REVIEW



Unexpected guests in the tumor microenvironment: microbiome in cancer

Abigail Wong-Rolle¹, Haohan Karen Wei², Chen Zhao^{1⊠}, Chengcheng Jin^{2⊠}

🖂 Correspondence: chen.zhao@nih.gov (C. Zhao), Chengcheng.Jin@Pennmedicine.upenn.edu (C. Jin)

Received August 23, 2020 Accepted November 1, 2020

ABSTRACT

Although intestinal microbiome have been established as an important biomarker and regulator of cancer development and therapeutic response, less is known about the role of microbiome at other body sites in cancer. Emerging evidence has revealed that the local microbiota make up an important part of the tumor microenvironment across many types of cancer, especially in cancers arising from mucosal sites, including the lung, skin and gastrointestinal tract. The populations of bacteria that reside specifically within tumors have been found to be tumor-type specific, and mechanistic studies have demonstrated that tumor-associated microbiota may directly regulate cancer initiation, progression and responses to chemo- or immuno-therapies. This review aims to provide a comprehensive review of the important literature on the microbiota in the cancerous tissue, and their function and mechanism of action in cancer development and treatment.

KEYWORDS microbiome, tumor, lung cancer, immune system, tumor-associated microbiota, cancer immunotherapy

INTRODUCTION

The commensal microbiome contains at least 100-fold more unique genes than the human host genome (Ley et al., 2006; Human Microbiome Project, 2012; Belkaid and Hand, 2014; Dzutsev et al., 2017). The primary habitat of the human commensal microbiota is the gut, but thriving microbial populations exist throughout much of the body including the skin, oral, respiratory, and genital tracts (Human Microbiome Project, 2012). These microbes influence many of our important physiological functions including our metabolism and immune system. This metaorganism, made up of human host and microbial symbionts, senses environmental cues and adapts accordingly (Belkaid and Naik, 2013; Dzutsev et al., 2017). The complex interaction between the host and microbes extends to cancer; microorganisms are implicated in 20% of human malignancies (de Martel et al., 2012). Commensal bacteria directly affect tumorigenesis, progression and responses to treatment. Disruption of commensal gut microbiota in mice can affect the response of tumors to immunotherapy and chemotherapy (lida et al., 2013; Viaud et al., 2013; Sivan et al., 2015; Vétizou et al., 2015). This dependence of treatment responsiveness on the gut microbiome has also been observed in cancer patients. Antibiotic treatment is associated with reduced response to immune checkpoint inhibitors whereas the presence or increased abundance of some gut bacteria strains correlates with better outcomes (Gopalakrishnan et al., 2018; Matson et al., 2018; Routy et al., 2018). The systemic effect of the gut microbiome in regulating cancers in the gastrointestinal tract, as well as in distal sites, has been well reviewed previously (Garrett, 2015; Dzutsev et al., 2017; Shalapour and Karin, 2020). The focus of this review is the role of microbiota in the tumor-bearing tissue, especially intratumor microbes.

The distribution of commensal microbes across anatomical sites reveals distinct microbial communities (Costello et al., 2009; Belkaid and Hand, 2014). The commensal microbiota of tumor-bearing tissue makes up an important part of the tumor microenvironment, affecting tumorigenesis and tumor progression on a more local scale (Garrett, 2015).

¹ Thoracic and Gastrointestinal Malignancies Branch, National Cancer Institute, National Institutes of Health, 4 Memorial Drive, Bethesda, MD 20892, USA

² Department of Cancer Biology, Perelman School of Medicine, University of Pennsylvania, 421 Curie Blvd., Philadelphia, PA 19104, USA

Abigail Wong-Rolle and Haohan Karen Wei have contributed equally.

Additionally, the populations of bacteria that reside specifically within tumors have been found to be tumor-type specific, suggesting an association with tumor development (Nejman et al., 2020). In a very recent study, Nejman et al. profiled over 1500 tumors across seven cancer types and discovered that intratumor and intracellular microbial compositions are distinct between tumor types; further analysis revealed correlations between microbial metabolic pathways and clinical features (Nejman et al., 2020). The distinction between the role of commensal microbes in tumor-bearing tissue and the role of intratumor or tumor-associated microbes has not always been made and cannot always be readily distinguished due to technical challenges. This review aims to provide an overview of the microbiota of the cancerous tissue, and their role in tumorigenesis and tumor progression across different cancer types (Table 1).

MECHANISMS

The link between intratumor microbes and cancer development has been correlatively established and three primary mechanisms have been demonstrated as potential modes of action (Garrett, 2015; Dzutsev et al., 2017; Ramirez-Labrada et al., 2020): (1) direct facilitation of tumorigenesis via increasing mutagenesis, (2) regulation of oncogenes or oncogenic pathways, and (3) reduction or enhancement of tumor progression via modulation of host immune system (Fig. 1).

Many microbes have evolved to produce compounds that can cause damage to DNA, cell cycle arrest, and genetic instability. The presence of bacteria producing such compounds in what will become the tumor microenvironment could directly increase mutagenesis of the occupied tissue. Colibactin, produced via the *pks* locus in some *Escherichia coli* and other *Enterobacteriaceae*, exemplifies the potential of microbiota to directly promote oncogenesis (Nougayrede et al., 2006; Putze et al., 2009): it causes double stranded DNA damage and thereby facilitates tumorigenesis in colorectal cancer (Guerra et al., 2011; Garrett, 2015).

Aside from direct damage to DNA, commensal microbial products can engage with host oncogenic pathways. Upregulation or activation of pathways leading to

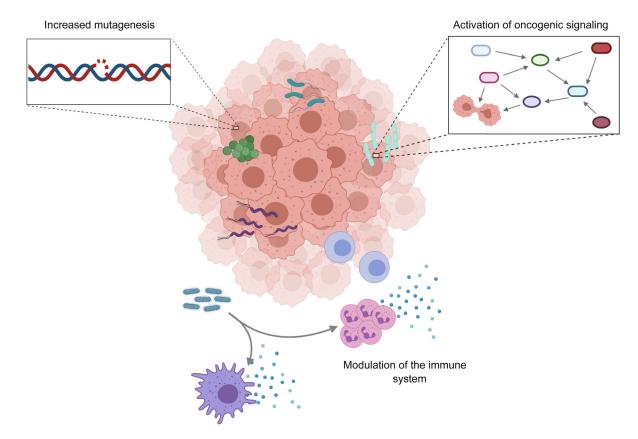


Figure 1. Interactions between microbes and tumor cells. Microbial communities in the tumor-bearing tissue and intratumor microbes are associated with cancer, but their exact role is not fully understood. Three main mechanisms have been proposed to explain how local microbes affect carcinogenesis and cancer progression: direct facilitation of DNA damage and increased mutagenesis (top left), activation of oncogenic signaling pathways (top right), and reduction or enhancement of tumorigenesis and tumor progression through interaction with the host immune system (bottom).

