Research Article

Use of Airborne Hyperspectral Imagery to Map Soil Properties in Tilled Agricultural Fields

W. Dean Hively,¹ Gregory W. McCarty,² James B. Reeves III,³ Megan W. Lang,⁴ Robert A. Oesterling,⁵ and Stephen R. Delwiche⁶

¹U.S. Geological Survey, Eastern Geographic Science Center, Reston, VA, USA

² U.S. Department of Agriculture (USDA), Agricultural Research Service (ARS), Hydrology and Remote Sensing Laboratory, Beltsville, MD, USA

³ USDA-ARS Environmental Management and Byproducts Utilization Laboratory, Beltsville, MD, USA

⁴ USDA Forest Service, Northern Research Station, Beltsville, MD, USA

⁵ University of Maryland, Department of Geography, College Park, MD, USA

⁶ USDA-ARS Food Quality Laboratory, Beltsville, MD, USA

Correspondence should be addressed to W. Dean Hively, whively@usgs.gov

Received 10 January 2011; Revised 27 March 2011; Accepted 3 May 2011

Academic Editor: Mehrez Zribi

Copyright © 2011 W. Dean Hively et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Soil hyperspectral reflectance imagery was obtained for six tilled (soil) agricultural fields using an airborne imaging spectrometer (400–2450 nm, ~10 nm resolution, 2.5 m spatial resolution). Surface soil samples (n = 315) were analyzed for carbon content, particle size distribution, and 15 agronomically important elements (Mehlich-III extraction). When partial least squares (PLS) regression of imagery-derived reflectance spectra was used to predict analyte concentrations, 13 of the 19 analytes were predicted with $R^2 > 0.50$, including carbon (0.65), aluminum (0.76), iron (0.75), and silt content (0.79). Comparison of 15 spectral math preprocessing treatments showed that a simple first derivative worked well for nearly all analytes. The resulting PLS factors were exported as a vector of coefficients and used to calculate predicted maps of soil properties for each field. Image smoothing with a 3×3 low-pass filter prior to spectral data extraction improved prediction accuracy. The resulting raster maps showed variation associated with topographic factors, indicating the effect of soil redistribution and moisture regime on in-field spatial variability. High-resolution maps of soil analyte concentrations can be used to improve precision environmental management of farmlands.

1. Introduction

Spatial assessment of soil properties is important for understanding the dynamics of agricultural ecosystems. Site specific data can provide information that is critical to maintaining healthy soils and adequate nutrient supply for crop production, preventing losses of nutrients and sediments to the environment, and evaluating the transfer of elements such as carbon between land and atmosphere. Research has demonstrated that soil properties such as carbon content are correlated with field topography, soil texture, electrical conductivity, and soil reflectance [1–4]. A study by Venteris et al. [5] documented accumulation of carbon in low areas of fields following soil translocation from higher areas, with resulting carbon loss and soil degradation in elevated areas, and Thompson et al. [6] used soil-landscape modeling techniques to evaluate topographic distribution of soil texture and carbon content. These geographic approaches accounted for 28% to 68% of variation in measured carbon and demonstrated the complexity of environmental and management practices that affect soil characteristics. Recent research into soil health and sustainable cropping systems has demonstrated the potential of improved systems management based on knowledge of distributed soil properties [7]. Contemporary farm management relies on moderate resolution soil maps derived from photo and topographic interpretation. Accurate mapping of soil properties is made difficult due to high spatial variability observed within agricultural fields, errors in spatial assessment of soil properties can result from inadequate or biased sampling of the landscape, and the high cost associated with collecting and analyzing soil samples often limits the amount information available to farmers and land managers. However, advances in remote sensing technology are now providing tools to support geospatial mapping of soil properties, with applications in agricultural and environmental management.

Diffuse reflectance spectroscopy offers a rapid and nondestructive means for measurement of soil properties based on the reflectance spectra of illuminated soil [8-10]. A growing body of literature supports the use of spectral reflectance to determine soil properties, mostly using laboratory instrumentation to measure soil reflectance in the visible (400-700 nm) near infrared (700-2500 nm) and midinfrared (2500-25,000 nm) wavelengths. Partial least squares (PLS) regression has emerged as a successful chemometric method for extracting predictive information from spectral reflectance datasets [10-12]. The PLS method, characterizes high leverage orthogonal factors within observed spectral variance and matches them to similar factors that describe observed variance within measurements of a corresponding dependant variable. It has been successfully used to predict the results of soil laboratory analysis for carbon content [13, 14], particle size distribution [13, 15, 16] and elemental nutrient content [14, 17, 18], with results sometimes approaching the analytical accuracy of laboratory tests [12, 19]. A review of 44 studies [20] documented R² associated with prediction of soil carbon that ranged from 0.45 to 0.98, with a median of 0.86. Prediction accuracy depends on the signal: noise associated with the spectral data, and, like most analytical calibrations, is also highly influenced by the distribution of values in the measured dataset. The PLS-derived predictive equations, like most analytical calibrations, are most effective when the unknowns fall with the range of observations used to create the predictive equation, and the best success is obtained when an adequate number of locally obtained samples are included in the calibration data set [12, 21].

Advances in sensor technology have enabled satellite and airborne collection of hyperspectral imagery, allowing the acquisition of spectrally detailed geospatial reflectance data at field and landscape scales. By combining PLS regression of soil properties with reflectance data derived from airborne imagery, high-resolution maps of soil properties can be developed, thus overcoming the inaccuracies associated with geospatial interpolation of soil test data. Reports in the literature, for example, [16, 19, 22], indicate great potential for remote sensing approaches to map surface soil properties. However, additional research is needed to optimize data analysis procedures and improve prediction ability [19]. Separation of signal from noise is an important part of spectral data processing. Geometric and atmospheric adjustments are first required to derive a geospatially representative map of soil reflectance spectra. The imagery can then be smoothed spatially by averaging adjacent pixels, for example using a 3×3 low-pass filter. This can reduce the noise that results from random signal variability within the detector array while increasing the signal associated with number of

observations. It is also common to smooth the spectra in various ways, often by averaging adjacent wavebands or by calculation of first and second derivatives. Numerous math pretreatments have been evaluated for application of PLS to spectral reflectance data obtained from agricultural soils [8, 14, 23, 24].

In this paper, we have three objectives: (1) to evaluate 30 combinations of spectral math pretreatments and imagery smoothing techniques to identify most effective methods of preparing remote sensing data for partial least squares (PLS) analysis of soil properties (2) to develop and validate PLS predictions of soil concentrations for 19 laboratory analytes based on data extracted from airborne hyperspectral imagery and (3) to export resulting PLS vectors to geospatial imagery processing software and calculate high-resolution raster maps of predicted soil characteristics. Six recently tilled agricultural fields were intensively sampled to provide the calibration data set.

