological Opportunity, and Evolutionary  
 937 Determinism. *The American Naturalist* **175**:623–639.



- 938 MacColl, A. D. C. 2011. The ecological causes of evolution. *Trends in Ecology and Evolution*  
 939 **26**:514–522.
- 940 Matthews, B., S. Hausch, C. Winter, and C. Suttle. 2011*a*. Contrasting Ecosystem-Effects  
 941 of Morphologically Similar Copepods. *PLoS ONE* **6**:e26700.
- 942 Matthews, B., A. Narwani, S. Hausch, E. Nonaka, H. Peter, M. Yamamichi, K. E. Sullam,  
 943 K. C. Bird, M. K. Thomas, T. C. Hanley, and C. B. Turner. 2011*b*. Toward an integration  
 944 of evolutionary biology and ecosystem science. *Ecol. Lett.* **14**:690–701.
- 945 McIntyre, P. B., A. S. Flecker, M. J. Vanni, J. M. Hood, B. W. Taylor, and S. A. Thomas.  
 946 2008. Fish distributions and nutrient cycling in streams: can fish create biogeochemical  
 947 hotspots? *Ecology* **89**:2335–2346.
- 948 Meysman, F., J. J. Middelburg, and C. H. R. Heip. 2006. Bioturbation: a fresh look at  
 949 Darwin’s last idea. *Trends in Ecology and Evolution* **21**:688–695.
- 950 Mizuno, A. N., and M. Kawata. 2009. The effects of the evolution of stoichiometry-related  
 951 traits on population dynamics in plankton communities. *Journal of Theoretical Biology*  
 952 **259**:209–218.
- 953 Morran, L. T., O. G. Schmidt, I. A. Gelarden, R. C. I. Parrish, and C. M. Lively. 2011.  
 954 Running with the Red Queen: Host-Parasite Coevolution Selects for Biparental Sex.  
 955 *Science* **333**:216–218.

- 956 Mueller, U. G., and N. Gerardo. 2002. Fungus-farming insects: multiple origins and diverse  
 957 evolutionary histories. *Proceedings of the National Academy of Sciences of the United*  
 958 *States of America* **99**:15247–15249.
- 959 Nosil, P. 2012. *Ecological Speciation*. Oxford University Press.
- 960 Nosil, P., L. J. Harmon, and O. Seehausen. 2009. Ecological explanations for (incomplete)  
 961 speciation. *Trends in Ecology and Evolution* **24**:145–156.
- 962 Odling-Smee, F., K. Laland, and M. Feldman. 2003. *Niche Construction: The Neglected*  
 963 *Process in Evolution*. Princeton University Press.
- 964 Odling-Smee, F. J., K. N. Laland, and M. W. Feldman. 1996. Niche construction. *The*  
 965 *American Naturalist* **147**:641–648.
- 966 Odling-Smee, J., D. H. Erwin, E. P. Palcovacs, M. W. Feldman, and K. N. Laland. 2013.  
 967 Niche construction theory: a practical guide for ecologists. *Quarterly Review of Biology*  
 968 **88**:4–28.
- 969 Okasha, S. 2005. On niche construction and extended evolutionary theory. *Biology and*  
 970 *Philosophy* **20**:1–10.
- 971 Olf, H., D. Alonso, M. P. Berg, B. K. Eriksson, M. Loreau, T. Piersma, and N. Rooney.  
 972 2009. Parallel ecological networks in ecosystems. *Philosophical Transactions of the Royal*  
 973 *Society B- Biological Sciences* **364**:1755–1779.

- 974 Palkovacs, E., and D. Post. 2008. Eco-evolutionary interactions between predators and prey:  
 975 can predator-induced changes to prey communities feed back to shape predator foraging  
 976 traits? *Evolutionary Ecology Research* **10**:699–720.
- 977 Palkovacs, E. P., and D. M. Post. 2009. Experimental evidence that phenotypic divergence  
 978 in predators drives community divergence in prey. *Ecology* **90**:300–305.
- 979 Pearce, T. 2011. Ecosystem engineering, experiment, and evolution. *Biology and Philosophy*  
 980 **26**:793–812.
- 981 Post, D. M., and E. P. Palkovacs. 2009. Eco-evolutionary feedbacks in community and  
 982 ecosystem ecology: interactions between the ecological theatre and the evolutionary play.  
 983 *Philosophical Transactions of the Royal Society B- Biological Sciences* **364**:1629–1640.
- 984 Reznick, D., and J. A. Endler. 1982. The impact of predation on life history evolution in  
 985 Trinidadian guppies (*Poecilia Reticulata*). *Evolution* **36**:160–177.
- 986 Roughgarden, J. 1976. Resource partitioning among competing species—a coevolutionary  
 987 approach. *Theoretical Population Biology* **9**:388–424.
- 988 Scheffer, M., S. Carpenter, J. A. Foley, C. Folke, and B. Walker. 2001. Catastrophic shifts  
 989 in ecosystems. *Nature* **413**:591–596.
- 990 Scheffer, M., and S. R. Carpenter. 2003. Catastrophic regime shifts in ecosystems: linking  
 991 theory to observation. *Trends in Ecology and Evolution* **18**:648–656.