Cancer type	Name of the identified tumor-associated microbiome	Proposed mechanism
Lung cancer	Veillonella (Yan et al., 2015; Lee et al., 2016; Tsay et al., 2018)	N/A
	<i>Streptococcus</i> (Cameron et al., 2017; Tsay et al., 2018)	N/A
	Acinetobacter (Cameron et al., 2017; Gomes et al., 2019)	N/A
	Capnocytophaga (Liu et al., 2018)	N/A
	Thermus and Legionella (Yu et al., 2016)	N/A
	Megasphaera (Lee et al., 2016)	N/A
	Granulicatella adiacens, Enterococcus, and Escherichia coli (Cameron et al., 2017)	N/A
	Brevundimonas, Propionibacterium, and Enterobacter (Gomes et al., 2019)	N/A
	Prevotella and Rothia (Tsay et al., 2018)	N/A
	Acidovorax (Greathouse et al., 2018)	N/A
Colorectal cancer	<i>Fusobacterium nucleatum</i> (Castellarin et al., 2012; Kostic et al., 2012, 2013; Rubinstein et al., 2013; Bullman et al., 2017; Yu et al., 2017; Garrett, 2019)	Binding of <i>F. nucleatum</i> adhesin molecule to the cell surface motifs on cancer cells or immune cells, which leads to the downstream oncogenic or immunosuppressive signaling (Garrett, 2019).
	Enterotoxigenic <i>Bacteroides fragilis</i> (ETBF) (Dejea et al., 2018)	Tumor-coating ETBF recruits other bacteria as well as immune cells to the tumor site and boosts IL-17- mediated inflammation (Dejea et al., 2018).
	<i>Escherichia coli</i> expressing the genomic island polyketide synthase (pks+ <i>E. coli</i>) (Arthur et al., 2012; Dejea et al., 2018)	pks+ <i>E. coli</i> -derived colibactin alkylates DNA and produces DNA adducts, resulting in DNA damage in colonic epithelial cells (Wilson et al., 2019).
	Bifidobacterium (Shi et al., 2020)	N/A
Pancreatic ductal adenocarcinoma	<i>Proteobacteria</i> (Geller et al., 2017; Pushalkar et al., 2018)	Proteobacteria lead to T cell anergy in a Toll-like receptor-dependent manner, accelerating tumor progression (Pushalkar et al., 2018).
	Malassezia globose (Aykut et al., 2019)	Contributes to tumorigenesis, tumor growth, and gemcitabine resistance via mannose-binding lectin-C3 axis (Aykut et al., 2019).
	Pseudoxanthomonas, Saccharopolyspora, and Streptomyces spp. (Riquelme et al., 2019)	N/A
Esophageal cancer	Fusobacterium nucleatum (Yamamura et al., 2016)	Contributes to tumor infiltration of Treg lymphocytes in a chemokine (especially CCL20)-dependent fashion, promoting aggressive tumor behaviors (Yamamura et al., 2016).

Table 1. An overview of the microbiota in the cancerous tissue, and their role in tumorigenesis and tumor progression across different cancer types.

carcinogenesis has been reported, notably including the Wnt/ β -catenin signaling pathway (Garrett, 2015). Altered β -catenin signaling can promote transcription of oncogenes like *c-Myc* and *CyclinD-1* and advance carcinogenesis and tumor progression (Shang et al., 2017). Activation or modulation of β -catenin by local commensal microbiota has been reported in gastric and colorectal cancers (Sears, 2009; Kostic et al., 2012, 2013; Rubinstein et al., 2013; Abreu and Peek, 2014).

Immune modulation by commensal microbiota is well documented, as are its effects on carcinogenesis and cancer progression (Belkaid and Naik, 2013; Belkaid and Hand, 2014; Garrett, 2015; Dzutsev et al., 2017). The interaction between commensals and host immune system is well-balanced and homeostatic in a state of health. The host immune system is able to tolerate commensal microbial communities and respond appropriately to potentially harmful pathogens. However, perturbation of this balance and dysregulation of the microbiome can lead to the creation of microenvironments that promote tumor initiation and progression (Belkaid and Hand, 2014; Garrett, 2015). Many studies have shown that dysbiosis in local bacterial communities can cause a chronic, pro-inflammatory immune response, and thereby promote cancer growth. For example, this may occur through microbe activation of NF- $\kappa\beta$, a key regulator of cancer-associated inflammation (DiDonato et al., 2012; Elinav et al., 2013; Kostic et al., 2013; Garrett, 2015). Aside from inflammation, local microbes have been shown to modulate local immune surveillance by suppressing the antitumoral immune response. This type of immunosuppression has been observed in colon cancer patients, as well as in mouse models of lung cancer (Garrett, 2015; Gur et al., 2015; Le Noci et al., 2018), suggesting that the local commensal microbiota may be a potential target for cancer treatment, though further investigation is required.

LUNG CANCER

As the mucosal site with the largest surface area in the body and a major interface with the external environment, the lung presents a unique opportunity for exposure to microbes and environmental challenges. Though traditionally thought to be sterile, the lung harbors a diverse array of microbes (Charlson et al., 2011; Erb-Downward et al., 2011). The myth of lung sterility stems from an inability to culture bacteria from lower respiratory tract samples (Baughman et al., 1987; Thorpe et al., 1987). However, with the advent of culture independent 16S rRNA sequencing technologies, diverse microbial populations have been discovered (Dickson et al., 2015; Yu et al., 2016). The lung microbiota is distinct from that of the gut and skin. In the healthy lung, Prevotella, Streptococcus, Veillonella, Neisseria, Haemophilus, and Fusobacterium are among the most abundant bacteria genera (Yu et al., 2016; Sommariva et al., 2020). Microbe prevalence is dictated by the immigration of new bacteria, mechanical and immune based elimination, and replicative success governed by local environmental conditions (Dickson et al., 2014; Sommariva et al., 2020). The lung microbiota has been shown to be in a state of dysregulation in pulmonary diseases like chronic obstructive pulmonary disease (COPD) and cystic fibrosis (Hilty et al., 2010; Erb-Downward et al., 2011; Dickson et al., 2013; O'Dwyer et al., 2016). Increasing evidence from both human and mouse studies have linked bacterial dysbiosis with lung cancer (Lee et al., 2016; Yu et al., 2016; Cameron et al., 2017; Liu et al., 2018; Ramirez-Labrada et al., 2020).