2. Materials and Methods

2.1. Field Sampling. On 10 April, 2007, we collected 315 soil samples from six fields (Figure 1) located on working grain farms on the Eastern Shore of the Chesapeake Bay (Delmarva Peninsula, near Easton, MD). Each of the fields (Temple 1S, 7.3 ha; Temple 1N, 7.1 ha; Temple 2, 8.9 ha; Temple 3, 18.1 ha; Mason, 14.6 ha; Schrader 9.8 ha) was chosen to provide uniform, smooth, bare-soil conditions, had been recently tilled (moldboard plow, field cultivator, and disk), and had little to no vegetation or plant residue. Soil conditions were moderately dry at the time of sampling, with six days of warm spring weather since the previous substantial rainfall (25 mm on 04 April, 2007). All fields were relatively flat (0% to 5% slope). Soil types included moderately well-drained silt loams (Pineyneck PiA, Mattapex-Butlertown MtA), poorly drained silt loams (Othello Ot), and well-drained sandy loams (Indleside IgB). Although the majority of Eastern Shore farms are managed using no-till practices, the tilled fields were otherwise typical of regional cash grain crop management strategies.

Sampling locations (315 total) were established at approximately 40 m intervals in transects across each field. Transects were established by using a tractor to pull a chisel plow shank through the soil at 20 cm depth. All sampling occurred on 10 April, 2007, during one long day of fieldwork with a large sampling crew. The soil was already well mixed from moldboard plowing, and this fresh tillage created an area of lightly disturbed soil behind the chisel plow shank from which surface soil samples (~400 g) were collected at each location. Because the chisel plow shank did not invert the soil, this sampling method approximated conditions at the bare soil surface that were observed by the imaging spectrophotometer. Number of samples ranged from 30 to 86 per field. Global positioning system (GPS) points were established for each sampling location using a handheld Trimble Geo-XT unit with submeter accuracy, calculated as the average of >20 sequential coordinate readings.

Soil samples were air dried (>48 hr) and ground to pass through a 2 mm sieve. Sand, silt, and clay content were



FIGURE 1: Field sites with soil sampling locations (black points) and SSURGO soil boundaries. See Materials and Methods for soil type designations.

determined using the hydrometer method of particle size analysis [25]. Standard soil nutrient analysis was performed at the University of Delaware Soil Testing Laboratory (http://ag.udel.edu/other_websites/DSTP/), using Mehlich III analysis [26] for potassium (K), calcium (Ca), magnesium (Mg), manganese (Mn), zinc (Zn), iron (Fe), nitrogen (N), phosphorus (P), copper (Cu), boron (B), sulfur (S), and aluminum (Al). Percent phosphorus saturation was calculated as a ratio of phosphorus to iron and aluminum content. Organic matter content (OM) was determined by loss on ignition. Subsamples were roller-milled for 12 hours prior to elemental analysis for carbon (C) and nitrogen (N) content by dry combustion using a TruSpec CN analyzer (Leco Corp, St. Joesph Mich, USA). None of the samples contained significant inorganic carbon. On the same day as field sampling for soils, airborne spectral imagery was acquired.

2.2. *Imagery Collection*. The airborne hyperspectral imaging spectrometer (HyperSpecTIR) used in this study was a push broom sensor developed by the SpecTIR Corporation (Reno Nevada, USA). It measured irradiance in 178 spectral

channels between 400 and 2450 nm with approximately 10-nm resolution [27]. Orthorectification was established to within one pixel (<2.5 m) tolerance. An upward facing radiation sensor measured incoming solar radiation which was used to calibrate imagery to ground reflectance and limit atmospheric effects to the space beneath the aircraft.

The plane was flown at 1800 m altitude with a ground speed of 210 km hr^{-1} , between 10:00 and 14:00 hrs, on 10 April, 2007. At this altitude, the imagery covered a swath 800 m wide (320 adjacent 2.5 m pixels). Flight paths were flown parallel to the principle plane of the sun. A correction for cross-track illumination was tested but did not improve results and was therefore not adopted. A number of sensor errors were identified where faulty detector elements produced erroneous results within particular wavebands, resulting in along-track striping in the field maps of predicted analyte concentrations. The 15 most obvious of these errors were corrected with spatial smoothing by replacing each faulty reflectance value with the average value of the two neighboring detector elements within the particular faulty waveband. While it was clear from visual inspection of the

predicted imagery that bad detector elements remained, only the 15 most obvious errors were corrected.

Soil spectra associated with each sampling location were extracted from the imagery by overlaying GPS point shapefiles (<1 m geopositioning error) of sampling locations and selecting the data associated with each underlying pixel. This was done for both the original imagery (1-pixel extraction) and spatially smoothed imagery to which a 3×3 low-pass filter had been applied (9-pixel extraction). Imagery processing was conducted using ENVI 4.7. Elevation data for each pixel were derived from a 3-m LIDAR digital elevation map that was resampled at 2.5 m resolution, with a vertical accuracy of 0.20 m. A 2.5 m resolution wetness index was also calculated from the LIDAR data, using SAGA software to apply two consecutive iterations of an enhanced lee 3×3 filter. This measure approximates the cumulative influence of upslope contributing area. Near infrared reflectance spectra for each of the 315 dried, ground soil samples were also obtained in the laboratory using a bench spectrometer and controlled light source, with methods and PLS results reported in McCarty et al. [28].

2.3. Spectral Data Processing. Fifteen spectral math pretreatments were evaluated using a SAS Ver. 9.12 program modified for hyperspectral data processing [23]. These included untransformed spectra, first derivatives with gap ranging from 1 to 64, and second derivatives with gap ranging from 1 to 64. Each of these math pretreatments was applied separately to the smoothed and unsmoothed spectra, resulting in a total of 30 data combinations that were used independently to calculate PLS predictions for each laboratory analyte using all 315 samples. Both the spectral data and the analyte values were mean centered prior to PLS analysis. The number of factors used in each PLS regression was chosen by the PRESS algorithm [24] within SAS Proc PLS and ranged from four to ten. Testing for outliers was not performed, and all observed values were included in the analysis. Goodness of fit was determined using repeated leave-one-out cross validation, with results presented as coefficient of determination (R^2) . Ranking analysis was employed to test for significant differences among math treatments, for all analytes predicted with $R^2 > 0.50$.

Once the best math treatment was chosen, PLS Toolbox Ver. 4.0 (Eigenvector Research, Wenatchee, Wash, USA), operating within the Matlab (Ver. 7.0) environment, was used to perform PLS analysis using mean-centered spectral and analyte data from five of the six fields (269 samples). Goodness of fit was determined using repeated leave-one-out cross validation, with results presented as coefficient of determination (R^2) and residual mean square error (RMSE). Data from the remaining field (Temple 1S, 46 samples) were treated as unknown samples and predicted from PLS coefficients, with prediction accuracy reported as bias (mean of predicted values minus mean of observed values) divided by the mean of predicted values (bias/mean(pred)) as well as standard deviation of validation [24, Section 18.8] divided by standard error of prediction (sd/se pred).

The use of Matlab allowed the mathematical flattening of the PLS factors into a 178 band vector of coefficients, and

the scalar product of this vector with each pixel's imageryderived spectrum was used to calculate predicted analyte concentrations. This calculation was made for each pixel of the hyperspectral imagery by using IDL code run within the ENVI 4.7 programming environment to calculate the appropriate band math. In this manner, geospatial field maps of predicted analyte concentrations were produced.