- 992 Schluter, D. 2000. *The Ecology of Adaptive Radiation*. Oxford University Press.
- 993 Schluter, D. 2009. Evidence for ecological speciation and its alternative. *Science*  
 994 **323**:737–741.
- 995 Schmitz, O. 2010. *Resolving Ecosystem Complexity*. Princeton University Press, New Jersey.
- 996 Schoener, T. W. 2011. The newest synthesis: understanding the interplay of evolutionary  
 997 and ecological dynamics. *Science* **331**:426–429.
- 998 Schröder, A., L. Persson, and A. M. de Roos. 2005. Direct experimental evidence for  
 999 alternative stable states: a review. *Oikos* **110**:3–19.
- 1000 Siepielski, A. M., J. D. DiBattista, and S. M. Carlson. 2009. It's about time: the temporal  
 1001 dynamics of phenotypic selection in the wild. *Ecol. Lett.* **12**:1261–1276.
- 1002 Sterner, R., and J. Elser. 2002. *Ecological stoichiometry: the biology of elements from*  
 1003 *molecules to the ecosystem*. Princeton University Press.
- 1004 Stinchcombe, J., and M. Rausher. 2002. The evolution of tolerance to deer herbivory:  
 1005 modifications caused by the abundance of insect herbivores. *Proceedings of the Royal*  
 1006 *Society B: Biological Sciences* **269**:1241–1246.
- 1007 Stomp, M., J. Huisman, F. de Jongh, A. J. Veraart, D. Gerla, M. Rijkeboer, B. W. Ibelings,  
 1008 U. I. A. Wollenzien, and L. J. Stal. 2004. Adaptive divergence in pigment composition  
 1009 promotes phytoplankton biodiversity. *Nature* **432**:104–107.

- 1010 Thompson, J. 2005. *The Geographic Mosaic Of Coevolution*. University of Chicago Press.
- 1011 Thompson, J. N. 1998. Rapid evolution as an ecological process. *Trends in Ecology and*  
 1012 *Evolution* **13**:329–332.
- 1013 Turner, J. S. 2002. *The Extended Organism: The Physiology of Animal-Built Structures*.  
 1014 Harvard University Press.
- 1015 Urban, M. C., M. Leibold, P. Amarasekare, L. DeMeester, R. Gomulkiewicz, M. E. Hochberg,  
 1016 C. A. Klausmeier, N. Loeuille, C. Demazancourt, and J. Norberg. 2008. The evolutionary  
 1017 ecology of metacommunities. *Trends in Ecology and Evolution* **23**:311–317.
- 1018 Van Dyken, J. D., and M. J. Wade. 2012. Origins of Altruism Diversity Ii: Runaway  
 1019 Coevolution of Altruistic Strategies via “Reciprocal Niche Construction”. *Evolution*  
 1020 **66**:2498–2513.
- 1021 Vanni, M. J., C. D. Layne, and S. E. Arnott. 1997. “Top–Down” Trophic Interactions in  
 1022 Lakes: Effects of Fish on Nutrient Dynamics. *Ecology* **78**:1–20.
- 1023 Vanormelingen, P., W. Vyverman, D. De Bock, K. Van der Gucht, and L. De Meester. 2009.  
 1024 Local genetic adaptation to grazing pressure of the green alga *Desmodesmus armatus* in  
 1025 a strongly connected pond system. *Limnology and Oceanography* **54**:503.
- 1026 Verreydt, D., L. De Meester, E. Decaestecker, M.-J. Villena, K. Van Der Gucht,  
 1027 P. Vannormelingen, W. Vyverman, and S. A. J. Declerck. 2012. Dispersal-mediated

1028 trophic interactions can generate apparent patterns of dispersal limitation in aquatic  
 1029 metacommunities. *Ecol. Lett.* **15**:218–226.