Among cancers, lung cancer is the leading cause of cancer related mortality and second most common cancer type, contributing to one quarter of all US cancer deaths (Siegel et al., 2019). An estimated 90% of all lung cancer cases are attributed to smoking, with tobacco smoke, air pollution, and other carcinogens all established risk factors, though exact mechanisms are not well understood (Gustafson et al., 2010). Although there are controversies (Greathouse et al., 2020), the intratumor and intracellular

bacterial taxa of smokers show enrichment in degradation pathways for chemicals in cigarette smoke, pointing towards an association between intratumor microbiome and cancer etiology (Nejman et al., 2020). More generally, the lung commensal microbiota has been linked to cancer. For example, Yu et al. found a correlation between increased Thermus genus abundance and advanced stage cancer, as well as between Legionella and metastases. Additionally, a decrease in the alpha diversity of bacterial community in tumor tissues was reported in comparison to non-malignant lung tissues in lung cancer patients (Yu et al., 2016). Though methods and samples vary, other studies have recapitulated an enrichment of specific bacterial taxa and reduced diversity in association with lung cancer (Yan et al., 2015; Lee et al., 2016; Cameron et al., 2017; Greathouse et al., 2018; Tsay et al., 2018; Gomes et al., 2019). Differences between non-malignant and tumor tissue point towards the question: what is the role of tumor-associated microbiota, especially the intratumor microbiota, in tumorigenesis and progression of lung cancer?

The lung microbiota can directly affect the growth of lung cancer cells. Of the three main mechanisms we have presented for the role of local commensal microbes in carcinogenesis and cancer progression, regulation of local immune environment, and oncogenic pathways have been implicated in lung cancer. Dysregulation of lung microbial communities has been suggested to facilitate changes in oncogenic pathways, potentially through specific microbial components (Ramirez-Labrada et al., 2020). Findings by Greathouse et al. suggested an association between TP53 and lung microbiota dysbiosis. The genus Acidovorax was found to be enriched in squamous cell carcinoma lung biopsy samples; this same taxa was found to be further enriched in lung biopsies of TP53 mutated squamous cell carcinoma patients, though further mechanistic analysis remains to be pursued (Greathouse et al., 2018). Using airway brushings from diagnostic bronchoscopies, Tsay et al. found that patients with lung cancer had increased oral taxa, specifically Streptococcus and Veillonella, compared to controls. Increased prevalence of oral taxa was associated with PI3K and ERK upregulation. In vitro experiments exposing airway epithelial cells to Veillonella, Prevotella, and Streptococcus also resulted in ERK and PI3K pathway upregulation (Tsay et al., 2018). The PI3K pathway has been implicated as an early event in lung carcinogenesis (Gustafson et al., 2010), and therefore upregulation of this pathway by commensal microbiota dvsbiosis facilitates carcinogenesis.

Lung microbiota has been shown to alter the immune microenvironment to promote tumor progression. Chronic inflammation has been closely linked to cancer; cytokines, chemokines, and other pro-inflammatory factors can facilitate tumor growth and spread (Garrett, 2015; Sommariva et al., 2020). The lung, due to its extensive exposure to the external environment, is a critical site of immune–microbiota interaction and exists in homeostasis maintained by lung resident immune cells (Pilette et al., 2001; Huffnagle et al.,

2017; Lloyd and Marsland, 2017; Sommariva et al., 2020). Jin et al. demonstrated the importance of microbiota-immune crosstalk in promoting inflammation and the development of lung cancer in an autochthonous mouse model. Specifically, they found that certain bacterial families such as Herbaspirillum and Sphingomonadaceae were enriched in tumor-bearing lung tissues compared to healthy lungs, while other taxa including Aggregatibacter and Lactobacillus were enriched in healthy lungs. The increased local bacterial burden and altered composition of lung microbiota stimulated Myd88-dependent IL-1 β and IL-23 production from myeloid cells. These cytokines induced the activation and proliferation of Vy6+V δ 1+y δ T cells, which produced IL-17, promoting inflammation and neutrophil infiltration. Additionally, these $\gamma\delta$ T cells produced IL-22 and other effector molecules promoting tumor cell proliferation. Germfree (GF) mice or antibiotics-treated mice had significantly reduced lung tumor growth, demonstrating commensal bacteria significantly promoted lung cancer development (Jin et al., 2019). Similarly, Le Noci et al. utilized aerosolized antibiotics to demonstrate that decreased bacterial biomass was linked to an enhancement of antitumoral immune response via T cell and NK cell activation and reduction of immunosuppressive regulatory T cells. Additionally, the probiotic Lactobacillus rhamnosus was found to overcome immunosuppression and inhibit lung tumor implantation, and tumor metastases were reduced under both antibiotic and probiotic conditions (Le Noci et al., 2018). Altogether, these findings support the notion that local microbiota plays a key role in lung cancer development by modulating the local immune response and targeting tumor-associated microbiota presents a potential new avenue for lung cancer prevention and treatment.

GI CANCER

Colorectal cancer

The gastrointestinal tract harbors the vast majority of commensal microbiomes in the human body (Dzutsev et al., 2017), and bi-directional crosstalk between the host and the microbiome communities has emerged as a central aspect of both tumor progression and the therapeutic response to various types of GI cancers (Garrett, 2015; Dzutsev et al., 2017; Shalapour and Karin, 2020).