3. Results and Discussion

3.1. Soil Test Results. Observed analyte concentrations for the six sampled fields are described in Table 1. Soils were dry at the time of sampling, with moisture content ranging from 10% to 23% for the majority (91%) of samples (Table 1). Overall, the distribution of observed carbon content values was not large, ranging from 0.6 to 2.0% (Table 1). The Mason site, which had recently transitioned from long term conventional grain production to organic grain production, exhibited low soil carbon (mean 0.7%) content relative to the other fields (means of 1.2 to 1.4%). The Schrader site had received long-term applications of dairy manure and exhibited somewhat increased soil C (mean 1.4%) relative to the other fields. The remaining four fields (Table 1: Temple 1S, Temple 1N, Temple 2, and Temple 3) were managed by one farmer, and were under similar management (conventional corn-wheat/soybean rotation). Overall, carbon content was low, as is typical of Maryland Eastern Shore farmland. Concentrations of the remaining analytes were within the normal range for agricultural soils, although variability among sites was not great.

3.2. Choice of Math Treatment. A SAS "shotgun approach" for hyperspectral data processing has previously been used to provide a factorial comparison of the effects of various math pretreatments on PLS analysis of soil spectra [23]. This method was used to compare a total of 15 different spectral pretreatments including 1st and 2nd derivatives with various gap widths (Table 2). Results showed that there were no statistical differences among the majority of the treatments, with the simplest treatments (no derivative, first derivative gap 2) often resulting in the best fit. This led to the conclusion that the PLS data mining techniques are capable of extracting the majority of signal information from the untransformed spectral data, without the need for math pretreatment.

There is some argument to be made that different math treatments are more appropriate for particular analytes, due to the physical interaction of light with those particular constituents. Some evidence for this was shown for potassium, for which a first derivative gap 8 increased R^2 from 0.514 (first derivative gap 2) to 0.578 (Table 2). However, caution must be taken to prevent overfitting of datasets, and further study is needed to justify the selection of a diversity of math treatments for use with particular analytes. Ultimately, the first derivative gap 2 was selected as the best math overall treatment even though it was slightly, but not significantly, outperformed in several cases (Table 2).

3.3.Partial Least Squares Regression: Calibration Dataset. Once the best math treatment was decided upon, PLS regression TABLE 1: Observed laboratory analyte concentrations for the six bare soil fields that were sampled on 10 April, 2007, with number of samples collected (n). See Section 2 for analyte descriptions.

			~	5	_		33	4	~1		~	~	_		4	4	_	L_	4	S	~		6	9			4	3	_
	ssəntəW	7.2	12.	10.	1.1	7.1	13.	$10.^{-1}$	1.2	6.8	12.	9.6	1.0	5.7	12.	10.5	1.0	6.4	13.	10.	<u>.</u> ;	7.5	11.	10.0	0.5	5.7	13.	10.	1.0
	m noitsvəl∃	-5.0	2.7	0.0	1.5	-4.8	2.7	0.0	1.5	-5.0	1.5	0.0	1.2	-5.9	1.0	0.0	0.5	-2.7	2.9	0.0	1.0	-2.7	2.9	0.0	1.0	-5.9	2.9	0.0	1.1
	Moisture%	5	35	15	4	11	20	16	2	11	42	17	5	×	34	14	4	9	18	10	ŝ	9	23	15	3	ß	42	15	4
	84\gm lA	348	798	503	119	299	960	557	148	306	648	463	70	352	1049	652	188	567	1033	803	125	541	822	699	66	299	1049	598	127
	និង/ឱ្យ ខ	20.5	82.0	41.4	12.5	27.7	75.0	46.3	13.9	26.6	108.8	48.2	19.2	23.8	147.3	37.1	14.5	17.1	29.8	21.7	3.4	28.1	44.1	35.6	3.4	17.1	147.3	39.5	12.3
	ց աց/չց	0.21	0.79	0.40	0.14	0.19	0.74	0.37	0.14	0.17	0.67	0.41	0.13	0.35	4.42	0.93	0.77	0.29	0.61	0.42	0.08	0.44	0.85	0.62	0.10	0.17	4.42	0.57	0.30
	ջя/ջm uD	1.8	6.0	3.3	0.8	1.9	5.0	3.3	0.6	1.9	5.9	3.3	0.9	2.0	9.1	4.0	1.1	2.2	21.2	4.4	3.4	3.5	12.9	6.6	1.8	1.8	21.2	4.1	1.3
	P mg/kg	23	585	95	110	22	175	67	29	11	265	49	39	22	243	95	51	34	320	148	71	86	232	133	35	11	585	93	53
	N mg/kg	0.03	0.12	0.07	0.02	0.02	0.12	0.06	0.02	0.03	0.11	0.07	0.02	0.03	0.11	0.07	0.01	0.02	0.07	0.03	0.01	0.04	0.09	0.07	0.01	0.02	0.12	0.06	0.01
	gy/gm lA	327	798	501	121	299	960	559	146	306	648	463	68	352	1049	652	188	567	1033	803	125	541	822	699	66	299	1049	598	168
	និង/ឱុា ១។	97	409	205	75	115	360	207	71	129	403	231	74	91	380	183	77	71	131	102	17	120	410	177	62	71	410	190	77
	ջя/Ցա uZ	1.1	6.9	2.5	1.1	1.3	3.9	2.5	0.7	0.8	4.4	1.9	0.7	1.4	8.7	3.0	1.2	2.5	12.2	5.7	1.9	4.1	8.6	6.0	1.0	0.8	12.2	3.4	1.8
	និង/ឱុញ ៣ស្រ	8	111	34	31	6	115	32	29	4	22	10	3	8	112	34	23	20	94	51	20	22	111	52	19	4	115	34	26
	Q., Q., Q.,		5	5	•	(2	7	, c	2	0	9	~	4	8	9	+		3	3	1	~	2	0	2	10	8	9	0
	Me me/ke	55	24	14	35	69	25	13	46	10	42	25	63	13	48	22	9	75	22	13	37	6	21	16	27	55	48	18	7
	Salygm 6.	378	1214	817	171	501	1252	785	192	482	1423	953	185	615	1857	992	271	357	775	540	111	478	1066	801	113	357	1857	855	238
	84/8ա Я	94	974	247	154	83	493	226	89	43	571	164	103	71	445	210	70	43	213	111	42	253	600	449	89	43	974	236	136
	%WO	1.70	2.70	2.22	0.25	1.30	3.00	2.12	0.33	1.40	3.10	2.17	0.35	1.50	3.40	2.50	0.37	0.50	2.20	0.97	0.39	2.10	3.50	2.65	0.34	0.50	3.50	2.21	0.56
	Hq	4.6	6.4	5.7	0.4	4.7	6.3	5.3	0.4	5.4	7.0	6.3	0.4	5.6	7.2	6.3	0.3	5.7	6.8	6.3	0.3	4.9	6.8	6.1	0.4	4.6	7.2	6.0	0.5
	%yalO	3.3	12.5	9.0	2.0	5.8	16.6	9.6	2.2	3.9	17.3	9.8	2.4	6.3	19.7	12.2	2.9	1.3	10.9	6.9	2.5	6.0	18.7	10.4	2.7	1.3	19.7	10.1	2.9
	%ilis	28.9	51.8	42.4	6.3	24.6	55.7	40.9	6.7	24.7	55.4	42.5	6.2	31.4	57.9	54.9	7.1	9.5	43.4	23.7	7.9	18.0	45.9	37.7	6.7	9.5	57.9	43.1	11.2
	%pues	36.5	55.4	18.6	7.3	32.3	6.96	19.5	7.6	34.1	58.2	17.6	7.6	0.61	59.5 (32.9	8.0	1 7.6	87.7	59.4	9.7	37.2	73.9 4	51.9	8.6	19.0	87.7	46.8 ⁴	13.1
	%Э	1.07	1.63 (1.32	0.15	0.89	1.75 (1.21	0.17	0.88	1.91	1.29	0.22	1.00	2.04	1.32	0.18	0.55 4	1.40	0.75 (0.19	1.09	1.88	1.41	0.18	0.55	2.04	1.26	0.25
		u u	XI	can	Ŭ	u (XI	an	Ŭ	u	XI	can	Ŭ	u u	XI	an	Ŭ	с и	XI	can (Ŭ	ц	XI	can	0	u (XI	an	Ŭ
10110		e mi	m	6) Mt	sd	e mi	m	2) m(sd	e mi	m	5) mé	sd	e mi	m	(9) m¢	a sd	imi r	0) mí	a mé	sd	ler mi	6) mé	mé	sd	l mi	15)mé	a mé	sd
diment	Field	Templ	1S	(n = 4	7.3 ha	Templ	lN	(n = 5)	7.1 ha	Templ	5	(n = 5)	8.9 ha	Templ	3	(n = 8)	18.1 hí	Masor	(n = 3)	14.6 hi		Schrac	(n = 4	9.8 ha		Overal	(n = 3)	65.8 hi	