1030 Vincent, T., and J. Brown. 2005. *Evolutionary Game Theory, Natural Selection, and*  
 1031 *Darwinian Dynamics*. Cambridge University Press, Cambridge.

1032 Wade, M. J., and S. Kalisz. 1990. The causes of natural selection. *Evolution* **44**:1947–1955.

1033 Walsh, M. R. 2013. The evolutionary consequences of indirect effects. *Trends in Ecology*  
 1034 *and Evolution* **28**:23–29.

1035 Walsh, M. R., J. P. DeLong, T. C. Hanley, and D. M. Post. 2012. A cascade of evolutionary  
 1036 change alters consumer-resource dynamics and ecosystem function. *Proceedings of the*  
 1037 *Royal Society B: Biological Sciences* **279**:3184–3192.

1038 Werner, E. E., and S. Peacor. 2003. A review of trait-mediated indirect interactions in  
 1039 ecological communities. *Ecology* **84**:1083–1100.

1040 Wright, J., P. Gribben, and J. Byers. 2012. Invasive Ecosystem Engineer Selects for Different  
 1041 Phenotypes of an Associated Native Species. *Ecology* **93**:1262–1268.

1042 Wright, J. P., and C. G. Jones. 2006. The Concept of Organisms as Ecosystem Engineers  
 1043 Ten Years On: Progress, Limitations, and Challenges. *Bioscience* **56**:203.

1044 Yoder, J. B., E. Clancey, S. Des Roches, J. M. Eastman, L. Gentry, W. Godsoe, T. J. Hagey,  
 1045 D. Jochimsen, B. P. Oswald, J. Robertson, B. A. J. Sarver, J. J. Schenk, S. F. Spear, and

1046 L. J. Harmon. 2010. Ecological opportunity and the origin of adaptive radiations. Journal  
1047 of Evolutionary Biology **23**:1581–1596.

esa  
preprint



Table 1: Gray shading illustrates the primary focus of each research field. For each study (row), the first column summarizes how the putative niche constructor (*organisms in italics*) affects selection pressures on a recipient of niche construction (*organisms in bold*). The second column indicates whether there is support for each criterion, coming either from only comparative (number in parentheses) or both comparative and experimental evidence. The next four columns identify the modifiable ( $M$  subscript) characteristics of the environment, as illustrated in Figure 2 (i.e. Chemical: C; Physical: P; Consumer-Resource interactions: CR, and Non-Trophic Direct Interactions NTD). The columns 'NC' (niche constructor) and 'Recipient' indicate which organisms are subject to changes in the selection pressures caused by the organisms identified as doing the environmental modification. We identify of two research gaps in the main text, labelled here as GAP I and GAP II.

<i>Niche constructor</i> : <b>Recipient</b>	Criteria	$C_M$	$P_M$	$CR_M$	NTD $_M$	NC	Recipient	Citation
<b>Niche construction theory</b>								
Vascular <i>plants</i> decreased CO <sub>2</sub> by accelerating weathering and carbon burial	1,(2,3)	x	x			x	x	(Beerling, 2005)

*Continued on next page*

[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	$C_M$	$P_M$	$CR_M$	NTD <sub>M</sub>	NC	Recipient	Citation
<b><i>Cowbirds</i></b> socially manufacture compliant hosts to increase parasitic egg survival	1,2,(3)				x	x		(Hoover and Robinson, 2007)
<b><i>Earthworms</i></b> modify soil structure and water holding capacity to maintain osmotic balance	1,(2,3)	x	x			x		(Turner, 2002)
<b><i>Ants</i></b> culture fungi to avoid cellulose metabolism	1,(2,3)			x	x	x		(Mueller and Gerardo, 2002)
<b><i>Bioturbators</i></b> of sediments promote radiations of <b>Cambrian species</b>	1,(2,3)	x	x			x	x	(Erwin and Tweedt, 2011)

*Continued on next page*

[ *Niche constructor* ] : [ **Recipient** ]      Criteria       $C_M$        $P_M$        $CR_M$        $NTD_M$       NC      Recipient      Citation

Sequential speciation whereby *host*      1,(2,3)      x      x      x      (Forbes et al., 2009)

diversity promotes diversity of

**parasites**

Stabilization of social networks by      1,(2,3)      x      x      (Flack et al., 2006)

policing increases infant survival

rates of *mammals*

Heat production during      1,2,3      x      x      (Goddard, 2008)

fermentation by *Saccharomyces*

*cerevisiae* increases fitness

relative to other yeasts

*Continued on next page*

[ *Niche constructor* ] : [ **Recipient** ]      Criteria       $C_M$        $P_M$        $CR_M$        $NTD_M$       NC      Recipient      Citation