Numerous studies have established the role of commensal bacteria in colorectal cancer. Mechanistically, gut microbiota has been shown to promote mutagenesis by causing double-stranded DNA damage in host, or through activating oncogenic signaling like the Wnt/ β -catenin pathway, or promoting inflammation by upregulating NF-kB signaling via increased engagement of pattern recognition receptors (Arthur et al., 2012; Kostic et al., 2013; Rubinstein et al., 2013). For example, *Fusobacterium* was first identified to be associated with colorectal carcinoma using unbiased genomic analyses (Castellarin et al., 2012; Kostic et al., 2012). Subsequent study further demonstrates the persistent colonization of Fusobacterium nucleatum and its associated anaerobes in colorectal tumors (Bullman et al., 2017). Bullman and colleagues, using both in vitro and in vivo humanderived colorectal cancer (CRC) xenograft models, showed that F. nucleatum promotes tumor cell proliferation or tumor growth (Bullman et al., 2017). A possible mechanism explaining this finding is the binding of F. nucleatum adhesin molecule to the cell surface motifs on cancer cells or immune cells, leading to the downstream oncogenic or immunosuppressive signaling (Garrett, 2019). Other commensal bacteria associated with colorectal cancer include enterotoxigenic Bacteroides fragilis (ETBF) and Escherichia coli expressing the genomic island polyketide synthase (pks+ E. coli) (Arthur et al., 2012; Dejea et al., 2018). It is discovered that the pks+ E. coli-derived colibactin alkylates DNA and produces DNA adducts, resulting in DNA damage in colonic epithelial cells (Wilson et al., 2019). Dejea et al. found that cocolonization by ETBF and pks+ E. coli, as shown by FISH and microbiology culture analysis, can accelerate the onset of colon cancer and increase mortality in mouse models of CRC (Dejea et al., 2018). In addition, tumor-coating ETBF has been shown to recruit other bacteria, as well as immune cells, to the tumor site and boosts IL-17-mediated inflammation (Dejea et al., 2018). Interestingly, a recent study discovered that mutant p53 is switched from tumor-suppressive to oncogenic by the gut microbiota in the distal gut i. e., ileum and colon (Kadosh et al., 2020).

Aside from tumorigenesis, intratumor microbiota plays an important role in modulating the response to cancer therapies. For instance, *F. nucleatum* was found to be abundant in the CRC tumorous tissues in patients with recurrence post chemotherapy (Yu et al., 2017). Yu et al. showed that *F. nucleatum* promotes chemoresistance by activation autophagy pathway through downregulation of microRNAs (miR-18a and miR4802) (Yu et al., 2017). In another study that investigated the role of intratumor microbiota in CD47-based cancer immunotherapy, Shi et al. found that colonic *Bifidobacterium* accumulates in tumor sites and facilitates local anti-CD47 treatment via the STING pathway (Shi et al., 2020).

OTHER GI CANCERS

Pancreatic cancer is one of the deadliest cancers and has a very poor prognosis, with a five-year survival rate of 8% (Siegel et al., 2018). The pancreas was historically considered to be sterile, yet emerging evidence demonstrates the presence of intratumor microbes and their impact on pancreatic cancer progression and therapeutic efficacy (Balachandran et al., 2017; Geller et al., 2017; Pushalkar et al., 2018; Aykut et al., 2019; Riquelme et al., 2019; Nejman et al., 2020). Geller and colleagues (Geller et al., 2017) incidentally found that 76% of the human pancreatic ductal adenocarcinoma (PDAC) samples were positive for bacteria, with *Gammaproteobacteria* being the dominant taxa.

Bacteria cultures from 14 out of 15 (93%) fresh human PDAC tumors were able to confer resistance to chemotherapeutic drug gemcitabine on tested human colon cancer cell lines. Pushalkar et al. (Aykut et al., 2019) discovered an increased abundance of select bacteria (45% of which were Proteobacteria) in pancreatic tumors from genetically engineered mice models. It was demonstrated that these intratumor bacteria led to T cell anergy in a Toll-like receptor (TLR)-dependent manner, and transfer of selective or bulk fecal bacteria from PDAC-bearing mice, but not control mice, accelerated tumor progression. In addition to bacteria, intratumor fungi like Malassezia globosa in pancreatic cancer were also found to contribute to tumorigenesis, tumor growth, and gemcitabine resistance in a mannose-bindinglectin-dependent manner (Aykut et al., 2019). On the other hand, some intratumor microbiomes were reported to boost the anti-tumor immunity. For example, Riquelme and colleagues (Riquelme et al., 2019) found that the abundance of intratumor bacteria Pseudoxanthomonas, Saccharopolyspora, and Streptomyces spp. was highly predictive of long-term survival in pancreatic cancer patients, and an overall more diverse composition of the tumor microbiome was observed in long-term survivors; the more diverse microbiome in the tumor contributes to the anti-tumor immune response by favoring recruitment and activation of CD8⁺ T cells. Furthermore, Balachandran et al. (2017) discovered that the enhancement of immune infiltrates was associated with the presence of intratumoral neoantigen MUC16 in long-term survivors. The authors speculated that intratumor microbiomes may enhance immune identification of relevant neoantigens (Balachandran et al., 2017).

In esophageal cancer, the presence of *F. nucleatum* in esophageal cancer tissues was correlated with a poor prognosis (Yamamura et al., 2016). Mechanistically, Yamamura et al. (2016) suggested that *F. nucleatum* contributed to tumor infiltration by Treg lymphocytes in a chemokine (especially CCL20)-dependent fashion, thereby promoting aggressive tumor behaviors.

As commensal microbiota resides in the GI tract at the steady state, an important question is posed as the origin of tumor-associated bacteria. Consistent with the hypothesis that constant interactions exist between the pancreas, liver, and intestine, the development of pancreatic and liver cancer has been associated with the dysregulation and mislocation of gut microbiota (Bullman et al., 2017; Geller et al., 2017; Pushalkar et al., 2018; Vitiello et al., 2019). Geller et al. (2017) suggested that PDAC-associated bacteria could be sourced from the gastrointestinal tract in a retrograde manner. Pushalkar et al. (2018) showed evidence of bacteria migration from the gut to the pancreas, as well as a timedependent association between gut dysbiosis and Kras activation in PDAC. Gut dysbiosis can also directly promote oncogenic signaling in the pancreas (Vitiello et al., 2019). Commensal microbiome obtained from long-term PDAC survivors was highly capable of enhancing immune infiltration and anti-tumor immunity (Riquelme et al., 2019). As in

liver cancer, Bullman et al. (2017) found that the relative abundance of *Fusobacterium* was significantly increased in colorectal cancer-derived liver metastases in comparison to primary liver hepatocellular carcinoma. The crosstalk between these organs points to a need to elucidate the origin of the observed intratumor microbiomes in pancreatic and liver cancer, and the relative contribution of microbiome in the distal intestine or in the local cancerous tissue to cancerassociated immune response.