TABLE 2: Partial least squares (PLS) prediction model goodness of fit (R^2) associated with each of 15 math treatments, for the 13 analytes that predicted with $R^2 > 0.5$, calculated using data from all 315 soil sampling locations¹. The first derivative gap two (1Dg2, depicted in bold) was selected as the overall most successful model although it was occasionally outperformed (italic). See Section 2 for analyte descriptions.

Derivative	Gap	С	Sand	Silt	Clay	pН	ОМ	Κ	Са	Mg	Mn	Zn	Fe	Al	average
		R^2													
NON	0	0.578	0.762	0.763	0.585	0.442	0.685	0.555	0.669	0.692	0.642	0.670	0.754	0.777	0.659
1ST	1	0.555	0.770	0.761	0.565	0.517	0.706	0.522	0.654	0.719	0.624	0.606	0.707	0.799	0.654
1ST	2	0.591	0.763	0.763	0.617	0.549	0.717	0.514	0.676	0.708	0.638	0.647	0.740	0.782	0.670
1ST	4	0.595	0.754	0.740	0.596	0.451	0.692	0.513	0.630	0.616	0.565	0.675	0.703	0.773	0.639
1ST	8	0.584	0.748	0.739	0.600	0.413	0.668	0.578	0.635	0.642	0.580	0.636	0.637	0.727	0.630
1ST	16	0.588	0.740	0.744	0.550	0.442	0.672	0.547	0.610	0.619	0.567	0.591	0.666	0.725	0.620
1ST	32	0.525	0.730	0.722	0.554	0.381	0.640	0.520	0.648	0.619	0.542	0.580	0.669	0.765	0.607
1ST	64	0.338	0.623	0.619	0.470	0.317	0.537	0.413	0.519	0.580	0.488	0.568	0.636	0.669	0.521
2ND	1	0.542	0.585	0.597	0.424	0.361	0.526	0.484	0.708	0.705	0.529	0.599	0.665	0.729	0.573
2ND	2	0.507	0.681	0.682	0.372	0.499	0.566	0.493	0.665	0.664	0.566	0.591	0.673	0.769	0.595
2ND	4	0.576	0.714	0.724	0.409	0.449	0.660	0.484	0.672	0.669	0.606	0.613	0.749	0.752	0.621
2ND	8	0.536	0.698	0.689	0.533	0.472	0.602	0.528	0.595	0.631	0.542	0.638	0.743	0.753	0.612
2ND	16	0.580	0.729	0.725	0.573	0.487	0.661	0.498	0.590	0.640	0.598	0.617	0.680	0.712	0.622
2ND	32	0.564	0.715	0.698	0.609	0.432	0.631	0.548	0.622	0.592	0.548	0.606	0.639	0.688	0.607
2ND	64	0.525	0.596	0.589	0.421	0.156	0.599	0.511	0.556	0.464	0.501	0.538	0.602	0.614	0.513
1			2.												

¹Analytes that predicted poorly R^2 (<0.5) included: N (<0.303), P (<.355), Cu (<0.358), B (<0.169), S (<0.282), and P saturation (<0.127).

was performed in Matlab using a calibration dataset of 269 samples (all sampling locations from five of the six fields), with results reported in Table 3. It should be noted that spectral data for the 315 samples used to calculate the SAS PLS in Table 2 were extracted from an earlier version of image processing output in which the 15 band errors had not vet been corrected and to which cross-track illumination had been applied. This discrepancy resulted in slightly lower R^2 being used in Table 2 than in Table 3, but was not thought to have affected the relative performance of math treatments. Using Matlab PLS on 269 samples, 13 of the 19 analytes were predicted with $R^2 > 0.50$ (Table 3(a)), and the remaining seven were predicted with $R^2 < 0.40$: N (<0.30), P (<0.36), Cu (<0.36), B (<0.17), S (<0.28), and P saturation (<0.13). A comparison of observed and predicted values for a selection of analytes (carbon, silt, aluminum, and iron) is shown in Figure 2.

Prediction accuracy for carbon (R^2 =0.65) fell well within the range of results (0.45 to 0.98) found in a survey of 44 studies [20] but was somewhat poor in comparison with results found in some other studies for example, [11, 14, 15]. The somewhat poor R^2 for carbon might lead to the conclusion that the sensor did not capture a good reflectance signal due to the increased noise often associated with airborne sensors that derives from atmospheric effects and variations in sensor-soil-sun geometry across the imagery. Indeed, a number of studies have found remote sensing spectroscopy to have reduced signal : noise relative to laboratory-based measurements [24]. However, the carbon PLS results reported in McCarty et al. [28], calculated for all 315 soil samples in this data set, using repeated leaveone-out correlation to determine goodness of fit, showed that spectra from the airborne sensor were as effective $(R^2 = 0.67)$ as spectra from a near-infrared benchtop laboratory spectrophotometer $(R^2 = 0.64)$ in predicting soil carbon concentrations. For the remaining 12 analytes under consideration, the airborne sensor exhibited decreased accuracy relative to the laboratory spectrophotometer in three cases (change in R^2 of -0.03 to -0.08) and exhibited increased accuracy in nine cases (change in R^2 of 0.01 to 0.20).