Evolution of seed dispersal in      1,2      x      x      x      (Donohue et al., 2005)

***Arabidopsis*** modifies environmental conditions and relative fitness of seed genotypes

*Caullerpa taxifolia* alters selection      1, 2      x      x      (Wright et al., 2012)

gradients of the co-occurring bivalve

**Extended phenotype**

***Gallwasps*** create oak galls to      1, (2,3)      x      x      (Bailey et al., 2009)

avoid parasitoids

*Continued on next page*

[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	$C_M$	$P_M$	$CR_M$	$NTD_M$	NC	Recipient	Citation
<b><i>Trematodes</i></b> cause limb malformation that increase amphibian susceptibility to predation	1,2,(3)		x	x	x	x		(Goodman and Johnson, 2011)
<b><i>Baculovirus</i></b> changes host behavior to increase transmission	1,(2,3)				x	x		(Hoover et al., 2011)
<b>Evolutionarily focussed studies</b>								
<b>Coevolution</b>								
Coevolving <i>pathogens</i> favor the evolution of sex in <i>nematodes</i>	1,2,3				x	x	x	(Morran et al., 2011)
<b>Diffuse co-evolution</b>								

Continued on next page

[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	C <sub>M</sub>	P <sub>M</sub>	CR <sub>M</sub>	NTD <sub>M</sub>	NC	Recipient	Citation
Pine cone <i>consumers</i> shape	1,2,3			x			x	(Benkman et al., 2010)
selection pressures on <i>crossbills</i>								
<i>Herbivore</i> community affects	1,2,(3)				x		x	(Stinchcombe and Rausher, 2002)
evolution of <i>plant</i> resistance to grazing								
<b>Character Displacement</b>		GAP I						
Competition for seeds drive	1,(2,3)			x		x	x	(Grant and Grant, 2006)
divergence in <i>finch</i> beak shape								
<b>Speciation</b>		GAP I						
Ecological speciation of	1,2,3			x	x	x	x	(Schluter, 2009)
<i>stickleback</i> via divergent natural selection								

*Continued on next page*

[ *Niche constructor* ] : [ **Recipient** ]      Criteria     $C_M$      $P_M$      $CR_M$      $NTD_M$     NC    Recipient      Citation

[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	$C_M$	$P_M$	$CR_M$	$NTD_M$	NC	Recipient	Citation
<b>Ecologically focussed studies</b>								
<b>Ecosystem engineering</b>								
<i>Riparian plants</i> structure fluvial landscapes	1		x					(Corenblit et al., 2009)
<i>Beavers</i> build dams and flood landscapes	1		x					(Jones et al., 1994)
<i>Spartina</i> modifies sediment accumulation and size structure	1		x					(Bouma et al., 2010)
<i>Sticklebacks</i> modify light environments of aquatic ecosystems	1		x					(Harmon et al., 2009)
<i>Earthworm</i> invasion changes ecosystem properties	1		x					(Hendrix, 2006)

*Continued on next page*



[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	$C_M$	$P_M$	$CR_M$	NTD <sub>M</sub>	NC	Recipient	Citation
<b>Ecological stoichiometry</b>								
Variation in stoichiometry of <i>predators</i> affects nutrient cycling	1	x				GAP II		(McIntyre et al., 2008)
Selection on growth rate of <b>consumers</b> increases their P-demand	1							(Sterner and Elser, 2002)
Selection on stoichiometry of <b>primary producers</b>	1							(Mizuno and Kawata, 2009)
<b>Ecosystem effects of organisms</b>								
Phenotypic variation of <i>consumers</i> affect ecosystem properties and functions	1	x	x	x	x			(Schmitz, 2010)

*Continued on next page*

[ <i>Niche constructor</i> ] : [ <b>Recipient</b> ]	Criteria	$C_M$	$P_M$	$CR_M$	$NTD_M$	NC	Recipient	Citation
Variation in <i>consumer</i> biomass affects ecosystem properties and functions	1	x	x	x	x			(Dickman et al., 2008)
<i>Predators</i> drive trophic cascades	1			x	x			(Carpenter and Kitchell, 1993)
<i>Organism</i> -mediated shifts between alternative stable states	1,(2,3)	x	x	x	x			(Scheffer et al., 2001)
<b>Species Turnover</b>								
<i>Predators</i> affect community assembly of prey	1			x	x			(Chase et al., 2009)