SKIN CANCER

The human skin is home to thriving populations of diverse microbes, with an estimated one million bacteria and hundreds of different species per square centimeter (Chen and Tsao, 2013). As in other body sites, changes in the composition of the skin microbiome are associated with cancer. In a study of the microbiota of normal versus melanotic pig skin, the melanoma samples were found to be enriched in Fusobacterium and Trueperella genera (Mrázek et al., 2019). In a cell culture study, a strain of skin commensal microbes, Staphylococcus epidermidis, had a protective effect against skin cancer. These strains of Staphylococcus epidermidis produce 6-N-hydroxyaminopurine, an inhibitor of DNA polymerase activity, which stops tumor line proliferation in culture (Nakatsuji et al., 2018). Investigation of the relationship between host immune system, local microbial communities, and cancer has also begun in the context of skin cancer. Hoste et al. utilized a wound-induced skin cancer mouse model to interrogate the mechanisms through which the skin microbiota promotes inflammation and tumorigenesis. In the presence of skin microbes, elimination of several innate immune sensors, including TLR-5, protects from tumorigenesis, while inflammation correlates with tumor incidence. Treatment with antibiotics inhibits tumor formation in a TLR-5 dependent manner (Hoste et al., 2015). A better understanding about the role of the skin microbiome in skin cancer is necessary, as it could potentially provide further insight into the different roles tissue-specific microbiota in cancer initiation, progression, and potentially treatment.

OTHER CANCERS

In addition to the major mucosal organs described above, the vaginal microbiome has also been associated with cervical cancer (Shannon et al., 2017; Norenhag et al., 2020). Aside from tumors that arise from the mucosal sites that host tissue-resident commensals, it is interesting to note that intratumor bacteria has also been identified in other types of cancer, including breast cancer, ovarian cancer, bone cancer, and glioblastoma multiforme (GBM) (Hieken et al., 2016; Urbaniak et al., 2016; Banerjee et al., 2018; Nejman et al., 2020). However, there is very limited experimental evidence for association of distinct microbes with these types of cancer. A recent landmark study by Nejman et al. (2020) showed the presence of intratumor bacteria in ovarian and bone cancers, as well as in GBM. Distinct microbial compositions and metabolic functions encoded by intratumor bacteria were identified across different types of cancer (Nejman et al., 2020). Moreover, different bacterial signatures were associated with subtypes of breast cancer, in agreement with findings from previous articles (Urbaniak et al., 2016; Banerjee et al., 2018). In addition, it was discovered that breast tumor-associated microbiomes had the highest level of diversity and abundance among all tumor types tested (Nejman et al., 2020). Importantly, this study confirmed the presence of metabolically active bacteria using *ex vivo* bacterial isolation from fresh breast tissue and fluorescently labeled *D*-alanine.

CONCLUSION AND DISCUSSION

The microbiota is gaining increasing attention as a key player in the tumor microenvironment that modulates tumor progression and influences cancer prognosis. The microbial communities of different body sites play a role in tumorigenesis and tumor progression of their respective cancers. In many cases, direct changes in mutagenesis, regulation of oncogenic pathways, and modulation of the immune system can provide potential explanations for the link between local microbes and cancer (Garrett, 2015; Dzutsev et al., 2017; Ramirez-Labrada et al., 2020). Notably, the commensal microbiota exerts an effect upon the immune microenvironment, frequently promoting inflammation or dampening antitumor immunity. Crosstalk between the host immune system and local microbes affects cancer growth and spread (Belkaid and Hand, 2014; Garrett, 2015; Ramirez-Labrada et al., 2020).

Despite our growing knowledge of microbiota in cancer and cancer treatments, the role of tumor-associated bacteria requires further investigation. Untangling the complex relationships between microbes, the tumor microenvironment, and cancer cells could provide valuable insight into potential cancer treatments and the way existing cancer treatments might perform in individual cancer patients. While the gut microbiota has been extensively studied in the context of carcinogenesis and tumor response to immunotherapy (Gopalakrishnan et al., 2018; Matson et al., 2018; Routy et al., 2018), the microbiota of the lung and other less well studied body sites requires better understanding.

Questions remain to be addressed in this growing field. For example, do tumor-associated microbes originate from the local expansion of normal tissue-resident commensals, or are they recruited from elsewhere as tumors progress? If tumor-associated microbial communities represent a shift from homeostasis, how does this shift occur spatially and temporally? Do the mechanisms of commensal microbe action differ between distinct types of cancer, and how do they differ? Other than microbial products, what is the contribution of the microbial metabolite within and around the tumor? One of the key challenges in the field is to selectively manipulate the microbiota of a specific body or tumor site without affecting the gut microbiota and the relatively low microbial presence in organs like the lung (Jin et al., 2019). Overall, future study of the intratumor microbiota is needed to explore its role in cancer and to find new therapeutic strategies for cancer treatment, including targeted and individualized treatments to maximize the efficacy of anti-tumor therapies.

ACKNOWLEDGEMENTS

Figures are created with BioRender. C.J. is supported in part by a R00 Award from NIH/NCI (CA226400), an Emerson Collective Cancer Research Fund, and a Lung Cancer Research Foundation (LCRF) Pilot Grant. C.Z. is supported in part by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research, SITC-AstraZeneca Immunotherapy in Lung Cancer (Early Stage NSCLC) Clinical Fellowship Award.

ABBREVIATIONS

COPD, chronic obstructive pulmonary disease; CRC, colorectal cancer; ETBF, enterotoxigenic *Bacteroides fragilis*; GBM, glioblastoma multiforme; GF, germfree; GI, gastrointestinal; PDAC, pancreatic ductal adenocarcinoma; pks+ *E. coli, Escherichia coli* expressing the genomic island polyketide synthase; TLR, Toll-like receptor

COMPLIANCE WITH ETHICS GUIDELINES

Abigail Wong-Rolle, Haohan Karen Wei, Chen Zhao, Chengcheng Jin declare that they have no conflict of interest.

This article does not contain any studies with human or animal subjects performed by any of the authors.

OPEN ACCESS

This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creativecommons.org/licenses/by/4.0/.