These observations led to the conclusion that the airborne sensor provides a viable option for mapping soil properties and that the somewhat poor prediction accuracies observed in this experiment apparently stemmed from features associated with the local soil environment or the calibration dataset, rather than the effectiveness of the airborne sensor. The distribution and range of observed analyte concentrations within a calibration dataset can have a substantial impact on prediction accuracy, and the somewhat low prediction accuracy for carbon content observed in this study ($R^2 = 0.65$) is likely associated with the limited range and low values of soil carbon contents found within the calibration dataset (0.6% to 2.0%).

For the other analytes, prediction accuracies (Table 3(a)) were comparable with those found in other studies [8, 15, 17, 18]. Using principal components regression, Chang et al. [18] predicted Melich III analyte concentrations, and, similarly to this study, found Cu and P to have poor predictions, silt and clay content to predict with $R^2 \sim 0.8$, and a number of other analytes, including Fe, Mg, Mn, K, and pH, predicting with $R^2 > 0.6$. It is not always known whether successful predictions are the direct action of the

TABLE 3: Partial least squares (PLS) model accuracy in predicting soil analyte concentrations¹ for (a) the 269 calibration samples using repeated leave-one-out cross validation and (b) 46 validation samples from the field (Temple 1S) that had been left apart from the calibration. Results were derived using a first derivative gap 2 math pretreatment, using spectra derived from unsmoothed imagery (2.5 m^2 pixel size) or from spatially smoothed imagery to which a 3×3 low pass filter had been applied. See Section 2 for analyte descriptions. Units refer to analyte residual mean squared error (RMSE) values. Bold indicates sd/sepred >1.20.

	С%	Sand%	Silt%	Clav%	рH	OM%	K	Ca	Mg	Mn	Zn	Fe	Al
Factors no.	0,0	ouriu / o	one, o	Giuj /o	P**	011170	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg	mg/kg
	8	9	10	9	10	10	4	8	9	10	8	10	10
(a) 269 calibration same	ples colle	cted from	five agr	ricultural	fields								
using unsmoothed in	nagery												
R^2	0.65	0.79	0.79	0.66	0.51	0.75	0.59	0.69	0.69	0.62	0.64	0.75	0.76
RMSE	0.19	7.9	6.9	2.2	0.4	0.4	89.5	166.1	50.3	19.6	1.4	49.3	104.7
using smoothed imag	gery												
R^2	0.64	0.80	0.80	0.68	0.58	0.77	0.61	0.71	0.71	0.67	0.67	0.78	0.81
RMSE	0.18	7.2	6.2	2.0	0.4	0.3	86.1	151.8	45.5	17.8	1.2	43.8	89.0
(b) 46 validation samples	collected	l from the	e remain	ing agric	ultural fi	eld (Tem	ple 1S)						
using unsmoothed ima	igery												
² se pred	0.20	9.0	7.3	2.4	0.5	0.4	147.1	178.1	51.4	27.5	1.2	52.9	123.9
³ bias	0.10	-2.88	1.42	1.00	0.03	0.34	24.92	40.45	31.84	3.23	0.45	2.12	107.33
bias/mean (pred)	0.08	-0.06	0.03	0.11	0.01	0.15	0.10	0.05	0.22	0.09	0.18	0.01	0.21
⁴ sd/se pred	0.74	0.81	0.86	0.84	0.89	0.63	1.05	0.96	0.76	1.15	0.92	1.41	0.98
using smoothed image	ery												
se pred	0.15	9.5	7.9	2.9	0.4	0.3	154.9	233.6	58.5	38.3	1.3	98.0	170.4
bias	-0.10	-0.87	-0.66	0.95	-0.25	-0.13	-23.06	-2.46	-2.07	-0.38	0.10	-5.26	77.66
bias/mean (pred)	-0.08	-0.02	-0.02	0.10	-0.05	-0.06	-0.10	0.00	-0.02	-0.01	0.04	-0.03	0.14
sd/se pred	0.86	1.29	1.26	1.07	0.97	0.74	1.04	1.34	1.41	1.25	0.98	1.91	1.29

¹Analytes that predicted poorly ($R^2 < 0.5$) in the set of 315 samples (N, P, Cu, B, S, and P saturation) are not included here

²se pred: sd(observed-predicted)

³bias: mean(pred)-mean(obs)

⁴sd: standard deviation.

analyte upon the reflectance signal or instead the covariation of analyte concentrations with other factors which are influencing the spectral response [29]. Phosphorus, for example, has no expected reflectance resonance and is generally poorly predicted (e.g., 0.36 in this study, 0.40 in [18], and as low as 0.10 in other studies [8, 29]), and yet occasionally will be predicted with considerable accuracy [8, 29], likely due to covariation with spectrally responsive factors associated with labile organic matter.

3.4. Validation. Chemometric predictions of soil properties are typically validated by predicting analyte concentrations for samples that were not included in the calibration data set, and comparing the predicted results to observed values. This can be achieved by calculating repeated leave-oneout or leave-ten-out predictions, or by selecting a distinct set of samples either randomly, spatially, or based on even sample distribution within the range of observed values [24, Section 18]. We chose to adopt a rigorous validation by removing an entire agricultural field comprising 46 samples (Temple 1S) from the data set, leaving 269 samples for calibration. This field was one of four Temple fields that received similar crop management (corn-soy/wheat rotation with full tillage and no use of cover crops) and it exhibited analyte concentrations in the mid-range of the six fields (Table 1, Figure 2). When predicted values were calculated for the 46 validation samples and compared to observed values, bias ranged from 1% to 22% of observed analyte concentrations (Table 3(b)). Seven of the analytes exhibited sd/se pred >1.20 (bold text in Table 3(b)) using the spectra extracted from smoothed imagery, indicating that the prediction was useful for those elements (sand, silt, Ca, Mg, Mn, Fe, and Al), while only one analyte (Fe) exhibited sd/se pred > 1.20 using the spectra extracted from unsmoothed imagery, indicating that spatial smoothing helped to increase signal : noise in the spectral data set.