1048 **Figure Legends**

1049 **Figure 1:** A Venn diagram showing which modules of biotic (square) and abiotic (circles)  
 1050 entities, which are connected by evolutionary (dashed lines) and non-evolutionary  
 1051 effects (solid lines), are associated with different major concepts in ecology and  
 1052 evolution (bounded by labelled shaded boxes). Non-evolutionary effects include  
 1053 organism-mediated effects on both biotic and abiotic conditions (e.g. ecological  
 1054 effects shown in Figure 2A), and evolutionary effects include evolutionary responses  
 1055 to selection. The asterisk denotes effects on the physical state of the abiotic  
 1056 environment, to distinguish ecosystem engineering (yellow box) from effects on other  
 1057 abiotic conditions (e.g. the chemical environment). The minimum condition for  
 1058 evolution by niche construction to occur is to have a pathway that starts and ends  
 1059 with an organism (i.e. a niche constructor and a recipient of niche construction), and  
 1060 has at least two connections with an evolutionary effect beyond the first connection.  
 1061 Starting from the left of each pathway the red dashed arrow defines where evolution  
 1062 by niche construction has occurred.

1063 **Figure 2:** (A) A partitioning of how organisms can modify their biotic and abiotic  
 1064 environments. (B) An elaboration of how organism mediated environmental  
 1065 modifications can affect the fitness of another organism (e.g. potentially a recipient of  
 1066 niche construction), through a variety of pathways (abbreviated following Figure 2A:

1067 Physical [P]; Chemical [C], Consumer-Resource [CR], Non-trophic direct [NTD]).

1068 Niche construction can occur when organism-mediated environmental modifications

1069 alter the evolutionary response of organisms relative to other environmental drivers of

1070 selection (e.g. unmodifiable environment).

1071 **Figure 3:** Four examples of comparative tests of the niche construction (Criteria 1 and 2).

1072 (A) Tests for relationships between unmodified and modifiable environmental

1073 properties in the presence and absence of a candidate niche constructor (Criteria 1).

1074 Differences in such relationships (e.g. line slopes) could be associated with

1075 organism-mediated environmental modifications. (B) Tests for the relationship

1076 between the abundance of an agent and variation in a modified component of the

1077 environment that is unexplained by other environmental conditions (Criteria 1). The

1078 indicated relationship could occur if the organism's effects on the modified

1079 component of the environment are linearly related with the abundance of the

1080 organism. (C) Tests of whether the relative fitness of two organisms with different

1081 phenotypes differ between two environments that are either unmodified or modified

1082 by a putative niche constructor (Criteria 2). (D) Tests for a relationship (for example

1083 among sites) between selection gradients and the degree of environmental

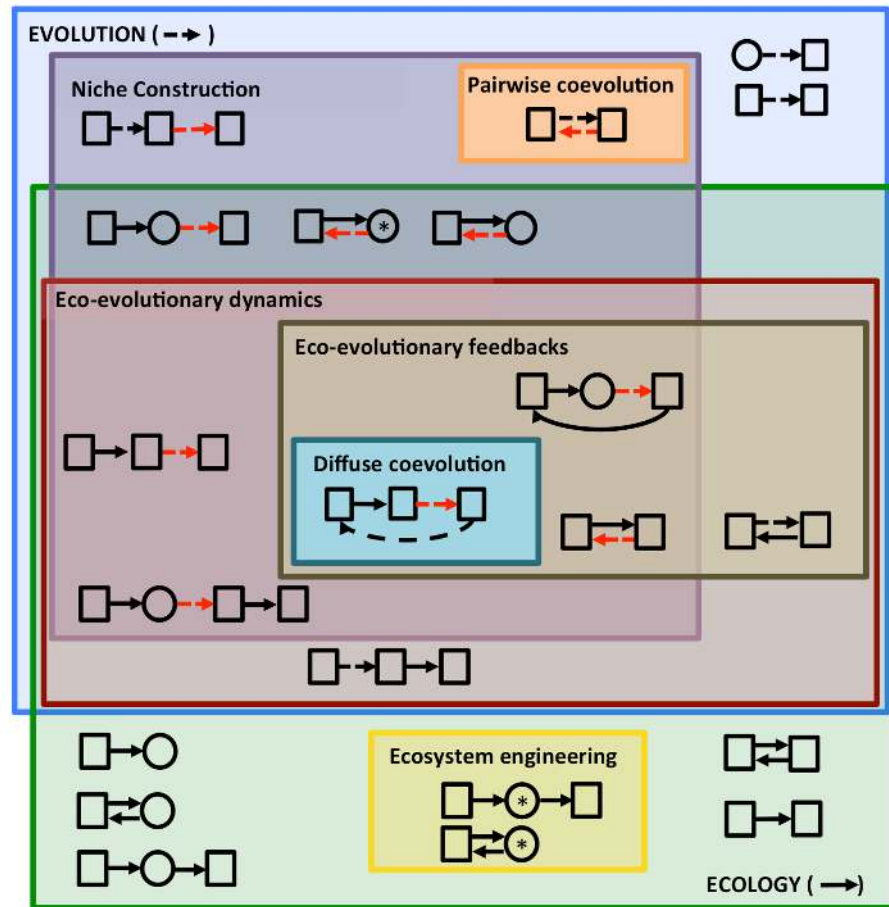
1084 modification of a niche constructor (Criteria 2). See Odling-Smee et al. (2003) for a