REFERENCES

- Abreu MT, Peek RM Jr (2014) Gastrointestinal malignancy and the microbiome. Gastroenterology 146:1534-1546.e1533
- Arthur JC, Perez-Chanona E, Muhlbauer M, Tomkovich S, Uronis JM, Fan TJ, Campbell BJ, Abujamel T, Dogan B, Rogers AB et al (2012) Intestinal inflammation targets cancer-inducing activity of the microbiota. Science 338:120–123

- Aykut B, Pushalkar S, Chen R, Li Q, Abengozar R, Kim JI, Shadaloey SA, Wu D, Preiss P, Verma N et al (2019) The fungal mycobiome promotes pancreatic oncogenesis via activation of MBL. Nature 574:264–267
- Balachandran VP, Łuksza M, Zhao JN, Makarov V, Moral JA, Remark R, Herbst B, Askan G, Bhanot U, Senbabaoglu Y et al (2017) Identification of unique neoantigen qualities in long-term survivors of pancreatic cancer. Nature 551:512–516
- Banerjee S, Tian T, Wei Z, Shih N, Feldman MD, Peck KN, DeMichele AM, Alwine JC, Robertson ES (2018) Distinct microbial signatures associated with different breast cancer types. Front Microbiol 9:951
- Baughman RP, Thorpe JE, Staneck J, Rashkin M, Frame PT (1987) Use of the protected specimen brush in patients with endotracheal or tracheostomy tubes. Chest 91:233–236
- Belkaid Y, Hand TW (2014) Role of the microbiota in immunity and inflammation. Cell 157:121–141
- Belkaid Y, Naik S (2013) Compartmentalized and systemic control of tissue immunity by commensals. Nat Immunol 14:646–653
- Bullman S, Pedamallu CS, Sicinska E, Clancy TE, Zhang X, Cai D, Neuberg D, Huang K, Guevara F, Nelson T et al (2017) Analysis of *Fusobacterium* persistence and antibiotic response in colorectal cancer. Science 358:1443–1448
- Cameron SJS, Lewis KE, Huws SA, Hegarty MJ, Lewis PD, Pachebat JA, Mur LAJ (2017) A pilot study using metagenomic sequencing of the sputum microbiome suggests potential bacterial biomarkers for lung cancer. PLoS ONE 12:e0177062
- Castellarin M, Warren RL, Freeman JD, Dreolini L, Krzywinski M, Strauss J, Barnes R, Watson P, Allen-Vercoe E, Moore RA et al (2012) *Fusobacterium nucleatum* infection is prevalent in human colorectal carcinoma. Genome Res 22:299–306
- Charlson ES, Bittinger K, Haas AR, Fitzgerald AS, Frank I, Yadav A, Bushman FD, Collman RG (2011) Topographical continuity of bacterial populations in the healthy human respiratory tract. Am J Respir Crit Care Med 184:957–963
- Chen YE, Tsao H (2013) The skin microbiome: current perspectives and future challenges. J Am Acad Dermatol 69:143–155
- Costello EK, Lauber CL, Hamady M, Fierer N, Gordon JI, Knight R (2009) Bacterial community variation in human body habitats across space and time. Science (New York NY) 326:1694–1697
- de Martel C, Ferlay J, Franceschi S, Vignat J, Bray F, Forman D, Plummer M (2012) Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. Lancet Oncol 13:607–615
- Dejea CM, Fathi P, Craig JM, Boleij A, Taddese R, Geis AL, Wu X, DeStefano Shields CE, Hechenbleikner EM, Huso DL et al (2018)
 Patients with familial adenomatous polyposis harbor colonic biofilms containing tumorigenic bacteria. Science 359:592–597
- Dickson RP, Erb-Downward JR, Freeman CM, McCloskey L, Beck JM, Huffnagle GB, Curtis JL (2015) Spatial variation in the healthy human lung microbiome and the adapted island model of lung biogeography. Ann Am Thorac Soc 12:821–830
- Dickson RP, Erb-Downward JR, Huffnagle GB (2013) The role of the bacterial microbiome in lung disease. Expert Rev Respir Med 7:245–257

- Dickson RP, Martinez FJ, Huffnagle GB (2014) The role of the microbiome in exacerbations of chronic lung diseases. Lancet 384:691–702
- DiDonato JA, Mercurio F, Karin M (2012) NF-kappaB and the link between inflammation and cancer. Immunol Rev 246:379–400
- Dzutsev A, Badger JH, Perez-Chanona E, Roy S, Salcedo R, Smith CK, Trinchieri G (2017) Microbes and cancer. Annu Rev Immunol 35:199–228
- Elinav E, Nowarski R, Thaiss CA, Hu B, Jin C, Flavell RA (2013) Inflammation-induced cancer: crosstalk between tumours, immune cells and microorganisms. Nat Rev Cancer 13:759–771
- Erb-Downward JR, Thompson DL, Han MK, Freeman CM, McCloskey L, Schmidt LA, Young VB, Toews GB, Curtis JL, Sundaram B et al (2011) Analysis of the lung microbiome in the "healthy" smoker and in COPD. PLoS ONE 6:e16384

Garrett WS (2015) Cancer and the microbiota. Science 348:80-86

Garrett WS (2019) The gut microbiota and colon cancer. Science 364:1133–1135

- Geller LT, Barzily-Rokni M, Danino T, Jonas OH, Shental N, Nejman D, Gavert N, Zwang Y, Cooper ZA, Shee K et al (2017) Potential role of intratumor bacteria in mediating tumor resistance to the chemotherapeutic drug gemcitabine. Science 357:1156–1160
- Gomes S, Cavadas B, Ferreira JC, Marques PI, Monteiro C, Sucena M, Sousa C, Vaz Rodrigues L, Teixeira G, Pinto P et al (2019)
 Profiling of lung microbiota discloses differences in adenocarcinoma and squamous cell carcinoma. Sci Rep 9:12838
- Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, Prieto PA, Vicente D, Hoffman K, Wei SC et al (2018) Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. Science 359:97–103
- Greathouse KL, Stone JK, Harris CC (2020) Cancer-type-specific bacteria: freeloaders or partners? Cancer Cell 38:158–160
- Greathouse KL, White JR, Vargas AJ, Bliskovsky VV, Beck JA, von Muhlinen N, Polley EC, Bowman ED, Khan MA, Robles AI et al (2018) Interaction between the microbiome and TP53 in human lung cancer. Genome Biol 19:123
- Guerra L, Guidi R, Frisan T (2011) Do bacterial genotoxins contribute to chronic inflammation, genomic instability and tumor progression? FEBS J 278:4577–4588
- Gur C, Ibrahim Y, Isaacson B, Yamin R, Abed J, Gamliel M, Enk J, Bar-On Y, Stanietsky-Kaynan N, Coppenhagen-Glazer S et al (2015) Binding of the Fap2 protein of *Fusobacterium nucleatum* to human inhibitory receptor TIGIT protects tumors from immune cell attack. Immunity 42:344–355
- Gustafson AM, Soldi R, Anderlind C, Scholand MB, Qian J, Zhang X,
 Cooper K, Walker D, McWilliams A, Liu G et al (2010) Airway
 PI3K pathway activation is an early and reversible event in lung
 cancer development. Sci Transl Med 2:26ra25
- Hieken TJ, Chen J, Hoskin TL, Walther-Antonio M, Johnson S, Ramaker S, Xiao J, Radisky DC, Knutson KL, Kalari KR et al (2016) The microbiome of aseptically collected human breast tissue in benign and malignant disease. Sci Rep 6:30751
- Hilty M, Burke C, Pedro H, Cardenas P, Bush A, Bossley C, Davies J, Ervine A, Poulter L, Pachter L et al (2010) Disordered microbial communities in asthmatic airways. PLoS ONE 5:e8578

- Hoste E, Arwert EN, Lal R, South AP, Salas-Alanis JC, Murrell DF, Donati G, Watt FM (2015) Innate sensing of microbial products promotes wound-induced skin cancer. Nat Commun 6:5932
- Huffnagle GB, Dickson RP, Lukacs NW (2017) The respiratory tract microbiome and lung inflammation: a two-way street. Mucosal Immunol 10:299–306
- Human Microbiome Project Consortium (2012) Structure, function and diversity of the healthy human microbiome. Nature 486:207– 214
- lida N, Dzutsev A, Stewart CA, Smith L, Bouladoux N, Weingarten RA, Molina DA, Salcedo R, Back T, Cramer S et al (2013) Commensal bacteria control cancer response to therapy by modulating the tumor microenvironment. Science 342:967–970
- Jin C, Lagoudas GK, Zhao C, Bullman S, Bhutkar A, Hu B, Ameh S, Sandel D, Liang XS, Mazzilli S et al (2019) Commensal microbiota promote lung cancer development via gammadelta T cells. Cell 176(998–1013):e1016
- Kadosh E, Snir-Alkalay I, Venkatachalam A, May S, Lasry A, Elyada
 E, Zinger A, Shaham M, Vaalani G, Mernberger M et al (2020)
 The gut microbiome switches mutant p53 from tumour-suppressive to oncogenic. Nature 586:133–138
- Kostic AD, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, Clancy TE, Chung DC, Lochhead P, Hold GL et al (2013) *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. Cell Host Microbe 14:207–215
- Kostic AD, Gevers D, Pedamallu CS, Michaud M, Duke F, Earl AM, Ojesina AI, Jung J, Bass AJ, Tabernero J et al (2012) Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. Genome Res 22:292–298
- Le Noci V, Guglielmetti S, Arioli S, Camisaschi C, Bianchi F, Sommariva M, Storti C, Triulzi T, Castelli C, Balsari A et al (2018) Modulation of pulmonary microbiota by antibiotic or probiotic aerosol therapy: a strategy to promote immunosurveillance against lung metastases. Cell Rep 24:3528–3538
- Lee SH, Sung JY, Yong D, Chun J, Kim SY, Song JH, Chung KS, Kim EY, Jung JY, Kang YA et al (2016) Characterization of microbiome in bronchoalveolar lavage fluid of patients with lung cancer comparing with benign mass like lesions. Lung Cancer 102:89–95
- Ley RE, Peterson DA, Gordon JI (2006) Ecological and evolutionary forces shaping microbial diversity in the human intestine. Cell 124:837–848
- Liu HX, Tao LL, Zhang J, Zhu YG, Zheng Y, Liu D, Zhou M, Ke H, Shi MM, Qu JM (2018) Difference of lower airway microbiome in bilateral protected specimen brush between lung cancer patients with unilateral lobar masses and control subjects. Int J Cancer 142:769–778
- Lloyd CM, Marsland BJ (2017) Lung homeostasis: influence of age, microbes, and the immune system. Immunity 46:549–561
- Matson V, Fessler J, Bao R, Chongsuwat T, Zha Y, Alegre M-L, Luke JJ, Gajewski TF (2018) The commensal microbiome is associated with anti-PD-1 efficacy in metastatic melanoma patients. Science 359:104–108
- Mrázek J, Mekadim C, Kučerová P, Švejstil R, Salmonová H, Vlasáková J, Tarasová R, Čížková J, Červinková M (2019)