3.5. Prediction Maps. For each analyte, a PLS regression vector of 178 coefficients was exported from Matlab and applied to the mean-centered first derivative reflectance spectra associated with each pixel of the hyperspectral imagery to calculate a 2.5-m raster map of predicted analyte concentrations. Prediction maps for a selection of analytes (carbon, silt, iron, and aluminum) were derived in this manner using both unsmoothed and smoothed imagery (Figure 3). Ben-Dor et al. [19] have discussed the need for attention to imagery processing methods to extract maximum information from spectroscopic imagery. In this study, smoothing



FIGURE 2: Comparison of observed analyte concentrations with values predicted using partial least squares (PLS) regression on reflectance spectra extracted from smoothed imagery to which a 3×3 low-pass filter had been applied, for (a) soil carbon, (b) silt, (c) iron, and (d) aluminum content. The field containing 41 sampling points that were left out for validation (Temple 1S) is depicted with solid circles.

the hyperspectral imagery with a 3×3 low band filter prior to spectral data extraction resulted in somewhat improved prediction accuracies in comparison to unsmoothed imagery (Table 3(a)). While this result implies that the smoothed extent of nine adjacent pixels (56.2 m²) provided a better average representation of soil characteristics at each sampling point than did individual 6.25 m² pixels, it is more likely that the increased accuracy associated with the smoothed imagery is attributable to improvement of signal and reduction in sensor detector element variability (noise) that results from coaddition of the nine adjacent spectra. Smoothing also generally improved the range and distribution of predicted analyte values found within each field image (Table 4), likely due to the correction of aberrations within the detector elements. However, it should be noted that discontinuities (striping) along the plant's flight path were evident in both smoothed and unsmoothed predicted imagery (Figure 3), indicating the effects of variable detector element sensitivity



(b)

FIGURE 3: Maps of predicted soil carbon content calculated from (a) unsmoothed imagery (1-pixel data extraction) and (b) spatially smoothed imagery (9-pixel data extraction). Two of the six sampled fields are depicted here, with Temple 3 on the left and Schrader on the right. Predicted values ranged from 0.4% (black) to 2.5% (white).

along the sensor array and the need for continued improvement in sensor technology. Several additional imagery adjustments were considered, including cross-track illumination, transformation from reflectance (R) to log(1/R), and also spectral smoothing (not shown). However, none of these operations improved results and they were therefore avoided.

3.6. Topographic Analysis. Visual interpretation of field topographic features relative to the predicted analyte maps revealed accumulation of carbon, silt, iron, and aluminum in low areas (Figure 4). This result is in agreement with previous studies in Iowa that have linked increased soil C carbon contents in low areas with soil redistribution processes [3], and with a study by Terra et al. [1] who detected a correlation of soil properties with topographic indices and electrical conductivity that explained 50% of observed variability in car-

bon content. Other studies have evaluated the links between topography and soil constituents such as organic carbon at the landscape scale, showing accumulation at the bottom of slopes and in valley bottoms [30]. This study's work was conducted at the field scale, reflecting microtopographic soil distribution processes within a relatively flat elevation gradient (0%–5% slope), as well as the formative effects of soil moisture distribution and spatially variable biological processes resulting from soil heterogeneity and the balance of redoxomorphic status between oxic and anoxic conditions. Understanding distribution patterns of soil analytes, and the influence of hydrogeomorphic controls on these patterns, can provide greater certainty about the influence of soil erosion and soil ecology on the fate of carbon and nutrients.

To investigate potential causes of spatial distribution in predicted analyte values, a 2.5 m elevation map (re-sampled

TABLE 4: Predicted concentrations of select analytes (C, Silt, Al, Fe) derived from *n* pixels of near infrared hyperspectral imagery for each of the six bare soil fields using either unsmoothed (1-pixel) imagery with 2.5 m resolution, or spatially smoothed (9-pixel) imagery to which a 3×3 low pass filter had been applied¹. See Section 2 for analyte descriptions.

		(С	S	ilt	1	A1	Η	Fe
Field		1-pixel%	9-pixel%	1-pixel%	9-pixel%	1-pixel mg/kg	9-pixel mg/kg	1-pixel mg/kg	9-pixel mg/kg
Temple 1S	min	0.97	1.13	14.6	14.4	34	245	40	56
n = 11731	max	1.86	1.94	80.1	58.7	1017	947	474	394
	mean	1.41	1.48	42.1	41.1	633	631	194	192
	sd	0.10	0.10	5.9	5.0	98	92	46	44
Temple 1N	min	0.85	0.96	1.0	5.6	-31	147	37	64
n = 11436	max	1.95	2.04	68.9	59.1	1304	897	452	429
	mean	1.23	1.25	43.6	42.1	589	555	200	210
	sd	0.11	0.11	5.8	4.9	115	112	60	65
Temple 2	min	-0.48	0.81	14.6	8.7	34	-389	40	54
n = 14221	max	2.07	2.92	80.1	75.7	1017	1013	474	529
	mean	1.27	1.28	42.1	38.2	633	512	194	226
	sd	0.25	0.18	5.9	7.1	98	146	46	63
Temple 3	min	0.39	0.60	0.2	1.2	-137	0	-288	-6
<i>n</i> = 28955	max	2.29	2.48	80.8	74.9	1173	1031	465	474
	mean	1.27	1.27	52.3	51.7	632	639	173	170
	sd	0.17	0.16	7.9	7.5	154	145	71	63
Mason	min	0.41	0.44	-7.6	-7.5	-198	290	0	0
<i>n</i> = 23351	max	2.27	1.94	64.9	48.5	1231	1180	391	391
	mean	0.81	0.81	27.7	28.7	755	776	111	108
	sd	0.11	0.10	6.4	7.1	96	100	32	27
Schrader	min	0.91	0.98	12.3	13.4	255	296	58	66
n = 15684	max	2.08	2.05	61.6	56.9	996	971	346	315
	mean	1.45	1.48	38.6	38.3	669	681	177	172
	sd	0.13	0.15	6.0	5.8	87	81	35	34
Overall	min	0.97	1.13	14.6	14.4	255	296	58	66
n = 105378	max	1.86	1.94	61.6	48.5	996	897	346	315
	mean	1.20	1.22	41.3	40.5	660	648	168	171

¹See Table 3 for the accuracy (R^2) associated with each analyte prediction model.

TABLE 5: Correlation between predicted analyte concentrations and topographic indices including (a) relative elevation data derived from 3 m LIDAR DEM, and (b) wetness index calculated from 3 m LIDAR DEM, for one of the six sampled fields (Mason). Predicted values were calculated for unsmoothed imagery (2.5 m pixel size) and for smoothed imagery to which a 3×3 low band filter had been applied. See Section 2 for analyte descriptions.

	С	Silt	Al	Fe
	R^2	R^2	R^2	R^2
(a) correlation with normalized	d elevation			
unsmoothed	0.100	0.000	0.232	0.013
smoothed	0.051	0.007	0.386	0.000
(b) correlation with wetness in	dex			
unsmoothed	0.183	0.008	0.161	0.091
smoothed	0.178	0.021	0.317	0.037





FIGURE 4: Map of predicted values for select analytes (C, Silt, Fe, Al), overlaid on a high-resolution digital elevation map, for one of the six sampled fields (Mason). Predicted values were derived from imagery that had been smoothed with a 3×3 low pass filter.

from 3-m LIDAR DEM) was used to calculate a normalized elevation value (observed-field mean elevation) and a soil wetness index (two consecutive iterations of a SAGA enhanced lee 3×3 filter) for each pixel of the predicted analyte maps. The normalized elevation variable was somewhat correlated (Table 5(a)) with predicted concentrations of aluminum ($R^2 = 0.23$, 0.39 for unsmoothed and spatially smoothed imagery, respectively) and was not well correlated with predicted concentrations of silt, iron, and carbon. The wetness index was similarly correlated with predicted aluminum concentration ($R^2 = 0.16, 0.32$) and was a better predictor for soil carbon, explaining 17% of observed variability (Table 5(b)). In all cases, the topographic variables were better correlated with spatially smoothed predictions than those made from unsmoothed imagery. The comparatively poor correlation with topographic indices observed in this study might indicate that soil carbon and elemental nutrient content is more related to variability in field management and manure application than to elevation and soil redistribution, or that a more complex set of topography-influenced ecological and physical processes than was measured is at play. On low relief (<2% slope), ditch drained, Coastal Plain

soils such as were sampled in this study, soil redistribution due to erosion is likely limited to short distances (<1–10 m), and prior converted wetland areas within fields can exhibit retained capacity for redoxomorphic activities that affect iron and aluminum transformations and increase denitrification and carbon accumulation. In this landscape, spatial distribution of soil carbon and nutrient content is likely influenced by a complex interaction of drainage status and land use history that is not easily characterized [5], and ultimately it may be more feasible to map surficial soil parameters using remote sensing technology than to predict them based on measurement and understanding of site specific processes.