1085 description of counteractive and inceptive niche construction.

1086 **Figure 4:** (A) Experimental design of a common gardening experiment used to measure  
 1087 how potential niche constructors can have contrasting effects on ecosystems (e.g.  
 1088 organism 1 modifies the environment from square to star, while organism 2 modifies  
 1089 the environment from square to hexagon), relative to unmodified ecosystems (i.e.  
 1090 squares). The difference between the modified and the control ecosystems is the effect  
 1091 size for a given ecosystem metric, as shown in the panel on the right. Dotted lines  
 1092 delineate where the effect size is not significantly different from zero (dashed line).  
 1093 (B) Same approach as in (A) except the ecosystem metrics are measured multiple  
 1094 times (e.g.  $t_1, t_2$ ) after the organism doing the ecosystem modification is removed.  
 1095 (C) Four different ways to quantify evolutionary changes in common gardening  
 1096 experiments. In (i) and (ii) genetic properties of populations or individuals can be  
 1097 measured along a gradient of ecosystem modification, and, following the removal of a  
 1098 potential niche constructor, one can measure how selection gradients (iii: times refer  
 1099 to Panel B) change through time (iv).

1100 **Figure 5:** A schematic emphasizing how macrophyte communities can have  
 1101 non-evolutionary effects (shown as solid arrows) on the abiotic environment (circles)  
 1102 of shallow lakes in general, and on the physical (indicated by an asterisk) and  
 1103 chemical conditions in particular. Such effects could lead to both evolutionary effects  
 1104 (shown as dashed arrows) and ecological effects (arrows not shown for clarity but

1105 follow the same paths as evolutionary effects) on organisms (circles) that play a role  
1106 in the transition between turbid and clear states (shown as a double-headed arrow).  
1107 Following figure 1, macrophytes could drive evolution by niche construction by a  
1108 variety of mechanisms in shallow lakes. Such evidence could come from pathways of  
1109 effect that start with macrophytes, pass through an abiotic environmental condition,  
1110 and end with an evolutionary effect on a recipient organism. Ecological and  
1111 evolutionary effects between organisms (e.g. trophic interactions) are also left out for  
1112 clarity, but are also very important for understanding transitions between stable  
1113 states in shallow lakes.