Melanoma-related changes in skin microbiome. Folia Microbiol (Praha) 64:435–442

- Nakatsuji T, Chen TH, Butcher AM, Trzoss LL, Nam SJ, Shirakawa KT, Zhou W, Oh J, Otto M, Fenical W et al (2018) A commensal strain of *Staphylococcus epidermidis* protects against skin neoplasia. Sci Adv 4:eaao4502
- Nejman D, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, Rotter-Maskowitz A, Weiser R, Mallel G, Gigi E et al (2020) The human tumor microbiome is composed of tumor type-specific intracellular bacteria. Science 368:973–980
- Norenhag J, Du J, Olovsson M, Verstraelen H, Engstrand L, Brusselaers N (2020) The vaginal microbiota, human papillomavirus and cervical dysplasia: a systematic review and network meta-analysis. BJOG 127:171–180
- Nougayrede JP, Homburg S, Taieb F, Boury M, Brzuszkiewicz E, Gottschalk G, Buchrieser C, Hacker J, Dobrindt U, Oswald E (2006) *Escherichia coli* induces DNA double-strand breaks in eukaryotic cells. Science 313:848–851
- O'Dwyer DN, Dickson RP, Moore BB (2016) The lung microbiome, immunity, and the pathogenesis of chronic lung disease. J Immunol 196:4839–4847
- Pilette C, Ouadrhiri Y, Godding V, Vaerman JP, Sibille Y (2001) Lung mucosal immunity: immunoglobulin—a revisited. Eur Respir J 18:571–588
- Pushalkar S, Hundeyin M, Daley D, Zambirinis CP, Kurz E, Mishra A, Mohan N, Aykut B, Usyk M, Torres LE et al (2018) The pancreatic cancer microbiome promotes oncogenesis by induction of innate and adaptive immune suppression. Cancer Discov 8:403–416
- Putze J, Hennequin C, Nougayrede JP, Zhang W, Homburg S, Karch H, Bringer MA, Fayolle C, Carniel E, Rabsch W et al (2009) Genetic structure and distribution of the colibactin genomic island among members of the family Enterobacteriaceae. Infect Immun 77:4696–4703
- Ramirez-Labrada AG, Isla D, Artal A, Arias M, Rezusta A, Pardo J, Galvez EM (2020) The influence of lung microbiota on lung carcinogenesis, immunity, and immunotherapy. Trends Cancer 6:86–97
- Riquelme E, Zhang Y, Zhang L, Montiel M, Zoltan M, Dong W, Quesada P, Sahin I, Chandra V, San Lucas A et al (2019) Tumor microbiome diversity and composition influence pancreatic cancer outcomes. Cell 178:795-806.e712
- Routy B, Le Chatelier E, Derosa L, Duong CPM, Alou MT, Daillere R, Fluckiger A, Messaoudene M, Rauber C, Roberti MP et al (2018)
 Gut microbiome influences efficacy of PD-1-based immunotherapy against epithelial tumors. Science 359:91–97
- Rubinstein MR, Wang X, Liu W, Hao Y, Cai G, Han YW (2013) *Fusobacterium nucleatum* promotes colorectal carcinogenesis by modulating E-cadherin/β-catenin signaling via its FadA adhesin. Cell Host Microbe 14:195–206
- Sears CL (2009) Enterotoxigenic *Bacteroides fragilis*: a rogue among symbiotes. Clin Microbiol Rev 22:349–369 (**Table of Contents**)
- Shalapour S, Karin M (2020) Cruel to be kind: epithelial, microbial, and immune cell interactions in gastrointestinal cancers. Annu Rev Immunol 38:649–671

- Shang S, Hua F, Hu Z-W (2017) The regulation of β -catenin activity and function in cancer: therapeutic opportunities. OncoTarget 8:33972–33989
- Shannon B, Yi TJ, Perusini S, Gajer P, Ma B, Humphrys MS, Thomas-Pavanel J, Chieza L, Janakiram P, Saunders M et al (2017) Association of HPV infection and clearance with cervicovaginal immunology and the vaginal microbiota. Mucosal Immunol 10:1310–1319
- Shi Y, Zheng W, Yang K, Harris KG, Ni K, Xue L, Lin W, Chang EB, Weichselbaum RR, Fu YX (2020) Intratumoral accumulation of gut microbiota facilitates CD47-based immunotherapy via STING signaling. J Exp Med 217(5):e20192282
- Siegel RL, Miller KD, Jemal A (2018) Cancer statistics, 2018. CA Cancer J Clin 68:7–30
- Siegel RL, Miller KD, Jemal A (2019) Cancer statistics, 2019. CA Cancer J Clin 69:7–34
- Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, Benyamin FW, Lei YM, Jabri B, Alegre ML et al (2015) Commensal *Bifidobacterium* promotes antitumor immunity and facilitates anti-PD-L1 efficacy. Science 350:1084–1089
- Sommariva M, Le Noci V, Bianchi F, Camelliti S, Balsari A, Tagliabue E, Sfondrini L (2020) The lung microbiota: role in maintaining pulmonary immune homeostasis and its implications in cancer development and therapy. Cell Mol Life Sci 77:2739–2749
- Thorpe JE, Baughman RP, Frame PT, Wesseler TA, Staneck JL (1987) Bronchoalveolar lavage for diagnosing acute bacterial pneumonia. J Infect Dis 155:855–861
- Tsay JJ, Wu BG, Badri MH, Clemente JC, Shen N, Meyn P, Li Y, Yie TA, Lhakhang T, Olsen E et al (2018) Airway microbiota is associated with upregulation of the PI3K pathway in lung cancer. Am J Respir Crit Care Med 198:1188–1198
- Urbaniak C, Gloor GB, Brackstone M, Scott L, Tangney M, Reid G (2016) The microbiota of breast tissue and its association with breast cancer. Appl Environ Microbiol 82:5039–5048

- Vétizou M, Pitt JM, Daillère R, Lepage P, Waldschmitt N, Flament C, Rusakiewicz S, Routy B, Roberti MP, Duong CPM et al (2015) Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. Science 350:1079–1084
- Viaud S, Saccheri F, Mignot G, Yamazaki T, Daillere R, Hannani D, Enot DP, Pfirschke C, Engblom C, Pittet MJ et al (2013) The intestinal microbiota modulates the anticancer immune effects of cyclophosphamide. Science 342:971–976
- Vitiello GA, Cohen DJ, Miller G (2019) Harnessing the microbiome for pancreatic cancer immunotherapy. Trends Cancer 5:670–676
- Wilson MR, Jiang Y, Villalta PW, Stornetta A, Boudreau PD, Carrá A, Brennan CA, Chun E, Ngo L, Samson LD et al (2019) The human gut bacterial genotoxin colibactin alkylates DNA. Science 363: eaar7785
- Yamamura K, Baba Y, Nakagawa S, Mima K, Miyake K, Nakamura K, Sawayama H, Kinoshita K, Ishimoto T, Iwatsuki M et al (2016) Human microbiome *Fusobacterium nucleatum* in esophageal cancer tissue is associated with prognosis. Clin Cancer Res 22:5574–5581
- Yan X, Yang M, Liu J, Gao R, Hu J, Li J, Zhang L, Shi Y, Guo H, Cheng J et al (2015) Discovery and validation of potential bacterial biomarkers for lung cancer. Am J Cancer Res 5:3111– 3122
- Yu G, Gail MH, Consonni D, Carugno M, Humphrys M, Pesatori AC, Caporaso NE, Goedert JJ, Ravel J, Landi MT (2016) Characterizing human lung tissue microbiota and its relationship to epidemiological and clinical features. Genome Biol 17:163
- Yu T, Guo F, Yu Y, Sun T, Ma D, Han J, Qian Y, Kryczek I, Sun D, Nagarsheth N et al (2017) *Fusobacterium nucleatum* promotes chemoresistance to colorectal cancer by modulating autophagy. Cell 170:548-563.e516