Soil moisture can become an interfering variable in remote sensing chemometric analyses, as it mutes the soil's reflectance signal in a number of specific wavebands that are associated with water absorption bands [31]. Soil moisture for the 315 samples included in this study ranged between 5% and 42%, with most samples falling near the mean of 15% moisture. When observed moisture content was incorporated with the first derivative spectra of soil samples as a 177th predictor variable (following appropriate mean centering and variation normalization), the prediction accuracies associated with PLS models for C, silt, Al, and Fe did not improve (change in R^2 of <0.02). Similarly, when silt was included as an additional predictor variable, to possibly account for interferences between soil particle size distribution and spectral reflectance, the prediction accuracies associated with PLS models for C, silt, Al, and Fe did not improve. These results verify that PLS analysis is a robust method that can successfully extract predictive information from remotely sensed imagery of *in situ* agricultural soils.

4. Conclusions

Aircraft-based acquisition of hyperspectral reflectance imagery can currently provide the necessary data to map soil properties in an efficient and rapid manner, and future improvements in sensor technology are expected to improve the signal: noise and spatial resolution associated with remote sensing imagery. Results of this remote sensing study, conducted on relatively flat, moderately well drained, Coastal Plain silt loam soils, showed that the PLS framework is robust, and spectral math pretreatments can be kept simple, with a first derivative gap 2 providing good results for all analytes. Spatial smoothing of reflectance imagery using a 3×3 low-band filter improved results, likely by reducing noise inherent to variability within the spectrophotometer's detector array. Fifteen of the 19 tested analytes predicted with $R^2 > 0.50$ (with R^2 from 0.51 to 0.79 for C, sand, silt, clay, pH, OM, K, Ca, Mg, Mn, Zn, Fe, and Al content; Table 3). Prediction accuracy for carbon was not particularly good ($R^2 = 0.64$) for this selection of sampled fields, perhaps owing to the low concentrations and limited range of observed variability (0.5% to 2%) in the calibration data set, but prediction accuracies for other analytes were on par with those found in other studies [8, 15, 18]. Predicted map values of select analytes, particularly aluminum, were correlated to field topography, indicating the influence of environmental processes on soil properties. Further research is needed to understand the interplay of measurable topographic and hydrological variation inherent in this relatively flat agricultural landscape with management variables such as tillage, crop rotation, and additions of manures and fertilizers that also affect soil chemical and physical properties.

Remote sensing approaches that use multispectral or hyperspectral imagery to predict soil properties are necessarily limited to analysis of plowed fields with very little crop residue or vegetation cover (bare soil). As a result, techniques that extrapolate soil properties data based on correlations derived from more easily measured parameters, such as topography, wetness, and cropping pattern may provide. The fields used in this study were carefully selected to have negligible amounts of surface residue, green vegetation, or hydrologically active areas. If this technique were put in practice on a larger scale, band filtering could be employed to remove such non-soil areas from analysis based on thresholding of spectral indices that correlate well with biomass (e.g., normalized difference vegetation index [32]), crop residue (e.g., cellulose adsorption index [33]), or soil moisture (e.g., water adsorption [13, 31]).

As hyperspectral imagery becomes more readily available and at lower cost, the application of partial least squares (PLS) regression to soil spectral reflectance data can provide an effective method for calculating high-resolution raster maps of important soil properties including texture, pH, and carbon and nutrient content. This information can then be used to inform farmer decision making, support precision environmental management of agricultural fields, increase sustainable crop production, and help to reduce nutrient, sediment, and carbon losses from agricultural systems.

Acknowledgments

This paper has benefitted greatly from the comments of two anonymous reviewers, and the project is also indebted to input from known scientific colleagues, with thanks going out to Drs. Craig Daughtry, Terry Slonecker, Guy Serbin, and the late Paul Doraiswamy. Technical assistance was provided by Kusuma Prabhakara and Dr. Beth Gardner, and laboratory and field work by Alex White and Antonio Pereira. Thanks to the University of Delaware Soil Test Laboratory for efficient sample processing. Thanks to Oliver Weatherbee and the SpecTIR corporation for airborne imagery acquisition. Support for the this project was provided by the USDA Choptank River Conservation Effects Assessment Project. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

References

- J. A. Terra, J. N. Shaw, D. W. Reeves, R. L. Raper, E. Van Santen, and P. L. Mask, "Soil carbon relationships with terrain attributes, electrical conductivity, and a soil survey in a coastal plain landscape," *Soil Science*, vol. 169, no. 12, pp. 819–831, 2004.
- [2] G. C. Simbahan and A. Dobermann, "Sampling optimization based on secondary information and its utilization in soil carbon mapping," *Geoderma*, vol. 133, no. 3-4, pp. 345–362, 2006.
- [3] J. C. Ritchie, G. W. McCarty, E. R. Venteris, and T. C. Kaspar, "Soil and soil organic carbon redistribution on the landscape," *Geomorphology*, vol. 89, no. 1-2, pp. 163–171, 2007.
- [4] U. W. A. Vitharana, M. Van Meirvenne, D. Simpson, L. Cockx, and J. De Baerdemaeker, "Key soil and topographic properties to delineate potential management classes for precision agriculture in the European loess area," *Geoderma*, vol. 143, no. 1-2, pp. 206–215, 2008.
- [5] E. R. Venteris, G. W. McCarty, J. C. Ritchie, and T. Gish, "Influence of management history and landscape variables on soil organic carbon and soil redistribution," *Soil Science*, vol. 169, no. 11, pp. 787–795, 2004.
- [6] J. A. Thompson, E. M. Pena-Yewtukhiw, and J. H. Grove, "Soil-landscape modeling across a physiographic region: topographic patterns and model transportability," *Geoderma*, vol. 133, no. 1-2, pp. 57–70, 2006.
- [7] O. J. Idowu, H. M. Van Es, G. S. Abawi et al., "Use of an integrative soil health test for evaluation of soil management impacts," *Renewable Agriculture and Food Systems*, vol. 24, no. 3, pp. 214–224, 2009.
- [8] R. A. Viscarra Rossel, D. J. J. Walvoort, A. B. McBratney, L. J. Janik, and J. O. Skjemstad, "Visible, near infrared, mid infrared or combined diffuse reflectance spectroscopy for simultaneous

assessment of various soil properties," *Geoderma*, vol. 131, no. 1-2, pp. 59–75, 2006.

- [9] J. B. Reeves III, G. W. McCarty, and W. D. Hively, "Midversus near-infrared spectroscopy for on-site analysis of soils," in *Proximal Soil Sensing*, R. A. Viscarra Rossel, A. McBratney, and B. Minasny, Eds., chapter 11, pp. 133–142, Springer, New York, USA, 2010.
- [10] D. J. Brown, K. D. Shepherd, M. G. Walsh, M. D. Mays, and T. G. Reinsch, "Global soil characterization with VNIR diffuse reflectance spectroscopy," *Geoderma*, vol. 132, no. 3-4, pp. 273–290, 2006.
- [11] H. M. Bartholomeus, M. E. Schaepman, L. Kooistra, A. Stevens, W. B. Hoogmoed, and O. S. P. Spaargaren, "Spectral reflectance based indices for soil organic carbon quantification," *Geoderma*, vol. 145, no. 1-2, pp. 28–36, 2008.
- [12] D. J. Brown, R. S. Bricklemyer, and P. R. Miller, "Validation requirements for diffuse reflectance soil characterization models with a case study of VNIR soil C prediction in Montana," *Geoderma*, vol. 129, no. 3-4, pp. 251–267, 2005.
- [13] C. W. Chang, D. A. Laird, and C. R. Hurburgh, "Influence of soil moisture on near-infrared reflectance spectroscopic measurement of soil properties," *Soil Science*, vol. 170, no. 4, pp. 244–255, 2005.
- [14] G. W. McCarty, J. B. Reeves, V. B. Reeves, R. F. Follett, and J. M. Kimble, "Mid-infrared and near-infrared diffuse reflectance spectroscopy for soil carbon measurement," *Soil Science Society of America Journal*, vol. 66, no. 2, pp. 640–646, 2002.
- [15] G. W. McCarty and J. B. Reeves, "Comparison of near infrared and mid infrared diffuse reflectance spectroscopy for fieldscale measurement of soil fertility parameters," *Soil Science*, vol. 171, no. 2, pp. 94–102, 2006.
- [16] T. Selige, J. Böhner, and U. Schmidhalter, "High resolution topsoil mapping using hyperspectral image and field data in multivariate regression modeling procedures," *Geoderma*, vol. 136, no. 1-2, pp. 235–244, 2006.
- [17] J. Wetterlind, B. Stenberg, and M. Söderström, "Increased sample point density in farm soil mapping by local calibration of visible and near infrared prediction models," *Geoderma*, vol. 156, no. 3-4, pp. 152–160, 2010.
- [18] C. W. Chang, D. A. Laird, M. J. Mausbach, and C. R. Hurburgh, "Near-infrared reflectance spectroscopy—principal components regression analyses of soil properties," *Soil Science Society of America Journal*, vol. 65, no. 2, pp. 480–490, 2001.
- [19] E. Ben-Dor, S. Chabrillat, J. A. M. Demattê et al., "Using imaging spectroscopy to study soil properties," *Remote Sensing of Environment*, vol. 113, no. 1, pp. 538–555, 2009.
- [20] G. M. Vasques, S. Grunwald, and J. O. Sickman, "Comparison of multivariate methods for inferential modeling of soil carbon using visible/near-infrared spectra," *Geoderma*, vol. 146, no. 1-2, pp. 14–25, 2008.
- [21] C. Guerrero, R. Zornoza, I. Gómez, and J. Mataix-Beneyto, "Spiking of NIR regional models using samples from target sites: effect of model size on prediction accuracy," *Geoderma*, vol. 158, no. 1-2, pp. 66–77, 2010.
- [22] C. Gomez, R. A. Viscarra, and A. B. McBratney, "Soil organic carbon prediction by hyperspectral remote sensing and field vis-NIR spectroscopy: an Australian case study," *Geoderma*, vol. 146, pp. 403–411, 2008.
- [23] J. B. Reeves and S. R. Delwiche, "SAS partial least squares regression for analysis of spectroscopic data," *Journal of Near Infrared Spectroscopy*, vol. 11, no. 6, pp. 415–431, 2003.
- [24] "ASTM International. Standard practices for infrared multivariate quantitative analysis," E 1655-05, ASTM International, West Cronshocken, Pa, USA, 2005.

- [25] G. W. Gee and J. W. Bauder, "Particle Size Analysis," in Methods of Soil Analysis, Part 1: Physical and Minearological Methods, A. Klute, Ed., pp. 404–408, American Society of Agronomy, Madison, Wisconsin, USA, 2nd edition, 1986.
- [26] A. Mehlich, "Mehlich 3 soil test extractant: a modification of Mehlich 2 extractant," *Communications in Soil Science and Plant Analysis*, vol. 15, no. 12, pp. 1409–1416, 1984.
- [27] C. M. Jengo and J. LaVeigne, "Sensor performance comparison of hyperspecTIR instruments 1 and 2," in *Proceedings of IEEE Aerospace Conference*, pp. 1799–1805, March 2004.
- [28] G. W. McCarty, W. D. Hively, J. B. Reeves, M. Lang, E. Lund, and O. Weatherbee, "Infrared sensors to map soil carbon in agricultural ecosystems," in *Proximal Soil Sensing*, R. A. Viscarra Rossel, Ed., vol. 1 of *Progress in Soil Science*, pp. 165–176, Springer, 2010.
- [29] B. Stenberg and R. A. Viscarra Rossel, "Diffuse reflectance spectroscopy for high-resolution soil sensing," in *Proximal Soil Sensing*, R. A. Viscarra Rossel, A. M. McBratney, and B. Minasny, Eds., Progress in Soil Science Book Series, Chapter 3, Springer, New York, NY, USA, 2010.
- [30] W. Schwanghart and T. Jarmer, "Linking spatial patterns of soil organic carbon to topography—a case study from southeastern Spain," *Geomorphology*, vol. 126, pp. 252–263, 2011.
- [31] C. Y. Wu, A. R. Jacobson, M. Laba, and P. Baveye, "Alleviating moisture content effects on the visible near-infrared diffusereflectance sensing of soils," *Soil Science*, vol. 174, no. 8, pp. 456–465, 2009.
- [32] C. Tucker, "Red and photographic infrared linear combinations for monitoring vegetation," *Remote Sensing of Environment*, vol. 8, no. 2, pp. 127–150, 1979.
- [33] G. Serbin, C. S. T. Daughtry, E. R. Hunt, J. B. Reeves, and D. J. Brown, "Effects of soil composition and mineralogy on remote sensing of crop residue cover," *Remote Sensing of Environment*, vol. 113, no. 1, pp. 224–238, 2009.





The Scientific World Journal Journal of Environmental and Public Health











International Journal of Atmospheric Sciences



Submit your manuscripts at http://www.hindawi.com



International Journal of Oceanography





International Journal of Biodiversity





Journal of Computational Environmental Sciences









Environmental Soil Science



Advances in Meteorology



Journal of Climatology