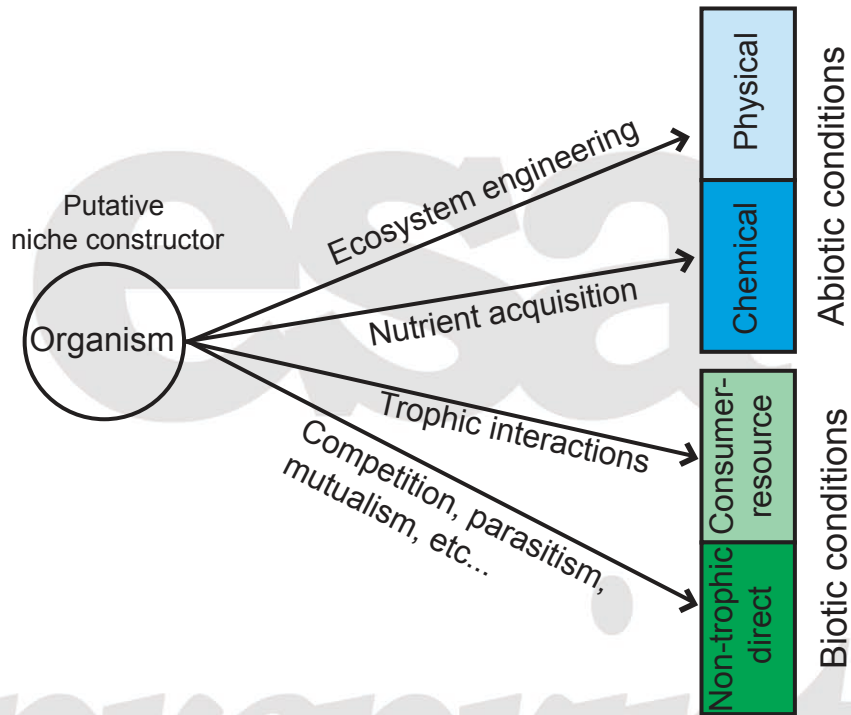


1.pdf

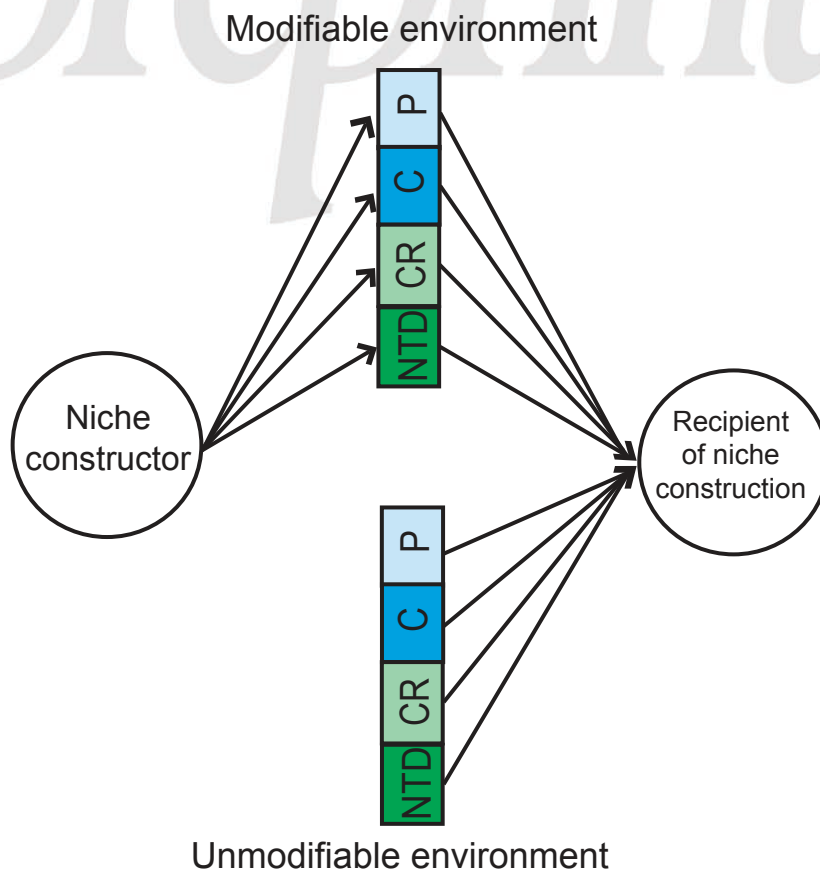
Figure 1:

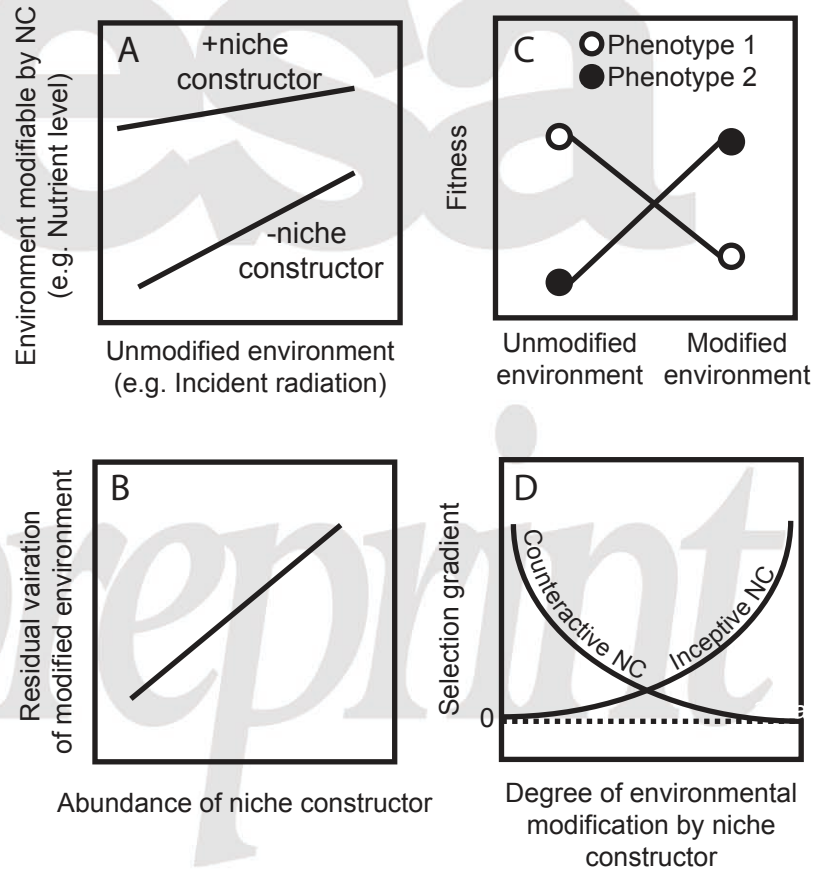


(A)



(B)

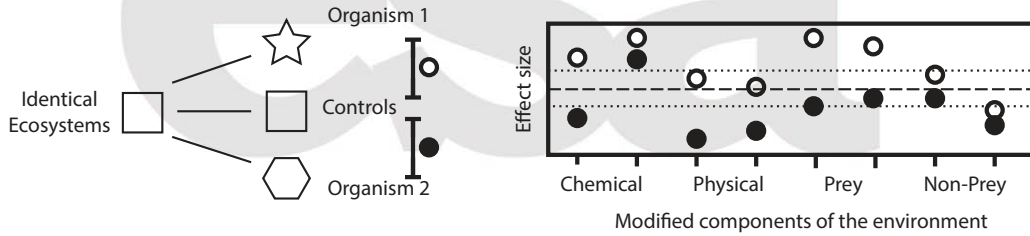




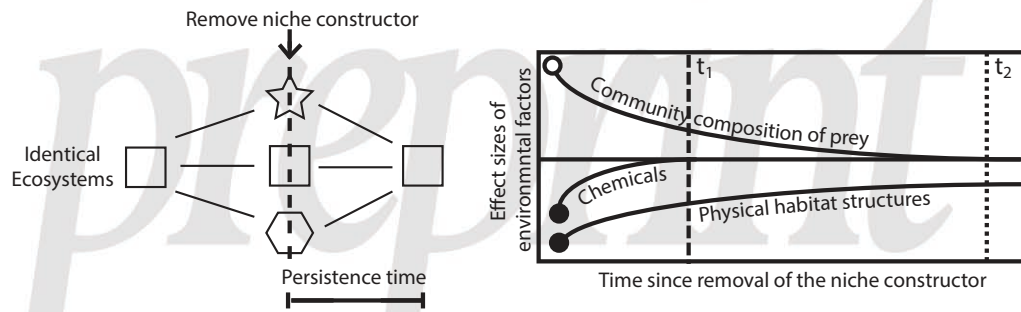
3.pdf

Figure 3:

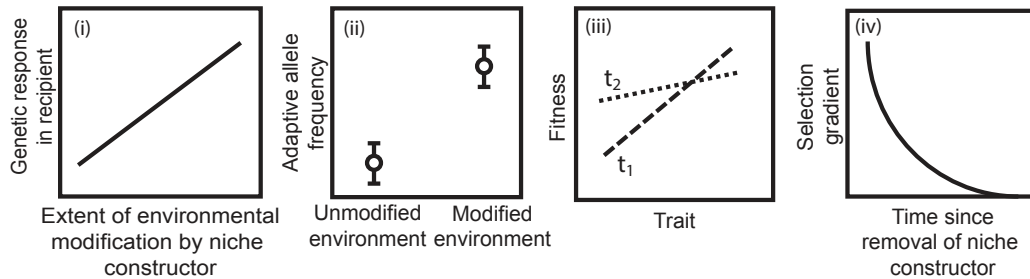
A: Measuring the effect-size distribution of organism-mediated environmental modifications



B: Exploring the persistence time of environmental modifications



C: Evolutionary responses of recipients (y-axis) caused by niche constructors (x-axis)



4.pdf

Figure 4:

